



Habitat Partitioning in the Marine Sector of Karst Subterranean Estuaries and Bermuda's Marine Caves: Benthic Foraminiferal Evidence

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Karst subterranean estuaries (KSEs) are created from the two- and three-way mixing of saline groundwater, rain, and oceanic water in the subsurface on carbonate landscapes, and this hydrographic framework promotes unique physical processes, biogeochemical cycling, and biological communities. Here we provide evidence that the source and quantity of particulate organic matter (POM) that is delivered to the benthos strongly correlates to benthic habitat partitioning in the oxygenated marine sectors of KSEs. A dataset of benthic foraminifera at 128 different locations from several large flooded cave systems in Bermuda were compiled and evaluated against common environmental characteristics (e.g., tidal exposure, substrate particle size, bulk organic matter, C:N, total organic carbon, and $\delta^{13}C_{org}$). Benthic areas receiving more carbon isotopically depleted organic matter sources (mean $\delta^{13}C_{org}$ values < -23.2%, C:N ratios >11), most likely from the terrestrial surface and some marine plankton, were dominated by Trochammina inflata, Bolivina spp., and Helenina anderseni. In contrast, benthic areas receiving more carbon isotopically enriched organic matter sources (mean $\delta^{13}C_{org}$ values > -21.6‰, C:N ratios <10), most likely from marine plankton transported through marine cave openings cave from adjacent coastal waters, were dominated by Spirophthalmidium emaciatum, Spirillina vivipara, Patellina corrugata, and Rotaliella arctica. The benthic foraminifera most distal from any cave entrances were dominated by taxa also known from the deep-sea (e.g., Rotaliella, Spirophthalmidium) in sediment with the lowest bulk organic matter content (mean: 6%), or taxa that prefer hard substrates and are potentially living attached to cave walls (Patellina, Spirillina). While physical groundwater characteristics (e.g., salinity, dissolved oxygen) are expected drivers of benthic ecosystems in KSEs, these results suggest that POM source, quantity, and delivery mechanisms (e.g., groundwater-seawater circulation mechanisms, terrestrial flux) play an important role in benthic habitat partitioning and the spatial variability of biogeochemical cycles in the oxygenated marine sector of KSEs.

Keywords: anchialine, marine cave, submarine cave, Bermuda, karst, subterranean estuaries, foraminifera, subterranean ecology

INTRODUCTION

Karst subterranean estuaries (KSEs) are an environment created from the two- and three-way mixing of saline groundwater, rain, and oceanic water in the subsurface on carbonate landscapes, and this hydrographic framework promotes unique physical processes, biogeochemical cycling, and biological communities. Research on subterranean estuaries has significantly increased in the last 20 years (Moore, 1999; Gonneea et al., 2014; Brankovits et al., 2017, 2018; Pain et al., 2019, 2020; Duque et al., 2020). Traditionally, the biologic-focused study (fauna and ecosystems) of caves that are flooded by a KSE have been described with the adjectives anchialine or marine cave environments (Iliffe et al., 1983, 1984; Stock et al., 1986; Bishop et al., 2015). Since the groundwater-ocean dynamics that create KSEs are linked to sea level, KSEs also vertically migrate in the subsurface in response to orbitally-forced changes in eustatic sea level (van Hengstum et al., 2019) and this is represented in the fossil record (Proctor and Smart, 1991; van Hengstum et al., 2009; Rosso et al., 2015, 2018; Guido et al., 2017). Despite advances in the last decades, there still is considerable uncertainty regarding ecosystem functioning in modern KSEs, including the impact of organic carbon on benthic community ecology (Figure 1).

Benthic foraminifera are useful for investigating environmental variability and benthic habitat partitioning in KSEs in both modern and ancient settings (Balduzzi and Cattaneo, 1985; Proctor and Smart, 1991; van Hengstum et al., 2008, 2009; Omori et al., 2010; Guido et al., 2017; Bergamin et al., 2018, 2020; Romano et al., 2018, 2020). Benthic foraminifera are eukaryotic protists that live among the meiofauna (0.062-0.5 mm in size) in most marine settings, and they exhibit ecological zonation from physical environmental gradients (e.g., salinity, organic matter flux, dissolved oxygen). In the deep-sea, benthic foraminifera they can account for 50% of eukaryotic biomass in sediment (Gooday et al., 1992). Foraminifera are heterotrophs that eat phytodetritus, terrestrial detritus, prokaryotes, among other trophic strategies, with only a few species living in symbiotic relationships with photosynthetic algae (Gooday, 1988, 1993; Goldstein and Corliss, 1994; Linke et al., 1995; Nomaki et al., 2005; Dupuy et al., 2010). Individual foraminifera produce a simple shell, or test, that remains within the sediment after death, so statistically significant populations can be sampled by SCUBA divers during routine ecological surveys, or their fossil remains can preserve a record of prehistoric KSEs.

Here we document how benthic foraminiferal communities correlate with particulate organic carbon source, quantity, and delivery mechanisms in Bermuda's horizontally expansive submerged caves, which represent areas of KSEs where salinity and dissolved oxygen are not limiting factors. Bermuda is a \sim 35 million year old volcanic seamount in the North Atlantic Ocean, that is capped by 30-80 m of limestone interspaced by paleosols (Pirsson and Vaughn, 1917; Woolard and Ewing, 1939; Plummer et al., 1976; Palmer et al., 1977; Vacher et al., 1995). The limestone units deposited during Quaternary sea-level high stands have weathered into a mature karst landscape (Mylroie et al., 1995), with abundant cave-related karst development in the oldest carbonate formations that are concentrated in the northeastern part of the island (Land et al., 1967; Plummer et al., 1976; Palmer et al., 1977; Vacher and Rowe, 1997). Bermuda is an ideal location for this study because the flooded caves are an established biodiversity hot spot of endemic anchialine fauna (Iliffe et al., 1983), and its surrounding marine environments have been the focus of long-term oceanographic study (e.g., Woolard and Ewing, 1939; Neumann, 1965; Morris et al., 1977) and foraminiferal work (Carmen, 1927; Steinker and Clem, 1984; Javaux and Scott, 2003). This study provides the broadest geographic perspective yet available for benthic foraminiferal distributions in the KSE habitat for any global island by combining foraminiferal census data from 28 new samples from





the 2nd largest flooded cave (Palm Cave) into a new dataset with 100 samples from previously studied caves in Bermuda (Deep Blue, Cow Cave, Green Bay Cave).

MATERIALS AND METHODS

Research Design and Study Sites

Submerged caves in Bermuda are most abundant in the oldest geologic formations (e.g., Walsingham Formation) between Castle Harbor and Harrington Sound, and between North Shore Lagoon and Harrington Sound (**Figure 2**). The volcanic-limestone contact has not yet been observed in any flooded cave, and cave passages are generally shallower than ~23 m below sea level. This study focuses on two large caves with physical connections into Harrington Sound (i.e., Green Bay Cave, Palm Cave), and two cave entrances that are slightly more inland (Cow Cave, Deep Blue). Harrington Sound is a shallow coastal embayment that is ~5 km², with a mean depth of 14.5 m below sea level, and a maximum depth of ~25 m below sea level in the southwest (Parsons et al., 2015). It is estimated that the seawater residence time in the mixed layer of Harrington Sound is ~30



FIGURE 2 | (A) Digital elevation model of Bermuda in the North Atlantic Ocean. **(B)** Detailed map of Harrington Sound plotted in ArcGIS. Red stars indicate study site locations.

days from tidal exchange of seawater through the narrow channel at Flatts Inlet (Morris et al., 1977), but effective tidal mixing can be as long as 4 months (Parsons et al., 2015). Harrington Sound experiences seasonal hypoxia or anoxia from water column stratification in summer (Coull, 1969; Bates, 2017), which causes benthic sedimentary and ecological zonation: a sandy shallow zone with sea grass and macroalgae (0-10 m), the Oculina coral zone (10-19 m), and the subthermocline zone (19-26 m)(Neumann, 1965). Seasonal dysoxia or anoxia have not been observed or documented in Bermuda's caves, and previous work documents that surficial foraminiferal distributions in Bermuda's caves are not typical of those observed in open ocean hypoxic to anoxic settings (Sen Gupta et al., 1996; Bernhard and Sen Gupta, 1997; Levin et al., 2009). This suggests that tidal pumping of seawater from the mixed layer in Harrington Sound into the caves is the dominant source of seawater. Tidal pumping also causes microtidal variability (<0.5 m) in the local groundwater table elevation (i.e., water table, Martin et al., 2012), which can be measured in the shallow pools that provide access to the flooded caves (Palmer et al., 1977; van Hengstum et al., 2015; Little and van Hengstum, 2019).

Bermuda's caves offer a great location for investigating the marine sector of a KSE on a carbonate platform that is nearly flooded by eustatic sea-level rise. The commonly-studied flooded caves in Bermuda do not discharge a meteoric lens toward the ocean like caves on larger carbonate landscapes in the Mediterranean area (Radolović et al., 2015; Romano et al., 2020) or Yucatan Peninsula (Suárez-Morales et al., 2004; Chávez-Solís et al., 2020). This is in part because Bermuda is currently in a late stage of carbonate platform flooding relative to the position of global sea level (van Hengstum et al., 2019), and local meteoric water masses in Bermuda are spatially diminished. On the carbonate platforms of The Bahamas, for example, landmass area and mean annual rainfall alone can be used to predict meteoric lens thickness on different islands (Cant and Weech, 1986). Bermuda does have five modest meteoric lenses located in geologically-recent lithologies (Vacher, 1978), but these are not located in the primary area of cave development on the isthmus between Castle Harbor and Harrington Sound (Walsingham Formation, highest porosity: Land et al., 1967). In northeastern Bermuda where the Walsingham Formation is located, only a thin (<50 cm), seasonal and ephemeral brackish cap occurs in some cave-collapse entrances, and a limited meteoric lens can form near the "Letterbox" area of Green Bay Cave that does not directly contact the sediment-water interface, or discharge water through marine exit into Harrington Sound (see map in van Hengstum and Scott, 2011). The mineralogy of sediment in Bermuda's caves is dominated by carbonates, along with localized deposits of paleosol eroding into cave from the terrestrial surface above. Lastly, while large carbonate platforms in the Bahamas and the Yucatan frequently have large sinkholes that host extensive primary productivity and are interconnected to flooded caves (Schmitter-Soto et al., 2002; Tamalavage et al., 2018), the most comparable geomorphologic feature in Bermuda is Cliff Pool Sinkhole in the Green Bay Cave System. Even then, Cliff Pool still only supports a thin (<0.5 m) brackish water mass whose salinity depends upon seasonal rainfall and aridity.

Two of the largest cave systems in Bermuda, Palm Cave (primary entrance: 32.34°N, 64.71°W) and Green Bay Cave (primary entrance: 32.33°N, 64.74°W), were targeted for analysis of recent foraminiferal distributions (uppermost 5 cm of sediment) and compared to the impact of tidal variability on foraminiferal distributions at cave entrances to the Walsingham Cave System at Deep Blue (32.347°N, 64.711°W) and the entrance to Cow Cave (32.34°N, 64.71°W). Green Bay Cave also has the largest direct connection to the sea at Harrington Sound. According to measurements of Iliffe (2008), 1,960,000 m³ of seawater flows into Green Bay Cave at a cross sectional area of 25.5 m² during a flood tidal cycle. In contrast, Palm Cave has multiple, small subaerial entrances to the forest landscape in the Walsingham Nature Preserve, and one direct connection to Harrington Sound at Cripple Gate (cross sectional area: 0.6 m²) that permits \sim 1,000 m³ of seawater exchange during a flood tidal cycle (Iliffe, 2008). Both Palm Cave and Green Bay Cave have seagrass meadows (e.g., Thalassia testudinium, Syringodium filiforme) within 100 m of their marine entrances (Iliffe, 2008), in the shallow sandy benthic ecological zone in Harrington Sound (Neumann, 1965).

Benthic Foraminiferal Processing From Palm Cave

28 new surface (<5 cm depth, $>30 \text{ cm}^3$ of sediment) sediment samples were collected from various depths and locations throughout Palm Cave in August 2015 (Figure 3). At each sample station, a separate 50 mL centrifuge tube was filled with sediment by a SCUBA diver employing advanced technical cave diving procedures. Collecting this volume of sediment permitted separate sediment sub-samples from each station to be individually processed for benthic foraminifera, textural analysis, Loss-on-Ignition (LOI), and bulk organic geochemistry. The total assemblages of benthic foraminifera were analyzed at each sampling station, as some workers perceive this to be most reflective of long-term distributions (Buzas, 1968; Scott and Medioli, 1980; Debenay and Guillou, 2002). Benthic foraminifera were concentrated by wet sieving sediment sub-samples (0.63-5 cm³) over a standard 63 μ m screen mesh, with the remaining coarse sediment residues sub-divided into representative aliquots using a wet splitter to enable representative census counts of \sim 300 individuals per sample (Patterson and Fishbein, 1989; Scott and Hermelin, 1993). While van Hengstum and Scott (2011) used a $45\,\mu m$ sieve to concentrate benthic foraminifera in their study of Green Bay Cave, Little and van Hengstum (2019) documented negligible difference between resultant foraminiferal communities from Bermudian caves using 45 vs. 63 µm mesh sizes. Individual benthic foraminifera were wet picked onto micropaleontological slides and identified to the species level in most cases. Taxonomy was confirmed by comparing scanning electron microscopy of representative individuals to literature comparisons (Carman, 1933; Bermúdez, 1949; Loeblich and Tappan, 1987; van Hengstum and Scott, 2011; Little and van Hengstum, 2019). This produced a new dataset for benthic foraminifera from Palm Cave of 28 samples x 111 taxonomic observations (see Supplementary Material).

Sediment Texture and Organic Geochemistry

Sediment texture (i.e., mean particle size) in Palm Cave was examined in separate sediment sub-samples using standard laser diffraction particle size analysis and loss on ignition (LOI) procedures. Organic matter was recorded as percent-lost relative to the original total sample weight after LOI at 550°C for 4.5 h (Heiri et al., 2001). Mean particle size for sediment sub-samples was measured with a Malvern Mastersizer 2000S laser particle size analyzer (Sperazza et al., 2004), with individual particles disaggregated in a sodium hexametaphosphate dispersant prior to analysis. The stable carbon isotopic value ($\delta^{13}C_{org}$) and C:N ratio of bulk sediment was measured on separate sediment sub-samples from the new sediment samples collected from Palm Cave (n = 28). In general, δ^{13} Corg values of bulk sedimentary organic matter reflect the relative contributions of marine, terrestrial and pelagic organic carbon sources in coastal environments (Lamb et al., 2006; Khan et al., 2019). Terrestrial plants that preferentially use the C3 photosynthetic pathway produce organic tissues that are ¹³C-depleted and nitrogen-poor, with $\delta^{13}C_{org}$ values generally ranging between -32 and -21%(Lamb et al., 2006). Sea grasses and macroalgae are common in the shallow waters from 0 to 10 m of Harrington Sound (sandy zone with seagrass and macroalgae of Neumann, 1965) where Palm Cave and Green Bay Cave have entrances (Wefer and Killingley, 1986; Fourqurean et al., 2015). Seagrasses are also important contributors of carbon isotopically enriched organic matter to the sediment in nearshore environments relative to marine plankton (Fry et al., 1977; Anderson et al., 1992; Kennedy et al., 2010). Based on measurements on plant tissues around Bermuda coastal waters, seagrasses (e.g., Syringodium and Thalassia: mean of -7.4%, n = 455) and macro algae like Halimeda (mean of -14.3%, n = 60) also have relatively more ¹³C-enriched $\delta^{13}C_{org}$ values (see Table 1 in Burgett et al., 2018). The $\delta^{13}C_{org}$ values of particulate organic carbon (POC) in the Sargasso Sea are similar to elsewhere in the tropical North Atlantic and range from \sim -21 to -23% (Jeffrey et al., 1983; Altabet, 1990), and these values primarily reflect the ranges of marine planktonic algae (France, 1995). For samples from Palm Cave, carbonates were first digested with 1.0 M HCl for 12 h, and the remaining sediment residue was rinsed, desiccated, powdered, and weighed into silver capsules. Stable carbon isotope ratios ($\delta^{13}C_{org}$) were measured against international standards at Baylor University Stable Isotope Laboratory on a Thermo-Electron Delta V Advantage Isotope Ratio Mass Spectrometer. Final isotopic ratios are reported relative to the standard delta (δ) notation relative to Vienna Pee Dee Belemnite (VPDB) for carbon (expressed as parts per mil: %) with an uncertainty of \pm 0.1‰, with the C:N ratio and total organic carbon (TOC, weight %) calculated from the analysis of the residual sediment fraction after acidification.

Data Analysis

A final dataset was compiled for further multivariate statistical analysis that contained 128 total samples: (a) the 28 new samples from Palm Cave (this study), 74 samples from



Green Bay Cave (van Hengstum and Scott, 2011), 18 samples from Cow Cave (Little and van Hengstum, 2019), and 7 samples from Deep Blue (Little and van Hengstum, 2019). From Little and van Hengstum (2019), we use the $>45 \,\mu m$ database, which has higher census counts and thus likely better represents the sampled areas. All taxonomic observations were grouped to the generic level to limit the impact of taxonomic bias imparted by different microscopists at the species level. The exception was the brackish water indicator Trochammina inflata, which was left at specific level to differentiate from co-generic marine taxa (e.g., T. globularis). Also, the final dataset amalgamated the taxonomically similar genera Pattelina, Heteropatellina, and Patellinoides (all members of the order Spirillinida). These genera were differentiated by Little and van Hengstum (2019, Deep Blue and Cow Cave), but lumped together by van Hengstum and Scott (2011, Green Bay Cave). The final combined dataset was

128 samples by 89 taxonomic units at the generic level (see **Supplementary Material**).

Numerical analyses were completed in R Studio software (version 1.1.383; R Studio, 2016). The final combined dataset of benthic foraminifera was used to calculate rarefied species richness (*S*) and the Shannon-Wiener Diversity Index (H), using the "rarefy" and "diversity" functions in the R package "vegan" (Gotelli and Colwell, 2011; Oksanen et al., 2013), and raw counts were converted to proportional abundances using the "decostand" function in the "vegan" package. Unconstrained Q-mode cluster analysis was used to help divide the dataset into ecologically meaningful benthic foraminiferal communities by grouping statistically similar populations (Legendre and Legendre, 1998). Prior to cluster analysis, the proportional abundances were square root transformed to minimize the impact of dominant species and to better compare community structure (Legendre and Legendre, 1998). Samples

were then subjected to hierarchical cluster analysis using an unweighted paired group averaging algorithm and the Bray-Curtis dissimilarity index using the "simprof" function in the "clustsig" package (Clarke et al., 2008; Whitaker and Christman, 2014), and the "vegdist" function in the "vegan" package (Oksanen et al., 2013). The proportion of dominant taxa in the dendrogram (5% proportional abundance in at least 1 sample) were illustrated between the different samples by generating a vertical stratigraphic plot using the "stratplot" function in the "rioja" package (Juggins, 2015). Finally, physical environmental parameters were graphically summarized in box-and-whisker plots: $\delta^{13}C_{org}$, C:N, bulk organic matter-LOI, TOC, mean particle size, and mean groundwater depth, and compared to the benthic foraminiferal communities grouped by unconstrained Q-mode cluster analysis.

RESULTS: BENTHIC FORAMINIFERAL COMMUNITIES

Similar to other Bermudian caves investigated (Green Bay Cave, Deep Blue, and Cow Cave), Palm Cave has abundant benthic foraminifera with a mean concentration of 7256 individuals per cm³ of sediment (range: 458-14,603). No new endemic species of foraminifera are described, but the identified benthic foraminiferal communities differ from those found in other Bermudian coastal environments (e.g., mangroves, carbonate lagoons, or reefs: see Javaux and Scott, 2003). Seven communities of benthic foraminifera can be recognized at a dissimilarity index of 0.44 on the dendrogram produced from unconstrained Qmode cluster analysis (Figure 4), but this division increases to nine groupings if Community 5 is subdivided at a dissimilarity of index of ~0.36 (discussed below). Two samples were plotted at the bottom of the dendrogram and were deemed most dissimilar to the other samples: ISC-72 and CC5-ITA. From the original dataset, however, the very high relative abundance of Spirophthalmidium emaciatum (70%) in ISC-72 suggests this sample is most closely related to Community 6. Similarly, CC5-ITA had a very high abundance of Trochammina, Entzia, and Miliammina relative to other samples in Community 1. With currently available data, we consider that the samples CC5-ITA and ISC-72 are just endmember examples in Community 6 and 1, respectively.

Community 1 characterizes the intertidal zone of Cow Cave, and this lower diversity (H = 2.2) community is dominated by taxa known to tolerate periodic aerial exposure related to tides, high quantities of bulk organic matter in the sediment, and lower salinities. Dominant taxa include *Trochammina inflata* (mean: 18.3%), *Miliammina fusca* (mean: 13.1%), and *Entzia macrescens* (mean: 8.7%; **Table 1**). Tidal variability of the water table causes periodic exposure of these shallow benthic areas (**Figure 5**), and this community likely also experiences repeated brackish conditions from rain and surface runoff (Little and van Hengstum, 2019). Community 1 is characterized by the highest bulk organic matter content (mean: 25.5%), which is predominantly derived from terrestrial sources based on the mean $\delta^{13}C_{org}$ values and C:N ratio (mean: -25.9‰, mean C:N ratio: 18.3; **Figure 6**).

Community 2 includes subtidal areas of Deep Blue in the saline groundwater mass that are exposed to normal sunlight. The benthos has lower bulk organic matter (mean: 18.8%) than Community 1, but the organic carbon remains primarily sourced from the adjacent terrestrial surface (mean: -26.1%, C:N ratio mean: 11.1; **Table 1**). This community has higher species richness (S = 28) and diversity (H = 2.7) than Community 1, which is most likely because these benthic area are constantly submerged. This community is dominated by the infaunal genus *Bolivina* (mean: 16.6%), *T. inflata* (mean: 12.2%), and *H. anderseni* (mean: 7.5%, **Table 1**). *Rosalina* is also dominant (mean: 10.5%), which is primarily *Rosalina globularis*, and small-sized *Triloculina oblonga* (mean: 12.2, **Supplementary Material**).

Communities 3 and 4 occur in a transect of increasing depth into the saline groundwater at Cliff Pool Sinkhole within the Green Bay Cave (AC-04 to AC-11 of van Hengstum and Scott, 2011), and include the shallowest samples from Cliff Pool Sinkhole (ML-1, ML-2, ML3). The shallow sinkhole itself receives direct sunlight, and shallow sampling sites have taxa that must be tolerant of brackish conditions. Communities 3 and 4 start in open water, and form a transect down into the aphotic zone of Green Bay Cave. Community 3 is dominated by Bolivina spp. (mean: 20.2%), H. anderseni (mean: 21.4%), and Rosalina spp. (mean: 11%). While Ammonia (mean: 11.6%) has been grouped to the generic level, van Hengstum and Scott (2011) observed that Community 3 was characterized by Ammonia beccarii var. tepida. In contrast, Community 4 has higher species richness (S = 30) and diversity (H = 2.6), and Bolivina spp. (mean: 26%) and Rosalina spp. (mean: 11%) remain dominant (Figure 4; Table 1). However, Community 4 also includes other taxa like phytodetritus feeding Melonis barleeanum (mean: 5.8%), Siphogenerina columellaris (mean: 4.9%), Siphonina (mean: 4.2%), and Nonionella (mean: 4.7%, Figure 4). Community 3 contains a mean of 14.1% of bulk organic matter, and a mean $\delta^{13}C_{org}$ value of -24.2% and a mean C:N ratio of 11 (Table 1), whereas Community 5 has higher bulk organic matter (mean 17.5%) content.

Most of the samples collected from throughout Palm Cave, Cow Cave, and Green Bay Cave collectively form Community 5 at a Bray-Curtis dissimilarity index of 0.44. Community 5 is located in benthic areas that: (i) are only impacted by saline groundwater, (ii) are not subaerially exposed anytime during the tidal cycle, and (iii) are in the aphotic zone. While all the areas characterized by Community 5 samples have similarities, important differences emerge with the physical environmental characteristics that warrant dividing further at a Bray-Curtis dissimilarity index of 0.36 (Figure 4). For example, Communities 5A, 5B, and 5C show a trend toward more enriched $\delta^{13}C_{org}$ values and decreasing bulk organic matter content (Figure 5). Community 5A is located in the subtidal areas of the roofed Cow Cave, and is located in shallow subtidal pools that are proximal to the adjacent terrestrial landscape. Except for the samples collected specifically in Cliff Pool Sinkhole (ML-1, ML-2, and ML-3), most of Communities 5B and 5C are located deeper in the saline groundwater. Community 5A is

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TABLE 1 | Selected summary statistics for environmental characteristics, and arithmetic mean of the percent abundance of dominant taxa in each benthic foraminiferal community.

	Dominated by Terrestrial Organic Matter (TOM) Sources				Dominated by Marine Organic Matter (MOM) Sources				
Benthic foraminiferal	1 Intertidal	2 TOM shallow subtidal	3 TOM high carbon	4 TOM lower carbon	5A MOM shallow subtidal	5B MOM high carbon–finer	5C MOM high carbon- coarser	6 Low carbon marine	7 Marine Cave entrance
community Community name									
Sample count	6	7	13	8	12	33	28	11	10
δ ¹³ C _{org} (‰) average	-25.9	-26.1	-24.2	-23.2	-21.6	-18.6	-20	-17.6	-18.6
$\delta^{13}C_{org}$ (‰) standard deviation	1.2	1.6	1.5	0.7	3.2	0.2	1.7	0.4	0.5
$\delta^{13}C_{\text{org}}$ (‰) range	-27.5 to -24	-27.7 to -23	-26.7 to -20.4	-24.4 to -21.7	-25.8 to -17.8	-19.2 to -17.8	-23.7 to -15.1	-18.6 to -16.8	-19.6 to -17.9
C:N average	18.3	11.1	11.2	11	10.1	8.2	9.1	8.6	8.1
C:N standard deviation	11.7	7.3	1.7	0.9	2.1	0.3	1.3	0.5	0.4
C:N range	3.4-28.7	1.1-18.6	8.5-15.1	9.5-12.1	6-12.4	7-8.7	7.8-14.2	8.1-10.2	7.5-8.6
Bulk organic matter (% LOI) average	25.5	18.7	14.1	17.5	13.4	8.1	8.6	6	10
Bulk organic matter (% LOI) standard deviation	6.5	6.1	8.8	4.4	4.8	1.9	2.6	2.3	6
Bulk organic matter (% LOI) range	18.2–36.3	11.6–28.2	6.7–40.8	11–23.2	9–20.5	4–12	2.4–12.5	2.5–9.3	3.2–20
TOC (%) average	7.4	4.5	23.9	12.5	5.5	15.3	12.4	5.1	30
TOC (%) standard deviation	5.1	2.4	8.9	6.3	1.2	4.5	5.7	2.2	4.2
TOC (%) range	2.6-16.2	1.5-7.8	11.4-41.3	4.1-22.6	4-8.4	5.8-26.7	2.6-25	1.8-9.6	24.3-35.4
Particle size (mm) average	16.2	43.8	271.2	149.6	14.3	20.7	39.7	5.9	311.3
Particle size (mm) standard deviation	6	27.4	52.8	109	2.6	43.6	34.9	1.3	113.9
Particle size (mm) range	92-247	23-99.3	190 5-370 4	13 5-333 8	97-196	1 1-218 1	13-169	4 1–7 9	86 7-459 4
Depth (m) average	-0.1	0.5	3	17.5	0.7	15.9	13.5	15.4	53
Depth (m) standard deviation	0	0.2	3.2	1.6	0.3	3.4	7	3	2.4
Depth (m) range	-0.3 to -0.1	0 2-0 9	0 2-10 7	13 7-18 7	0 1-1 2	5 2-20 1	2 4-22 6	96-186	2 7-10
Foraminiferal concentration (tests per cm^3) average	1811.8	595.4	322.3	1521.6	3507.2	3227.7	7415.5	2569.5	3480
Shannon-Weiner Diversity Index (<i>H</i>) average	2.2	2.7	2.1	2.6	3	2.9	2.9	2.1	2.5
Rarefied species richness average	21	28	18	30	34	35	34	23	30
Dominant taxa-percent at	oundance (%, a	verage)							
Ammodiscus	-	1.3	-	-	1.3	2.1	-	2.3	-
Ammonia	-	-	11.6	1.4	-	2.2	-	-	9.7
Bolivina	37	16.6	20.2	29.2	12.5	11.4	54	4.3	11
Cibicides	-	-			-	11	1.8	-	1.9
Cvcloavra	3.9	-	-	-	28	1.6	2.5	-	-
Elphidium	-	_	19	-	1	-		_	47
Entria	87	1.8	-	_	-	_	_	_	
Globocassidulina	0.1	2	_	3.1	23	2	2.4	_	_
Helenina	2.0	75	21.4	5.1	2.0	2	2.4	-	-
Loxostomum		-	-	2.5	-	-	-	-	-
Melonis	-	7	-	5.8	51	1.6	1.9	-	-
Miliammina	13.1	1	-	-	-	-	-	-	-
Average of Miliolinella	-	-	1	-	4	2.2	7.2	-	1.6
Mychostomina	-	-	-	-	2.9	-	1.6	1.7	-
Nonionella	-	1.7	-	4.7	2.8	1.4	-	-	-

(Continued)

TABLE 1 | Continued

	Dominated by Terrestrial Organic Matter (TOM) Sources				Dominated by Marine Organic Matter (MOM) Sources				
Benthic foraminiferal	1 Intertidal	2 TOM shallow subtidal	3 TOM high carbon	4 TOM lower carbon	5A MOM shallow subtidal	5B MOM high carbon–finer	5C MOM high carbon– coarser	6 Low carbon marine	7 Marine Cave entrance
Community Community name									
Peneroplis	-	-	-	-	-	-	-	-	3
Pseudo- <i>Patellina</i> (<i>Pattelina</i> -like taxa)	-	-	-	-	3.3	5.6	7.8	7.2	1.6
Quinqueloculina	1.4	3.1	1.5	-	9.2	4.1	8.2	-	27.5
Reophax	-	1.4	1.2	1.4	-	3.1	-	-	1
Rosalina	11.2	10.5	12.5	10.5	9.5	3.7	5.4	1.6	4.8
Rotaliella	-	-	-	-	1	1.6	-	2.5	-
Sigmoilina	-	-	-	-	1.4	1.8	1.2	5.9	-
Siphogenerina	-	-	-	4.9	1.8	-	-	-	-
Siphonina	-	-	-	4.2	1	1	1.5	-	-
Spirillina	1.1	-	1.4	-	5.8	10.9	13	24.3	1.1
Spirophthalmidium	-	-	-	2.2	7.6	8.3	10	31.5	-
Textularia	-	1.7	3.9	3.3	-	-	-	-	3.2
Trichohyalus	4.7	2.2	6.3	-	-	-	-	-	-
Triloculina	14.3	12.2	7.7	2.2	5.5	13	3.7	1.1	12.9
Trochammina	-	3.6	-	-	1.2	2.3	4.6	2.4	-
Trochammina inflata	18.3	12.2	-	-	-	-	-	-	-

Species with a mean percent abundance of <1% are marked with a dash to emphasize dominant taxa. The data from samples CC5-ITA and ISC-72 were averaged into Community 1 and 6, respectively. TOC, total organic carbon; LOI, Loss-on-Ignition.

also a transition between benthic foraminiferal communities characterized by more depleted $\delta^{13}C_{org}$ values (Communities 1–4) vs. more enriched $\delta^{13}C_{org}$ values (Communities 5B, 5C, 6, and 7), and similarly with C:N ratios (**Figure 5**). Despite a general linear trend in the $\delta^{13}C_{org}$ values of the sediment (**Figure 5**), there is a distinct change in the benthic foraminiferal communities from the dominance of *Bolivina* and *Rosalina* in Community 5A to *Spirophthalmidium*, *Spirillina*, and *Pattelina*like genera (*Patellina*, *Heteropatellina*, *Patellinoides*) dominating Communities 5B and 5C (**Figure 4**; **Table 1**).

Community 6 is located in areas most distal to any cave exit (either terrestrial or marine). Relative to most other areas, Community 6 is characterized by enriched $\delta^{13}C_{org}$ values (mean: -18.5%) and lower C:N ratios (mean: 8.6). The sediment where Community 6 is found has the lowest total organic carbon content (mean: 5.1 ± 2.2%) and bulk organic matter observed (mean: 6 ± 2.3%). While all cave areas are dominated by carbonate and organic matter, Community 6 is dominated by fine-grained carbonate particles (fine silt: 5.9 ± 1.3 µm, **Table 1**). Community 6 is less diverse than Community 5 (*S* = 22, *H* = 2.1), and is dominated by *Spirophthalmidium emaciatum* (mean 20.4%), *Spirillina vivipara* (mean: 19.6%), *Patellina* (mean: 9.6%), and *Bolivina* (mean: 4.3%).

Community 7 occurs in a transect from open water in the shallow (0–10 m) sandy ecological zone in Harrington Sound (E-47, E-48, E-49, and E-50), through the marine cave entrance into Green Bay Cave and stops at the Rat Trap area (see **Figure 2** in

van Hengstum and Scott, 2011). This community is characterized by lower bulk organic matter on average (mean: 10%), but high variability (range: 3.2–20%) depending upon the spatial variability in carbonate sand content (mean particle size: 311 μ m, **Table 1**). The δ^{13} Corg values (mean: –18.6‰) of the sediment are more enriched relative to Communities 1–4, with a lower C:N ratio (mean: 8.1). This community of benthic foraminifera has a higher diversity (H = 2.5) and species richness (S =29), and is dominated by common carbonate lagoon genera *Quinqueloculina* (mean: 26%), but also *Bolivina* (mean: 12%), and *Triloculina* (mean: 11.4%).While *Ammonia* (mean: 9.7%) has been grouped to the generic level, van Hengstum and Scott (2011) observed that Community 7 was characterized by *Ammonia beccarii* var. parkinsoniana.

DISCUSSION: DRIVERS OF FORAMINIFERAL DISTRIBUTIONS

Our previous work (i.e., van Hengstum and Scott, 2011) divided the benthic foraminiferal communities in Green Bay Cave into those that were termed anchialine (terrestrially-dominated) vs. submarine (marine-dominated). This was based on the ideas of "terrestrial vs. marine influence" that were originally presented in Stock et al. (1986, more specifically), see the footnotes on page 91). This partly reflects the challenges faced by early workers with evaluating environmental and biological differences



between: marine flooded caves like those found in northeastern Bermuda that have negligible meteoric lenses, vs. (ii) those with extensive meteoric lens development such as observed in caves the Yucatan Peninsula. However, a "terrestrial vs. marine" distinction is too simple for describing both benthic and pelagic habitat partitioning in subterranean estuaries, and the dependance of physical processes, biological communities, and biogeochemical cycles on specific groundwater masses (Brankovits et al., 2017, 2018; Chávez-Solís et al., 2020) that must vertically migrate through geologic time (Riedl and Ozretić, 1969; van Hengstum et al., 2019). For example, recent work with Typhlatya shrimp surveyed over a large area in the Yucatan Peninsula clearly documents pelagic habitat partitioning with respect to groundwater masses and carbon sources (see Figure 2 in Chávez-Solís et al., 2020). After applying broader environmental concepts of a KSE (Figure 1), the sampled marine caves of Bermuda include only the marine sector of KSEs that are not oxygen or salinity limited. As discussed below, modern benthic foraminiferal communities in the marine sector of KSEs in Bermuda's can be correlated with exposure caused by tidal variability of the water column, and the quantity, source, and transport mechanisms of particulate organic matter that is delivered to the benthos.

Marine Benthic Habitats Dominated by Terrestrial-Sourced Organic Carbon

In general, benthic foraminiferal Community 1 (mean: -25.9%), 2 (mean: -26.1%), 3 (mean: -24.2%), and 4 (mean: -23.2%) all occupy sediment that has more depleted $\delta^{13}C_{org}$ values and lower C:N ratios (**Figures 5**, **6**). Based on a comparison with literature values, this likely indicates that terrestrial organic matter is the dominant source of organic carbon delivered to these areas. In opposite, Communities 5, 6, and 7 occupied sediment with more enriched $\delta^{13}C_{org}$ values (mean: -21.6 to -17.6%, **Table 1**), which suggests that marine carbon sources are more dominant. The proximity between sample stations and cave





openings (i.e., exits) to the forest landscape is clearly impacting organic matter delivery to the benthos. Detrital carbon is a key resource for benthic foraminifera in marine environments (Gooday, 1993; Jorissen et al., 1995), so it is unsurprising that foraminifera are also responding to different carbon sources in flooded marine caves. Cave exits provide a point source for terrestrial sedimentation into an otherwise marine environment, and diverse organic carbon pools increase the opportunity for resource partitioning and foraminiferal diversity. While terrestrial detritus eroding into the caves from the adjacent landscapes is likely a dominant carbon source for Communities 1–4 ($\delta^{13}C_{org}$ values: –26.1 to –23.2‰), we cannot rule out the contribution of organic matter from marine plankton in the water column at Cliff Pool Sinkhole (Green Bay Cave) as also impacting the proximal Communities 3 and 4. Since marine POC from marine plankton has a $\delta^{13}C_{org}$ value of –21 to –23‰ (Jeffrey et al., 1983; Altabet, 1990; France, 1995), the $\delta^{13}C_{org}$ value of benthic sediment alone cannot differentiate the relative contributions of carbon from terrestrial detritus vs. marine plankton. Deposition of POC from marine plankton

can also explain the modestly lower and stabilized C:N values in Communities 3 and 4 relative to Communities 1 and 2 (Figure 5). This is because vascular terrestrial detritus typically has much higher C:N ratios (Lamb et al., 2006), which is likely to collect in cave pools closer to the forest, whereas the deeper water column at Cliff Pool Sinkhole can support higher primary productivity. The uncertainty in these results is an opportunity to use more specific organic biomarkers to resolve carbon source contributions to the benthos. Overall, the more carbon isotopically depleted samples suggest an increased proportion of terrestrial detritus in Communities 1–4 relative to Communities 5–7, as all areas likely receive inputs of POC from marine plankton in response to active seawatergroundwater circulation between Harrington Sound and the caves (discussed below).

Similar to the result of Little and van Hengstum (2019), benthic habitats dominated by terrestrial organic carbon deposition can be further separated by tidal exposure: Community 1 is exposed to tidal variability of the water column, whereas Communities 2, 3, and 4 are all subtidal (Figure 5). The micro-tidal changes to the water table elevation at Cow Cave are sufficient to expose Community 1 during low tides (Figure 5, Little and van Hengstum, 2019). Community 1 is dominated by agglutinated, brackish-tolerant species Trochammina inflata, Miliammina fusca, Entzia macrescens, and Helenina anderseni (Table 1). These taxa are common in marshes, which also experience tidal exposure (Hayward et al., 1999; Javaux and Scott, 2003; Camacho et al., 2015). Community 1 likely also experiences intermittent brackish conditions from runoff during pluvial events given the close proximity of Community 1 to the terrestrial landscape (Figure 7). Taxa like E. macrescens and *Miliammina fusca* are useful sea-level indicators due to their significant correlation with tidal elevation (Scott and Medioli, 1978; Guilbault et al., 1995; Gehrels and van de Plassche, 1999). In the studied Bermuda caves, *Miliammina fusca* is the primary taxon in intertidal areas dominated by terrestrial organic carbon, with *E. macrescens* occurring in both intertidal (Community 1) and subtidal (Community 2) areas (**Figure 4**).

Subtidal benthic areas dominated by the deposition of terrestrially-sourced organic carbon could be further divided by light exposure, depth into the subsurface, and particle size (Communities 2, 3, and 4, Figure 7). None of the recovered benthic foraminifera in the caves employ photosymbionts (e.g., Amphistegina), therefore, foraminifera are all feeding on materials delivered to, or otherwise available in, the substrate (e.g., algae, bacteria, detritus, or dissolved organic matter). While these communities are all dominated by Rosalina and Bolivina (Table 1), differences in the secondary taxa are relevant to the community structure. It is important to note that Cliff Pool Sinkhole has a water column open to sunlight, and eyewitnesses report that the sinkhole can appear seasonally greenish from primary productivity. This potentially explains faunal differences between Cliff Pool Sinkhole (increased abundance of Trichohyalus aguayoi and Helenina in Community 3) vs. Deep Blue (increased abundance of Trochammina and Entzia in Community 2, Figure 4), with the potential for increased resource partitioning and foraminiferal diversity deeper into the saline groundwater mass at Cliff Pool Sinkhole (Communities 3 and 4). Other secondary taxa that are also present include Melonis barleeanum (Caralp, 1989) and Globocassidulina subglobosa, both of which are known to respond favorably to seasonal delivery of phytodetritus in the open ocean. Their increased



FIGURE 7 | Conceptual model of idealized benthic foraminiferal communities in the marine sector of karst subterranean estuaries in Bermuda, along with idealized changes in stable carbon isotopic value of bulk organic matter ($\delta^{13}C_{org}$), organic matter quantity, mean particle size of substrate, tidal exposure in subaerial pools, and the magnitude of nutrient and particle delivery from ocean-groundwater exchange through marine cave openings. Representative values of $\delta^{13}C_{org}$ values for source organic matter summarized from the literature (Goericke and Fry, 1994; Kennedy et al., 2010; Fourqurean et al., 2019), and ecological zones in Harrington Sound from Neumann (1965).

percent abundance in Communities 3 and 4 proximal to Cliff Pool Sinkhole is likely related this additional food resource produced in the dimly-lit water column, and is supported by modestly lower C:N ratios (Figure 5). This likely indicates that additional environmental variables are impacting the benthic foraminiferal communities that are not quantified in this study (i.e., dissolved nutrients at the sediment-groundwater interface, pore water chemistry). Based on experimental cultures, the epifaunal Rosalina globularis prefers warm and saltier waters (optimal: 27°C, >25 psu) for reproduction (Saraswat et al., 2011, 2015), which are found in the relatively shallow (<20 mdepth) marine habitats in Bermuda's caves. The most abundant genus is the infaunal Bolivina, which is likely capitalizing on any available food resource in the benthos. While the impact of particle size on benthic foraminifera in other environments remains a debated issue (Debenay et al., 2001; Diz et al., 2004; Armynot du Châtelet et al., 2009), our results suggest that substrate texture in KSEs may also be influencing benthic habitat variability and foraminiferal community structure. Community 3 was associated with coarser substrates (mean particle size: $>271.2 \,\mu$ m) compared with Community 4 (mean particle size: 149.6 µm).

These results now clearly inform the environmental conditions of Pleistocene-aged KSEs that were colonized by benthic foraminifera, which have been subsequently preserved in fossil cave deposits. For example, Proctor and Smart (1991) documented fossil benthic foraminifera in elevated caves in England, which are most likely dated Marine Isotope Stage 11 (424,00-374,000 years ago). Detailed foraminiferal counts were not provided, but fine textured sediment contained echinoid spines, Ammonia beccarii, Cassidulina, and Bolivina. On the Yucatan Peninsula in Mexico, van Hengstum et al. (2009) described Bolivina-dominated foraminiferal communities in Carwash Cave (older than Last Glacial Maximum 20,000 years ago), and there are Helenina- and Bolivina-dominated Pleistocene-aged foraminiferal deposits in Maya Blue Cave and Vaca-Ha (van Hengstum and E. Reinhardt, unpublished observations). Based on modern benthic foraminiferal communities in Bermuda, these examples of fossil benthic foraminifera were all living in sediment that was dominated by terrestrial organic matter inputs, with additional taxonomic variability in the Pleistocene foraminiferal community most likely related to the salinity of the contemporaneous groundwater mass flooding that specific cave.

Marine Benthic Habitats Dominated by Marine-Sourced Organic Carbon

Communities 5, 6, and 7 occur in benthic habitats that primarily receive marine sources of organic carbon, which is most likely from marine plankton based on the $\delta^{13}C_{org}$ values from -22.3 to -15.1% and C:N ratios of 8–10 (Figures 6, 7). Marine particulate organic carbon generally has values from -18 to -23% in lower latitudes (Jeffrey et al., 1983; Altabet, 1990; Goericke and Fry, 1994). An important component of particulate organic matter is marine plankton, which has a much lower C:N ratio (<10) relative to terrestrial organic

carbon (Lamb et al., 2006; Figure 6). Seagrasses and macroalgae are abundant in the shallow sandy ecological zone of Harrington Sound (upper 10 m; Neumann, 1965), and consistent with global trends (Kennedy et al., 2010), seagrasses in Harrington Sound have enriched $\delta^{13}C_{org}$ values and C:N ratios exceeding 18 (Fourgurean et al., 2019). Two locally common seagrass species, for example, Thalassia testudium has a mean $\delta^{13}C_{org}$ value of -7.5% (n = 351) and a mean C:N ratio of 21 (n = 364), and Syringodium filiforme has a mean $\delta^{13}C_{org}$ value of -6.3% (n =369) and a mean C:N ratio of 18 (n = 362) (Fourgurean et al., 2019; Figure 6). Sediment below seagrass in Harrington Sound is estimated to have a C:N ratio of 11, and $\delta^{13}C_{org}$ value of -13to -16%, based on a global mean $\Delta^{13}C_{\text{seagrass-sediment}}$ value of 6.3‰ (Kennedy et al., 2010). While some contribution of organic matter from seagrasses and macroalgae cannot be ignored, the low C:N ratios associated with Communities 5, 6, and 7 (\sim 8– 10, Figure 7; Table 1) indicates that a more nitrogen-rich organic matter source like marine plankton is likely being delivered to these communities.

Community 7 is located in the main tunnel in Green Bay Cave that is closest to Harrington Sound, and extends from open water into an area colloquially referred to as the "Rat Trap" (see map in van Hengstum and Scott, 2011). This tunnel is characterized by attenuating light availability (disphotic zone), and past the Rat Trap is the aphotic zone. This tunnel is dominated by Quinqueloculina, Ammonia, Elphidium, and Peneroplis (Figure 6), which are common taxa in the shallow carbonate lagoons throughout the tropical North Atlantic Ocean (Javaux and Scott, 2003). While this study grouped all taxa at the genus level, van Hengstum and Scott (2011) noted that Community 7 is associated with Ammonia beccarii var. parkinsoniana. This tunnel coincides with areas that receive classic taxonomic investigation in marine caves, where strong gradients in light and tidal currents exist. The coarser substrate is likely promoted by winnowing-away of finer sediment particles in response to the large volume of seawater circulating through this tunnel during each tidal cycle (\sim 1,960,000 m³: Iliffe, 2008). Coarser substrates also occur in other Mediterranean marine caves with higher current velocities from tidal action (Radolović et al., 2015). It is likely that tidal and storm currents also have transported some benthic foraminifera tests from Harrington Sound into the cave tunnel, however, current velocities notably decrease through the rest of Green Bay Cave past the narrowing of cave passage at the Rat Trap. Interestingly, PCS-15 from Palm Cave was not similar to Community 7, despite its proximity to the opening with Harrington Sound (i.e., Cripple Gate). Perhaps this is related to the much smaller connection of Palm Cave to Harrington Sound at "Cripple Gate," and the much lower volume of water exchanged with Harrington Sound during tidal cycles impacting benthic habitat conditions.

Community 6 is most distal from any cave entrance (or exit, **Figure 7**), is associated with areas where endemic anchialine cave animals (e.g., *Procaris chacei*) are more likely to be observed, and the fine-grained substrate (mean particle size: $5.3 \,\mu$ m) has the lowest bulk organic matter content (mean: 6%) and TOC (mean: 5%). Community 6 has the highest percent abundance of *Spirophthalmidium emaciatum*, members of the order Sprillinida

(e.g., Patellina, Spirillina, and Mychostomina), and Rotaliella arctica. Observation of Spirophthalmidium emaciatum outside of marine caves is limited, and includes New Guinea (Haig, 1988), the Mediterranean (Kuhnt et al., 2007), and the deep Pacific (Brady, 1884). In general, the thin-walled and fragile test of S. emaciatum is unlikely to endure taphonomic reworking. This taxon is very closely related (or a ecophenotypic variant) of S. acutimargo, which constitutes up to 3% of communities in deepmarine carbonate oozes (>2000 m water depth) from the North Atlantic Ocean near the Azores (Hermelin and Scott, 1985). While common in Community 6 (mean water depth: 15.4 m), Rotaliella arctica is also observed at 4450 m water depth on the Bermuda Rise (Pawlowski, 1991), and at 1980 m water depth in the Arctic (Scott and Vilks, 1991). Interestingly, dominant taxa in Community 6 (*Rotaliella*, Spirillinids, and *Spirophthalmidium*) are not common in carbonate sediments from shallower locations (300-1000 m) off the Little Bahama Bank in The Bahamas (Martin, 1988), or on the shallow (10-40 m) flooded carbonate Serranilla Bank (Triffleman et al., 1991) elsewhere in the Tropical North Atlantic Ocean. In caves, a deep-sea affinity is wellknown in higher taxonomic groups like crustaceans (Iliffe et al., 1983). In the Mediterranean, S. emaciatum has been recently described in benthic foraminiferal communities from a marine cave with carbonate sedimentation on the Murica coastal region in Spain (Cueva Gemelas: Bergamin et al., 2020), but not in marine caves on eastern Sardinia with siliciclastic sedimentation and evidence for the discharge of meteoric water (Romano et al., 2018, 2020). On the island of Cozumel (Mexico) where Aerolito Cave is located (Mejía-Ortiz et al., 2007a,b; CalderóN-GutiéRrez et al., 2018), caves passages flooded with oxygenated saline groundwater and benthic carbonate sediment also host populations of S. emaciatum (van Hengstum, personal observation). Murray (2006) states that Spirillina and Patellina prefer hard substrates, so it remains possible that the cave walls/ceiling may be providing an ideal natural hard substrate for these taxa. However, our research design is unable to assess this possibility (sampling benthic sediment, not cave walls). As suggested by Fichez (1991), perhaps complex organic matter is a critical resource for Community 6, which can only be evaluated by resolving biogeochemical cycling of nutrients in these areas (Fichez, 1990, 1991; Airoldi and Cinelli, 1996; Jian et al., 2020). Finally, Community 6 also occurs very close to the shoreline in Green Bay Cave (samples ISC-61 or ISC-73 See fig 2 in van Hengstum and Scott, 2011), yet does not receive enough nutrients to promote higher diversity and species richness like Community 5 from direct seawater transfer through the limestone pores. From a conservation perspective, these results indicate that any introduced physical cave openings from anthropogenic activity will change nutrient supply and communities of benthic meiofauna in marine sectors of KSEs.

Community 5 (including: 5A, 5B, and 5C) is colonizing benthic habitats experiencing more regular tidal groundwaterseawater exchange and likely higher sedimentation rates (Community 5B and 5C) or some terrestrial input (Community 5A). This is likely providing increased nutrient availability, and in turn is promoting higher species richness and diversity than benthic habitats colonized by Community 6 (**Table 1**).

Community 5 is not primarily influenced by: (i) strong tidal currents and attenuating light availability like Community 7, (ii) exposure from water table variability like Community 1, or (iii) increased organic matter input from the adjacent terrestrial landscape like Communities 2-4. Dominant taxa in Community 5 include Globocassidulina subglobosa that commonly occurs in carbonate sediments >1000 deep of the Little Bahama Bank (Martin, 1988), and Melonis barleeanum who is successful in areas with regular delivery of marine organic matter (Caralp, 1989). In Green Bay Cave, Community 5 is proximal to the large entrance (exit) into Harrington Sound. Tintinnid loricas (pelagic ciliates) are also deposited in the sediment with Community 5 in Green Bay Cave, and they were likely transported into the cave and settled out of the suspended sedimentary load. This observation supported the hypothesis that Community 5 in Green Bay Cave receives increased deposition of suspended sediments as the velocity of groundwater currents caused by tidal flow attenuate in the subsurface (van Hengstum and Scott, 2011), similar to what has been observed in marine caves on the Mediterranean coast of France (Fichez, 1991). An important taxonomic caveat is that small miliolids identified as Triloculina in Bermuda caves are not well-developed adult specimens with clear phenotypic characteristics like those common in subtidal marine carbonate lagoons (Javaux and Scott, 2003). Thus, it cannot be ruled-out that these are juvenile Quinqueloculina (Schnitker, 1967). Overall, these smaller cave miliolids are perhaps responding to a nutritional deficiency that is not constrained in this study.

Further evaluation of the physical environmental characteristics (Table 1; Figures 5-7) supports a tripartite division of the dendrogram (Figure 4, Bray-Curtis Similarity Index: 0.36). As previously discussed, the subtidal benthic foraminifera in Cow Cave (Community 5A, Cow Cave) is transitional between the foraminiferal communities that favor primary inputs from marine vs. terrestrial organic matter (Figure 5). While these samples are from shallow subtidal settings, benthic areas in Cow Cave receive protection from direct terrestrial sedimentation by the rock roof of the cave (i.e., geomorphology). Faunistically, samples from Palm Cave (Community 5C) closely compare with benthic areas in Green Bay Cave (Community 5B, Figure 4). However, finer substrates (Community 5B) had a greater abundance of Reophax spp. (mean: 3.1%) and Bolivina spp. (mean: 11.4%), while coarser substrates (Community 5C) had a greater abundance of Spirillina vivipara (mean: 13%) and Patellina-like taxa (mean: 7.8%). However, a careful evaluation of the cross plot between $\delta^{13}C_{org}$ and C:N (Figure 6) indicates that organic matter in Palm Cave (Community 5C) is slightly more carbon isotopically depleted with higher C:N values than Green Bay Cave (Community 5B). At least two hypotheses emerge to explain these differences between Community 5C and 5B that are related to cave morphology: either (i) proportionally less carbon isotopically enriched organic matter (e.g., seagrass, algae) is transported into Palm Cave through the smaller opening at Cripple Gate vs. the large marine opening of Green Bay Cave (Figure 3), or (ii) more ¹³C-depleted terrestrial organic matter is eroding into Palm Cave through its more numerous openings (exits) to the terrestrial landscape above (**Figure 2**). This suggests that differences in tidal exchange of seawater and particulate organic carbon transport through Cripple Gate (Palm Cave) and Green Bay Entrance (Green Bay Cave) may be influencing benthic habitat partitioning between caves, but other geochemical proxies are required to resolve these uncertainties. The collective role of cave geometry and volumetric exchange of seawater during tidal cycles between saline groundwater and Harrington Sound appear important for generating differences in the benthic foraminiferal community structures, and by corollary, benthic habitats in the oxygenated marine sectors of KSEs.

CONCLUSIONS

- Benthic foraminifera in the marine sector of karst subterranean estuaries (KSEs) in Bermuda can be grouped into 9 different communities that are correlated to organic matter source and quantity, tidal exposure, texture of the carbonate sediment substrate (i.e., mean particle size size), proximity to cave exits, and the relationship between cave morphology and tidally-forced circulation between coastal seawater and saline groundwater.
- The source and quantity of particulate organic carbon delivered to the benthos is significantly correlated to benthic foraminiferal community structure. A primary division emerges in foraminiferal communities correlated with organic carbon inputs primarily from the terrestrial surface with some marine plankton (mean $\delta^{13}C_{org}$ values <-23.2‰, C:N ratios >11), vs. organic carbon inputs dominantly from marine plankton (mean $\delta^{13}C_{org}$ values >-21.6‰, C:N ratios <10) that are transported into the caves from groundwater-seawater circulation from the adjacent carbonate lagoon (i.e., Harrington Sound).
- Benthic areas with more depleted $\delta^{13}C_{org}$ values of sediment are dominated by *Rosalina*, *Bolivina*, and *Triloculina*. In contrast, benthic areas with more enriched $\delta^{13}C_{org}$ values of sediment are dominated by *Patellina*, *Spirillina*, *Rotaliella*, *Miliolinella*, and *Spirophthalmidium*.
- Benthic areas with the least amount of organic carbon in the sediment (mean: TOC 5.1%, mean bulk organic content: 6%) are the least diverse, and dominated by *Spirophthalmidium emaciatum* (mean: 31%, maximum: 70%).
- Differences in the size and morphology of direct connections between Harrington Sound and Palm Cave

REFERENCES

- Airoldi, L., and Cinelli, F. (1996). Variability of fluxes of particulate material in a submarine cave with chemolithoautotrophic inputs of organic carbon. *Mar. Ecol. Prog. Ser.* 139, 205–217. doi: 10.3354/meps139205
- Altabet, M. A. (1990). Organic C, N, and stable isotopic composition of particulate matter colelcted on glass-fiber and aluminum oxide filters. *Limnol. Oceanogr.* 35, 902–909. doi: 10.4319/lo.1990.35.4.0902
- Anderson, B., Scalan, R. S., Behrens, E. W., and Parker, P. L. (1992). Stable carbon isotope variations in sediment from Baffin Bay, Texas, U.S.A.: evidence for

vs. Green Bay Cave are impacting benthic foraminiferal community structure.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/**Supplementary Material**, further inquiries can be directed to the corresponding author.

AUTHOR CONTRIBUTIONS

PvH designed the research. JC collected the samples from Palm Cave and completed laboratory activities. PvH and JC jointly interpreted that data, generated the artwork, and co-wrote the final manuscript. All authors contributed to the article and approved the submitted version.

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cyclic changes in organic matter source. *Chem. Geol. Isotope Geosci. Sect.* 101, 223–233. doi: 10.1016/0009-2541(92)90004-O

- Armynot du Châtelet, É., Bout-Roumazeilles, V., Riboulleau, A., and Trentesaux, A. (2009). Sediment (grain size and clay mineralogy) and organic matter quality control on living benthic foraminifera. *Rev. Micropaléontol.* 52, 75–84. doi: 10.1016/j.revmic.2008.10.002
- Balduzzi, A., and Cattaneo, R. (1985). Estimation par relèvements photographiques de la distribution de *Miniacina miniacea* (Sarcomastigophora, Foraminifera) dans une grotte sous-marine du Golfe de Naples. *Rapp. P. V. Comm. Int. Explor. Sci. Mer Méditerr.* 29, 127–129.

- Bates, N. R. (2017). Twenty years of marine carbon cycle observations at Devils Hole Bermuda provide insights into seasonal hypoxia, coral reef calcification, and ocean acidification. *Front. Mar. Sci.* 4:372. doi: 10.3389/fmars.2017.00036
- Bergamin, L., Marassich, A., Provenzani, P., and Romano, E. (2018). Foraminiferal ecozones in two submarine caves of the Orosei Gulf (Sardinia, Italy). *Rend. Lincei Sci. Fis. Nat.* 29, 547–557. doi: 10.1007/s12210-018-0700-0
- Bergamin, L., Taddei Ruggiero, E., Pierfranceschi, G., Andres, B., Costantino, R., Crovato, C., et al. (2020). Benthic foraminifera and brachiopods from a marine cave in Spain: environmental significance. *Mediterr. Mar. Sci.* 21, 506–518. doi: 10.12681/mms.23482
- Bermúdez, P. J. (1949). Tertiary Smaller Foraminifera of the Dominican Republic. Cushman Laboratory for Foraminiferal Research Special Publication, 25, 1–322.
- Bernhard, J. M., and Sen Gupta, B. K. (1997). "Foraminifera of oxygen-depleted environments," in *Modern Foraminifera*, ed B. K. Sen Gupta (Boston, MA: Kluwer Academic Press), 201–216. doi: 10.1007/0-306-48104-9_12
- Bishop, R. E., Humphreys, W. F., Cukrov, N., Žic, V., Boxshall, G. A., Cukrov, M., et al. (2015). "Anchialine" redefined as a subterranean estuary in a crevicular or cavernous geological setting. J. Crust. Biol. 35, 511–514. doi: 10.1163/1937240X-00002335
- Brady, H. B. (1884). Report on the foraminifera dredged by the H.M.S. Challenger during the years 18731–1876: reports on the scientific results of the Voyage of the H.M.S. Challenger during the years 18731–1876. *Zoology* 9:814.
- Brankovits, D., Pohlman, J. W., Ganju, N. K., Iliffe, T. M., Lowell, N., Roth, E., et al. (2018). Hydrologic controls of methane dynamics in karst subterranean estuaries. *Global Biogeochem. Cycles* 32, 1759–1775. doi: 10.1029/2018GB006026
- Brankovits, D., Pohlman, J. W., Niemann, H., Leigh, M. B., Leewis, M. C., Becker, K. W., et al. (2017). Methane- and dissolved organic carbon-fueled microbial loop supports a tropical subterranean estuary ecosystem. *Nat. Commun.* 8:1835. doi: 10.1038/s41467-017-01776-x
- Burgett, C. M., Burkholder, D. A., Coates, K. A., Fourqurean, V. L., Kenworthy, W. J., Manuel, S. A., et al. (2018). Ontogenetic diet shifts of green sea turtles (*Chelonia mydas*) in a mid-ocean developmental habitat. *Mar. Biol.* 165:33. doi: 10.1007/s00227-018-3290-6
- Buzas, M. A. (1968). On the spatial distribution for for a minifera. Contrib. Cushman Found. For a miniferal Res. 19, 1–11.
- Calderón-GutiéRrez, F., SáNchez-Ortiz, C. A., and Huato-Soberanis, L. (2018). Ecological patterns in anchialine caves. *PLoS ONE* 3:e0202909. doi: 10.1371/journal.pone.0202909
- Camacho, S., Moura, D., Connor, S., Scott, D., and Boski, T. (2015). Ecological zonation of benthic foraminifera in the lower Guadiana Estuary (southeastern Portugal). *Mar. Micropaleontol.* 114, 1–18. doi: 10.1016/j.marmicro.2014.10.004
- Cant, R. V., and Weech, P. S. (1986). A review of the factors affecting the development of Ghyben-Hertzberg lenses in the Bahamas. J. Hydrol. 84, 333–343. doi: 10.1016/0022-1694(86)90131-9
- Caralp, M. H. (1989). Size and morphology of the benthic foraminifera *Melonis* barleeanum: relationships with marine organic matter. J. Foraminiferal Res. 19, 235–245. doi: 10.2113/gsjfr.19.3.235
- Carman, K. W. (1933). The Shallow-Water Foraminifera of Bermuda. Cambridge, MA: Massachusetts Institute of Technology.
- Carmen, K. W. (1927). The Shallow-Water Foraminifera of Bermuda. Cambridge, MA: Massachusetts Institute of Technology.
- Chávez-Solís, M., Solís, C., Simões, N., and Mascaró, M. (2020). Distribution patterns, carbon sources, and niche partitioning in cave shrimps (Atyidae: Typhlatya). Sci. Rep. 10:12812. doi: 10.1038/s41598-020-69562-2
- Clarke, K. R., Somerfield, P. J., and Gorley, R. N. (2008). Testing of null hypotheses in exploratory community analyses: similarity profiles and biota-environment linkage. J. Exp. Mar. Biol. Ecol. 366, 56–69. doi: 10.1016/j.jembe.2008.07.009
- Coull, B. (1969). Hydrographic control of meiobenthos in Bermuda. Oecologia 14, 953–957. doi: 10.4319/lo.1969.14.6.0953
- Debenay, J.-P., Tsakiridis, E., Soulard, R., and Grossel, H. (2001). Factors determining the distribution of foraminiferal assemblages in Port Joinville Harbor (Ile d'Yeu, France): the influence of pollution. *Mar. Micropaleontol.* 43, 75–118. doi: 10.1016/S0377-8398(01)00023-8

- Debenay, J. P., and Guillou, J. J. (2002). Ecological transitions indicated by foraminiferal assemblages in paralic environment. *Estuaries* 6A, 1107–1120. doi: 10.1007/BF02692208
- Diz, P., Francés, G., Costas, S., Souto, C., and Alejo, I. (2004). Distribution of benthic foraminifera in coarse sediments, Ría de Vigo, NW Iberian margin. J. Foraminiferal Res. 34, 258–275. doi: 10.2113/34.4.258
- Dupuy, C., Rossignol, L., Geslin, E., and Pascal, P. Y. (2010). Predation of mudflat meio-macrofaunal metazoans by a clacareous foraminifer, *Ammonia tepida* (Cushman 1926). J. Foraminiferal Res. 40, 305–3012. doi: 10.2113/gsjfr.40.4.305
- Duque, C., Michael, H. A., and Wilson, A. M. (2020). The subterranean estuary: technical term, simple analogy, or source of confusion? *Water Resour. Res.* 56:e2019WR026554. doi: 10.1029/2019WR026554
- Fichez, R. (1990). Decrease in allochthonous organic inputs in dark submarine caves, connection with lowering in benthic community richness. *Hydrobiologia* 207, 61–69. doi: 10.1007/BF00041441
- Fichez, R. (1991). Suspended particulate organic matter in a Mediterranean submarine cave. Mar. Biol. 108, 167–174. doi: 10.1007/BF01313485
- Fourqurean, J. W., Manuel, S. A., Coates, K. A., Kenworthy, W. J., and Boyer, J. N. (2015). Water quality, isoscapes, and stoichioscapes of seagrasses indicate general P limitation and unique N cycling in shallow water benthos of Bermuda. *Biogeosciences* 12, 6235–6249. doi: 10.5194/bg-12-6235-2015
- Fourqurean, J. W., Manuel, S. A., Coates, K. A., Massey, S. C., and Kenworthy, W. J. (2019). Decadal monitoring in Bemruda shows a widespread loss of seagrasses attributable to overgrazing by the green sea turtle *Chelonia mydas. Estuaries Coasts* 42, 1524–1540. doi: 10.1007/s12237-019-00587-1
- France, R. L. (1995). Carbon-13 enrichment in benthic compared to planktonic algae: foodweb implications. *Mar. Ecol. Prog. Ser.* 124, 307–312. doi: 10.3354/meps124307
- Fry, B., Scalan, R. S., and Parker, P. L. (1977). Stable carbon isotope evidence for two sources of organic matter in coastal sediments: seagrasses and plankton. *Geochem. Cosmochim. Acta* 41, 1875–1877. doi: 10.1016/0016-7037(77) 90218-6
- Gehrels, W. R., and van de Plassche, O. (1999). The use of Jadammina macrescens (Brady) and Balticammina pseudomacrescens Brönnimann, Lutze, and Whittaker (Protozoa: Foraminiferida) as sea-level indicators. *Palaeogeogr. Palaeoclimatol. Palaeoecol.* 149, 89–101. doi: 10.1016/S0031-0182(98) 00194-1
- Goericke, R., and Fry, B. (1994). Variations of marine plankton delta-13-C with latitude, temperature and dissolved CO2 in the world ocean. *Global Biogeochem. Cycles* 8, 85–90. doi: 10.1029/93GB03272
- Goldstein, S. T., and Corliss, B. H. (1994). Deposit feeding in selected deepsea and shallow-water benthic foraminifera. *Deep Sea Res.* 41, 229–241. doi: 10.1016/0967-0637(94)90001-9
- Gonneea, M. E., Charette, M. A., Liu, Q., Herrera-Silveira, J. A., and Morales-Ojeda, S. M. (2014). Trace element geochemistry of groundwater in a karst subterranean estuary (Yucatan Peninsula, Mexico). *Geochim. Cosmochim. Acta* 132, 31–49. doi: 10.1016/j.gca.2014.01.037
- Gooday, A. J. (1988). A response by benthic foraminifera to the deposition of pytodetritus in the deep sea. *Nature* 332, 70–73. doi: 10.1038/332070a0
- Gooday, A. J. (1993). Deep-sea benthic foraminiferal species which exploit phytodetritus-characteristic features and controls on distribution. *Mar. Micropaleonol.* 22, 187–205. doi: 10.1016/0377-8398(93)90043-W
- Gooday, A. J., Levin, L. A., Linke, P., and Heeger, T. (1992). The role of benthic foraminifera in deep-sea food webs and carbon cycling. *Deep Sea Food Chains Global Carbon Cycle* 360, 63–91. doi: 10.1007/978-94-011-2452-2_5
- Gotelli, N. J., and Colwell, R. K. (2011). "Estimating species richness," in *Biological Diversity: Frontiers in Measurement and Assessment*, eds A. E. Magurran and B. J. Mcgill (Oxford: Oxford University Press), 39–54.
- Guido, A., Rosso, A., Sanfilippo, R., Russo, F., and Mastandrea, A. (2017). Microbial biomineralization in biotic crusts from a pleistocene marine cave (NW Sicily, Italy). *Geomicrobiol. J.* 34, 864–872. doi: 10.1080/01490451.2017.1284283
- Guilbault, J.-P., Clague, J. J., and Lapointe, M. (1995). Amount of subsidence during a late Holocene earthquake—evidence from fossil tidal marsh foraminifera at Vancouver Island, west coast of Canada. *Palaeogeogr. Palaeoclimatol. Palaeoecol.* 118, 49–71. doi: 10.1016/0031-0182(94)0 0135-U

- Haig, D. W. (1988). Miliolid foraminfiera from the inner neritic sand and mud facies of the Papuan Lagoon, New Guinea. J. Foraminiferal Res. 18, 203–236. doi: 10.2113/gsjfr.18.3.203
- Hayward, B. W., Grenfell, H. R., and Scott, D. B. (1999). Tidal range of marsh foraminifera for determining former sea-level heights in New Zealand. N. Z. J. Geol. Geophys. 42, 395–413. doi: 10.1080/00288306.1999.9514853
- Heiri, O., Lotter, A. F., and Lemcke, G. (2001). Loss on ignition as a method for estimating organic and carbonate content in sediments: reproducibility and comparability of results. *J. Paleolimnol.* 25, 101–110. doi: 10.1023/A:1008119611481
- Hermelin, J. O. R., and Scott, D. B. (1985). Recent benthic foraminifera from the central North Atlantic. *Micropaleontology* 31, 199–220. doi: 10.2307/1485542
- Iliffe, T. M. (2008). Groundwater pollution and its impact on Bermuda's reefs and inshore water, NOAA Coral Reef Conservation Grant–Final Report, National Oceanic and Atmospheric Administration.
- Iliffe, T. M., Hart, C. W. J., and Manning, R. B. (1983). Biogeography and the caves of Bermuda. Nature 302, 141–142. doi: 10.1038/302141a0
- Iliffe, T. M., Wilkens, H., Parzefall, J., and Williams, D. (1984). Marine lava cave fauna: composition, biogeography, and origins. *Science* 225, 309–311. doi: 10.1126/science.225.4659.309
- Javaux, E., and Scott, D. B. (2003). Illustration of modern benthic foraminifera from Bermuda and remarks on distributions in other subtropical/tropical areas. *Paleontol. Electron.* 6:29. Available online at: http://palaeo-electronica. org/paleo/2003_1/benthic/issue1_03.htm
- Jeffrey, A. W. A., Pflaum, R. C., Brooks, J. M., and Sackett, W. M. (1983). Vertical trends in particulate organic carbon 13C:12C ratios in the upper water column. *Deep Sea Res.* 30, 971–983. doi: 10.1016/0198-0149(83)90052-3
- Jian, S., Zhang, Y., Jin, J., Wu, Y., Wei, Y., Wang, X., et al. (2020). Organic carbon in a seepage face of a subterranean estuary: turnover and microbial interrelations. *Sci. Tot. Environ.* 725:138220. doi: 10.1016/j.scitotenv.2020.138220
- Jorissen, F. J., De Stigter, H. C., and Widmark, J. G. V. (1995). A conceptual model explaining benthic foraminifera microhabitats. *Mar. Micropaleontol.* 26, 3–15. doi: 10.1016/0377-8398(95)00047-X
- Juggins, S. (2015). *Rioja: Analysis of Quaternary Science Data. R-Package 0.9-5.* Available online at: https://cran.r-project.org/web/packages/rioja/index.html
- Kennedy, H., Beggins, J., Duarte, C. M., Fourqurean, J. W., Holmer, M., Marba, N., et al. (2010). Seagrass sediments as a global carbon sink: Isotopic constraints. *Global Biogeochem. Cycles* 24:2010GB4026. doi: 10.1029/2010GB0 03848
- Khan, N. S., Vane, C. H., Englehart, S. E., Kendrick, C., and Horton, B. P. (2019). The application of δ13C, TOC, and C/N geochemistry of mangrove sediments to reconstruct Holocene paleoenvironments and relative sea levels, Puerto Rico. *Mar. Geol.* 415:105963. doi: 10.1016/j.margeo.2019.105963
- Kuhnt, T., Schmiedl, G., Ehrmann, W., Hamann, Y., and Hemleben, C. (2007). Deep-sea ecosystem variability of the Aegean Sea during the past 22 kyr as revealed by benthic foraminifera. *Mar. Micropaleontol.* 64, 141–162. doi: 10.1016/j.marmicro.2007.04.003
- Lamb, A. L., Wilson, G. P., and Leng, M. J. (2006). A review of coastal palaeoclimate and relative sea-level reconstructions using δ13C and C/N ratios in organic material. *Earth Sci. Rev.* 75, 29–57. doi: 10.1016/j.earscirev.200 5.10.003
- Land, L. S., Mackenzie, F. T., and Gould, S. J. (1967). Pleistocene history of Bermuda. Geol. Soc. Am. Bull. 78, 993–1006. doi: 10.1130/0016-7606(1967)78[993:PHOB]2.0.CO;2
- Legendre, P., and Legendre, L. (1998). Numerical ecology: developments in environmental modelling. *Dev. Environ. Model.* 20, 1–853.
- Levin, L. A., Ekau, W., Gooday, A. J., Jorissen, F. J., Middelburg, J. J., Naqvi, S. W. A., et al. (2009). Effects of natural and human-induced hypoxia on coastal benthos. *Biogeosciences* 6, 2063–2098. doi: 10.5194/bg-6-2063-2009
- Linke, P., Altenbach, A. V., Graf, G., and Herger, T. (1995). Response of deep-sea benthic foraminifera to a simulated sedimentation event. J. Foraminiferal Res. 25, 75–82. doi: 10.2113/gsjfr.25.1.75
- Little, S. N., and van Hengstum, P. J. (2019). Intertidal and subtidal benthic foraminifera in flooded caves: implications for reconstructing coastal karst aquifers and cave paleoenvironments. *Mar. Micropaleontol.* 149, 19–34. doi: 10.1016/j.marmicro.2019.03.005
- Loeblich Jr, A. R., and Tappan, H. (1987). Foraminiferal Genera and Their Classification. Boston, MA: Springer.

- Martin, J. B., Gulley, J., and Spellman, P. (2012). Tidal pumping of water between Bahamian blue holes, aquifers, and the ocean. J. Hydrol. 416–417, 28–38. doi: 10.1016/j.jhydrol.2011.11.033
- Martin, R. E. (1988). Benthic foraminiferal zonation in deep-water carbonate platform margin environments, northern Little Bahama Bank. J. Paleontol. 62, 1–8. doi: 10.1017/S0022336000017935
- Mejía-Ortiz, L. M., Yánez, G., and López-Mejía, M. (2007a). Echinoderms in an anchialine cave in Mexico. *Mar. Ecol.* 28, 31–34. doi: 10.1111/j.1439-0485.2007.00174.x
- Mejía-Ortiz, L. M., Yánez, G., López-Mejía, M., and Zarza-Gónzales, E. (2007b). Cenotes (anchialine caves) on Cozumel Island, Quintana Roo, Mexico. J. Cave Karst Stud. 69, 250–255.
- Moore, W. S. (1999). The subterranean estuary: a reaction zone of ground water and sea water. *Mar. Chem.* 65, 111–125. doi: 10.1016/S0304-4203(99)00014-6
- Morris, B., Barnes, J., Brown, F., and Markham, J. (1977). The Bermuda Marine Environment: A Report of the Bermuda Inshore Waters Investigations 1976– 1977. St. Georges: Bermuda Biological Station.
- Murray, J. (2006). *Ecology and Applications of Benthic Foraminifera*. London: Cambridge University Press.
- Mylroie, J. E., Carew, J. L., and Vacher, H. L. (1995). "Karst development in the Bahamas and Bermuda," in *Terrestrial and Shallow Marine Geology of the Bahamas and Bermuda*, eds H. A. Curran and B. White (Boulder, CO: Geologic Society of America), 251–267.
- Neumann, A. C. (1965). Processes of recent carbonate sedimentation in Harrington Sound, Bermuda. Bull. Mar. Sci. 15, 987–1035.
- Nomaki, H., Heinz, P., Hemleben, C., and Kitazato, H. (2005). Behavior and response of deep-sea benthic foraminifera to freshly supplied organic matter: a laboratory feeding experiment in microcosm environments. *J. Foraminiferal Res.* 35, 103–113. doi: 10.2113/35.2.103
- Oksanen, J., Blanchet, F. G., Kindt, R., Legendre, P., Minchin, P. R., O'hara, R. B., et al. (2013). *Vegan: Community Ecology Package. R Package Version 3*. Available online at: https://cran.r-project.org/web/packages/vegan/index.html
- Omori, A., Kitamura, A., Fujita, K., Honda, K., and Yamamoto, N. (2010). Reconstruction of light conditions within a submarine cave during the past 7000 years based on the temporal and spatial distribution of algal symbiontbearing large benthic foraminifers. *Palaeogeogr. Palaeoclimatol. Palaeoecol.* 292, 443–452. doi: 10.1016/j.palaeo.2010.04.004
- Pain, A. J., Martin, J. B., and Young, C. R. (2019). Sources and sinks of CO2 and CH4 in siliciclastic subterranean estuaries. *Limnol. Oceanogr.* 64, 1500–1514. doi: 10.1002/lno.11131
- Pain, A. J., Martin, J. B., Young, C. R., Valle-Levinson, A., and Mariño-Tapia, I. (2020). Carbon and phosphorus processing in a carbonate karst aquifer and delivery to the coastal ocean. *Geochimi. Cosmochim. Acta* 269, 484–495. doi: 10.1016/j.gca.2019.10.040
- Palmer, A., Mv, P., and Jm, Q. (1977). "Geology and origin of the caves in Bermuda," in *Proceeding of the 7th International Speleological Congress* (Sheffield), 336–339.
- Parsons, R. J., Nelson, C. A., Denman, C. C., Andersson, A. J., Kledzik, A. L., Vergin, K. L., et al. (2015). Marine bacterioplankton community turnover within seasonally hypoxic waters of a subtropical sound: Devil's Hole, Bermuda. *Environ. Microbiol.* 17, 3481–3499. doi: 10.1111/1462-2920. 12445
- Patterson, R. T., and Fishbein, E. (1989). Re-examination of the statistical methods used to determine the number of point counts needed for micropaleontological quantitative research. J. Paleontol. 63, 245–248. doi: 10.1017/S0022336000019272
- Pawlowski, J. (1991). Distribution and taxonomy of some benthic tiny foraminifers from the Bermuda rise. *Micropaleontology* 37, 163–172. doi: 10.2307/14 85556
- Pirsson, L. V., and Vaughn, T. W. (1917). A deep Boring in Bermuda Island. Am. J. Sci. Ser. s4-36, 70-71. doi: 10.2475/ajs.s4-36.211.70
- Plummer, L. N., Vacher, H. L., Mckenzie, F. T., Bricker, O. P., and Land, L. S. (1976). Hydrogeochemistry of Bermuda: a case history of groundwater diagenesis of biocalcarenites. *Geol. Soc. Am. Bull.* 87, 1301–1316. doi: 10.1130/0016-7606(1976)87<1301:HOBACH>2.0.CO;2
- Proctor, C. J., and Smart, P. L. (1991). A dated sediment record of Pleistocene transgressions on Berry Head, Southwest England. J. Quat. Sci. 6, 233–244. doi: 10.1002/jqs.3390060306

- Radolović, M., Bakran-Petricioli, T., Petricioli, D., Surić, M., and Perica, D. (2015). Biological response to geochemical and hydrological processes in a shallow submarine cave. *Mediterr. Mar. Sci.* 16, 305–324. doi: 10.12681/mms.1146
- Riedl, R., and Ozretić, B. (1969). Hydrobiology of marginal caves. Part 1: general problems and introduction. *Int. Rev. Gesamten Hydrobiol. Hydrogr.* 54, 661–683. doi: 10.1002/iroh.19690540503
- Romano, E., Bergamin, L., Di Bella, L., Frezza, V., Marassich, A., Pierfranceschi, G., et al. (2020). Benthic foraminifera as proxies of marine influence in the Orosei marine caves (Sardinia, Italy). *Aquat. Conserv. Mar. Freshwater Ecosyst.* 30, 701–716. doi: 10.1002/aqc.3288
- Romano, E., Bergamin, L., Pierfranceschi, G., Provenzani, C., and Marassich, A. (2018). The distribution of benthic foraminifera in Bel Torrente submarine cave (Sardinia, Italy) and their environmental significance. *Mar. Environ. Res.* 133, 114–127. doi: 10.1016/j.marenvres.2017.12.014
- Rosso, A., Sanfilippo, R., Bonfiglio, L., Richards, D. A., and Nita, D. C. (2018). Exceptional Pleistocene vermetid crusts preserved in a cave located 130 m above sea level near Taormina (NE Sicily). *Boll. Soc. Paleontol. Ital.* 57:134. doi: 10.4435/BSPI.2018.09
- Rosso, A., Sanfilippo, R., Ruggieri, R., Maniscalco, R., and Vertino, A. (2015). Exceptional record of submarine cave communities from the Pleistocene of Sicily (Italy). *Lethaia* 48, 133–144. doi: 10.1111/let.12094
- Saraswat, R., Kouthanker, M., Kurtarkar, S. R., Nigam, R., Naqvi, S. W. A., and Linshy, V. N. (2015). Effect of salinity induced pH/alkalinity changes on benthic foraminifera: a laboratory culture experiment. *Estuar. Coast. Shelf Sci.* 153, 96–107. doi: 10.1016/j.ecss.2014.12.005
- Saraswat, R., Nigam, R., and Pachkhande, S. (2011). Difference in optimum temperature for growth and reproduction in benthic foraminifer Rosalina globularis: implications for paleoclimatic studies. J. Exp. Mar. Biol. Ecol. 405, 105–110. doi: 10.1016/j.jembe.2011.05.026
- Schmitter-Soto, J. J., Comín, F. A., Escobar-Briones, E., Herrera-Silveira, J., Alcocer, J., Suárez-Morales, E., et al. (2002). Hydrogeochemical and biological characteristics of cenotes in the Yucatan Peninsula (SE Mexico). *Hydrobiologia* 467, 215–228. doi: 10.1023/A:1014923217206
- Schnitker, D. (1967). Variation in Test Morphology of Triloculina linneiana d'Orbigny in Laboratory Cultures. Contributions from the Cushman Foundation for Foraminiferal Research, Vol. 18, 85–86.
- Scott, D. B., and Hermelin, J. (1993). A device for precision splitting of micropaleontological samples in liquid suspension. J. Paleontol. 67, 151–154. doi: 10.1017/S0022336000021302
- Scott, D. B., and Medioli, F. S. (1978). Verticle zonation of marsh foraminifera as accurate indicators of former sea levels. *Nature* 272, 528–531. doi: 10.1038/272528a0
- Scott, D. B., and Medioli, F. S. (1980). Living vs. total foraminiferal populations: their relative usefulness in paleoecology. J. Paleontol. 54, 814–813.
- Scott, D. B., and Vilks, G. (1991). Benthonic foraminifera in the surface sediments of the deep-sea Arctic Ocean. J. Foraminiferal Res. 21, 20–38. doi: 10.2113/gsjfr.21.1.20
- Sen Gupta, B. K., Turner, R. E., and Rabalais, N. N. (1996). Seasonal oxygen depletion in continental-shelf waters of Louisiana: historical record of benthic foraminifers. *Geology* 24, 227–230. doi: 10.1130/0091-7613(1996)024<0227:SODICS>2.3.CO;2
- Sperazza, M., Moore, J. N., and Hendrix, M. S. (2004). High-resolution particle size analysis of naturally occurring very fine-grained sediment through laser diffractometry. J. Sediment. Res. 74, 736–743. doi: 10.1306/031104740736
- Steinker, D. C., and Clem, K. V. (1984). Some nearshore foraminiferal assemblages from phytal substrates and bottom sediments, Bermuda. *Compass* 61, 98–115.
- Stock, J. H., Iliffe, T. M., and Williams, D. (1986). The concept "anchialine" reconsidered. *Stygologia* 2, 90–92.
- Suárez-Morales, E., Reid, J. W., Fiers, F., and Iliffe, T. M. (2004). Historical biogeography and distribution of the freshwater cyclopine copepods (Copepoda, Cyclopoida, Cyclopinae) of the Yucatan Peninsula, Mexico. *J. Biogeogr.* 31, 1051–1063. doi: 10.1111/j.1365-2699.2004.01053.x

- Tamalavage, A. E., van Hengstum, P. J., Louchouarn, P., Kaiser, K., Donnelly, J. P., Albury, N. A., et al. (2018). Organic matter sources and lateral sedimentation in a Bahamian karst basin (sinkhole) over the late Holocene: influence of local vegetation and climate. *Palaeogeogr. Palaeoclimatol. Palaeoecol.* 506, 70–83. doi: 10.1016/j.palaeo.2018.06.014
- Triffleman, N. J., Hallock, P., Hine, A. C., and Peebles, M. W. (1991). Distribution of foraminfieral tests in sediments of Serranilla Bank, Nicaraguan Rise, Southwestern Caribbean. J. Foraminiferal Res. 21, 39–47. doi: 10.2113/gsjfr.21.1.39
- Vacher, H., Hearty, P., and Rowe, M. (1995). Terrestrial and Shallow Marine Geology of the Bahamas and Bermuda. Boulder, CO: Geological Society of America.
- Vacher, H., and Rowe, M. (1997). *Geology and Hydrogeology of Carbonate Islands*. Amsterdam: Elsevier.
- Vacher, H. L. (1978). Hydrogeology of Bermuda—significance of an across-the-island variation in permeability. J. Hydrol. 39, 207–226. doi: 10.1016/0022-1694(78)90001-X
- van Hengstum, P. J., Cresswell, J. N., Milne, G. A., and Iliffe, T. M. (2019). Development of anchialine cave habitats and karst subterranean estuaries since the last ice age. *Sci. Rep.* 9:11907. doi: 10.1038/s41598-019-48058-8
- van Hengstum, P. J., Donnelly, J. P., Kingston, A. W., Williams, B. E., Scott, D. B., Reinhardt, E. G., et al. (2015). Low-frequency storminess signal at Bermuda linked to cooling events in the North Atlantic region. *Paleoceanography* 30, 52–76. doi: 10.1002/2014PA002662
- van Hengstum, P. J., Reinhardt, E. G., Beddows, P. A., Huang, R. J., and Gabriel, J. J. (2008). Thecamoebians (testate amoebae) and foraminifera from three anchialine cenotes in Mexico: Low salinity (1.5–4.5 psu) faunal transitions. J. Foraminiferal Res. 38, 305–317. doi: 10.2113/gsjfr.38.4.305
- van Hengstum, P. J., Reinhardt, E. G., Beddows, P. A., Schwarcz, H. P., and Garbriel, J. J. (2009). Foraminifera and testate amoebae (thecamoebians) in an anchialine cave: surface distributions from Aktun Ha (Carwash) cave system, Mexico. *Limnol. Oceanogr.* 54, 391–396. doi: 10.4319/lo.2009.54. 1.0391
- van Hengstum, P. J., and Scott, D. B. (2011). Ecology of foraminifera and habitat variability in an underwater cave: distinguishing anchialine versus submarine cave environments. *J. Foraminiferal Res.* 41, 201–229. doi: 10.2113/gsjfr.41.3.201
- van Hengstum, P. J., and Scott, D. B. (2012). Sea-level rise and coastal circulation controlled Holocene groundwater development and caused a meteoric lens to collapse 1600 years ago in Bermuda. *Mar. Micropaleonol.* 90–91, 29–43. doi: 10.1016/j.marmicro.2012.02.007
- Wefer, G., and Killingley, J. S. (1986). Carbon isotopes in organic matter from a benthic alga *Halimeda incrassata* (Bermuda): effect of light intensity. *Chem. Geol. Isotope Geosci. Sect.* 59, 321–326. doi: 10.1016/0168-9622(86)90081-3
- Whitaker, D., and Christman, M. (2014). Sigclust: Significant Cluster Analysis. R Package Version 1, 2.4–2. Available online at: https://cran.r-project.org/web/ packages/sigclust/
- Woolard, G. P., and Ewing, M. (1939). Structural geology of the Bermuda Islands. *Nature* 143:898. doi: 10.1038/143898a0

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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