#### Check for updates

#### OPEN ACCESS

EDITED BY Peiyuan Yin, Dalian Medical University, China

REVIEWED BY Rishat Ruzi, First Affiliated Hospital of Xinjiang Medical University, China Priyanka Singh, Indian Institute of Technology Jodhpur, India

\*CORRESPONDENCE Yong-Jiang Li [yongjiangli@dlut.edu.cn](mailto:yongjiangli@dlut.edu.cn) Zheng-Nan Gao [gao2008@dlut.edu.cn](mailto:gao2008@dlut.edu.cn)

RECEIVED 05 March 2024 ACCEPTED 21 August 2024 PUBLISHED 03 September 2024

#### CITATION

Zhang Y-Y, Li Y-J, Xue C-D, Li S, Gao Z-N and Qin K-R (2024) Effects of T2DM on cancer progression: pivotal precipitating factors and underlying mechanisms. Front. Endocrinol. 15:1396022. [doi: 10.3389/fendo.2024.1396022](https://doi.org/10.3389/fendo.2024.1396022)

#### COPYRIGHT

© 2024 Zhang, Li, Xue, Li, Gao and Qin. This is an open-access article distributed under the terms of the [Creative Commons Attribution](http://creativecommons.org/licenses/by/4.0/) [License \(CC BY\).](http://creativecommons.org/licenses/by/4.0/) The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# [Effects of T2DM on cancer](https://www.frontiersin.org/articles/10.3389/fendo.2024.1396022/full) [progression: pivotal](https://www.frontiersin.org/articles/10.3389/fendo.2024.1396022/full) [precipitating factors and](https://www.frontiersin.org/articles/10.3389/fendo.2024.1396022/full) [underlying mechanisms](https://www.frontiersin.org/articles/10.3389/fendo.2024.1396022/full)

Yu-Yuan Zhang<sup>1,2</sup>, Yong-Jiang Li<sup>1,2\*</sup>, Chun-Dong Xue<sup>1,2</sup>, Shen Li<sup>1</sup>, Zheng-Nan Gao<sup>1\*</sup> and Kai-Rong Qin<sup>1,2</sup>

1 Institute of Cardio-Cerebrovascular Medicine, Central Hospital of Dalian University of Technology, Dalian, Liaoning, China, 2School of Biomedical Engineering, Faculty of Medicine, Dalian University of Technology, Dalian, Liaoning, China

Type 2 diabetes mellitus (T2DM) is a chronic metabolic disorder affecting people worldwide. It is characterized by several key features, including hyperinsulinemia, hyperglycemia, hyperlipidemia, and dysbiosis. Epidemiologic studies have shown that T2DM is closely associated with the development and progression of cancer. T2DM-related hyperinsulinemia, hyperglycemia, and hyperlipidemia contribute to cancer progression through complex signaling pathways. These factors increase drug resistance, apoptosis resistance, and the migration, invasion, and proliferation of cancer cells. Here, we will focus on the role of hyperinsulinemia, hyperglycemia, and hyperlipidemia associated with T2DM in cancer development. Additionally, we will elucidate the potential molecular mechanisms underlying their effects on cancer progression. We aim to identify potential therapeutic targets for T2DM-related malignancies and explore relevant directions for future investigation.

KEYWORDS

T2DM, cancer, molecular mechanism, hyperinsulinemia, hyperglycemia, hyperlipidemia

# 1 Introduction

Type 2 diabetes mellitus (T2DM) is a chronic metabolic disorder primarily characterized by insulin resistance and relative insulin deficiency ([1](#page-11-0)). This condition leads to persistent hyperglycemia ([2](#page-11-0)–[4](#page-11-0)). T2DM accounts for approximately 90% of all diabetes cases worldwide. It is often associated with various systemic complications, including cardiovascular disease, nephropathy, neuropathy, and retinopathy ([3](#page-11-0), [4](#page-11-0)). Hyperglycemia is a hallmark of T2DM, resulting from the body's inability to efficiently use glucose due to impaired insulin action [\(5](#page-11-0)–[10\)](#page-11-0). This chronic elevation of blood glucose levels is a key factor in the development of T2DM-related complications ([9](#page-11-0)–[11](#page-11-0)).

In addition to hyperglycemia, T2DM is often associated with hyperinsulinemia. This condition is an early compensatory response to insulin resistance, where the pancreas produces more insulin to overcome reduced insulin sensitivity [\(12,](#page-11-0) [13](#page-11-0)). Over time, hyperinsulinemia can lead to pancreatic  $\beta$ -cell dysfunction, further exacerbating hyperglycemia. T2DM is also linked with dyslipidemia, characterized by elevated levels of triglycerides and low-density lipoprotein (LDL) cholesterol, along with decreased levels of high-density lipoprotein (HDL) cholesterol [\(14](#page-11-0)–[16](#page-11-0)). This lipid imbalance further contributes to the increased risk of cardiovascular disease in T2DM patients.

Epidemiologic studies consistently show that individuals with T2DM have an increased risk of developing several malignancies ([17](#page-11-0)–[20\)](#page-11-0). These include lung, colorectal, liver, breast, gastric, and pancreatic cancers ([21\)](#page-11-0). The pathological features of T2DM, particularly hyperinsulinemia, hyperglycemia, and hyperlipidemia, are believed to contribute to this increased cancer risk (Figure 1). Hyperglycemia promotes cancer cell proliferation by providing a readily available source of glucose, fueling rapid cell division [\(22](#page-11-0), [23\)](#page-12-0). Hyperinsulinemia promotes tumor growth and progression through its mitogenic effects by activating insulin and insulin-like growth factor (IGF) signaling pathways ([24,](#page-12-0) [25](#page-12-0)). Dyslipidemia contributes to chronic

inflammation and oxidative stress, creating a pro-tumorigenic environment ([26](#page-12-0)–[28\)](#page-12-0).

Despite these associations, the precise molecular mechanisms linking T2DM-related features to cancer progression remain incompletely understood. In this review, we will provide an overview of the epidemiologic associations between T2DM and cancer. We will also explore the potential molecular mechanisms involved. Specifically, we will examine how hyperglycemia, hyperinsulinemia, and hyperlipidemia may accelerate cancer development and progression in patients with T2DM.

# 2 Epidemiological association between T2DM-related features and cancer

In recent years, clinical researchers have conducted extensive epidemiological studies to elucidate the relationship between these two diseases ([29](#page-12-0)). The majority of the findings from these studies



hyperinsulinemia, hyperglycemia, hyperlipidemia, and other factors. T2DM, Type 2 diabetes mellitus.

consistently show a significantly increased risk of cancer among individuals with T2DM. To gain a clearer understanding of the association between T2DM and cancer, this section compiles and summarizes epidemiological research on the three key features of T2DM: hyperinsulinemia, hyperglycemia, and hyperlipidemia in relation to cancer.

Numerous epidemiological studies have consistently shown that the increased risk of cancer among people with T2DM is significantly influenced by hyperinsulinemia. Researchers have discovered that hyperinsulinemia increases the risk of colorectal, breast, endometrial, hepatocellular, and prostate cancers through assessments of the relationship between the empirical dietary index for hyperinsulinemia (EDIH) and specific cancer risks ([30](#page-12-0)–[34\)](#page-12-0). Furthermore, hyperinsulinemia increases the likelihood of cancer recurrence in individuals with prostate cancer (PCa), breast cancer, and curative hepatocellular carcinoma (HCC) [\(35](#page-12-0)–[37](#page-12-0)). While being unrelated to survival rates in individuals with late-stage colon cancer, a prospective investigation found that a higher EDIH associated with hyperinsulinemia may increase the incidence risk of colon cancer ([38](#page-12-0)). Moreover, one of the contributing factors to hastening mortality in cancer patients is hyperinsulinemia ([39\)](#page-12-0). Specific research results have indicated that hyperinsulinemia increases the probability of dying from colorectal, gastroenteric, and hepatic malignancies by 1.51-fold, 1.61-fold, and 2.72-fold, respectively [\(40](#page-12-0)–[42](#page-12-0)). It is noteworthy that hyperinsulinemia raises the risk of death among cancer patients regardless of whether individuals fall into the obese category ([43](#page-12-0)). Opting for meals with a lower insulinemic potential may be a useful strategy to improve general health and prevent early death [\(44](#page-12-0)). The findings of the previous study demonstrate that hyperinsulinemia may independently influence cancer incidence and mortality rates.

While glycemic control has received significant attention in the context of T2DM, the importance of hyperglycemia as a key component in tumor growth is sometimes overlooked. Nevertheless, epidemiological research has established a connection between hyperglycemia and cancer incidence, progression, and mortality rates [\(7\)](#page-11-0). For instance, in a study investigating the impact of blood sugar on cancer risk in women from the 1987–1992 ORDET cohort, conditional logistic regression was employed to determine the rate ratios (RR). The results indicated that women with blood sugar levels in the highest quartile had a significantly higher risk of breast cancer compared to those in the lowest quartile (RR 1.63; 95% CI: 1.14-2.32; trend  $p$ -value = 0.003) [\(45\)](#page-12-0). Additionally, a recent systematic study sheds light on the major influence of diabetes and hyperglycemia on the occurrence of colon cancer [\(46](#page-12-0)). Although several cohort studies have emphasized the link between diabetes and cancer risk, only a limited number of studies have directly examined the underlying impact of high blood sugar on cancer risk [\(7](#page-11-0)). At present, clinical studies are the main focus of research on the relationship between hyperglycemia and cancer ([47\)](#page-12-0). Clinical research has demonstrated that elevated blood glucose is associated with poor clinical outcomes in patients with T2DM and cancer ([5\)](#page-11-0). Conversely, the prognosis for cancer in diabetic people improves when plasma glucose levels return to normal. This suggests that T2DM and cancer share common signaling pathways that hyperglycemia may activate. Therefore, regulating plasma glucose levels in people with cancer and T2DM may improve the prognosis for cancer.

Hyperlipidemia primarily develops as a result of dysregulated lipid metabolism. Lipids serve crucial functions in cellular signaling, chemical transport, and energy storage and they are essential components of cell membranes. An increasing body of evidence from recent studies indicates that abnormal lipid metabolism increases the risk of tumorigenesis, disease progression, and treatment resistance ([28,](#page-12-0) [48](#page-12-0)). Excess lipid and cholesterol accumulation within cancer cells is stored in lipid droplets, and higher lipid droplet levels and stored cholesterol ester content are now considered indicators of cancer invasion [\(49\)](#page-12-0). Furthermore, lipid metabolism reprogramming has emerged as a new factor in cancer development ([28](#page-12-0)). Recent epidemiological studies have highlighted the critical role of lipid metabolism in the progression of various cancers, including breast cancer [\(50](#page-12-0)), thyroid cancer [\(51](#page-12-0)), prostate cancer [\(52](#page-12-0)), and ovarian cancer ([53](#page-12-0)). Elevated serum cholesterol levels have been linked to an increased risk of colon, rectal, prostate, and testicular cancers [\(54\)](#page-12-0). Interestingly, low cholesterol levels have occasionally been associated with cancer. A prospective study based on the Japan Public Health Center found a significant correlation between low cholesterol levels and the incidence of primary liver and stomach malignancies [\(55\)](#page-12-0). Recent research has also linked low cholesterol to colon and lung malignancies [\(54](#page-12-0), [56](#page-12-0)). However, due to the conflicting epidemiological results mentioned above, it is still unclear how cholesterol contributes to the development of cancer. According to some recent research, low cholesterol levels may be linked to impaired immune function, increasing one's vulnerability to cancer [\(57\)](#page-12-0). In contrast, low cholesterol levels may lower the risk of cancer by activating the nuclear factor-kB (NF-kB), which inhibits cell proliferation and promotes cell differentiation ([58](#page-12-0)). Furthermore, extracellular vesicle proteins and lipids could serve as biomarkers for lung cancer diagnosis and prognosis, as well as therapeutic targets for drug development ([59\)](#page-12-0). While epidemiological research examines the impact of aberrant blood lipid levels on cancer, the heterogeneity of study results makes it challenging to establish causal correlations. Uncovering the specific pathways by which hyperlipidemia promotes cancer remains a critical task.

### 3 Hyperinsulinemia and cancer progression

In patients with T2DM, hyperinsulinemia is associated with an increased risk of cancer. It can also accelerate cancer progression through both direct and indirect mechanisms [\(Figure 2\)](#page-3-0). These mechanisms involve the activation of key signaling pathways and the modulation of extracellular factors that influence tumor growth.

#### 3.1 Direct mechanisms

Under normal conditions, insulin primarily regulates glucose metabolism with limited direct involvement in cancer progression. However, in individuals with hyperinsulinemia, the excessive

<span id="page-3-0"></span>

interleukin-10; IGF-1R, IGF-1 receptor; MEK, mitogen-activated protein kinase; ERK, extracellular signal-regulated kinase.

activation of insulin signaling pathways, including the phosphatidylinositol 3-kinase/protein kinase B (PI3K/AKT), mTOR, and Ras/mitogen-activated protein kinase (MAPK) pathways, significantly enhances tumor growth ([60](#page-12-0)–[65\)](#page-12-0). 1) PI3K/ AKT Pathway: In hyperinsulinemic states, the PI3K/AKT pathway is persistently activated due to chronic insulin receptor (IR) stimulation, leading to enhanced tumor cell proliferation, migration, and survival ([66](#page-12-0)). This pathway also inhibits apoptosis by downregulating pro-apoptotic proteins such as B-cell lymphoma-2 (Bcl-2)-associated death promoter and rapidly accelerated fibrosarcoma-1 (Raf-1), thereby promoting tumor cell survival [\(67](#page-12-0)). 2) mTOR Pathway: The mTOR pathway is activated by abnormal insulin signaling, enhancing synthetic metabolism, cell proliferation, and inhibiting autophagy ([68](#page-12-0)). This contributes to tumor growth by facilitating the accumulation of biomolecules essential for cancer cell proliferation. 3) Ras/MAPK Pathway: Hyperinsulinemia also activates the Ras/MAPK pathway, which regulates cell division, migration, and proliferation ([62\)](#page-12-0). This pathway is particularly important in promoting the aggressive spread of tumors and is implicated in various cancers, including prostate and endometrial cancers ([69\)](#page-12-0).

### 3.2 Indirect mechanisms

Normally, systemic factors are balanced to regulate cell growth. In hyperinsulinemia, elevated insulin levels indirectly promote tumorigenesis. This occurs through alterations in the insulin-like growth factor-1 (IGF-1) and sex hormone-binding globulin (SHBG) system [\(13,](#page-11-0) [65\)](#page-12-0). Insulin increases IGF-1 synthesis in the liver and upregulating growth hormone receptors. This leads to elevated IGF-1 levels in the bloodstream ([70](#page-12-0)–[72](#page-13-0)). In hyperinsulinemic states, these elevated IGF-1 levels result in enhanced proliferation, migration, and anti-apoptosis in HCC cell lines, such as SK-Hep1 and HepG2. Additionally, there is increased resistance to sorafenib. These effects are mediated by the regulation of the PI3K/AKT and RAS/Raf/ERK signaling pathways ([24](#page-12-0)). In addition to increasing cancer risk through IGF-1, insulin also influences carcinogenesis and progression by lowering sex hormone-binding globulin (SHBG) levels [\(73](#page-13-0)). Normally, sex hormone levels are tightly regulated to maintain physiological balance. However, during conditions like menopause or hormonal imbalances, elevated sex hormones can increase the risk of certain cancers. For example, prolonged exposure to high estrogen levels is closely linked to the development of breast and endometrial cancers. In individuals with hyperinsulinemia, elevated insulin levels reduce SHBG. Lower SHBG levels lead to higher circulating levels of free estrogen and testosterone. This increase in active sex hormones may accelerating the onset and progression of hormonerelated cancers, such as endometrial and breast cancers ([74\)](#page-13-0). Notably, recent research suggests a close relationship between sex hormones, such as estrogen and testosterone, and the expression of the insulin-degrading enzyme (IDE). High levels of these hormones can enhance the negative feedback of IDE on itself. Additionally, elevated hormone levels may increase the risk of hormonedependent malignancies ([75](#page-13-0)).

#### 3.3 Inflammation and immune modulation

The inflammatory milieu associated with hyperinsulinemia is distinct from that of normal individuals, contributing to a more pro-tumorigenic environment. Persistent high levels of insulin can promote generation of an inflammation environment, which plays a key role in tumor initiation and progression [\(12\)](#page-11-0). Studies have shown that hyperinsulinemic patients secrete more proinflammatory cytokines, including interleukin-6 (IL-6) and tumor necrosis factor-alpha (TNF- $\alpha$ ), compared to normal individuals. Under inflammatory stimuli such as transforming growth factor- $\beta$ (TGF-b), IL-6, and interleukin-10 (IL-10), tumor-associated macrophages (TAMs) are more likely to transition from the antitumor M1 phenotype to the pro-tumor M2 phenotype [\(76\)](#page-13-0). Within the tumor microenvironment (TME), M2 macrophages contribute to tumor promotion by facilitating immune escape ([77](#page-13-0)). Moreover, M2 macrophages can influence the outcomes of radiation, chemotherapy, and immunotherapy by impacting the drug resistance of tumor cells [\(78\)](#page-13-0). Other immune subsets, such as neutrophils and dendritic cells, also play roles within the TME.

In conclusion, cancer can develop in both individuals with normal insulin levels and those with hyperinsulinemia. However, hyperinsulinemia significantly accelerates cancer progression and alters the tumor microenvironment. This leads to more aggressive cancer phenotypes. These distinctions underscore the importance of understanding the specific mechanisms by which hyperinsulinemia influences cancer to develop targeted interventions.

# 4 Hyperglycemia and cancer progression

In patients with T2DM, high blood glucose levels, or hyperglycemia, can influence the development and progression of cancer through various mechanisms, as illustrated in [Figure 3.](#page-5-0) These include increased oxidative stress, chronic inflammation, and the activation of specific signaling pathways like the insulinlike growth factor (IGF) and advanced glycation end products (AGEs)/receptor for AGEs (RAGE) pathways.

#### 4.1 Increased oxidative stress

Normally, the oxidative environment in the human body maintains a dynamic balance. However, elevated glucose levels in the bloodstream intensify intracellular glucose metabolism, leading to excessive production of reactive oxygen species (ROS). This increased ROS disrupts the electron transport chain, impairs adenosine triphosphate (ATP) synthesis, and promotes mitochondrial dysfunction, further elevating ROS levels ([79](#page-13-0)–[81\)](#page-13-0).The accumulation of ROS due to mitochondrial malfunction degrades cellular components and stimulates tumor formation through deoxyribonucleic acid (DNA) mutations and genomic instability ([81](#page-13-0)–[83\)](#page-13-0). Moreover, hyperglycemia activates NADPH oxidase, a key ROS generator, thereby exacerbating oxidative stress, damaging DNA, activating oncogenes, and suppressing tumor suppressor genes, all of which contribute to tumor growth and progression ([82](#page-13-0)–[84\)](#page-13-0).

Moreover, hyperglycemia induces the production of inflammatory cytokines, such as TNF- $\alpha$  and IL-6 [\(85](#page-13-0), [86](#page-13-0)), which further elevate ROS levels by activating NADPH oxidase ([87](#page-13-0)). These inflammatory cytokines, along with increased ROS, contribute to chronic inflammation and create a tumor-promoting microenvironment. Normally, antioxidant enzymes neutralize harmful reactive molecules, but hyperglycemia reduces the activity of these enzymes, like catalase and superoxide dismutase (SOD), resulting in ROS accumulation and cellular oxidative damage [\(88\)](#page-13-0).

Molecular regulation plays a crucial role in the connection between cancer and hyperglycemia-induced oxidative stress. For instance, oxidative stress activates the transcription factor GATA1, which upregulates the expression of von Willebrand Factor, thereby promoting tumor metastasis [\(89\)](#page-13-0). While the complex interactions between these molecular components are not yet fully understood, it is clear that oxidative stress driven by hyperglycemia can lead to the formation of DNA adducts and chromosomal abnormalities, contributing to genetic instability and cancer development [\(90,](#page-13-0) [91\)](#page-13-0).

In conclusion, hyperglycemia impacts tumor development and progression in individuals with T2DM. This effect is more pronounced compared to individuals with normal glucose levels. Hyperglycemia increases oxidative stress, which contributes to cellular damage, genetic mutations, and abnormal cellular development

### 4.2 Warburg effect

The Warburg effect was first discovered by Otto Warburg in the 1950s ([92](#page-13-0)). It describes the tendency of cancer cells to prefer lactate production through glycolysis. This preference occurs even when sufficient oxygen is available for ATP synthesis via oxidative phosphorylation. In normal, well-differentiated cells, energy is primarily generated through a combination of the tricarboxylic acid cycle (TCA) and oxidative phosphorylation, yielding up to 36 mol of ATP from 1 mol of glucose [\(93\)](#page-13-0). However, in cancer cells, glycolysis predominates even in aerobic conditions, leading to production of lactate rather than ATP [\(94\)](#page-13-0).

<span id="page-5-0"></span>

#### FIGURE 3

Mechanisms underlying the promotion of cancer progression by hyperglycemia. HIF1α, hypoxia-inducible factor 1α; LDHA, lactate dehydrogenase A; PI3K, phosphatidylinositol 3-kinase; AKT, protein kinase B; MAPK, mitogen-activated protein kinase; JAK, Janus kinase; STAT, signal transducer and activator of transcription; FAK, focal adhesion kinase; Erk, extracellular signal-regulated kinase; Elk1, comprising E26 transformation-specific domaincontaining protein Elk-1; EGR1, early growth response protein 1; Src, sarcoma; IGFBP-2, Insulin-like growth factor binding protein-2; IL-1ß, interleukin-1 $\beta$ ; IL-6, interleukin-6; TNF-a, tumor necrosis factor-a; IL-18, interleukin-18; TGF- $\beta$ , transforming growth factor- $\beta$ ; Sp1, specificity Protein 1; MMPs, matrix metalloproteinases; VEGF, vascular endothelial growth factor; Ang-2, angiopoietin-2; COX-2, cyclooxygenase-2; iNOS, inducible nitric oxide synthase; BcL-X, B-cell lymphoma-extra large; BcL-2, B-cell lymphoma-2; XIAP, X-linked inhibitor of apoptosis protein.

In individuals with normal blood sugar levels, cancer cells may still exhibit the Warburg effect, but this metabolic shift is less pronounced due to the limited availability of glucose. However, in the context of T2DM and associated hyperglycemia, elevated glucose levels significantly amplify the Warburg effect. High glucose levels increase aerobic glycolysis in cancer cells, promoting cancer growth ([95](#page-13-0), [96](#page-13-0)). Glucose enters cells via the glucose transporter 1 and triggers various metabolic pathways, some of which increase the invasive potential of cancer cells ([97\)](#page-13-0). For instance, high glucose conditions have been shown to upregulate key glycolytic enzymes, such as hexokinase II and pyruvate kinase in breast cancer cells [\(98\)](#page-13-0) and enolase 1 in gastric cancer cells [\(99](#page-13-0)).

In pancreatic cancer, hyperglycemia induces the accumulation of hypoxia-inducible factor  $1\alpha$ , leading to increased lactate dehydrogenase A (LDHA) activity and expression [\(100,](#page-13-0) [101\)](#page-13-0). This upregulation elevates the glycolytic rate, accelerating cancer progression and worsening disease outcomes ([100](#page-13-0), [101](#page-13-0)). The upregulation of LDHA activity converts pyruvate to lactate, which is extruded through monocarboxylate transporter 4, contributing to the formation of an acidic TME [\(102](#page-13-0)). Lactate, once considered a metabolic waste, has intriguingly been discovered to enhance the metabolism of regulatory T cells that infiltrate tumors, allowing cancer cells to evade immune cytotoxicity [\(103](#page-13-0)).

In summary, the Warburg effect is a common feature of cancer cells. In individuals with T2DM, hyperglycemia significantly enhances this effect. This leads to increased glycolysis and lactate production. Consequently, the cancer phenotype becomes more aggressive.

### 4.3 Chronic inflammation

Chronic inflammation plays a pivotal role in cancer development and progression through multiple pathways. The chemokines and growth factors generated during the inflammatory response can enhance the development, invasion, and metastasis of cancer cells while preventing normal cell death [\(104](#page-13-0)). In individuals with T2DM, hyperglycemia significantly amplifies chronic inflammation compared to normal glucose levels with individuals, accelerating cancer progression ([105](#page-13-0)). In individuals with hyperglycemic, prolonged exposure to high glucose levels increases the formation of AGEs, which interact with the receptor for AGEs (RAGE). This interaction activates several signaling pathways, including NF-kB, MAPK, and Janus kinase/signal transducer and activator of transcription (JAK/ STAT), leading to heightened production of inflammatory cytokines like Interleukin-1  $\beta$  (IL-1 $\beta$ ), IL-6, and TNF- $\alpha$  [\(18](#page-11-0)). These cytokines

exacerbate the inflammatory response, creating a microenvironment that significantly promotes cancer cell proliferation and survival.

Hyperglycemia also increases oxidative stress, leading to the activation of transcription factors such as NF-kB and activator protein-1 (AP-1), and the upregulating genes related to inflammation. This results in the release of inflammatory cytokines like IL-1 $\beta$ , Interleukin-18 (IL-18), and IL-6 ([106\)](#page-13-0). The free radicals generated by oxidative stress in hyperglycemic conditions cause more severe DNA damage, contributing to DNA mutations and chromosomal abnormalities underlying cancer development. Moreover, hyperglycemia increases the production of protein kinase C (PKC) isoforms, including PKC- $\alpha$ , PKC- $\beta$ 1, PKC- $\beta$ 2, and PKC- $\delta$  ([107,](#page-13-0) [108](#page-13-0)). The increased activity of PKC can further activate NF-kB and enhance the expression of inflammatory molecules, including TNF- $\alpha$  and TGF- $\beta$  [\(109](#page-13-0)).

Additionally, hyperglycemia disrupts lipid metabolism, causing adipose tissue to produce excess free fatty acids. Elevated free fatty acid levels can induce inflammatory reactions through various signaling pathways, such as the activation of pro-inflammatory serine/threonine protein kinase cascades, which encourage the release of IL-6 and stimulate C-reactive protein production ([110\)](#page-13-0). Excess free fatty acids also induce endoplasmic reticulum stress and suppress glucose metabolism, further increasing inflammation ([111](#page-13-0), [112\)](#page-13-0). Furthermore, high levels of free fatty acids may suppress the expression of genes involved in glucose metabolism, such as glucose transporters, reducing intracellular glucose uptake and increasing the risk of inflammation in hyperglycemic situations ([97,](#page-13-0) [113\)](#page-13-0).

The sustained and intensified inflammatory response in individuals with hyperglycemia not only accelerates cancer cell growth but also impairs the immune system's ability to detect and eliminate malignant cells [\(114](#page-13-0), [115\)](#page-13-0). This altered immune function, combined with the chronic inflammatory environment, fosters the rapid progression and spread of cancer. On the contrary, normal people have a healthier physiological environment and do not experience long-term inflammatory reactions, which may be one of the reasons for the low risk of cancer. Therefore, a comprehensive approach that includes glycemic control, reduction of oxidative stress, and modulation of inflammatory responses are essential strategies to reduce risk of cancer in patients with T2DM.

#### 4.4 The IGF signaling pathway

IGF serves as a critical mediator of growth, development, and survival. It also contributes to an increased risk of cancer. IGF promotes cancer cell proliferation and inhibits apoptosis ([116\)](#page-13-0). In normoglycemic individuals, insulin secretion and IGF-1R activation are tightly regulated, resulting in a more controlled rate of cancer cell growth. However, in hyperglycemia individuals, regulation of the IGF signaling pathway influences cancer progression ([117\)](#page-13-0). Specifically, elevated glucose levels stimulate the release of more insulin by pancreatic  $\beta$ -cells, leading to higher insulin levels in the bloodstream. This increase in insulin can directly or indirectly impact the IGF-1R, which belongs to the receptor tyrosine kinase family. The activation of IGF-1R by insulin triggers several signaling

pathway, including PI3K/AKT, MAPK, JAK/STAT, Sarcoma (Src), and focal adhesion kinase (FAK), all of which collectively enhance cancer cell proliferation, survival, and migration [\(25](#page-12-0)). Moreover, insulin may affect the expression of insulin-like growth factor binding protein (IGFBP) to regulate the bioavailability and transport of IGF-1, indirectly affecting its efficacy. For instance, in differentiated 3T3-L1 adipocytes, insulin can stimulate the transcription of the IGFBP-2 gene and increase its secretion ([118\)](#page-13-0). Early cancer cell studies have also found that the insulin signaling pathway regulates IGFBP-2 transcription, consistent with the aforementioned results ([119\)](#page-13-0). With elevated IGFBP-2 concentrations, the bioavailability of IGF-1 decreases, impeding its transport and potentially weakening its impact on tumor progression. However, epidemiological studies have shown that overexpression of IGFBP-2 is associated with aggressive phenotypes in various human cancers, including glioma, ovarian cancer, prostate cancer, pancreatic cancer, breast cancer, lung cancer, colorectal cancer, melanoma, liver cancer, gastric cancer, rhabdomyosarcoma, and leukemia ([120\)](#page-13-0). Exogenous IGFBP-2 can promote the activation of the integrin  $\beta$ 1/FAK/ERK/Elk1/EGR1 pathway, thereby stimulating HCC cell proliferation ([121\)](#page-13-0). Interestingly, some studies have found that insulin further inhibits the secretion of IGFBP-1 and IGFBP-2 ([122\)](#page-13-0), a result in stark contrast to previous findings. Therefore, the action of IGF-1 might be minimally affected by IGFBP but rather influenced by the activation of IGF-1R and its downstream signaling pathways to impact cancer progression.

#### 4.5 The AGEs/RAGE signaling pathway

Hyperglycemia leads to the formation of AGEs, which are nonenzymatic glycation products, resulting from the interaction between the aldehyde group of reducing sugars and macromolecules such as proteins, amino acids, lipids, and nucleic acids. In normoglycemic individuals, the formation of AGEs is significantly lower. This reduces the likelihood of AGE accumulation. Consequently, the detrimental effects of AGEs on cellular function are minimized. RAGE, a pattern-recognition receptor in the immunoglobulin superfamily, interacts with extracellular AGEs. RAGE is constitutively expressed in immune cells, lung tissues, and certain cancer cells, including pancreatic cancer ([123](#page-13-0)), non-small cell lung cancer [\(124](#page-13-0)), gastric cancer ([125\)](#page-14-0), and breast cancer [\(126](#page-14-0)). The interaction between AGEs and RAGE activates multiple signaling pathways that critical for tumor growth, angiogenesis, and invasion, such as the PI3K/AKT/mTOR, MAPK, MMPs, vascular endothelial growth factor (VEGF), NF-kB, JAK/ STAT, and p53 [\(127](#page-14-0)).

In hyperglycemic conditions, the AGE-RAGE interaction is significantly enhanced due to the elevated levels of glucose and subsequent increase in AGE formation. This leads to a more aggressive activation of signaling pathways that promote cancer progression. For example, in PCa-3 cells, the interaction between AGEs and RAGE activates the PI3K/AKT pathway, increasing the phosphorylation of downstream Retinoblastoma protein (Rb) and decreasing overall Rb levels, ultimately enhancing cell proliferation ([128\)](#page-14-0). Similarly, hyperglycemia-induced AGE accumulation activates RAGE, leading to elevated ERK phosphorylation and increased expression of MMP-2 and MMP-9, which drive oral cancer cell migrate and further deterioration [\(129](#page-14-0)). In gastric cancer, the RAGE/ERK/Sp1/MMP-2 pathway is significantly activated under hyperglycemic conditions, promoting invasion and metastasis [\(130](#page-14-0)). Furthermore, the binding of AGEs to RAGE improves transcription of NF-kB and AP-1, resulting in upregulation of VEGF and ang2 mRNA levels, promoting angiogenesis ([131\)](#page-14-0), which plays an important role in the cancer development.

The activation of the AGE/RAGE axis induces a strong proinflammatory response. This leads to increased leukocyte activation and apoptosis, accelerated desmoplastic responses, and the recruitment of stromal cells into the TME ([132](#page-14-0)). In hyperglycemic individuals, the chronic elevation of AGEs continuously stimulates RAGE. This exacerbates oxidative stress and promotes NF-kB activation. Consequently, there is an increase in the synthesis and secretion of cytokines, chemokines, and adhesion molecules ([133\)](#page-14-0). This hyperactivation contributes to a tumor-promoting microenvironment. It leads to elevated levels of pro-inflammatory cytokines, pro-angiogenic factors, and antiapoptotic signals. Specifically, the transcription and translation of pro-inflammatory cytokines such as TNF-a, cyclooxygenase-2, inducible nitric oxide synthase, IL-1, and IL-6 are enhanced. Additionally, pro-angiogenic factors and anti-apoptotic signals, including B-cell lymphoma-extra-large (Bcl-xL), Bcl-2, and Xlinked inhibitor of apoptosis protein (XIAP), are upregulated ([132\)](#page-14-0).

The AGEs/RAGE signaling pathway plays a crucial role in glioma-associated microglia, TAM, and breast cancer-associated fibroblasts (CAFs). The TAM and glioma-associated microglia contribute to tumor development, invasion, and angiogenesis by secreting VEGF and pro-inflammatory cytokines [\(134\)](#page-14-0). In MDA-MB-231 breast cancer cells, the CAFs promote migration and induce phenotypic alterations linked to invasion by upregulating interleukin-8 (IL-8) levels and activating the IL-8/C-X-C chemokine receptor type 1/2 paracrine signaling pathway [\(135\)](#page-14-0). Therefore, the AGEs/ RAGE signaling pathway can serve as a significant target for cancer therapy. Currently, inhibition of RAGE signaling has effectively suppressed the growth, migration, and invasion of cancer cells ([126](#page-14-0), [136\)](#page-14-0). In clinical practice, metformin is a commonly used medicine for the treatment of T2DM. Additionally, it has shown therapeutic advantages in those with T2DM and concomitant malignancy. As a result, the combination of metformin and AGE/ RAGE inhibitors may be a useful therapeutic approach to treating T2DM-related cancer.

### 5 Hyperlipidemia and cancer progression

Dyslipidemia, characterized by increased triglycerides (TG), total cholesterol (TC), and LDL levels, as well as decreased HDL concentrations, plays a complex role in cancer progression ([16\)](#page-11-0). The interaction between hyperlipidemia and cancer involves several

mechanisms that are not fully understood. The processes through which hyperlipidemia promotes cancer development are shown in [Figure 4.](#page-8-0) LDL and its oxidized form, Ox-LDL, promote cancer cell proliferation, angiogenesis, invasion, and metastasis through interactions with receptors such as LDLR, LOX-1, and CD36 ([137\)](#page-14-0). Furthermore, HDL, especially when oxidized or glycated in conditions like T2DM, can also enhance cancer cell migration and proliferation ([138\)](#page-14-0). These processes are driven by aberrant lipid metabolism and cholesterol accumulation in cancer cells [\(139](#page-14-0)). In summary, hyperlipidemia promotes tumor growth in T2DM patients through various mechanisms, which we will further investigate in the following chapters.

#### 5.1 Cellular processes

In normal individuals, lipid levels are tightly regulated, preventing excessive intracellular cholesterol, which increases the risk of cancer. However, in hyperlipidemic conditions, abnormal alterations in the TG, TC, LDL, and HDL levels can disrupt cellular signaling pathways, encouraging tumor cell proliferation, invasion, migration, and anti-apoptosis. Interestingly, in prostate cancer, cancer cells can accumulate intracellular cholesterol levels by disrupting normal mitochondria, thereby promoting their own proliferation and migration ([140,](#page-14-0) [141](#page-14-0)). Additionally, breast, prostate, and pancreatic malignant tumor cells are encouraged to proliferate and invade by LDL cholesterol signaling, which activates the AKT, ERK, and STAT3 pathways ([142,](#page-14-0) [143\)](#page-14-0). The LDLR superfamily member low-density lipoprotein receptor-related protein 1 promotes tumor cell motility and invasion by modulating MMP-2 and MMP-9 production. It also suppresses cell apoptosis by modulating IR, serine/threonine protein kinase signaling pathways, and cysteine-aspartic acid protease-3 expression [\(15\)](#page-11-0).

### 5.2 Tumor microenvironment, angiogenesis and immunosuppression

The tumor microenvironment (TME) is a dynamic system of cells, chemicals, and stromal components that support tumor growth, invasion, and metastasis. In hyperlipidemic conditions, elevated lipid levels further exacerbate these processes compared to individuals with normal lipid levels. Angiogenesis, a critical factor in tumor progression, is particularly enhanced under hypoxic conditions. HDL contributes to this by stimulating the PI3K/Akt pathway via the scavenger receptor class B-I (SR-BI), leading to the accumulation of hypoxia-inducible factor-1 $\alpha$  (HIF-1 $\alpha$ ) and the activation of angiogenic protein transcription ([144](#page-14-0), [145\)](#page-14-0). Additionally, HDL increases the phosphorylation of the vascular endothelial growth factor receptor 2, which in turn activates the downstream pathways ERK1/2 and p38 MAPK to promote angiogenesis [\(146](#page-14-0)). However, HDL can also modulate macrophages within the inflammatory TME, potentially inhibiting pathological angiogenesis [\(146\)](#page-14-0).

Cholesterol plays a pivotal role in modulating the immune response within the TME. It activates the ER stress sensor X-box binding protein 1 (XBP1) in CD8+ T cells, leading to the

<span id="page-8-0"></span>

transcription of proteins like programmed cell death protein-1 (PD-1) and CD244, contributing to T cell exhaustion [\(147](#page-14-0)). Cholesterol efflux in TAMs also enhances their tumor-promoting properties by increasing IL-4 signaling and reducing IFN- $\gamma$ -induced gene expression [\(148](#page-14-0)).

### 5.3 The LDL and Ox-LDL signal pathways

LDL is a key plasma lipoprotein that primarily transports cholesterol throughout the body. Upon oxidation, LDL forms Ox-LDL [\(149](#page-14-0)), which plays a significant role in various pathological conditions, including cancer progression. In normal individuals, where blood lipid levels remain in a dynamic balance, LDL and OxLDL signaling contribute to a low risk of cancer. However, in individuals with hyperlipidemia, elevated LDL levels significantly alter cancer progression dynamics, leading to more aggressive tumor growth and resistance to therapy.

Normally, LDL binds to LDLR on the cell surface, where it is internalized and processed to release cholesterol for cellular use ([150\)](#page-14-0). This process is tightly regulated by mechanisms involving sterol regulatory element-binding protein-2 (SREBP-2), SREBP cleavage-activating protein, and proprotein convertase subtilisin/ kexin 9 ([151\)](#page-14-0). However, cancer cells can exploit this pathway by upregulating LDLR expression, leading to increased cholesterol uptake, which supports rapid cell proliferation and tumor growth ([152](#page-14-0)). Although this mechanism is present in normolipidemic individuals, its impact is amplified in hyperlipidemic conditions.

In such cases, excess LDL supplies a larger amount of cholesterol and lipids for energy, further fueling cancer cell metabolism. Dysregulated lipid metabolism can result in lipotoxicity and elevated oxidative stress, which enhances the susceptibility of LDL to oxidation, forming Ox-LDL [\(137,](#page-14-0) [153](#page-14-0)). Ox-LDL binds to receptors such as LOX-1 and CD36, triggering cascades of oncogenic signals that are more pronounced in hyperlipidemic conditions. This leads to mutations, promotion the epitheliummesenchyme transition, initiation protective autophagy, and production of growth factors, cytokines, and pro-inflammatory markers. The resulting increase in ROS and pro-inflammatory markers significantly contributes to cancer progression and resistance to chemotherapy [\(137](#page-14-0)).

Research into the connection between LDL and Ox-LDL and their effects on cancer is still in its early stages. In MDA-MB-231 breast cancer cells, an increase in LDL levels (greater than 1.5 times that of the control group) led to upregulation of 147 mapped genes (including pERK, pAKT, and pJNK) and downregulation of 95 mapped genes (including CD226, Claudin7, Ocludin, and  $integrin\beta8$ ), while reducing cell adhesion and promoting cell migration and proliferation ([142,](#page-14-0) [154](#page-14-0)–[157](#page-14-0)). LDL activates STAT3 and JAK1, JAK2, and Src in prostate and pancreatic tumor cells, promoting cancer cell proliferation, migration, invasion, and the up-regulation of numerous oncogene products [\(143](#page-14-0)). Furthermore, increased LDL cholesterol in kidney carcinoma activates the PI3K/ AKT signaling pathway, which inhibits the anticancer effects of tyrosine kinase inhibitors ([158\)](#page-14-0). It's important to note that cancer cells may be able to evade immune surveillance at high LDL levels. When V9γδ2 T cells are activated and express LDLR, uptake of lowdensity lipoprotein cholesterol leads to decreased mitochondrial mass and reduced ATP production, resulting in inhibition of the antitumor function of V9 $\gamma$  $\delta$ 2 T cells ([159\)](#page-14-0).

Ox-LDL has been demonstrated to induce mutagenesis and enhance cancer cell proliferation, migration, invasion, and treatment resistance, in addition to the impact of LDL on cancer cells ([137](#page-14-0), [160\)](#page-14-0). In an experimental investigation on primary rat hepatocytes, Ox-LDL, the principal component of 4 hydroxynonenal and lipotoxic, encouraged micronucleus formation, chromosomal aberration, and increased sister chromatid exchange frequency at concentrations ranging from  $0.1$ -10  $\mu$ M, resulting in enhanced DNA damage and induced mutation [\(161](#page-14-0)). Similarly, up-regulation of microRNA-210 and HIF-1 by Ox-LDL increases the risk of vascular disease and cancer ([162,](#page-14-0) [163](#page-14-0)). Furthermore, Ox-LDL receptors such as LOX-1 and CD36, along with associated downstream signaling cascades, play an important role in cancer progression. Recent investigations have demonstrated that the Ox-LDL/LOX-1 axis facilitates the migration of cancer cells by attracting neutrophils to tumor endothelial cells ([164](#page-14-0)). Additionally, the interaction of Ox-LDL with LOX-1 activates NF-kB target genes like VEGF, MMP-2, and MMP-9 to encourage the growth, invasion, and angiogenesis of cancer cells [\(137](#page-14-0), [160](#page-14-0)). In prostate cancer cells, Ox-LOL-induced overexpression of LOX-1 resulted in epithelial-mesenchymal transformation and promoted cancer cell invasion and migration by reducing the expression of epithelial markers (such as cadherin and platelet globin) and increasing the expression of mesenchymal

markers (like vimentin, N-cadherin, snails, slugs, MMP-2, and MMP-9) [\(165](#page-14-0)).

CD36 is another Ox-LDL receptor expressed in various cell types, including monocyte macrophages, microvascular endothelial cells, dendritic cells, and tumor cells. CD36 primarily regulates cellular lipid metabolism but also mediates lipid uptake, immune recognition, inflammation, molecular adhesion, and apoptosis ([166,](#page-14-0) [167\)](#page-14-0). CD36-mediated Ox-LDL uptake causes CD8+ T cell lipid peroxidation, which inhibits IFN- $\gamma$  and TNF production via p38 kinase activation, favoring cancer cell proliferation [\(166\)](#page-14-0). Ox-LDL increase the association of CD36 with JAK2, resulting in increased bladder cancer dryness by promoting JAK2 phosphorylation and activating the STAT3 signaling cascade [\(168\)](#page-14-0). Neurite outgrowth inhibitor-B (Nogo-B) expression is directly upregulated by CD36 mediated Ox-LDL uptake. Nogo-B interacts with autophagy-related 5 (ATG5) to promote autophagy. These process leads to lysophosphatidic acid-enhanced yes-associated protein carcinogenic activity ([169](#page-14-0)). Another significant mechanism through which Ox-LDL promotes cancer progression is autophagy. Research indicates that Ox-LDL can activate the critical metabolic enzyme OPLINe oxidase, increasing autophagy in cancer cells via pathways involving Ox-LDL and peroxisome proliferator-activated receptor [\(137](#page-14-0)).

In conclusion, LDL and Ox-LDL signaling pathways contribute to cancer progression in both normolipidemic and hyperlipidemic individuals. However, the effects are significantly more aggressive in those with hyperlipidemia. Elevated levels of LDL and Ox-LDL in hyperlipidemic individuals lead to increased cancer cell proliferation, invasion, and resistance to therapy. This makes hyperlipidemia a critical factor in cancer development. Therefore, lowering LDL and Ox-LDL levels could be a promising therapeutic strategy for preventing and managing cancer, particularly in individuals with T2DM.

### 5.4 The HDL signal pathways

HDL, often referred to as "good" cholesterol, plays a critical role in maintaining lipid homeostasis and has been shown to exert antiinflammatory, anti-oxidative, and anti-tumor effects in the TME [\(170](#page-14-0)– [172](#page-14-0)). In individuals with normal lipid levels, HDL functions optimally. It facilitates reverse cholesterol transport [\(173](#page-15-0)), reducing oxidative stress, and mitigating inflammation. Together, these actions contribute to a reduced risk of cancer progression [\(171](#page-14-0), [172\)](#page-14-0). However, in T2DM patients, HDL undergoes structural and compositional changes, resulting in dysfunctional lipoproteins ([174,](#page-15-0) [175](#page-15-0)). Elevated cholesteryl ester transfer protein (CETP) activity and reduced lecithin cholesterol acyltransferase (LCAT) activity lead to altered in HDL particle size and composition, resulting in dysfunctional HDL. This dysfunction is marked by a decrease in key proteins such as apolipoprotein A-I and paraoxonase 1, along with an increase in triglycerides and serum amyloid A (SAA) [\(176,](#page-15-0) [177\)](#page-15-0). These alterations negatively impact HDL's ability to counteract oxidative stress and inflammation, crucial mechanisms that normally accelerate cancer development.

The accumulation of SAA in HDL particles enhances its binding to Toll-like receptor 2 (TLR2), triggering the NF-kB signaling pathway, which promotes production of proinflammatory cytokines. This pro-inflammatory state creates a conducive environment for cancer initiation and progression, contrasting with the protective anti-inflammatory role of HDL in individuals with normal lipid profiles [\(178](#page-15-0), [179\)](#page-15-0). Additionally, lower levels of apolipoprotein M in HDL lead to elevated free sphingosine-1-phosphate (S1P) levels, which further exacerbate tumor growth, migration, and angiogenesis through the AKT phosphorylation pathway ([176\)](#page-15-0). Moreover, the glycation and oxidation of HDL, particularly prevalent in T2DM patients, have been implicated in the progression of cancers such as breast cancer ([180\)](#page-15-0). Oxidized and glycated HDL affects MDA-MB-231 breast cancer cells through the AKT/ERK signaling pathway, increasing integrin expression and enhancing cancer cell growth, migration, and invasion [\(181](#page-15-0)). Additionally, through enhancing lipid internalization and cholesterol intake, SR-BI overexpression in cancer cells encourages cell growth and proliferation [\(182](#page-15-0), [183\)](#page-15-0).

In conclusion, HDL generally protects against cancer in individuals with normal lipid levels. However, in hyperlipidemic conditions, its functionality is compromised, leading to a higher risk of cancer development. These differences in HDL signaling pathways highlight the importance of managing lipid levels to reduce cancer risks, especially in T2DM patients.

### 6 Other factors and cancer progression

The connection between T2DM and cancer is significantly influenced by the gut microbiome, a rapidly emerging area of research. The gut microbiota, consisting 500–1000 species and  $10^{14}$  bacteria—ten times more abundant than human cells—acts as a complex endocrine organ. It plays vital roles in digestion, nutrient absorption, and reinforcement of the intestinal immune system. Additionally, it maintains the intestinal mucosal barrier, preventing harmful substance infiltration, synthesizes beneficial compounds like vitamins, regulates host metabolism, and reduces cancer risk while enhancing anti-tumor responses [\(184](#page-15-0)–[190\)](#page-15-0).

Studies have demonstrated a strong connection between T2DM and the gut microbiota. Research by Larsen in 2010 revealed significant changes in gut microbial composition between individuals with and without T2DM [\(191\)](#page-15-0). Ma's subsequent summary highlighted a reduction in beneficial bacteria and an increase in harmful and potentially pathogenic bacteria in T2DM patients [\(184\)](#page-15-0). Dysbiosis, or an imbalance in the gut microbiota, has been associated with various illnesses, including diabetes, metabolic syndrome, non-alcoholic fatty liver disease, and even mental disorders such as depression and multiple sclerosis [\(192](#page-15-0)). Moreover, abnormal gut microbiota changes have been linked to a higher risk of several cancers, including colorectal cancer [\(193\)](#page-15-0), hepatocellular carcinoma [\(194](#page-15-0)), non-small cell lung cancer [\(195\)](#page-15-0), and prostate cancer [\(196\)](#page-15-0).

The gut microbiota plays a crucial role in human health and disease development. It influences immune cell activity and cancer risk through its remarkable metabolic abilities ([197\)](#page-15-0). Through the metabolic byproducts it produces, the gut microbiota may be able to indirectly cause the development of cancer in distant organs such as the pancreas, liver, breast, lung, prostate, and stomach ([198\)](#page-15-0). The primary metabolic products of the gut microbiota are short-chain fatty acids (SCFAs). Historically, SCFAs have been primarily studied for their anti-inflammatory properties. Recent research, however, has uncovered a unique carcinogenic pathway called the "gut-prostate axis", where SCFAs produced by specific bacteria (e.g., Rikenellaceae, Alistipes, and Lachnospira) promote PCa growth ([199](#page-15-0)). Additionally, the gut microbiota indirectly impacts PCa progression through other metabolic products such as testosterone, estrogen, folate, and phenylacetylglutamine [\(196\)](#page-15-0).

The gut microbiota also influences tumor immune responses through various mechanisms, including stimulating regulatory T cell proliferation, inducing IgA expression, regulating antimicrobial peptides and systemic inflammation, and affecting bacterial translocation [\(196](#page-15-0)). The intestinal mucosal barrier, composed of tightly bound epithelial cells, usually separates the gut microbiota from immune cells. However, specific changes in gut microbial composition can stimulate the mucosal immune system, leading to chronic inflammation and mucosal damage. This imbalance in intestinal mucosal immunity may cause cellular and DNA damage, genetic mutations, activation of tumor-associated signaling pathways, and ultimately contribute to tumor development and progression [\(197](#page-15-0), [200\)](#page-15-0).

The relationship between gut microbiota, T2DM, and cancer is complex and remains an active area of research. While current data suggest a strong connection, there is insufficient evidence to definitively conclude that gut microbiota dysbiosis directly causes cancer in T2DM patients or to fully elucidate the specific mechanisms involved. Therefore, further research is necessary to better understand these interactions and underlying mechanisms.

# 7 Conclusion

In normal individuals, cancer development is typically driven by genetic factors, environmental exposures (such as smoking or radiation), and chronic inflammation. These individuals usually maintain balanced metabolic states, including normal blood sugar, insulin, and lipid levels. However, in T2DM patients, cancer progression is accelerated due to hyperglycemia, hyperinsulinemia, and hyperlipidemia. Hyperglycemia increases oxidative stress and chronic inflammation, leading to DNA damage and promoting the Warburg effect, which fuels cancer cell metabolism. Hyperinsulinemia activates critical signaling pathways, such as PI3K/AKT, mTOR, and Ras/MAPK, which drive cell proliferation, survival, and resistance to apoptosis. Additionally, hyperinsulinemia lowers SHBG levels, increasing the availability of sex hormones like estrogen, thereby heightening the risk of hormone-dependent cancers. Hyperlipidemia contributes by altering lipid metabolism, influencing cellular signaling within the tumor microenvironment, and promoting angiogenesis and immune suppression, all of which support tumor growth and metastasis.

These metabolic abnormalities do not act in isolation but are interrelated, creating a pro-tumorigenic environment through synergistic interactions. Hyperinsulinemia and hyperglycemia, often exacerbated by T2DM, induce insulin resistance and elevate <span id="page-11-0"></span>IGF-1 levels, further activating the PI3K/AKT pathway and supporting tumor cell proliferation. Hyperglycemia and hyperlipidemia provide essential metabolic substrates that fuel cancer cell glycolysis and lipid synthesis, accelerating tumor growth. Moreover, the chronic inflammation and oxidative stress associated with these conditions activate pro-inflammatory pathways like NF- $\kappa$ B and transcription factors such as HIF-1 $\alpha$ , enhancing tumor angiogenesis, metastasis, and overall progression.

In summary, the metabolic abnormalities in T2DM significantly accelerate cancer progression. Understanding the complex interplay between these metabolic states and cancer is crucial for developing targeted interventions. Future research should focus on elucidating the precise molecular mechanisms underlying these interactions and exploring the potential of insulin sensitizers, metabolic modulators, and anti-inflammatory agents as therapeutic strategies. Personalized approaches that integrate metabolic management with conventional cancer therapies may offer improved treatment efficacy and outcomes for patients with T2DM-related malignancies. Continued investigation into these areas could pave the way for novel therapies that address both metabolic disorders and cancer progression.

### Author contributions

YZ: Writing – original draft, Visualization, Conceptualization. YL: Writing – review & editing, Visualization. CX: Writing – review & editing, Visualization. SL: Writing – review & editing, Visualization. ZG: Writing – review & editing, Funding acquisition. KQ: Writing – review & editing, Supervision, Funding acquisition, Conceptualization.

### Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This work was supported by the National Natural Science Foundation of China (Grant No. 12372304).

### Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

### Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

### References

1. Kasera H, Shekhawat RS, Yadav P, Singh P. Gene expression profiling and protein– protein network analysis revealed prognostic hub biomarkers linking cancer risk in type 2 diabetic patients. Sci Rep. (2023) 13:22605. doi: [10.1038/s41598-023-49715-9](https://doi.org/10.1038/s41598-023-49715-9)

2. Ahmad E, Lim S, Lamptey R, Webb DR, Davies MJ. Type 2 diabetes. Lancet. (2022) 400:1803–20. doi: [10.1016/S0140-6736\(22\)01655-5](https://doi.org/10.1016/S0140-6736(22)01655-5)

3. Joshi S, Liu M, Turner N. Diabetes and its link with cancer: providing the fuel and spark to launch an aggressive growth regime. BioMed Res Int. (2015) 2015:e390863. doi: [10.1155/2015/390863](https://doi.org/10.1155/2015/390863)

4. Chatterjee S, Khunti K, Davies MJ. Type 2 diabetes. Lancet. (2017) 389:2239–51. doi: [10.1016/S0140-6736\(17\)30058-2](https://doi.org/10.1016/S0140-6736(17)30058-2)

5. Supabphol S, Seubwai W, Wongkham S, Saengboonmee C. High glucose: an emerging association between diabetes mellitus and cancer progression. J Mol Med. (2021) 99:1175–93. doi: [10.1007/s00109-021-02096-w](https://doi.org/10.1007/s00109-021-02096-w)

6. Qiu J, Zheng Q, Meng X. Hyperglycemia and chemoresistance in breast cancer: from cellular mechanisms to treatment response. Front Oncol. (2021) 11:628359. doi: [10.3389/fonc.2021.628359](https://doi.org/10.3389/fonc.2021.628359)

7. Ramteke P, Deb A, Shepal V, Bhat MK. Hyperglycemia associated metabolic and molecular alterations in cancer risk, progression, treatment, and mortality. Cancers. (2019) 11:1402. doi: [10.3390/cancers11091402](https://doi.org/10.3390/cancers11091402)

8. Li W, Zhang X, Sang H, Zhou Y, Shang C, Wang Y, et al. Effects of hyperglycemia on the progression of tumor diseases. J Exp Clin Cancer Res. (2019) 38:327. doi: [10.1186/s13046-019-1309-6](https://doi.org/10.1186/s13046-019-1309-6)

9. Sun X-F, Shao Y-B, Liu M-G, Chen Q, Liu Z-J, Xu B, et al. High-concentration glucose enhances invasion in invasive ductal breast carcinoma by promoting Glut1/MMP2/MMP9 axis expression. Oncol Lett. (2017) 13:2989–95. doi: [10.3892/ol.2017.5843](https://doi.org/10.3892/ol.2017.5843)

10. Saengboonmee C, Seubwai W, Pairojkul C, Wongkham S. High glucose enhances progression of cholangiocarcinoma cells via STAT3 activation. Sci Rep. (2016) 6:18995. doi: [10.1038/srep18995](https://doi.org/10.1038/srep18995)

11. Karamian M, Moossavi M, Hemmati M. From diabetes to renal aging: the therapeutic potential of adiponectin. J Physiol Biochem. (2021) 77:205–14. doi: [10.1007/](https://doi.org/10.1007/s13105-021-00790-4) [s13105-021-00790-4](https://doi.org/10.1007/s13105-021-00790-4)

12. Zhang AMY, Wellberg EA, Kopp JL, Johnson JD. Hyperinsulinemia in obesity, inflammation, and cancer. Diabetes Metab J. (2021) 45:285–311. doi: [10.4093/](https://doi.org/10.4093/dmj.2020.0250) [dmj.2020.0250](https://doi.org/10.4093/dmj.2020.0250)

13. Arcidiacono B, Iiritano S, Nocera A, Possidente K, Nevolo MT, Ventura V, et al. Insulin resistance and cancer risk: an overview of the pathogenetic mechanisms. Exp Diabetes Res. (2012) 2012:789174. doi: [10.1155/2012/789174](https://doi.org/10.1155/2012/789174)

14. Nuovo J. Hyperlipidemia in patients with type 2 diabetes. afp. (1999) 59:1666–71.

15. Xing P, Liao Z, Ren Z, Zhao J, Song F, Wang G, et al. Roles of low-density lipoprotein receptor-related protein 1 in tumors. Chin J Cancer. (2016) 35:6. doi: [10.1186/s40880-015-0064-0](https://doi.org/10.1186/s40880-015-0064-0)

16. Dong S, Yu J, Chen X, Shen K. Association of serum lipid levels and clinical outcomes in early breast cancer patients. Ther Adv Med Oncol. (2023) 15:175883592311770. doi: [10.1177/17588359231177004](https://doi.org/10.1177/17588359231177004)

17. Shlomai G, Neel B, LeRoith D, Gallagher EJ. Type 2 diabetes mellitus and cancer: the role of pharmacotherapy. JCO. (2016) 34:4261–9. doi: [10.1200/JCO.2016.67.4044](https://doi.org/10.1200/JCO.2016.67.4044)

18. Waghela BN, Vaidya FU, Ranjan K, Chhipa AS, Tiwari BS, Pathak C. AGE-RAGE synergy influences programmed cell death signaling to promote cancer. Mol Cell Biochem. (2021) 476:585–98. doi: [10.1007/s11010-020-03928-y](https://doi.org/10.1007/s11010-020-03928-y)

19. Muthyalaiah YS, Jonnalagadda B, John CM, Arockiasamy S. Impact of Advanced Glycation End products (AGEs) and its receptor (RAGE) on cancer metabolic signaling pathways and its progression. Glycoconj J. (2021) 38:717–34. doi: [10.1007/s10719-021-](https://doi.org/10.1007/s10719-021-10031-x)  $10031 - x$ 

20. Ruze R, Song J, Yin X, Chen Y, Xu R, Wang C, et al. Mechanisms of obesity- and diabetes mellitus-related pancreatic carcinogenesis: a comprehensive and systematic review. Sig Transduct Target Ther. (2023) 8:1–41. doi: [10.1038/s41392-023-01376-w](https://doi.org/10.1038/s41392-023-01376-w)

21. Li Z, Long T, Wang R, Feng Y, Hu H, Xu Y, et al. Plasma metals and cancer incidence in patients with type 2 diabetes. Sci Total Environ. (2021) 758:143616. doi: [10.1016/j.scitotenv.2020.143616](https://doi.org/10.1016/j.scitotenv.2020.143616)

22. Han L, Ma Q, Li J, Liu H, Li W, Ma G, et al. High glucose promotes pancreatic cancer cell proliferation via the induction of EGF expression and transactivation of EGFR. PLoS One. (2011) 6:e27074. doi: [10.1371/journal.pone.0027074](https://doi.org/10.1371/journal.pone.0027074)

<span id="page-12-0"></span>23. Kang X, Kong F, Wu X, Ren Y, Wu S, Wu K, et al. High glucose promotes tumor invasion and increases metastasis-associated protein expression in human lung epithelial cells by upregulating heme oxygenase-1 via reactive oxygen species or the TGF-b1/PI3K/akt signaling pathway. Cell Physiol Biochem. (2015) 35:1008–22. doi: [10.1159/000373928](https://doi.org/10.1159/000373928)

24. Cai W, Ma Y, Song L, Cao N, Gao J, Zhou S, et al. IGF-1R down regulates the sensitivity of hepatocellular carcinoma to sorafenib through the PI3K / akt and RAS / raf / ERK signaling pathways. BMC Cancer. (2023) 23:87. doi: [10.1186/s12885-023-](https://doi.org/10.1186/s12885-023-10561-7) [10561-7](https://doi.org/10.1186/s12885-023-10561-7)

25. Hua H, Kong Q, Yin J, Zhang J, Jiang Y. Insulin-like growth factor receptor signaling in tumorigenesis and drug resistance: a challenge for cancer therapy. J Hematol Oncol. (2020) 13:64. doi: [10.1186/s13045-020-00904-3](https://doi.org/10.1186/s13045-020-00904-3)

26. Siasos G, Tousoulis D, Oikonomou E, Zaromitidou M, Stefanadis C, G. Papavassiliou A. Inflammatory markers in hyperlipidemia: from experimental models to clinical practice. CPD. (2011) 17:4132–46. doi: [10.2174/138161211798764780](https://doi.org/10.2174/138161211798764780)

27. Papapanagiotou A, Siasos G, Kassi E, Gargalionis A, Papavassiliou A. Novel inflammatory markers in hyperlipidemia: clinical implications. CMC. (2015) 22:2727– 43. doi: [10.2174/0929867322666150520095008](https://doi.org/10.2174/0929867322666150520095008)

28. Yin X, Xu R, Song J, Ruze R, Chen Y, Wang C, et al. Lipid metabolism in pancreatic cancer: emerging roles and potential targets. Cancer Commun. (2022) 42:1234–56. doi: [10.1002/cac2.12360](https://doi.org/10.1002/cac2.12360)

29. Mirmiran P, Bahadoran Z, Ghasemi A, Hosseinpanah F. Type 2 diabetes and cancer: an overview of epidemiological evidence and potential mechanisms. CRO. (2019) 24:223–33. doi: [10.1615/CritRevOncog.2019031153](https://doi.org/10.1615/CritRevOncog.2019031153)

30. Yang W, Sui J, Zhao L, Ma Y, Tabung FK, Simon TG, et al. Association of inflammatory and insulinemic potential of diet and lifestyle with risk of hepatocellular carcinoma. Cancer Epidemiology Biomarkers Prev. (2021) 30:789–96. doi: [10.1158/](https://doi.org/10.1158/1055-9965.EPI-20-1329) [1055-9965.EPI-20-1329](https://doi.org/10.1158/1055-9965.EPI-20-1329)

31. Jin Q, Shi N, Lee DH, Rexrode KM, Manson JE, Balasubramanian R, et al. Hyperinsulinemic and pro-inflammatory dietary patterns and metabolomic profiles are associated with increased risk of total and site-specific cancers among postmenopausal women. Cancers. (2023) 15:1756. doi: [10.3390/cancers15061756](https://doi.org/10.3390/cancers15061756)

32. Aroke D, Folefac E, Shi N, Jin Q, Clinton SK, Tabung FK. Inflammatory and insulinemic dietary patterns: influence on circulating biomarkers and prostate cancer risk. Cancer Prev Res. (2020) 13:841–52. doi: [10.1158/1940-6207.CAPR-20-0236](https://doi.org/10.1158/1940-6207.CAPR-20-0236)

33. Fu BC, Tabung FK, Pernar CH, Wang W, Gonzalez-Feliciano AG, Chowdhury-Paulino IM, et al. Insulinemic and inflammatory dietary patterns and risk of prostate cancer. Eur Urol. (2021) 79:405–12. doi: [10.1016/j.eururo.2020.12.030](https://doi.org/10.1016/j.eururo.2020.12.030)

34. Mokhtari E, Jamshidi S, Daftari G, Farhadnejad H, Teymoori F, Momeni SA, et al. The relationship between the insulinemic potential of diet and lifestyle and risk of breast cancer: a case-control study among Iranian adult women. Arch Public Health. (2023) 81:1–9. doi: [10.1186/s13690-022-01016-9](https://doi.org/10.1186/s13690-022-01016-9)

35. Ruggieri L, Moretti A, Berardi R, Cona MS, Dalu D, Villa C, et al. Host-related factors in the interplay among inflammation, immunity and dormancy in breast cancer recurrence and prognosis: an overview for clinicians. Int J Mol Sci. (2023) 24:4974. doi: [10.3390/ijms24054974](https://doi.org/10.3390/ijms24054974)

36. Langlais CS, Graff RE, Van Blarigan EL, Kenfield SA, Neuhaus J, Tabung FK, et al. Postdiagnostic inflammatory, hyperinsulinemic, and insulin-resistant diets and lifestyles and the risk of prostate cancer progression and mortality. Cancer Epidemiology Biomarkers Prev. (2022) 31:1760–8. doi: [10.1158/1055-9965.EPI-22-0147](https://doi.org/10.1158/1055-9965.EPI-22-0147)

37. Imai K, Takai K, Miwa T, Maeda T, Hanai T, Shiraki M, et al. Increased visceral adipose tissue and hyperinsulinemia raise the risk for recurrence of non-B non-C hepatocellular carcinoma after curative treatment. Cancers. (2021) 13:1542. doi: [10.3390/cancers13071542](https://doi.org/10.3390/cancers13071542)

38. Cheng E, Zhang S, Ou F-S, Mullen B, Ng K, Saltz LB, et al. The diet of higher insulinemic potential is not associated with worse survival in patients with stage III colon cancer (Alliance). Cancer Epidemiology Biomarkers Prev. (2020) 29:1692–5. doi: [10.1158/1055-9965.EPI-19-1454](https://doi.org/10.1158/1055-9965.EPI-19-1454)

39. Kira S, Ito C, Fujikawa R, Misumi M. Increased cancer mortality among Japanese individuals with hyperinsulinemia. Metab Open. (2020) 7:100048. doi: [10.1016/](https://doi.org/10.1016/j.metop.2020.100048) [j.metop.2020.100048](https://doi.org/10.1016/j.metop.2020.100048)

40. Tabung FK, Noonan A, Lee DH, Song M, Clinton SK, Spakowicz D, et al. Postdiagnosis dietary insulinemic potential and survival outcomes among colorectal cancer patients. BMC Cancer. (2020) 20:817. doi: [10.1186/s12885-020-07288-0](https://doi.org/10.1186/s12885-020-07288-0)

41. Perseghin G, Calori G, Lattuada G, Ragogna F, Dugnani E, Garancini MP, et al. Insulin resistance/hyperinsulinemia and cancer mortality: the Cremona study at the 15th year of follow-up. Acta Diabetol. (2012) 49:421–8. doi: [10.1007/s00592-011-0361-2](https://doi.org/10.1007/s00592-011-0361-2)

42. Balkau B, Kahn HS, Courbon D, Eschwège E, Ducimetière P. Hyperinsulinemia Predicts Fatal Liver Cancer but Is Inversely Associated With Fatal Cancer at Some Other Sites: The Paris Prospective Study. Diabetes Care. (2001) 24:843–9. doi: [10.2337/](https://doi.org/10.2337/diacare.24.5.843) [diacare.24.5.843](https://doi.org/10.2337/diacare.24.5.843)

43. Tsujimoto T, Kajio H, Sugiyama T. Association between hyperinsulinemia and increased risk of cancer death in nonobese and obese people: A population-based observational study. Int J Cancer. (2017) 141:102–11. doi: [10.1002/ijc.30729](https://doi.org/10.1002/ijc.30729)

44. Wan Y, Tabung FK, Lee DH, Fung TT, Willett WC, Giovannucci EL. Dietary insulinemic potential and risk of total and cause-specific mortality in the nurses' Health study and the health professionals follow-up study. Diabetes Care. (2021) 45:451–9. doi: [10.2337/dc21-1530](https://doi.org/10.2337/dc21-1530)

45. Sieri S, Muti P, Claudia A, Berrino F, Pala V, Grioni S, et al. Prospective study on the role of glucose metabolism in breast cancer occurrence. Int  $\overline{J}$  Cancer. (2012) 130:921–9. doi: [10.1002/ijc.26071](https://doi.org/10.1002/ijc.26071)

46. Cheng H-C, Chang T-K, Su W-C, Tsai H-L, Wang J-Y. Narrative review of the influence of diabetes mellitus and hyperglycemia on colorectal cancer risk and oncological outcomes. Trans Oncol. (2021) 14:101089. doi: [10.1016/](https://doi.org/10.1016/j.tranon.2021.101089) [j.tranon.2021.101089](https://doi.org/10.1016/j.tranon.2021.101089)

47. Hammer M, Storey S, Hershey DS, Brady VJ, Davis E, Mandolfo N, et al. Hyperglycemia and cancer: A state-of-the-science review. Oncol Nurs Forum. (2019) 46:459–72. doi: [10.1188/19.ONF.459-472](https://doi.org/10.1188/19.ONF.459-472)

48. Hoy AJ, Nagarajan SR, Butler LM. Tumour fatty acid metabolism in the context of therapy resistance and obesity. Nat Rev Cancer. (2021) 21:753–66. doi: [10.1038/](https://doi.org/10.1038/s41568-021-00388-4) [s41568-021-00388-4](https://doi.org/10.1038/s41568-021-00388-4)

49. Beloribi-Djefaflia S, Vasseur S, Guillaumond F. Lipid metabolic reprogramming in cancer cells. Oncogenesis. (2016) 5:e189–9. doi: [10.1038/oncsis.2015.49](https://doi.org/10.1038/oncsis.2015.49)

50. Islam D, Islam MS, Jesmin. Association of hypertension, hyperlipidemia, obesity, and demographic risk factors with breast cancer in Bangladeshi women. Medicine. (2022) 101:e31698. doi: [10.1097/MD.0000000000031698](https://doi.org/10.1097/MD.0000000000031698)

51. Zhao J, Tian Y, Yao J, Gu H, Zhang R, Wang H, et al. Hypercholesterolemia is an associated factor for risk of differentiated thyroid cancer in chinese population. Front Oncol. (2021) 10:508126. doi: [10.3389/fonc.2020.508126](https://doi.org/10.3389/fonc.2020.508126)

52. Han HH, Sung JS, Song DW, Park CK, Cho NH, Choi YD, et al. Abstract 2270: Hyperlipidemia promotes aggressive variant prostate cancer via RNA-binding protein Quaking. Cancer Res. (2022) 82:2270. doi: [10.1158/1538-7445.AM2022-2270](https://doi.org/10.1158/1538-7445.AM2022-2270)

53. Zhang D, Xi Y, Feng Y. Ovarian cancer risk in relation to blood lipid levels and hyperlipidemia: a systematic review and meta-analysis of observational epidemiologic studies. Eur J Cancer Prev. (2021) 30:161. doi: [10.1097/CEJ.0000000000000597](https://doi.org/10.1097/CEJ.0000000000000597)

54. Ding X, Zhang W, Li S, Yang H. The role of cholesterol metabolism in cancer. Am J Cancer Res. (2019) 9:219–27.

55. Iso H, Ikeda A, Inoue M, Sato S, Tsugane S. Serum cholesterol levels in relation to the incidence of cancer: The JPHC study cohorts. Int J Cancer. (2009) 125:2679–86. doi: [10.1002/ijc.24668](https://doi.org/10.1002/ijc.24668)

56. Ravnskov U, McCully KS, Rosch PJ. The statin-low cholesterol-cancer conundrum. QJM: Int J Med. (2012) 105:383–8. doi: [10.1093/qjmed/hcr243](https://doi.org/10.1093/qjmed/hcr243)

57. Heiniger HJ, Brunner KT, Cerottini JC. Cholesterol is a critical cellular component for T-lymphocyte cytotoxicity. Proc Natl Acad Sci USA. (1978) 75:5683– 7. doi: [10.1073/pnas.75.11.5683](https://doi.org/10.1073/pnas.75.11.5683)

58. Calleros L, Lasa M, Toro MJ, Chiloeches A. Low cell cholesterol levels increase NFkB activity through a p38 MAPK-dependent mechanism. Cell Signalling. (2006) 18:2292–301. doi: [10.1016/j.cellsig.2006.05.012](https://doi.org/10.1016/j.cellsig.2006.05.012)

59. Hsu M-T, Wang Y-K, Tseng YJ. Exosomal proteins and lipids as potential biomarkers for lung cancer diagnosis, prognosis, and treatment. Cancers. (2022) 14:732. doi: [10.3390/cancers14030732](https://doi.org/10.3390/cancers14030732)

60. Fruman DA, Chiu H, Hopkins BD, Bagrodia S, Cantley LC, Abraham RT. The PI3K pathway in human disease. Cell. (2017) 170:605–35. doi: [10.1016/](https://doi.org/10.1016/j.cell.2017.07.029) [j.cell.2017.07.029](https://doi.org/10.1016/j.cell.2017.07.029)

61. Draznin B, Miles P, Kruszynska Y, Olefsky J, Friedman J, Golovchenko I, et al. Effects of insulin on prenylation as a mechanism of potentially detrimental influence of hyperinsulinemia. Endocrinology. (2000) 141:1310–6. doi: [10.1210/endo.141.4.7411](https://doi.org/10.1210/endo.141.4.7411)

62. Sarkar PL, Lee W, Williams ED, Lubik AA, Stylianou N, Shokoohmand A, et al. Insulin enhances migration and invasion in prostate cancer cells by up-regulation of FOXC2. Front Endocrinol (Lausanne). (2019) 10:481. doi: [10.3389/fendo.2019.00481](https://doi.org/10.3389/fendo.2019.00481)

63. Wang G, Yin L, Peng Y, Gao Y, Gao H, Zhang J, et al. Insulin promotes invasion and migration of KRASG12D mutant HPNE cells by upregulating MMP-2 gelatinolytic activity via ERK- and PI3K-dependent signalling. Cell Proliferation. (2019) 52:e12575. doi: [10.1111/cpr.12575](https://doi.org/10.1111/cpr.12575)

64. Hopkins BD, Goncalves MD, Cantley LC. Insulin–PI3K signalling: an evolutionarily insulated metabolic driver of cancer. Nat Rev Endocrinol. (2020) 16:276–83. doi: [10.1038/s41574-020-0329-9](https://doi.org/10.1038/s41574-020-0329-9)

65. Poloz Y, Stambolic V. Obesity and cancer, a case for insulin signaling. Cell Death Dis. (2015) 6:e2037–7. doi: [10.1038/cddis.2015.381](https://doi.org/10.1038/cddis.2015.381)

66. Wu K, Chen H, Fu Y, Cao X, Yu C. Insulin promotes the proliferation and migration of pancreatic cancer cells by up-regulating the expression of PLK1 through the PI3K/AKT pathway. Biochem Biophys Res Commun. (2023) 648:21–7. doi: [10.1016/](https://doi.org/10.1016/j.bbrc.2023.01.061) [j.bbrc.2023.01.061](https://doi.org/10.1016/j.bbrc.2023.01.061)

67. Kumar M, Bansal N. Implications of phosphoinositide 3-kinase-akt (PI3K-akt) pathway in the pathogenesis of alzheimer's disease. Mol Neurobiol. (2022) 59:354–85. doi: [10.1007/s12035-021-02611-7](https://doi.org/10.1007/s12035-021-02611-7)

68. Ray A, Alalem M, Ray BK. Insulin signaling network in cancer. Indian J Biochem Biophys. (2014) 51:493–8.

69. Mu N, Zhu Y, Wang Y, Zhang H, Xue F. Insulin resistance: A significant risk factor of endometrial cancer. Gynecologic Oncol. (2012) 125:751–7. doi: [10.1016/](https://doi.org/10.1016/j.ygyno.2012.03.032) [j.ygyno.2012.03.032](https://doi.org/10.1016/j.ygyno.2012.03.032)

70. Bock G, Goode J eds. Biology of IGF-1: Its Interaction with Insulin in Health and Malignant States: Novartis Foundation Symposium. 1st ed. Chichester: Wiley (2004). p. 262. doi: [10.1002/0470869976](https://doi.org/10.1002/0470869976)

<span id="page-13-0"></span>71. Leung K-C, Doyle N, Ballesteros M, Waters MJ, Ho KKY. Insulin regulation of human hepatic growth hormone receptors: divergent effects on biosynthesis and surface translocation1. J Clin Endocrinol Metab. (2000) 85:4712–20. doi: [10.1210/](https://doi.org/10.1210/jcem.85.12.7017) [jcem.85.12.7017](https://doi.org/10.1210/jcem.85.12.7017)

72. Calle EE, Kaaks R. Overweight, obesity and cancer: epidemiological evidence and proposed mechanisms. Nat Rev Cancer. (2004) 4:579–91. doi: [10.1038/nrc1408](https://doi.org/10.1038/nrc1408)

73. Kaaks R, Lukanova A. Energy balance and cancer: the role of insulin and insulinlike growth factor-I. Proc Nutr Soc. (2001) 60:91–106. doi: [10.1079/PNS200070](https://doi.org/10.1079/PNS200070)

74. Makari-Judson G, Viskochil R, Katz D, Barham R, Mertens WC. Insulin resistance and weight gain in women treated for early stage breast cancer. Breast Cancer Res Treat. (2022) 194:423–31. doi: [10.1007/s10549-022-06624-1](https://doi.org/10.1007/s10549-022-06624-1)

75. Lesire L, Leroux F, Deprez-Poulain R, Deprez B. Insulin-degrading enzyme, an under-estimated potential target to treat cancer? Cells. (2022) 11:1228. doi: [10.3390/](https://doi.org/10.3390/cells11071228) [cells11071228](https://doi.org/10.3390/cells11071228)

76. Denk D, Greten FR. Inflammation: the incubator of the tumor microenvironment. Trends Cancer. (2022) 8:901–14. doi: [10.1016/j.trecan.2022.07.002](https://doi.org/10.1016/j.trecan.2022.07.002) 77. Murray PJ. Macrophage polarization. Annu Rev Physiol. (2017) 79:541–66.

doi: [10.1146/annurev-physiol-022516-034339](https://doi.org/10.1146/annurev-physiol-022516-034339)

78. Wang J, Long R, Han Y. The role of exosomes in the tumour microenvironment on macrophage polarisation. Biochim Biophys Acta (BBA) - Rev Cancer. (2022) 1877:188811. doi: [10.1016/j.bbcan.2022.188811](https://doi.org/10.1016/j.bbcan.2022.188811)

79. Ghanbari Movahed Z, Rastegari-Pouyani M, Mohammadi MH, Mansouri K. Cancer cells change their glucose metabolism to overcome increased ROS: One step from cancer cell to cancer stem cell? Biomedicine Pharmacotherapy. (2019) 112:108690. doi: [10.1016/j.biopha.2019.108690](https://doi.org/10.1016/j.biopha.2019.108690)

80. Yang Y, Wu Y, Sun X-D, Zhang Y. Reactive oxygen species, glucose metabolism, and lipid metabolism. In: Huang C, Zhang Y, editors. Oxidative Stress: Human Diseases and Medicine. Springer, Singapore (2021). p. 213–35. doi: [10.1007/978-981-16-0522-2\\_9](https://doi.org/10.1007/978-981-16-0522-2_9)

81. Zhang Z, Huang Q, Zhao D, Lian F, Li X, Qi W. The impact of oxidative stressinduced mitochondrial dysfunction on diabetic microvascular complications. Front Endocrinol (Lausanne). (2023) 14:1112363. doi: [10.3389/fendo.2023.1112363](https://doi.org/10.3389/fendo.2023.1112363)

82. Choi S-W, Benzie IFF, Ma S-W, Strain JJ, Hannigan BM. Acute hyperglycemia and oxidative stress: Direct cause and effect? Free Radical Biol Med. (2008) 44:1217–31. doi: [10.1016/j.freeradbiomed.2007.12.005](https://doi.org/10.1016/j.freeradbiomed.2007.12.005)

83. Kang D-H. Oxidative stress, DNA damage, and breast cancer. AACN Clin Issues. (2002) 13:540–9. doi: [10.1097/00044067-200211000-00007](https://doi.org/10.1097/00044067-200211000-00007)

84. Arfin S, Jha NK, Jha SK, Kesari KK, Ruokolainen J, Roychoudhury S, et al. Oxidative stress in cancer cell metabolism. Antioxidants (Basel). (2021) 10:642. doi: [10.3390/antiox10050642](https://doi.org/10.3390/antiox10050642)

85. Pothiwala P, Jain SK, Yaturu S. Metabolic syndrome and cancer. Metab Syndrome Related Disord. (2009) 7:279–88. doi: [10.1089/met.2008.0065](https://doi.org/10.1089/met.2008.0065)

86. Esposito K, Nappo F, Marfella R, Giugliano G, Giugliano F, Ciotola M, et al. Inflammatory cytokine concentrations are acutely increased by hyperglycemia in humans. Circulation. (2002) 106:2067–72. doi: [10.1161/01.CIR.0000034509.14906.AE](https://doi.org/10.1161/01.CIR.0000034509.14906.AE)

87. Park HS, Chun JN, Jung HY, Choi C, Bae YS. Role of NADPH oxidase 4 in lipopolysaccharide-induced proinflammatory responses by human aortic endothelial cells. Cardiovasc Res. (2006) 72:447–55. doi: [10.1016/j.cardiores.2006.09.012](https://doi.org/10.1016/j.cardiores.2006.09.012)

88. Matough FA, Budin SB, Hamid ZA, Alwahaibi N, Mohamed J. The role of oxidative stress and antioxidants in diabetic complications. Sultan Qaboos Univ Med J. (2012) 12:5–18. doi: [10.12816/0003082](https://doi.org/10.12816/0003082)

89. Jeong H-S, Lee D-H, Kim S-H, Lee C-H, Shin HM, Kim H-R, et al. Hyperglycemia-induced oxidative stress promotes tumor metastasis by upregulating vWF expression in endothelial cells through the transcription factor GATA1. Oncogene. (2022) 41:1634–46. doi: [10.1038/s41388-022-02207-y](https://doi.org/10.1038/s41388-022-02207-y)

90. Giacco F, Brownlee M, Schmidt AM. Oxidative stress and diabetic complications. Circ Res. (2010) 107:1058–70. doi: [10.1161/CIRCRESAHA.110.223545](https://doi.org/10.1161/CIRCRESAHA.110.223545)

91. Alhmoud JF, Woolley JF, Al Moustafa A-E, Malki MI. DNA damage/repair management in cancers. Cancers (Basel). (2020) 12:1050. doi: [10.3390/cancers12041050](https://doi.org/10.3390/cancers12041050)

92. Warburg O. On the origin of cancer cells. Science. (1956) 123:309–14. doi: [10.1126/science.123.3191.309](https://doi.org/10.1126/science.123.3191.309)

93. Vander Heiden MG, Cantley LC, Thompson CB. Understanding the warburg effect: the metabolic requirements of cell proliferation. Science. (2009) 324:1029–33. doi: [10.1126/science.1160809](https://doi.org/10.1126/science.1160809)

94. Kim J, Dang CV. Cancer's molecular sweet tooth and the warburg effect. Cancer Res. (2006) 66:8927–30. doi: [10.1158/0008-5472.CAN-06-1501](https://doi.org/10.1158/0008-5472.CAN-06-1501)

95. Reczek CR, Chandel NS. ROS promotes cancer cell survival through calcium signaling. Cancer Cell. (2018) 33:949–51. doi: [10.1016/j.ccell.2018.05.010](https://doi.org/10.1016/j.ccell.2018.05.010)

96. Cheung EC, Vousden KH. The role of ROS in tumour development and progression. Nat Rev Cancer. (2022) 22:280–97. doi: [10.1038/s41568-021-00435-0](https://doi.org/10.1038/s41568-021-00435-0)

97. Liberti MV, Locasale JW. The warburg effect: how does it benefit cancer cells? Trends Biochem Sci. (2016) 41:211–8. doi: [10.1016/j.tibs.2015.12.001](https://doi.org/10.1016/j.tibs.2015.12.001)

98. Santos JM, Hussain F. Higher glucose enhances breast cancer cell aggressiveness. Nutr Cancer. (2020) 72:734–46. doi: [10.1080/01635581.2019.1654527](https://doi.org/10.1080/01635581.2019.1654527)

99. Xu X, Chen B, Zhu S, Zhang J, He X, Cao G, et al. Hyperglycemia promotes Snail-induced epithelial–mesenchymal transition of gastric cancer via activating ENO1 expression. Cancer Cell Int. (2019) 19:344. doi: [10.1186/s12935-019-1075-8](https://doi.org/10.1186/s12935-019-1075-8)

100. Liu Z, Jia X, Duan Y, Xiao H, Sundqvist K-G, Permert J, et al. Excess glucose induces hypoxia-inducible factor-1 $\alpha$  in pancreatic cancer cells and stimulates glucose metabolism and cell migration. Cancer Biol Ther. (2013) 14:428–35. doi: [10.4161/](https://doi.org/10.4161/cbt.23786) cht.23786

101. Cheng L, Qin T, Ma J, Duan W, Xu Q, Li X, et al. Hypoxia-inducible factor-1a Mediates hyperglycemia-induced pancreatic cancer glycolysis. Anti-Cancer Agents Medicinal Chem. (2019) 19:1503–12. doi: [10.2174/1871520619666190626120359](https://doi.org/10.2174/1871520619666190626120359)

102. Goswami KK, Banerjee S, Bose A, Baral R. Lactic acid in alternative polarization and function of macrophages in tumor microenvironment. Hum Immunol. (2022) 83:409–17. doi: [10.1016/j.humimm.2022.02.007](https://doi.org/10.1016/j.humimm.2022.02.007)

103. Watson MJ, Vignali PDA, Mullett SJ, Overacre-Delgoffe AE, Peralta RM, Grebinoski S, et al. Metabolic support of tumour-infiltrating regulatory T cells by lactic acid. Nature. (2021) 591:645–51. doi: [10.1038/s41586-020-03045-2](https://doi.org/10.1038/s41586-020-03045-2)

104. Landskron G, de la Fuente M, Thuwajit P, Thuwajit C, Hermoso MA. Chronic inflammation and cytokines in the tumor microenvironment. J Immunol Res. (2014) 2014:e149185. doi: [10.1155/2014/149185](https://doi.org/10.1155/2014/149185)

105. Chang S-C, Yang W-CV. Hyperglycemia, tumorigenesis, and chronic inflammation. Crit Rev Oncology/Hematology. (2016) 108:146–53. doi: [10.1016/](https://doi.org/10.1016/j.critrevonc.2016.11.003) [j.critrevonc.2016.11.003](https://doi.org/10.1016/j.critrevonc.2016.11.003)

106. Forrester SJ, Kikuchi DS, Hernandes MS, Xu Q, Griendling KK. Reactive oxygen species in metabolic and inflammatory signaling. Circ Res. (2018) 122:877–902. doi: [10.1161/CIRCRESAHA.117.311401](https://doi.org/10.1161/CIRCRESAHA.117.311401)

107. Hempel A, Maasch C, Heintze U, Lindschau C, Dietz R, Luft FC, et al. High glucose concentrations increase endothelial cell permeability via activation of protein kinase Ca. Circ Res. (1997) 81:363–71. doi: [10.1161/01.RES.81.3.363](https://doi.org/10.1161/01.RES.81.3.363)

108. Geraldes P, King GL. Activation of protein kinase C isoforms and its impact on diabetic complications. Circ Res. (2010) 106:1319–31. doi: [10.1161/](https://doi.org/10.1161/CIRCRESAHA.110.217117) [CIRCRESAHA.110.217117](https://doi.org/10.1161/CIRCRESAHA.110.217117)

109. Wang H, Xu Y, Xu A, Wang X, Cheng L, Lee S, et al. PKC $\beta$ /NF-KB pathway in diabetic atrial remodeling. J Physiol Biochem. (2020) 76:637–53. doi: [10.1007/s13105-](https://doi.org/10.1007/s13105-020-00769-7) [020-00769-7](https://doi.org/10.1007/s13105-020-00769-7)

110. Ellulu MS, Patimah I, Khaza'ai H, Rahmat A, Abed Y. Obesity and inflammation: the linking mechanism and the complications. Arch Med Sci. (2017) 13:851–63. doi: [10.5114/aoms.2016.58928](https://doi.org/10.5114/aoms.2016.58928)

111. Chen X, Cubillos-Ruiz JR. Endoplasmic reticulum stress signals in the tumour and its microenvironment. Nat Rev Cancer. (2021) 21:71–88. doi: [10.1038/s41568-020-](https://doi.org/10.1038/s41568-020-00312-2) [00312-2](https://doi.org/10.1038/s41568-020-00312-2)

112. Hotamisligil GS. Endoplasmic reticulum stress and the inflammatory basis of metabolic disease. Cell. (2010) 140:900–17. doi: [10.1016/j.cell.2010.02.034](https://doi.org/10.1016/j.cell.2010.02.034)

113. Boden G. Effects of free fatty acids (FFA) on glucose metabolism: significance for insulin resistance and type 2 diabetes. Exp Clin Endocrinol Diabetes. (2003) 111:121–4. doi: [10.1055/s-2003-39781](https://doi.org/10.1055/s-2003-39781)

114. Shomali N, Mahmoudi J, Mahmoodpoor A, Zamiri RE, Akbari M, Xu H, et al. Harmful effects of high amounts of glucose on the immune system: An updated review. Biotechnol Appl Biochem. (2021) 68:404–10. doi: [10.1002/bab.1938](https://doi.org/10.1002/bab.1938)

115. Duan Q, Li H, Gao C, Zhao H, Wu S, Wu H, et al. High glucose promotes pancreatic cancer cells to escape from immune surveillance via AMPK-Bmi1-GATA2-MICA/B pathway. J Exp Clin Cancer Res. (2019) 38:192. doi: [10.1186/s13046-019-1209-9](https://doi.org/10.1186/s13046-019-1209-9)

116. Moschos SJ, Mantzoros CS. The role of the IGF system in cancer: from basic to clinical studies and clinical applications. Oncology. (2002) 63:317–32. doi: [10.1159/](https://doi.org/10.1159/000066230) [000066230](https://doi.org/10.1159/000066230)

117. Lopez R, Arumugam A, Joseph R, Monga K, Boopalan T, Agullo P, et al. Hyperglycemia Enhances the Proliferation of Non-Tumorigenic and Malignant Mammary Epithelial Cells through Increased leptin/IGF1R Signaling and Activation of AKT/mTOR. PLoS One. (2013) 8:e79708. doi: [10.1371/journal.pone.0079708](https://doi.org/10.1371/journal.pone.0079708)

118. Li Z, Miard S, Laplante M, Sonenberg N, Picard F. Insulin stimulates IGFBP-2 expression in 3T3-L1 adipocytes through the PI3K/mTOR pathway. Mol Cell Endocrinol. (2012) 358:63–8. doi: [10.1016/j.mce.2012.02.022](https://doi.org/10.1016/j.mce.2012.02.022)

119. Martin JL, Baxter RC. Expression of insulin-like growth factor binding protein-2 by MCF-7 breast cancer cells is regulated through the phosphatidylinositol 3-kinase/ AKT/mammalian target of rapamycin pathway. Endocrinology. (2007) 148:2532–41. doi: [10.1210/en.2006-1335](https://doi.org/10.1210/en.2006-1335)

120. Wei L-F, Weng X-F, Huang X-C, Peng Y-H, Guo H-P, Xu Y-W. IGFBP2 in cancer: Pathological role and clinical significance (Review). Oncol Rep. (2021) 45:427– 38. doi: [10.3892/or.2020.7892](https://doi.org/10.3892/or.2020.7892)

121. Ma Y, Cui D, Zhang Y, Han C, Wei W. Insulin-like growth factor binding protein-2 promotes proliferation and predicts poor prognosis in hepatocellular carcinoma. Onco Targets Ther. (2020) 13:5083–92. doi: [10.2147/OTT.S249527](https://doi.org/10.2147/OTT.S249527)

122. Hoeflich A, Russo VC. Physiology and pathophysiology of IGFBP-1 and IGFBP-2 – Consensus and dissent on metabolic control and Malignant potential. Best Pract Res Clin Endocrinol Metab. (2015) 29:685–700. doi: [10.1016/](https://doi.org/10.1016/j.beem.2015.07.002) [j.beem.2015.07.002](https://doi.org/10.1016/j.beem.2015.07.002)

123. Kang R, Tang D, Schapiro NE, Livesey KM, Farkas A, Loughran P, et al. The receptor for advanced glycation end products (RAGE) sustains autophagy and limits apoptosis, promoting pancreatic tumor cell survival. Cell Death Differ. (2010) 17:666-76. doi: [10.1038/cdd.2009.149](https://doi.org/10.1038/cdd.2009.149)

124. Mukherjee TK, Malik P, Hoidal JR. Receptor for advanced glycation end products (RAGE) and its polymorphic variants as predictive diagnostic and

<span id="page-14-0"></span>prognostic markers of NSCLCs: a perspective. Curr Oncol Rep. (2021) 23:12. doi: [10.1007/s11912-020-00992-x](https://doi.org/10.1007/s11912-020-00992-x)

125. Kuniyasu H, Oue N, Wakikawa A, Shigeishi H, Matsutani N, Kuraoka K, et al. Expression of receptors for advanced glycation end-products (RAGE) is closely associated with the invasive and metastatic activity of gastric cancer. J Pathol. (2002) 196:163–70. doi: [10.1002/path.1031](https://doi.org/10.1002/path.1031)

126. Kwak T, Drews-Elger K, Ergonul A, Miller PC, Braley A, Hwang GH, et al. Targeting of RAGE-ligand signaling impairs breast cancer cell invasion and metastasis. Oncogene. (2017) 36:1559–72. doi: [10.1038/onc.2016.324](https://doi.org/10.1038/onc.2016.324)

127. El-Far AH, Sroga G, Al Jaouni SK, Mousa SA. Role and mechanisms of RAGEligand complexes and RAGE-inhibitors in cancer progression. Int J Mol Sci. (2020) 21:3613. doi: [10.3390/ijms21103613](https://doi.org/10.3390/ijms21103613)

128. Bao J-M, He M-Y, Liu Y-W, Lu Y-J, Hong Y-Q, Luo H-H, et al. AGE/RAGE/ Akt pathway contributes to prostate cancer cell proliferation by promoting Rb phosphorylation and degradation. Am J Cancer Res. (2015) 5:1741–50.

129. Ko S-Y, Ko H-A, Shieh T-M, Chang W-C, Chen H-I, Chang S-S, et al. Cell migration is regulated by AGE-RAGE interaction in human oral cancer cells in vitro. PLoS One. (2014) 9:e110542. doi: [10.1371/journal.pone.0110542](https://doi.org/10.1371/journal.pone.0110542)

130. Deng R, Mo F, Chang B, Zhang Q, Ran H, Yang S, et al. Glucose-derived AGEs enhance human gastric cancer metastasis through RAGE/ERK/Sp1/MMP2 cascade. Oncotarget. (2017) 8:104216–26. doi: [10.18632/oncotarget.22185](https://doi.org/10.18632/oncotarget.22185)

131. Okamoto T, Yamagishi S, Inagaki Y, Amano S, Koga K, Abe R, et al. Angiogenesis induced by advanced glycation end products and its prevention by cerivastatin. FASEB J. (2002) 16:1928–30. doi: [10.1096/fj.02-0030fje](https://doi.org/10.1096/fj.02-0030fje)

132. Rojas A, Schneider I, Lindner C, Gonzalez I, Morales MA. The RAGE/ multiligand axis: a new actor in tumor biology. Bioscience Rep. (2022) 42: BSR20220395. doi: [10.1042/BSR20220395](https://doi.org/10.1042/BSR20220395)

133. Lange-Sperandio B, Sperandio M, Nawroth P, Bierhaus A. RAGE signaling in cell adhesion and inflammation. Curr Pediatr Rev. (2007) 3:1–9. doi: [10.2174/](https://doi.org/10.2174/157339607779941598) [157339607779941598](https://doi.org/10.2174/157339607779941598)

134. Chen X, Zhang L, Zhang IY, Liang J, Wang H, Ouyang M, et al. RAGE expression in tumor-associated macrophages promotes angiogenesis in glioma. Cancer Res. (2014) 74:7285–97. doi: [10.1158/0008-5472.CAN-14-1240](https://doi.org/10.1158/0008-5472.CAN-14-1240)

135. Santolla MF, Talia M, Cirillo F, Scordamaglia D, De Rosis S, Spinelli A, et al. The AGEs/RAGE transduction signaling prompts IL-8/CXCR1/2-mediated interaction between cancer-associated fibroblasts (CAFs) and breast cancer cells. Cells. (2022) 11:2402. doi: [10.3390/cells11152402](https://doi.org/10.3390/cells11152402)

136. Li R, Song Y, Zhou L, Li W, Zhu X. Downregulation of RAGE inhibits cell proliferation and induces apoptosis via regulation of PI3K/AKT pathway in cervical squamous cell carcinoma. OTT. (2020) 13:2385-97. doi: [10.2147/OTT.S240378](https://doi.org/10.2147/OTT.S240378)

137. Deng C-F, Zhu N, Zhao T-J, Li H-F, Gu J, Liao D-F, et al. Involvement of LDL and ox-LDL in cancer development and its therapeutical potential. Front Oncol. (2022) 12:803473. doi: [10.3389/fonc.2022.803473](https://doi.org/10.3389/fonc.2022.803473)

138. Cedó L, Reddy ST, Mato E, Blanco-Vaca F, Escolà-Gil JC. HDL and LDL: potential new players in breast cancer development. J Clin Med. (2019) 8:853. doi: [10.3390/jcm8060853](https://doi.org/10.3390/jcm8060853)

139. Ganjali S, Ricciuti B, Pirro M, Butler AE, Atkin SL, Banach M, et al. Highdensity lipoprotein components and functionality in cancer: state-of-the-art. Trends Endocrinol Metab. (2019) 30:12–24. doi: [10.1016/j.tem.2018.10.004](https://doi.org/10.1016/j.tem.2018.10.004)

140. Zhuang L, Kim J, Adam RM, Solomon KR, Freeman MR. Cholesterol targeting alters lipid raft composition and cell survival in prostate cancer cells and xenografts. J Clin Invest. (2005) 115:959–68. doi: [10.1172/JCI19935](https://doi.org/10.1172/JCI19935)

141. Tosi MR, Tugnoli V. Cholesteryl esters in Malignancy. Clinica Chimica Acta. (2005) 359:27–45. doi: [10.1016/j.cccn.2005.04.003](https://doi.org/10.1016/j.cccn.2005.04.003)

142. Rodrigues Dos Santos C, Domingues G, Matias I, Matos J, Fonseca I, De Almeida JM, et al. LDL-cholesterol signaling induces breast cancer proliferation and invasion. Lipids Health Dis. (2014) 13:16. doi: [10.1186/1476-511X-13-16](https://doi.org/10.1186/1476-511X-13-16)

143. Jung YY, Ko J-H, Um J-Y, Chinnathambi A, Alharbi SA, Sethi G, et al. LDL cholesterol promotes the proliferation of prostate and pancreatic cancer cells by activating the STAT3 pathway. J Cell Physiol. (2021) 236:5253–64. doi: [10.1002/jcp.30229](https://doi.org/10.1002/jcp.30229)

144. Tan JTM, Prosser HCG, Vanags LZ, Monger SA, Ng MKC, Bursill CA. Highdensity lipoproteins augment hypoxia-induced angiogenesis via regulation of post-translational modulation of hypoxia-inducible factor 1a. FASEB J. (2014) 28:206–17. doi: [10.1096/fj.13-233874](https://doi.org/10.1096/fj.13-233874)

145. Nakayama K, Frew IJ, Hagensen M, Skals M, Habelhah H, Bhoumik A, et al. Siah2 regulates stability of prolyl-hydroxylases, controls  $HIF1\alpha$  Abundance, and modulates physiological responses to hypoxia. Cell. (2004) 117:941–52. doi: [10.1016/](https://doi.org/10.1016/j.cell.2004.06.001) [j.cell.2004.06.001](https://doi.org/10.1016/j.cell.2004.06.001)

146. Zhao T-J, Zhu N, Shi Y-N, Wang Y-X, Zhang C-J, Deng C-F, et al. Targeting HDL in tumor microenvironment: New hope for cancer therapy. J Cell Physiol. (2021) 236:7853–73. doi: [10.1002/jcp.30412](https://doi.org/10.1002/jcp.30412)

147. Ma X, Bi E, Lu Y, Su P, Huang C, Liu L, et al. Cholesterol induces CD8+ T cell exhaustion in the tumor microenvironment. Cell Metab. (2019) 30:143–156.e5. doi: [10.1016/j.cmet.2019.04.002](https://doi.org/10.1016/j.cmet.2019.04.002)

148. Xu H, Zhou S, Tang Q, Xia H, Bi F. Cholesterol metabolism: New functions and therapeutic approaches in cancer. Biochim Biophys Acta Rev Cancer. (2020) 1874:188394. doi: [10.1016/j.bbcan.2020.188394](https://doi.org/10.1016/j.bbcan.2020.188394)

149. Guan X, Liu Z, Zhao Z, Zhang X, Tao S, Yuan B, et al. Emerging roles of lowdensity lipoprotein in the development and treatment of breast cancer. Lipids Health Dis. (2019) 18:137. doi: [10.1186/s12944-019-1075-7](https://doi.org/10.1186/s12944-019-1075-7)

150. Zanoni P, Velagapudi S, Yalcinkaya M, Rohrer L, Von Eckardstein A. Endocytosis of lipoproteins. Atherosclerosis. (2018) 275:273–95. doi: [10.1016/](https://doi.org/10.1016/j.atherosclerosis.2018.06.881) [j.atherosclerosis.2018.06.881](https://doi.org/10.1016/j.atherosclerosis.2018.06.881)

151. Zhang Y, Ma KL, Ruan XZ, Liu BC. Dysregulation of the low-density lipoprotein receptor pathway is involved in lipid disorder-mediated organ injury. Int J Biol Sci. (2016) 12:569–79. doi: [10.7150/ijbs.14027](https://doi.org/10.7150/ijbs.14027)

152. Siemianowicz K, Gminski J, Stajszczyk M, Wojakowski W, Goss M, Machalski M, et al. Serum LDL cholesterol concentration and lipoprotein electrophoresis pattern in patients with small cell lung cancer. Int J Mol Med. (2000) 5:55–7. doi: [10.3892/](https://doi.org/10.3892/ijmm.5.1.55) [ijmm.5.1.55](https://doi.org/10.3892/ijmm.5.1.55)

153. Rani V, Deep G, Singh RK, Palle K, Yadav UCS. Oxidative stress and metabolic disorders: Pathogenesis and therapeutic strategies. Life Sci. (2016) 148:183–93. doi: [10.1016/j.lfs.2016.02.002](https://doi.org/10.1016/j.lfs.2016.02.002)

154. Ma J-J, Jiang L, Tong D-Y, Ren Y-N, Sheng M-F, Liu H-C. CXCL13 inhibition induce the apoptosis of MDA-MB-231 breast cancer cells through blocking CXCR5/ ERK signaling pathway. Eur Rev Med Pharmacol Sci. (2018) 22:8755–62. doi: [10.26355/](https://doi.org/10.26355/eurrev_201812_16641) [eurrev\\_201812\\_16641](https://doi.org/10.26355/eurrev_201812_16641)

155. Filippi A, Picot T, Aanei CM, Nagy P, Szöllősi J, Campos L, et al. Epigallocatechin-3-O-gallate alleviates the Malignant phenotype in A-431 epidermoid and SK-BR-3 breast cancer cell lines. Int J Food Sci Nutr. (2018) 69:584– 97. doi: [10.1080/09637486.2017.1401980](https://doi.org/10.1080/09637486.2017.1401980)

156. Wang J, Zhang C, Chen K, Tang H, Tang J, Song C, et al. ERb1 inversely correlates with PTEN/PI3K/AKT pathway and predicts a favorable prognosis in triplenegative breast cancer. Breast Cancer Res Treat. (2015) 152:255–69. doi: [10.1007/](https://doi.org/10.1007/s10549-015-3467-3) [s10549-015-3467-3](https://doi.org/10.1007/s10549-015-3467-3)

157. Varghese E, Samuel S, Varghese S, Cheema S, Mamtani R, Büsselberg D. Triptolide decreases cell proliferation and induces cell death in triple negative MDA-MB-231 breast cancer cells. Biomolecules. (2018) 8:163. doi: [10.3390/biom8040163](https://doi.org/10.3390/biom8040163)

158. Naito S, Makhov P, Astsaturov I, Golovine K, Tulin A, Kutikov A, et al. LDL cholesterol counteracts the antitumour effect of tyrosine kinase inhibitors against renal cell carcinoma. Br J Cancer. (2017) 116:1203–7. doi: [10.1038/bjc.2017.77](https://doi.org/10.1038/bjc.2017.77)

159. Rodrigues NV, Correia DV, Mensurado S, Nóbrega-Pereira S, deBarros A, Kyle-Cezar F, et al. Low-density lipoprotein uptake inhibits the activation and antitumor functions of human V $\gamma$ 9V $\delta$ 2 T cells. *Cancer Immunol Res*. (2018) 6:448–57. doi: [10.1158/2326-6066.CIR-17-0327](https://doi.org/10.1158/2326-6066.CIR-17-0327)

160. Bitorina AV, Oligschlaeger Y, Shiri-Sverdlov R, Theys J. Low profile high value target: The role of OxLDL in cancer. Biochim Biophys Acta (BBA) - Mol Cell Biol Lipids. (2019) 1864:158518. doi: [10.1016/j.bbalip.2019.158518](https://doi.org/10.1016/j.bbalip.2019.158518)

161. Esterbauer H, Eckl P, Ortner A. Possible mutagens derived from lipids and lipid precursors. Mutat Research/Reviews Genet Toxicol. (1990) 238:223–33. doi: [10.1016/](https://doi.org/10.1016/0165-1110(90)90014-3) [0165-1110\(90\)90014-3](https://doi.org/10.1016/0165-1110(90)90014-3)

162. Liu Y, Wang Y, Xu Q, Zhou X, Qin Z, Chen C, et al. Prognostic evaluation of microRNA-210 in various carcinomas. Med (Baltimore). (2017) 96:e8113. doi: [10.1097/](https://doi.org/10.1097/MD.0000000000008113) [MD.0000000000008113](https://doi.org/10.1097/MD.0000000000008113)

163. Chen K-C, Liao Y-C, Wang J-Y, Lin Y-C, Chen C-H, Juo S-HH. Oxidized lowdensity lipoprotein is a common risk factor for cardiovascular diseases and gastroenterological cancers via epigenomical regulation of microRNA-210. Oncotarget. (2015) 6:24105–18. doi: [10.18632/oncotarget.4152](https://doi.org/10.18632/oncotarget.4152)

164. Tsumita T, Maishi N, Annan DA-M, Towfik MA, Matsuda A, Onodera Y, et al. The oxidized-LDL/LOX-1 axis in tumor endothelial cells enhances metastasis by recruiting neutrophils and cancer cells. Int J Cancer. (2022) 151:944–56. doi: [10.1002/ijc.34134](https://doi.org/10.1002/ijc.34134)

165. González-Chavarría I, Fernandez E, Gutierrez N, González-Horta EE, Sandoval F, Cifuentes P, et al. LOX-1 activation by oxLDL triggers an epithelial mesenchymal transition and promotes tumorigenic potential in prostate cancer cells. Cancer Lett. (2018) 414:34–43. doi: [10.1016/j.canlet.2017.10.035](https://doi.org/10.1016/j.canlet.2017.10.035)

166. Liao X, Yan S, Li J, Jiang C, Huang S, Liu S, et al. CD36 and its role in regulating the tumor microenvironment. Curr Oncol. (2022) 29:8133–45. doi: [10.3390/curroncol29110642](https://doi.org/10.3390/curroncol29110642)

167. Wang J, Li Y. CD36 tango in cancer: signaling pathways and functions. Theranostics. (2019) 9:4893–908. doi: [10.7150/thno.36037](https://doi.org/10.7150/thno.36037)

168. Yang L, Sun J, Li M, Long Y, Zhang D, Guo H, et al. Oxidized low-density lipoprotein links hypercholesterolemia and bladder cancer aggressiveness by promoting cancer stemness. Cancer Res. (2021) 81:5720–32. doi: [10.1158/0008-](https://doi.org/10.1158/0008-5472.CAN-21-0646) [5472.CAN-21-0646](https://doi.org/10.1158/0008-5472.CAN-21-0646)

169. Tian Y, Yang B, Qiu W, Hao Y, Zhang Z, Yang B, et al. ER-residential Nogo-B accelerates NAFLD-associated HCC mediated by metabolic reprogramming of oxLDL lipophagy. Nat Commun. (2019) 10:3391. doi: [10.1038/s41467-019-11274-x](https://doi.org/10.1038/s41467-019-11274-x)

170. Jomard A, Osto E. High density lipoproteins: metabolism, function, and therapeutic potential. Front Cardiovasc Med. (2020) 7:39. doi: [10.3389/](https://doi.org/10.3389/fcvm.2020.00039) [fcvm.2020.00039](https://doi.org/10.3389/fcvm.2020.00039)

171. Tabet F, Rye K-A. High-density lipoproteins, inflammation and oxidative stress. Clin Sci. (2008) 116:87–98. doi: [10.1042/CS20080106](https://doi.org/10.1042/CS20080106)

172. Ossoli A, Wolska A, Remaley AT, Gomaraschi M. High-density lipoproteins: A promising tool against cancer. Biochim Biophys Acta (BBA) - Mol Cell Biol Lipids. (2022) 1867:159068. doi: [10.1016/j.bbalip.2021.159068](https://doi.org/10.1016/j.bbalip.2021.159068)

<span id="page-15-0"></span>173. Trajkovska KT, Topuzovska S. High-density lipoprotein metabolism and reverse cholesterol transport: strategies for raising HDL cholesterol. Anatol J Cardiol. (2017) 18:149–54. doi: [10.14744/AnatolJCardiol.2017.7608](https://doi.org/10.14744/AnatolJCardiol.2017.7608)

174. Pedersen KM, Çolak Y, Bojesen SE, Nordestgaard BG. Low high-density lipoprotein and increased risk of several cancers: 2 population-based cohort studies including 116,728 individuals. J Hematol Oncol. (2020) 13:129. doi: [10.1186/s13045-](https://doi.org/10.1186/s13045-020-00963-6) [020-00963-6](https://doi.org/10.1186/s13045-020-00963-6)

175. Bonilha I, Zimetti F, Zanotti I, Papotti B, Sposito AC. Dysfunctional highdensity lipoproteins in type 2 diabetes mellitus: molecular mechanisms and therapeutic implications. JCM. (2021) 10:2233. doi: [10.3390/jcm10112233](https://doi.org/10.3390/jcm10112233)

176. Ganjali S, Banach M, Pirro M, Fras Z, Sahebkar A. HDL and cancer - causality still needs to be confirmed? Update 2020. Semin Cancer Biol. (2021) 73:169–77. doi: [10.1016/j.semcancer.2020.10.007](https://doi.org/10.1016/j.semcancer.2020.10.007)

177. Mihajlovic M, Gojkovic T, Vladimirov S, Miljkovic M, Stefanovic A, Vekic J, et al. Changes in lecithin: cholesterol acyltransferase, cholesteryl ester transfer protein and paraoxonase-1 activities in patients with colorectal cancer. Clin Biochem. (2019) 63:32–8. doi: [10.1016/j.clinbiochem.2018.11.010](https://doi.org/10.1016/j.clinbiochem.2018.11.010)

178. Niu X, Yin L, Yang X, Yang Y, Gu Y, Sun Y, et al. Serum amyloid A 1 induces suppressive neutrophils through the Toll-like receptor 2–mediated signaling pathway to promote progression of breast cancer. Cancer Sci. (2022) 113:1140-53. doi: [10.1111/](https://doi.org/10.1111/cas.15287) [cas.15287](https://doi.org/10.1111/cas.15287)

179. Cheng N, He R, Tian J, Ye PP, Ye RD. Cutting Edge: TLR2 is a functional receptor for acute-phase serum amyloid A. J Immunol. (2008) 181:22–6. doi: [10.4049/](https://doi.org/10.4049/jimmunol.181.1.22) [jimmunol.181.1.22](https://doi.org/10.4049/jimmunol.181.1.22)

180. Kontush A, Chapman MJ. Why is HDL functionally deficient in type 2 diabetes? Curr Diabetes Rep. (2008) 8:51–9. doi: [10.1007/s11892-008-0010-5](https://doi.org/10.1007/s11892-008-0010-5)

181. Pan B, Ren H, He Y, Lv X, Ma Y, Li J, et al. HDL of patients with type 2 diabetes mellitus elevates the capability of promoting breast cancer metastasis. Clin Cancer Res. (2012) 18:1246–56. doi: [10.1158/1078-0432.CCR-11-0817](https://doi.org/10.1158/1078-0432.CCR-11-0817)

182. Karimi N, Karami Tehrani FS. Expression of SR-B1 receptor in breast cancer cell lines, MDAMB-468 and MCF-7: Effect on cell proliferation and apoptosis. Iran J Basic Med Sci. (2021) 24:1069–77. doi: [10.22038/ijbms.2021.56752.12674](https://doi.org/10.22038/ijbms.2021.56752.12674)

183. Wang D, Huang J, Gui T, Yang Y, Feng T, Tzvetkov NT, et al. SR-BI as a target of natural products and its significance in cancer. Semin Cancer Biol. (2022) 80:18–38. doi: [10.1016/j.semcancer.2019.12.025](https://doi.org/10.1016/j.semcancer.2019.12.025)

184. Ma Q, Li Y, Li P, Wang M, Wang J, Tang Z, et al. Research progress in the relationship between type 2 diabetes mellitus and intestinal flora. Biomedicine Pharmacotherapy. (2019) 117:109138. doi: [10.1016/j.biopha.2019.109138](https://doi.org/10.1016/j.biopha.2019.109138)

185. Yang G, Wei J, Liu P, Zhang Q, Tian Y, Hou G, et al. Role of the gut microbiota in type 2 diabetes and related diseases. Metabolism. (2021) 117:154712. doi: [10.1016/](https://doi.org/10.1016/j.metabol.2021.154712) [j.metabol.2021.154712](https://doi.org/10.1016/j.metabol.2021.154712)

186. Nicholson JK, Holmes E, Kinross J, Burcelin R, Gibson G, Jia W, et al. Host-gut microbiota metabolic interactions. Science. (2012) 336:1262–7. doi: [10.1126/](https://doi.org/10.1126/science.1223813) [science.1223813](https://doi.org/10.1126/science.1223813)

187. Hooper LV, Littman DR, Macpherson AJ. Interactions between the microbiota and the immune system. Science. (2012) 336:1268–73. doi: [10.1126/science.1223490](https://doi.org/10.1126/science.1223490)

188. O'Hara AM, Shanahan F. The gut flora as a forgotten organ. EMBO Rep. (2006) 7:688–93. doi: [10.1038/sj.embor.7400731](https://doi.org/10.1038/sj.embor.7400731)

189. Jandhyala SM, Talukdar R, Subramanyam C, Vuyyuru H, Sasikala M, Reddy DN. Role of the normal gut microbiota. World J Gastroenterol. (2015) 21:8787–803. doi: [10.3748/wjg.v21.i29.8787](https://doi.org/10.3748/wjg.v21.i29.8787)

190. Kamada N, Seo S-U, Chen GY, Núñez G. Role of the gut microbiota in immunity and inflammatory disease. Nat Rev Immunol. (2013) 13:321–35. doi: [10.1038/nri3430](https://doi.org/10.1038/nri3430)

191. Larsen N, Vogensen FK, van den Berg FW, Nielsen DS, Andreasen AS, Pedersen BK, et al. Gut microbiota in human adults with type 2 diabetes differs from non-diabetic adults. PLoS One. (2010) 5:e9085. doi: [10.1371/journal.pone.0009085](https://doi.org/10.1371/journal.pone.0009085)

192. Biedermann L, Rogler G. The intestinal microbiota: its role in health and disease. Eur J Pediatr. (2015) 174:151–67. doi: [10.1007/s00431-014-2476-2](https://doi.org/10.1007/s00431-014-2476-2)

193. Cheng Y, Ling Z, Li L. The intestinal microbiota and colorectal cancer. Front Immunol. (2020) 11:615056. doi: 10.3389/fi[mmu.2020.615056](https://doi.org/10.3389/fimmu.2020.615056)

194. Wang T, Huang S, Wu C, Wang N, Zhang R, Wang M, et al. Intestinal microbiota and liver diseases: insights into therapeutic use of traditional chinese medicine. Evidence-Based Complementary Altern Med. (2021) 2021:e6682581. doi: [10.1155/2021/6682581](https://doi.org/10.1155/2021/6682581)

195. Wang X, Hou L, Cui M, Liu J, Wang M, Xie J. The traditional Chinese medicine<br>and non-small cell lung cancer: from a gut microbiome perspective. Front Cell Infect Microbiol. (2023) 13:1151557. doi: [10.3389/fcimb.2023.1151557](https://doi.org/10.3389/fcimb.2023.1151557)

196. Zha C, Peng Z, Huang K, Tang K, Wang Q, Zhu L, et al. Potential role of gut microbiota in prostate cancer: immunity, metabolites, pathways of action? Front Oncol. (2023) 13:1196217. doi: [10.3389/fonc.2023.1196217](https://doi.org/10.3389/fonc.2023.1196217)

197. Bhatt AP, Redinbo MR, Bultman SJ. The role of the microbiome in cancer development and therapy. CA: A Cancer J Clin. (2017) 67:326–44. doi: [10.3322/](https://doi.org/10.3322/caac.21398) [caac.21398](https://doi.org/10.3322/caac.21398)

198. Picardo SL, Coburn B, Hansen AR. The microbiome and cancer for clinicians. Crit Rev Oncology/Hematology. (2019) 141:1–12. doi: [10.1016/j.critrevonc.2019.06.004](https://doi.org/10.1016/j.critrevonc.2019.06.004)

199. Matsushita M, Fujita K, Motooka D, Hatano K, Fukae S, Kawamura N, et al. The gut microbiota associated with high-Gleason prostate cancer. Cancer Sci. (2021) 112:3125–35. doi: [10.1111/cas.14998](https://doi.org/10.1111/cas.14998)

200. Marchesi JR, Dutilh BE, Hall N, Peters WHM, Roelofs R, Boleij A, et al. Towards the human colorectal cancer microbiome. PLoS One. (2011) 6:e20447. doi: [10.1371/journal.pone.0020447](https://doi.org/10.1371/journal.pone.0020447)