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Treg cells as a protective factor for Hashimoto`s thyroiditis: a Mendelian randomization study

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Background and objectives: Hashimoto's thyroiditis (HT), a chronic autoimmune disorder impacting thyroid function, is a growing public health concern. The relationship between Treg cells and HT has been extensively studied, with Treg cells considered crucial in suppressing HT progression. However, these studies have mainly been observational, limiting our understanding of Treg cells' impact on HT risk. Leveraging large datasets, we utilized Mendelian randomization (MR) analysis to examine the causal association between Treg cell biomarkers and HT, providing additional validation for these relationships.

Methods: Comprehensive two-sample Mendelian randomization analysis was performed to determine the causal association between Treg cells signatures and HT in this study. Based on publicly available genetic data, we explored causal associations between 165 Treg cells signatures and HT risk.

Results: The European cohort study has identified five Treg cell phenotypes that causally protect against HT risk. Resting Treg %CD4 (OR = 0.975, 95% CI = 0.954~0.998, P = 0.030); CD4 on resting Treg (OR = 0.938, 95% CI = 0.882~0.997, P = 0.041; CD28- CD8dim %CD8dim (OR = 0.983, 95% CI = 0.969~0.998, P = 0.030); CD25 on CD39+ resting Treg (OR = 0.926, 95% CI = 0.864~0.991, P = 0.026); 5) CD28 on activated & secreting Treg (OR = 0.969, 95% CI = 0.942~0.996, P = 0.025). The Asian cohort study has identified four Treg cell phenotypes negatively correlated with the risk of HT. CD25hi %T cell (OR = 0.635, 95% CI = 0.473~852, P = 0.002); CD4 Treg %CD4 (OR = 0.829, 95% CI = 0.687~1.000, P = 0.050); CD127-CD8br %T cell (OR = 0.463, 95% CI = 0.311~0.687, P < 0.001); CD3 on resting Treg (OR = 0.786, 95% CI = 0.621~0.994, P = 0.044).

Conclusion: Our study has demonstrated the close connection between Treg cells and HT by genetic means, thus providing foundational basis for future research.

KEYWORDS

Hashimoto's thyroiditis, Treg cells, immunity, causal inference, MR analysis

1 Introduction

Hashimoto's Thyroiditis (HT) is a common chronic autoimmune disease affecting thyroid function (1, 2), named after the Japanese scientist Hashimoto, who first described it as a type of autoimmune thyroiditis typically resulting in hypothyroidism (3). The primary pathological changes include extensive enlargement of the thyroid gland, follicular hyperplasia, and the presence of plasma cells and lymphocytes (4). The incidence of HT has been rapidly increasing worldwide in recent years (4), with rates ranging between 27 and 448 per 100,000 (5). Consequently, HT has become a critical public health issue. And studies have shown that, HT is a known risk factor for thyroid cancer development (6). Therefore, Precise diagnosis and effective treatment of HT are essential for enhancing patients' quality of life and reducing their risk of thyroid cancer (7).

Despite the ongoing debate regarding the etiology and pathogenesis of HT, there is a prevailing agreement that autoimmunity maintains a crucial role in its pathogenesis (2, 8). Recently, regulatory T cells (Treg) have emerged as crucial factors in HT. Treg cells, a CD4 + T cell population, exert immunosuppressive effects on excessive immune responses (9-11). Animal experiments (12-14) indicate that the lack of Tregs could result in the spontaneous advancement of autoimmune thyroiditis. Clinical evidence (10, 15) further substantiates that Treg frequency in circulation is reduced among HT patients. These findings suggest that a decrease in Treg numbers or dysfunction leads to impaired immune tolerance towards autoantigenic stimuli, contributing to the development of Hashimoto's thyroiditis. Consequently, Treg immunomodulation has emerged as a crucial matter in recent HT immunology research. However, conflicting findings exist in other studies. Glick et al. reported no significant difference in CD4+CD25 + Tregs frequency in HT (16). Even one study found a significantly higher proportion of CD4+Foxp3+ Tregs in autoimmune thyroid disease (AITD) patients (17). The variation may be ascribed to the dual regulatory function of the immune system and the heterogeneity induced by the multiple Treg cell subtypes (18). Therefore, investigating the association between Treg cell subtypes and HT could potentially have a crucial role in precise immunotherapy for HT.

Mendelian randomization (MR) utilizes single nucleotide polymorphisms (SNPs) as instrumental variables (IVs) to infer potential causal relationships between exposure and outcomes (19, 20). follows the Mendelian gamete random allocation principle as well as the principle of free combination. This approach effectively mitigates the impact of confounding factors and reverse causation commonly encountered in conventional observational studies (21). MR is widely used in genetic epidemiology (22). With the large datasets currently available, MR analysis may be a good tool for exploring novel biomarkers in Treg cells that have a causal impact on the risk of Hashimoto's thyroiditis. Therefore, this study employs a two-sample Mendelian randomization approach, utilizing large-scale GWAS databases to explore the causal relationship between immune cell phenotypes and HT risk, To our knowledge, there haven't been previous studies using MR to assess the causal impact of Treg cell traits on the risk of HT.

2 Materials and methods

2.1 Study design

The current study applied a two-sample MR method to analyze causal relationships between 165 Treg traits and HT. Therefore, to ensure the validity of causal inference, the IVs relies on three fundamental assumptions: 1) Genetic variants have a direct connection with the exposure; 2) Genetic variants are not associated with potential confounding factors between exposure and outcome; and 3) Genetic variants do not influence the outcome through pathways unrelated to the exposure. All MR analyses were conducted using publicly available summary statistics, thus eliminating the need for additional ethical approval or informed consent.

2.2 Data source

The GWAS summary statistics for a diverse range of Treg traits were retrieved from the GWAS Catalog (23), the GWAS Catalog Accession Numbers are detailed in Supplementary Table 1. The original GWAS for immune traits utilized data from 3,757 European individuals, these datasets encompass an extensive collection of 165 Treg traits. The GWAS summary statistics for HT were sourced from the GWAS Catalog, encompassing datasets from both European (accession number GCST90018855) (24) and Asian populations (accession number GCST90018855). The initial GWAS analysis for the European cohort involved 395,640 individuals, including 15,654 cases and 379,986 controls. Likewise, the original GWAS analysis for the Asian cohort included 173,193 participants of East Asian descent, predominantly of Japanese origin, with 537 cases and 172,656 controls.

2.3 Instrumental variable selection

Genetic variants were utilized as IVs in this study. Significant SNPs that were associated with the Treg cell phenotype with genomewide significance $(P < 1 \times 10^{-5})$ (25) were selected based on recent research. To minimize bias due to linkage disequilibrium (LD), the following conditions were imposed: $R^2 < 0.001$ and distance = 10,000 kb (26). IVs with an F<10 were excluded to ensure strong correlation of SNPs with HT risk (Supplementary Table 2). The formula for calculating F is: $F = \mathbb{R}^2 \times \frac{K-2}{1-\mathbb{R}^2}$, where K is the sample size, R^2 is the proportion of variance explained by SNPs in the exposure database (27). The formula for calculating R^2 is: $R^2 =$ $\frac{2 \times \beta \times \text{eaf} \times (1-\text{eaf})}{2 \times \beta^2 \times \text{eaf} \times (1-\text{eaf}) + 2 \times \text{se}^2 \times \text{K} \times \text{eaf} \times (1-\text{eaf})}, \text{ where eaf is the effect allele}$ frequency, β is the allele effect value, K is the sample size, and se is the Standard Error. The exposure and outcome datasets were merged, and incompatible alleles and palindromic SNPs were eliminated. The phenotypes associated with the remaining SNPs were identified utilizing the Human Genotype-Phenotype Association Database (http://www.phenoscanner.medschl.cam.ac.uk/) (28). The remaining SNPs were ultimately utilized as instrumental variables for exposure.

2.4 Statistical analysis

The univariable MR analysis was performed using the R packages "TwoSampleMR". Our analysis primarily utilized the inverse variance weighting (IVW) method (29). We employed four additional methods to assess the reliability and consistency of our results. These methods include MR-Egger regression (30), the weighted median estimator (WME) (31), mode-based simple estimation (32), and mode-based weighted estimation. All causal estimates were converted to odds ratios (ORs) for the outcome which was a dichotomous phenotype.

Heterogeneity among the individual estimates of genetic variation was evaluated using The Cochran Q test. No significant heterogeneity was observed when P > 0.05. We used the MR-Egger intercept to assess the effect of horizontal pleiotropy (33). Sensitivity analysis was conducted using the "leave-one-out" method to assess the impact of individual SNPs on the causal relationship. The Mendelian Randomization Pleiotropy RESidual Sum and Outlier (MR-PRESSO) method were utilized as indicators to evaluate and correct the level of pleiotropy (34), the MR-PRESSO was conducted using the R package "MRPRESSO" (Figure 1).

3 Results

3.1 The MR analysis of results from European cohorts

The results of the European cohorts study revealed nine Treg cell phenotypes significantly linked with HT risk. Figure 2 presents these findings. Of the identified Treg cell features, we found five that were protective against HT risk: 1) CD25 on CD39+ resting Treg (IVW OR = 0.926, 95% CI = 0.864~0.991, P = 0.026); 2) CD28- CD8dim % CD8dim (IVW OR = 0.983, 95% CI = 0.969~0.998, P = 0.030); 3) CD28 on activated & secreting Treg (IVW OR = 0.969, 95% CI = $0.942 \sim 0.996$, P = 0.025; 4)CD4 on resting Treg (IVW OR = 0.938, 95% CI = 0.882~0.997, P = 0.041); 5) Resting Treg %CD4 (IVW OR = 0.975, 95% CI = 0.954~0.998, P = 0.030), as well as four other Treg cell subtypes (CD3 on CD39+ secreting Treg, CD3 on CD4 Treg, CD3 on CD45RA+ CD4+, CD39+ activated Treg %activated Treg). Reverse MR analysis revealed a negative causal relationship between HT and CD25 on CD39+ resting Treg (IVW OR = 0.891, 95% CI = $0.798 \sim 0.996$, P = 0.042) (see Figure 3). The sensitivity test results are displayed in Table 1. Additionally, the leave-one-out analysis corroborated this finding (see Supplementary Figure 1). Causal effect estimates were depicted using scatter plots (see Supplementary Figure 2). Moreover, funnel plots, providing a visual appraisal, revealed no heterogeneity in the causal effect (see Supplementary Figure 3).

3.2 The MR analysis of results from East Asian cohorts

The study's findings in the East Asian cohorts indicate that seven Treg cell phenotypes are significantly linked to HT risk. Figure 4 demonstrates these associations. Four Treg cell features were identified as beneficial in mitigating HT risk: 1) CD127-CD8br %T cell (OR = 0.463, 95% CI = $0.311 \sim 0.687$, P < 0.001); 2) CD25hi % T cell (OR = 0.635, 95% CI = $0.473 \sim 852$, P = 0.002); 3) CD3 on resting Treg (OR = 0.786, 95% CI = $0.621 \sim 0.994$, P = 0.044); 4) CD4 Treg %CD4 (OR = 0.829, 95% CI = $0.687 \sim 1.000$, P = 0.050). And 3 other Treg cell subtypes (CD25 on CD39+activated Treg, CD39 +CD8br %CD8br, CD39+CD8br %T cell).



Randomization Pleiotropy RESidual Sum and Outlier

exposure	nsnp	method	pval	FDR		OR(95% CI)
CD25 on CD39+ resting Treg	15	Inverse variance weighted	0.026	0.03357656	•	0.926 (0.864 to 0.991)
	15	MR Egger	0.443	NA	H H H	0.950 (0.836 to 1.079)
	15	Weighted median	0.018	NA	Her.	0.889 (0.807 to 0.980
	15	Simple mode	0.181	NA		0.891 (0.759 to 1.046
	15	Weighted mode	0.134	NA	•••	0.891 (0.773 to 1.027
CD28- CD8dim %CD8dim	16	Inverse variance weighted	0.030	0.03357656	•	0.983 (0.969 to 0.998
	16	MR Egger	0.048	NA	•	0.982 (0.965 to 0.998
	16	Weighted median	0.053	NA	•	0.983 (0.967 to 1.000
	16	Simple mode	0.474	NA		0.979 (0.925 to 1.036
	16	Weighted mode	0.049	NA		0.984 (0.970 to 0.999
CD28 on activated & secreting Treg	24	Inverse variance weighted	0.025	0.03357656		0.969 (0.942 to 0.996
	24	MR Egger	0.280	NA	•	0.980 (0.946 to 1.016
	24	Weighted median	0.188	NA		0.972 (0.932 to 1.014
	24	Simple mode	0.423	NA		0.973 (0.910 to 1.040
	24	Weighted mode	0.087	NA		0.968 (0.933 to 1.003
CD3 on CD39+ secreting Treg	28	Inverse variance weighted	0.002	0.01522485		1.063 (1.023 to 1.105
	28	MR Egger	0.046	NA		1.074 (1.005 to 1.147
	28	Weighted median	0.005	NA		1 082 (1 025 to 1 143
	28	Simple mode	0 791	NA	H	1 014 (0 917 to 1 120
	28	Weighted mode	0.025	NA		1 072 (1 012 to 1 136
CD3 on CD4 Trea	10	Inverse variance weighted	0.010	0.03357656		1.075 (1.018 to 1.135
	10	MR Egger	0.215	NA		1.076 (0.967 to 1.196
	10	Weighted median	0.002	NA		1.096 (1.034 to 1.161
	10	Simple mode	0.002	NA		1.107 (0.993 to 1.233
	10	Weighted mode	0.010	NA		1.107 (0.000 to 1.200
CD2 on CD45BA+ CD4+	21	Inverse upriance weighted	0.010	0.02257656		1.041 (1.009 to 1.076
CD3 OF CD43IO(* CD4+	21	MR Engor	0.012	0.03331030		1.039 (0.003 to 1.075
	31	Weighted median	0.101	NA		1.030 (0.992 to 1.080
	31	Simple meda	0.101	NA		1.035 (0.955 to 1.000
	31	Simple mode	0.333	1924	T.	1.030 (0.900 to 1.111
CD201 estimated Taxa % estimated Taxa	31	weighted mode	0.010	0.03357656		1.049 (1.011 to 1.087
CD39+ activated heg %activated heg	8	MD Farms	0.030	0.03337636		1.081 (1.008 to 1.181
	8	MR Egger	0.195	NA		1.097 (0.969 to 1.241
	8	vveignted median	0.265	NA		1.054 (0.961 to 1.155
	8	Simple mode	0.853	NA	-	1.012 (0.898 to 1.140
	8	Weighted mode	0.685	NA	Her	1.023 (0.920 to 1.138
CD4 on resting Treg	14	Inverse variance weighted	0.041	0.04084317		0.938 (0.882 to 0.997
	14	MR Egger	0.516	NA	H ^e l	0.966 (0.872 to 1.070
	14	Weighted median	0.178	NA		0.940 (0.859 to 1.029
	14	Simple mode	0.148	NA	•	0.901 (0.788 to 1.029)
	14	Weighted mode	0.384	NA	••	0.951 (0.854 to 1.060
Resting Treg %CD4	34	Inverse variance weighted	0.030	0.03357656	•	0.975 (0.954 to 0.998
	34	MR Egger	0.041	NA	•	0.970 (0.943 to 0.998
	34	Weighted median	0.012	NA	•	0.967 (0.941 to 0.993)
	34	Simple mode	0.549	NA	+	0.984 (0.934 to 1.037)
	34	Weighted mode	0.012	NA		0.972 (0.951 to 0.992)

FIGURE 2

Forest plots showing significantly causal relationships between 9 Treg cell traits and HT (European cohorts). nsnp, number of single-nucleotide polymorphism; OR, odds ratio; CI, confidence interval.

Reverse MR analysis revealed no significant causal relationship between HT and Treg cell phenotype. The results of the sensitivity test are shown in Table 2. In the same vein, the leave-one-out test also supported this observation (Supplementary Figure 4). The scatter plots see Supplementary Figure 5, funnel plots see Supplementary Figure 6.

3.3 Cross-validation results comparing the European and East Asian cohorts

Mutual validation was performed between European groups and East Asian groups. The MR results showed a total of five results for both datasets that showed a similar propensity to protect as the preliminary analysis (IVW OR<1), but the significance did not reach statistical significance, as shown in Figure 5. Specifically included were CD25 on CD39+ resting Treg (East Asian OR = 0.759, and European OR = 0.926), CD28- CD8dim %CD8dim (East Asian OR = 0.991, and European OR = 0.983), CD28 on activated & secreting Treg (East Asian OR = 0.991, and European OR = 0.969), Resting Treg %CD4 (East Asian OR = 0.857, and European OR = 0.975), and CD4 Treg %CD4 (East Asian OR = 0.829, and European OR = 0.964). And 4 other Treg cell subtypes (CD4 on resting Treg, CD127-CD8br %T cell, CD25hi %T cell, and CD3 on resting Treg).

In conclusion, through MR analysis, we identified five treg cell characteristics of interest that were protective against the risk of HT. They include Resting Treg %CD4, CD4 Treg %CD4, CD28 on activated & secreting Treg, CD28- CD8dim %CD8dim, and CD25

	exposure	outcome	nsnp	method	pval		OR(95% CI)
	Hashimoto's thyroiditis	CD25 on CD39+ resting Treg	10	MR Egger	0.931		1.015 (0.730 to 1.412)
		CD25 on CD39+ resting Treg	10	Weighted median	0.023	101	0.847 (0.733 to 0.978)
		CD25 on CD39+ resting Treg	10	Inverse variance weighted	0.042	•	0.891 (0.798 to 0.996)
		CD25 on CD39+ resting Treg	10	Simple mode	0.117		0.791 (0.606 to 1.031)
		CD25 on CD39+ resting Treg	10	Weighted mode	0.107	⊷ →	0.794 (0.616 to 1.022)
					Ó	0.5 1 1.5 2	
JRE 3							
st plots showing reverse	MR analysis result	s (European coho	rts). nsnj	o, number of sir	ngle-ni	ucleotide	polymorphism

TABLE 1 Result of sensitivity analysis (European cohorts).

			(Cochran	s Q	Pleic	otropy	
Exposure	Outcome	Method	Q	Q_df	Q_pval	Egger intercept	SE	pval
		MR Egger	14.26	13	0.356	-0.0067	0.014	0.641
CD25 on CD39+ resting Treg	Hashimoto`s thyroiditis	IVW	14.51	14	0.412			
		MR-PRESSO						0.457
		MR Egger	18.33	14	0.192	0.0044	0.008	0.607
CD28- CD8dim %CD8dim	Hashimoto`s thyroiditis	IVW	18.70	15	0.228			
		MR-PRESSO					0.008 0.007 0.007 0.010 0.010 0.019 0.019 0.008	0.425
		MR Egger	17.89	22	0.713	-0.0081	0.007	0.298
CD28 on activated & secreting Treg	Hashimoto`s thyroiditis	IVW	19.02	23	0.700			
		MR-PRESSO						0.758
		MR Egger	31.89	26	0.197	-0.0035	0.010	0.734
CD3 on CD39+ secreting Treg	Hashimoto`s thyroiditis	IVW	32.04	27	0.231			
		MR-PRESSO						0.226
		MR Egger	11.95	8	0.153	-0.0004	0.019	0.985
CD3 on CD4 Treg	Hashimoto`s thyroiditis	IVW	11.95	9	0.216			
		MR-PRESSO						0.311
		MR Egger	39.70	29	0.089	0.0016	0.008	0.835
CD3 on CD45RA+ CD4+	Hashimoto`s thyroiditis	IVW	39.77	30	0.110			
		MR-PRESSO						0.104
		MR Egger	7.28	6	0.296	-0.0056	0.020	0.790
CD39+ activated Treg %activated Treg	Hashimoto`s thyroiditis	IVW	7.37	7	0.391			
		MR-PRESSO						0.452
		MR Egger	11.91	12	0.453	-0.0081	0.012	0.499
CD4 on resting Treg	Hashimoto`s thyroiditis	IVW	12.39	13	0.496			
		MR-PRESSO						0.527
		MR Egger	47.07	32	0.042	0.0065	0.010	0.508
Resting Treg %CD4	Hashimoto`s thyroiditis	IVW	47.73	33	0.047			
		MR-PRESSO						0.070
		MR Egger	4.74	8	0.785	-0.0221	0.027	0.435
Hashimoto`s thyroiditis	CD25 on CD39+ resting Treg	IVW	5.41	9	0.797			
		MR-PRESSO						0.708

IVW, Inverse variance weighted; MR-PRESSO, MR-Pleiotropy Residual Sum and Outlier.

on CD39+ resting Treg. These features exhibit a negative correlation with the risk of HT, suggesting a potential protective role against HT.

4 Discussion

HT is an autoimmune disease in which Treg cells play a crucial role in preventing the onset and progression by suppressing the

production of harmful autoantibodies targeting thyroglobulin (TG) and thyroid peroxidase (TPO) (35) by autoreactive B cells, thus preventing the destruction caused by these autoreactive B cells (36, 37). Furthermore, Treg cells protect the thyroid tissue from attack by the release of immunosuppressive molecules such as IL-10 and TGF- β (38). In addition to their role in immune homeostasis, Treg cells play indispensable roles in self-tolerance (39). Numerous animal experiments (12, 13, 40) have demonstrated that the absence of Tregs can trigger the spontaneous development of autoimmune

exposure	nsnp	method	pval	FDR		OR(95% CI)
CD127- CD8br %T cell	6	Inverse variance weighted	<0.001	0.000929733	⊷ ;	0.463 (0.311 to 0.687)
	6	MR Egger	0.134	NA	H O	0.145 (0.019 to 1.095)
	6	Weighted median	0.013	NA	H	0.518 (0.308 to 0.872)
	6	Simple mode	0.020	NA	H 1	0.267 (0.124 to 0.575)
	6	Weighted mode	0.106	NA		0.532 (0.284 to 0.997)
CD25 on CD39+ activated Treg	6	Inverse variance weighted	0.047	0.049882062	→	1.516 (1.005 to 2.288)
	6	MR Egger	0.152	NA	⊢ →	10.396 (0.772 to 139.96
	6	Weighted median	0.064	NA	→	1.667 (0.972 to 2.860)
	6	Simple mode	0.136	NA	•	1.847 (0.937 to 3.640)
	6	Weighted mode	0.141	NA	⊢ →	1.855 (0.929 to 3.704)
CD25hi %T cell	10	Inverse variance weighted	0.002	0.008592031	+++	0.635 (0.473 to 0.852)
	10	MR Egger	0.019	NA	++	0.193 (0.064 to 0.581)
	10	Weighted median	0.008	NA	H H -1	0.585 (0.393 to 0.872)
	10	Simple mode	0.065	NA		0.507 (0.270 to 0.955)
	10	Weighted mode	0.054	NA	H	0.527 (0.300 to 0.928)
CD3 on resting Treg	20	Inverse variance weighted	0.044	0.049882062	++	0.786 (0.621 to 0.994)
	20	MR Egger	0.818	NA		0.922 (0.466 to 1.824)
	20	Weighted median	0.265	NA		0.826 (0.591 to 1.156)
	20	Simple mode	0.403	NA		0.791 (0.463 to 1.353)
	20	Weighted mode	0.239	NA		0.731 (0.442 to 1.211)
CD39+ CD8br %CD8br	6	Inverse variance weighted	0.045	0.049882062	→ →	1.578 (1.011 to 2.463)
	6	MR Egger	0.326	NA	→ →	3.426 (0.395 to 29.684
	6	Weighted median	0.173	NA	→	1.492 (0.839 to 2.653)
	6	Simple mode	0.533	NA	·	1.308 (0.595 to 2.874)
	6	Weighted mode	0.312	NA		1.458 (0.756 to 2.811)
CD39+ CD8br %T cell	8	Inverse variance weighted	0.036	0.049882062) 	1.411 (1.023 to 1.946)
	8	MR Egger	0.471	NA	⊢ • • • • • • • • • • • • • • • • • • •	1.274 (0.688 to 2.358)
	8	Weighted median	0.313	NA	· · · · ·	1.254 (0.808 to 1.949)
	8	Simple mode	0.448	NA	→	1.262 (0.716 to 2.225)
	8	Weighted mode	0.353	NA		1.237 (0.813 to 1.882)
CD4 Treg %CD4	9	Inverse variance weighted	0.050	0.049882062	H H	0.829 (0.687 to 1.000)
	9	MR Egger	0.730	NA		0.941 (0.674 to 1.313)
	9	Weighted median	0.258	NA	H	0.873 (0.689 to 1.105)
	9	Simple mode	0.539	NA		0.878 (0.591 to 1.306)
	9	Weighted mode	0.286	NA	н е ни	0.869 (0.684 to 1.106)

FIGURE 4

Forest plots showing significantly causal relationships between 7 Treg cell traits and HT (East Asian cohorts). nsnp, number of single-nucleotide polymorphism; OR, odds ratio; CI, confidence interval.

TABLE 2	Result of	sensitivity	analysis	(East	Asian	cohorts).
	nesure or	SCHSICIVICY	anacysis	(Last	Asian	conorca).

				Cochran's	; Q	Pleiotropy			
Exposure	Outcome	Method	Q	Q_df	Q_pval	Egger intercept	SE	pval	
		MR Egger	4.17	4	0.38	0.19	0.165	0.32	
CD127- CD8br %T cell	Hashimoto`s thyroiditis	IVW	5.53	5	0.35				
		MR-PRESSO						0.45	
		MR Egger	1.74	4	0.78	-0.28	0.19	0.22	
CD25 on CD39+ activated Treg	Hashimoto`s thyroiditis	IVW	3.90	5	0.56				
		MR-PRESSO						0.6	
		MR Egger	3.59	8	0.89	0.20	0.093	0.06	
CD25hi %T cell	Hashimoto`s thyroiditis	IVW	8.42	9	0.49				
		MR-PRESSO						0.54	
		MR Egger	13.95	18	0.73	-0.03	0.061	0.63	
CD3 on resting Treg	Hashimoto`s thyroiditis	IVW	14.19	19	0.77				
		MR-PRESSO						0.78	
		MR Egger	4.85	6	0.56	0.03	0.066	0.72	
CD39+ CD8br %T cell	Hashimoto`s thyroiditis	IVW	5.00	7	0.66				
		MR-PRESSO						0.71	
CD39+ CD8br %CD8br	Hashimoto`s thyroiditis	MR Egger	0.50	4	0.97	0.01	0.023	0.72	

(Continued)

TABLE 2 Continued

				Cochran's	Q	Pleiotropy			
Exposure	Outcome	Method	Q	Q_df	Q_pval	Egger intercept	SE	pval	
		IVW	1.01	5	0.96				
		MR-PRESSO						0.95	
		MR Egger	3.71	7	0.81	-0.04	0.046	0.4	
CD4 Treg %CD4	Hashimoto`s thyroiditis	IVW	4.51	8	0.81				
		MR-PRESSO						0.79	

IVW, Inverse variance weighted; MR-PRESSO, MR-Pleiotropy Residual Sum and Outlier.

thyroiditis. Meanwhile, clinical evidence (10) suggests a decrease in the frequency of circulating Tregs among individuals diagnosed with HT. However, there are also reports indicating that Treg cells may contribute to the progression of HT (16, 17). Such discrepancies may be attributed to different detection markers or subsets with distinct phenotypes. Considering the dissection of Tregs into subpopulations and limited data reported in autoimmune thyroiditis, in order to delve deeper into the potential Treg cell phenotypes associated with reduced risk of HT, we comprehensively examined the causal relationship between 165 Treg cell phenotypes and HT utilizing public genetic data and employing Mendelian randomization methods (41). MR employs genetic variants as instrumental variables, which are fixed at conception, to perform causal inferences regarding the effects of modifiable risk factors. This approach can effectively mitigate certain types of confounding (42). Notably, this study represented a pioneering analysis utilizing Mendelian randomization to scrutinize the causal association between multiple Treg cell traits and HT. In our study, five protective factors were ultimately identified.

The first trait is Resting Treg %CD4. Previous research has demonstrated that by utilizing FoxP3 and CD45RA, three distinct subpopulations can be differentiated based on both phenotypic and functional characteristics: the CD45RA + FoxP3lo resting Treg (rTreg), the CD45RA-FoxP3hi effector Treg (eTreg), and the CD45RA-FoxP3lo cells (Fraction III, FIII). Resting Treg cells have the ability to suppress autoimmune reactions (43), which prevents the immune system from launching attacks against its own tissues. Resting-phase Treg has been demonstrated to decrease in autoimmune disease, indicating a potential role for Resting Treg in mitigating autoimmune disease (44, 45). Research on resting Tregs in HT is limited; currently, only one clinical study has reported no significant differences in the proportion of resting Tregs between HT patients and healthy volunteers, and there is no direct research establishing a link between resting Treg %CD4 and HT. However, HT is considered an autoimmune disease, and our MR analysis results also indicate a negative correlation between the percentage of resting Treg %CD4 and the risk of HT, suggesting a potential protective role of resting Treg %CD4 in HT.

The second protective trait is CD4 Treg %CD4. CD4 is a membrane protein expressed extensively on the surface of T cells, which is indispensable for recognizing most antigens in the body. The CD4 co-receptor augments the T cell sensitivities to antigens by 30- to 100-fold (46), and its primary function is to bind with MHC-II (major histocompatibility complex class II) (47) molecules that are present on the surfaces of antigen-presenting cells, mainly associating with the β chain binding. This association allows CD4+ T cells to closely associate with antigen-presenting cells, which initiates T cell activation and guides their immune response (48, 49). This process is crucial for regulating the immune system's normal functionality and preventing excessive immune responses. Hence, the activation and execution of Treg cells' biological functions are integrally linked to CD4 molecules. Experimental in vivo studies have indicated a significant reduction in the proportion of CD4CD25 Treg cells in patients with HT compared to healthy controls (HCs) (50). One meta-analysis showed that newly

id.outcome	exposure	nsnp	method	pval		OR(95% CI)
GCST90018635	CD25 on CD39+ resting Treg	6	Inverse variance weighted	0.268		0.759 (0.466 to 1.236)
GCST90018855		15	Inverse variance weighted	0.026	•	0.926 (0.864 to 0.991)
GCST90018635	CD28- CD8dim %CD8dim	9	Inverse variance weighted	0.610	•	0.991 (0.959 to 1.025)
GCST90018855		16	Inverse variance weighted	0.030	•	0.983 (0.969 to 0.998)
GCST90018635	CD28 on activated & secreting Treg	13	Inverse variance weighted	0.930	H	0.991 (0.813 to 1.209)
GCST90018855		24	Inverse variance weighted	0.025	•	0.969 (0.942 to 0.996)
GCST90018635	CD4 on resting Treg	6	Inverse variance weighted	0.260	⊢ • • •	1.350 (0.801 to 2.274)
GCST90018855		14	Inverse variance weighted	0.041		0.938 (0.882 to 0.997)
GCST90018635	Resting Treg %CD4	9	Inverse variance weighted	0.435	H.	0.857 (0.581 to 1.263)
GCST90018855		34	Inverse variance weighted	0.030	•	0.975 (0.954 to 0.998)
GCST90018635	CD127- CD8br %T cell	6	Inverse variance weighted	<0.001	H H H	0.463 (0.311 to 0.687)
GCST90018855		15	Inverse variance weighted	0.387	H H H	1.053 (0.937 to 1.182)
GCST90018635	CD25hi %T cell	10	Inverse variance weighted	0.002	HH-1	0.635 (0.473 to 0.852)
GCST90018855		20	Inverse variance weighted	0.419	•	1.019 (0.973 to 1.067)
GCST90018635	CD3 on resting Treg	20	Inverse variance weighted	0.044	++-	0.786 (0.621 to 0.994)
GCST90018855		37	Inverse variance weighted	0.110	•	1.029 (0.993 to 1.066)
GCST90018635	CD4 Treg %CD4	9	Inverse variance weighted	0.050	H H -1	0.829 (0.687 to 1.000)
GCST90018855		19	Inverse variance weighted	0.074	•	0.964 (0.925 to 1.004)

FIGURE 5

The MR analysis results of mutual validation was performed between European groups and East Asian groups. nsnp, number of SNP. CI, confidence interval.

diagnosed HT patients have lower levels of Tregs compared to HCs (51). Another meta-analysis investigated the percentage of peripheral blood Tregs among CD4+ T cells in patients with AITDs. The results revealed that the proportions of Tregs among CD4+ T cells in untreated AITDs were significantly lower than those in HCs. However, unfortunately, no significant differences were observed in untreated HT patients (52). Although our findings suggest a negative correlation between CD4 Treg %CD4 and HT, further studies are warranted to establish and validate this relationship, considering the limited research on the association between CD4 Treg %CD4 and HT.

The third and fourth involve CD28 on activated & secreting Treg, and CD28- CD8dim %CD8dim. CD28 is a type I transmembrane glycoprotein receptor typically expressed on most CD4⁺ T cells (53). It serves as a co-stimulatory receptor expressed differently at specific stages of T cell differentiation and within specific T cell subgroups to regulate immune responses (54). T cell activation involves two signals. The first signal involves the T cell receptor (TCR) recognizing and binding specifically to the antigenic peptide-major histocompatibility complex (MHC) on antigen-presenting cells. The second signal results from the interaction between T cells and a range of co-stimulatory proteins expressed on the surface of antigen-presenting cells, generating co-stimulatory signals (55). It is worth noting that the most crucial interaction among these proteins is the binding of CD28 to CD80/CD86 (56). There is ongoing clinical development of a solitary CD28 agonist antibody, specifically TAB 08 (TGN 1412) (57, 58). However, the potential risks and benefits of this immunological agonist remain unclear and require further study (59). Additionally, CD28 superagonistic monoclonal antibodies (CD28SAB) have been found to preferentially activate and expand immuno-suppressive regulatory T (Treg) cells. In preclinical tests, CD28SABs have demonstrated the ability to alleviate the pathogenesis, progression, and clinical course of autoimmune diseases (60). Although our results revealed a negative association between CD28 on activated & secreting Treg and the risk of HT, which is consistent with previous findings, it is noteworthy that the CD28- CD8dim %CD8dim phenotype was also negatively associated with the risk of HT. Previous studies indicated that since CD28 was expressed not only on Treg but also on effector T cells, over-activation of effector T cells aggravated the progression of thyroid tissue injury or inflammation (61). Hyperreactivity of B7/CD28 signaling is closely linked with the development of autoimmune diseases (62). Cytotoxic T-Lymphocyte Antigen-4 (CTLA-4) is an inhibitory protein that competitively binds to B7 molecules (CD80 and CD86) in competition with CD28 (63), exerting immune co-suppressive responses (64), making it a highly regarded therapeutic target in the treatment of autoimmune diseases. It has been reported that using CTLA-4 fusion protein to block the interaction between CD28 and B7 can inhibit the autoimmune response in experimental rheumatoid arthritis (RA) (65). Additionally, inhibiting the CD80-CD28-mediated signaling pathway via specific RNAi and antibodies to block the interaction between CD28 and B7 reduced immune cell activation and reversed the damage in a pathological lupus nephritis model (66). Therefore, We believe that solely activating or inhibiting the CD28 molecule may not be ideal for improving HT.

The final parameter is CD25 on CD39+ resting Treg. The α -chain of the interleukin-2 receptor (IL-2R) is CD25, which is expressed

mainly on immune-related cells' surfaces, including activated T cells, B cells, and NK cells. It is another Treg surface marker that interacts with the ligand IL-2 and is involved in a variety of immune responses (67, 68). In 1995, Sakaguchi et al. identified a subset of CD4+ T lymphocytes expressing abundant CD25 in thymectomized mice aged 3 days and other autoimmune models, demonstrating their capacity to impede disease progression (69). Similarly, the elevated counts of CD4+ CD25 high HLA-DR+ cells in HT patients may suggest compensatory expansion of this Treg subpopulation to suppress the immune response (70). In an in vivo study (71), CD4 +CD25+Foxp3+ Tregs were revealed to play a role in the pathogenesis of experimental autoimmune thyroiditis (EAT), a murine model of HT. CD25 expression is indispensable for Tregs' differentiation and adaptive proliferation. Mice lacking IL2R α or IL2R β chains manifest severe systemic autoimmunity, a condition ameliorated by transferring CD4 + CD25 + T cells from wild-type mice (72). Furthermore, Mice with deficiencies in IL-2 or the IL-2 receptor exhibit diminished Treg generation, precipitating autoimmune disorders (73). Our findings indicate a negative correlation between HT risk and CD25 on CD39 + resting Treg, consistent with previous research. However, the reverse MR analysis revealed a significant causal relationship between HT risk and CD25 on CD39+ resting Treg, which may be related to unknown confounding factors. Our MR analysis attempted to exclude confounding factors, but there may still be unidentified factors that could impact the results.

Our study has inherent limitations. First, we cannot entirely exclude the potential influence of other confounding factors on this association. Considering the possible presence of additional variables, further exploration of the precise role of these Treg cell phenotypes in the pathogenesis of HT is warranted based on our study findings. Second, despite leveraging GWAS data from European and Asian populations, the generalizability of our findings to other ethnic groups remains subject to constraints. Third, the absence of individual-specific data precluded a more nuanced, stratified analysis of the population, introducing potential inaccuracies into our speculative conclusions. Finally, Our study reported on the Treg cell phenotypes that may decrease the risk of HT in both European and Asian populations. We conducted mutual validation in two study cohorts and applied a more lenient FDR correction for p-values. Although this effectively reduces the occurrence of false-negative results, it inevitably leads to an increased probability of type I errors.

5 Conclusions

In summary, we have established a causal association between five distinct Treg cell phenotypes and HT through an extensive twoway Mendelian randomization analysis, elucidating the intricate interplay between the immune system and HT. Furthermore, our research has effectively mitigated the impact of inevitable confounding factors, reverse causality, and other variables. This may serve as a foundational basis for researchers to delve into Treg cells as potential immunotherapeutic targets for HT.

Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material. Further inquiries can be directed to the corresponding author.

Author contributions

JG: Data curation, Methodology, Software, Visualization, Writing – original draft. GS: Investigation, Methodology, Software, Validation, Visualization, Writing – review & editing. FS: Conceptualization, Formal Analysis, Funding acquisition, Project administration, Supervision, Writing – review & editing.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fendo.2024.1347695/ full#supplementary-material

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