Check for updates

OPEN ACCESS

EDITED BY Weihao Wang, Peking University, China

REVIEWED BY Zhiqiang Huang, Nanjing University, China Yao Ma, Chinese Academy of Medical Sciences and Peking Union Medical College, China

*CORRESPONDENCE Chunguang Xie 💌 xiecg@cdutcm.edu.cn Xiyu Zhang 💌 zhangxiyutcm@126.com

RECEIVED 22 August 2024 ACCEPTED 30 December 2024 PUBLISHED 21 January 2025

CITATION

Tang R, Xie C and Zhang X (2025) NOD1: a metabolic modulator. *Front. Endocrinol.* 15:1484829. doi: 10.3389/fendo.2024.1484829

COPYRIGHT

© 2025 Tang, Xie and Zhang. This is an openaccess article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

NOD1: a metabolic modulator

Ruobing Tang¹, Chunguang Xie^{1,2,3*} and Xiyu Zhang^{1,2,3*}

¹Hospital of Chengdu University of Traditional Chinese Medicine, Chengdu, Sichuan, China, ²TCM Regulating Metabolic Diseases Key Laboratory of Sichuan Province, Chengdu, Sichuan, China, ³Department of Endocrinology, Hospital of Chengdu University of Traditional Chinese Medicine, Chengdu, Sichuan, China

Nucleotide-binding oligomerization domain 1 (NOD1) is an intracellular pattern recognition receptor that detects injury signals and initiates inflammatory responses and host defense. Furthermore, NOD1 serves as a metabolic mediator by influencing the metabolism of various tissues, including adipose tissue, liver, cardiovascular tissue, pancreatic β cells, adrenal glands, and bones through diverse mechanisms. It has been discovered that activated NOD1 is associated with the pathological mechanisms of certain metabolic diseases. This review presents a comprehensive summary of the impact of NOD1 on tissue-specific metabolism.

KEYWORDS

NOD1, metabolism, endocrine, inflammation, immunity

1 Introduction

Nucleotide-binding oligomerization domain-1 (NOD1), a member of the NOD-like receptor family, is an intracellular immunoregulatory protein (1, 2). NOD1 is widely distributed in antigen-presenting cells and epithelial cells (3). It recognizes damage signals and triggers an inflammatory response to mediate host defense (4). It has been demonstrated that NOD1 and its downstream pathways activated in the adipose tissue of individuals diagnosed with metabolic syndrome, and gestational diabetes mellitus, as well as those who are overweight or obese (5–7). Additionally, there is a significant increase in NOD1 expression in the myocardium of individuals diagnosed with type 2 diabetes mellitus (8). These studies indicate that NOD1 may be involved in metabolic processes and could play a role in metabolic diseases. This article provides a comprehensive review of the impact of NOD1 on metabolism across various tissues and organs, highlighting its potential significance in understanding metabolic disorders. Furthermore, it introduces the potential for NOD1 to serve as a risk assessment marker for metabolic diseases.

2 NOD1

The NOD1 protein consists of three domains. The C-terminus contains leucine-rich repeats, which are capable of recognizing specific ligands. The central nucleotide-binding domain exhibits ATPase activity and undergoes conformational changes upon ligand activation, thus promoting the oligomerization of NOD1. The N-terminal contains a

caspase-activation-and-recruitment-domain (CARD), which is used to recruit downstream effectors (9–11).

NOD1 is a classical cytoplasmic pattern recognition receptor capable of recognizing pathogen-associated molecular patterns (1, 12). NOD1 specifically recognizes the peptidoglycan (PGN) fragment γ -Dglutamyl-meso-diaminopimelic acid (iE-DAP) on the bacterial cell wall (1, 13). Additionally, NOD1 was found activated during viral, fungal, and parasitic infections. For example, the hepatitis C virus synthesizes dsRNA through a nonstructural protein 5B RNA-dependent RNA polymerase. dsRNA interacts with NOD1 to promote its activation (12). The parasitic nematode L. sigmodontis has been identified as a host of the intracellular symbiont Wolbachia, which possesses all the necessary enzymes for synthesizing lipid II, the precursor of bacterial peptidoglycan. Consequently, L. sigmodontis is capable of activating NOD1 (14). Furthermore, there have been reports of human cytomegalovirus (15), Aspergillus fumigatus (16, 17), Trypanosoma cruzi (18), and Plasmodium berghei (19) activating NOD1. However, the precise mechanism of their activation remains unclear. In addition to recognizing pathogen-associated molecules, NOD1 is also capable of recognizing danger-associated molecular signals. A deficiency in NODs has been observed to attenuate the inflammatory response by DTT, an endoplasmic reticulum (ER) stress inducer (20). Peptidoglycanindependent Brucella abortus has been demonstrated to induce ER stress through the type IV secretion system, thereby activating the NOD1/RIP2 signaling pathway (20). The aforementioned evidence suggests that ER stress may serve as one of the signals for NOD1 activation (20). Interestingly, thapsigargin, an ER stress agonist and a sarcoplasmic reticulum Ca²⁺ ATPase inhibitor has been demonstrated to induce ER stress by mediating calcium influx. It subsequently leads to the internalization of peripheral micro-peptidoglycan and activates cytoplasmic NOD1. Indicates that calcium influx contributes to NOD1 activation (21). Further investigation is required to elucidate the independent role of calcium influx and ER stress in NOD1 activation. Moreover, NOD1 detects aberrant expression of small Rho GTPases triggered by perturbations in host cell function caused by bacterial virulence factors (22). In this way, NOD1 identifies the pathogenic factors of Salmonella enterica (23) and Shigella flexneri (24). Recently, it has been found that the activated small Rho GTPase Rac1 activates intracellular NOD1 during hematopoietic stem and progenitor cell differentiation. NOD1-mediated "Developmental Inflammation" induces hematopoietic stem and progenitor cells to differentiate into hemogenic endothelium (25). It again demonstrates the ability of NOD1 to sense intracellular perturbations and signals from small Rho GTPases, suggesting that NOD1 influences the physiological function of cell fate. However, further study is needed to determine whether there is selectivity in NOD1 recognition of such signals.

After NOD1 is activated, it interacts through the CARD domain to bind with receptor-interacting kinase 2 (RIP2) receptors to form protein complexes (26). E3 ligases, such as inhibitors of apoptosis proteins (IAP) like c-IAP1, c-IAP2, and XIAP (27, 28), mediate the conjugation of RIP2 to the K63 ubiquitin chain (29). Polyubiquitylated RIP2 recruits TAK1 and interacts with the I κ B kinase (IKK) subunit NEMO (IKK γ) to recruit the IKK complex. The IKKs are subsequently activated by TAK1 (29). In addition, the formation of the TAK1 kinase complex leads to the activation of MKK6, which in turn activates mitogen-activated protein kinase. Finally, it triggers downstream signaling pathways including nuclear factor kappa B (NF- κ B) and mitogen-activated protein kinase to promote cytokines release (9, 26, 30). Another signal pathway has identified in epithelial cell lines. NOD1 recruits RIP2, binds to TNF receptor-associated factor 3, and activates the IFN regulatory factor 7. Induces the production of IFN- β and activates IFN-stimulated gene factor 3 for type I IFN signaling. This pathway has a mucosal protective function (31).

Independent of the NOD1/RIP2 pathway, NOD1 induces autophagy-related16-like 1 (ATG16L1) translocation to the plasma membrane in response to bacterial infection. ATG16L1 exerts a negative regulatory influence on the assembly of the NOD1/RIP2 complex (32, 33). Furthermore, ubiquitin competitively regulates the binding of NOD1 to RIP2 and ATG16L1 (34). Additionally, NOD1 interacts with caspase-1 to mediate IL-18 maturation and IL-1b secretion (35, 36). Previous research has shown that NOD1 enhances procaspase-9 activation and caspase-9-mediated apoptosis (9). However, it remains unclear whether this process is independent of NOD1/RIP2. The signaling pathway of NOD1 summarized in Figure 1.

3 NOD1, commensal microbiota, metabolism

In mammals, a great deal of microbiota coexist with host cells, and their status is closely related to host health (37). In the intestine, a barrier system consists of the intestinal epithelium and mucosa (38). This system separates the commensal bacteria and metabolites within the intestinal tract from the body's internal environment effectively reducing potential disease risks (39). However, under conditions such as high sugar intake, high-fat consumption, obesity, and inflammation, an imbalance in intestinal flora occurs along with increased intestinal permeability. Consequently, intestinal bacteria and metabolites are released into the bloodstream, causing chronic low-grade inflammation and metabolic dysfunction within target organs (40–42).

NOD1 is a protector of intestinal barrier function (43, 44). It monitors danger signals in intestinal epithelial cells and mediates the immune response to pathogen invasion (22). NOD1-deficient mice exhibit increased levels of inflammatory mediators in the intestines, altered immune characteristics (such as decreased expression of NOD2, secretory mucin, host defense peptide, and keratinocytes), and enhanced intestinal permeability (43, 45). The impact of NOD1 on the abundance of gut microbiota remains controversial. A previous study did not find evidence that NOD1 deficiency affects the microbiota in the ileum and cecum of mice (45). However, a recent study revealed an increased abundance of Parabacteroides, Rikenella, Prevotella, and Helicobacter in the intestines of NOD1-/mice (46). It suggests that while maintaining intestinal homeostasis, NOD1 may regulate the composition of intestinal flora. The discrepancy between these findings may attributed to differences in approaches used for microbiota analysis in the two studies.

The specific ligand of NOD1, iE-DAP, is derived from PGN. It has been demonstrated that the majority of circulating PGN originates from the host microbiota (47). The serum levels of



PGN in specific pathogen-free mice are approximately 0.18-0.3 μ g ml⁻¹, while in germ-free mice, they are undetectable (47). It has been established that the intestine harbors a vast array of colonizing microorganisms (48). Intestine exists an iE-DAP production pathway. The iE-DAP fragments are cleaved from PGN by lysozyme secreted by intestinal Paneth cells (49). In conclusion, iE-DAP primarily derived from host microbes, with the intestinal tract representing a significant source.

During the process of metabolism, NOD1 receptors present in target tissues recognize ligand signals from the host microbiota, promoting tissue metabolism and facilitating communication between host microorganisms and metabolism. Activation of the NOD1 receptor in pancreatic β cells by intestinal-derived ligands enhance insulin vesicle transport, stimulate insulin secretion, and maintain blood glucose homeostasis (49). Similarly, NOD1 receptors located within the dense core granules of mice adrenal chromaffin cells are capable of recognizing ligands derived from the intestinal tract. They recruit Rab2a, which then mediate the storage and secretion of epinephrine and chromogranin A (50). In addition, the function of intestinal flora in reducing bone cortical thickness and promoting bone resorption (enhanced expression of RANKL and TNFa) in mice is under regulation of NOD1. The bone metabolism in NOD-1- mice remains insensitive to alterations in intestinal flora due to different feeding conditions (51). Overall, while maintaining the intestinal barrier function, NOD1 may contribute to the homeostasis of

intestinal flora. Additionally, serving as a sensor, NOD1 detects host microbial signals and is involved in metabolism.

4 Activation of NOD1 is regulated by diet

Different diets lead to differences in nutrient intake, which affects the intestinal environment and metabolism (52).

A high-fat diet (HFD) is characterized by high fat intake. It's a well-known model of abnormalities in glycolipid metabolism (53). The concentration of NOD1 ligands in the circulation showed a significant increase in mice fed an HFD (54). The activity level of NOD1 ligands increased with prolonged exposure to an HFD (41). With the up-regulation of NOD1 expression in adipose tissue, skeletal muscle, liver, and spleen, the expression of inflammatory cytokines also increased (41, 55, 56). HFD may affect NOD1 expression through several factors independently or synergistically with its factors. HFD feeding gradually impairs intestinal barrier function in mice, resulting in enhanced translocation of microbial metabolites, and consequently elevated circulating levels of NOD1 ligands (41, 57). Moreover, it has been demonstrated that HFD contributes to gut microbiota dysbiosis (57-59), characterized by a decrease in Bacteroidetes and an increase in Firmicutes (52). Given the variations in subject, timing of intervention, and composition of

HFD, no study has conclusively demonstrated that an HFD directly influences NOD1 by modifying the gut microbiota. However, this dysbacteriosis associated with HFD affects the production of metabolites such as short-chain fatty acids and bile acids (52). Alterations in these metabolites may affect NOD1.

A study found that saturated fatty acid intake was negatively associated with insulin sensitivity in young healthy people with the NOD1 (Glu266Lys) Lys/Lys genotype (60). It suggests that single nucleotide polymorphisms in NOD1, in combination with dietary factors, synergistically increase the risk of metabolic abnormalities. Dietary saturated fatty acids are found in vegetable oils and animal fats. Because of their pro-inflammatory effects, excessive intake has been demonstrated to increase the risk of metabolic disorders (61). Saturated fatty acids (lauric acid) activate the NOD1/NF-KB signaling pathway in human colonic epithelial cells HCT116 to induce inflammation (62). However, NOD1-deficient macrophages are not subjected to inflammatory stimulation by saturated fatty acids (palmitic acid) (54). Both lauric acid and iE-DAP-mediated cellular inflammation were inhibited by polyunsaturated fatty acids (docosahexaenoic acid) (62). These studies support that NOD1 is one of the targets of saturated fatty acid-induced inflammation. Unsaturated fatty acids may be protective against such inflammation. In summary, the dietary structure represented by the HFD model has a notable influence on the expression of NOD1. Dietary fatty acids may be one of the key factors in the involvement of NOD1 in metabolic diseases.

Notably, a study found that, following four weeks of HFD consumption, NOD1^{-/-} mice did not exhibit abnormal glucose tolerance, unlike their wild-type (WT) counterparts (63). It suggests that NOD1 may play a crucial role in the metabolic damage induced by an HFD and that there is a complex interaction between NOD1 and the pathogenicity of an HFD.

5 How does NOD1 mediate the metabolism of various tissues

NOD1 is expressed in various tissues including the human heart, lungs, skeletal muscle, liver, kidney, pancreas, spleen, and others (9). The widespread expression of NOD1 significantly contributes to its involvement in the metabolism of multiple organs and tissues throughout the body. The role of NOD1 in mediating metabolism in tissues is summarized in Figure 2.

5.1 NOD1 and insulin

Insulin is a peptide hormone synthesized and secreted by pancreatic β cells. It is an essential mediator in energy metabolism (64). NOD1 functions as a signal receiver for microbial signals within the pancreatic islets, helping efficient glucose-stimulated insulin secretion (49). Activation of NOD1 in insulin secretory granules by ligands of intestinal microbial origin recruits RIP2 and Rab1a, which then mediates the intracellular transport of the secretory granules, driving them away from the nucleus towards the plasma membrane to complete the secretion of insulin (49). It has been established that NOD1 ligands in normaldiet WT mice are adequate to support insulin secretion. More ligand does not enhance its pro-secretory function (49).

Insulin binds to the insulin receptors in the target cell, activating tyrosine kinases within the receptor. Receptors recruit and phosphorylate tyrosine residues of the receptor substrate, activating downstream signaling and thus building a complex insulin signaling pathway (64, 65). It has been found that NF- κ B and MAPK signaling can be crosstalk with insulin signaling (65), at the same time, both signals can activated by NOD1 (4). It establishes a link between NOD1 and insulin signaling. NOD1 can inhibit IRS-1-mediated metabolism-related insulin signaling (5, 64-66). Phosphorylation of JNK in human adipocytes increased after iE-DAP intervention that stimulated phosphorylation of IRS-1 Ser307. Insulin-induced phosphorylation of IRS-1 tyrosine, Akt Ser473, and Akt Thr308 inhibited under the influence of iE-DAP. Ultimately, interfering with insulin signaling reduces insulin-induced glucose uptake in adipocytes (6). In addition to JNK, protein kinases such as ERK1/2 and cytokines such as TNF- α and IL-6 have been reported may be involved in the interference of insulin signaling by NOD1 (5, 6, 67). However, further research is needed to clarify the mechanisms of these NOD1 downstream signaling molecules that affect insulin signaling, to understand the role NOD1 plays in this process.

Insulin regulates the synthesis of glycogen, lipids, and proteins to maintain smooth blood glucose and energy metabolism homeostasis in the body (68). The interference of insulin signaling by NOD1 leads to reduced insulin sensitivity in NOD1expressing tissues, triggering specific metabolic abnormalities in the corresponding tissues.

5.2 NOD1 and adipose tissue

Adipose tissue is essential for energy metabolism, endocrine and immune functions and helps to maintain metabolic balance in the body (69).

5.2.1 Inflammation, oxidative stress, interfering with insulin signaling

NOD1 in adipocytes activates downstream NF- κ B signaling, enhances secretion of proinflammatory chemokines, and enhances inflammatory signaling in adipose tissue (66, 67). In addition, Oxidative stress enhanced by activated NOD1 also contributes to adipose inflammation (70). NOD1 activates NOX4, NOX1, and protein kinase C δ (PKC δ) in adipocytes. PKC δ promoted ROS generation by NOX1 and NOX4 while inhibiting the expression of antioxidant enzymes. It also activates the JNK pathway and NF- κ B pathway (70).

Insulin acts in adipose tissue to promote glucose uptake, enhance lipid synthesis, and inhibit lipolysis (64). However, as we explored in section 5.1, NOD1 regulation of the IRS1/Akt pathway hinders insulin signaling, inducing insulin resistance. NOD1 reduces insulin sensitivity in adipose tissue, decreases glucose uptake (6, 67), and increases lipolysis in adipose tissue (70–72). As insulin resistance progresses within adipose tissue, excess lipolysis occurs, which leads



to increased production of free fatty acids and ectopic accumulation of fat within the organs that are sensitive to insulin. Results in further exacerbation of insulin resistance (64, 73).

5.2.2 Enhance lipolysis

Lipolysis is a process in which lipase releases fatty acids from fat and generates energy (74). Abnormal lipolysis is associated with the pathogenesis of obesity, type 2 diabetes, non-alcoholic fatty liver, and other diseases (74). Insulin is an inhibitor of lipolysis (75). Insulin inhibits lipolysis by activating phosphodiesterase 3B and suppressing the cAMP and protein kinase A (PKA) (75). When insulin signaling is blocked in adipose tissue, the anti-lipolytic effect of insulin is impaired. However, the activation of NOD1 in adipocytes hinders the insulin signaling pathway, potentially enhancing lipolysis. NOD1 enhances lipolysis both in mouse white adipose tissue (71) and 3T3-L1 adipocytes (70-72). NOD1 regulates the enzymatic activity of hormone-sensitive lipase (HSL) to affect lipolysis (71, 72). HSL is an intracellular lipase widely expressed and primarily involved in adipocyte lipolysis, steroidogenesis, and spermatogenesis processes (76). NOD1 promotes phosphorylation of HSL at Ser563 by PKA to modulate its enzyme activity, and it also facilitates phosphorylation of Plin at Ser517. Which is necessary for efficient lipolysis (72). By mediating the synergistic effects of ERK, PKA, and NF-κB pathways, NOD1 potentiates the HSL-mediated lipolysis program (71).

During the lipolysis process, diacylglycerol accumulates as an intermediate product within cells leading to the activation of PKC δ and subsequent enhancement of oxidative stress levels (70). Meanwhile, PKC δ triggers cascade activation of interleukin-1 receptor-related kinase IRAK1/4, upregulating expression of proinflammatory cytokines including IL-1 β , IL-18, IL-6, TNF- α , and MCP-1 enhanced inflammation in adipocytes (77).

5.2.3 Endocrine/paracrine

Adipocytes secrete a variety of adipokines, lipids, metabolites, and Exosomal microRNAs. Thus, adipose tissue can regulate the metabolic processes of other metabolic tissues (78). In a study, human hepatocellular carcinoma cells cultured in a conditioned medium from 3T3-L1 adipocytes intervened in advance by NOD1 ligand (iE-DAP) showed lipid accumulation was enhanced (79). It was attributed to the secreted factors such as fatty acids and inflammatory mediators produced by NOD1-involved adipose inflammation and lipolysis, which enhance hepatocyte lipid metabolism. It suggests that NOD1 regulates hepatic lipid metabolism with endocrine/paracrine functions of adipose tissue (79).

5.2.4 Reduce adipocyte energy expenditure

Compared to white fat, brown adipose tissue (BAT) cells are characterized by abundant mitochondria and iron content, as well as multilocular lipid droplets that enhance energy metabolism and thermogenesis in response to cold and other stimuli (80). The classical mechanism of BAT thermogenesis involves the uncoupling of mitochondrial respiration mediated by uncoupling protein 1 (UCP-1) (80). NOD1 inhibits the formation and function of BAT, thereby affecting adipose energy expenditure (81). Activation of NOD1 in mouse embryonic mesenchymal stromal cells C3H10T1/2 and immortalized brown preadipocytes inhibited their differentiation into brown adipose tissue. This inhibition may be attributed to the mechanism where NOD1 induces activation of the NF-kB pathway while inhibiting trans-activation of Peroxisome proliferator-activated receptor gamma (PPARy) (82). Activated NOD1 suppressed the UCP-1 promoter activity in brown adipocytes, resulting in restricted expression of UCP-1 under both basal and isoproterenol treatment conditions. As a consequence, the oxygen consumption rate of brown adipocytes was reduced in both conditions (81). These findings demonstrate the inhibitory effect of NOD1 on the thermogenic function of BAT.

The impact of NOD1 on thyroid hormone homeostasis may serve as a potential mechanism through which NOD1 suppresses UCP-1 expression in mice (46). After consuming HFD, NOD1deficient mice exhibited decreased triiodothyronine levels in the liver and BAT, reduced hepatic Pnpla3 expression, and diminished UCP1 expression in the BAT. This led to improved insulin resistance but earlier onset of obesity (46). It has been demonstrated that elevated triiodothyronine levels stimulate lipolysis in animal models, resulting in a reduction in adipose tissue mass (83). The reduction of T3 leads to a deficiency of PNPLA3, an enzyme with lipase activity that is responsible for triglyceride hydrolysis; this deficiency impairs triglyceride breakdown and disrupts phospholipid remodeling (84, 85). The downregulation of triiodothyronine also suppresses the expression of UCP1 in BAT, leading to a decrease in thermogenesis and energy expenditure. This facilitates the development of obesity (86). However, the effects of physiologically expressed and overexpressed NOD1 on thyroid hormone remain unknown. Further investigation is required to elucidate the mechanism by which NOD1 affects thyroid hormone homeostasis.

5.2.5 Cell differentiation

The inflammation in the microenvironment inhibits the transcriptional activity of adipocyte differentiation regulators in adipose stem cells. Additionally, inflammation influences insulin signaling, impairing triglyceride synthesis and enhanced hydrolysis. Ultimately, the adipose differentiation in adipose stem cells is inhibited, while endothelial cell differentiation is promoted (87). It has been shown that activated NOD1 triggers activation of the NF- κ B pathway in human adipose-derived stem cells and 3T3-L1 cells, leading to the inhibition of PPAR γ and C/EBP α levels and attenuating adipocyte differentiation (88). The pro-cellular differentiation capacity of NOD1 has also been demonstrated in hematopoietic stem cells and mesenchymal stem cells (25, 89).

5.3 NOD1 and liver: inflammation, endoplasmic reticulum stress, insulin signaling

The liver is a vital organ responsible for metabolic and immune functions. Hepatic dysfunction contributes to dysregulation of glucolipid metabolism (90, 91).

Activated NOD1 increases the activation of NF- κ B and MAPK pathways in hepatocytes, promotes the release of chemokines CCL5 and CXCL1 in an NF- κ B-dependent manner, and together with cytokines such as IFN- γ mediates an increase in iNOS production and induces nitric oxide production (92). The pro-inflammatory response of NOD1 in the liver is affected by ER stress (55). ER stress facilitates the activation of NOD1 (20). Moreover, the concomitant occurrence of ER stress and NOD1 activation may synergistically exacerbate tissue inflammation.

Insulin inhibits gluconeogenesis and glycogenolysis while promoting glycogen synthesis and lipogenesis in hepatocytes (64). Similar to adipocytes, NOD1 interferes with insulin signaling in hepatocytes (42, 55, 66). NOD1 ligand reduces insulin-stimulated Akt phosphorylation in hepatocytes. Mice intervened with by NOD1 ligand have reduced insulin sensitivity and reduced phosphorylation of Forkhead-O1 in liver tissue (66). Forkhead-O1, a critical transcription factor for hepatic metabolism, promotes gluconeogenesis and affects intrahepatic lipid metabolism (93, 94). As a downstream target of the PI3K/Akt insulin signaling pathway, Forkhead-O1 phosphorylation is enhanced by Akt, leading to a reduction in its transcriptional activity, which in turn reduces the expression of genes associated with Forkhead-O1-induced gluconeogenesis (93, 94). It again demonstrates that NOD1 interferes with insulin signaling, reduces insulin sensitivity in peripheral tissues, and may decrease the inhibitory effect of insulin on gluconeogenesis.

In addition, NOD1-mediated inflammation and lipolysis products enhance hepatocyte lipid metabolism by enhancing cellular fatty acid uptake, mediating the expression of markers on the triglyceride synthesis pathway and output pathway, enhancing hepatocyte inflammation, and hindering insulin signaling (79). It suggests a remote regulatory role of NOD1 from adipocytes on hepatocytes.

5.4 NOD1 and cardiovascular system

The inflammatory mechanism of NOD1 hinders insulin signaling in the tissues, resulting in inefficient glucose utilization, being retained in the circulation and elevating blood glucose (67, 95). In response to abnormal blood glucose levels, the pancreas compensates by secreting more insulin, creating hyperinsulinemia (96). NOD1 promotes lipolysis, releasing inflammatory mediators and free fatty acids into circulation and enhances hepatic lipid metabolism (79). Finally, hyperglycemia (8) and high levels of insulin (97) are all involved in inducing activation of NOD1 in the cardiovascular system and triggering cardiovascular dysfunction (98).

5.4.1 Inflammation

NOD1 is a key mediator of vascular inflammation at multiple sites throughout the body, with the cardiac macrovascular inflammatory response being the most intense (99). Activation of NOD1 induces multiple vascular inflammations throughout the body in mice, characterized by infiltration of neutrophils and macrophages mainly in the aortic root (99). This finding indicated the cardiovascular predisposition associated with NOD1-dependent vascular inflammation and the capacity of NOD1 to mobilize leukocyte subsets. The NOD1/RIP2/NF-κB signaling cascade in endothelial cells (ECs) upregulates VCAM-1 expression and promotes aggregation of monocytes and neutrophils in large vessels (100). Under HFD conditions, NOD1-deficient mice exhibit elevated serum levels of CCL2, CXCL1, and CXCL2, leading to the accumulation of immune cells in the circulation but reduced aggregation at the site of atherosclerotic lesions (56). These studies demonstrate that the recruitment of immune cells by NOD1 is closely related to its function in regulating chemokine expression. In addition, ECs (101-104), Vascular Smooth Muscle Cells (VSMCs) (97, 105), and cardiomyocytes (8) were all affected by NOD1mediated inflammation. It can be reasonably assumed that persistent inflammation represents the primary mechanism through which NOD1 contributes to cardiovascular dysfunction.

5.4.2 Cell transdifferentiation

Injury induces cellular reprogramming in tissues, causing cells with the ability to remodel to change their cellular identity known as cell trans-differentiation (106, 107). Endothelial-to-mesenchymal transition (EndMT) is a typical pattern of transdifferentiation characterized by decreased endothelial properties and enhanced mesenchymal properties (108). EndMT can be triggered by inflammation-related cytokines such as TGFB, IL-33, IL-1B, TNF- α , etc., and mediated reprogramming by enforced transcription factors like Spi1 (109, 110). In vitro experiments have shown that NOD1 induces EndMT by activating the Akt/NF-κB pathway to create persistent inflammation in Human umbilical vein endothelial cells (111). This demonstrates that the inflammatory pathway downstream of NOD1 is involved in EndMT. Apart from inflammatory pathways, metabolic abnormalities such as hyperglycemia and oxidized OxLDL also activate EndMT (109). Interestingly, both activators of EndMT equally activate NOD1 (8, 100, 105), linking metabolic abnormalities, NOD1 activation, and cell trans-differentiation. Essentially, EndMT is a self-healing mechanism. However, in pathological states, it promotes endothelial dysfunction and contributes to pathological changes in cardiovascular diseases such as atherosclerosis, cardiac fibrosis, and pulmonary hypertension (108).

6 NOD1 and metabolic disease

6.1 Insulin resistance

Insulin resistance is a decrease in the sensitivity of insulin target tissues to insulin and the inability of normal levels of insulin to mediate appropriate levels of glucose-lowering action. Insulin resistance contributes to diseases such as obesity, type 2 diabetes, and metabolic syndrome (64, 68).

NOD1 inhibits the IRS and interferes with the PI3K/Akt pathway, which is the pathway by which insulin regulates metabolism (5, 6, 66, 67). This results in weakened insulin signaling in insulin-sensitive tissues. NOD1 mediates reduced glucose uptake and enhanced lipolysis in adipocytes, and adipose-secreting factors subjected to NOD1 enhance lipid deposition in hepatocytes (6, 79). NOD1 attenuates insulin-mediated phosphorylation of Forkhead-O1 in hepatocytes and may attenuate the inhibition of gluconeogenesis (66).

In addition, NOD1 expression in immune cells may contribute to insulin resistance. A clinical trial found that NOD1 and NOD2 mRNA expression in peripheral blood mononuclear cells from people with type 2 diabetes was positively correlated with insulin resistance and glycemic abnormalities (112). Moreover, NOD1 deficiency in immune cells protected mice from HFD-induced impairment of glycemic homeostasis. This may be attributed to reduced pro-inflammatory polarisation of macrophages in white adipose tissue and reduced recruitment of neutrophils (54).

6.2 Cardiometabolic disease

Metabolic abnormalities exacerbate cardiovascular damage and subsequently increase the risk of endpoint events in cardiovascular disease, a pathophysiological process in which metabolic risk factors and cardiovascular disease synergistically cause disease, also known as cardiometabolic syndrome or cardiometabolic disease (113, 114).

6.2.1 Atherosclerotic lesions

Atherosclerotic lesions are characterized by inflammation and the deposition of lipid and fibrous substances in the intima of blood vessels (115, 116).

NOD1/RIP2/NF-κB pathway activation mediates endothelial inflammation and endothelial dysfunction, promoting the aggregation of leukocyte subsets into large blood vessels (56, 100). NOD1 activates EndMT (111). Fibroblast-like cells transdifferentiated from endothelial cells are involved in plaque formation. These fibroblast-like cells exhibit a phenotype characterized by reduced collagen expression and upregulation of destabilizing collagen-MMP expression. It suggests that EndMT enhances plaque instability (115). NOD1-deficient mice had reduced leukocyte subpopulations in the aortic root intima, decreased apoptosis, proliferation of VSMCs, and increased percentage of mature fiber type I collagen in the plaque, forming a solid plaque fibrous cap. It suggests that NOD1 activation promotes plaque generation and increases the risk of plaque rupture (105). Moreover, insulin resistance induces compensatory hyperinsulinemia (96). Activation of the NOD1 receptor in VSMCs by high concentrations of insulin increases the secretion of migration regulator IL-8 and inflammatory factor IL-1 β in VSMCs (97). The migration of VSMCs to the intima enhances the accumulation of smooth muscle cells in atherosclerotic plaques (115).

The regulation of iron metabolism is likely one of the mechanisms through which NOD1 regulates atherosclerotic heart disease. Spleen, liver, and heart tissues of NOD1-deficient mice under HFD conditions exhibit significantly reduced iron content. In addition, genes related to iron metabolism show differential expression in the macrophages of these mice (117). In cases of iron deficiency, there is an increase in collagen fiber production (118), and increased macrophage glycolysis and lipid droplet. Iron overload favors M1-type differentiation (119). M1 macrophages amplify the inflammatory response at atherosclerotic sites and contribute to necrotic core formation and plaque instability (120).

6.2.2 Diabetic cardiomyopathy

Diabetes mellitus mediates myocardial metabolic disorders, damages the cardiac microvascular circulation, causes cardiomyocyte dysfunction, promotes myocardial fibrosis, impairs systolic and diastolic function, and eventually leads to the development of congestive heart failure, a condition known as diabetic cardiomyopathy (121). NOD1 is capable of promoting the development of diabetic cardiomyopathy. The NOD1/NF-κB pathway and key proteins of apoptosis are highly expressed in heart tissues of type 2 diabetic mice (db/db mice), and the expression levels of key proteins in the NOD1/NF-κB pathway are further upregulated after intervention with an activator for NOD1 (8). Meanwhile, activation of the NOD1/NF-KB pathway promotes activation of the TGF- β pathway in db/db mice cardiac fibroblasts (122). TGF- β is a cytokine involved in embryonic development and tissue repair. The activation and proliferation of fibroblasts are regulated by TGF-B, which promotes myocardial fibrosis (122). In vitro data also support that high glucose induces diabetic cardiomyopathy by stimulating the NOD1/NF-KB pathway, promoting cardiac cell apoptosis, and enhancing myocardial fibrosis (123).

6.2.3 Blood pressure

Wistar rats with NOD1 agonist intervention showed increased expression of NOS2 and elevated nitric oxide production, resulting in lower blood pressure by vasodilation. The rats also exhibited symptoms of tachycardia, impaired renal function, and stimulated coagulation. Additionally, the aorta incubated with NOD1 agonist demonstrated reduced response to vasopressin (124). *In vitro* experiments with NOD1 similarly revealed enhanced nitric oxide release from VSMCs (125).

Different from Wistar rats (124), spontaneously hypertensive rats (SHRs) exhibit endothelial dysfunction (126). As SHRs age, there is an increase in blood pressure levels, myocardial NOD1/ RIP2 expression levels, and myocardial remodeling levels (127). NOD1 inhibitor delayed vascular remodeling in SHRs (127, 128). The activation of NOD1 in the cardiovascular system promotes inflammation, induces EndMT (111), and increases angiogenesis in ECs (129). Multiple pathophysiological mechanisms jointly mediate myocardial and vascular remodeling and affect blood pressure levels (130).

7 Inhibition of NOD1-mediated metabolic damage

7.1 Interfering with NOD1 signaling

Previous studies have found that HFD and saturated fatty acids contribute to NOD1 activation (41, 62). Docosahexaenoic acid is an n-3 polyunsaturated fatty acid that can be obtained from marine fish and fish oil (131). It inhibits cellular inflammation mediated by lauric acid and iE-DAP (62). Need to verify if it has the same NOD1 inhibitory function *in vivo*.

Troxerutin, commonly found in the daily diet, is a hydroxyethylated compound of rutin, a natural flavonoid glycoside. It has been proven to have therapeutic effects on metabolic syndromes such as diabetes and cardiovascular disease (132). It has the efficacy of inhibiting ER stress, regulating the NOD1 pathway, and inhibiting HFD-enhanced hepatic gluconeogenesis (55).

Several natural monomers/extracts have been reported to ameliorate NOD1-mediated metabolic damage. Ginsenoside Rg3 (Rg3), an active ingredient in red ginseng, has been shown to inhibit intimal hyperplasia caused by inflammation. Rg3 inhibits NF- κ B nuclear translocation and suppresses Akt/NF- κ B signaling by upregulating miR-139-5p expression, contributing to the amelioration of EndMT in human umbilical vein endothelial cells induced by NOD1 (111). Moreover, purple sweet potato pigments ameliorate hepatic inflammation induced by an HFD (133), and Osthole exhibits cardioprotective effects (134). These effects are probably related to the inhibition of NOD1 and downstream pathway activation. However, further research is required to confirm the mechanism of action.

Tauroursodeoxycholic acid (TUDCA), a hydrophilic bile acid with a cytoprotective effect, has been approved for clinical use in the treatment of primary biliary cholangitis (135). It inhibits ER stress and the NOD1 pathway, improving glucose metabolism in HFD-fed mice (55).

PPAR γ is a ligand-activated transcription factor that is highly expressed in adipocytes (136). PPAR γ reduces the inhibitory effect of NOD1 on adipose tissue browning during adipocyte differentiation (82). It also regulates the expression of miR-125a, which inhibits the expression level of NOD1 and its proinflammatory and pro-angiogenic function in ECs (129). Thiazolidinediones are PPAR γ receptor agonists that increase insulin sensitivity (136). However, thiazolidinediones have not been studied to see how this affects the NOD1.

The Nuclear factor of activated T cells (NFAT) is highly expressed in monocytes of individuals diagnosed with type 2 diabetes and is positively correlated with NOD1, insulin resistance, and blood glucose levels (97). NFAT regulates NOD1 transcription in VSMCs, promotes cell proliferation, and induces phenotypic transformation of VSMCs under hyperinsulinemic conditions (97). Targeted inhibition of NFAT can regulate the NOD1 pathway and alleviate the proliferation and differentiation of VSMCs. However, NFAT consists of five subtypes, each with different structures and functions (137). It is necessary to explore the subtypes of NFAT associated with NOD1 and verify the efficacy and safety of selective inhibition.

7.2 Targeting RIP2

Another strategy is to target RIP2 and interfere with NOD/RIP2 signaling. Tyrosine kinase inhibitors (TKIs) are currently in clinical use for cancer treatment. Some TKIs can control blood glucose but the mechanism is unclear (138). TKIs targeting RIP2 may ameliorate NOD1-mediated metabolic abnormalities (138). For example, gefitinib, which has been used in the clinic, reduced NOD1 ligand-induced metabolic damage in both cellular and animal experiments (138). However, because of the inhibitory properties of TKIs on a variety of kinases, multiple potential mechanisms exist to ameliorate metabolic abnormalities (139). NOD1/RIP2 is not a critical metabolic regulatory pathway for gefitinib (139). The effect and mechanism of the inhibitory effect of gefitinib on the NOD1/RIP2 pathway may require further evaluation.

RIP2 is a downstream signal molecule common to NOD1 and NOD2 (140). Considering that the two have opposite functions on insulin sensitivity (140), direct inhibition of RIP2 may reduce the glucose-lowering effect of NOD2. Furthermore, it is not known whether direct inhibition of NOD1 has an effect on the body's immune function. Perhaps being able to target NOD1 expression in target tissues is a better strategy.

8 Risk assessment indicators for cardiometabolic disease

In a study, NOD1 has been used as one of the risk prediction classifiers for acute myocardial infarction associated with cellular pyroptosis. Due to the small sample size of this study, future validation through *in vivo*, *in vitro*, and clinical trials is needed (141).

Mechanistically, NOD1 disrupts insulin signaling and mediates insulin resistance (5, 6, 66, 67). In addition, NOD1-induced inflammation mediates EC, VSMC, and cardiomyocyte dysfunction. Insulin resistance increases the risk of cardiovascular disease (142). It has been found that people with the NOD1 (Glu266Lys) Lys/Lys genotype are affected by saturated fatty acids, increasing the risk of insulin resistance (60). We envisage that NOD1 has the potential to contribute to the assessment of the risk of cardiometabolic diseases associated with endothelial damage such as atherosclerosis. This is particularly applicable to insulinresistant populations or those with a Lys/Lys genotype with NOD1 (Glu266Lys). Considering that the mechanism by which NOD1 mediates atherosclerosis is related to its function in promoting the aggregation of leukocyte subpopulations (100), assessing NOD1 expression on leukocyte subpopulations in the peripheral circulation of subjects may be a feasible approach.

9 Summary

NOD1 is involved in endocrine and metabolic processes in several tissues throughout the body. As a sensitive intracellular sensor, NOD1 can rapidly recognize host microbial signals, sense disturbances in the intracellular environment, and transmit downstream signals. NOD1 induces oxidative stress, ER stress, EndMT, and apoptosis to respond to the challenges. Inflammatory response is the most central mechanism of NOD1. Under physiological conditions, NOD1 functions in immune defense, determines cell differentiation direction, promotes hormone secretion, and maintains internal environment stability. Under pathological conditions, NOD1 is abnormally expressed, leading to metabolic dysfunction in tissues and organs. NOD1 triggers abnormal cellular metabolism, including glycolipid metabolism, and is involved in the development and progression of insulin resistance and cardiometabolic disease. NOD1 not only increases cardiovascular disease risk factors, but also mediates damage to cardiovascular endothelium, vascular smooth muscle, and myocardium. Therefore, NOD1 has the potential to be an indicator for assessing cardiometabolic diseases. There are several issues for further study (1) The effect of NOD1 on pancreatic islet function under conditions where abnormalities of glucolipid metabolism have already occurred. (2) Whether NOD1 interferes with other insulin receptor substrates. (3) Signal transduction between NOD1 signaling downstream and insulin signaling. (4) Further studies are needed to investigate the metabolic role of NOD1 in liver and skeletal muscle in physiopathological states.

Author contributions

RT: Conceptualization, Writing – original draft. CX: Supervision, Writing – review & editing. XZ: Conceptualization, Funding acquisition, Supervision, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This study was supported by the National Natural Science Foundation of China (82104813), the Natural Science Foundation of Sichuan Provincial Science and Technology Department (2023NSFSC0689).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated

References

1. Chamaillard M, Hashimoto M, Horie Y, Masumoto J, Qiu S, Saab L, et al. An essential role for NOD1 in host recognition of bacterial peptidoglycan containing diaminopimelic acid. *Nat Immunol.* (2003) 4:702–7. doi: 10.1038/ni945

2. Ting JP, Lovering RC, Alnemri ES, Bertin J, Boss JM, Davis BK, et al. The NLR gene family: a standard nomenclature. *Immunity*. (2008) 28:285–7. doi: 10.1016/j.immuni.2008.02.005

3. Strober W, Murray PJ, Kitani A, Watanabe T. Signalling pathways and molecular interactions of NOD1 and NOD2. *Nat Rev Immunol.* (2006) 6:9–20. doi: 10.1038/nri1747

4. Caruso R, Warner N, Inohara N, Nunez G. NOD1 and NOD2: signaling, host defense, and inflammatory disease. *Immunity*. (2014) 41:898–908. doi: 10.1016/j.immuni.2014.12.010

5. Lappas M. NOD1 expression is increased in the adipose tissue of women with gestational diabetes. J Endocrinol. (2014) 222:99-112. doi: 10.1530/JOE-14-0179

 Zhou YJ, Zhou H, Li Y, Song YL. NOD1 activation induces innate immune responses and insulin resistance in human adipocytes. *Diabetes Metab.* (2012) 38:538– 43. doi: 10.1016/j.diabet.2012.08.001

7. Zhou YJ, Liu C, Li CL, Song YL, Tang YS, Zhou H, et al. Increased NOD1, but not NOD2, activity in subcutaneous adipose tissue from patients with metabolic syndrome. *Obes (Silver Spring).* (2015) 23:1394–400. doi: 10.1002/oby.21113

8. Prieto P, Vallejo-Cremades MT, Benito G, Gonzalez-Peramato P, Frances D, Agra N, et al. NOD1 receptor is up-regulated in diabetic human and murine myocardium. *Clin Sci (Lond).* (2014) 127:665–77. doi: 10.1042/CS20140180

9. Inohara N, Koseki T, del Peso L, Hu Y, Yee C, Chen S, et al. Nod1, an Apaf-1-like activator of caspase-9 and nuclear factor-kappaB. *J Biol Chem.* (1999) 274:14560–7. doi: 10.1074/jbc.274.21.14560

10. Chou WC, Jha S, Linhoff MW, Ting JP. The NLR gene family: from discovery to present day. *Nat Rev Immunol.* (2023) 23:635–54. doi: 10.1038/s41577-023-00849-x

11. Girardin SE, Boneca IG, Carneiro LA, Antignac A, Jehanno M, Viala J, et al. Nod1 detects a unique muropeptide from gram-negative bacterial peptidoglycan. *Science.* (2003) 300:1584–7. doi: 10.1126/science.1084677

12. Vegna S, Gregoire D, Moreau M, Lassus P, Durantel D, Assenat E, et al. NOD1 participates in the innate immune response triggered by hepatitis C virus polymerase. *J Virol.* (2016) 90:6022–35. doi: 10.1128/JVI.03230-15

13. Laroui H, Yan Y, Narui Y, Ingersoll SA, Ayyadurai S, Charania MA, et al. L-Alagamma-D-Glu-meso-diaminopimelic acid (DAP) interacts directly with leucine-rich region domain of nucleotide-binding oligomerization domain 1, increasing phosphorylation activity of receptor-interacting serine/threonine-protein kinase 2 and its interaction with nucleotide-binding oligomerization domain 1. *J Biol Chem.* (2011) 286:31003–13. doi: 10.1074/jbc.M111.257501

14. Ajendra J, Specht S, Ziewer S, Schiefer A, Pfarr K, Parcina M, et al. NOD2 dependent neutrophil recruitment is required for early protective immune responses against infectious Litomosoides sigmodontis L3 larvae. *Sci Rep.* (2016) 6:39648. doi: 10.1038/srep39648

15. Fan YH, Roy S, Mukhopadhyay R, Kapoor A, Duggal P, Wojcik GL, et al. Role of nucleotide-binding oligomerization domain 1 (NOD1) and its variants in human cytomegalovirus control *in vitro* and in *vivo*. *Proc Natl Acad Sci U S A*. (2016) 113: E7818–E27. doi: 10.1073/pnas.1611711113

16. Gresnigt MS, Jaeger M, Subbarao Malireddi RK, Rasid O, Jouvion G, Fitting C, et al. The absence of NOD1 enhances killing of aspergillus fumigatus through modulation of dectin-1 expression. *Front Immunol.* (2017) 8:1777. doi: 10.3389/fimmu.2017.01777

17. Zhang Y, Wu J, Xin Z, Wu X. Aspergillus fumigatus triggers innate immune response via NOD1 signaling in human corneal epithelial cells. *Exp Eye Res.* (2014) 127:170–8. doi: 10.1016/j.exer.2014.07.025

18. Silva GK, Gutierrez FR, Guedes PM, Horta CV, Cunha LD, Mineo TW, et al. Cutting edge: nucleotide-binding oligomerization domain 1-dependent responses account for murine resistance against Trypanosoma cruzi infection. *J Immunol.* (2010) 184:1148–52. doi: 10.4049/jimmunol.0902254

19. Finney CA, Lu Z, LeBourhis L, Philpott DJ, Kain KC. Disruption of Nod-like receptors alters inflammatory response to infection but does not confer protection in experimental cerebral malaria. *Am J Trop Med Hyg.* (2009) 80:718–22. doi: 10.4269/ajtmh.2009.80.718

20. Keestra-Gounder AM, Byndloss MX, Seyffert N, Young BM, Chavez-Arroyo A, Tsai AY, et al. NOD1 and NOD2 signalling links ER stress with inflammation. *Nature*. (2016) 532:394–7. doi: 10.1038/nature17631

organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

21. Molinaro R, Mukherjee T, Flick R, Philpott DJ, Girardin SE. Trace levels of peptidoglycan in serum underlie the NOD-dependent cytokine response to endoplasmic reticulum stress. *J Biol Chem.* (2019) 294:9007–15. doi: 10.1074/jbc.RA119.007997

22. Keestra AM, Baumler AJ. Detection of enteric pathogens by the nodosome. *Trends Immunol.* (2014) 35:123–30. doi: 10.1016/j.it.2013.10.009

23. Keestra AM, Winter MG, Auburger JJ, Frassle SP, Xavier MN, Winter SE, et al. Manipulation of small Rho GTPases is a pathogen-induced process detected by NOD1. *Nature*. (2013) 496:233–7. doi: 10.1038/nature12025

24. Fukazawa A, Alonso C, Kurachi K, Gupta S, Lesser CF, McCormick BA, et al. GEF-H1 mediated control of NOD1 dependent NF-kappaB activation by Shigella effectors. *PloS Pathog.* (2008) 4:e1000228. doi: 10.1371/journal.ppat.1000228

25. Cheng X, Barakat R, Pavani G, Usha MK, Calderon R, Snella E, et al. Nod1dependent NF-kB activation initiates hematopoietic stem cell specification in response to small Rho GTPases. *Nat Commun.* (2023) 14:7668. doi: 10.1038/s41467-023-43349-1

26. Inohara N, Koseki T, Lin J, del Peso L, Lucas PC, Chen FF, et al. An induced proximity model for NF-kappa B activation in the Nod1/RICK and RIP signaling pathways. *J Biol Chem.* (2000) 275:27823–31. doi: 10.1074/jbc.M003415200

27. Krieg A, Correa RG, Garrison JB, Le Negrate G, Welsh K, Huang Z, et al. XIAP mediates NOD signaling via interaction with RIP2. *Proc Natl Acad Sci U S A.* (2009) 106:14524–9. doi: 10.1073/pnas.0907131106

28. Bertrand MJ, Doiron K, Labbe K, Korneluk RG, Barker PA, Saleh M. Cellular inhibitors of apoptosis cIAP1 and cIAP2 are required for innate immunity signaling by the pattern recognition receptors NOD1 and NOD2. *Immunity*. (2009) 30:789–801. doi: 10.1016/j.immuni.2009.04.011

29. Hasegawa M, Fujimoto Y, Lucas PC, Nakano H, Fukase K, Nunez G, et al. A critical role of RICK/RIP2 polyubiquitination in Nod-induced NF-kappaB activation. *EMBO J.* (2008) 27:373–83. doi: 10.1038/sj.emboj.7601962

30. Philpott DJ, Sorbara MT, Robertson SJ, Croitoru K, Girardin SE. NOD proteins: regulators of inflammation in health and disease. *Nat Rev Immunol.* (2014) 14:9–23. doi: 10.1038/nri3565

 Watanabe T, Asano N, Fichtner-Feigl S, Gorelick PL, Tsuji Y, Matsumoto Y, et al. NOD1 contributes to mouse host defense against Helicobacter pylori via induction of type I IFN and activation of the ISGF3 signaling pathway. J Clin Invest. (2010) 120:1645–62. doi: 10.1172/JCI39481

32. Travassos LH, Carneiro LA, Ramjeet M, Hussey S, Kim YG, Magalhaes JG, et al. Nod1 and Nod2 direct autophagy by recruiting ATG16L1 to the plasma membrane at the site of bacterial entry. *Nat Immunol.* (2010) 11:55–62. doi: 10.1038/ni.1823

33. Sorbara MT, Ellison LK, Ramjeet M, Travassos LH, Jones NL, Girardin SE, et al. The protein ATG16L1 suppresses inflammatory cytokines induced by the intracellular sensors Nod1 and Nod2 in an autophagy-independent manner. *Immunity*. (2013) 39:858–73. doi: 10.1016/j.immuni.2013.10.013

34. Ver Heul AM, Fowler CA, Ramaswamy S, Piper RC. Ubiquitin regulates caspase recruitment domain-mediated signaling by nucleotide-binding oligomerization domain-containing proteins NOD1 and NOD2. J Biol Chem. (2013) 288:6890–902. doi: 10.1074/jbc.M12.413781

35. Tran LS, Ying L, D'Costa K, Wray-McCann G, Kerr G, Le L, et al. NOD1 mediates interleukin-18 processing in epithelial cells responding to Helicobacter pylori infection in mice. *Nat Commun.* (2023) 14:3804. doi: 10.1038/s41467-023-39487-1

36. Yoo NJ, Park WS, Kim SY, Reed JC, Son SG, Lee JY, et al. Nod1, a CARD protein, enhances pro-interleukin-1beta processing through the interaction with pro-caspase-1. *Biochem Biophys Res Commun.* (2002) 299:652–8. doi: 10.1016/s0006-291x(02)02714-6

37. Pickard JM, Zeng MY, Caruso R, Nunez G. Gut microbiota: Role in pathogen colonization, immune responses, and inflammatory disease. *Immunol Rev.* (2017) 279:70–89. doi: 10.1111/imr.12567

38. Lewis CV, Taylor WR. Intestinal barrier dysfunction as a therapeutic target for cardiovascular disease. *Am J Physiol Heart Circ Physiol.* (2020) 319:H1227–H33. doi: 10.1152/ajpheart.00612.2020

39. Peterson LW, Artis D. Intestinal epithelial cells: regulators of barrier function and immune homeostasis. *Nat Rev Immunol.* (2014) 14:141–53. doi: 10.1038/nri3608

40. Winer DA, Luck H, Tsai S, Winer S. The intestinal immune system in obesity and insulin resistance. Cell Metab. (2016) 23:413-26. doi: 10.1016/j.cmet.2016.01.003

41. Sharma A, Singh S, Mishra A, Rai AK, Ahmad I, Ahmad S, et al. Insulin resistance corresponds with a progressive increase in NOD1 in high fat diet-fed mice. *Endocrine*. (2022) 76:282–93. doi: 10.1007/s12020-022-02995-z

42. Schertzer JD, Klip A. Give a NOD to insulin resistance. *Am J Physiol Endocrinol Metab.* (2011) 301:E585–6. doi: 10.1152/ajpendo.00362.2011

43. Chen GY, Shaw MH, Redondo G, Nunez G. The innate immune receptor Nod1 protects the intestine from inflammation-induced tumorigenesis. *Cancer Res.* (2008) 68:10060–7. doi: 10.1158/0008-5472.CAN-08-2061

44. Zhan Y, Seregin SS, Chen J, Chen GY. Nod1 limits colitis-associated tumorigenesis by regulating IFN-gamma production. *J Immunol.* (2016) 196:5121–9. doi: 10.4049/jimmunol.1501822

45. Robertson SJ, Zhou JY, Geddes K, Rubino SJ, Cho JH, Girardin SE, et al. Nod1 and Nod2 signaling does not alter the composition of intestinal bacterial communities at homeostasis. *Gut Microbes*. (2013) 4:222–31. doi: 10.4161/gmic.24373

46. Gonzalez-Ramos S, Paz-Garcia M, Fernandez-Garcia V, Portune KJ, Acosta-Medina EF, Sanz Y, et al. NOD1 deficiency promotes an imbalance of thyroid hormones and microbiota homeostasis in mice fed high fat diet. *Sci Rep.* (2020) 10:12317. doi: 10.1038/s41598-020-69295-2

47. Huang Z, Wang J, Xu X, Wang H, Qiao Y, Chu WC, et al. Antibody neutralization of microbiota-derived circulating peptidoglycan dampens inflammation and ameliorates autoimmunity. *Nat Microbiol.* (2019) 4:766-73. doi: 10.1038/s41564-019-0381-1

48. de Vos WM, Tilg H, Van Hul M, Cani PD. Gut microbiome and health: mechanistic insights. *Gut.* (2022) 71:1020–32. doi: 10.1136/gutjnl-2021-326789

49. Zhang Q, Pan Y, Zeng B, Zheng X, Wang H, Shen X, et al. Intestinal lysozyme liberates Nod1 ligands from microbes to direct insulin trafficking in pancreatic beta cells. *Cell Res.* (2019) 29:516–32. doi: 10.1038/s41422-019-0190-3

50. Xiang C, Chen P, Zhang Q, Li Y, Pan Y, Xie W, et al. Intestinal microbiota modulates adrenomedullary response through Nod1 sensing in chromaffin cells. *iScience.* (2021) 24:102849. doi: 10.1016/j.isci.2021.102849

51. Ohlsson C, Nigro G, Boneca IG, Backhed F, Sansonetti P, Sjogren K. Regulation of bone mass by the gut microbiota is dependent on NOD1 and NOD2 signaling. *Cell Immunol.* (2017) 317:55–8. doi: 10.1016/j.cellimm.2017.05.003

52. Beam A, Clinger E, Hao L. Effect of diet and dietary components on the composition of the gut microbiota. *Nutrients.* (2021) 13:2795. doi: 10.3390/nu13082795

53. Buettner R, Scholmerich J, Bollheimer LC. High-fat diets: modeling the metabolic disorders of human obesity in rodents. *Obes (Silver Spring)*. (2007) 15:798-808. doi: 10.1038/oby.2007.608

54. Chan KL, Tam TH, Boroumand P, Prescott D, Costford SR, Escalante NK, et al. Circulating NOD1 activators and hematopoietic NOD1 contribute to metabolic inflammation and insulin resistance. *Cell Rep.* (2017) 18:2415–26. doi: 10.1016/j.celrep.2017.02.027

55. Zhang Z, Wang X, Zheng G, Shan Q, Lu J, Fan S, et al. Troxerutin attenuates enhancement of hepatic gluconeogenesis by inhibiting NOD activation-mediated inflammation in high-fat diet-treated mice.*Int J Mol Sci.* (2016) 18: 31. doi: 10.3390/ ijms18010031

56. Fernandez-Garcia V, Gonzalez-Ramos S, Avendano-Ortiz J, Martin-Sanz P, Gomez-Coronado D, Delgado C, et al. High-fat diet activates splenic NOD1 and enhances neutrophil recruitment and neutrophil extracellular traps release in the spleen of ApoE-deficient mice. *Cell Mol Life Sci.* (2022) 79:396. doi: 10.1007/s00018-022-04415-x

57. Yang J, Wei H, Zhou Y, Szeto CH, Li C, Lin Y, et al. High-fat diet promotes colorectal tumorigenesis through modulating gut microbiota and metabolites. *Gastroenterology*. (2022) 162:135–49 e2. doi: 10.1053/j.gastro.2021.08.041

58. Murphy EF, Cotter PD, Healy S, Marques TM, O'Sullivan O, Fouhy F, et al. Composition and energy harvesting capacity of the gut microbiota: relationship to diet, obesity and time in mouse models. *Gut.* (2010) 59:1635–42. doi: 10.1136/gut.2010.215665

59. Wang J, Tang H, Zhang C, Zhao Y, Derrien M, Rocher E, et al. Modulation of gut microbiota during probiotic-mediated attenuation of metabolic syndrome in high fat diet-fed mice. *ISME J.* (2015) 9:1–15. doi: 10.1038/ismej.2014.99

60. Cuda C, Badawi A, Karmali M, El-Sohemy A. Effects of polymorphisms in nucleotidebinding oligomerization domains 1 and 2 on biomarkers of the metabolic syndrome and type II diabetes. *Genes Nutr.* (2012) 7:427–35. doi: 10.1007/s12263-012-0287-5

61. Dicks LMT. How important are fatty acids in human health and can they be used in treating diseases? *Gut Microbes*. (2024) 16:2420765. doi: 10.1080/19490976.2024.2420765

62. Zhao L, Kwon MJ, Huang S, Lee JY, Fukase K, Inohara N, et al. Differential modulation of Nods signaling pathways by fatty acids in human colonic epithelial HCT116 cells. J Biol Chem. (2007) 282:11618–28. doi: 10.1074/jbc.M608644200

63. Amar J, Chabo C, Waget A, Klopp P, Vachoux C, Bermudez-Humaran LG, et al. Intestinal mucosal adherence and translocation of commensal bacteria at the early onset of type 2 diabetes: molecular mechanisms and probiotic treatment. *EMBO Mol Med.* (2011) 3:559–72. doi: 10.1002/emmm.201100159

64. Okuma H, Tsuchiya K. Tissue-specific activation of insulin signaling as a potential target for obesity-related metabolic disorders. *Pharmacol Ther.* (2024) 262:108699. doi: 10.1016/j.pharmthera.2024.108699

65. Le TKC, Dao XD, Nguyen DV, Luu DH, Bui TMH, Le TH, et al. Insulin signaling and its application. *Front Endocrinol (Lausanne)*. (2023) 14:1226655. doi: 10.3389/ fendo.2023.1226655

66. Schertzer JD, Tamrakar AK, Magalhaes JG, Pereira S, Bilan PJ, Fullerton MD, et al. NOD1 activators link innate immunity to insulin resistance. *Diabetes*. (2011) 60:2206–15. doi: 10.2337/db11-0004

67. Zhao L, Hu P, Zhou Y, Purohit J, Hwang D. NOD1 activation induces proinflammatory gene expression and insulin resistance in 3T3-L1 adipocytes. *Am J Physiol Endocrinol Metab.* (2011) 301:E587–98. doi: 10.1152/ajpendo.00709.2010

68. Petersen MC, Shulman GI. Mechanisms of insulin action and insulin resistance. *Physiol Rev.* (2018) 98:2133–223. doi: 10.1152/physrev.00063.2017

69. Sakers A, De Siqueira MK, Seale P, Villanueva CJ. Adipose-tissue plasticity in health and disease. *Cell.* (2022) 185:419–46. doi: 10.1016/j.cell.2021.12.016

70. Sharma A, Singh S, Ahmad S, Gulzar F, Schertzer JD, Tamrakar AK. NOD1 activation induces oxidative stress via NOX1/4 in adipocytes. *Free Radic Biol Med.* (2021) 162:118–28. doi: 10.1016/j.freeradbiomed.2020.11.036

71. Chi W, Dao D, Lau TC, Henriksbo BD, Cavallari JF, Foley KP, et al. Bacterial peptidoglycan stimulates adipocyte lipolysis via NOD1. *PloS One.* (2014) 9:e97675. doi: 10.1371/journal.pone.0097675

72. Purohit JS, Hu P, Chen G, Whelan J, Moustaid-Moussa N, Zhao L. Activation of nucleotide oligomerization domain containing protein 1 induces lipolysis through NF-kappaB and the lipolytic PKA activation in 3T3-L1 adipocytes. *Biochem Cell Biol.* (2013) 91:428–34. doi: 10.1139/bcb-2013-0049

73. Qiu Y, Gan M, Wang X, Liao T, Chen Q, Lei Y, et al. The global perspective on peroxisome proliferator-activated receptor gamma (PPARgamma) in ectopic fat deposition: A review. *Int J Biol Macromol.* (2023) 253:127042. doi: 10.1016/ jibiomac.2023.127042

74. Grabner GF, Xie H, Schweiger M, Zechner R. Lipolysis: cellular mechanisms for lipid mobilization from fat stores. *Nat Metab.* (2021) 3:1445–65. doi: 10.1038/s42255-021-00493-6

75. Sancar G, Liu S, Gasser E, Alvarez JG, Moutos C, Kim K, et al. FGF1 and insulin control lipolysis by convergent pathways. *Cell Metab.* (2022) 34:171-83 e6. doi: 10.1016/j.cmet.2021.12.004

76. Albert JS, Yerges-Armstrong LM, Horenstein RB, Pollin TI, Sreenivasan UT, Chai S, et al. Null mutation in hormone-sensitive lipase gene and risk of type 2 diabetes. *N Engl J Med.* (2014) 370:2307–15. doi: 10.1056/NEJMoa1315496

77. Sharma A, Maurya CK, Arha D, Rai AK, Singh S, Varshney S, et al. Nod1mediated lipolysis promotes diacylglycerol accumulation and successive inflammation via PKCdelta-IRAK axis in adipocytes. *Biochim Biophys Acta Mol Basis Dis.* (2019) 1865:136–46. doi: 10.1016/j.bbadis.2018.10.036

78. Ghesmati Z, Rashid M, Fayezi S, Gieseler F, Alizadeh E, Darabi M. An update on the secretory functions of brown, white, and beige adipose tissue: Towards therapeutic applications. *Rev Endocr Metab Disord*. (2024) 25:279–308. doi: 10.1007/s11154-023-09850-0

79. Gulzar F, Ahmad S, Singh S, Kumar P, Sharma A, Tamrakar AK. NOD1 activation in 3T3-L1 adipocytes confers lipid accumulation in HepG2 cells. *Life Sci.* (2023) 316:121400. doi: 10.1016/j.lfs.2023.121400

80. Cohen P, Kajimura S. The cellular and functional complexity of thermogenic fat. *Nat Rev Mol Cell Biol.* (2021) 22:393–409. doi: 10.1038/s41580-021-00350-0

81. Bae J, Ricciardi CJ, Esposito D, Komarnytsky S, Hu P, Curry BJ, et al. Activation of pattern recognition receptors in brown adipocytes induces inflammation and suppresses uncoupling protein 1 expression and mitochondrial respiration. *Am J Physiol Cell Physiol.* (2014) 306:C918–30. doi: 10.1152/ajpcell.00249.2013

82. Bae J, Chen J, Zhao L. Chronic activation of pattern recognition receptors suppresses brown adipogenesis of multipotent mesodermal stem cells and brown pre-adipocytes. *Biochem Cell Biol.* (2015) 93:251–61. doi: 10.1139/bcb-2014-0139

83. Samuels MH, Kolobova I, Antosik M, Niederhausen M, Purnell JQ, Schuff KG. Thyroid function variation in the normal range, energy expenditure, and body composition in L-T4-treated subjects. *J Clin Endocrinol Metab.* (2017) 102:2533–42. doi: 10.1210/jc.2017-00224

84. Calvo RM, Obregon MJ. Tri-iodothyronine upregulates adiponutrin mRNA expression in rat and human adipocytes. *Mol Cell Endocrinol.* (2009) 311:39–46. doi: 10.1016/j.mce.2009.07.006

85. Pingitore P, Romeo S. The role of PNPLA3 in health and disease. *Biochim Biophys Acta Mol Cell Biol Lipids*. (2019) 1864:900-06. doi: 10.1016/ j.bbalip.2018.06.018

86. Liu S, Shen S, Yan Y, Sun C, Lu Z, Feng H, et al. Triiodothyronine (T3) promotes brown fat hyperplasia via thyroid hormone receptor alpha mediated adipocyte progenitor cell proliferation. *Nat Commun.* (2022) 13:3394. doi: 10.1038/s41467-022-31154-1

87. Ye J, Gimble JM. Regulation of stem cell differentiation in adipose tissue by chronic inflammation. *Clin Exp Pharmacol Physiol.* (2011) 38:872–8. doi: 10.1111/j.1440-1681.2011.05596.x

88. SP J, Hu P, Burke SJ, Collier JJ, Chen J, Zhao L. The effects of NOD activation on adipocyte differentiation. *Obes (Silver Spring)*. (2013) 21:737–47. doi: 10.1002/ oby.20275

89. Kim HS, Shin TH, Yang SR, Seo MS, Kim DJ, Kang SK, et al. Implication of NOD1 and NOD2 for the differentiation of multipotent mesenchymal stem cells derived from human umbilical cord blood. *PloS One*. (2010) 5:e15369. doi: 10.1371/journal.pone.0015369

90. Trefts E, Gannon M, Wasserman DH. The liver. Curr Biol. (2017) 27:R1147-R51. doi: 10.1016/j.cub.2017.09.019

91. Okin D, Medzhitov R. The effect of sustained inflammation on hepatic mevalonate pathway results in hyperglycemia. *Cell.* (2016) 165:343–56. doi: 10.1016/j.cell.2016.02.023

92. Scott MJ, Chen C, Sun Q, Billiar TR. Hepatocytes express functional NOD1 and NOD2 receptors: a role for NOD1 in hepatocyte CC and CXC chemokine production. *J Hepatol.* (2010) 53:693–701. doi: 10.1016/j.jhep.2010.04.026

93. Benchoula K, Arya A, Parhar IS, Hwa WE. FoxO1 signaling as a therapeutic target for type 2 diabetes and obesity. *Eur J Pharmacol.* (2021) 891:173758. doi: 10.1016/j.ejphar.2020.173758

94. Yang Z, Roth K, Agarwal M, Liu W, Petriello MC. The transcription factors CREBH, PPARa, and FOXO1 as critical hepatic mediators of diet-induced metabolic dysregulation. *J Nutr Biochem.* (2021) 95:108633. doi: 10.1016/j.jnutbio.2021.108633

95. Taniguchi CM, Emanuelli B, Kahn CR. Critical nodes in signalling pathways: insights into insulin action. *Nat Rev Mol Cell Biol.* (2006) 7:85–96. doi: 10.1038/ nrm1837

96. Wang CC, Goalstone ML, Draznin B. Molecular mechanisms of insulin resistance that impact cardiovascular biology. *Diabetes.* (2004) 53:2735-40. doi: 10.2337/diabetes.53.11.2735

97. Shiny A, Regin B, Mohan V, Balasubramanyam M. Coordinated augmentation of NFAT and NOD signaling mediates proliferative VSMC phenotype switch under hyperinsulinemia. *Atherosclerosis*. (2016) 246:257-66. doi: 10.1016/j.atherosclerosis.2016.01.006

98. Fernandez-Velasco M, Prieto P, Terron V, Benito G, Flores JM, Delgado C, et al. NOD1 activation induces cardiac dysfunction and modulates cardiac fibrosis and cardiomyocyte apoptosis. *PloS One.* (2012) 7:e45260. doi: 10.1371/journal.pone.0045260

99. Nishio H, Kanno S, Onoyama S, Ikeda K, Tanaka T, Kusuhara K, et al. Nod1 ligands induce site-specific vascular inflammation. *Arterioscler Thromb Vasc Biol.* (2011) 31:1093–9. doi: 10.1161/ATVBAHA.110.216325

100. Gonzalez-Ramos S, Paz-Garcia M, Rius C, Del Monte-Monge A, Rodriguez C, Fernandez-Garcia V, et al. Endothelial NOD1 directs myeloid cell recruitment in atherosclerosis through VCAM-1. *FASEB J.* (2019) 33:3912–21. doi: 10.1096/fj.201801231RR

101. Opitz B, Forster S, Hocke AC, Maass M, Schmeck B, Hippenstiel S, et al. Nod1mediated endothelial cell activation by Chlamydophila pneumoniae. *Circ Res.* (2005) 96:319–26. doi: 10.1161/01.RES.0000155721.83594.2c

102. Cho KA, Jun YH, Suh JW, Kang JS, Choi HJ, Woo SY. Orientia tsutsugamushi induced endothelial cell activation via the NOD1-IL-32 pathway. *Microb Pathog.* (2010) 49:95–104. doi: 10.1016/j.micpath.2010.05.001

103. Opitz B, Puschel A, Beermann W, Hocke AC, Forster S, Schmeck B, et al. Listeria monocytogenes activated p38 MAPK and induced IL-8 secretion in a nucleotide-binding oligomerization domain 1-dependent manner in endothelial cells. *J Immunol.* (2006) 176:484–90. doi: 10.4049/jimmunol.176.1.484

104. Wan M, Liu J, Ouyang X. Nucleotide-binding oligomerization domain 1 regulates Porphyromonas gingivalis-induced vascular cell adhesion molecule 1 and intercellular adhesion molecule 1 expression in endothelial cells through NF-kappaB pathway. *J Periodontal Res.* (2015) 50:189–96. doi: 10.1111/jre.12192

105. Gonzalez-Ramos S, Fernandez-Garcia V, Recalde M, Rodriguez C, Martinez-Gonzalez J, Andres V, et al. Deletion or inhibition of NOD1 favors plaque stability and attenuates atherothrombosis in advanced atherogenesis (dagger). *Cells.* (2020) 9:2067. doi: 10.3390/cells9092067

106. Wang H, Yang Y, Liu J, Qian L. Direct cell reprogramming: approaches, mechanisms and progress. *Nat Rev Mol Cell Biol.* (2021) 22:410–24. doi: 10.1038/ s41580-021-00335-z

107. Merrell AJ, Stanger BZ. Adult cell plasticity *in vivo*: de-differentiation and transdifferentiation are back in style. *Nat Rev Mol Cell Biol.* (2016) 17:413–25. doi: 10.1038/nrm.2016.24

108. Kovacic JC, Dimmeler S, Harvey RP, Finkel T, Aikawa E, Krenning G, et al. Endothelial to mesenchymal transition in cardiovascular disease: JACC state-of-the-art review. J Am Coll Cardiol. (2019) 73:190–209. doi: 10.1016/j.jacc.2018.09.089

109. Cho JG, Lee A, Chang W, Lee MS, Kim J. Endothelial to mesenchymal transition represents a key link in the interaction between inflammation and endothelial dysfunction. *Front Immunol.* (2018) 9:294. doi: 10.3389/fimmu.2018.00294

110. Zhang Y, Kang Z, Liu M, Wang L, Liu F. Single-cell omics identifies inflammatory signaling as a trans-differentiation trigger in mouse embryos. *Dev Cell.* (2024) 59:961–78 e7. doi: 10.1016/j.devcel.2024.02.010

111. Lee A, Yun E, Chang W, Kim J. Ginsenoside Rg3 protects against iE-DAPinduced endothelial-to-mesenchymal transition by regulating the miR-139-5p-NFkappaB axis. J Ginseng Res. (2020) 44:300–07. doi: 10.1016/j.jgr.2019.01.003

112. Shiny A, Regin B, Balachandar V, Gokulakrishnan K, Mohan V, Babu S, et al. Convergence of innate immunity and insulin resistance as evidenced by increased nucleotide oligomerization domain (NOD) expression and signaling in monocytes from patients with type 2 diabetes. *Cytokine*. (2013) 64:564–70. doi: 10.1016/j.cyto.2013.08.003

113. Govindarajan G, Whaley-Connell A, Mugo M, Stump C, Sowers JR. The cardiometabolic syndrome as a cardiovascular risk factor. *Am J Med Sci.* (2005) 330:311–8. doi: 10.1097/00000441-200512000-00009

114. Tahir UA, Gerszten RE. Molecular biomarkers for cardiometabolic disease: risk assessment in young individuals. *Circ Res.* (2023) 132:1663–73. doi: 10.1161/CIRCRESAHA.123.322000

115. Libby P, Buring JE, Badimon L, Hansson GK, Deanfield J, Bittencourt MS, et al. Atherosclerosis. *Nat Rev Dis Primers*. (2019) 5:56. doi: 10.1038/s41572-019-0106-z

116. Cheng J, Huang H, Chen Y, Wu R. Nanomedicine for diagnosis and treatment of atherosclerosis. *Adv Sci (Weinh)*. (2023) 10:e2304294. doi: 10.1002/advs.202304294

117. Fernandez-Garcia V, Gonzalez-Ramos S, Avendano-Ortiz J, Martin-Sanz P, Delgado C, Castrillo A, et al. NOD1 splenic activation confers ferroptosis protection and reduces macrophage recruitment under pro-atherogenic conditions. *BioMed Pharmacother*. (2022) 148:112769. doi: 10.1016/j.biopha.2022.112769

118. Gardi C, Arezzini B, Fortino V, Comporti M. Effect of free iron on collagen synthesis, cell proliferation and MMP-2 expression in rat hepatic stellate cells. *Biochem Pharmacol.* (2002) 64:1139–45. doi: 10.1016/s0006-2952(02)01257-1

119. Pereira M, Chen TD, Buang N, Olona A, Ko JH, Prendecki M, et al. Acute iron deprivation reprograms human macrophage metabolism and reduces inflammation. *In Vivo Cell Rep.* (2019) 28:498–511 e5. doi: 10.1016/j.celrep.2019.06.039

120. Theofilis P, Oikonomou E, Tsioufis K, Tousoulis D. The role of macrophages in atherosclerosis: pathophysiologic mechanisms and treatment considerations. *Int J Mol Sci.* (2023) 24:9568. doi: 10.3390/ijms24119568

121. Jia G, DeMarco VG, Sowers JR. Insulin resistance and hyperinsulinaemia in diabetic cardiomyopathy. *Nat Rev Endocrinol.* (2016) 12:144–53. doi: 10.1038/ nrendo.2015.216

122. Val-Blasco A, Prieto P, Gonzalez-Ramos S, Benito G, Vallejo-Cremades MT, Pacheco I, et al. NOD1 activation in cardiac fibroblasts induces myocardial fibrosis in a murine model of type 2 diabetes. *Biochem J.* (2017) 474:399–410. doi: 10.1042/BCJ20160556

123. Wang X, Jin H, Jiang S, Xu Y. MicroRNA-495 inhibits the high glucose-induced inflammation, differentiation and extracellular matrix accumulation of cardiac fibroblasts through downregulation of NOD1. *Cell Mol Biol Lett.* (2018) 23:23. doi: 10.1186/s11658-018-0089-x

124. Cartwright N, Murch O, McMaster SK, Paul-Clark MJ, van Heel DA, Ryffel B, et al. Selective NOD1 agonists cause shock and organ injury/dysfunction. *vivo Am J Respir Crit Care Med.* (2007) 175:595–603. doi: 10.1164/rccm.200608-1103OC

125. Moreno L, McMaster SK, Gatheral T, Bailey LK, Harrington LS, Cartwright N, et al. Nucleotide oligomerization domain 1 is a dominant pathway for NOS2 induction in vascular smooth muscle cells: comparison with Toll-like receptor 4 responses in macrophages. *Br J Pharmacol.* (2010) 160:1997–2007. doi: 10.1111/j.1476-5381.2010.00814.x

126. Masodsai K, Lin YY, Lin SY, Su CT, Lee SD, Yang AL. Aging additively influences insulin- and insulin-like growth factor-1-mediated endothelial dysfunction and antioxidant deficiency in spontaneously hypertensive rats. *Biomedicines*. (2021) 9: 676. doi: 10.3390/biomedicines9060676

127. Liu FY, Fang BQ, Sun LM, Zhang XZ, Liu JL, Yang Y, et al. The role of the NOD1/rip2 signaling pathway in myocardial remodeling in spontaneously hypertensive rats. *Med Sci Monit*. (2020) 26:e924748. doi: 10.12659/MSM.924748

128. Zhang J, Fang B, Sun L, Zhang X, Liu J, Yang Y, et al. Roles of NOD1/Rip2 signal pathway in carotid artery remodelling in spontaneous hypertensive rats. *Gen Physiol Biophys.* (2022) 41:31–42. doi: 10.4149/gpb_2021042

129. Kang H, Park Y, Lee A, Seo H, Kim MJ, Choi J, et al. Negative regulation of NOD1 mediated angiogenesis by PPARgamma-regulated miR-125a. *Biochem Biophys Res Commun.* (2017) 482:28–34. doi: 10.1016/j.bbrc.2016.11.032

130. Touyz RM, Alves-Lopes R, Rios FJ, Camargo LL, Anagnostopoulou A, Arner A, et al. Vascular smooth muscle contraction in hypertension. *Cardiovasc Res.* (2018) 114:529–39. doi: 10.1093/cvr/cvy023

131. Borgonovi SM, Iametti S, Di Nunzio M. Docosahexaenoic acid as master regulator of cellular antioxidant defenses: A systematic review. *Antioxidants (Basel)*. (2023) 12:1283. doi: 10.3390/antiox12061283

132. Zamanian M, Bazmandegan G, Sureda A, Sobarzo-Sanchez E, Yousefi-Manesh H, Shirooie S. The protective roles and molecular mechanisms of troxerutin (Vitamin P4) for the treatment of chronic diseases: A mechanistic review. *Curr Neuropharmacol.* (2021) 19:97–110. doi: 10.2174/1570159X18666200510020744

133. Wang X, Zhang ZF, Zheng GH, Wang AM, Sun CH, Qin SP, et al. The inhibitory effects of purple sweet potato color on hepatic inflammation is associated with restoration of NAD(+) levels and attenuation of NLRP3 inflammasome activation in high-fat-diet-treated mice. *Molecules*. (2017) 22:1315. doi: 10.3390/molecules22081315

134. Duan J, Yang Y, Liu H, Dou PC, Tan SY. Osthole ameliorates acute myocardial infarction in rats by decreasing the expression of inflammatory-related cytokines, diminishing MMP-2 expression and activating p-ERK. *Int J Mol Med.* (2016) 37:207–16. doi: 10.3892/ijmm.2015.2402

135. Kusaczuk M. Tauroursodeoxycholate-bile acid with chaperoning activity: molecular and cellular effects and therapeutic perspectives. *Cells.* (2019) 8:1471. doi: 10.3390/cells8121471

136. Staels B, Butruille L, Francque S. Treating NASH by targeting peroxisome proliferator-activated receptors. J Hepatol. (2023) 79:1302–16. doi: 10.1016/j.jhep.2023.07.004

137. Kitamura N, Kaminuma O. Isoform-selective NFAT inhibitor: potential usefulness and development. Int J Mol Sci. (2021) 22:2725. doi: 10.3390/ijms22052725

138. Duggan BM, Foley KP, Henriksbo BD, Cavallari JF, Tamrakar AK, Schertzer JD. Tyrosine kinase inhibitors of Ripk2 attenuate bacterial cell wall-mediated lipolysis, inflammation and dysglycemia. *Sci Rep.* (2017) 7:1578. doi: 10.1038/s41598-017-01822-0

139. Duggan BM, Cavallari JF, Foley KP, Barra NG, Schertzer JD. RIPK2 dictates insulin responses to tyrosine kinase inhibitors in obese male mice. *Endocrinology*. (2020) 161:bqaa086. doi: 10.1210/endocr/bqaa086

140. Cavallari JF, Barra NG, Foley KP, Lee A, Duggan BM, Henriksbo BD, et al. Postbiotics for NOD2 require nonhematopoietic RIPK2 to improve blood glucose and metabolic inflammation in mice. *Am J Physiol Endocrinol Metab.* (2020) 318:E579–E85. doi: 10.1152/ajpendo.00033.2020

141. Guo K, Zhuo Z, Chen P, Wu H, Yang Q, Li J, et al. Construction of a pyroptosis-related classifier for risk prediction of acute myocardial infarction. *Rev Cardiovasc Med.* (2022) 23:52. doi: 10.31083/j.rcm2302052

142. Yun JS, Ko SH. Current trends in epidemiology of cardiovascular disease and cardiovascular risk management in type 2 diabetes. *Metabolism*. (2021) 123:154838. doi: 10.1016/j.metabol.2021.154838