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[Unraveling the genetic basis of](https://www.frontiersin.org/articles/10.3389/fendo.2024.1344917/full) [the causal association between](https://www.frontiersin.org/articles/10.3389/fendo.2024.1344917/full) infl[ammatory cytokines](https://www.frontiersin.org/articles/10.3389/fendo.2024.1344917/full) [and osteonecrosis](https://www.frontiersin.org/articles/10.3389/fendo.2024.1344917/full)

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Background: Previous studies have reported that the occurrence and development of osteonecrosis is closely associated with immune-inflammatory responses. Mendelian randomization was performed to further assess the causal correlation between 41 inflammatory cytokines and osteonecrosis.

Methods: Two-sample Mendelian randomization utilized genetic variants for osteonecrosis from a large genome-wide association study (GWAS) with 606 cases and 209,575 controls of European ancestry. Another analysis included drug-induced osteonecrosis with 101 cases and 218,691 controls of European ancestry. Inflammatory cytokines were sourced from a GWAS abstract involving 8,293 healthy participants. The causal relationship between exposure and outcome was primarily explored using an inverse variance weighting approach. Multiple sensitivity analyses, including MR-Egger, weighted median, simple model, weighted model, and MR-PRESSO, were concurrently applied to bolster the final results.

Results: The results showed that bFGF, IL-2 and IL2-RA were clinically causally associated with the risk of osteonecrosis (OR=1.942, 95% CI=1.13-3.35, p=0.017; OR=0.688, 95% CI=0.50-0.94, p=0.021; OR=1.386, 95% CI=1.04-1.85, p = 0.026). there was a causal relationship between SCF and drug-related osteonecrosis (OR=3.356, 95% CI=1.09-10.30, p=0.034).

Conclusion: This pioneering Mendelian randomization study is the first to explore the causal link between osteonecrosis and 41 inflammatory cytokines. It conclusively establishes a causal association between osteonecrosis and bFGF, IL-2, and IL-2RA. These findings offer valuable insights into osteonecrosis pathogenesis, paving the way for effective clinical management. The study suggests bFGF, IL-2, and IL-2RA as potential therapeutic targets for osteonecrosis treatment.

KEYWORDS

bone, necrosis, immune, inflammation, mendelian randomization, GWAS

Introduction

Osteonecrosis, characterized by bone cell and marrow death, encompasses a group of diseases [\(1\)](#page-7-0). Triggered by factors like trauma, drugs, alcohol, or blood disorders, it induces an immune response inhibiting bone repair, leading to osteonecrosis ([2](#page-7-0)–[6\)](#page-7-0). Although osteonecrosis often occurs in weight-bearing joints, including the hip, knee, and humerus, it can occur in any bone or location, even the craniofacial area ([7,](#page-7-0) [8](#page-7-0)). It is a progressive disease that imposes a substantial burden on society, especially since it affects young people in their prime working years, and can lead to disability in important weight-bearing joints [\(9](#page-8-0), [10\)](#page-8-0).

The specific etiology and pathophysiology of osteonecrosis remain not fully understood due to its highly heterogeneous nature. Nonetheless, past research underscores the crucial role of the immune inflammatory response in its initiation and progression. Anomalies in immune responses and immune cell infiltration in osteonecrotic tissue often leads to uncontrolled inflammation. While inflammation is integral to bone tissue repair by recruiting immune cells and bone marrow mesenchymal stem cells for assistance [\(11,](#page-8-0) [12](#page-8-0)), persistent inflammation can impede repair and exacerbate failures, particularly in the case of osteonecrosis ([6,](#page-7-0) [13\)](#page-8-0). Nevertheless, we still know very little about the specific inflammatory cytokines involved in osteonecrosis development. Several observational studies in the past have attempted to explore the relationship between inflammatory cytokines and osteonecrosis. Li et al. explored potential inflammation-related biomarkers of osteonecrosis using the Gene Expression Omnibus (GEO) database [\(14\)](#page-8-0), and Zou et al. explored the relationship between TH17 and IL-17 with osteonecrosis ([15\)](#page-8-0). However, the sample sizes of these studies were usually small, resulting in low confidence in the results. In addition, their results may be affected by unforeseen confounding variables or reverse causality because confounding factors were not excluded, which also poses a challenge in establishing a clear causal relationship [\(16](#page-8-0)).

Mendelian randomization (MR) analysis methods use genetic variation in nonexperimental data to infer the causal effect of

exposure on outcome. Because alleles are randomly assigned during meiosis, MR reduces conventional confounding variables and reverse causality, thus providing better evidence for causal inference [\(17\)](#page-8-0). Twosample MR analysis allows researchers to assess instrument-exposure and instrument-outcome associations in two separate population samples, thereby improving the applicability and validity of the test. In this study, we extracted validated genetic variants for 41 inflammatory cytokines for the first time from published genomewide association study (GWAS) pooled data and innovatively used Mendelian randomization analysis to investigate the association between inflammatory cytokines and osteonecrosis, aiming to fill the gaps in the understanding of the etiology and pathophysiology of osteonecrosis, which utilizes genetic variation as an instrument for the estimation of causality variables, which can alleviate the problem of confounding variables and reverse causality and provide stronger evidence for causal inference.

Method

Data resources

The research design is illustrated in Figure 1, incorporating osteonecrosis cases derived from two distinct meta-analysis studies focused on individuals of European ancestry. The first dataset (finn-b-M13 OSTEONECROSIS) comprises 604 cases and 209,575 controls, while the second dataset (finn-b-OSTEON_DRUGS) involves 101 cases and 218,691 controls. Genetic predictors of and genetic associations with cytokines and other systemic inflammatory modulators were obtained from the most recent GWAS of 41 systemic inflammatory modulators in 8293 Finns from three multicenter studies. The three populationbased cohorts were: the Cardiovascular Risk in Young Finns Study (mean age of men: 37.4 years; mean age of women: 37.5 years) and the "FINRISK1997" study (mean age of men: 48.3 years; mean age of women: 47.3 years). FINRISK1997" study (mean age of men: 48.3 years;

correlation, independence, and exclusionary restrictions, are illustrated in this causally directed acyclic graph.

mean age of women: 47.3 years) and "FINRISK2002" study (mean age of men: 60.4 years; mean age of women: 60.1 years). The studies measured circulating inflammatory modulators using Bio-Rad's premixed Bio-Plex Pro Human Cytokine 27-plex Assay and 21-plex Assay and a Bio-Plex 200 reader with Bio-Plex 6.0 software. Only measurements within the detectable range for each cytokine were considered in the analysis; any cytokine with more than 90% missing values (48 out of 7) was excluded from the analysis. Genetic associations were adjusted for age, sex, body mass index, and the top 10 genetic principal components and corrected for genomic controls. All study participants provided informed consent [\(18\)](#page-8-0).

Selection of cytokine SNPs

Mendelian Randomization (MR) analysis relies on three fundamental assumptions: correlation, independence, and exclusion restriction ([19](#page-8-0)). The premise is that the chosen genetic variants exhibit an association with risk factors (correlation) but not with any confounding factors in the risk factor-outcome relationship (independence). Additionally, it assumes that these genetic variants are not linked to the outcome through any pathway other than the targeted risk factor (exclusion restriction). This two-way study integrates data from three Genome-Wide Association Studies (GWAS) encompassing 41 inflammatory cytokines, osteonecrosis, and druginduced osteonecrosis. The SNP selection process involved applying a genome-wide significance threshold of $p < 5 \times 10^{\text{A}}-8$ to identify SNPs strongly associated with osteonecrosis/drug-induced osteonecrosis and inflammatory cytokines. To mitigate linkage disequilibrium, SNPs were clustered with a threshold of $kb = 10,000$ and $r^2 = 0.001$. Palindromic SNPs were excluded, considering the challenges in identifying these SNPs in the exposure and outcome GWASs where systemic inflammatory regulators were aligned in the same direction. The R^2 value of each SNP was utilized to compute the proportion of variance in exposure, while the F statistic was employed to estimate instrumental strength, thereby avoiding weak instrumental bias [\(20,](#page-8-0) [21\)](#page-8-0). Finally, we will replace the unavailable SNPs in the result summary with the proxy SNPs ($r^2 > 0.8$) from LDlink (LDlink | An Interactive Web Tool for Exploring Linkage Disequilibrium in Population Groups (nih.gov)) [\(22\)](#page-8-0).

Statistical analysis

Given the varying number of SNPs associated with each cytokine, our primary MR analysis for cytokines with only one SNP employed the Wald ratio [\(23\)](#page-8-0). For those with two or more SNPs, we opted for inverse variance weighting (IVW) as the primary MR analysis [\(24](#page-8-0)). This approach aimed to investigate the potential pathogenic role of inflammatory cytokines in relation to the risk of osteonecrosis. To assess the presence of heterogeneity in our primary MR analysis (IVW), we conducted a Cochrane Q test. Most outcomes exhibited no significant heterogeneity ($p > 0.05$), though a few displayed heterogeneity. It's important to note that since IVW was our primary analysis and may inherently contain heterogeneity, the presence of heterogeneity in individual outcomes does not significantly impact the overall prediction of causality [\(25](#page-8-0)).

To further evaluate causality and explore potential pleiotropy, we implemented additional checks, including MR Egger regression and MR-PRESSO [\(26](#page-8-0)). Additionally, a Leave-one-out analysis was performed to examine the influence of individual SNPs on the overall MR analysis. To eliminate potential confounding effects, we utilized PhenoScanner to investigate potential dimorphic phenotypes associated with the evaluated individual SNPs. Much of this analytical work was carried out using R analysis software (version 4.0.3) and relevant R packages, encompassing two-sample MRs, data arrays, and others.

Result

[Supplementary Table 1](#page-7-0) provides comprehensive details about the study and dataset. Notably, all participants in the study were of European descent (100%), a deliberate choice aimed at mitigating the impact of ethnic differences on the research outcomes. This meticulous consideration ensures a homogenous participant pool and enhances the study's reliability by eliminating potential confounding factors associated with ethnic diversity.

Selection of instrumental variables

A flowchart outlining the full-text logic is provided in [Figure 1](#page-1-0), and raw data for inflammatory cytokines are available from the link (Cytokines GWAS results - Datasets - data.bris). Genetic variants were screened against screening criteria (P<5×10-8, r^2<0.001, kb=10,000) as described previously. The associations of inflammatory cytokines on osteonecrosis and and drug-induced osteonecrosis are summarized in [Supplementary Tables 2](#page-7-0)–7, including chromosomal location, gene, effector allele (EA), other allele and effector allele frequency (EAF). In addition, estimates of the association between each SNP and inflammatory cytokines and osteonecrosis are given, including beta values, standard errors (SE) and P values.

Causal relationship with osteonecrosis

Our findings suggest a potential involvement of cyclic bFGF, IL-2RA, and IL-2 in the risk of osteonecrosis development, as indicated by the IVW approach (refer to [Figure 2](#page-3-0)). Utilizing the IVW method, we observed that elevated genetic prediction levels of cyclic bFGF correlated with an increased risk of osteonecrosis (OR=1.942, 95% $CI=1.13-3.35$, $p=0.017$ per 1 standard deviation (SD)) (see [Supplementary Figure 1\)](#page-7-0). Cochran's Q test did not reveal any heterogeneity (P=0.858), and directional polymorphisms were absent (MR Egger-intercept=0.038, P=0.717 for MR Eggerintercept; P=0.858 for MR PRESSO global test).

Furthermore, heightened IL-2RA levels were associated with an elevated risk of osteonecrosis (OR=1.386, 95% CI=1.04-1.85, p=0.026), while increased IL-2 levels were linked to a decreased risk (OR=0.688, 95% CI=0.50-0.94, p=0.021, per 1 standard deviation (SD) increase). Cochran's Q test indicated no heterogeneity (p=0.523; p=0.681), and no directional polymorphism was found (MR Egger-intercept = -0.085, p =

0.135 for MR Egger-intercept, $p = 0.568$ for MR PRESSO global test; MR Egger-intercept = -0.024 , p = 0.689 for MR Egger-intercept, p = 0.736 for MR PRESSO global test) (refer to [Supplementary Tables 2](#page-7-0)–4). With the exception of bFGF, IL-2RA, and IL-2, other cytokines such as VEGF, GRO-a, Trail, MIG, IL-7, and IL-17 did not demonstrate any association with osteonecrosis risk in the IVW primary MR analysis or secondary analyses. Notably, our heterogeneity test revealed significant heterogeneity in IL-4 (p=0.015), MIG (p=0.036), and PDGFbb (p=0.042), whereas most other cytokines exhibited significant non-heterogeneity. Aside from MIP1b (p=0.045 for MR Egger-intercept), MR-Egger regression did not identify any polymorphisms in p-values for all cytokines. Our MR-PRESSO assay, serving as an additional robustness test, did not identify any outliers, except for IL-4 ($p = 0.01$) and MIG ($p < 0.04$).

Causal relationship with druginduced osteonecrosis

As previously mentioned, various factors can contribute to osteonecrosis, with steroid-induced osteonecrosis being a prominent cause of nontraumatic occurrences in nontraumatic fractures. Additionally, osteonecrosis of the jaw induced by bisphosphonates has emerged as a significant area of research interest in recent years. Despite differing mechanisms of action, these drugs share a common outcome—they induce osteonecrosis by disrupting the delicate balance between bone resorption and formation, ultimately interfering with the bone remodeling process. To enhance the robustness of our study, we delved into the causal effects of inflammatory cytokines on the risk of drug-induced osteonecrosis. Notably, bFGF with genetically predicted levels of IL-2RA exhibited a positive, though not statistically significant, causal effect on drug-induced osteonecrosis (OR=2.324, 95% CI=0.62- 8.72, p=0.211 per 1 standard deviation (SD); OR=1.053, 95% CI=0.51- 2.19, per 1 standard deviation (SD) P=0.888). Similarly, the genetic prediction level of IL-2 demonstrated a negative, albeit not statistically significant, causal effect with drug-induced osteonecrosis (OR=0.507, 95% CI=0.17-1.55, per 1 standard deviation (SD) P=0.232). This trend aligns with the earlier study results, ensuring consistency [\(Figures 3](#page-4-0), [4](#page-5-0)). Furthermore, in drug-induced osteonecrosis, a higher genetic prediction level of cyclic SCF was associated with an elevated risk of osteonecrosis according to the IVW approach (OR=3.356, 95% CI=1.09-10.30, P=0.034 per 1 standard deviation (SD))

[\(Supplementary Figure 2\)](#page-7-0). Importantly, Cochran's Q test did not reveal heterogeneity (p=0.667), and directional polymorphisms were absent (MR Egger-intercept=0.071, p=0.655 for MR Egger-intercept; p=0.673 for MR PRESSO global test) [\(Supplementary Tables 5](#page-7-0)–7).

Discussion

In our current investigation, our findings strongly indicate a causal relationship between elevated levels of bFGF/IL-2RA and an increased risk of osteonecrosis. Moreover, higher circulating levels of IL-2 appear to be suggestively associated with a reduced risk of osteonecrosis. Additionally, we identified a positive causal effect of SCF on drug-induced osteonecrosis. These results contribute novel evidence to the etiological understanding of osteonecrosis, underscoring the advantages of employing the MR approach. It is noteworthy that our study represents the inaugural application of MR analysis to ascertain whether heightened levels of inflammatory cytokines are correlated with an elevated risk of osteonecrosis. This exploration is based on genetic data extracted from publicly available databases, marking a significant contribution to the field by leveraging robust and unbiased genetic information.

The immune-inflammatory response plays a pivotal role in both the development and progression of osteonecrosis, representing a prominent feature of the condition [\(27](#page-8-0), [28](#page-8-0)). Within the necrotic bone tissue, the sustained production of inflammatory cytokines serves as a constant attractant for both innate immune cells, including macrophages, neutrophils, and dendritic cells, and adaptive immune cells like T and B cells. This creates a positive feedback loop, wherein these immune cells release additional inflammatory cytokines, thereby amplifying the overall inflammatory response [\(29](#page-8-0)–[31](#page-8-0)). Furthermore, chronic inflammation not only overstimulates bone resorption but also hinders bone formation, thereby contributing to the pathogenesis of osteonecrosis. This disruption of the normal coordination between proinflammatory activation and anti-inflammatory silencing during bone repair forms the pathophysiological basis of osteonecrosis [\(Figure 5](#page-5-0)).

Numerous studies have delved into the relationship between osteonecrosis and the immune-inflammatory response. However, the causal direction and the extent of association between inflammatory cytokines and the risk of osteonecrosis remain unclear, with observational studies constituting the primary mode of investigation. These studies are susceptible to confounding, reverse causality bias, and measurement errors, which limit their ability to provide causal estimates of exposure-outcome effects and, consequently, hinder their utility in informing prevention and treatment strategies. In contrast, MR analysis employs specific genetic variants that meet instrumental variable assumptions to systematically address causality issues in epidemiological studies, thereby mitigating the potential for inherent bias. Moreover, MR analysis is a more cost-effective and feasible approach compared to randomized controlled trials. According to our present MR analysis, bFGF in conjunction with IL-2RA may contribute to the development of osteonecrosis, while IL-2 exhibits an opposing

FIGURE 4

Causal estimation of bFGF, IL-2, IL-2RA and SCF on osteonecrosis and its drug-induced subtype. Forest plots depict the causal estimates of bFGF, IL-2, IL-2RA and SCF on osteonecrosis and its drug-induced subtypes. The dominance ratio (OR) was estimated using the fixed-effects IVW method. Horizontal bars indicate 95% confidence intervals (CI).

inflammation excessively activates bone resorption, inhibits bone formation, and drives osteonecrosis.

effect. Additionally, our findings underscore the significance of SCF as a crucial factor in promoting the onset of drug-induced osteonecrosis. These insights gleaned from MR analysis offer a valuable contribution to understanding the complex interplay between inflammatory cytokines and osteonecrosis, offering potential avenues for further research and therapeutic exploration.

IL-2 (interleukin-2) is a regulator of the immune and inflammatory response that participates in the regulation of immune cell activation and proliferation [\(32\)](#page-8-0). IL-2RA (CD25) is the alpha subunit of the IL-2 receptor and is likewise involved in immune inflammatory responses ([33,](#page-8-0) [34\)](#page-8-0). IL-2 plays a multifaceted regulatory role in the development of bone tissue, activating Treg cells to regulate the activity of innate and adaptive immune system cells and suppress the inflammatory response, which reduces the occurrence of local inflammatory responses, protects blood vessels and bone cells from damage [\(35](#page-8-0)–[37](#page-8-0)). IL-2 plays a pivotal role in mitigating osteonecrosis and fostering the healing of bone tissue Lu et al. [10.3389/fendo.2024.1344917](https://doi.org/10.3389/fendo.2024.1344917)

by impacting various cellular processes. Specifically, it influences osteoblast proliferation and differentiation, promotes osteoclast activity, and inhibits the activity of bone-resorbing cells. Osteoblasts, being crucial for bone repair, are significantly influenced by IL-2, as evidenced by Reyes-Botella et al.'s study, which demonstrated that IL-2 fosters an increase in the number of osteoblasts ([38\)](#page-8-0). Furthermore, a vitro research by Sartori et al. suggests that IL-2 can stimulate the upregulation of glucocorticoid receptors, indirectly fostering osteoblast proliferation [\(39](#page-8-0)). Additionally, a finding by Sun et al. suggests that IL-2 can hinder osteoclastogenesis, thus indirectly safeguarding bone tissue and aiding in the alleviation of osteonecrosis [\(40](#page-8-0)). Due to the limited number of observational clinical studies or meta-analyses linking IL-2RA to osteoimmunity, little is known about the role of IL-2RA in bone tissue healing. IL-2RA has been shown to be associated with the development of autoimmune inflammatory diseases [\(41,](#page-8-0) [42](#page-8-0)), and immune disorders affecting the skeleton may be an important cause of osteonecrosis, with a strong association between abnormal immune responses and immune cell infiltration in osteonecrosis tissue, often resulting in necrotic bone tissue showing signs of uncontrolled inflammation [\(43](#page-8-0), [44](#page-8-0)). Rheumatoid arthritis (RA) is an autoimmune inflammatory disease that leads to persistent synovitis and severe bone and cartilage destruction, which is closely associated with osteonecrosis [\(45\)](#page-8-0). Lakshmanan et al. showed that IL-2RA may be clearly linked to the pathogenesis of arthritis ([46\)](#page-8-0), and Hinks et al. stated in an earlier study that IL-2RA was juvenile idiopathic arthritis sensitive gene locus, which may reveal another important pathogenesis of osteonecrosis ([47\)](#page-8-0). However, the precise mechanisms of IL-2 and IL-2RA in this regard demand further research and exploration to provide a more precise scientific basis and clinical application outlook for the treatment of diseases associated with the skeletal field.

Fibroblast growth factor (FGF) comprises a peptide family with 22 distinct peptides [\(48](#page-8-0)), of which basic fibroblast growth factor (bFGF) is a key coordinating factor that affects bone homeostasis. Kawaguchi et al. demonstrated that bFGF, acting on osteoblasts at high concentrations, not only efficiently stimulates bone formation via COX-2 induction and production of prostaglandins, but also stimulates bone resorption, whereas at low concentrations it acts directly on mature osteoclasts to resorb bone [\(49](#page-8-0)). In a follow-up study, Kawaguchi demonstrated that bFGF affects bone resorption by acting directly on osteoclasts through activation of FGF receptor 1 and p42/p44 MAP kinase, proving the previous study [\(50](#page-8-0)). In addition, a study by Taketomi et al. found that bFGF stimulation induced high levels of Sprouty2 expression, inhibited the expression of markers of osteoblast differentiation, and also inhibited osteoblast matrix mineralization [\(51](#page-8-0)). Previous investigations have highlighted bFGF's capability to induce the differentiation of bone marrow mesenchymal stem cells into lipogenic cells [\(52](#page-8-0), [53\)](#page-8-0). This induction leads to intraosseous fat deposition, triggering structural alterations in bone that impact blood supply and mechanical properties, ultimately culminating in osteonecrosis ([54,](#page-8-0) [55](#page-8-0)). In addition, the above arguments are supported by Ganguly et al. who showed that myeloid-specific TGF- β signaling, which promotes bFGF expression, is strongly associated with the development of osteolytic bone lesions, and Lee et al.'s study suggests that bFGF plays a key role in the onset and development of RA, which provides support for the above argument [\(56,](#page-8-0) [57\)](#page-8-0). Interestingly, however, it has been shown that bFGF in combination with BMP-2 and VEGF promotes osteogenic differentiation of bone marrow MSCs ([58\)](#page-9-0). It has also been consequently investigated as a promising agent for promoting bone tissue healing [\(59](#page-9-0), [60](#page-9-0)), contributing to the development of novel materials to promote bone tissue healing. As a versatile factor, bFGF is anticipated to emerge as a new therapeutic target for preventing osteonecrosis and promoting bone tissue healing. This multifaceted role positions bFGF as an active participant in the development and prognosis of osteonecrosis, presenting potential avenues for therapeutic interventions.

Our study found an interesting association between stem cell factor (SCF) and a higher risk of developing pharmacological osteonecrosis (OR=3.356, 95% CI=1.09-10.30, p=0.034 per 1 standard deviation (SD)). Earlier studies have demonstrated the potent enhancing effect of SCF on osteoclast activity in vitro [\(61\)](#page-9-0), and glucocorticoids, one of the causative agents of pharmacologic osteonecrosis, also increase the expression of SCF in various ways [\(62\)](#page-9-0), which may reveal one of the key pathogenetic mechanisms of drug-induced osteonecrosis. Furthermore, it is worth noting that Molfetta et al. demonstrated that sustained stimulation of SCF promotes the proliferation of connective tissue-like mast cell subpopulations and maintains a pro-inflammatory microenvironment in the body, which may also be one of the potential factors contributing to the development of osteonecrosis lesions [\(63](#page-9-0)). Interestingly, however, the positive effects of SCF on osteogenesis, possibly through the c-Kit-Akt signaling pathway, have been recognized in recent years, which may also suggest that SCF may have beneficial effects on osteonecrosis repair ([64,](#page-9-0) [65\)](#page-9-0). Additionally, studies have shown that SCF can stimulate cell migration and proliferation, promote blood vessel formation and maturation, and play an important role in angiogenesis and tissue engineering, which may be a potential therapeutic target for osteonecrosis [\(66,](#page-9-0) [67\)](#page-9-0). While only a limited number of clinical observational studies or meta-analyses have investigated the correlation between SCF and osteonecrosis or drug-induced osteonecrosis, our current MR analysis presents compelling evidence suggesting that elevated levels of circulating SCF are indeed associated with an increased risk of osteonecrosis. This insight, derived from the unique perspective of MR analysis, contributes valuable information to the existing body of research and emphasizes the importance of considering SCF as a potential factor in the development of osteonecrosis.

The current study has successfully established a causal relationship between bFGF, IL-2, IL-RA, and osteonecrosis from a genetic perspective. Although SCF did not exhibit any statistically significant causal association with osteonecrosis, it remains a potential key player in the mechanism of drug-induced osteonecrosis development. This study holds several notable advantages. Firstly, it is the inaugural MR study to illuminate the relationship between inflammatory cytokines and the risk of osteonecrosis. Secondly, in contrast to observational studies, our approach substantially mitigates the impact of confounders and reverse causality, enhancing the reliability of causality inferences. Thirdly, leveraging publicly available GWAS databases and a substantial amount of original study data contributes to the robustness of our findings. Fourthly, compared to the time-consuming nature of randomized controlled trials (RCTs), our study provides cost-effective insights. Nevertheless, the study does have certain limitations. Firstly, owing to the European origin of the database, the results may not be fully generalizable to other populations. Secondly, the study faced constraints due to the limited dataset of cases and non-cases with osteonecrosis. Given that genetic variation typically exerts a modest effect on exposure, necessitating large sample sizes for statistically significant results, our dataset size might be limiting. In addition, this study only does not have validation against other databases, which may have some impact on the usability of our findings. Lastly, the publicly available pooled meta-analyses lacked comprehensive information on sex and age, and therefore did not allow for subgroup analyses based on demographic or clinical factors, nor did they allow for a comprehensive assessment of potential nonlinear associations between circulating levels of inflammatory factors and osteonecrosis, which may have led to less than comprehensive study results.

Conclusion

In conclusion, this Mendelian randomization study stands as the inaugural investigation into the causal relationship between osteonecrosis and 41 distinct inflammatory cytokines. Notably, the study unequivocally establishes a causal association between bFGF, IL-2, and IL-2RA with osteonecrosis. These findings not only contribute significant insights into the pathogenesis of osteonecrosis but also pave the way for the development of effective management strategies in clinical settings. The identification of bFGF, IL-2, and IL-2RA as causally linked to osteonecrosis suggests promising avenues for potential therapeutic interventions. This novel perspective positions these cytokines as potential targets for the treatment of osteonecrosis, offering hope for the development of more targeted and effective therapeutic approaches in the future. However, it's noted that due to the European origin of the database, the generalizability of the findings to other populations, beyond individuals of European ancestry, may be limited.

Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material. Further inquiries can be directed to the corresponding authors.

Author contributions

YL: Conceptualization, Writing – original draft. YP: Conceptualization, Writing – original draft, Data curation. Y-MG:

References

2. Zhu T, Cui Y, Zhang M, Zhao D, Liu G, Ding J. Engineered three-dimensional scaffolds for enhanced bone regeneration in osteonecrosis. Bioact Mater. (2020) 5:584– 601. doi: [10.1016/j.bioactmat.2020.04.008](https://doi.org/10.1016/j.bioactmat.2020.04.008)

3. Li Z, Yang B, Weng X, Tse G, Chan MTV, Wu WKK. Emerging roles of micrornas in osteonecrosis of the femoral head. Cell Prolif.. (2018) 51(1):e12405. doi: [10.1111/cpr.12405](https://doi.org/10.1111/cpr.12405)

4. Gupta A, Tripathi L, Pandey S, Dwivedi D. Biology of bone morphogenetic proteins in skeleton disease: osteonecrosis in sickle cell disease patients. Curr Protein Pept Sci. (2022) 23:264–70. doi: [10.2174/1389203723666220530104442](https://doi.org/10.2174/1389203723666220530104442)

Investigation, Methodology, Writing – original draft. F-FZ: Writing – review & editing. LW: Writing – review & editing, Conceptualization. YZ: Conceptualization, Writing – review & editing.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: [https://www.frontiersin.org/articles/10.3389/fendo.2024.1344917/](https://www.frontiersin.org/articles/10.3389/fendo.2024.1344917/full#supplementary-material) [full#supplementary-material](https://www.frontiersin.org/articles/10.3389/fendo.2024.1344917/full#supplementary-material)

^{5.} Jones JP Jr. Coagulopathies and osteonecrosis. Acta orthopaedica Belgica. (1999) 65 Suppl 1:5–8.

^{6.} Goodman SB, Maruyama M. Inflammation, bone healing and osteonecrosis: from bedside to bench. J Inflamm Res. (2020) 13:913–23. doi: [10.2147/jir.S281941](https://doi.org/10.2147/jir.S281941)

^{7.} Cohen-Rosenblum A, Cui Q. Osteonecrosis of the femoral head. Orthopedic Clinics North America. (2019) 50:139–49. doi: [10.1016/j.ocl.2018.10.001](https://doi.org/10.1016/j.ocl.2018.10.001)

^{8.} Yoon BH, Mont MA, Koo KH, Chen CH, Cheng EY, Cui Q, et al. The 2019 revised version of association research circulation osseous staging system of osteonecrosis of the femoral head. *J Arthroplasty*. (2020) 35:933-40. doi: [10.1016/](https://doi.org/10.1016/j.arth.2019.11.029) [j.arth.2019.11.029](https://doi.org/10.1016/j.arth.2019.11.029)

9. Cui Q, Jo WL, Koo KH, Cheng EY, Drescher W, Goodman SB, et al. Arco consensus on the pathogenesis of non-traumatic osteonecrosis of the femoral head. J Korean Med Sci. (2021) 36:e65. doi: [10.3346/jkms.2021.36.e65](https://doi.org/10.3346/jkms.2021.36.e65)

10. Rezus E, Tamba BI, Badescu MC, Popescu D, Bratoiu I, Rezus C. Osteonecrosis of the femoral head in patients with hypercoagulability-from pathophysiology to therapeutic implications. Int J Mol Sci. (2021) 22(13):6801. doi: [10.3390/ijms22136801](https://doi.org/10.3390/ijms22136801)

11. Loi F, Córdova LA, Pajarinen J, Lin TH, Yao Z, Goodman SB. Inflammation, fracture and bone repair. Bone. (2016) 86:119–30. doi: [10.1016/j.bone.2016.02.020](https://doi.org/10.1016/j.bone.2016.02.020)

12. Oishi Y, Manabe I. Macrophages in inflammation, repair and regeneration. Int Immunol. (2018) 30:511–28. doi: [10.1093/intimm/dxy054](https://doi.org/10.1093/intimm/dxy054)

13. Maruyama M, Rhee C, Utsunomiya T, Zhang N, Ueno M, Yao Z, et al. Modulation of the inflammatory response and bone healing. Front Endocrinol (Lausanne). (2020) 11:386. doi: [10.3389/fendo.2020.00386](https://doi.org/10.3389/fendo.2020.00386)

14. Li T, Huang C, Ma J, Ding R, Zhang Q, Wang W. Identification of inflammationrelated genes and exploration of regulatory mechanisms in patients with osteonecrosis of the femoral head. BioMed Res Int. (2022) 2022:4501186. doi: [10.1155/2022/4501186](https://doi.org/10.1155/2022/4501186)

15. Zou D, Zhang K, Yang Y, Ren Y, Zhang L, Xiao X, et al. Th17 and il-17 exhibit higher levels in osteonecrosis of the femoral head and have a positive correlation with severity of pain. Endokrynol Pol. (2018) 69:283–90. doi: [10.5603/EP.a2018.0031](https://doi.org/10.5603/EP.a2018.0031)

16. Burgess S, Butterworth A, Thompson SG. Mendelian randomization analysis with multiple genetic variants using summarized data. Genet Epidemiol. (2013) 37:658– 65. doi: [10.1002/gepi.21758](https://doi.org/10.1002/gepi.21758)

17. Burgess S, Scott RA, Timpson NJ, Davey Smith G, Thompson SG. Using published data in mendelian randomization: A blueprint for efficient identification of causal risk factors. Eur J Epidemiol. (2015) 30:543–52. doi: [10.1007/s10654-015-0011-z](https://doi.org/10.1007/s10654-015-0011-z)

18. Ahola-Olli AV, Würtz P, Havulinna AS, Aalto K, Pitkänen N, Lehtimäki T, et al. Genome-wide association study identifies 27 loci influencing concentrations of circulating cytokines and growth factors. Am J Hum Genet. (2017) 100:40–50. doi: [10.1016/j.ajhg.2016.11.007](https://doi.org/10.1016/j.ajhg.2016.11.007)

19. Emdin CA, Khera AV, Kathiresan S. Mendelian randomization. Jama. (2017) 318:1925–6. doi: [10.1001/jama.2017.17219](https://doi.org/10.1001/jama.2017.17219)

20. Pierce BL, Ahsan H, Vanderweele TJ. Power and instrument strength requirements for mendelian randomization studies using multiple genetic variants. Int J Epidemiol. (2011) 40:740–52. doi: [10.1093/ije/dyq151](https://doi.org/10.1093/ije/dyq151)

21. Palmer TM, Lawlor DA, Harbord RM, Sheehan NA, Tobias JH, Timpson NJ, et al. Using multiple genetic variants as instrumental variables for modifiable risk factors. Stat Methods Med Res. (2012) 21:223–42. doi: [10.1177/0962280210394459](https://doi.org/10.1177/0962280210394459)

22. Machiela MJ, Chanock SJ. Ldlink: A web-based application for exploring population-specific haplotype structure and linking correlated alleles of possible functional variants.
Bioinf (Oxford England). (2015) 31:3555-7. doi: [10.1093/bioinformatics/btv402](https://doi.org/10.1093/bioinformatics/btv402)

23. Perry BI, Burgess S, Jones HJ, Zammit S, Upthegrove R, Mason AM, et al. The potential shared role of inflammation in insulin resistance and schizophrenia: A bidirectional two-sample mendelian randomization study. PloS Med. (2021) 18: e1003455. doi: [10.1371/journal.pmed.1003455](https://doi.org/10.1371/journal.pmed.1003455)

24. Lawlor DA, Harbord RM, Sterne JA, Timpson N, Davey Smith G. Mendelian randomization: using genes as instruments for making causal inferences in epidemiology. Stat Med. (2008) 27:1133–63. doi: [10.1002/sim.3034](https://doi.org/10.1002/sim.3034)

25. Burgess S, Thompson SG. Avoiding bias from weak instruments in mendelian randomization studies. Int J Epidemiol. (2011) 40:755–64. doi: [10.1093/ije/dyr036](https://doi.org/10.1093/ije/dyr036)

26. Cui Z, Tian Y. Using genetic variants to evaluate the causal effect of serum vitamin D concentration on covid-19 susceptibility, severity and hospitalization traits: A mendelian randomization study. J Trans Med. (2021) 19:300. doi: [10.1186/s12967-](https://doi.org/10.1186/s12967-021-02973-5) [021-02973-5](https://doi.org/10.1186/s12967-021-02973-5)

27. Leuti A, Fazio D, Fava M, Piccoli A, Oddi S, Maccarrone M. Bioactive lipids, inflammation and chronic diseases. Adv Drug Deliv Rev. (2020) 159:133–69. doi: [10.1016/j.addr.2020.06.028](https://doi.org/10.1016/j.addr.2020.06.028)

28. Panigrahy D, Gilligan MM, Serhan CN, Kashfi K. Resolution of inflammation: an organizing principle in biology and medicine. Pharmacol Ther. (2021) 227:107879. doi: [10.1016/j.pharmthera.2021.107879](https://doi.org/10.1016/j.pharmthera.2021.107879)

29. Tsukasaki M, Takayanagi H. Osteoimmunology: evolving concepts in boneimmune interactions in health and disease. Nat Rev Immunol. (2019) 19:626–42. doi: [10.1038/s41577-019-0178-8](https://doi.org/10.1038/s41577-019-0178-8)

30. Walsh MC, Takegahara N, Kim H, Choi Y. Updating osteoimmunology: regulation of bone cells by innate and adaptive immunity. Nat Rev Rheumatol. (2018) 14:146–56. doi: [10.1038/nrrheum.2017.213](https://doi.org/10.1038/nrrheum.2017.213)

31. Phipps MC, Huang Y, Yamaguchi R, Kamiya N, Adapala NS, Tang L, et al. In vivo monitoring of activated macrophages and neutrophils in response to ischemic osteonecrosis in a mouse model. J Orthop Res. (2016) 34:307–13. doi: [10.1002/jor.22952](https://doi.org/10.1002/jor.22952)

32. Wu R, Li N, Zhao X, Ding T, Xue H, Gao C, et al. Low-dose interleukin-2: biology and therapeutic prospects in rheumatoid arthritis. Autoimmun Rev. (2020) 19:102645. doi: [10.1016/j.autrev.2020.102645](https://doi.org/10.1016/j.autrev.2020.102645)

33. Maier LM, Anderson DE, Severson CA, Baecher-Allan C, Healy B, Liu DV, et al. Soluble il-2ra levels in multiple sclerosis subjects and the effect of soluble il-2ra on immune responses. J Immunol (Baltimore Md 1950). (2009) 182:1541–7. doi: [10.4049/](https://doi.org/10.4049/jimmunol.182.3.1541) [jimmunol.182.3.1541](https://doi.org/10.4049/jimmunol.182.3.1541)

34. Sadovnik I, Hoelbl-Kovacic A, Herrmann H, Eisenwort G, Cerny-Reiterer S, Warsch W, et al. Identification of cd25 as stat5-dependent growth regulator of leukemic

stem cells in ph+ Cml. Clin Cancer Res an Off J Am Assoc Cancer Res. (2016) 22:2051– 61. doi: [10.1158/1078-0432.Ccr-15-0767](https://doi.org/10.1158/1078-0432.Ccr-15-0767)

35. Arenas-Ramirez N, Woytschak J, Boyman O. Interleukin-2: biology, design and application. Trends Immunol. (2015) 36:763–77. doi: [10.1016/j.it.2015.10.003](https://doi.org/10.1016/j.it.2015.10.003)

36. Dikiy S, Rudensky AY. Principles of regulatory T cell function. Immunity. (2023) 56:240–55. doi: [10.1016/j.immuni.2023.01.004](https://doi.org/10.1016/j.immuni.2023.01.004)

37. Huang F, Wong P, Li J, Lv Z, Xu L, Zhu G, et al. Osteoimmunology: the correlation between osteoclasts and the th17/treg balance in osteoporosis. J Cell Mol Med. (2022) 26:3591–7. doi: [10.1111/jcmm.17399](https://doi.org/10.1111/jcmm.17399)

38. Reyes-Botella C, Vallecillo-Capilla MF, Ruiz C. Effect of different growth factors on human cultured osteoblast-like cells. Cell Physiol Biochem. (2002) 12:353–8. doi: [10.1159/000067905](https://doi.org/10.1159/000067905)

39. Sartori ML, Masera RG, Staurenghi A, Racca S, Angeli A. Interleukin 2 upregulates glucocorticoid receptor number in human peripheral blood mononuclear cells and the osteosarcoma cell line saos-2 in vitro. Steroids. (1998) 63:349–51. doi: [10.1016/s0039-128x\(98\)00023-3](https://doi.org/10.1016/s0039-128x(98)00023-3)

40. Sun H, Zhao Y, Wang K, Zhu L, Dong J, Zhao J, et al. Low dose il-2 suppress osteoclastogenesis in collagen-induced arthritis via jnk dependent pathway. Immun Inflamm Dis. (2020) 8:727–35. doi: [10.1002/iid3.364](https://doi.org/10.1002/iid3.364)

41. Cavanillas ML, Alcina A, Núñez C, de las Heras V, Fernández-Arquero M, BartoloméM, et al. Polymorphisms in the il2, il2ra and il2rb genes in multiple sclerosis risk. Eur J Hum Genet EJHG. (2010) 18:794–9. doi: [10.1038/ejhg.2010.15](https://doi.org/10.1038/ejhg.2010.15)

42. Bouzid D, Amouri A, Fourati H, Marques I, Abida O, Tahri N, et al. Polymorphisms in the il2ra and il2rb genes in inflammatory bowel disease risk. Genet testing Mol Biomarkers. (2013) 17:833–9. doi: [10.1089/gtmb.2013.0291](https://doi.org/10.1089/gtmb.2013.0291)

43. Yu R, Zhang J, Zhuo Y, Hong X, Ye J, Tang S, et al. Arg2, map4k5 and tsta3 as diagnostic markers of steroid-induced osteonecrosis of the femoral head and their correlation with immune infiltration. Front Genet. (2021) 12:691465. doi: [10.3389/](https://doi.org/10.3389/fgene.2021.691465) [fgene.2021.691465](https://doi.org/10.3389/fgene.2021.691465)

44. Ma H, Zhang W, Shi J. Differentially expressed genes reveal the biomarkers and molecular mechanism of osteonecrosis. J Healthc Eng. (2022) 2022:8684137. doi: [10.1155/2022/8684137](https://doi.org/10.1155/2022/8684137)

45. de Molon RS, Hsu C, Bezouglaia O, Dry SM, Pirih FQ, Soundia A, et al. Rheumatoid arthritis exacerbates the severity of osteonecrosis of the jaws (Onj) in mice. A randomized, prospective, controlled animal study. J Bone mineral Res Off J Am Soc Bone Mineral Res. (2016) 31:1596–607. doi: [10.1002/jbmr.2827](https://doi.org/10.1002/jbmr.2827)

46. Lakshmanan DK, Murugesan S, Rajendran S, Ravichandran G, Elangovan A, Raju K, et al. Brassica juncea (L.) czern. Leaves alleviate adjuvant-induced rheumatoid arthritis in rats via modulating the finest disease targets - il2ra, il18 and vegfa. J Biomol Struct Dyn. (2022) 40:8155–68. doi: [10.1080/07391102.2021.1907226](https://doi.org/10.1080/07391102.2021.1907226)

47. Hinks A, Ke X, Barton A, Eyre S, Bowes J, Worthington J, et al. Association of the il2ra/cd25 gene with juvenile idiopathic arthritis. Arthritis rheumatism. (2009) 60:251– 7. doi: [10.1002/art.24187](https://doi.org/10.1002/art.24187)

48. Mossahebi-Mohammadi M, Quan M, Zhang JS, Li X. Fgf signaling pathway: A key regulator of stem cell pluripotency. Front Cell Dev Biol. (2020) 8:79. doi: [10.3389/](https://doi.org/10.3389/fcell.2020.00079) [fcell.2020.00079](https://doi.org/10.3389/fcell.2020.00079)

49. Kawaguchi H, Chikazu D, Nakamura K, Kumegawa M, Hakeda Y. Direct and indirect actions of fibroblast growth factor 2 on osteoclastic bone resorption in cultures. J Bone mineral Res Off J Am Soc Bone Mineral Res. (2000) 15:466–73. doi: [10.1359/](https://doi.org/10.1359/jbmr.2000.15.3.466) [jbmr.2000.15.3.466](https://doi.org/10.1359/jbmr.2000.15.3.466)

50. Chikazu D, Hakeda Y, Ogata N, Nemoto K, Itabashi A, Takato T, et al. Fibroblast growth factor (Fgf)-2 directly stimulates mature osteoclast function through activation of fgf receptor 1 and P42/P44 map kinase. J Biol Chem. (2000) 275:31444–50. doi: [10.1074/jbc.M910132199](https://doi.org/10.1074/jbc.M910132199)

51. Taketomi T, Onimura T, Yoshiga D, Muratsu D, Sanui T, Fukuda T, et al. Sprouty2 is involved in the control of osteoblast proliferation and differentiation through the fgf and bmp signaling pathways. Cell Biol Int. (2018) 42:1106–14. doi: [10.1002/cbin.10876](https://doi.org/10.1002/cbin.10876)

52. Aguirre JI, Leal ME, Rivera MF, Vanegas SM, Jorgensen M, Wronski TJ. Effects of basic fibroblast growth factor and a prostaglandin E2 receptor subtype 4 agonist on osteoblastogenesis and adipogenesis in aged ovariectomized rats. J Bone mineral Res Off J Am Soc Bone Mineral Res. (2007) 22:877–88. doi: [10.1359/jbmr.070313](https://doi.org/10.1359/jbmr.070313)

53. Song X, Li Y, Chen X, Yin G, Huang Q, Chen Y, et al. Bfgf promotes adipocyte differentiation in human mesenchymal stem cells derived from embryonic stem cells. Genet Mol Biol. (2014) 37:127–34. doi: [10.1590/s1415-47572014000100019](https://doi.org/10.1590/s1415-47572014000100019)

54. Lin X, Zhu D, Wang K, Luo P, Rui G, Gao Y, et al. Activation of aldehyde dehydrogenase 2 protects ethanol-induced osteonecrosis of the femoral head in rat model. Cell Prolif. (2022) 55:e13252. doi: [10.1111/cpr.13252](https://doi.org/10.1111/cpr.13252)

55. Gillet C, Dalla Valle A, Gaspard N, Spruyt D, Vertongen P, Lechanteur J, et al. Osteonecrosis of the femoral head: lipotoxicity exacerbation in msc and modifications of the bone marrow fluid. Endocrinology. (2017) 158:490–502. doi: [10.1210/en.2016-1687](https://doi.org/10.1210/en.2016-1687)

56. Ganguly SS, Daft PG, Cao J, Meng X, Zhong ZA, Vander Ark A, et al. Loss of myeloid-specific tgf-Β Signaling decreases cthrc1 to downregulate bfgf and the development of H1993-induced osteolytic bone lesions. Cancers. (2018) 10(12):463. doi: [10.3390/cancers10120463](https://doi.org/10.3390/cancers10120463)

57. Lee CJ, Moon SJ, Jeong JH, Lee S, Lee MH, Yoo SM, et al. Kaempferol targeting on the fibroblast growth factor receptor 3-ribosomal S6 kinase 2 signaling axis prevents the development of rheumatoid arthritis. Cell Death Dis. (2018) 9:401. doi: [10.1038/](https://doi.org/10.1038/s41419-018-0433-0) [s41419-018-0433-0](https://doi.org/10.1038/s41419-018-0433-0)

58. Bai Y, Li P, Yin G, Huang Z, Liao X, Chen X, et al. Bmp-2, vegf and bfgf synergistically promote the osteogenic differentiation of rat bone marrow-derived mesenchymal stem cells. Biotechnol Lett. (2013) 35:301–8. doi: [10.1007/s10529-012-](https://doi.org/10.1007/s10529-012-1084-3) [1084-3](https://doi.org/10.1007/s10529-012-1084-3)

59. Guo Y, Hu Z, Chen J, Zhang Z, Liu Q, Li J, et al. Injectable tg-linked recombinant human collagen hydrogel loaded with bfgf for rat cranial defect repair. *Int J Biol* Macromol. (2023) 236:123864. doi: [10.1016/j.ijbiomac.2023.123864](https://doi.org/10.1016/j.ijbiomac.2023.123864)

60. Li S, Zhang L, Liu C, Kim J, Su K, Chen T, et al. Spontaneous immunomodulation and regulation of angiogenesis and osteogenesis by sr/cuborosilicate glass (Bsg) bone cement to repair critical bone defects. Bioact Mater. (2023) 23:101–17. doi: [10.1016/j.bioactmat.2022.10.021](https://doi.org/10.1016/j.bioactmat.2022.10.021)

61. van't Hof RJ, von Lindern M, Nijweide PJ, Beug H. Stem cell factor stimulates chicken osteoclast activity in vitro. FASEB J Off Publ Fed Am Societies Exp Biol. (1997) 11:287–93. doi: [10.1096/fasebj.11.4.9068618](https://doi.org/10.1096/fasebj.11.4.9068618)

62. Da Silva CA, Kassel O, Lebouquin R, Lacroix EJ, Frossard N. Paradoxical early glucocorticoid induction of stem cell factor (Scf) expression in inflammatory conditions. Br J Pharmacol. (2004) 141:75–84. doi: [10.1038/sj.bjp.0705598](https://doi.org/10.1038/sj.bjp.0705598)

63. Molfetta R, Lecce M, Milito ND, Putro E, Pietropaolo G, Marangio C, et al. Scf and il-33 regulate mouse mast cell phenotypic and functional plasticity supporting a pro-inflammatory microenvironment. Cell Death Dis. (2023) 14:616. doi: [10.1038/](https://doi.org/10.1038/s41419-023-06139-7) $\overline{}$ [s41419-023-06139-7](https://doi.org/10.1038/s41419-023-06139-7)

64. Yang L, Wu Z, Yin G, Liu H, Guan X, Zhao X, et al. Stem cell factor (Scf) protects osteoblasts from oxidative stress through activating C-kit-akt signaling. Biochem Biophys Res Commun. (2014) 455:256–61. doi: [10.1016/j.bbrc.2014.11.002](https://doi.org/10.1016/j.bbrc.2014.11.002)

65. Wu X, Yang S, Duan D, Liu X, Zhang Y, Wang J, et al. A combination of granulocyte colony-stimulating factor and stem cell factor ameliorates steroidassociated osteonecrosis in rabbits. J Rheumatol. (2008) 35:2241–8. doi: [10.3899/](https://doi.org/10.3899/jrheum.071209) irheum.071209

66. Takematsu E, Massidda M, Howe G, Goldman J, Felli P, Mei L, et al. Transmembrane stem factor nanodiscs enhanced revascularization in a hind limb ischemia model in diabetic, hyperlipidemic rabbits. Sci Rep. (2024) 14:2352. doi: [10.1038/s41598-024-52888-6](https://doi.org/10.1038/s41598-024-52888-6)

67. Mu X, Liu H, Yang S, Li Y, Xiang L, Hu M, et al. Chitosan tubes inoculated with dental pulp stem cells and stem cell factor enhance facial nerve-vascularized regeneration in rabbits. ACS Omega. (2022) 7:18509–20. doi: [10.1021/](https://doi.org/10.1021/acsomega.2c01176) [acsomega.2c01176](https://doi.org/10.1021/acsomega.2c01176)