Check for updates

OPEN ACCESS

EDITED BY Rachida Guennoun, Institut National de la Santé et de la Recherche Médicale (INSERM), France

REVIEWED BY Claire L. Gibson

University of Nottingham, United Kingdom Silvia Giatti, University of Milan, Italy Alejandro Federico De Nicola, CONICET Institute of Biology and Experimental Medicine (IBYME), Argentina

*CORRESPONDENCE Meharvan Singh Smsingh@luc.edu

RECEIVED 30 August 2023 ACCEPTED 15 January 2024 PUBLISHED 26 February 2024

CITATION

Singh M, Krishnamoorthy VR, Kim S, Khurana S and LaPorte HM (2024) Brainderived neuerotrophic factor and related mechanisms that mediate and influence progesterone-induced neuroprotection. *Front. Endocrinol.* 15:1286066. doi: 10.3389/fendo.2024.1286066

COPYRIGHT

© 2024 Singh, Krishnamoorthy, Kim, Khurana and LaPorte. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Brain-derived neuerotrophic factor and related mechanisms that mediate and influence progesterone-induced neuroprotection

Meharvan Singh*, Vignesh R. Krishnamoorthy, Seongcheol Kim, Saira Khurana and Heather M. LaPorte

Department of Cell and Molecular Physiology, Stritch School of Medicine, Loyola University Chicago, Maywood, IL, United States

Historically, progesterone has been studied significantly within the context of reproductive biology. However, there is now an abundance of evidence for its role in regions of the central nervous system (CNS) associated with such nonreproductive functions that include cognition and affect. Here, we describe mechanisms of progesterone action that support its brain-protective effects, and focus particularly on the role of neurotrophins (such as brain-derived neurotrophic factor, BDNF), the receptors that are critical for their regulation, and the role of certain microRNA in influencing the brain-protective effects of progesterone. In addition, we describe evidence to support the particular importance of glia in mediating the neuroprotective effects of progesterone. Through this review of these mechanisms and our own prior published work, we offer insight into why the effects of a progestin on brain protection may be dependent on the type of progestin (e.g., progesterone versus the synthetic, medroxyprogesterone acetate) used, and age, and as such, we offer insight into the future clinical implication of progesterone treatment for such disorders that include Alzheimer's disease, stroke, and traumatic brain injury.

KEYWORDS

progesterone, brain-derived neurotrophic factor (BDNF), microRNA (miRNA), neuroprotection, stroke, brain

The biology of progesterone

Progesterone is a natural progestin and an important gonadal hormone synthesized in mainly in the ovaries of females and both the adrenal cortex and testes of males. Despite the fact that progesterone levels are overall higher in females, it is known that there is recorded similarities between male progesterone levels and those of females specifically during the

follicular phase in female menstrual cycle, indicating a definite and important role of progesterone in males (1). Historically, progesterone has primarily been considered with regards solely to the reproductive functions. For example, it plays vital role in not only female fertility and embryo implantation, but also in the maintenance of the uterus and production of inflammatory mediators in the uterine cavity during pregnancy to prevent miscarriage or preterm labor (2). And though our knowledge of progesterone and related progestins with regards to their role in reproductive biology is quite extensive, we also now appreciate there are rather important effects from progesterone in multiple organ systems including the brain, in which progesterone has demonstrated protective effects. The mechanisms associated with the protective effects of progesterone are complex, and as suggested below, may require the right complement of progesterone receptors (e.g., the classical progesterone receptor (PR) and membrane progesterone receptors), may involve the simultaneous influence on multiple cell types (e.g., glia and neurons), as well as regulation of certain classes of growth factors (e.g., neurotrophins).

The receptor pharmacology of progesterone

Brain regions such as the cerebral cortex and hippocampus express not just the classical progesterone receptors (PR) (3, 4), but also express membrane progesterone receptors (see (5) for review). The "classical" PR is generally described as a nuclear transcription factor, which when bound to specific progesterone response elements (PRE) within the promoter region of target genes, regulates the transcription of such genes. PR-A and PR-B, the two main isoforms of the classical PR, are transcribed from the same gene, and transcription is usually estrogen-dependent. A third isoform, known as PR-C, also exists, but its exact function is unclear (6).

Recently it has been proposed that there are membrane receptors for progesterone. While it has only been recently that the membrane-associated progesterone receptors have been cloned, they have been suggested for years originating from the knowledge and observation of specific and displaceable binding sites found in preparations of synaptosome membranes (7, 8). Zhu et al., discovered a novel membrane-associated progesterone receptor which is termed mPR (9) that is predicted to couple to Gi/o class of G-proteins (10). Additional progesterone membrane receptors include Progesterone receptor membrane component 1 (Pgmrc1) (11–13) which is implicated in abundant features of cellular function ranging from the regulation of reproductive behavior (11), steroidogenesis (14), growth regulation of triple-negative breast cancer (15), to neuronal development (16) and potentially neuroprotective effects of progesterone as described above.

Pgrmc1's involvement in the cellular functions associated with cytoprotection may be attributed to its role as a positive regulator of many cytochrome P450 catalyzed reactions that are imperative for intracellular sterol metabolism (17). The Singh laboratory has previously demonstrated that, through our experimental

conditions showing progesterone-induced neuroprotection, the classical PR, mPR-alpha, mPR-beta and Pgrmc1 are expressed, while we recently determined that progesterone's ability to elicit an increase in Brain-derived neurotorphic factor (BDNF) expression is dependent on the classical PR (18). With respect to Pgrmc1, however, selective knockdown completely abolishes the ability of progesterone to elicit an increase in BDNF release (19), an effect that we believe to be critical to progesterone's neuroprotective effects (20). Based on these observations, and at least from the standpoint of neuroprotection, we suggest that the classical PR and the Pgrmc1 are critical, although perhaps not exclusive, mediators of progesterone's effects on cell viability.

Progesterone and neuroprotection

Progesterone has been reported to be protective against a variety of insults relevant to brain aging or indeed, various neurodegenerative diseases, that include stroke, traumatic brain injury (TBI), stroke and Alzheimer's disease (AD). For example, progesterone, at physiologically relevant concentrations, has been shown to significantly ameliorate oxidative and/or excitotoxic injury resulting from glutamate treatment (20–23), glucose deprivation (24), as well as FeSO₄- and amyloid β -peptide-induced toxicity (24).

With regards to animal models of stroke, progesterone has been shown to be protective, as exemplified by the study by Jiang et al., which illustrated that treatment with progesterone prior to middle cerebral artery occlusion (MCAO) resulted in a significant reduction in cerebral infarction as well as the functional impairments that resulted from the occlusion (25). Interestingly, administration of progesterone following ischemia was also found to be protective (26-28), and resulted in improvements in various functional outcomes, including the rotarod test, and adhesivebacked somatosensory and neurological scores (29). Progesterone has even been shown to provide protection against ischemiainduced visual impairments, as shown by the work of Allen et al. (2015), who demonstrated that post-ischemic administration of progesterone protected against MCAO-induced retinal ganglion cell (RGC) loss, glutamine synthetase upregulation, and glial fibrillary acidic protein (GFAP) upregulation (30). The protective effects of progesterone following insult suggest that both rapid/ immediate and long-term mechanisms of progesterone action may underlie the protective effects of progesterone.

In experimental models of traumatic brain injury (TBI), progesterone is also protective. In such models, progesterone administration has been shown to reduce cerebral edema for up to 24 hours following injury. Further, progesterone was found to reduce complement factor C3, glial fibrillary acidic protein (GFAP), and nuclear factor kappa beta (NF κ B) in a rodent model of medial frontal cortex impact injury (31), all of which can be interpreted as protective mechanisms. Progesterone not only facilitates cognitive improvement while reducing secondary neuronal loss caused by edema in ovariectomized female rats after TBI, but also elicits similar effects in male rats (32). Lipid peroxidation was also decreased following treatment with progesterone when administered post-TBI in male rats (33).

The neuroprotective effects of progesterone have also been investigated in the context of experimental systems that simulate the pathology of such neurodegenerative diseases as Alzheimer's disease (AD) and Parkinson's disease (PD). For instance, a study by Qin et al. (2015) demonstrated the neuroprotective properties of progesterone against amyloid beta (Aβ)₂₅₋₃₅-mediated neuronal cell death by alleviating mitochondrial membrane potential loss (34). The neuroprotective effects of progesterone were also noted in animal models of AD, where progesterone improved cognitive performance and glucose uptake in neurons in two separate animal models of AD (35, 36). The protection afforded by progesterone seemed to also be generalizable to other models of neurodegeneration, including Parkinson's disease. For example, progesterone elicited neuroprotective effects in the murine 1methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) model of Parkinson's disease. Of note, and similar to that described in an animal model of stroke, progesterone was protective at administration time points both pre- and post-MPTP treatment (37, 38).

While the anatomical focus of the brain disorders/diseases referenced above were the hippocampus and cortex, it is worth noting that progesterone has also been shown to be protective in other regions of the central nervous system. For instance, work by Thomas et al., highlighted the beneficial effects of progesterone on spinal cord contusion injuries, by showing a reduction in the size of the lesion and a prevention of secondary neuronal loss with progesterone treatment (39). Additionally, progesterone's protective actions have been shown in the Wobbler mouse, an animal model of spinal cord degeneration, where progesterone treatment promoted morphological and functional recovery (40, 41). Re-myelination can also be induced by progesterone as evidenced by the increased expression of myelin proteins in the damaged sciatic nerves of both young adult rats and old (22-24 months of age) males (42). Based on these findings, progesterone may be of potential therapeutic benefit in diseases where demyelination is an important component that contributes to pathogenesis.

While the studies described above were derived from animal models and cell or tissue culture models, it is noteworthy that a phase II, randomized, double-blind, placebo-controlled clinical trial assessing the efficacy of progesterone treatment for acute traumatic brain injury yielded promising results. Data from this study suggested that progesterone treatment can improve functional recovery, at least in those with moderate, but not severe, traumatic brain injury (43-47). However, the results from the Phase III study (ProTECTIII) failed to corroborate earlier findings, potentially due to suboptimal dosing (48). Assessment of "early dosing" with progesterone, as conducted by Wright et al. (2014), also failed to show a benefit of progesterone administration over control despite treatment within 4 hours of injury (49). Given that TBI in the clinical setting is heterogeneous, unlike the very reproducible injury that is sustained in pre-clinical (animal) models of TBI (which have consistently shown a protective effect of

progesterone), the protective effects of progesterone may be evident only in a subset of TBI patients. Indeed, studies like that by Soltani et al. (2017), which focused on diffuse axonal injury (50), extend the earlier suggestion that progesterone may be protective in patients with TBI.

Many studies that have investigated the protective effects of progesterone have done so within the context of estrogen treatment. More specifically, a significant proportion of these studies have evaluated the protective effects of estrogen alone in comparison to combined treatment with estrogen and progesterone. More recently, however, researchers have addressed the influence of progesterone alone. Although growing evidence suggests that treatment with progesterone alone is neuroprotective, the influence of progesterone on estrogen's neuroprotective effects is more equivocal. Some studies suggest that progesterone does not interfere with the effects of estrogens (E) (21, 51, 52), while other studies have argued that progesterone or synthetic progestins antagonize the effects of estrogen (53-58). Specifically, the work of Murphy and Segal demonstrated that progesterone antagonizes the effect of 17β -estradiol (E2) on hippocampal spine density (59). Additionally, McEwen and Woolley showed that in both adult and developing brains, progesterone contributed to the loss of hippocampal spines and spine synapses noted across the estrous cycle (60), although progesterone did result initially (within the first 6 hours) in an increase in hippocampal dendritic spine density (61). In contrast, a positive effect of progesterone, similar to that of E2 in the hippocampus of a rat stroke model, was reported by Zhao et al. (62), while Foy et al. (63) described that progesterone enhanced long-term potentiation (LTP) and long-term depression (LTD) in rat hippocampus. Future studies will help clarify the biological basis of this apparent discrepancy, which may be the result of multiple factors, that include the experimental model used (reflecting the types of receptors expressed in the model), the chosen concentrations/doses of progesterone, the timing of the progesterone relative to that of estrogen, the timing of progesterone relative to the insult, or potentially, regional differences in the effects of combined estrogen and progesterone.

BDNF as a neuroprotectant

BDNF belongs to the family of neurotrophins (that include NGF, NT-3 and NT4), and plays a vital role in maintaining brain health by supporting cell viability and synaptic plasticity (64, 65). The initial synthesis of BDNF occurs in both neurons and glia (66, 67) as a glycosylated precursor (pre-pro-BDNF), which is processed into a 35 kDa pro-BDNF, and can then be converted into the 14 kDa mature BDNF intracellularly or extracellularly (68, 69). Released BDNF can exert its functions on target cells by binding to TrkB (a tropomysin related kinase family (Trk) of receptors) or p75 neurotrophin receptor (p75^{NTR}) receptor (67, 68, 70, 71). Differentiating the effects of pro- versus mature BDNF is of high importance as they often exert contrary biological functions: while mature BDNF binds to the TrkB receptor to influence neuronal survival, differentiation, and promote long-term potentiation

(LTP), pro-BDNF binds preferentially to p75^{NTR} and can promote neuronal apoptosis and long-term depression (LTD) (see (69) and references cited therein), particularly in the absence of TrkB. Furthermore, it is hypothesized that neuronal dysfunction or atrophy that occurs as a consequence of aging or age-associated diseases may result not only from a decrease in mature neurotrophin expression or function (72–75), but potentially also from *increased* accumulation of the pro-neurotrophins. Consistent with this premise, Fahnestock and colleagues have described that the pro-NGF, the pro-neurotrophin for NGF, is increased in brains from individuals diagnosed with AD (76).

As suggested above, there is strong evidence to support the role of BDNF in synaptic plasticity and cognitive function (77, 78), and as such, alteration in its function and/or expression has been implicated in the pathophysiology of aged-related neurodegenerative diseases including AD and PD (79-81), and conversely, restoring BDNF expression and/or function may be therapeutic. In stroke, BDNF has been largely shown to play a protective role. The delivery of BDNF has been shown to promote tissue reparative processes in various animal models of stroke (82, 83) and interventions that improve functional recovery are often associated with increased BDNF levels in the peri-infarct area. For example, Lazarovici et al., reported that pituitary adenylate cyclase activating peptide (PACAP) is protective in a rat model of stroke by inducing BDNF expression and release, as well as activating TrkB receptor (84). Ishrat et al., also showed that the protective effects of progesterone are mediated by BDNF, helping to reduce ischemic lesion size and edema in rats experiencing permanent focal cerebral ischemia (85). Furthermore, gene delivery of BDNF, using a recombinant adeno-associated virus (rAAV), decreased cell death in a rat focal ischemic lesion (86). Conversely, attenuating BDNF levels or its effects following cerebral ischemia often reduces recovery of function (87, 88). It is noteworthy that BDNF is also produced in non-neuronal cells (such as astrocytes, microglia and endothelial cells) after ischemic stroke (89), and therefore these cells may contribute significantly to the BDNFdependent recovery. BDNF has also been implicated as a neuroprotectant in TBI (for review, see (90) and references cited therein). In the hippocampus, conditional knock out of BDNF increased death of adult-born immature neurons following TBI (91). Conversely, therapeutic improvement and recovery of function after TBI is associated with an induction of BDNF and its associated proteins (92, 93). Changes in BDNF signaling have also been implicated in Alzheimer's disease and work by Zheng et al., 2010, shows that A β reduces mature BDNF expression *in vitro* (94). Conversely, BDNF gene delivery has shown to reverse synapse loss and protect against neuronal death of entorhinal neurons in mouse models of AD (95).

The mechanisms by which BDNF-mediated protection occurs may be elicited through multiple mechanisms including the activation of specific signaling pathways, including the MAPK and phosphoinositide inositol-3 kinase (PI-3K) pathways. For example, exogenous BDNF protects primary cortical neurons from apoptosis in a dose-dependent manner (96). While BDNF increases the phosphorylation of PI3K and Ak Strain Transforming (Akt) (as an indicator of their activation), pharmacological inhibition of the PI-3K pathway by LY294002 prevents the neuroprotective effects of BDNF in primary cortical neurons (96). Likewise, extracellular-signal regulated kinase (ERK)1/2 phosphorylation/activation is significantly increased by BDNF, while pharmacological inhibition of the ERK1/2 pathway by PD98059 greatly reduces BDNF's neuroprotective effects. In addition, direct administration of BDNF, via intracerebroventricular administration, in postnatal day 7, rats resulted in phosphorylation of ERK1/2 and Akt within minutes (97), while pharmacological inhibition of ERK inhibited the ability of BDNF to block hypoxia/ischemia-induced caspase-3 activation and tissue loss (97). Based on progesterone's ability to rapidly activate both the ERK/MAPK and Akt signaling pathways in the CNS (21, 23, 98), and increase the expression of BDNF as well, either the direct activation of these signaling pathways (i.e., direct coupling with activated progesterone receptors) or the regulation of BDNF synthesis and release may be relevant mechanisms that are, at least partially, responsible for progesterone's protective effects.

BDNF as a mediator of progesterone's protective effects

Our laboratory (20, 99) and that of others (100, 101) have shown in various experimental systems, including explants of the cerebral cortex, the injured spinal cord, and in degenerating Wobbler motor neurons, that BDNF expression is increased by progesterone. For instance, a study by Meyer et al., 2013, shows that progesterone promotes upregulation of BDNF in the hippocampus, dentate gyrus, and Cornu Ammonis (CA)3 pyramidal regions of the Wobbler mouse in comparison to vehicle groups (102). Additionally, progesterone treatment has been shown to increase BDNF expression in an animal model of TBI (103). Our lab has shown that in cortical explants, treatment with a physiologically relevant concentration of progesterone (100 nM) for 24 hours induces an approximately 75% increase in both BDNF mRNA and protein expression, an effect that appeared to be consistent with the protective effects of progesterone (20). Furthermore, the principal mediator of the effect of progesterone on BDNF expression was determined to be the "classical" intracellular/ nuclear PR, since this effect was inhibited by the pharmacological inhibitor of the PR, RU486, and was lost in PR knockout mice (18). The membrane progesterone receptor (mPR) has also been reported to be potentially involved in promoting expression of BDNF as shown by a study by Castelnovo and Thomas (2022), in which activation of the mPR α in human adipose stem cells differentiated into Schwann cell-like cells (SLC-ASC) led to an upregulation of BDNF expression (104).

The regulation of cell signaling pathways consequent to BDNF action (through interaction with the TrkB receptor) may be an important way by which progesterone elicits its effects, and as such, while synthesis is important, the ability to elicit the <u>release</u> of BDNF is also important. Recently, we have shown that the release of BDNF from glia is triggered by progesterone (19). Interestingly, this release was mediated through a novel membrane-associated progesterone receptor, Pgrmc1, as opposed to the classical PR, which was not

involved in this process as our cultured astrocytes lacked the expression of PR (19). Furthermore, this was associated with activation of the ERK5 signaling cascade (19). Based on these findings, our laboratory proposes that the receptor mechanisms of both Pgrmc1 and PR are required to afford sustainable protective effects, since we have shown that the classical PR mediates the effect of progesterone on BDNF expression, and Pgrmc1 appears to mediate the effect of BDNF on release. Specifically, we posit that progesterone, through the PR, replenishes BDNF stores while simultaneously, through Pgrmc1, promotes the release, and thus, availability of BDNF to surrounding cells.

Progesterone-mediated BDNF regulation may also be relevant to progesterone's cognitive enhancing role. Indeed, numerous studies implicate BDNF in regulating synaptic plasticity in the brain, including LTP (105, 106). Specifically, "late" LTP, a component of LTP that requires de novo synthesis of mRNA and protein, appears to require BDNF (107-109). Mechanistically, BDNF is thought to mediate LTP via N-methyl-D aspartic acid (NMDA) receptor phosphorylation, which in turn, alters the function of the receptor. Indeed, BDNF-mediated signaling has been shown to target both the NR1 (110) and the NR2B (111) subunits of the NMDA receptor. Moreover, functional changes in the receptor, such as increasing the open probability of the NMDA receptor channel, are associated with this BDNF-mediated phosphorylation of the NMDA receptor (112, 113). Studies have also shown that signaling pathways elicited by progesterone are implicated in regulating LTP. Specifically, pharmacological or genetic inhibition/disruption of these pathways can inhibit LTPrelevant NMDA receptor phosphorylation (114), BDNF-induced increase in field excitatory postsynaptic potentials (fEPSPs) (115) and produce frank deficits in hippocampal LTP (116). Accordingly, progesterone may regulate LTP (thereby enhancing cognitive function) by direct activation of specific cell signaling pathways consequent to binding to progesterone receptors, or alternatively, influence LTP through the induction of BDNF release, which in turn, activates the signaling pathways that phosphorylate NMDA receptors.

As previously discussed, the ratio of relative abundance of the pro- and mature forms of the neurotrophin may govern the protective effects of synthesized neurotrophins, since proneurotrophins preferentially bind to the p75 "pan" neurotrophin receptor to promote cell death, while mature neurotrophins preferentially bind to their cognate Trk receptor and elicit signaling events consistent with cell survival. The laboratory of Dr. Donald Stein has recently reported the expression of pro- versus mature neurotrophins can be differentially regulated by progesterone. Specifically, progesterone treatment led to a decrease in the proapoptotic, pro-NGF, while increasing the level of mature NGF in a model of traumatic brain injury (TBI). Although this effect on NGF appears consistent with the protective effects of progesterone, the observed effects of progesterone on pro- versus mature BDNF were not. In fact, progesterone not only decreased the expression of pro-BDNF, but also reduced the expression of mature BDNF and its cognate receptor, TrkB (117). However, recent studies from our lab show that P4 is protective, and that the efficacy of P4's protective effects was enhanced by inhibiting the microRNA, let-7i (118). Future studies are warranted to help clarify this apparent discrepancy.

Metabolites of progesterone and their influence on BDNF

When exploring the neuroprotective properties of progesterone, it is crucial to acknowledge the potential involvement of its metabolite, allopregnanolone (3 α , 5 α tetrahydroprogesterone or THP). This consideration broadens our understanding and opens up new avenues for research into the mechanisms by which progesterone exerts its beneficial effects on brain health. Allopregnanolone, a positive allosteric modulator of the gammaamino butyric acid-A (GABA-A) receptor, has been shown to have neuroprotective effects by reducing excitotoxicity caused by brain injury or insult (119). There is strong evidence from multiple studies suggesting that allopregnanolone treatment offers substantial benefits in reducing various deficits and consequences associated with traumatic brain injury. Examples of a potential benefit of allopregnanolone is its ability to reduce levels of inflammatory cytokines (120), minimize cell death, and alleviate cognitive deficits (120-125) following traumatic brain injury. It has been postulated that allopregnanolone plays a pivotal role in mediating the protective effects of progesterone. In addition to its other benefits, allopregnanolone has been found to have a significant impact on neurogenesis (refer to the study cited in reference (126) and the accompanying sources). Moreover, extensive research has demonstrated the therapeutic benefits of allopregnanolone in treating Alzheimer's disease by promoting neurogenesis, enhancing cognitive function and memory, reducing neuroinflammation and beta-amyloid build-up, as well as improving bioenergetics deficits in 3xTgAD mice (127-130). Interestingly, studies have demonstrated that allopregnanolone can exert its protective benefits by modulating BDNF (refer to reference (125) and the relevant citations therein), while the exact method by which allopregnanolone triggers BDNF remains uncertain (i.e., what receptor(s) allopregnanolone interact with). However, noting recent reports that allopregnanolone is a ligand for the PAQR family of membrane progesterone receptors (131), the involvement of multiple classes of membrane progesterone receptors (i.e., to include mPRs and Pgrmc1) in mediating the effect of metabolites of progesterone is indeed emerging.

And while there has been considerable attention to the neuroprotective effects of allopregnanolone, its precursor, 5α -dihydroprogesterone, has also been shown to protect neurons against excitotoxic insults (120, 132, 133). Thus, the protective effects of progesterone may not only be attributed to the abundance of progesterone and its cognate receptors, but also the abundance of the synthetic enzymes responsible for the conversion of progesterone to its neuroprotective metabolites, and in turn, the relatively more recently described receptors associated with their action (e.g., mPR and Pgrmc1).

Do all clinically used progestins have similar effects on cytoprotection and BDNF?

Medroxyprogesterone Acetate (MPA) is a synthetic progestin often used in conjunction with estrogens to reduce the risk of such cancers that include uterine cancer, associated with unopposed estrogen therapy (134, 135). However, we and others have equivalent effects within the context of their cytoprotective effects. For example, progesterone, but not MPA, was protective against glutamate toxicity in both explants derived from the cerebral cortex (20) and in primary dissociated hippocampal neurons (21). This disparity between the effects of P4 and MPA have also been noted in vivo. For example, a study illustrated that the combined administration of estrogen and progesterone in rhesus monkeys protects against coronary vasospasm, whereas the coadministration of MPA with estrogen failed to elicit this protection (136). This difference between progesterone and MPA is also evident in humans, where progesterone administration to post-menopausal women enhanced the protective effects of estrogen on exerciseinduced myocardial ischemia, whereas MPA did not (137).

While the discrepancy between the protective effects of progesterone and MPA may be attributed to a variety of factors that include differential regulation of ERK translocation (23), antiapoptotic protein regulation (21), and calcium homeostasis (21), it appears that a fundamental difference in the regulation of BDNF may also underlie the difference between progesterone and MPA. Notably, our laboratory showed that that while progesterone increased the expression of BDNF in cerebral cortical cultures, MPA suppressed BDNF levels (18).

More recently, the progestin component of a recently FDAapproved contraceptive vaginal ring, segesterone acetate, was shown to have neuroprotective effects (138–140) in the MCAO model of stroke in male rats. Given the preferential affinity of this compound for the PR over other steroid hormone receptors, such data bolster the importance of the classical PR in mediating progesterone's neuroprotective effects. Further, this study also supports the potential utility of PR-engaging compounds (progesterone and related progestins) in eliciting cytoprotective effects in males as well.

miRNA and their influence of progesterone's biological effects

MicroRNAs (miRNAs) are a class of small non-coding RNAs (approximately 20-22 nucleotides in length) that play a major role in regulating gene expression (141). miRNAs were first discovered in *Caenorhabditis elegans*, in which a short RNA sequence produced from the *lin-14* gene downregulated *lin-14* translation (142). miRNAs are initially synthesized as primary-miRNAs (primiRNA), a hairpin structure that is predominantly transcribed by the enzyme RNA polymerase II (141). These pri-miRNAs can then

undergo RNA editing by adenosine deaminase acting on RNA (ADAR), which can convert adenosine into inosine and potentially modifies cleavage products in succeeding stages of miRNA processing (141, 143). Pre-miRNAs are subsequently produced following cleavage of pri-miRNAs by a microprocessor complex composed of an RNAse III enzyme, Drosha, and the protein DiGeorge syndrome critical region 8 (DGCR8) (141, 144, 145). After these processing steps, pre-miRNAs can then be exported out of the nucleus and into the cytosol through the involvement of exportin 5 (141). Subsequently, pre-miRNAs then undergo an additional modification step in which the RNAse III enzyme, Dicer, binds and cleaves the pre-miRNA to produce a double stranded mature-miRNA duplex (141). Once processed by Dicer, the mature miRNA duplex gets associated with endonucleases belonging to the Argonaute (Ago) family of proteins to form the pre-RNA-induced silencing complex (pre-RISC) (141, 146). Within the pre-RISC, one of the strands of the miRNA duplex, the "passenger strand", is removed, leaving the single stranded "guide strand" to form the mature RISC complex (146). The RISC complex can bind to the 3' UTR region of a target mRNA and regulate gene expression through either translation repression or mRNA decay (141, 147). Through these two mechanisms, miRNAs play a major role in regulating numerous genes and various cellular processes. It is estimated that the human genome encodes nearly 2,300 miRNAs, each of which targets about 100 mRNA transcripts (148).

The cellular effects of gonadal steroid hormones, including progesterone, estrogen, and testosterone, have been shown to be influenced by miRNAs. For instance, miRNAs have been shown to negatively regulate the expression of estrogen receptor- α and estrogen receptor- β (149–152), and a study by Epis et al., 2009, suggests that miR-331-3p regulates androgen receptor (AR)mediated signaling in prostate cancer cell lines (153). As it pertains specifically to progesterone, less is known regarding effects of miRNAs on progesterone in comparison to that of estrogen. However, studies have shown potential miRNA binding sites on progesterone receptor (PR) mRNA that can result in changes in PR expression in breast cancer cell lines and in endometrial carcinogenesis (154-157). Specifically, Gilam et al., 2017, showed that miR-181a, miR-23a, and miR-26b reduced PR expression in ER-positive breast cancer (158). In addition to the classical PR, the influence of miRNAs has been demonstrated on Pgrmc1. Wendler et al., 2010, found that the Pgrmc1 mRNA 3' untranslated region (UTR) contains a binding site for let7/miR-98, and accordingly, transfection of let-7i in SKOV-3 ovarian cancer cell lines resulted in a decrease in relative Pgrmc1 expression (159). Our laboratory also showed that Let-7i decreases the expression of Pgrmc1 (118). Apart from effects on receptor expression, the release of progesterone has also been shown to be regulated by miRNAs, as evidenced by the study of Sirotkin et al., 2009, where thirty six out of eighty tested miRNA constructs resulted in inhibition of progesterone release from granulosa cells (160, 161). Additionally, transfection of antisense constructs for two of the tested miRNAs resulted in an increase in progesterone release (160, 161). Apart from release, miRNAs have also been shown to regulate progesterone synthesis. Work from An et al., 2020, shows that that transfection of miR-101-3p promotes progesterone synthesis in granulosa cells via inhibition of STC1, which functions to inhibit hCG-stimulated progesterone synthesis, in goat granulosa cells (162). Furthermore, transfection of miR-101-3p results in an increase in mRNA and protein levels of cytochrome P450 family 11 subfamily A member 1 (Cyp11a1) and 3 β -hydroxysteroid dehydrogenase (3 β -HSD), two enzymes involved in the synthesis of progesterone from cholesterol (162). Collectively, these studies underscore the diagnostic potential of miRNAs in pathological conditions and illustrate miRNAs as a possible therapeutic target for the treatment of various cancers and diseases.

Work from our laboratory has highlighted the therapeutic potential of targeting miRNAs in experimental models of ischemic stroke. We have previously shown that Pgrmc1 is a key mediator in promoting the release of BDNF from cortical astrocytes (19, 163). Based on findings that the miRNA let-7i downregulates expression of Pgrmc1 in peripheral tissue (159), and that the antagomir to let-7f, a sister miRNA to let7i, is neuroprotective in models of ischemic stroke (164), we hypothesized that administration of an antagomir to let-7i, when combined with progesterone, could promote enhanced BDNF release and provide protection in experimental models of ischemic stroke (118). Our findings show that overexpression of let-7i results in downregulation of mRNA and protein levels of Pgrmc1 and BDNF expression in primary cortical astrocytes (118). Furthermore, let-7i was found to inhibit progesterone-mediated BDNF release (118). Additional experiments revealed that treatment of primary neurons with conditioned media from progesterone-treated astrocytes led to an increase in the synaptogenic marker, synaptophysin (SYP) (118). However, SYP expression was greatly diminished in neurons treated with conditioned media from astrocytes treated with let-7i and progesterone, but restored when treated with conditioned media from anti-let7i and progesterone treated astrocytes (118). These results were further validated using an animal model of ischemic stroke, the middle cerebral artery occlusion (MCAo) model, where animals treated with the combination of progesterone and the let-7i antagomir showed a robust increase in mature-BDNF protein levels, significantly reduced infarct sizes, enhanced SYP expression, and improved functional recovery compared to vehicle-treated animals (118). Collectively, these results suggest that inhibition of let-7i in experimental models of stroke promotes neuroprotection by increasing BDNF release, a process mediated by Pgrmc1. These significant findings pave the way for future studies in other disease models in which miRNA dysregulation has been implicated.

The influence of age on progesterone's brainprotective efficacy

Aging is related to a decline in circulating gonadal hormone levels, which is more pronounced in women due to menopause. While experimental evidence has shown that estrogen has neuroprotective effects against various insults in animal models, clinical trials such as the Women's Health Initiative (WHI) have yielded inconsistent results regarding the cognitive benefits of hormone therapy in postmenopausal women. The research conducted in the post-WHI era has yielded compelling evidence that supports the neuroprotective effects of estradiol in animal models of various CNS disorders, including Alzheimer's disease, Parkinson's disease, stroke, multiple sclerosis, chronic hypertension, and traumatic brain injury (165-171). Recent findings, however, have raised crucial questions about the circumstances in which hormone therapy after menopause can be beneficial. The effectiveness of hormone therapy may depend on various factors, such as the length of the postmenopausal period before hormone intervention and the subject's age (172-174). The concept of a therapeutic "window of opportunity" for hormone therapy after menopause has gained traction as a potential explanation for the differing outcomes observed in animal studies and clinical trials. Although the therapeutic potential of estrogens in the brain has been explored to some extent, there is a lack of research on whether a similar limited window of opportunity exists for progesterone and its related metabolite. The remarkable benefits of progesterone in reducing the size of stroke-induced lesions have been observed in young adult (3-month-old) ovariectomized (OVX) C57Bl/6 mice; however, its impact on the neurological outcome seems to be limited when tested on old (12-month-old) OVX mice (175). Such limited information warrants a more comprehensive evaluation of the brain-protective efficacy of progesterone as a function of age and in different models of brain aging and disease.

Summary

This review has offered information supporting the potential for progesterone as being protective in the brain and recognizing the regulation of BDNF and its associated signaling in this protection. A schematic of our overall working model by which progesterone elicits its neuroprotective efficacy, focusing significantly on BDNF and its associated signaling, and the factors that may alter progesterone's cytoprotective efficacy, is provided in Figure 1. While progesterone regulates the intracellular content of BDNF, the protein and mRNA, as an effect facilitated by the classical progesterone receptor, it also promotes the release of BDNF through the activation of a putative membrane progesterone receptor. Thus, the release of BDNF leads to the activation of TrkB receptors which are positioned on the surface of the adjacent cells, activating a cascade of pro-survival cell signaling pathways which include the ERK/MAPK and PI3/ Akt signaling pathways. Remarkably so, the ability of progestin to use neuroprotective properties seems to be associated with its ability to increase BDNF levels. Not only does this association support the important function of BDNF in these protective effects but emphasizes that not all progestins are made identically or equally, especially with respect to their impact on brain function. This awareness, including the key receptor targets of progesterone that relate to various mechanisms that support cell viability,



potentially direct future advancement and capability of developing more effective treatments for symptoms of the menopause, and those (neurodegenerative) diseases whose risk or incidence increases during the post-menopausal period.

Author contributions

neuroprotective effects.

MS: Writing – original draft, Writing – review & editing. VK: Writing – original draft, Writing – review & editing. SeK: Writing – original draft, Writing – review & editing. SaK: Writing – original draft, Writing – review & editing. HL: Writing – original draft, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This study

References

1. Strauss J, Barbieri R. Yen and Jaffe's reproductive endocrinology: Physiology, Pathophysiology, and Clinical Management, 5th edn. Philadelphia: Elsevier Saunders (2004). p. 1042.

was supported, in part, by a grant from the National Institute of Aging (AG027956).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The author(s) declared that they were an editorial board member of Frontiers, at the time of submission. This had no impact on the peer review process and the final decision.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

4. Kato J, Hirata S, Nozawa A, Mouri N. The ontogeny of gene expression of progestin receptors in the female rat brain. *J Steroid Biochem Mol Biol.* (1993) 47:173–82. doi: 10.1016/0960-0760(93)90072-5

5. Brinton RD, Thompson RF, Foy MR, Baudry M, Wang J, Finch CE, et al. Progesterone receptors: form and function in brain. *Front Neuroendocrinol.* (2008) 29:313–39. doi: 10.1016/j.yfrne.2008.02.001

6. Medina-Laver Y, Rodriguez-Varela C, Salsano S, Labarta E, Dominguez F. What do we know about classical and non-classical progesterone receptors in the human female reproductive tract? a review. *Int J Mol Sci.* (2021) 22(20):11278. doi: 10.3390/ijms222011278

^{2.} Arab H, Alharbi AJ, Oraif A, Sagr E, Al Madani H, Abduljabbar H, et al. The role of progestogens in threatened and idiopathic recurrent miscarriage. *Int J Womens Health*. (2019) 11:589–96. doi: 10.2147/IJWH.S224159

^{3.} MacLusky NJ, McEwen BS. Progestin receptors in rat brain: distribution and properties of cytoplasmic progestin-binding sites. *Endocrinology*. (1980) 106:192–202. doi: 10.1210/endo-106-1-192

7. Ke FC, Ramirez VD. Binding of progesterone to nerve cell membranes of rat brain using progesterone conjugated to 125I-bovine serum albumin as a ligand. *J Neurochem.* (1990) 54:467–72. doi: 10.1111/j.1471-4159.1990.tb01895.x

8. Towle AC, Sze PY. Steroid binding to synaptic plasma membrane: differential binding of glucocorticoids and gonadal steroids. *J Steroid Biochem*. (1983) 18:135–43. doi: 10.1016/0022-4731(83)90079-1

9. Zhu Y, Bond J, Thomas P. Identification, classification, and partial characterization of genes in humans and other vertebrates homologous to a fish membrane progestin receptor. *Proc Natl Acad Sci USA*. (2003) 100:2237–42. doi: 10.1073/pnas.0436133100

10. Zhu Y, Rice CD, Pang Y, Pace M, Thomas P. Cloning, expression, and characterization of a membrane progestin receptor and evidence it is an intermediary in meiotic maturation of fish oocytes. *Proc Natl Acad Sci USA*. (2003) 100:2231–6. doi: 10.1073/pnas.0336132100

11. Krebs CJ, Jarvis ED, Chan J, Lydon JP, Ogawa S, Pfaff DW. A membraneassociated progesterone-binding protein, 25-Dx, is regulated by progesterone in brain regions involved in female reproductive behaviors. *Proc Natl Acad Sci USA*. (2000) 97:12816–21. doi: 10.1073/pnas.97.23.12816

12. Meyer C, Schmid R, Scriba PC, Wehling M. Purification and partial sequencing of high-affinity progesterone-binding site(s) from porcine liver membranes. *Eur J Biochem.* (1996) 239:726–31. doi: 10.1111/j.1432-1033.1996.0726u.x

13. Falkenstein E, Schmieding K, Lange A, Meyer C, Gerdes D, Welsch U, et al. Localization of a putative progesterone membrane binding protein in porcine hepatocytes. *Cell Mol Biol (Noisy-le-grand)*. (1998) 44:571–8.

14. Min L, Takemori H, Nonaka Y, Katoh Y, Doi J, Horike N, et al. Characterization of the adrenal-specific antigen IZA (inner zone antigen) and its role in the steroidogenesis. *Mol Cell Endocrinol.* (2004) 215:143–8. doi: 10.1016/j.mce.2003.11.025

15. Pedroza DA, Rajamanickam V, Subramani R, Bencomo A, Galvez A, Lakshmanaswamy R. Progesterone receptor membrane component 1 promotes the growth of breast cancers by altering the phosphoproteome and augmenting EGFR/PI3K/AKT signalling. *Br J Cancer*. (2020) 123:1326–35. doi: 10.1038/s41416-020-0992-6

16. Sakamoto H, Ukena K, Takemori H, Okamoto M, Kawata M, Tsutsui K. Expression and localization of 25-Dx, a membrane-associated putative progesteronebinding protein, in the developing Purkinje cell. *Neuroscience*. (2004) 126:325–34. doi: 10.1016/j.neuroscience.2004.04.003

17. Mauvais-Jarvis F, Lange CA, Levin ER. Membrane-initiated estrogen, and rogen, and progesterone receptor signaling in health and disease. *Endocr Rev.* (2022) 43:720–42. doi: 10.1210/endrev/bnab041

 Jodhka PK, Kaur P, Underwood W, Lydon JP, Singh M. The differences in neuroprotective efficacy of progesterone and medroxyprogesterone acetate correlate with their effects on brain-derived neurotrophic factor expression. *Endocrinology*. (2009) 150:3162–8. doi: 10.1210/en.2008-1247

19. Su C, Cunningham RL, Rybalchenko N, Singh M. Progesterone increases the release of brain-derived neurotrophic factor from glia via progesterone receptor membrane component 1 (Pgrmc1)-dependent ERK5 signaling. *Endocrinology*. (2012) 153(9):4389–400. doi: 10.1210/en.2011-2177

20. Kaur P, Jodhka PK, Underwood WA, Bowles CA, de Fiebre NC, de Fiebre CM, et al. Progesterone increases brain-derived neuroptrophic factor expression and protects against glutamate toxicity in a mitogen-activated protein kinase- and phosphoinositide-3 kinase-dependent manner in cerebral cortical explants. *J Neurosci Res.* (2007) 85:2441–9. doi: 10.1002/jnr.21370

21. Nilsen J, Brinton RD. Impact of progestins on estrogen-induced neuroprotection: synergy by progesterone and 19-norprogesterone and antagonism by medroxyprogesterone acetate. *Endocrinology*. (2002) 143:205–12. doi: 10.1210/ endo.143.1.8582

22. Nilsen J, Brinton RD. Impact of progestins on estradiol potentiation of the glutamate calcium response. *Neuroreport.* (2002) 13:825–30. doi: 10.1097/00001756-200205070-00018

23. Nilsen J, Brinton RD. Divergent impact of progesterone and medroxyprogesterone acetate (Provera) on nuclear mitogen-activated protein kinase signaling. *Proc Natl Acad Sci USA*. (2003) 100:10506–11. doi: 10.1073/pnas.1334098100

24. Goodman Y, Bruce AJ, Cheng B, Mattson MP. Estrogens attenuate and corticosterone exacerbates excitotoxicity, oxidative injury, and amyloid beta-peptide toxicity in hippocampal neurons. *J Neurochem.* (1996) 66:1836–44. doi: 10.1046/j.1471-4159.1996.66051836.x

25. Jiang N, Chopp M, Stein D, Feit H. Progesterone is neuroprotective after transient middle cerebral artery occlusion in male rats. *Brain Res.* (1996) 735:101–7. doi: 10.1016/0006-8993(96)00605-1

26. Kumon Y, Kim SC, Tompkins P, Stevens A, Sakaki S, Loftus CM. Neuroprotective effect of postischemic administration of progesterone in spontaneously hypertensive rats with focal cerebral ischemia. *J Neurosurg.* (2000) 92:848–52. doi: 10.3171/jns.2000.92.5.0848

27. Morali G, Letechipia-Vallejo G, Lopez-Loeza E, Montes P, Hernandez-Morales L, Cervantes M. Post-ischemic administration of progesterone in rats exerts neuroprotective effects on the hippocampus. *Neurosci Lett.* (2005) 382:286–90. doi: 10.1016/j.neulet.2005.03.066

28. Frechou M, Zhang S, Liere P, Delespierre B, Soyed N, Pianos A, et al. Intranasal delivery of progesterone after transient ischemic stroke decreases mortality and

provides neuroprotection. Neuropharmacology. (2015) 97:394-403. doi: 10.1016/j.neuropharm.2015.06.002

29. Chen J, Chopp M, Li Y. Neuroprotective effects of progesterone after transient middle cerebral artery occlusion in rat. *J Neurol Sci.* (1999) 171:24–30. doi: 10.1016/S0022-510X(99)00247-6

30. Allen RS, Olsen TW, Sayeed I, Cale HA, Morrison KC, Oumarbaeva Y, et al. Progesterone treatment in two rat models of ocular ischemia. *Invest Ophthalmol Vis Sci.* (2015) 56:2880–91. doi: 10.1167/iovs.14-16070

31. Pettus EH, Wright DW, Stein DG, Hoffman SW. Progesterone treatment inhibits the inflammatory agents that accompany traumatic brain injury. *Brain Res.* (2005) 1049:112–9. doi: 10.1016/j.brainres.2005.05.004

32. Roof RL, Duvdevani R, Braswell L, Stein DG. Progesterone facilitates cognitive recovery and reduces secondary neuronal loss caused by cortical contusion injury in male rats. *Exp Neurol.* (1994) 129:64–9. doi: 10.1006/exnr.1994.1147

33. Roof RL, Hall ED. Gender differences in acute CNS trauma and stroke: neuroprotective effects of estrogen and progesterone. *J Neurotrauma*. (2000) 17:367–88. doi: 10.1089/neu.2000.17.367

34. Qin Y, Chen Z, Han X, Wu H, Yu Y, Wu J, et al. Progesterone attenuates Abeta (25-35)-induced neuronal toxicity via JNK inactivation and progesterone receptor membrane component 1-dependent inhibition of mitochondrial apoptotic pathway. *J Steroid Biochem Mol Biol.* (2015) 154:302–11. doi: 10.1016/j.jsbmb.2015.01.002

35. Frye CA, Walf AA. Effects of progesterone administration and APPswe +PSEN1Deltae9 mutation for cognitive performance of mid-aged mice. *Neurobiol Learn Mem.* (2008) 89:17–26. doi: 10.1016/j.nlm.2007.09.008

36. Wu H, Wu ZG, Shi WJ, Gao H, Wu HH, Bian F, et al. Effects of progesterone on glucose uptake in neurons of Alzheimer's disease animals and cell models. *Life Sci.* (2019) 238:116979. doi: 10.1016/j.lfs.2019.116979

37. Litim N, Morissette M, Di Paolo T. Effects of progesterone administered after MPTP on dopaminergic neurons of male mice. *Neuropharmacology*. (2017) 117:209–18. doi: 10.1016/j.neuropharm.2017.02.007

38. Bourque M, Morissette M, Al Sweidi S, Caruso D, Melcangi RC, Di Paolo T. Neuroprotective effect of progesterone in MPTP-treated male mice. *Neuroendocrinology.* (2016) 103:300-14. doi: 10.1159/000438789

39. Thomas AJ, Nockels RP, Pan HQ, Shaffrey CI, Chopp M. Progesterone is neuroprotective after acute experimental spinal cord trauma in rats. *Spine*. (1999) 24:2134–8. doi: 10.1097/00007632-199910150-00013

40. Gonzalez Deniselle MC, Lopez Costa JJ, Gonzalez SL, Labombarda F, Garay L, Guennoun R, et al. Basis of progesterone protection in spinal cord neurodegeneration. *J Steroid Biochem Mol Biol.* (2002) 83:199–209. doi: 10.1016/S0960-0760(02)00262-5

41. Gonzalez Deniselle MC, Lopez-Costa JJ, Saavedra JP, Pietranera L, Gonzalez SL, Garay L, et al. Progesterone neuroprotection in the Wobbler mouse, a genetic model of spinal cord motor neuron disease. *Neurobiol Dis.* (2002) 11:457–68. doi: 10.1006/ nbdi.2002.0564

42. Ibanez C, Shields SA, El-Etr M, Leonelli E, Magnaghi V, Li WW, et al. Steroids and the reversal of age-associated changes in myelination and remyelination. *Prog Neurobiol.* (2003) 71:49–56. doi: 10.1016/j.pneurobio.2003.09.002

43. Wright DW, Kellermann AL, Hertzberg VS, Clark PL, Frankel M, Goldstein FC, et al. ProTECT: a randomized clinical trial of progesterone for acute traumatic brain injury. *Ann Emerg Med.* (2007) 49:391–402. doi: 10.1016/j.annemergmed.2006.07.932

44. Ma J, Huang S, Qin S, You C, Zeng Y. Progesterone for acute traumatic brain injury. *Cochrane Database Syst Rev.* (2016) 12(12):CD008409. doi: 10.1002/14651858.CD008409.pub4

45. Vandromme M, Melton SM, Kerby JD. Progesterone in traumatic brain injury: time to move on to phase III trials. *Crit Care*. (2008) 12:153. doi: 10.1186/cc6899

46. Wali B, Sayeed I, Stein DG. Improved behavioral outcomes after progesterone administration in aged male rats with traumatic brain injury. *Restor Neurol Neurosci.* (2011) 29:61–71. doi: 10.3233/RNN-2011-0579

47. Xiao G, Wei J, Yan W, Wang W, Lu Z. Improved outcomes from the administration of progesterone for patients with acute severe traumatic brain injury: a randomized controlled trial. *Crit Care*. (2008) 12:R61. doi: 10.1186/cc6887

48. Howard RB, Sayeed I, Stein DG. Suboptimal dosing parameters as possible factors in the negative phase III clinical trials of progesterone for traumatic brain injury. *J Neurotrauma*. (2017) 34:1915–8. doi: 10.1089/neu.2015.4179

49. Wright DW, Yeatts SD, Silbergleit R, Palesch YY, Hertzberg VS, Frankel M, et al. Very early administration of progesterone for acute traumatic brain injury. *N Engl J Med.* (2014) 371:2457–66. doi: 10.1056/NEJMoa1404304

50. Soltani Z, Shahrokhi N, Karamouzian S, Khaksari M, Mofid B, Nakhaee N, et al. Does progesterone improve outcome in diffuse axonal injury? *Brain Inj.* (2017) 31:16–23. doi: 10.1080/02699052.2016.1213421

51. Lorenz L, Dang J, Misiak M, Tameh Abolfazl A, Beyer C, Kipp M. Combined 17beta-oestradiol and progesterone treatment prevents neuronal cell injury in cortical but not midbrain neurones or neuroblastoma cells. *J Neuroendocrinol.* (2009) 21:841–9. doi: 10.1111/j.1365-2826.2009.01903.x

52. Mannella P, Sanchez AM, Giretti MS, Genazzani AR, Simoncini T. Oestrogen and progestins differently prevent glutamate toxicity in cortical neurons depending on prior hormonal exposure via the induction of neural nitric oxide synthase. *Steroids*. (2009) 74:650–6. doi: 10.1016/j.steroids.2009.02.011

53. Aguirre C, Jayaraman A, Pike C, Baudry M. Progesterone inhibits estrogenmediated neuroprotection against excitotoxicity by down-regulating estrogen receptorbeta. *J Neurochem*. (2010) 115:1277–87. doi: 10.1111/j.1471-4159.2010.07038.x

54. Aguirre CC, Baudry M. Progesterone reverses 17beta-estradiol-mediated neuroprotection and BDNF induction in cultured hippocampal slices. *Eur J Neurosci.* (2009) 29:447–54. doi: 10.1111/j.1460-9568.2008.06591.x

55. Carroll JC, Rosario ER, Pike CJ. Progesterone blocks estrogen neuroprotection from kainate in middle-aged female rats. *Neurosci Lett.* (2008) 445:229–32. doi: 10.1016/j.neulet.2008.09.010

56. Jayaraman A, Pike CJ. Progesterone attenuates oestrogen neuroprotection via downregulation of oestrogen receptor expression in cultured neurones. *J Neuroendocrinol.* (2009) 21:77-81. doi: 10.1111/j.1365-2826.2008.01801.x

57. Rosario ER, Ramsden M, Pike CJ. Progestins inhibit the neuroprotective effects of estrogen in rat hippocampus. *Brain Res.* (2006) 1099:206–10. doi: 10.1016/ j.brainres.2006.03.127

58. Yao J, Chen S, Cadenas E, Brinton RD. Estrogen protection against mitochondrial toxin-induced cell death in hippocampal neurons: antagonism by progesterone. *Brain Res.* (2011) 1379:2-10. doi: 10.1016/j.brainres.2010.11.090

59. Murphy DD, Segal M. Progesterone prevents estradiol-induced dendritic spine formation in cultured hippocampal neurons. *Neuroendocrinology*. (2000) 72:133–43. doi: 10.1159/000054580

60. McEwen BS, Woolley CS. Estradiol and progesterone regulate neuronal structure and synaptic connectivity in adult as well as developing brain. *Exp Gerontol.* (1994) 29:431–6. doi: 10.1016/0531-5565(94)90022-1

61. Woolley CS, McEwen BS. Roles of estradiol and progesterone in regulation of hippocampal dendritic spine density during the estrous cycle in the rat. *J Comp Neurol.* (1993) 336:293–306. doi: 10.1002/cne.903360210

62. Zhao Y, Wang J, Liu C, Jiang C, Zhao C, Zhu Z. Progesterone influences postischemic synaptogenesis in the CA1 region of the hippocampus in rats. *Synapse*. (2011) 65:880–91. doi: 10.1002/syn.20915

63. Foy MR, Akopian G, Thompson RF. Progesterone regulation of synaptic transmission and plasticity in rodent hippocampus. *Learn Mem.* (2008) 15:820–2. doi: 10.1101/lm.1124708

64. Cohen S, Greenberg ME. Communication between the synapse and the nucleus in neuronal development, plasticity, and disease. *Annu Rev Cell Dev Biol.* (2008) 24:183–209. doi: 10.1146/annurev.cellbio.24.110707.175235

65. Lu Y, Christian K, Lu B. BDNF: a key regulator for protein synthesis-dependent LTP and long-term memory? *Neurobiol Learn Mem.* (2008) 89:312–23. doi: 10.1016/j.nlm.2007.08.018

66. Causing CG, Gloster A, Aloyz R, Bamji SX, Chang E, Fawcett J, et al. Synaptic innervation density is regulated by neuron-derived BDNF. *Neuron*. (1997) 18:257–67. doi: 10.1016/S0896-6273(00)80266-4

67. Pruginin-Bluger M, Shelton DL, Kalcheim C. A paracrine effect for neuronderived BDNF in development of dorsal root ganglia: stimulation of Schwann cell myelin protein expression by glial cells. *Mech Dev.* (1997) 61:99–111. doi: 10.1016/ S0925-4773(96)00623-5

68. Matsumoto T, Rauskolb S, Polack M, Klose J, Kolbeck R, Korte M, et al. Biosynthesis and processing of endogenous BDNF: CNS neurons store and secrete BDNF, not pro-BDNF. *Nat Neurosci.* (2008) 11:131–3. doi: 10.1038/nn2038

69. Lu B, Pang PT, Woo NH. The yin and yang of neurotrophin action. *Nat Rev Neurosci.* (2005) 6:603–14. doi: 10.1038/nrn1726

70. Yoshii A, Constantine-Paton M. Postsynaptic BDNF-TrkB signaling in synapse maturation, plasticity, and disease. *Dev Neurobiol.* (2010) 70:304–22. doi: 10.1002/ dneu.20765

71. Minichiello L. TrkB signalling pathways in LTP and learning. *Nat Rev Neurosci.* (2009) 10:850–60. doi: 10.1038/nrn2738

72. Hock C, Heese K, Muller-Spahn F, Hulette C, Rosenberg C, Otten U. Decreased trkA neurotrophin receptor expression in the parietal cortex of patients with Alzheimer's disease. *Neurosci Lett.* (1998) 241:151–4. doi: 10.1016/S0304-3940(98)00019-6

73. Phillips HS, Hains JM, Armanini M, Laramee GR, Johnson SA, Winslow JW. BDNF mRNA is decreased in the hippocampus of individuals with Alzheimer's disease. *Neuron*. (1991) 7:695–702. doi: 10.1016/0896-6273(91)90273-3

74. Connor B, Young D, Yan Q, Faull RL, Synek B, Dragunow M. Brain-derived neurotrophic factor is reduced in Alzheimer's disease. *Brain Res Mol Brain Res.* (1997) 49:71–81. doi: 10.1016/S0169-328X(97)00125-3

75. Tanila H. The role of BDNF in Alzheimer's disease. *Neurobiol Dis.* (2017) 97:114-8. doi: 10.1016/j.nbd.2016.05.008

76. Fahnestock M, Michalski B, Xu B, Coughlin MD. The precursor pro-nerve growth factor is the predominant form of nerve growth factor in brain and is increased in Alzheimer's disease. *Mol Cell Neurosci.* (2001) 18:210–20. doi: 10.1006/mcne.2001.1016

77. Murer MG, Yan Q, Raisman-Vozari R. Brain-derived neurotrophic factor in the control human brain, and in Alzheimer's disease and Parkinson's disease. *Prog Neurobiol.* (2001) 63:71–124. doi: 10.1016/S0301-0082(00)00014-9

78. Kowianski P, Lietzau G, Czuba E, Waskow M, Steliga A, Morys J. BDNF: A key factor with multipotent impact on brain signaling and synaptic plasticity. *Cell Mol Neurobiol.* (2018) 38:579–93. doi: 10.1007/s10571-017-0510-4

79. Ferrer I, Marin C, Rey MJ, Ribalta T, Goutan E, Blanco R, et al. BDNF and fulllength and truncated TrkB expression in Alzheimer disease. Implications in therapeutic strategies. J Neuropathol Exp Neurol. (1999) 58:729–39. doi: 10.1097/00005072-199907000-00007

80. Tapia-Arancibia L, Aliaga E, Silhol M, Arancibia S. New insights into brain BDNF function in normal aging and Alzheimer disease. *Brain Res Rev.* (2008) 59:201–20. doi: 10.1016/j.brainresrev.2008.07.007

81. Zuccato C, Cattaneo E. Brain-derived neurotrophic factor in neurodegenerative diseases. *Nat Rev Neurol.* (2009) 5:311–22. doi: 10.1038/nrneurol.2009.54

82. Cook DJ, Nguyen C, Chun HN, Chiu AS, Machnicki M, Zarembinski TI, et al. Hydrogel-delivered brain-derived neurotrophic factor promotes tissue repair and recovery after stroke. *J Cereb Blood Flow Metab.* (2017) 37:1030–45. doi: 10.1177/0271678X16649964

83. Harris NM, Ritzel R, Mancini NS, Jiang Y, Yi X, Manickam DS, et al. Nanoparticle delivery of brain derived neurotrophic factor after focal cerebral ischemia reduces tissue injury and enhances behavioral recovery. *Pharmacol Biochem Behav*. (2016) 150-151:48–56. doi: 10.1016/j.pbb.2016.09.003

84. Lazarovici P, Cohen G, Arien-Zakay H, Chen J, Zhang C, Chopp M, et al. Multimodal neuroprotection induced by PACAP38 in oxygen-glucose deprivation and middle cerebral artery occlusion stroke models. *J Mol Neurosci MN*. (2012) 48:526–40. doi: 10.1007/s12031-012-9818-1

85. Ishrat T, Sayeed I, Atif F, Hua F, Stein DG. Progesterone is neuroprotective against ischemic brain injury through its effects on the phosphoinositide 3-kinase/ protein kinase B signaling pathway. *Neuroscience.* (2012) 210:442–50. doi: 10.1016/ j.neuroscience.2012.03.008

86. Zhang J, Yu Z, Yu Z, Yang Z, Zhao H, Liu L, et al. rAAV-mediated delivery of brain-derived neurotrophic factor promotes neurite outgrowth and protects neurodegeneration in focal ischemic model. *Int J Clin Exp Pathol.* (2011) 4:496–504.

87. Ploughman M, Windle V, MacLellan CL, White N, Dore JJ, Corbett D. Brainderived neurotrophic factor contributes to recovery of skilled reaching after focal ischemia in rats. *Stroke*. (2009) 40:1490–5. doi: 10.1161/STROKEAHA.108.531806

88. Chen J, Zacharek A, Zhang C, Jiang H, Li Y, Roberts C, et al. Endothelial nitric oxide synthase regulates brain-derived neurotrophic factor expression and neurogenesis after stroke in mice. *J Neurosci.* (2005) 25:2366–75. doi: 10.1523/JNEUROSCL5071-04.2005

89. Bejot Y, Prigent-Tessier A, Cachia C, Giroud M, Mossiat C, Bertrand N, et al. Time-dependent contribution of non neuronal cells to BDNF production after ischemic stroke in rats. *Neurochem Int.* (2011) 58:102–11. doi: 10.1016/j.neuint.2010.10.019

90. Kaplan GB, Vasterling JJ, Vedak PC. Brain-derived neurotrophic factor in traumatic brain injury, post-traumatic stress disorder, and their comorbid conditions: role in pathogenesis and treatment. *Behav Pharmacol.* (2010) 21:427–37. doi: 10.1097/FBP.0b013e32833d8bc9

91. Gao X, Chen J. Conditional knockout of brain-derived neurotrophic factor in the hippocampus increases death of adult-born immature neurons following traumatic brain injury. *J Neurotrauma*. (2009) 26:1325–35. doi: 10.1089/neu.2008.0744

92. Wu H, Lu D, Jiang H, Xiong Y, Qu C, Li B, et al. Simvastatin-mediated upregulation of VEGF and BDNF, activation of the PI3K/Akt pathway, and increase of neurogenesis are associated with therapeutic improvement after traumatic brain injury. *J Neurotrauma*. (2008) 25:130–9. doi: 10.1089/neu.2007.0369

93. Griesbach GS, Hovda DA, Molteni R, Wu A, Gomez-Pinilla F. Voluntary exercise following traumatic brain injury: brain-derived neurotrophic factor upregulation and recovery of function. *Neuroscience*. (2004) 125:129-39. doi: 10.1016/j.neuroscience.2004.01.030

94. Zheng Z, Sabirzhanov B, Keifer J. Oligomeric amyloid-beta inhibits the proteolytic conversion of brain-derived neurotrophic factor (BDNF), AMPA receptor trafficking, and classical conditioning. *J Biol Chem.* (2010) 285:34708–17. doi: 10.1074/ jbc.M110.150821

95. Nagahara AH, Merrill DA, Coppola G, Tsukada S, Schroeder BE, Shaked GM, et al. Neuroprotective effects of brain-derived neurotrophic factor in rodent and primate models of Alzheimer's disease. *Nat Med.* (2009) 15:331–7. doi: 10.1038/nm.1912

96. Hetman M, Kanning K, Cavanaugh JE, Xia Z. Neuroprotection by brain-derived neurotrophic factor is mediated by extracellular signal-regulated kinase and phosphatidylinositol 3-kinase. *J Biol Chem.* (1999) 274:22569–80. doi: 10.1074/jbc.274.32.22569

97. Han BH, Holtzman DM. BDNF protects the neonatal brain from hypoxicischemic injury in *vivo* via the ERK pathway. *J Neurosci.* (2000) 20:5775-81. doi: 10.1523/INEUROSCI.20-15-05775.2000

98. Singh M. Ovarian hormones elicit phosphorylation of Akt and extracellularsignal regulated kinase in explants of the cerebral cortex. *Endocrine*. (2001) 14:407–15. doi: 10.1385/ENDO:14:3:407

99. Singh M, Meyer EM, Simpkins JW. The effect of ovariectomy and estradiol replacement on brain-derived neurotrophic factor messenger ribonucleic acid expression in cortical and hippocampal brain regions of female Sprague-Dawley rats. *Endocrinology*. (1995) 136:2320–4. doi: 10.1210/endo.136.5.7720680

100. Gonzalez Deniselle MC, Garay L, Gonzalez S, Saravia F, Labombarda F, Guennoun R, et al. Progesterone modulates brain-derived neurotrophic factor and choline acetyltransferase in degenerating Wobbler motoneurons. *Exp Neurol.* (2007) 203:406–14. doi: 10.1016/j.expneurol.2006.08.019

101. Gonzalez SL, Labombarda F, Gonzalez Deniselle MC, Guennoun R, Schumacher M, De Nicola AF. Progesterone up-regulates neuronal brain-derived neurotrophic factor expression in the injured spinal cord. *Neuroscience*. (2004) 125:605–14. doi: 10.1016/j.neuroscience.2004.02.024

102. Meyer M, Gonzalez Deniselle MC, Gargiulo-Monachelli G, Lima A, Roig P, Guennoun R, et al. Progesterone attenuates several hippocampal abnormalities of the Wobbler mouse. *J Neuroendocrinol.* (2013) 25:235–43. doi: 10.1111/jne.12004

103. Ghadiri T, Vakilzadeh G, Hajali V, Khodagholi F. Progesterone modulates post-traumatic epileptogenesis through regulation of BDNF-TrkB signaling and cell survival-related pathways in the rat hippocampus. *Neurosci Lett.* (2019) 709:134384. doi: 10.1016/j.neulet.2019.134384

104. Castelnovo LF, Thomas P. Membrane progesterone receptor alpha (mPRalpha/ PAQR7) promotes survival and neurite outgrowth of human neuronal cells by a direct action and through Schwann cell-like stem cells. *J Mol Neurosci.* (2022) 72:2067–80. doi: 10.1007/s12031-022-02057-z

105. Schuman EM. Neurotrophin regulation of synaptic transmission. Curr Opin Neurobiol. (1999) 9:105–9. doi: 10.1016/S0959-4388(99)80013-0

106. Leal G, Comprido D, Duarte CB. BDNF-induced local protein synthesis and synaptic plasticity. *Neuropharmacology*. (2014) 76 Pt C:639–56. doi: 10.1016/j.neuropharm.2013.04.005

107. Bliss TV, Collingridge GL. A synaptic model of memory: long-term potentiation in the hippocampus. *Nature*. (1993) 361:31-9. doi: 10.1038/361031a0

108. Nguyen PV, Kandel ER. A macromolecular synthesis-dependent late phase of long-term potentiation requiring cAMP in the medial perforant pathway of rat hippocampal slices. *J Neurosci.* (1996) 16:3189–98. doi: 10.1523/JNEUROSCI.16-10-03189.1996

109. Panja D, Bramham CR. BDNF mechanisms in late LTP formation: A synthesis and breakdown. *Neuropharmacology*. (2014) 76 Pt C:664-76. doi: 10.1016/j.neuropharm.2013.06.024

110. Suen PC, Wu K, Levine ES, Mount HT, Xu JL, Lin SY, et al. Brain-derived neurotrophic factor rapidly enhances phosphorylation of the postsynaptic N-methyl-D-aspartate receptor subunit 1. *Proc Natl Acad Sci USA*. (1997) 94:8191–5. doi: 10.1073/pnas.94.15.8191

111. Lin SY, Wu K, Levine ES, Mount HT, Suen PC, Black IB. BDNF acutely increases tyrosine phosphorylation of the NMDA receptor subunit 2B in cortical and hippocampal postsynaptic densities. *Brain Res Mol Brain Res.* (1998) 55:20–7. doi: 10.1016/S0169-328X(97)00349-5

112. Levine ES, Kolb JE. Brain-derived neurotrophic factor increases activity of NR2B-containing N-methyl-D-aspartate receptors in excised patches from hippocampal neurons. J Neurosci Res. (2000) 62:357–62. doi: 10.1002/1097-4547 (20001101)62:3<357::AID-JNR5>3.0.CO;2-6

113. Crozier RA, Black IB, Plummer MR. Blockade of NR2B-containing NMDA receptors prevents BDNF enhancement of glutamatergic transmission in hippocampal neurons. *Learn Mem.* (1999) 6:257–66. doi: 10.1101/lm.6.3.257

114. Xu F, Plummer MR, Len GW, Nakazawa T, Yamamoto T, Black IB, et al. Brainderived neurotrophic factor rapidly increases NMDA receptor channel activity through Fynmediated phosphorylation. *Brain Res.* (2006) 1121:22–34. doi: 10.1016/j.brainres.2006.08.129

115. Ying SW, Futter M, Rosenblum K, Webber MJ, Hunt SP, Bliss TV, et al. Brainderived neurotrophic factor induces long-term potentiation in intact adult hippocampus: requirement for ERK activation coupled to CREB and upregulation of Arc synthesis. *J Neurosci.* (2002) 22:1532–40. doi: 10.1523/JNEUROSCI.22-05-01532.2002

116. Chen L, Miyamoto Y, Furuya K, Dai XN, Mori N, Sokabe M. Chronic DHEAS administration facilitates hippocampal long-term potentiation via an amplification of Src-dependent NMDA receptor signaling. *Neuropharmacology*. (2006) 51:659–70. doi: 10.1016/j.neuropharm.2006.05.011

117. Cekic M, Johnson SJ, Bhatt VH, Stein DG. Progesterone treatment alters neurotrophin/proneurotrophin balance and receptor expression in rats with traumatic brain injury. *Restor Neurol Neurosci.* (2012) 30:115–26. doi: 10.3233/RNN-2011-0628

118. Nguyen T, Su C, Singh M. Let-7i inhibition enhances progesterone-induced functional recovery in a mouse model of ischemia. *Proc Natl Acad Sci USA*. (2018) 115: E9668–77. doi: 10.1073/pnas.1803384115

119. Djebaili M, Hoffman SW, Stein DG. Allopregnanolone and progesterone decrease cell death and cognitive deficits after a contusion of the rat pre-frontal cortex. *Neuroscience.* (2004) 123:349–59. doi: 10.1016/j.neuroscience.2003.09.023

120. Ciriza I, Azcoitia I, Garcia-Segura LM. Reduced progesterone metabolites protect rat hippocampal neurones from kainic acid excitotoxicity in vivo. J Neuroendocrinol. (2004) 16:58–63. doi: 10.1111/j.1365-2826.2004.01121.x

121. Ardeshiri A, Kelley MH, Korner IP, Hurn PD, Herson PS. Mechanism of progesterone neuroprotection of rat cerebellar Purkinje cells following oxygen-glucose deprivation. *Eur J Neurosci.* (2006) 24:2567–74. doi: 10.1111/j.1460-9568.2006.05142.x

122. He J, Hoffman SW, Stein DG. Allopregnanolone, a progesterone metabolite, enhances behavioral recovery and decreases neuronal loss after traumatic brain injury. *Restor Neurol Neurosci.* (2004) 22:19–31.

123. Sayeed I, Parvez S, Wali B, Siemen D, Stein DG. Direct inhibition of the mitochondrial permeability transition pore: a possible mechanism for better neuroprotective effects of allopregnanolone over progesterone. *Brain Res.* (2009) 1263:165–73. doi: 10.1016/j.brainres.2009.01.045

124. Wang JM, Liu L, Irwin RW, Chen S, Brinton RD. Regenerative potential of allopregnanolone. *Brain Res Rev.* (2008) 57:398-409. doi: 10.1016/j.brainresrev.2007.08.010

125. Nin MS, Martinez LA, Pibiri F, Nelson M, Pinna G. Neurosteroids reduce social isolation-induced behavioral deficits: a proposed link with neurosteroid-mediated upregulation of BDNF expression. *Front Endocrinol.* (2011) 2:73. doi: 10.3389/fendo.2011.00073

126. Naert G, Maurice T, Tapia-Arancibia L, Givalois L. Neuroactive steroids modulate HPA axis activity and cerebral brain-derived neurotrophic factor (BDNF) protein levels in adult male rats. *Psychoneuroendocrinology*. (2007) 32:1062–78. doi: 10.1016/j.psyneuen.2007.09.002

127. Chen S, Wang JM, Irwin RW, Yao J, Liu L, Brinton RD. Allopregnanolone promotes regeneration and reduces beta-amyloid burden in a preclinical model of Alzheimer's disease. *PLoS One.* (2011) 6:e24293. doi: 10.1371/journal.pone.0024293

128. Irwin RW, Solinsky CM, Brinton RD. Frontiers in therapeutic development of allopregnanolone for Alzheimer's disease and other neurological disorders. *Front Cell Neurosci.* (2014) 8:203. doi: 10.3389/fncel.2014.00203

129. Irwin RW, Wang JM, Chen S, Brinton RD. Neuroregenerative mechanisms of allopregnanolone in Alzheimer's disease. *Front Endocrinol.* (2011) 2:117. doi: 10.3389/ fendo.2011.00117

130. Wang T, Yao J, Chen S, Mao Z, Brinton RD. Allopregnanolone reverses bioenergetic deficits in female triple transgenic Alzheimer's mouse model. *Neurotherapeutics*. (2020) 17:178-88. doi: 10.1007/s13311-019-00793-6

131. Thomas P, Pang Y. Anti-apoptotic actions of allopregnanolone and ganaxolone mediated through membrane progesterone receptors (PAQRs) in neuronal cells. *Front Endocrinol.* (2020) 11:417. doi: 10.3389/fendo.2020.00417

132. Roglio I, Bianchi R, Gotti S, Scurati S, Giatti S, Pesaresi M, et al. Neuroprotective effects of dihydroprogesterone and progesterone in an experimental model of nerve crush injury. *Neuroscience*. (2008) 155:673-85. doi: 10.1016/j.neuroscience.2008.06.034

133. Luchetti S, Liere P, Pianos A, Verwer RWH, Sluiter A, Huitinga I, et al. Disease stage-dependent changes in brain levels and neuroprotective effects of neuroactive steroids in Parkinson's disease. *Neurobiol Dis.* (2023) 183:106169. doi: 10.1016/j.nbd.2023.106169

134. Hirvonen E. Progestins. *Maturitas*. (1996) 23 Suppl:S13-8. doi: 10.1016/0378-5122(96)01005-5

135. Gambrell RD Jr. The role of hormones in the etiology and prevention of endometrial cancer. *Clin Obstet Gynaecol.* (1986) 13:695–723.

136. Miyagawa K, Vidgoff J, Hermsmeyer K. Ca2+ release mechanism of primate drug-induced coronary vasospasm. *Am J Physiol.* (1997) 272:H2645–54. doi: 10.1152/ ajpheart.1997.272.6.H2645

137. Rosano GM, Webb CM, Chierchia S, Morgani GL, Gabraele M, Sarrel PM, et al. Natural progesterone, but not medroxyprogesterone acetate, enhances the beneficial effect of estrogen on exercise-induced myocardial ischemia in postmenopausal women. *J Am Coll Cardiol.* (2000) 36:2154–9. doi: 10.1016/S0735-1097(00)01007-X

138. Nelson AL. Comprehensive overview of the recently FDA-approved contraceptive vaginal ring releasing segesterone acetate and ethinylestradiol: A new year-long, patient controlled, reversible birth control method. *Expert Rev Clin Pharmacol.* (2019) 12:953–63. doi: 10.1080/17512433.2019.1669448

139. Tanaka M, Ogaeri T, Samsonov M, Sokabe M. Nestorone exerts long-term neuroprotective effects against transient focal cerebral ischemia in adult male rats. *Brain Res.* (2019) 1719:288–96. doi: 10.1016/j.brainres.2018.09.022

140. Frechou M, Zhu X, Kumar N, Sitruk-Ware R, Schumacher M, Mattern C, et al. Sex differences in the cerebroprotection by Nestorone intranasal delivery following stroke in mice. *Neuropharmacology*. (2021) 198:108760. doi: 10.1016/ ineuropharm.2021.108760

141. Gebert LFR, MacRae IJ. Regulation of microRNA function in animals. *Nat Rev Mol Cell Biol.* (2019) 20:21–37. doi: 10.1038/s41580-018-0045-7

142. Wightman B, Ha I, Ruvkun G. Posttranscriptional regulation of the heterochronic gene lin-14 by lin-4 mediates temporal pattern formation in C. *elegans Cell.* (1993) 75:855–62. doi: 10.1016/0092-8674(93)90530-4

143. Nishikura K. A-to-I editing of coding and non-coding RNAs by ADARs. Nat Rev Mol Cell Biol. (2016) 17:83–96. doi: 10.1038/nrm.2015.4

144. Denli AM, Tops BB, Plasterk RH, Ketting RF, Hannon GJ. Processing of primary microRNAs by the Microprocessor complex. *Nature*. (2004) 432:231–5. doi: 10.1038/nature03049

145. Gregory RI, Yan KP, Amuthan G, Chendrimada T, Doratotaj B, Cooch N, et al. The Microprocessor complex mediates the genesis of microRNAs. *Nature*. (2004) 432:235–40. doi: 10.1038/nature03120

146. Ha M, Kim VN. Regulation of microRNA biogenesis. Nat Rev Mol Cell Biol. (2014) 15:509-24. doi: 10.1038/nrm3838

147. Iwakawa HO, Tomari Y. The functions of microRNAs: mRNA decay and translational repression. *Trends Cell Biol.* (2015) 25:651-65. doi: 10.1016/j.tcb.2015.07.011

148. Alles J, Fehlmann T, Fischer U, Backes C, Galata V, Minet M, et al. An estimate of the total number of true human miRNAs. *Nucleic Acids Res.* (2019) 47:3353–64. doi: 10.1093/nar/gkz097

149. Adams BD, Furneaux H, White BA. The micro-ribonucleic acid (miRNA) miR-206 targets the human estrogen receptor-alpha (ERalpha) and represses ERalpha messenger RNA and protein expression in breast cancer cell lines. *Mol Endocrinol.* (2007) 21:1132–47. doi: 10.1210/me.2007-0022

150. Pandey DP, Picard D. miR-22 inhibits estrogen signaling by directly targeting the estrogen receptor alpha mRNA. *Mol Cell Biol.* (2009) 29:3783–90. doi: 10.1128/ MCB.01875-08

151. Al-Nakhle H, Burns PA, Cummings M, Hanby AM, Hughes TA, Satheesha S, et al. Estrogen receptor beta1 expression is regulated by miR-92 in breast cancer. *Cancer Res.* (2010) 70:4778–84. doi: 10.1158/0008-5472.CAN-09-4104

152. Kalinina T, Kononchuk V, Alekseenok E, Obukhova D, Sidorov S, Strunkin D, et al. Expression of estrogen receptor- and progesterone receptor-regulating microRNAs in breast cancer. *Genes (Basel).* (2021) 12(4):582. doi: 10.3390/genes12040582

153. Epis MR, Giles KM, Barker A, Kendrick TS, Leedman PJ. miR-331-3p regulates ERBB-2 expression and androgen receptor signaling in prostate cancer. *J Biol Chem.* (2009) 284:24696–704. doi: 10.1074/jbc.M109.030098

154. Cui W, Li Q, Feng L, Ding W. MiR-126-3p regulates progesterone receptors and involves development and lactation of mouse mammary gland. *Mol Cell Biochem*. (2011) 355:17–25. doi: 10.1007/s11010-011-0834-1

155. Theis V, Theiss C. Progesterone effects in the nervous system. Anat Rec (Hoboken). (2019) 302:1276-86. doi: 10.1002/ar.24121

156. Panda H, Chuang TD, Luo X, Chegini N. Endometrial miR-181a and miR-98 expression is altered during transition from normal into cancerous state and target PGR, PGRMC1, CYP19A1, DDX3X, and TIMP3. *J Clin Endocrinol Metab.* (2012) 97: E1316–26. doi: 10.1210/jc.2012-1018

157. Maillot G, Lacroix-Triki M, Pierredon S, Gratadou L, Schmidt S, Benes V, et al. Widespread estrogen-dependent repression of micrornas involved in breast tumor cell growth. *Cancer Res.* (2009) 69:8332–40. doi: 10.1158/0008-5472.CAN-09-2206

158. Gilam A, Shai A, Ashkenazi I, Sarid LA, Drobot A, Bickel A, et al. MicroRNA regulation of progesterone receptor in breast cancer. *Oncotarget*. (2017) 8:25963–76. doi: 10.18632/oncotarget.15657

159. Wendler A, Keller D, Albrecht C, Peluso JJ, Wehling M. Involvement of let-7/ miR-98 microRNAs in the regulation of progesterone receptor membrane component 1 expression in ovarian cancer cells. *Oncol Rep.* (2011) 25:273–9. doi: 10.3892/ or_00001071

160. Sirotkin AV, Ovcharenko D, Grossmann R, Laukova M, Mlyncek M. Identification of microRNAs controlling human ovarian cell steroidogenesis via a genome-scale screen. J Cell Physiol. (2009) 219:415–20. doi: 10.1002/jcp.21689

161. Cochrane DR, Spoelstra NS, Richer JK. The role of miRNAs in progesterone action. *Mol Cell Endocrinol.* (2012) 357:50–9. doi: 10.1016/j.mce.2011.09.022

162. An X, Ma H, Liu Y, Li F, Song Y, Li G, et al. Effects of miR-101-3p on goat granulosa cells in *vitro* and ovarian development in *vivo via* STC1. J Anim Sci Biotechnol. (2020) 11:102. doi: 10.1186/s40104-020-00506-6

163. Sun F, Nguyen T, Jin X, Huang R, Chen Z, Cunningham RL, et al. Pgrmc1/ BDNF signaling plays a critical role in mediating glia-neuron cross talk. *Endocrinology*. (2016) 157:2067–79. doi: 10.1210/en.2015-1610

164. Selvamani A, Sathyan P, Miranda RC, Sohrabji F. An antagomir to microRNA Let7f promotes neuroprotection in an ischemic stroke model. *PLoS One.* (2012) 7: e32662. doi: 10.1371/journal.pone.0032662

165. Barreto G, Santos-Galindo M, Diz-Chaves Y, Pernia O, Carrero P, Azcoitia I, et al. Selective estrogen receptor modulators decrease reactive astrogliosis in the injured brain: effects of aging and prolonged depletion of ovarian hormones. *Endocrinology.* (2009) 150:5010–5. doi: 10.1210/en.2009-0352

166. Bourque M, Dluzen DE, Di Paolo T. Neuroprotective actions of sex steroids in Parkinson's disease. Front Neuroendocrinol. (2009) 30:142-57. doi: 10.1016/j.yfrne.2009.04.014

167. Etgen AM, Jover-Mengual T, Zukin RS. Neuroprotective actions of estradiol and novel estrogen analogs in ischemia: translational implications. *Front Neuroendocrinol.* (2011) 32:336–52. doi: 10.1016/j.yfrne.2010.12.005

168. Kipp M, Beyer C. Impact of sex steroids on neuroinflammatory processes and experimental multiple sclerosis. *Front Neuroendocrinol.* (2009) 30:188–200. doi: 10.1016/j.yfrne.2009.04.004

169. Pietranera L, Lima A, Roig P, De Nicola AF. Involvement of brain-derived neurotrophic factor and neurogenesis in oestradiol neuroprotection of the hippocampus of hypertensive rats. *J Neuroendocrinol.* (2010) 22:1082–92. doi: 10.1111/j.1365-2826.2010.02058.x

170. Pike CJ, Carroll JC, Rosario ER, Barron AM. Protective actions of sex steroid hormones in Alzheimer's disease. *Front Neuroendocrinol.* (2009) 30:239–58. doi: 10.1016/j.yfrne.2009.04.015

171. Suzuki S, Brown CM, Wise PM. Neuroprotective effects of estrogens following ischemic stroke. Front Neuroendocrinol. (2009) 30:201-11. doi: 10.1016/j.yfrne.2009.04.007

172. Bohacek J, Daniel JM. The beneficial effects of estradiol on attentional processes are dependent on timing of treatment initiation following ovariectomy in middle-aged rats. *Psychoneuroendocrinology*. (2010) 35:694–705. doi: 10.1016/j.psyneuen.2009.10.010

173. Rodgers SP, Bohacek J, Daniel JM. Transient estradiol exposure during middle age in ovariectomized rats exerts lasting effects on cognitive function and the hippocampus. *Endocrinology*. (2010) 151:1194–203. doi: 10.1210/en.2009-1245

174. Wu WW, Adelman JP, Maylie J. Ovarian hormone deficiency reduces intrinsic excitability and abolishes acute estrogen sensitivity in hippocampal CA1 pyramidal neurons. *J Neurosci.* (2011) 31:2638–48. doi: 10.1523/JNEUROSCI.6081-10.2011

175. Gibson CL, Coomber B, Murphy SP. Progesterone is neuroprotective following cerebral ischaemia in reproductively ageing female mice. *Brain*. (2011) 134:2125–33. doi: 10.1093/brain/awr132