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[Exploring the relationship](https://www.frontiersin.org/articles/10.3389/fendo.2024.1396032/full) [between air pollution, non](https://www.frontiersin.org/articles/10.3389/fendo.2024.1396032/full)[alcoholic fatty liver disease,](https://www.frontiersin.org/articles/10.3389/fendo.2024.1396032/full) [and liver function indicators: a](https://www.frontiersin.org/articles/10.3389/fendo.2024.1396032/full) [two-sample Mendelian](https://www.frontiersin.org/articles/10.3389/fendo.2024.1396032/full) [randomization analysis study](https://www.frontiersin.org/articles/10.3389/fendo.2024.1396032/full)

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Background and aims: Non-alcoholic fatty liver disease (NAFLD) is a common metabolic disorder worldwide, with an increasing incidence in recent years. While previous studies have suggested an association between the air pollutant PM2.5 and NAFLD, there is still considerable debate regarding the existence of a clear causal relationship between air pollution and NAFLD. This study aims to employ Mendelian randomization methods to evaluate the causal relationship between major air pollutants and NAFLD.

Method: We conducted Mendelian randomization analyses on a large-scale publicly available genome-wide association study (GWAS) dataset of European populations to dissect the association between air pollutants, NAFLD, and liver function indicators. We used five different analysis methods, including Inversevariance weighted (IVW), Weighted median, MR-Egger, Simple mode, and Weighted mode, to analyze the data. We also tested for pleiotropy, heterogeneity, and sensitivity of the results.

Results: This study utilized four common exposures related to air pollution and four outcomes related to NAFLD. The results regarding the association between air pollutants and NAFLD (PM2.5: P=0.808, 95% CI=0.37-3.56; PM10: P=0.238, 95% CI=0.33-1.31; nitrogen dioxide: P=0.629, 95% CI=0.40-4.61; nitrogen oxides: $P=0.123$, 95% CI=0.13-1.28) indicated no statistically significant correlation between them. However, notably, there was a causal relationship between PM10 and serum albumin (ALB) levels (P=0.019, 95% CI=1.02-1.27).

Conclusion: This MR study found no evidence of a causal relationship between air pollution and NAFLD in European populations. However, a statistically significant association was observed between PM10 and ALB levels, suggesting that the air pollutant PM10 may impact the liver's ability to synthesize proteins.

KEYWORDS

air pollution, non-alcoholic fatty liver disease, liver function indicators, PM2.5, causal relationship, Mendelian randomization study

1 Introduction

Non-alcoholic fatty liver disease (NAFLD) is a clinicalpathological syndrome characterized by hepatocellular steatosis and lipid accumulation ([1\)](#page-11-0). It includes a range of liver abnormalities, starting from simple steatosis (NAFL) to nonalcoholic steatohepatitis (NASH), with various disease progression patterns that can result in liver fibrosis, cirrhosis, and cancer ([2](#page-11-0), [3\)](#page-11-0). Epidemiological studies have shown a strong correlation between NAFLD and metabolic diseases such as obesity, diabetes, hypertension, and dyslipidemia ([4](#page-11-0)), leading many scholars in recent years to refer to it as metabolic dysfunction-associated fatty liver disease (MAFLD) to emphasize the impact of metabolism on the disease [\(5](#page-12-0)–[7](#page-12-0)).

Currently, experts estimate that NAFLD affects around 25% of the global population, and there has been a rising trend in recent years ([8](#page-12-0)). With its increasing prevalence, NAFLD has become a significant public health concern globally. Despite the high medical demand for NAFLD, no effective drugs targeting NAFLD have yet received approval from the United States Food and Drug Administration (FDA) and the National Medical Products Administration (NMPA) [\(9\)](#page-12-0), making lifestyle modifications still a recommended intervention ([10\)](#page-12-0). Therefore, it is crucial to identify factors that may influence the occurrence and progression of NAFLD and implement effective interventions to reduce the incidence of NAFLD.

Air pollutants primarily originate from human activities or natural events, including pollutants from burning fossil fuels and sources from natural disasters, mainly comprising particulate matter (PM2.5, PM10), sulfur dioxide, nitrogen dioxide, ozone, and nitrogen oxides [\(11](#page-12-0)). Prolonged exposure to air pollutants is bound to have adverse effects on human health. Evidence suggests that long-term exposure to air pollution or fine particulate matter PM2.5 can negatively impact human health, increasing the risk of cardiovascular events and diseases such as diabetes ([12](#page-12-0)–[15\)](#page-12-0). Furthermore, prolonged environmental exposure to fine particulate matter PM2.5 may be associated with an increased risk of NAFLD development ([16](#page-12-0)–[18\)](#page-12-0). However, these reports still face numerous contradictions and controversies ([19](#page-12-0)–[21](#page-12-0)), necessitating further investigation and validation. Based on this, we hypothesize: Is there a causal relationship between air pollutants and NAFLD?

Mendelian randomization (MR) is a widely used analytical method for exploring causal relationships. Based on the random allocation of genes from parents to offspring, MR uses differences in human genotypes as instrumental variables (IVs) to investigate the causal impact of exposures on outcomes ([22\)](#page-12-0). MR can minimize confounding factors to a great extent as genetic variations are randomly allocated to offspring and thus independent of environmental factors, which are typically confounders associated with exposure and outcome ([23](#page-12-0)). In conclusion, well-designed MR studies can provide more reliable evidence to guide clinical practice ([24](#page-12-0), [25\)](#page-12-0).

In this study, we utilized a large amount of publicly available GWAS data and conducted a two-sample MR analysis to elucidate the impact of air pollutants on the development of NAFLD, thereby further investigating the causal relationship between air pollution and NAFLD, providing new insights for NAFLD prevention.

2 Methods

2.1 Study design

Our design is based on the three core assumptions of MR [\(26\)](#page-12-0): assumption 1, the relevance assumption: strong associations exist between genetic variations and exposure factors; assumption 2, the independence assumption: genetic variations are independent of confounding factors that influence both exposure and outcome;

Abbreviations: NAFLD, Non-alcoholic fatty liver disease; MAFLD, Metabolic dysfunction-associated fatty liver disease; NAFL, Non-alcoholic simple fatty liver; NASH, non-alcoholic steatohepatitis; ALB, Serum albumin; FDA, Food and Drug Administration; NMPA, National Medical Products Administration; ALT, Alanine aminotransferase; AST, Aspartate aminotransferase; PM, particulate matter; NO₂, nitrogen dioxide; NOx, nitrogen oxides; IVs, instrumental variables; MR, Mendelian randomization; GWAS, genome-wide association studies; IVW, inverse variance weighted; SNPs, single nucleotide polymorphisms; CI, confidence interval; HR, hazard ratio.

assumption 3, the exclusion restriction assumption: genetic variations only affect outcomes through exposure and not through other pathways.

We utilized common air pollution indicators, namely PM2.5, PM10, nitrogen dioxide, and nitrogen oxides, as exposure factors. A diagnosis of Non-Alcoholic Fatty Liver Disease (NAFLD) was considered the outcome of Mendelian randomization analysis. Furthermore, given the primary characteristics of NAFLD are hepatic steatosis and liver dysfunction, we conducted a second Mendelian randomization analysis on alanine aminotransferase (ALT) levels, aspartate aminotransferase (AST) levels, serum albumin (ALB) levels, and liver fat percentage in relation to air pollutants to bolster the persuasiveness of our findings. The causal relationship between air pollution and NAFLD was assessed through two-sample Mendelian randomization analyses. The flowchart of the Mendelian randomization study and the fundamental hypotheses of this research are depicted in Figures 1, [2,](#page-3-0) respectively.

2.2 Data sources

The data used in this study were obtained from the Open GWAS database, as detailed in [Table 1](#page-3-0). The exposure factors of air pollutants (PM2.5, PM10, nitrogen dioxide, nitrogen oxides) were sourced from a prospective study involving over half a million participants in the UK, with phenotype and genetic details already published. We utilized European population GWAS samples for the study, including PM2.5 (GWAS ID: ukb-b-10,817), PM10 (GWAS ID: ukb-b-589), nitrogen dioxide (GWAS ID: ukb-b-2,618), and nitrogen oxides (GWAS ID: ukb-b-12,417). Additionally, NAFLD and liver indicators (ALT, AST, ALB, percent liver fat) were used as outcome measures, sourced from European populations: NAFLD (GWAS ID: ebi-a-GCST90091033), ALT (GWAS ID: ebi-a-GCST004940), AST (GWAS ID: ebi-a-GCST005064), ALB (GWAS ID: ebi-a-GCST90025992), Percent liver fat (GWAS ID: ebi-a-GCST90016673). Specific SNP information and corresponding $R²$ and F-statistics, are shown in [Supplementary Tables 1](#page-11-0)–[5](#page-11-0).

2.3 Selection of instrumental variables

We employed the following steps to select valid SNPs [\(1\)](#page-11-0): setting the genome-wide significance level at $P < 5 \times 10^{-8}$ to meet the first key assumption that these SNPs are significantly associated with the exposure ([2\)](#page-11-0). Linkage disequilibrium clustering $(r^2 < 0.001$, region size = 10,000 kb) to ensure the independence of SNPs ([3](#page-11-0)). Interpretation and strength of R^2 and F-statistic tests to eliminate low-strength SNPs (F-statistic < 10). $R^{2} = 2\times EAx(1-EAF)\times beta^{2}/(2\times EAF\times (1-EAF) \times beta^{2}) +$

 $2 \times EAF \times (1-EAF) \times SE \times N \times beta^2$, F = $R^2 \times (N-2)/(1-R^2)$ ([4\)](#page-11-0). Utilizing PhenoScanner V2 to query SNP phenotypes when necessary, excluding SNPs closely related to confounding factors to meet the second assumption of exclusivity [\(27](#page-12-0)). Initially setting the significance level at $P < 5 \times 10^{-6}$ revealed the presence of outliers and horizontal pleiotropy, with further analysis indicating no causal relationship between the two. To enhance result accuracy, we decided to uniformly set the P -value at $P <$ 5×10−⁸ , significantly reducing outliers and addressing pleiotropy, ensuring result reliability without altering statistical outcomes.

TABLE 1 Summary of the genome-wide association studies (GWAS) included in this two-sample MR study.

"NA" usually refers to "Not Available" or "Not Applicable". This indicates that a data point has no available data or is not applicable in a specific situation.

2.4 Mendelian randomization analysis

In this study, we employed five methods for data analysis, including Weighted median, MR-Egger, IVW, Simple mode, and Weighted mode. Among these, the IVW method played a predominant role. IVW is the primary method for conducting MR analysis, as it is the most commonly used and convincing MR statistical method when SNPs are valid and show no evidence of pleiotropy ([28](#page-12-0)). The IVW test selects a fixed or random effects model based on the presence of heterogeneity. The MR-Egger method allows for the intercept of the regression line to vary in the presence of pleiotropy in the IVs. It assesses the magnitude of pleiotropy between IVs using the intercept, while the slope serves as an estimate of the causal effect, providing consistent estimates even when all instrumental variables exhibit genetic pleiotropy ([29](#page-12-0)). The strength of the Weighted median method lies in its ability to consistently estimate causal relationships even with over 50% of invalid instrumental variables. Therefore, the study utilized MR-Egger regression and Weighted median as complementary methods. A significance level of $P < 0.05$ was considered statistically significant.

2.5 Sensitivity analysis

Furthermore, we conducted analyses on pleiotropy, heterogeneity, and sensitivity. Heterogeneity testing using Cochran's Q statistic, with no significant heterogeneity among the instrumental variables, led IVW to adopt fixed effects models uniformly. Outlier detection using the MR-PRESSO method revealed $P > 0.05$, indicating no outliers were detected. Horizontal pleiotropy testing with MR-Egger showed no evidence of horizontal pleiotropy, with $P > 0.05$ (Table 2). Additionally, stability assessment of MR results through leave-one-out analysis indicated that no single SNP significantly influenced the stability of the study results. Therefore, the MR results on the association between air pollutants and NAFLD and its liver indicators were deemed reliable.

2.6 Statistical analysis

All analyses were conducted using the "TwoSampleMR" and "MR-PRESSO" packages in R version 4.2.2. The statistical significance threshold for evidence was set at $P < 0.05$.

3 Results

3.1 Air pollutants and NAFLD

The results of the MR analysis are presented in [Table 3](#page-6-0), along with scatter plots [\(Figure 3](#page-7-0)), leave-one-out analysis plots [\(Figure 4\)](#page-8-0), forest plots ([Figure 5](#page-9-0)), and funnel plots [\(Figure 6\)](#page-10-0). In this study, four common air pollution-related exposures (PM2.5, PM10, nitrogen dioxide, and nitrogen oxides) were used for MR analysis with NAFLD as the outcome. The results between air pollutants and NAFLD showed no statistically significant correlation: PM2.5: P=0.808, 95% CI=0.37-3.56; PM10: P=0.238, 95%CI=0.33-1.31; nitrogen dioxide: P=0.629, 95%CI=0.40-4.61; nitrogen oxides: P=0.123, 95%CI=0.13- 1.28. The leave-one-out analysis also did not reveal any abnormal SNPs. The corresponding values of R^2 and F statistics can be found in [Supplementary Table 1.](#page-11-0)

3.2 Air pollutants and liver indicators

The MR analysis results are presented in Table 2. To further investigate the causal relationship between air pollution and NAFLD, we selected several liver indicators closely related to NAFLD (ALT, AST, ALB, percent liver fat). The IVW method results indicated no causal relationship between air pollutants and ALT (PM2.5: P=0.317; PM10: P=0.869; nitrogen dioxide: P=0.784; nitrogen oxides: P=0.081), nor between air pollutants and AST (PM2.5: P=0.150; PM10: P=0.143; nitrogen dioxide: P=0.317; nitrogen oxides: P=0.060), nor between air pollutants and ALB (PM2.5: P=0.068; nitrogen dioxide: P=0.298; nitrogen oxides: P=0.143), nor between air pollutants and percent liver fat (PM2.5: P=0.331; PM10: P=0.051; nitrogen dioxide: P=0.830; nitrogen oxides: P=0.868).

It is worth mentioning that, after controlling for heterogeneity and multiple effects, our study found a statistically significant association between PM10 and ALB (P=0.019, 95% CI=1.02- 1.27). The corresponding R^2 and F statistics values can be found in [Supplementary Tables 3](#page-11-0)–5.

4 Discussion

Previous studies have extensively investigated the causal relationship between air pollution and various diseases and related indicators using Mendelian randomization (MR) methods, such as cardiovascular diseases, diabetes, and thyroid diseases. However, existing evidence regarding the association between air pollution and NAFLD is primarily limited to cross-sectional and cohort studies, which have been subject to considerable controversy and skepticism. Therefore, it is essential to explore the specific relationship between air pollutants and NAFLD, as this understanding would play a crucial role in the early diagnosis and treatment of NAFLD. In this study, we utilized genetic data retrieved from genome-wide association studies (GWAS) databases and systematically evaluated the causal associations between major air pollutants and the onset of NAFLD. Our results revealed no causal relationship between major air pollutants (PM2.5, PM10, nitrogen dioxide, nitrogen oxides) and NAFLD.

Despite the increasing number of studies on the pathogenesis of NAFLD in recent years, the mechanisms underlying its development remain incompletely understood due to its complexity [\(30](#page-12-0)). The current pathogenesis of NAFLD is primarily explained by the "double-hit" and "multiple-hit" theories, suggesting that factors such as abnormal lipid metabolism, TABLE 2 Mendelian randomization (MR) analysis of air pollution (particulate matter, nitrogen dioxide, and nitrogen oxides, exposure) with Liver Indicators in NAFLD in the European population (IVW method).

"NA" usually refers to "Not Available" or "Not Applicable". This indicates that a data point has no available data or is not applicable in a specific situation.

oxidative stress, inflammatory stimuli, insulin resistance, mitochondrial dysfunction, and disrupted gut microbiota contribute to the occurrence of NAFLD.

Previous research has shown that air pollutants can increase fat inflammation and insulin resistance in diet-induced obese mouse models ([31](#page-12-0)), induce NASH-like phenotypes in mice, impair hepatic glucose metabolism in animal models ([32](#page-12-0), [33\)](#page-12-0), and disrupt liver glucose and lipid synthesis pathways [\(34,](#page-12-0) [35](#page-12-0)). Clinical studies have demonstrated a significant correlation between air pollution and increased diabetes prevalence in populations, particularly in young, overweight, or obese individuals ([36\)](#page-12-0). Air pollution has been identified as a risk factor for type 2 diabetes and is known to promote the development of diabetes and cardiovascular diseases, with cardiovascular diseases being the leading cause of death in NAFLD patients.

Therefore, it is crucial to pay close attention to the impact of air pollution on NAFLD. In contrast to our study, a cross-sectional study ([37\)](#page-12-0) analyzed data from 269,705 hospitalized patients diagnosed with TABLE 3 Mendelian randomization (MR) analysis of air pollution (particulate matter, nitrogen dioxide, and nitrogen oxides, exposure) with NAFLD outcome in the European population.

axis), with error bars corresponding to each standard error (SE). The slope of each line corresponds to the combined estimate using each method of the inverse variance weighted (light blue line), the MR-Egger (blue line), the simple mode (light green line), the weighted median (green line), and the weighted mode (pink line). (A) PM2.5; (B) PM10; (C) Nitrogen dioxide; (D) Nitrogen oxides.

NAFLD and estimated average annual PM2.5 exposure using a spatial exposure model to investigate the relationship between environmental PM2.5 exposure and hospitalized patients with NAFLD. The results indicated a significant association between NAFLD and PM2.5 exposure (P< 0.01, 95% CI 1.15-1.33), which was more pronounced in certain populations and regions. Additionally, a cohort study [\(38](#page-12-0)) analyzing medical examination data from 2005 to 2017 involving 17,106 hospital patients found a link between long-term environmental PM2.5 exposure and an increased risk of NAFLD, with females, lean individuals, and younger people being more susceptible to the effects of PM2.5. However, these retrospective studies are limited by recall bias and the inclusion of a limited number of cases and geographical regions, rendering their results less reliable.

Therefore, we chose the MR method to conduct gene-level causal analysis of common major air pollutants and NAFLD using single nucleotide polymorphisms (SNPs) closely associated with air pollution as instrumental variables (IVs) to enhance the accuracy and reliability of our study. Our research indicates a lack of statistically significant causal relationships between the current major air pollutants and NAFLD, reducing the likelihood of their clinical relevance and refuting the role of air pollution in the onset of NAFLD. However, in further MR analysis of air pollutants and liver indicators related to NAFLD, we found a statistical association between PM10 and ALB (Beta: 0.131, 95% CI: 1.02-1.27; $P = 0.019$).

Given the current lack of specific biomarkers for NAFLD, its clinical diagnosis primarily relies on liver tissue biopsy ([39,](#page-12-0) [40\)](#page-12-0).

However, due to the invasive nature of tissue biopsy, patients' willingness to undergo liver tissue biopsy is often low, posing challenges to the diagnosis and treatment of NAFLD. Furthermore, due to the heterogeneity of NAFLD, not all NAFLD patients exhibit obesity and high lipids; there are also many lean NAFLD patients. From the perspective of liver enzyme levels, ALT and AST are primarily present within liver cells, and their release into the blood occurs when liver cells are damaged. In patients with mild to moderate NAFLD, due to the strong compensatory capacity of liver cells, their liver enzyme levels may remain within normal

ranges, with significant liver enzyme level abnormalities typically seen in more severe cases of NASH. Therefore, it is imperative to approach these results with a rational mindset.

ALB, a crucial protein synthesized by the liver, constitutes approximately 60% of serum proteins and plays a vital role in maintaining acid-base balance, vascular permeability, colloid osmotic pressure, and combating oxidative stress ([41](#page-12-0)–[43\)](#page-12-0). Clinically, low albumin levels are often associated with chronic liver disease, malnutrition, and tumors, among others ([44\)](#page-12-0). Hypoalbuminemia (<3.5g/dL) is typically the result of hepatocyte

death and impaired albumin synthesis due to chronic liver disease. Under inflammatory conditions, ALB levels may also decrease ([45](#page-12-0), [46](#page-12-0)). Our study identified a significant association between environmental particulate matter PM10 and ALB (Beta: 0.131, 95% CI: 1.02-1.27; P=0.019). Currently, there is a lack of research on the relationship between fine particulate matter PM10 and ALB, but existing studies suggest that environmental PM10 can increase the production of pro-inflammatory cytokines, such as IL-1 β and IL-6 ([47](#page-12-0)). To some extent, this may explain our findings that

environmental PM10 might affect the synthesis of ALB in liver cells, but further in vivo and in vitro experiments are needed to validate this hypothesis.

4.1 Advantages and limitations

To the best of our knowledge, this is the first Mendelian randomization (MR) study analyzing the relationship between air

pollutants and non-alcoholic fatty liver disease (NAFLD) to elucidate the impact of air pollution on NAFLD. The main strength of this study lies in the utilization of large-scale genomewide association study (GWAS) data for MR analysis, which has increased the sample size and facilitated the identification of reliable causal relationships. The MR-Egger method reduces bias caused by reverse causation and confounding factors. The combination of the inverse-variance weighted (IVW) and MR-Egger methods enhances the reliability of this study.

However, this study also has some limitations. We only used data from individuals of European descent, and further validation is needed to determine if this relationship also exists in other populations. Additionally, although our MR study was based on results with a Pvalue of 5×10^{-8} , only a small number of instrumental variables (IVs) were identified, potentially reducing statistical power. Therefore, despite the F-statistic indicating no clear IV bias, caution should be exercised in interpreting these results. We also tested a threshold of adjusted P-value of 5×10^{-6} for SNP selection, but the results remained unchanged. Lastly, despite the removal of SNPs with confounding factors and the establishment of strict thresholds for horizontal pleiotropy, NAFLD is a broad disease category with challenging clinical diagnosis, and the biological functions of many current genetic variants remain unclear. In addition to natural environmental factors such as air pollutants, social environmental factors, including a patient's socioeconomic status, income level, and education level, exert varying degrees of influence on NAFLD. Research indicates that a lower socioeconomic status is independently associated with an increased risk of NAFLD [\(48,](#page-12-0) [49\)](#page-12-0). Thus, complete avoidance of horizontal pleiotropy may not be achievable.

5 Conclusion

In conclusion, our study indicates that major air pollutants (PM2.5, PM10, nitrogen dioxide, and nitrogen oxides) do not show a clear causal relationship with NAFLD. Furthermore, we are excited to report a statistically significant association between environmental particulate matter PM10 and ALB, but further experimental and mechanistic studies are needed to explore this relationship in depth.

Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material. Further inquiries can be directed to the corresponding authors.

Author contributions

QS: Writing – review & editing, Writing – original draft, Data curation. JP: Writing – review & editing, Writing – original draft. MP: Writing – review & editing, Methodology, Data curation. CZ: Writing – review & editing, Data curation. WF: Writing – review & editing, Data curation. JZ: Writing – review & editing, Data curation. DP: Writing – review & editing, Data curation. ZL: Writing – review & editing. HS: Writing – review & editing. YL: Writing – review & editing. QY: Writing – review & editing, Supervision, Resources, Project administration, Methodology. YZ: Writing – review & editing, Supervision, Resources, Project administration, Methodology.

References

1. Pierantonelli I, Svegliati-Baroni G. Nonalcoholic Fatty Liver Disease: Basic pathogenetic mechanisms in the progression from NAFLD to NASH[J. Transplantation. (2019) 103:e1–e13. doi: [10.1097/TP.0000000000002480](https://doi.org/10.1097/TP.0000000000002480)

2. Stefan N, Häring H, Cusi K. Non-alcoholic fatty liver disease: causes, diagnosis, cardiometabolic consequences, and treatment strategies. Lancet Diabetes Endocrinol. (2019) 7:313–24. doi: [10.1016/S2213-8587\(18\)30154-2](https://doi.org/10.1016/S2213-8587(18)30154-2)

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: [https://www.frontiersin.org/articles/10.3389/fendo.2024.1396032/](https://www.frontiersin.org/articles/10.3389/fendo.2024.1396032/full#supplementary-material) [full#supplementary-material](https://www.frontiersin.org/articles/10.3389/fendo.2024.1396032/full#supplementary-material)

3. Friedman SL, Neuschwander-Tetri BA, Rinella M, Sanyal AJ. Mechanisms of NAFLD development and therapeutic strategies. Nat Med. (2018) 24:908–22. doi: [10.1038/s41591-018-0104-9](https://doi.org/10.1038/s41591-018-0104-9)

4. Stefan N, Häring HU, Cusi K. Non-alcoholic fatty liver disease: causes, diagnosis, cardiometabolic consequences, and treatment strategies. Lancet Diabetes Endocrinol. (2019) 7:313–24. doi: [10.1016/S2213-8587\(18\)30154-2](https://doi.org/10.1016/S2213-8587(18)30154-2)

5. Gofton C, Upendran Y, Zheng M-H. MAFLD: How is it different from NAFLD? [J. Clin Mol Hepatol. (2023) 29:S17–31. doi: [10.3350/cmh.2022.0367](https://doi.org/10.3350/cmh.2022.0367)

6. Yoon EL, Jun DW. Changing the nomenclature from nonalcoholic fatty liver disease to metabolic dysfunction-associated fatty liver disease is more than a change in terminology[J. Clin Mol Hepatol. (2023) 29:371–3. doi: [10.3350/cmh.2023.0086](https://doi.org/10.3350/cmh.2023.0086)

7. Huang PL. A comprehensive definition for metabolic syndrome. Dis Models Mech. (2009) 2:231–7. doi: [10.1242/dmm.001180](https://doi.org/10.1242/dmm.001180)

8. Younossi ZM, Koenig AB, Abdelatif D. Global epidemiology of nonalcoholic fatty liver disease-Meta-analytic assessment of prevalence, incidence, and outcomes. Hepatol (Baltimore Md.). (2016) 64:73–84. doi: [10.1002/hep.28431](https://doi.org/10.1002/hep.28431)

9. Harrison SA, Allen AM, Dubourg J. Challenges and opportunities in NASH drug development. Nat Med. (2023) 29:562–73. doi: [10.1038/s41591-023-02242-6](https://doi.org/10.1038/s41591-023-02242-6)

10. Younossi Z, Tacke F, Arrese M. Global perspectives on nonalcoholic fatty liver disease and nonalcoholic steatohepatitis. Hepatology. (2019) 69:2672–82. doi: [10.1002/](https://doi.org/10.1002/hep.30251) [hep.30251](https://doi.org/10.1002/hep.30251)

11. Leung PB, Davis AM, Kumar S. Diagnosis and management of nonalcoholic fatty liver disease. JAMA. (2023) 330:1687–8. doi: [10.1001/jama.2023.17935](https://doi.org/10.1001/jama.2023.17935)

12. Rice M, Balmes J, Malhotra A. Outdoor air pollution and your health. Am J Respir Crit Care Med. (2021) 204:P13–4. doi: [10.1164/rccm.2046P13](https://doi.org/10.1164/rccm.2046P13)

13. Chen Y-C, Chin W-S, Pan S-C, et al. Long-term exposure to air pollution and the occurrence of metabolic syndrome and its components in Taiwan. Environ Health Perspect. (2023) 131:17001. doi: [10.1289/EHP10611](https://doi.org/10.1289/EHP10611)

14. Kim K-N, Ha B, Seog W, et al. Long-term exposure to air pollution and the blood lipid levels of healthy young men. Environ Int. (2022) 161:107119. doi: [10.1016/](https://doi.org/10.1016/j.envint.2022.107119) [j.envint.2022.107119](https://doi.org/10.1016/j.envint.2022.107119)

15. Yang Y, Ma X, Pang W, et al. Causal associations of PM2.5 and GDM: A twosample mendelian randomization study. Toxics Multidiscip Digital Publishing Institute. (2023) 11:171. doi: [10.3390/toxics11020171](https://doi.org/10.3390/toxics11020171)

16. Kim JM, Kim E, Song DK, et al. Causal relationship between particulate matter 2.5 and diabetes: two sample Mendelian randomization. Front Public Health. (2023) 11. doi: [10.3389/fpubh.2023.1164647](https://doi.org/10.3389/fpubh.2023.1164647)

17. Deng P, Tang H, Zhu L, et al. Association of long-term ambient fine particulate matter (PM2.5) and incident non-alcoholic fatty liver disease in Chinese adults. Environ pollut. (2023) 329:121666. doi: [10.1016/j.envpol.2023.121666](https://doi.org/10.1016/j.envpol.2023.121666)

18. Matthiessen C, Glaubitz L, Lucht S. Long-term exposure to air pollution and prevalent nonalcoholic fatty liver disease. Environ Epidemiol (Philadelphia Pa.). (2023) 7:e268. doi: [10.1097/EE9.0000000000000268](https://doi.org/10.1097/EE9.0000000000000268)

19. VoPham T, Kim NJ, Berry K. PM2.5 air pollution exposure and nonalcoholic fatty liver disease in the Nationwide Inpatient Sample. Environ Res. (2022) 213:113611. doi: [10.1016/j.envres.2022.113611](https://doi.org/10.1016/j.envres.2022.113611)

20. Guo B, Zhou J, Zhao X. Reply to: <Comment on <Exposure to air pollution is associated with an increased risk of metabolic dysfunction-associated fatty liver disease>>. J Hepatol. (2022) 77:260–2. doi: [10.1016/jjhep.2021.10.016](https://doi.org/10.1016/jjhep.2021.10.016)

21. Chen Y-S, Hung Y-M, Wei JC-C. Comment on "Exposure to air pollution is associated with an increased risk of metabolic dysfunction-associated fatty liver disease. J Hepatology Elsevier. (2022) 77:259–60. doi: [10.1016/j.jhep.2021.12.032](https://doi.org/10.1016/j.jhep.2021.12.032)

22. Guo B, Guo Y, Nima Q. Exposure to air pollution is associated with an increased risk of metabolic dysfunction-associated fatty liver disease. J Hepatol. (2022) 76:518–25. doi: [10.1016/j.jhep.2021.10.016](https://doi.org/10.1016/j.jhep.2021.10.016)

23. Davey Smith G, Hemani G. Mendelian randomization: genetic anchors for causal inference in epidemiological studies. Hum Mol Genet. (2014) 23:R89–98. doi: [10.1093/hmg/ddu328](https://doi.org/10.1093/hmg/ddu328)

24. Zuccolo L, Holmes MV. Commentary: Mendelian randomization-inspired causal inference in the absence of genetic data. Int J Epidemiol. (2017) 46:962–5. doi: [10.1093/ije/dyw327](https://doi.org/10.1093/ije/dyw327)

25. Bowden J, Davey Smith G, Burgess S. Mendelian randomization with invalid instruments: effect estimation and bias detection through Egger regression. Int J Epidemiol. (2015) 44:512–25. doi: [10.1093/ije/dyv080](https://doi.org/10.1093/ije/dyv080)

26. Hartwig FP, Davies NM, Hemani G. Two-sample Mendelian randomization: avoiding the downsides of a powerful, widely applicable but potentially fallible technique. Int J Epidemiol. (2016) 45:1717–26. doi: [10.1093/ije/dyx028](https://doi.org/10.1093/ije/dyx028)

27. Zhang X, Chen K, Yin S. Association of leisure sedentary behavior and physical activity with the risk of nonalcoholic fatty liver disease: a two-sample Mendelian
randomization study. *Front Nutr*. (2023) 10. doi: [10.3389/fnut.2023.1158810](https://doi.org/10.3389/fnut.2023.1158810)

28. Burgess S, Butterworth A, Thompson SG. Mendelian randomization analysis with multiple genetic variants using summarized data. Genet Epidemiol. (2013) 37:658– 65. doi: [10.1002/gepi.2013.37.issue-7](https://doi.org/10.1002/gepi.2013.37.issue-7)

29. Bowden J, Davey Smith G, Burgess S. Mendelian randomization with invalid instruments: effect estimation and bias detection through Egger regression. Int J Epidemiol. (2015) 44:512–25. doi: [10.1093/ije/dyv080](https://doi.org/10.1093/ije/dyv080)

30. Friedman SL, Neuschwander-Tetri BA, Rinella M, Sanyal AJ. Mechanisms of NAFLD development and therapeutic strategies. Nat Med. (2018) 24:908–22. doi: [10.1038/s41591-018-0104-9](https://doi.org/10.1038/s41591-018-0104-9)

31. Sun Q. Ambient air pollution exaggerates adipose inflammation and insulin resistance in a mouse model of diet-induced obesity. Circulation. (2009) 119:538–46. doi: [10.1161/CIRCULATIONAHA.108.799015](https://doi.org/10.1161/CIRCULATIONAHA.108.799015)

32. Zheng Z. Exposure to ambient particulate matter induces a NASH-like phenotype and impairs hepatic glucose metabolism in an animal model. J Hepatol. (2013) 58:148–54. doi: [10.1016/j.jhep.2012.08.009](https://doi.org/10.1016/j.jhep.2012.08.009)

33. Liu C. Air pollution-mediated susceptibility to inflammation and insulin resistance: influence of CCR2 pathways in mice. Environ Health Perspect. (2014) 122:17–26. doi: [10.1289/ehp.1306841](https://doi.org/10.1289/ehp.1306841)

34. Eze IC. Long-term exposure to ambient air pollution and metabolic syndrome in adults. PloS One. (2015) 10:e0130337–e0130337. doi: [10.1371/journal.pone.0130337](https://doi.org/10.1371/journal.pone.0130337)

35. Reyes-Caballero H, Rao X, Sun Q. Air pollution-derived particulate matter dysregulates hepatic Krebs cycle, glucose and lipid metabolism in mice. Sci Rep. (2019) 9:17423. doi: [10.1038/s41598-019-53716-y](https://doi.org/10.1038/s41598-019-53716-y)

36. Yang B-Y. Ambient air pollution in relation to diabetes and glucosehomoeostasis markers in China: a cross-sectional study with findings from the 33 Communities Chinese Health Study. Lancet Planet Health. (2018) 2:e64–73. doi: [10.1016/S2542-5196\(18\)30001-9](https://doi.org/10.1016/S2542-5196(18)30001-9)

37. VoPham T, Kim NJ, Berry K. PM2.5 air pollution exposure and nonalcoholic fatty liver disease in the Nationwide Inpatient Sample. Environ Res. (2022) 213:113611. doi: [10.1016/j.envres.2022.113611](https://doi.org/10.1016/j.envres.2022.113611)

38. Deng P, Tang H, Zhu L. Association of long-term ambient fine particulate matter (PM2.5) and incident non-alcoholic fatty liver disease in Chinese adults. Environ pollut. (2023) 329:121666. doi: [10.1016/j.envpol.2023.121666](https://doi.org/10.1016/j.envpol.2023.121666)

39. Vw W, La A V, de L. Noninvasive biomarkers in NAFLD and NASH - current progress and future promise. Nat Rev Gastroenterol hepatology Nat Rev Gastroenterol Hepatol. (2018) 15. doi: [10.1038/s41575-018-0014-9](https://doi.org/10.1038/s41575-018-0014-9)

40. Masoodi M, Gastaldelli A, Hyötyläinen T. Metabolomics and lipidomics in NAFLD: biomarkers and non-invasive diagnostic tests. Nat Rev Gastroenterol Hepatol. (2021) 18:835–56. doi: [10.1038/s41575-021-00502-9](https://doi.org/10.1038/s41575-021-00502-9)

41. Tanık VO, Çınar T, Karabağ Y, Şimşek B, Burak C, Çağdaş M, et al. The prognostic value of the serum albumin level for long-term prognosis in patients with acute pulmonary embolism. Clin Respir J. (2020) 14:578–85. doi: [10.1111/crj.13176](https://doi.org/10.1111/crj.13176)

42. Alcorta MD, Alvarez PC, Cabetas RN, MjA Martín, Valero M, Candela CG. The importance of serum albumin determination method to classify patients based on nutritional status. Clin Nutr ESPEN. (2018) 25:110–3. doi: [10.1016/j.clnesp.2018.03.124](https://doi.org/10.1016/j.clnesp.2018.03.124)

43. Cho SY, Han J, Cha SH, Yoon S. Structural basis of serum albumin recognition by SL335, an antibody Fab extending the serum half-life of protein therapeutics. Biochem Biophys Res Commun. (2020) 526:941–6. doi: [10.1016/j.bbrc.2020.03.133](https://doi.org/10.1016/j.bbrc.2020.03.133)

44. Sheinenzon A, Shehadeh M, Michelis R, Shaoul E, Ronen O. Serum albumin levels and inflammation. Int J Biol Macromol. (2021) 184:857–62. doi: [10.1016/j.ijbiomac.2021.06.140](https://doi.org/10.1016/j.ijbiomac.2021.06.140)

45. Małkowski P. Human albumin: old, new, and emerging applications. Ann Transplant. (2013) 18:205–17. doi: [10.12659/AOT.889188](https://doi.org/10.12659/AOT.889188)

46. Don BR, Kaysen G. Poor nutritional status and inflammation: serum albumin: relationship to inflammation and nutrition. Semin Dialysis. (2004) 17:432–7. doi: [10.1111/j.0894-0959.2004.17603.x](https://doi.org/10.1111/j.0894-0959.2004.17603.x)

47. Marın-Palma D, Tabares-Guevara JH, Zapata-Cardona MI, Zapata-Builes W, ́ Taborda N, Rugeles MT, et al. PM10 promotes an inflammatory cytokine response that may impact SARS-CoV-2 replication in vitro. Front Immunol. (2023) 14:1161135. doi: 10.3389/fi[mmu.2023.1161135](https://doi.org/10.3389/fimmu.2023.1161135)

48. Cho J, Lee I, Park DH, Kwak HB, Min K. Relationships between socioeconomic status, handgrip strength, and non-alcoholic fatty liver disease in middle-aged adults. Int J Environ Res Public Health. (2021) 18:1892. doi: [10.3390/ijerph18041892](https://doi.org/10.3390/ijerph18041892)

49. Orkin S, Brokamp C, Yodoshi T. Community socioeconomic deprivation and nonalcoholic fatty liver disease severity. J Pediatr Gastroenterol Nutr. (2020) 70:364–70. doi: [10.1097/MPG.0000000000002527](https://doi.org/10.1097/MPG.0000000000002527)