Check for updates

OPEN ACCESS

EDITED BY Guy A. Rutter, Imperial College London, United Kingdom

REVIEWED BY Sudhanshu Kumar Bharti, Patna University, India Supriya Krishnan, Patna University, India

*CORRESPONDENCE Ziwei Jiang jiangziwei1686@gzucm.edu.cn

RECEIVED 24 April 2024 ACCEPTED 30 September 2024 PUBLISHED 14 October 2024

CITATION

Zeng W, Jiang S, Cun D, Huang F and Jiang Z (2024) Tracing links between micronutrients and type 2 diabetes risk: the singular role of selenium. Front. Endocrinol. 15:1422796. [doi: 10.3389/fendo.2024.1422796](https://doi.org/10.3389/fendo.2024.1422796)

COPYRIGHT

© 2024 Zeng, Jiang, Cun, Huang and Jiang. This is an open-access article distributed under the terms of the [Creative Commons Attribution](http://creativecommons.org/licenses/by/4.0/) [License \(CC BY\).](http://creativecommons.org/licenses/by/4.0/) The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

[Tracing links between](https://www.frontiersin.org/articles/10.3389/fendo.2024.1422796/full) [micronutrients and type 2](https://www.frontiersin.org/articles/10.3389/fendo.2024.1422796/full) [diabetes risk: the singular](https://www.frontiersin.org/articles/10.3389/fendo.2024.1422796/full) [role of selenium](https://www.frontiersin.org/articles/10.3389/fendo.2024.1422796/full)

Wenxing Zeng¹, Shan Jiang², Dejun Cun¹, Feng Huang³ and Ziwei Jiang^{1,3*}

1 First Clinical Medical College of Guangzhou University of Chinese Medicine, Guangzhou, China, ²Guangzhou University of Chinese Medicine Huizhou Hospital, Huizhou, China, ³Department of Traumatology and Orthopedics, The First Affiliated Hospital of Guangzhou University of Chinese Medicine, Guangzhou, China

Background: Type 2 diabetes (T2D) is a growing global health concern. While micronutrients are crucial for physiological functions and metabolic balance, their precise links to T2D are not fully understood.

Methods: We investigated the causal relationships between 15 key micronutrients and T2D risk using both univariate and multivariate Mendelian randomization (MR) methods. Our analysis leveraged data from a large prospective cohort genome-wide association study (GWAS) on these micronutrients and T2D. We employed MR techniques such as inverse variance weighting (IVW), MR Egger, weighted median, and simple models. Multivariate analysis adjusted for diabetes-related factors like body mass index (BMI) and hypertension to assess the independent effects of micronutrients, particularly selenium, on T2D risk.

Results: Selenium intake was associated with an increased risk of T2D, with an odds ratio (OR) of 1.045, a 95% confidence interval (CI) ranging from 1.009 to 1.082, and a P-value of 0.015. This association was consistent in multivariate analyses, suggesting an independent effect of selenium on T2D risk after adjusting for confounders.

Conclusion: Our study presents novel evidence of a positive correlation between selenium intake and T2D risk, underscoring the importance of micronutrients in diabetes prevention and treatment strategies. Further research is necessary to confirm these findings and to clarify the specific biological mechanisms through which selenium influences diabetes risk.

KEYWORDS

selenium, type 2 diabetes, causality, Mendelian randomization, multivariate analysis, micronutrients

1 Introduction

T2D is a global chronic metabolic disorder characterized by insulin resistance and relative insulin deficiency ([1\)](#page-7-0). According to the latest projections from the International Diabetes Federation, approximately 425 million adults worldwide currently suffer from diabetes, with T2D comprising the majority. By 2045, this number is expected to rise to 700 million, with an estimated 4.2 million adults aged 20-79 succumbing to diabetes, accounting for 11.3% of all deaths, predominantly in low- and middle-income countries [\(2\)](#page-7-0). This increase correlates closely with rising obesity rates and lifestyle changes. Prolonged uncontrolled hyperglycemia can lead to serious complications such as cardiovascular disease, nephropathy, neuropathy, and retinopathy, imposing significant health burdens on patients and substantial economic pressures on healthcare systems [\(3](#page-7-0)).

While the genetic basis of T2D has been extensively studied, environmental factors, particularly micronutrient intake, appear equally critical in disease development. Micronutrients like calcium, magnesium, iron, and zinc play pivotal roles not only in maintaining metabolic health but also potentially in directly or indirectly regulating glucose metabolism through insulin secretion and action [\(4,](#page-7-0) [5\)](#page-7-0). For instance, magnesium, as a cofactor for over 300 enzymes, plays a crucial role in glucose metabolism and insulin signal transduction [\(4\)](#page-7-0). Despite widespread exploration of the relationship between micronutrients and T2D in epidemiological studies, existing evidence remains inconsistent due to potential confounding factors and reverse causation. For example, studies such as the Australian Longitudinal Study ([6\)](#page-7-0) and Nurses' Health Study ([7](#page-7-0)) have concluded that higher zinc intake, compared to the lowest quintile, correlates with lower T2D risk. However, in the Multi-Ethnic Study of Atherosclerosis, baseline dietary zinc or supplement use showed no association with T2D risk over a 10 year follow-up ([8](#page-8-0), [9](#page-8-0)).

MR emerges as a novel epidemiological tool to explore causal relationships within observational data [\(10\)](#page-8-0). This method utilizes genetic variants as instrumental variables to help address confounding and reverse causation issues inherent in traditional observational studies ([11](#page-8-0)). Our study aims to employ MR to comprehensively assess the potential causal relationships between 15 major micronutrients and T2D. By analyzing data from large prospective cohorts and GWAS, we will utilize MR to investigate the associations between genetic markers of these micronutrients and T2D risk. Additionally, we will employ multivariable analysis methods to control for other disease-related potential factors, thereby enhancing the accuracy of our causal inferences.

Our research not only promises new insights into the etiology of T2D but may also have profound implications for nutritional guidance, disease prevention, and public health policy formulation. By elucidating causal relationships between micronutrients and T2D, our study aims to better understand the complex pathogenesis of T2D and provide scientific foundations for future prevention and treatment strategies.

2 Materials and methods

2.1 Methods

This study utilizes MR to explore potential causal relationships between 15 micronutrients and T2D. Initially, genetic instrumental variables (SNPs) for these micronutrients are extracted from published GWAS. Subsequently, these instrumental variables are used in two-sample MR analyses to preliminarily screen for micronutrients significantly associated with T2D risk. Building on this, to control for the influence of other confounding factors on T2D, we incorporate BMI and hypertension—two closely related risk factors—into a multivariable MR analysis. This analysis aims to assess the independent association between selenium and other confounding factors with T2D risk. Additionally, MR design must meet three essential conditions ([12](#page-8-0)): (A) Genetic variants chosen as instrumental variables (IVs) should be closely associated with each of the 15 micronutrients; (B) The genetic instruments should be unrelated to the outcome of type 2 diabetes and independent of potential confounding factors; (C) Genetic variants should be specifically associated with T2D through micronutrients rather than other pathways. [Figure 1](#page-2-0) illustrates our workflow.

2.2 Data sources for GWAS

2.2.1 Data sources for 15 micronutrients

Elsworth et al. ([13\)](#page-8-0) synthesized data from MRC-IEU, providing comprehensive GWAS data on micronutrients. The study included 64,979 individuals from European cohorts, covering 9,851,867 SNPs. This dataset encompasses 12 micronutrients: calcium, carotenoids, folate, iron, magnesium, potassium, vitamins A, B12, B6, C, D, and E. Additionally, GWAS data for copper, selenium, and zinc were sourced from Evans et al. ([14](#page-8-0)), involving 2,603 individuals and 2,543,646 SNPs. All samples underwent rigorous quality control and are stored in IEU OpenGWAS ([https://gwas.mrcieu.ac.uk\)](https://gwas.mrcieu.ac.uk); specific GWAS IDs are detailed in [Supplementary Table S1](#page-7-0).

2.2.2 Outcome data and confounders data

Summary data on SNPs related to T2D are from the latest FinnGen (https://www.finngen.fi[/en/access_results](https://www.finngen.fi/en/access_results)), version R10, under finngen_R10_T2D. This dataset aggregates GWAS results from 400,197 individuals (65,085 T2D cases and 335,112 controls) ([15](#page-8-0)). T2D cases were diagnosed based on International Classification of Diseases (ICD)-8, ICD-9, and ICD-10 criteria by the Finnish Genetic Alliance. All statistical data used in our study are publicly accessible, thus ethical approval was not required. Detailed information on outcome data is provided in [Supplementary Table S2](#page-7-0).

For the multivariable MR study, BMI GWAS data encompassed 461,460 individuals of European descent, with 79,851,867 SNPs, sourced from IEU OpenGWAS under the ID ukb-b-19953. Hypertension GWAS data came from the Finnish database (finnb-I9_HYPERTENSION), including 55,955 hypertension cases and 162,837 healthy controls, totaling 16,380,466 SNPs.

2.3 Selection of instrumental variables

In this study, SNPs with initial analysis p-values below the genome-wide significance threshold $(5 \times 10^{6} - 6)$ were chosen as IVs to ensure comprehensive results and enhance IV sensitivity. Subsequently, all IVs underwent linkage disequilibrium clumping $(r^2 = 0.001;$ distance=10,000 kb) to minimize the influence of correlated SNPs. Furthermore, we utilized Phenoscanner ([http://](http://www.phenoscanner.medschl.cam.ac.uk/) [www.phenoscanner.medschl.cam.ac.uk/\)](http://www.phenoscanner.medschl.cam.ac.uk/) to screen for potential pleiotropic effects and exclude SNPs associated with outcomes (see [Supplementary Table S3](#page-7-0)). We calculated the F-statistic $\frac{R^2(N-2)}{N}$ $(1-R^2)$] for each instrument to assess its strength, where R^2 represents the proportion of variance explained by genetic instruments and N is the effective sample size of the GWAS. SNPs with an F-statistic greater than 10 were selected for subsequent MR analysis, providing reliable estimates of genetic variation [\(16](#page-8-0)).

2.4 Mendelian randomization analysis

We conducted two-sample MR analyses to investigate the causal relationships between 15 micronutrients and T2D. The methods employed include the IVW approach ([17](#page-8-0), [18](#page-8-0)), weighted median [\(19\)](#page-8-0), simple model, and MR Egger ([20](#page-8-0)). The IVW method utilizes the variance of each IV to determine its weight, thereby mitigating the impact of instruments with larger variances on the results. This method assumes all IVs are valid and independent of the outcome, providing unbiased estimates of the causal effects of exposure on the outcome, thus serving as the primary test method. MR Egger combines traditional IV regression analysis with a

method based on quantiles. It detects and corrects for potential issues with invalid instruments, such as measurement errors or correlations between IVs and confounding factors, making it a crucial tool for sensitivity analysis. The weighted median method reduces the influence of estimates with larger variances by assigning different weights based on their variances, aiming to provide a more robust causal estimate and minimize the impact of outliers or extreme values. It allows up to half of the SNPs to be invalid instruments or exhibit pleiotropy. Additionally, the weighted model and simple model serve as supplementary methods to IVW. Consistency among these five methods enhances the reliability of the results, ensuring all effect estimates (ORs) are in the same direction. Cochran's Q test [\(21\)](#page-8-0) evaluates heterogeneity among instruments. Furthermore, sensitivity analyses using MR pleiotropy residual sum and outlier (MR-PRESSO) ([22\)](#page-8-0) assess statistical significance and evidence of potential causal effects, excluding significance levels of $P < 0.05$. Leave-one-out sensitivity analysis ([23\)](#page-8-0) and MR Egger intercept can serve as supplementary indicators for sensitivity analysis, ensuring the reliability and robustness of our final results.

2.5 Multivariate Mendelian randomization analysis

As an extension of two-sample MR, multivariable MR estimates the combined causal effects of various risk factors on T2D susceptibility [\(24\)](#page-8-0) by including all exposures in the same model. To demonstrate the independent direct effect of selenium as a trace element on T2D risk, controlling for BMI and hypertension, and confirming this effect is not mediated through other exposures, we selected significantly associated SNPs and integrated them with existing exposure IVs. After excluding these duplicate SNPs, we obtained the effects of each SNP and their corresponding standard errors from exposure and outcome. The core analytical method

IVW was still employed to infer causal relationships in multivariable MR analysis.

3 Results

3.1 Selection of Ivs

[Supplementary Table S4](#page-7-0) provides detailed information on SNPs associated with the 15 micronutrients, including β -values, standard errors, effect alleles, and other relevant data. The F-statistic values for the selected SNPs range from 20.87 to 84.68, indicating robust instrumental strength and supporting the absence of weak instrumental bias in our analyses.

3.2 Causal association between 15 micronutrients and T2D mellitus

In our MR analysis, we assessed the potential causal relationships between 15 micronutrients and the risk of T2D using multiple MR methods, including IVW, MR Egger, weighted median, simple model, and weighted mode. For micronutrients such as calcium, iron, folate, and vitamin B6, the P-values exceeded 0.05, indicating no significant associations with T2D risk. Figure 2 provides comprehensive results for all 15 micronutrients. Notably, selenium exhibited a significant association, detailed in [Figure 3.](#page-4-0)

In a detailed two-sample MR analysis, selenium significantly increased T2D risk by 45% per standard deviation increase in its levels (95% CI: 1.009 to 1.082, $P = 0.015$). Results from MR Egger, WME, IVW, SM, and WM methods were as follows: MR Egger: OR = 1.016 (95% CI: 0.951-1.086, P = 0.660); WME: OR = 1.038 $(95\% \text{ CI: } 1.001 - 1.078, P = 0.046);$ IVW: OR = 1.040 (95% CI: 1.009-1.072, $P = 0.010$); SM: OR = 1.041 (95% CI: 0.987-1.099, $P = 0.202$); WM: OR = 1.032 (95% CI: 0.987-1.080, P = 0.219). All methods consistently indicated ORs above 1, supporting a positive association between selenium and T2D risk, detailed in [Figure 4.](#page-4-0) While copper showed a positive trend toward association with T2D risk, it did not reach statistical significance ($P = 0.922$). [Figure 5](#page-5-0) displays various MR plots for selenium, including scatterplot, forest plot, funnel plot, and leave-one-out method. Our pleiotropy analyses, primarily using MR-pleiotropy, indicated no significant bias ($P > 0.05$). The MR-Egger intercept, detecting horizontal pleiotropy, also supported the absence of such bias with all p-

FIGURE 2

Results of 15 micronutrients with T2D MR analysis (IVW & WME).

values above 0.05. Furthermore, MR-PRESSO analysis, assessing heterogeneity, detected no outliers, indicating robustness. Leaveone-out sensitivity analysis confirmed that no individual SNP disproportionately influenced the overall effect. Additionally, Cochran's Q test ($P = 0.36$) showed no significant heterogeneity, confirming the robustness of our findings and the consistency of complementary analyses.

3.3 Multivariate Mendelian randomization analysis

In our multivariate analyses, we rigorously adjusted for BMI and hypertension to assess the independent association of selenium with T2D risk. The positive correlation between selenium and T2D risk remained statistically significant even after controlling for these established diabetes risk factors (IVW: OR = 1.042, 95% CI: 1.013 to 1.072). This finding underscores the robustness of selenium's effect and confirms the significance of BMI and hypertension as T2D risk factors, detailed in [Figure 6](#page-6-0). Additionally, sensitivity analysis using MR-pleiotropy found a P-value of 0.510, indicating no significant pleiotropic interference with the instrumental variables selected for the three exposures. This result further strengthens the reliability of our findings, suggesting that observed associations are likely specific to selenium's effects on T2D risk and not confounded by horizontal pleiotropy.

4 Discussion

In this study, we employed MR to explore the potential causal relationship between 15 micronutrients and T2D. Our primary finding identified a significant positive association between selenium intake and the risk of T2D. This association persisted even after accounting for established risk factors such as BMI and hypertension in multivariate MR analyses. These results not only reinforce the understanding of selenium's role in T2D pathogenesis but also may influence future strategies for diabetes prevention.

In both univariate and multivariate MR analyses, we identified a significant association between selenium intake and an elevated risk of T2D, which aligns partly with findings from previous studies. For instance, Demircan et al. [\(25\)](#page-8-0) noted in their study on serum selenium and selenoprotein P levels a positive correlation with T2D among males, where a 1 mg/L increase in SELENOP corresponded to an

(OR) of 1.22 (95% CI: 1.00, 1.48), particularly pronounced in male subjects, and also noted an association with hypertension, suggesting a potentially risky role of selenium in T2D etiology. Selenium, an essential micronutrient previously studied for its potential as an anticancer agent, has been associated with higher incidence of T2D in individuals using selenium for cancer prevention. Kohler et al. [\(26\)](#page-8-0) conducted a cross-sectional analysis involving 1,727 participants from a randomized clinical trial on selenium supplementation for colorectal adenoma chemoprevention, finding that higher plasma selenium levels were significantly linked to T2D prevalence, which supports our findings. Two studies ([27,](#page-8-0) [28](#page-8-0)) utilizing NHANES data to explore the association between selenium and T2D in American participants, covering over 9,000 individuals, found that higher selenium concentrations were associated with increased incidence of T2D, with ORs of 1.57 (95% CI: 1.16, 2.13) and 7.64 (95% CI: 3.34, 17.46), respectively. Stranges et al. ([29](#page-8-0)) reported from the Olivetti Heart Study in Italy that participants in the highest tertile of baseline selenium levels had a higher proportion of diabetes cases compared to those in the lowest tertile. Zhang et al. [\(30\)](#page-8-0), comparing trace element levels among Chinese participants with and without diabetes history, found that those in the highest quartile of selenium content had an OR of 2.69 (95% CI: 1.31, 3.49) compared to others. In contrast to these findings linking selenium with T2D, studies from the Nord-Trøndelag Health Study in Norway [\(31](#page-8-0), [32](#page-8-0)) found no association. Simic et al. ([31](#page-8-0)) reported an OR of 1.13 (95% CI: 0.65, 1.96) for diabetes prevalence in participants with the highest tertile of whole blood selenium concentration compared to the lowest, while Hansen et al. ([32](#page-8-0)), focusing on early diabetes, reported an OR of 0.93 (95% CI: 0.50, 1.74). The reasons for discrepancies with Norwegian studies, where median selenium concentrations were measured within 100- 105 micrograms per liter, significantly lower than mainstream studies, remain unclear. Stranges et al. [\(33](#page-8-0)) indicated in the Nutritional Prevention of Cancer trial that increased odds of T2D were observed specifically in individuals with blood selenium levels ≥121.6 ng/mL, suggesting a potential threshold effect of selenium levels associated with increased susceptibility to T2D that warrants further investigation in future studies. Overall, these findings suggest that higher circulating selenium concentrations or selenium supplementation for chemoprevention may moderately increase the incidence of type 2 diabetes ([34](#page-8-0)).

The potential mechanistic link between selenium and T2D may be partially mediated through selenoprotein glutathione peroxidase 1 (GPx-1) [\(35](#page-8-0)). GPx-1, as the most abundant selenoprotein, primarily functions in the cytoplasm by converting hydrogen

peroxide to water, thereby preventing lipid and protein oxidation ([36](#page-8-0)–[38\)](#page-8-0). Mice with elevated GPx-1 expression exhibit a T2D-like phenotype characterized by obesity, hyperglycemia, hyperlipidemia, hyperinsulinemia, insulin resistance, increased pancreatic β -cell mass, glucose-stimulated insulin secretion, elevated plasma leptin levels, and hepatic lipogenesis [\(39\)](#page-8-0). This phenotype in GPx-1 overexpressing mice can be almost completely reversed under dietary selenium deficiency or alleviated with reduced food intake by 40% ([39](#page-8-0)). Prolonged GPx-1 activation in animal models may disrupt insulin signaling pathways ([40,](#page-8-0) [41\)](#page-8-0), and GPx-1 overexpression has been associated with obesity and insulin resistance in experimental settings ([42\)](#page-8-0), whereas its reduction tends to mitigate these effects ([43](#page-8-0)). Moreover, liver-derived selenoprotein P (SELENOP) plays a critical role in systemic selenium homeostasis. Studies indicate that increased hepatic SELENOP mRNA expression correlates with hyperglycemia and reduced glucose tolerance in humans [\(44\)](#page-8-0). Administration of purified SELENOP has been shown to impair insulin signaling, whereas anti-SELENOP antibodies enhance insulin signaling and glucose metabolism [\(45,](#page-8-0) [46](#page-8-0)). Individuals with T2D or prediabetes generally exhibit significantly higher serum SELENOP concentrations compared to those with normal glucose tolerance. Furthermore, SELENOP levels positively correlate with markers of insulin resistance, including body mass index, waist circumference, systolic blood pressure, plasma triglycerides, serum glucose, and glycated hemoglobin ([47\)](#page-8-0). Genetic studies involving targeted deletion and RNA interference-mediated knockdown of the Selenop gene in mice have demonstrated improved systemic insulin sensitivity and glucose tolerance, underscoring SELENOP's potential role in regulating insulin function ([45\)](#page-8-0). Another potential pathway involves selenium's essential function in the synthesis of deiodinase-2 (DIO2), a critical enzyme in thyroid hormone metabolism. Selenium status directly influences DIO2 activity, which in turn affects thyroid hormone levels. Thyroid dysfunction is linked to an increased risk of insulin resistance and T2D. Studies involving specific knockout of DIO2 in pituitary and adipose tissues in mice with normal thyroid function have shown improved glucose tolerance, insulin sensitivity, and reduced body fat [\(48,](#page-8-0) [49\)](#page-8-0). This suggests that DIO2 activity not only impacts thyroid hormone levels but may also directly or indirectly influence glucose metabolism and the risk of T2D [\(50](#page-8-0)).

Another proposed mechanism relates to oxidative stress, particularly under conditions of high selenite concentrations and its metabolic product, methylselenol, in the presence of persistent hyperglycemia. Such conditions significantly elevate intracellular oxidative stress ([51](#page-9-0)), leading to increased production of reactive oxygen species (ROS). If not adequately neutralized, these ROS can impair pancreatic β -cell function ([52\)](#page-9-0), which is particularly susceptible to oxidative damage. This impairment directly affects insulin secretion and contributes to diabetes progression ([53\)](#page-9-0). Selenium normally assists in neutralizing ROS through activation of the antioxidant enzyme system under physiological conditions, thereby protecting pancreatic cells from oxidative damage [\(54\)](#page-9-0). However, excessive selenium intake can exceed this protective threshold and act as a pro-oxidant, potentially increasing oxidative stress instead of mitigating it [\(55](#page-9-0)). Excess selenium may interact with cellular thiol groups, disrupting protein folding and compromising cellular function. In diabetic individuals, this effect could exacerbate existing oxidative stress conditions [\(56](#page-9-0)).

Although selenium discovery is the focus of this study, we must not overlook the potential roles of other micronutrients in diabetes. In this study, elements such as calcium, iron, and vitamin D did not exhibit significant associations with the risk of T2D. This may be due to their weaker biological effects on diabetes risk or potential modulation by other factors. For instance, calcium may primarily relate to bone health, while both excess and deficiency of iron can impact cellular metabolism and immune function. Vitamin D's role is more complex, influencing not only bone health but also immune system regulation and inflammatory responses. For example, in a cohort study ([57](#page-9-0)) of the Spanish population, elevated serum calcium levels were associated with increased risk of developing diabetes. Researchers suggested that abnormalities in calcium homeostasis may play a significant role in the development of type 2 diabetes (HR 2.87, 95% CI 1.18-6.96, $P = 0.02$). Therefore, future research should explore the intricate effects of these elements on diabetes risk across different biological backgrounds and environmental conditions.

While our study findings support a positive correlation between selenium and the risk of T2D, we must also recognize the limitations of the MR approach. Firstly, the selection of genetic instrumental variables may be influenced by population structure, especially across different ethnicities and geographical regions. Since our sample data are derived from European populations, the generalizability of our conclusions needs validation in subsequent studies. Secondly, the biological effects of selenium may be influenced by interactions with other nutrients, which may not have been fully accounted for. Additionally, the bioactivity of selenium may depend on its chemical form and bioavailability, factors that may not have been adequately considered in our analysis.

Our study results offer new insights for future research directions. Firstly, further mechanistic studies are needed to elucidate how selenium influences the onset and progression of diabetes. Secondly, clinical trials could help validate the potential effects of selenium supplementation on the prevention and treatment of diabetes, as well as determine the optimal selenium intake. Furthermore, future research should explore interactions between selenium and other micronutrients, and consider how individual genotypes affect selenium requirements and metabolism.

5 Conclusion

This study systematically evaluated potential causal relationships between 15 micronutrients and the risk of T2D using MR. Multivariate analyses adjusting for confounding factors such as BMI and hypertension confirmed significant results. Our primary finding reveals a positive correlation between selenium intake and T2D risk, consistently observed across various sensitivity analyses. This supports selenium's potential role in T2D onset and provides novel insights for future prevention and treatment strategies. These findings align with prior research linking selenium to T2D risk, while highlighting the complex nature of interpreting such associations, including selenium's biological mechanisms and interactions with other micronutrients. Although other micronutrients did not show significant associations with T2D risk in this study, our research underscores the importance of comprehensively considering micronutrients in T2D research. In summary, this study advances understanding of selenium's role in T2D development and suggests promising avenues for further research and clinical applications. These findings warrant validation in broader biological contexts.

Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material. Further inquiries can be directed to the corresponding author.

Author contributions

WZ: Writing – original draft. SJ: Conceptualization, Writing – original draft. DC: Writing – original draft, Methodology. FH: Writing – original draft, Methodology. ZJ: Writing – review & editing.

References

1. Hameed I, Masoodi SR, Mir SA, Nabi M, Ghazanfar K, Ganai BA. Type 2 diabetes mellitus: From a metabolic disorder to an inflammatory condition. World J diabetes. (2015) 6:598–612. doi: [10.4239/wjd.v6.i4.598](https://doi.org/10.4239/wjd.v6.i4.598)

3. Calcutt NA, Cooper ME, Kern TS, Schmidt AM. Therapies for hyperglycaemiainduced diabetic complications: from animal models to clinical trials. Nat Rev Drug discov. (2009) 8:417–29. doi: [10.1038/nrd2476](https://doi.org/10.1038/nrd2476)

Funding

The authors declare financial support was received for the research, authorship, and/or publication of this article. The authors disclose that they have no business or financial associations that could create potential conflicts of interest. This study was supported by the "Academic Reserve Talent Cultivation Programme for Double First-class High-level Universities" and the "National TCM Expert Workshop Construction Project" of Guangzhou University of Traditional Chinese Medicine (Huang Feng Workshop N75, 2022).

Acknowledgments

We express our sincere gratitude to the participants and researchers of the UK BioBank, IEU OpenGWAS, and Finnish databases for their valuable contributions.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: [https://www.frontiersin.org/articles/10.3389/fendo.2024.](https://www.frontiersin.org/articles/10.3389/fendo.2024.1422796/full#supplementary-material) [1422796/full#supplementary-material](https://www.frontiersin.org/articles/10.3389/fendo.2024.1422796/full#supplementary-material)

4. Pelczyńska M, Moszak M, Bogdański P. The role of magnesium in the pathogenesis of metabolic disorders. Nutrients. (2022) 14. doi: [10.3390/nu14091714](https://doi.org/10.3390/nu14091714) 5. Fields M. Nutritional factors adversely influencing the glucose/insulin system. J Am Coll Nutr. (1998) 17:317–21. doi: [10.1080/07315724.1998.10718768](https://doi.org/10.1080/07315724.1998.10718768)

6. Vashum KP, McEvoy M, Shi Z, Milton AH, Islam MR, Sibbritt D, et al. Is dietary zinc protective for type 2 diabetes? Results from the Australian longitudinal study on women's health. BMC endocrine Disord. (2013) 13:40. doi: [10.1186/1472-6823-13-40](https://doi.org/10.1186/1472-6823-13-40)

7. Sun Q, van Dam RM, Willett WC, Hu FB. Prospective study of zinc intake and risk of type 2 diabetes in women. Diabetes Care. (2009) 32:629–34. doi: [10.2337/dc08-](https://doi.org/10.2337/dc08-1913) [1913](https://doi.org/10.2337/dc08-1913)

^{2.} Saeedi P, Petersohn I, Salpea P, Malanda B, Karuranga S, Unwin N, et al. Global and regional diabetes prevalence estimates for 2019 and projections for 2030 and 2045: Results from the International Diabetes Federation Diabetes Atlas, 9(th) edition. Diabetes Res Clin practice. (2019) 157:107843. doi: [10.1016/j.diabres.2019.107843](https://doi.org/10.1016/j.diabres.2019.107843)

8. de Oliveira Otto MC, Alonso A, Lee DH, Delclos GL, Bertoni AG, Jiang R, et al. Dietary intakes of zinc and heme iron from red meat, but not from other sources, are associated with greater risk of metabolic syndrome and cardiovascular disease. J Nutr. (2012) 142:526–33. doi: [10.3945/jn.111.149781](https://doi.org/10.3945/jn.111.149781)

9. Song Y, Xu Q, Park Y, Hollenbeck A, Schatzkin A, Chen H. Multivitamins, individual vitamin and mineral supplements, and risk of diabetes among older U. S adults Diabetes Care. (2011) 34:108–14. doi: [10.2337/dc10-1260](https://doi.org/10.2337/dc10-1260)

10. Smith GD, Ebrahim S. 'Mendelian randomization': can genetic epidemiology contribute to understanding environmental determinants of disease? Int J Epidemiol. (2003) 32:1–22. doi: [10.1093/ije/dyg070](https://doi.org/10.1093/ije/dyg070)

11. Pingault JB, O'Reilly PF, Schoeler T, Ploubidis GB, Rijsdijk F, Dudbridge F. Using genetic data to strengthen causal inference in observational research. Nat Rev Genet. (2018) 19:566–80. doi: [10.1038/s41576-018-0020-3](https://doi.org/10.1038/s41576-018-0020-3)

12. Burgess S, Butterworth A, Malarstig A, Thompson SG. Use of Mendelian randomisation to assess potential benefit of clinical intervention. BMJ (Clinical Res ed). (2012) 345:e7325. doi: [10.1136/bmj.e7325](https://doi.org/10.1136/bmj.e7325)

13. Lyon MS, Andrews SJ, Elsworth B, Gaunt TR, Hemani G, Marcora E. The variant call format provides efficient and robust storage of GWAS summary statistics. Genome Biol. (2021) 22:32. doi: [10.1186/s13059-020-02248-0](https://doi.org/10.1186/s13059-020-02248-0)

14. Evans DM, Zhu G, Dy V, Heath AC, Madden PA, Kemp JP, et al. Genome-wide association study identifies loci affecting blood copper, selenium and zinc. Hum Mol Genet. (2013) 22:3998–4006. doi: [10.1093/hmg/ddt239](https://doi.org/10.1093/hmg/ddt239)

15. Kurki MI, Karjalainen J, Palta P, Sipilä TP, Kristiansson K, Donner KM, et al. FinnGen provides genetic insights from a well-phenotyped isolated population. Nature. (2023) 613:508–18. doi: [10.1038/s41586-022-05473-8](https://doi.org/10.1038/s41586-022-05473-8)

16. Daniel N, Bouras E, Tsilidis KK, Hughes DJ. Genetically predicted circulating concentrations of micronutrients and COVID-19 susceptibility and severity: A Mendelian randomization study. Front Nutr. (2022) 9:842315. doi: [10.3389/](https://doi.org/10.3389/fnut.2022.842315) [fnut.2022.842315](https://doi.org/10.3389/fnut.2022.842315)

17. Hartwig FP, Davey Smith G, Bowden J. Robust inference in summary data Mendelian randomization via the zero modal pleiotropy assumption. Int J Epidemiol. (2017) 46:1985–98. doi: [10.1093/ije/dyx102](https://doi.org/10.1093/ije/dyx102)

18. Burgess S, Small DS, Thompson SG. A review of instrumental variable estimators for Mendelian randomization. Stat Methods Med Res. (2017) 26:2333–55. doi: [10.1177/](https://doi.org/10.1177/0962280215597579) [0962280215597579](https://doi.org/10.1177/0962280215597579)

19. Bowden J, Davey Smith G, Haycock PC, Burgess S. Consistent estimation in Mendelian randomization with some invalid instruments using a weighted median estimator. Genet Epidemiol. (2016) 40:304–14. doi: [10.1002/gepi.21965](https://doi.org/10.1002/gepi.21965)

20. Bowden J, Davey Smith G, Burgess S. Mendelian randomization with invalid instruments: effect estimation and bias detection through Egger regression. Int J Epidemiol. (2015) 44:512–25. doi: [10.1093/ije/dyv080](https://doi.org/10.1093/ije/dyv080)

21. Greco MF, Minelli C, Sheehan NA, Thompson JR. Detecting pleiotropy in Mendelian randomisation studies with summary data and a continuous outcome. Stat Med. (2015) 34:2926–40. doi: [10.1002/sim.6522](https://doi.org/10.1002/sim.6522)

22. Verbanck M, Chen CY, Neale B, Do R. Detection of widespread horizontal pleiotropy in causal relationships inferred from Mendelian randomization between complex traits and diseases. Nat Genet. (2018) 50:693–8. doi: [10.1038/s41588-018-](https://doi.org/10.1038/s41588-018-0099-7) [0099-7](https://doi.org/10.1038/s41588-018-0099-7)

23. Burgess S, Bowden J, Fall T, Ingelsson E, Thompson SG. Sensitivity analyses for robust causal inference from Mendelian randomization analyses with multiple genetic variants. Epidemiol (Cambridge Mass). (2017) 28:30–42. doi: [10.1097/](https://doi.org/10.1097/ede.0000000000000559) [ede.0000000000000559](https://doi.org/10.1097/ede.0000000000000559)

24. Sanderson E, Davey Smith G, Windmeijer F, Bowden J. An examination of multivariable Mendelian randomization in the single-sample and two-sample summary data settings. Int J Epidemiol. (2019) 48:713–27. doi: [10.1093/ije/dyy262](https://doi.org/10.1093/ije/dyy262)

25. Demircan K, Hybsier S, Chillon TS, Vetter VM, Rijntjes E, Demuth I, et al. Sexspecific associations of serum selenium and selenoprotein P with type 2 diabetes mellitus and hypertension in the Berlin Aging Study II. Redox Biol. (2023) 65:102823. doi: [10.1016/j.redox.2023.102823](https://doi.org/10.1016/j.redox.2023.102823)

26. Kohler LN, Florea A, Kelley CP, Chow S, Hsu P, Batai K, et al. Higher plasma selenium concentrations are associated with increased odds of prevalent type 2 diabetes. J Nutr. (2018) 148:1333–40. doi: [10.1093/jn/nxy099](https://doi.org/10.1093/jn/nxy099)

27. Laclaustra M, Navas-Acien A, Stranges S, Ordovas JM, Guallar E. Serum selenium concentrations and diabetes in U.S. adults: National Health and Nutrition Examination Survey (NHANES) 2003-2004. Environ Health Perspect. (2009) 117:1409– 13. doi: [10.1289/ehp.0900704](https://doi.org/10.1289/ehp.0900704)

28. Laclaustra M, Stranges S, Navas-Acien A, Ordovas JM, Guallar E. Serum selenium and serum lipids in US adults: National Health and Nutrition Examination Survey (NHANES) 2003-2004. Atherosclerosis. (2010) 210:643–8. doi: [10.1016/](https://doi.org/10.1016/j.atherosclerosis.2010.01.005) [j.atherosclerosis.2010.01.005](https://doi.org/10.1016/j.atherosclerosis.2010.01.005)

29. Stranges S, Galletti F, Farinaro E, D'Elia L, Russo O, Iacone R, et al. Associations of selenium status with cardiometabolic risk factors: an 8-year follow-up analysis of the Olivetti Heart study. Atherosclerosis. (2011) 217:274–8. doi: [10.1016/j.atherosclerosis.2011.03.027](https://doi.org/10.1016/j.atherosclerosis.2011.03.027)

30. Zhang H, Yan C, Yang Z, Zhang W, Niu Y, Li X, et al. Alterations of serum trace elements in patients with type 2 diabetes. J Trace elements Med biology: Organ Soc Minerals Trace Elements (GMS). (2017) 40:91–6. doi: [10.1016/j.jtemb.2016.12.017](https://doi.org/10.1016/j.jtemb.2016.12.017)

31. SimićA, Hansen AF, Åsvold BO, Romundstad PR, Midthjell K, Syversen T, et al. Trace element status in patients with type 2 diabetes in Norway: The HUNT3 Survey. J Trace elements Med biology: Organ Soc Minerals Trace Elements (GMS). (2017) 41:91– 8. doi: [10.1016/j.jtemb.2017.03.001](https://doi.org/10.1016/j.jtemb.2017.03.001)

32. Hansen AF, SimićA, Åsvold BO, Romundstad PR, Midthjell K, Syversen T, et al. Trace elements in early phase type 2 diabetes mellitus-A population-based study. The HUNT study in Norway. J Trace elements Med biology: Organ Soc Minerals Trace Elements (GMS). (2017) 40:46–53. doi: [10.1016/j.jtemb.2016.12.008](https://doi.org/10.1016/j.jtemb.2016.12.008)

33. Stranges S, Marshall JR, Natarajan R, Donahue RP, Trevisan M, Combs GF, et al. Effects of long-term selenium supplementation on the incidence of type 2 diabetes: randomized trial. Ann Internal Med. (2007) 147:217–23. doi: [10.7326/0003-4819-147-](https://doi.org/10.7326/0003-4819-147-4-200708210-00175) [4-200708210-00175](https://doi.org/10.7326/0003-4819-147-4-200708210-00175)

34. Siddiqi SM, Sun C, Wu X, Shah I, Mehmood A. The correlation between dietary selenium intake and type 2 diabetes: A cross-sectional population-based study on North Chinese adults. BioMed Res Int. (2020) 2020:8058463. doi: [10.1155/2020/](https://doi.org/10.1155/2020/8058463) [8058463](https://doi.org/10.1155/2020/8058463)

35. Rayman MP, Stranges S. Epidemiology of selenium and type 2 diabetes: can we make sense of it? Free Radical Biol Med. (2013) 65:1557–64. doi: [10.1016/](https://doi.org/10.1016/j.freeradbiomed.2013.04.003) [j.freeradbiomed.2013.04.003](https://doi.org/10.1016/j.freeradbiomed.2013.04.003)

36. Cheng WH, Ho YS, Ross DA, Valentine BA, Combs GF, Lei XG. Cellular glutathione peroxidase knockout mice express normal levels of selenium-dependent plasma and phospholipid hydroperoxide glutathione peroxidases in various tissues. J Nutr. (1997) 127:1445–50. doi: [10.1093/jn/127.8.1445](https://doi.org/10.1093/jn/127.8.1445)

37. Cheng WH, Valentine BA, Lei XG. High levels of dietary vitamin E do not replace cellular glutathione peroxidase in protecting mice from acute oxidative stress. J Nutr. (1999) 129:1951–7. doi: [10.1093/jn/129.11.1951](https://doi.org/10.1093/jn/129.11.1951)

38. Fu Y, Cheng WH, Porres JM, Ross DA, Lei XG. Knockout of cellular glutathione peroxidase gene renders mice susceptible to diquat-induced oxidative stress. Free Radical Biol Med. (1999) 27:605–11. doi: [10.1016/s0891-5849\(99\)00104-5](https://doi.org/10.1016/s0891-5849(99)00104-5)

39. Yan X, Pepper MP, Vatamaniuk MZ, Roneker CA, Li L, Lei XG. Dietary selenium deficiency partially rescues type 2 diabetes-like phenotypes of glutathione peroxidase-1-overexpressing male mice. J Nutr. (2012) 142:1975–82. doi: [10.3945/](https://doi.org/10.3945/jn.112.164764) [jn.112.164764](https://doi.org/10.3945/jn.112.164764)

40. McClung JP, Roneker CA, Mu W, Lisk DJ, Langlais P, Liu F, et al. Development of insulin resistance and obesity in mice overexpressing cellular glutathione peroxidase. Proc Natl Acad Sci United States America. (2004) 101:8852–7. doi: [10.1073/](https://doi.org/10.1073/pnas.0308096101) [pnas.0308096101](https://doi.org/10.1073/pnas.0308096101)

41. Loh K, Deng H, Fukushima A, Cai X, Boivin B, Galic S, et al. Reactive oxygen species enhance insulin sensitivity. Cell Metab. (2009) 10:260–72. doi: [10.1016/](https://doi.org/10.1016/j.cmet.2009.08.009) [j.cmet.2009.08.009](https://doi.org/10.1016/j.cmet.2009.08.009)

42. Huang JQ, Zhou JC, Wu YY, Ren FZ, Lei XG. Role of glutathione peroxidase 1 in glucose and lipid metabolism-related diseases. Free Radical Biol Med. (2018) 127:108– 15. doi: [10.1016/j.freeradbiomed.2018.05.077](https://doi.org/10.1016/j.freeradbiomed.2018.05.077)

43. Chen S, Su X, Mi H, Dai X, Li S, Chen S, et al. Comprehensive analysis of glutathione peroxidase-1 (GPX1) expression and prognostic value in three different types of renal cell carcinoma. Trans andrology urol. (2020) 9:2737–50. doi: [10.21037/](https://doi.org/10.21037/tau-20-1398) [tau-20-1398](https://doi.org/10.21037/tau-20-1398)

44. Burk RF, Hill KE. Selenoprotein P: an extracellular protein with unique physical characteristics and a role in selenium homeostasis. Annu Rev Nutr. (2005) 25:215–35. doi: [10.1146/annurev.nutr.24.012003.132120](https://doi.org/10.1146/annurev.nutr.24.012003.132120)

45. Misu H, Takamura T, Takayama H, Hayashi H, Matsuzawa-Nagata N, Kurita S, et al. A liver-derived secretory protein, selenoprotein P, causes insulin resistance. Cell Metab. (2010) 12:483–95. doi: [10.1016/j.cmet.2010.09.015](https://doi.org/10.1016/j.cmet.2010.09.015)

46. Mita Y, Nakayama K, Inari S, Nishito Y, Yoshioka Y, Sakai N, et al. Selenoprotein P-neutralizing antibodies improve insulin secretion and glucose sensitivity in type 2 diabetes mouse models. Nat Commun. (2017) 8:1658. doi: [10.1038/s41467-017-01863-z](https://doi.org/10.1038/s41467-017-01863-z)

47. Yang SJ, Hwang SY, Choi HY, Yoo HJ, Seo JA, Kim SG, et al. Serum selenoprotein P levels in patients with type 2 diabetes and prediabetes: implications for insulin resistance, inflammation, and atherosclerosis. J Clin Endocrinol Metab. (2011) 96:E1325–9. doi: [10.1210/jc.2011-0620](https://doi.org/10.1210/jc.2011-0620)

48. Salvatore D, Harney JW, Larsen PR. Mutation of the Secys residue 266 in human type 2 selenodeiodinase alters 75Se incorporation without affecting its biochemical properties. Biochimie. (1999) 81:535–8. doi: [10.1016/s0300-9084\(99\)80106-0](https://doi.org/10.1016/s0300-9084(99)80106-0)

49. Fonseca TL, Werneck-De-Castro JP, Castillo M, Bocco BM, Fernandes GW, McAninch EA, et al. Tissue-specific inactivation of type 2 deiodinase reveals multilevel control of fatty acid oxidation by thyroid hormone in the mouse. Diabetes. (2014) 63:1594–604. doi: [10.2337/db13-1768](https://doi.org/10.2337/db13-1768)

50. Fonseca TL, Correa-Medina M, Campos MP, Wittmann G, Werneck-de-Castro JP, Arrojo e Drigo R, et al. Coordination of hypothalamic and pituitary T3 production regulates TSH expression. J Clin Invest. (2013) 123:1492–500. doi: [10.1172/jci61231](https://doi.org/10.1172/jci61231)

51. Bai J, Tan R, An Z, Xu Y. Quantitative estimation of intracellular oxidative stress in human tissues. Briefings Bioinf. (2022) 23. doi: [10.1093/bib/bbac206](https://doi.org/10.1093/bib/bbac206)

52. Wang P, Liu WC, Han C, Wang S, Bai MY, Song CP. Reactive oxygen species: Multidimensional regulators of plant adaptation to abiotic stress and development. J Integr Plant Biol. (2024) 66:330–67. doi: [10.1111/jipb.13601](https://doi.org/10.1111/jipb.13601)

53. Cuenco J, Dalmas E. Islet inflammation and β Cell dysfunction in type 2 diabetes. Handb Exp Pharmacol. (2022) 274:227-51. doi: [10.1007/164_2021_571](https://doi.org/10.1007/164_2021_571)

54. Tinggi U. Selenium: its role as antioxidant in human health. Environ Health Prev Med. (2008) 13:102–8. doi: [10.1007/s12199-007-0019-4](https://doi.org/10.1007/s12199-007-0019-4)

55. Rayman MP. Selenium intake, status, and health: a complex relationship. Hormones (Athens Greece). (2020) 19:9–14. doi: [10.1007/s42000-019-00125-5](https://doi.org/10.1007/s42000-019-00125-5)

56. Zoidis E, Seremelis I, Kontopoulos N, Danezis GP. Selenium-dependent antioxidant enzymes: actions and properties of selenoproteins. Antioxidants (Basel Switzerland). (2018) 7. doi: [10.3390/antiox7050066](https://doi.org/10.3390/antiox7050066)

57. Becerra-Tomás N, Estruch R, Bulló M, Casas R, Dı́ az-López A, Basora J, et al. Increased serum calcium levels and risk of type 2 diabetes in individuals at high cardiovascular risk. Diabetes Care. (2014) 37:3084–91. doi: [10.2337/dc14-](https://doi.org/10.2337/dc14-0898) [0898](https://doi.org/10.2337/dc14-0898)