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RECEIVED 06 August 2023 ACCEPTED 22 November 2023 PUBLISHED 13 December 2023

#### CITATION

Wang K, Li Y and Chen Y (2023) Androgen excess: a hallmark of polycystic ovary syndrome. *Front. Endocrinol.* 14:1273542. doi: 10.3389/fendo.2023.1273542

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# Androgen excess: a hallmark of polycystic ovary syndrome

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Polycystic ovarian syndrome (PCOS) is a metabolic, reproductive, and psychological disorder affecting 6–20% of reproductive women worldwide. However, there is still no cure for PCOS, and current treatments primarily alleviate its symptoms due to a poor understanding of its etiology. Compelling evidence suggests that hyperandrogenism is not just a primary feature of PCOS. Instead, it may be a causative factor for this condition. Thus, figuring out the mechanisms of androgen synthesis, conversion, and metabolism is relatively important. Traditionally, studies of androgen excess have largely focused on classical androgen, but in recent years, adrenal-derived 11-oxygenated androgen has also garnered interest. Herein, this Review aims to investigate the origins of androgen excess, androgen synthesis, how androgen receptor (AR) signaling mediates adverse PCOS traits, and the role of 11-oxygenated androgen in the pathophysiology of PCOS. In addition, it provides therapeutic strategies targeting hyperandrogenism in PCOS.

### KEYWORDS

androgen receptor, Hyperandrogenism, insulin resistance, polycystic ovarian syndrome, steroidogenesis

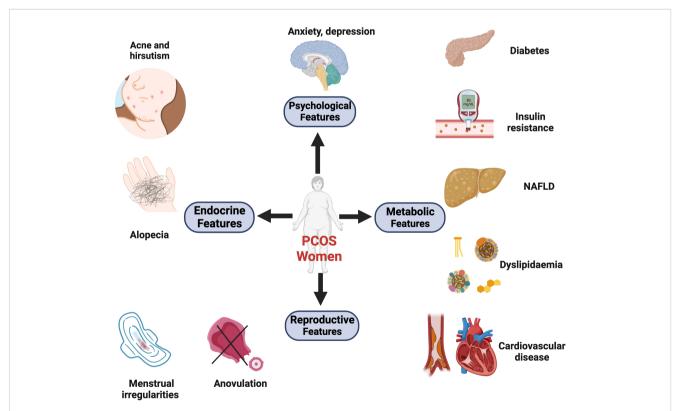
### 1 Introduction

Polycystic ovary syndrome (PCOS) is an endocrine, metabolic, reproductive, and psychological disorder affecting approximately 6%–20% of reproductive women worldwide, regardless of ethnicity. It is primarily characterized by ovulation dysfunction, clinical or biochemical hyperandrogenemia (HA), and polycystic ovarian morphology (PCOM) (1). According to the Rotterdam criteria, two of the three abovementioned features are required for the diagnosis of PCOS. Although this standard is widely accepted, the definition of PCOS by the National Institutes of Health is mainly focused on two aspects: HA and ovulatory dysfunction. As for the Androgen Excess Society criteria, the presence of HA is required along with either ovulatory dysfunction or PCOM or both. Hence, four PCOS phenotypes (A, B, C, and D) are ascertained under the Rotterdam criteria. Patients should be thoroughly assessed to exclude other conditions that cause symptoms similar to PCOS. Although approximately 75% of patients with PCOS exhibit insulin resistance (IR), IR is not recognized as a diagnostic criterion (2).

As PCOS is a highly heterogeneous disease, its exact cause remains unknown. Abundant evidence suggests that environment, prenatal, and genetic factors, as well as epigenetic changes, are interrelated with each other (3). Patients with PCOS may still experience poor metabolic outcomes regardless of whether their body mass index (BMI) is within the normal range. A previous study has demonstrated that the incidence of non-alcoholic fatty liver disease is correlated with HA, after adjusting for BMI (4). Furthermore, patients with PCOS with HA are more likely to develop type 2 diabetes later in life than those with normal androgen levels (5). Additionally, HA exposure leads to a higher prevalence of cardiometabolic complications, such as hypertension and obesity. This could be explained by the activation of the reninangiotensin system, deregulation of sympathetic nervous system activity, and upregulation of androgen receptors (ARs) (6). Moreover, excess maternal androgen exposure can impair the placental function, increase the risk of developing PCOS in female offspring, and reduce sperm quality in male generations (7) (Figure 1).

Under the Rotterdam classification, patients with phenotype D do not exhibit HA but only symptoms of oligo-anovulation and ultrasonographic evidence of PCOM. One study reviewed several recent publications and surprisingly determined that there was no evident difference in metabolic parameters between patients with phenotype D and controls, although some of these normoandrogenic patients from East Asian populations present with higher metabolic risks (8). Moreover, androgen levels decline dramatically with age in patients with phenotype D PCOS, and even decrease below the average levels of their healthy counterparts when they reach  $\geq$ 35 years. Gleicher et al. redefined this phenotype as the hyper-/hypoandrogenic phenotype (HH-PCOS). This phenotype may be driven by immune dysregulation and adrenal autoimmunity, indicating a different etiopathology than the other three phenotypes (9). Therefore, the application of the Rotterdam criteria has been controversial, and an alternative classification for the diagnosis of PCOS has been proposed in recent years (10). Overall, more individualized and tailored treatments targeting different PCOS phenotypes are required.

Considering that the majority (60%-80%) of women with PCOS have elevated androgen levels, we focused on the interactions between IR and HA, synthesis of androgen in target organs and tissues, action of androgen through ARs, and promising therapeutic interventions to relieve hyperandrogenic symptoms.



### FIGURE 1

The major clinical manifestations of PCOS. These symptoms can be divided into four categories: reproductive, endocrine, metabolic, and psychological comorbidities. (1) Reproductive features: the dysregulated hypothalamus-pituitary-gonadal (HPG) axis and neuroendocrine factors contribute to menstrual irregularities, anovulation, infertility, and increased risks of pregnancy complications. (2) Endocrine features: hormonal imbalances (ovarian and adrenal hyperandrogenemia). Symptoms like hirsutism, acne, and androgenic alopecia are induced by high levels of circulating androgens. (3) Metabolic features: 30% of lean and 70% of obese patients exhibit insulin resistance. Additionally, women with PCOS suffer from abdominal obesity, dyslipidemia, and non-alcoholic fatty liver disease (NAFLD). These metabolic abnormalities can result in long-term cardiometabolic sequelae, including type 2 diabetes, hypertension, and atherosclerotic disease. (4) Psychological features: PCOS is associated with an increased prevalence of depression, anxiety, and poor quality of life. Created with BioRender.com.

### 2 Insulin resistance

IR and hyperandrogenism are often interconnected and collectively account for the reproductive and metabolic characteristics of PCOS. It is widely argued that IR and subsequent compensatory hyperinsulinemia in PCOS are caused by post-receptor defects, which manifest as decreased tyrosine phosphorylation and increased serine phosphorylation of insulin receptors and insulin receptor substrate (IRS) (11). Hyperinsulinemia inhibits the secretion of the hepatic sex hormone-binding protein (SHBG), thereby increasing free testosterone (T) levels in circulation. Additionally, insulin and luteinizing hormone (LH) administration independently and synergistically stimulates androgen and progesterone production by upregulating the activities of 17-hydroxylase/17,20-lyase (CYP17) and steroidogenic acute regulatory protein (StAR) in ovarian theca cells (TCs). Moreover, insulin and human chorionic gonadotropin combination treatment upregulates CYP11A1 (P450scc) production in TCs (12). Additionally, acute insulin stimulation promotes adrenal steroidogenesis by amplifying the response to ACTH stimulation, with increased  $5\alpha$ -reductase enzyme activity (13). Moreover, one meta-analysis further concluded the increased peripheral 5αreductase activity in women with PCOS, and it is associated with IR, unrelated to BMI (14).

Insulin signaling pathways are divided into two categories: phosphatidylinositol-3-kinase/serine/threonine-specific protein kinase B (PI3K/AKT), which is related to metabolism, and mitogen-activated protein kinase (MEK)-extracellular signalregulated kinase (ERK), which is involved in cell growth and proliferation. A growing body of evidence has demonstrated that the aberrant PI3K/AKT pathway in patients with PCOS results in reduced translocation of glucose transporter 4 (GLUT4) from adipocyte and endometrial cells, thus impairing glucose uptake. Decreased endometrial Glut4 gene expression is also regulated by ARs through binding to the GLUT promoter in rat models (15). Interestingly, one three-dimensional culture of mouse follicles in vitro demonstrated that high concentrations of insulin-like growth factor-1 (IGF-1) could inhibit mouse follicular development and maturation, which provides a possible explanation for the involvement of IGF-1 in the pathogenesis of ovulatory disorders in phenotype D in the absence of HA (2, 16). Moreover, enhanced CYP17 activity in TCs is regulated by the PI3K/AKT pathway (17). Despite disrupted PI3K/AKT signaling, the mitogenic insulin receptor-activated pathway remains intact or is enhanced in skin fibroblasts and skeletal muscle from patients with PCOS, which is called selective IR (18, 19).

# **3** Neuroendrocine abnormalities

The hypothalamic-pituitary-gonadal axis regulates ovarian steroidogenesis. Negative feedback from progesterone and estradiol to the hypothalamus maintains sex hormone levels within the normal range. In women with PCOS, increased GnRH pulsatility stimulates greater LH secretion than follicle-stimulating hormone (FSH), thereby increasing the LH-to-FSH ratio. Consequently, it promotes LH-stimulated androgen production in ovarian TCs. Androgen is required for follicular growth in the early phases; however, excess androgen suppresses the expression of cumulus expansion-related genes and oocyte maturation-related genes, causing excess small growing follicles to be arrested at the antral stage, inhibiting the development of the dominant follicle (20). HA in PCOS also disrupts the preovulatory LH surge. Altogether, these findings suggest anovulation in patients with PCOS.

Ovarian anti-Müllerian hormone (AMH) is primarily synthesized by granulosa cells (GCs) from preantral and small antral follicles with a maximum diameter of 8 mm due to the inhibitory effect of E2. Therefore, AMH levels were 2-3 times higher in the serum and follicular fluid of patients with PCOS than in normal women (21). AMH inhibits the sensitivity of growing follicles to FSH. Moreover, E2 suppressed AMH and AMH-specific type 2 receptor (AMHR2) expression at the ovarian level (21). Excess dihydrotestosterone (DHT) directly increases AMH, whereas T upregulates ERa expression through the conversion to E2. High AMH levels activate downstream inhibitory SMAD-6/7 signaling and result in follicular arrest (22). Elevated T levels have a greater correlation with increased AHM levels than other types of androgens (23). Considering the close relationship between HA and AMH, AMH is used to evaluate the degree of HA and is considered an alternative to diagnosing PCOS, although the cutoff values are inconsistent (24). Additionally, AMH has been reported to stimulate GnRH and LH release; however, how AMH modulates GnRH function remains largely unknown. Recently, one study uncovered the role of AMH in regulating GnRH neuron activity in a murine model (25). The structure of the hypothalamic median eminence is altered after AMH binds to AMH2R; thus, the retracted tanycytes make it easier for GnRH neuron terminals to release GnRH into the blood capillaries.

Interestingly, central progesterone or estradiol receptors are not present in GnRH neurons in humans (26). Therefore, upstream KNDy neurons may collectively regulate the activity of GnRH neurons. KNDy neurons distributed in the arcuate nucleus (ARN) are composed of kisspeptin, neurokinin B (NKB), and dynorphin A. Kisspeptin and NKB stimulate kisspeptin release, which is considered a major GnRH pulse generator. Dynorphin A inhibits GnRH release.

HA has been reported to stimulate KNDy neurons, embodied by increased kisspeptin and NKB expression and downregulated DynA expression in rats, thus causing a vicious cycle of hypothalamic-pituitary-ovarian (HPO) dysfunction (27). Treatment with anti-epileptic drugs, which increases the gammaaminobutyric acid (GABA) concentration in the brain, increases the occurrence of PCOS. This supports the notion that dysfunctional GABA neurons also participate in regulating GnRH neuronal activity (28). The chronic activation of GABA neurons induced PCOS-like symptoms in healthy female rodents, causing elevated T levels and an impaired reproductive cycle. Prenatally androgenized (PNA) mice exhibited attenuated responsiveness to GABA stimulation (29). Mechanistically, when GABA binds to the GABA<sub>b</sub> receptor expressed in GnRH neurons, it promotes Cl<sup>-</sup> influx, thus exciting GnRH neurons. Overall, circuit remodeling is evident in GABAergic and GnRH neurons when they are exposed to excess androgens (30).

### 4 Sexual dimorphism

A Mendelian randomization analysis using statistics from the UK Biobank revealed the sex-specific association of testosterone in men and women. A genetically determined 1 standard deviation (SD) higher T level would result in a 15% lower risk of developing type 2 diabetes in men. In accordance with the study, previous research has demonstrated the positive impact of androgen on reducing fat mass in men (31). By contrast, high circulating T levels were detrimental to women. Every 1 SD increase in T levels would increase the occurrence of type 2 diabetes by 37%, and this was associated with a higher PCOS risk with an odds ratio (OR) of 1.51 in women. This study indicated that hyperandrogenism is not simply a feature or consequence but a causative factor in PCOS development (32). Moreover, high androgen levels determined approximately 20% of heritability in both men and women. A metabolomic analysis revealed a similar sexual dimorphism (33). Obesity increases the metabolites of serum branched chains and aromatic amino acids; however, this adverse effect only affects women, with no deleterious effect on men of normal weight or with obesity (33). Numerous studies have demonstrated that women with excess androgen and men with androgen deficiency exhibit overlapping metabolic traits, indicating sexual androgen dimorphism. A good example is PCOS in women and men with hypogonadism (34). Theoretically, females with high estrogen and low androgen levels are prone to gain subcutaneous fat rather than visceral fat and display increased glucose uptake, thus protecting them from adverse metabolic consequences. By contrast, androgens help males have higher muscle mass and reduce abdominal adiposity. Clinical studies have shown that men with androgen deficiency have worse metabolic phenotypes, such as impaired glucose tolerance and higher CVD risks. As for women with PCOS, excess male sex hormones favor abdominal visceral fat deposition, which is also known as the android fat pattern (35).

# 5 Gene polymorphisms

Accumulating evidence indicates that polymorphisms in multiple genes play important roles in PCOS susceptibility and pathogenesis. In recent decades, polymorphisms in several common genes have been identified in the development of PCOS, such as variants in *DEND1A*, *THADA*, *FSHR*, and *LHCGR* gene variants (36). Heidarzadehpilehrood et al. retrieved and summarized the roles of several critical gene mutations involved in steroidogenesis pathways, including *CYP11A1*, *CYP17A1*, and *CYP19A1*, in PCOS pathophysiology. For example, the rs743572 polymorphism in *CYP17* is associated with severe biochemical and clinical characteristics. Allele rs2414096 of the *CYP19* gene is linked to HA and reduced aromatase activity (37). Some findings may be inconsistent among different ethnic populations, and correlations

between polymorphisms in the CYP11A1 and CYP17 promoters and T levels in women with PCOS have been reported (38). One study has demonstrated that although polymorphisms in the AMH and AMHR2 genes were not associated with a higher risk of developing PCOS, patients with PCOS with AMH Ile(49)Ser (rs10407022) may represent a milder phenotype due to decreased AMH bioactivity (39). Interestingly, polymorphism rs10406324 (-210 A>G) in the AMH promoter region is associated with lower AMH levels in patients with PCOS (40). Women with a shorter CAG repeat length ( $\leq 17$ ) in the AR gene may have a higher risk of developing PCOS. In terms of SHBG, eight or more SHBG gene pentanucleotide TAAAA repeats (rs35785886) have been associated with low serum SHBG levels in women with PCOS (41). The association between AR gene polymorphism and polycystic ovary syndrome will be discussed later (in the AR section).

### 6 Androgen synthesis

### 6.1 Androgen synthesis in ovaries

Ovarian androgens are produced by TCs under LH stimulation, whereas estrogen is synthesized in GCs in response to FSH. Owing to the absence of CYP17A1 in GCs, parts of androgens spread from TCs to GCs to produce estrogen. The subtypes of estrogen are determined by different androgen substrates, with A4 converting to estrone (E1) and T aromatizing to estradiol (E2). Moreover, E1 could further be transformed into E2 by  $17\beta$ -hydroxysteroid dehydrogenase type1 ( $17\beta$ -HSD1). E1 has weaker bioactivity than E2 (42). Unlike the mixed results of E2 levels in classical and non-classical PCOS groups, nearly all patients with PCOS experience increased E1 levels, thus making E1 levels a reliable biomarker for distinguishing between patients with PCOS and healthy women (43, 44).

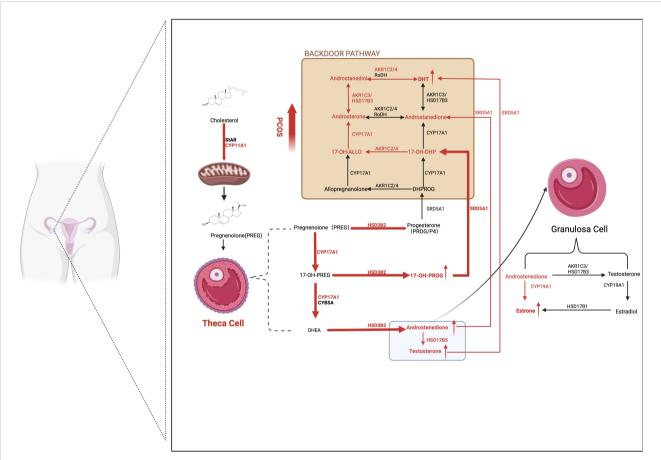
Androstenedione (A4) is the most abundant serum steroid metabolite in patients with PCOS. Additionally, higher androsterone, T, and 11-ketotestosterone (11-KT) levels and reduced progesterone (PROG/P4) levels were observed when comparing patients with PCOS with healthy matched groups. The synthesis of DHT through the backdoor pathway, which bypasses the formation of T, is enhanced in the ovaries of patients with PCOS. The conversion from 17OH-Prog to 17OH-dihydroprogesterone by 5 $\alpha$ -reductase 1 is the starting point of the backdoor pathway, and ultimately, DHT is produced by retinol dehydrogenase and 3 $\beta$ -HSD1/3(AKR1C2/4) from precursor androstanediol in GCs (45).

Multiple studies have reported that TCs in patients with PCOS are predisposed to generate more androgens due to overactive and overexpressed steroidogenic enzymes, such as CYP17A1, CYP11A1, HSD3B2, SRD5A1, and  $17\beta$ -HSD5 (also known as AKR1C3) (46). Patients with PCOS with the *DENND1A* splice variant (DENND1A.V2), which encodes the DENND1A protein, demonstrated increased CYP17A1 and CYP11A1 expression and androgen production. Knockout of DENND1A.V2 reduced *CYP17A1* and *CYP11A1* gene transcription and subsequent

androgen biosynthesis (47). Moreover, aromatase activity was downregulated in GCs in human polycystic ovaries due to hypomethylation of the CYP19A1 promoter influenced by HA. Additionally, FSH receptors are downregulated in women with PCOS, which may partly explain the lean or non-obese phenotype of PCOS (Figure 2).

In bovines, A4 accumulation inhibits GC proliferation and promotes apoptosis, resulting in an arrested cell cycle. Specifically, increased AMH levels induced by HA stimulate CTNNBIP1 expression, which in turn suppresses the interaction of CTNNBIP with Wnt, leading to dysregulated Wnt signaling (48). One study has demonstrated that AR indirectly binds to the promoter of klotho, a regulator of autophagy and aging, embodied with increased klotho levels. Subsequently, this induces the apoptosis of GCs (49). One study disclosed how a hyperandrogenic microenvironment affects a rodent's ovary. The expression of vascular cell adhesion molecule 1 (Vcam1) was upregulated in TCs and stroma cells identified by dual ARs and NR2F2 expression. Vcam1 is characteristic of Leydig cells and may be linked to immune attachment and inflammation. Interestingly, similar changes were not detected in the GCs (50).

Numerous previous studies have demonstrated that androgen excess disturbs the immune response in the ovary and other targeted organs, with an increased secretion of inflammatory cytokines and dysregulated immune cells (51). Chronic low-grade inflammation, in turn, modulates the expression of steroidogenic genes and initiates a series of physiological processes in theca-interstitial cells (52). For example, IFN- $\gamma$  expression is inversely associated with dehydroepiandrosterone (DHEA) levels in a dose- and time-dependent manner in GCs (53). DHT treatment increased nuclear AR abundance and thus activated TLR4-IRF-7-NF $\kappa$ B signaling, causing endometrial inflammation (54). One study treated lean and healthy women with DHEA and observed that such treatment stimulates TNF $\alpha$  release by mononuclear cells and AR mRNA in the fasting state. These alterations are positively correlated with increased androgen levels. When pretreated with flutamide, TNF $\alpha$  levels decreased (55).



#### FIGURE 2

Ovarian-derived androgen biosynthesis in PCOS women (the classical and backdoor pathway). A series of steroidogenic enzymes, including CYP11A1, CYP17A1, and HSD3B2 in the theca cells and CYP19A1 in granulosa cells, are reported to increase in PCOS patients. It is demonstrated that circulating A4 was preferentially elevated in women with PCOS. Furthermore, globally higher SRD5A1 activity is observed, resulting in increased downstream androstanedione and DHT activation from A4 and T, respectively. The backdoor pathway, which involves the production of DHT bypassing the formation from T, is enhanced. In detail, 17OHP4 is the starting point of the backdoor pathway, Because 17OH-Allo is more efficiently metabolized by CYP17A1 than 17OH-Preg, it preferentially synthesizes DHT through the backdoor pathway rather than the classic pathway. The red arrows represent the enhanced pathways. SRD5A1, 5a-reductase type I; A4, androstenedione; T, testosterone; DHT, dihydrotestosterone; 17OHP4, 17ahydroxy-progesterone; 17OH-Allo, 17-hydroxy-allopregnanolone.

### 6.2 Androgen synthesis in adipose tissue

Adipose tissues (ATs) are also the main source of androgen production with complete steroidogenic machinery, especially in women with obesity (56). ATs can be classified into two categories: white and brown AT, which are composed of visceral adipose tissue (VAT) and subcutaneous adipose tissue (SAT). These two types of ATs manifest significant characteristics in many aspects. In the SAT of women with PCOS, increased activity of aldo-keto reductase1C1-3 is observed, whereas the enzymes aromatase and  $5\alpha$ -reductase1 are downregulated, indicating increased T biosynthesis and DHT inactivation. Moreover, leptin levels are inversely correlated with DHT concentration (57). These findings are in line with those of a previous study reporting that insulin significantly stimulates A4 and T production in SAT (58). Conversely, in the VAT of women with obesity, increased aromatase activity, representing the hypertrophy of adipocyte, parallels the increased androgen catabolism via enhancing AKR1C2 function (59). In addition, the expression level of AR in the VAT may be higher than that in the SAT of women with PCOS, making the VAT more vulnerable to androgen exposure (35). Hence, in the VAT of women with PCOS, androgen tends to promote lipolysis, whereas insulin-induced lipolysis is suppressed. Thus, excessive free fatty acid efflux leads to ectopic fat deposition, including in the liver and skeletal muscles. Adipocytes become hyperplastic and hypertrophic when exposed to androgen, which triggers immune cell infiltration. Moreover, hyperandrogenism stimulates the transcription of AR in mononuclear cells to release more inflammatory factors (60). Consequently, this stimulates the abnormal secretion of adipocytokines by adipocytes and results in decreased adiponectin production and elevated leptin, visfatin, and resistin levels. The functions of different types of adipocytokines have been described well by various previous articles (61).

Even in normal-weight patients with PCOS, IR is present in AT and positively correlated with serum androgen levels. Androgen modulates the alteration in subcutaneous abdominal adipose stem cell (ASC) gene expression (62). Compared with abdominal SAT, the expression of genes involved in fat accumulation and angiogenesis in gluteofemoral SAT was downregulated. Along with the pro-inflammatory states and low gene expression of adipogenesis in ACSs of gluteofemoral SAT, these findings indicate the restricted adipose expansion in the lower body region. Moreover, the ASCs from gluteofemoral SAT are hypermethylated. The capacity for fat storage may be programmed at the early stages of life, thus promoting the predisposition toward abdominal fat accumulation (63). This opinion is supported by the finding of differential DNA methylation from the VAT of PNA rhesus monkeys (64).

The activity and volume of brown adipose tissue (BAT) are decreased in women with PCOS due to the downregulated expression of uncoupling protein 1 upon excess androgen stimulation, which may cause mitochondrial dysfunction and impaired thermogenesis, which would decrease lipolysis (65). Cold treatment and BAT transplant may be attractive strategies.

### 6.3 Androgen synthesis in the adrenal cortex

The adrenal cortex comprises three layers with different distributions of steroidogenic enzymes. Androgen biosynthesis occurs in the zona reticularis (ZR) (66).

ZR accounts for half of DHEA synthesis, with 20% occurring in the ovary and the remaining 30% being transformed by circulating DHEAS. In contrast to DHEA, its sulfated metabolite, DHEAS, comes exclusively from the adrenal gland, which is attributed to the high and specific expression of sulfotransferase 2A1 in the ZR. Because of the relatively low expression of HSD3B2 in the ZR, the ZR tends to produce DHEAS rather than A4. Thus, its downstream product T is negligible because of the small amount of adrenal HSD17B5. Therefore, DHEAS was used as a reliable marker to assess adrenal androgen excess (67). Genetic variants in the SULT2A1 gene, for example, SNP rs182420 identified in patients with PCOS, contribute to higher DHEAS levels (68). Interestingly, previous studies have linked low DHEAS concentrations to higher cardiovascular risks and poor lipid profiles in both sexes. DHEAS and DHEA mediate the relaxation of vascular smooth muscles via the mechanism of the openness of potassium channels, thus lowering blood pressure (69). Additionally, high circulating DHEAS helps to lower total cholesterol levels and prevent visceral fat accumulation (70). Similarly, women with PCOS with a high DHEA phenotype have lower carotid intima-media thickness. Moreover, DHEAS decreases with age at similar rates between women with PCOS and their healthy counterparts (71, 72). However, DHEAS measurement cannot accurately reflect the actual amounts of adrenal androgen. As precursors, DHEA and DHEAS can be converted to other forms of androgen in the periphery. Furthermore, women with PCOS may have exaggerated adrenal steroidogenesis in response to ACTH stimulation. One study used a dexamethasone suppression test to distinguish the sources of androgen in women with PCOS. Surprisingly, total testosterone levels decreased dramatically in 9 out of 51 patients after glucocorticoid suppression, indicating adrenal-derived androgen dominance. However, among them, only one patient demonstrated elevated baseline DHEAS levels (73). This study suggested that excess serum dehydroepiandrosterone sulfate does not parallel adrenal hyperandrogenism.

### 6.4 11-oxygenated androgen

Emerging evidence has demonstrated that adrenal-derived 11oxygenated androgens are also key players in the development of PCOS. 11-oxygenated androgens acquired their name because they share the same oxygen atom on the 11th carbon (74).

Because of the specific expression of CYP11B1 in the ZR, A4, and T serve as substrates and are then converted to 11-OH-androstenedione (11-OH-A4) and 11OH-testosterone (11-OH-T), respectively. 11-OH-A4 is the most abundant 11-oxygenated androgen in the adrenal cortex, considering the minor abundance of T for low HSD17B5 activity. In the kidney,  $11\beta$ -hydroxysteroid dehydrogenase type 2

(HSD11B2) is responsible for catalyzing 11-OH-A4 and 11-OH-T into 11-ketoandrostenedione (11-KA4) and 11-ketotestosterone (11-KT). Therefore, the majority of circulating 11-KT is converted from 11-KA4 by HSD17B5. Under 5 $\alpha$ -reductase stimulation, 11-KT is metabolized to 11-ketodihydrotestosterone (11-KDHT), which has similar bioactivity to DHT. Similarly, 11-KT has equivalent potency to T (75). However, 11 $\beta$ -hydroxy derivatives, including 11-OH-A4, 11-OH-T, and 11-OH-DHT, exhibit low or no androgenic bioactivity in mammalian cells (76). Considering the low affinity of 5 $\alpha$ -reductase for 11-KT, serum 11-KDHT only constitutes a minor proportion of the overall circulating androgen pool when compared with DHT (77). The backdoor pathway for 11-oxygenated androgen begins with P4 or 17 $\alpha$ hydroxyprogesterone (17OHP4). After a series of conversions, the end product, 11-KDHT, was obtained (66).

CYP11B1 expression is almost negligible in the ovary. Moreover, 11-oxygenated androgens are unfavorable substrates for the enzyme CYP19A1, which explains why 11-oxygenated estrogen does not exist (78). Consistently, one study measured the levels of 11-KT and its precursors and observed that although 11-KT levels were elevated in patients with PCOS, no distinct difference in 11-KT and 11-OHA4 levels was observed between the ovarian vein and periphery (79). Moreover, the administration of oral contraceptive does not completely block HA in women with PCOS. These findings indicate that the ovary is not the only source of excess androgens in PCOS.

As the 11-oxygenated androgens are predominantly secreted by adrenal glands rather than the ovary, they are therefore not controlled by the HPO axis but by the ACTH. They follow diurnal rhythms similar to cortisol (79). The levels of traditional androgen decline with age, whereas 11-oxygenated androgen levels are quite stable. Owing to the age-related degradation of ZR, the zonal boundary of HSD3B2 and CYB5A becomes less distinct. Hence, promoting the conversion from DHEA into A4 and thus downstream 11-oxygenated androgens through the action of HSD3B2 (80). This may partly explain why women with PCOS still have a higher cardiovascular risk even after menopause (81-83). As for adolescents, there is no evident increase in 11-KT in patients with PCOS, and metformin administration has no effect on 11-KT levels (84). One study compared the 11-oxygenated androgen levels between girls whose mothers have PCOS and obesity and healthy individuals. Unfortunately, no evident difference was observed between the two groups (85). The dominant circulating androgen during the adrenarche is 11-HT, and premature adrenarche may be the possible explanation (86).

Intriguingly, one study identified 11-oxygenated androgens as the predominant circulating androgens in women with PCOS, with 11-KT levels being more than twice as high as T (87). However, androgen levels vary among patients with PCOS (88). Another study supports BMI as a key player in determining steroid profiles (89). The 11-oxygenated androgens are considered to be higher in patients with obesity and PCOS than in healthy counterparts without obesity. They are positively correlated with IR, partly because HSD17B5 upregulates lipid accumulation in the obese group. Dumesic et al. have reported that the normal level of 11-oxygenated androgens in women of normal weight and with PCOS is associated with reduced cortisol levels and decreased HSD11B1 activity. This protective effect diminishes as visceral fat mass increases

(90). HSD17B5 (AKR1C3) is induced by the insulin with PI3K-AKTmTOR signaling (88). Moreover, one study has reported that excess 11-oxygenated androgen production is triggered by heightened adrenal insulin signaling (91). Specifically, insulin directly upregulates steroidogenic factor 1 (SF-1) expression to stimulate the synthesis of adrenal steroidogenic enzymes, including StAR, CCYP11A1, CYP11B1, CYB11B2, and HSD3B2, and suppresses FoxO1 nuclear translocation (92).

In addition, HSD17B5 has a higher affinity for 11-KA4 converting into 11-KT than classical androgen. 11-KT is further inactivated and transformed into 11-OH-T in the presence of HSD11B1 (93). Moreover, HSD17B5 has been reported to stabilize and activate AR without androgen binding and to upregulate fatty acid synthase, thus leading to excess lipogenesis and lipotoxicity (94). Therefore, the ratio of HSD17B5 to HSD11B1 plays a crucial role in determining the severity of hyperandrogenism in patients with PCOS (Figure 3).

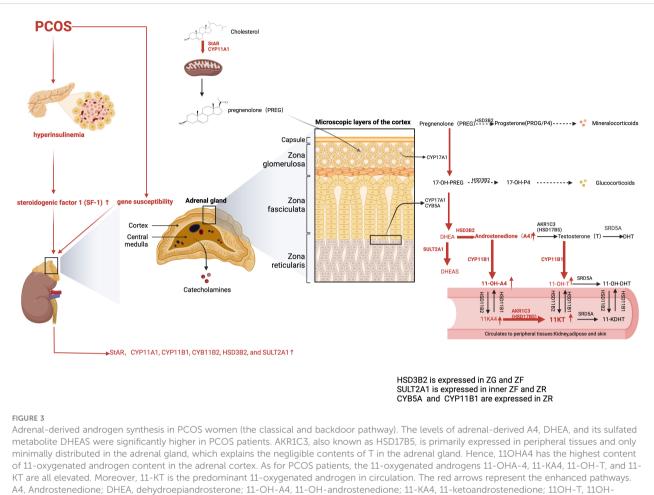
In conclusion, these studies strongly suggest the presence of diverse sources of androgen in circulation among women with PCOS. The detection of both classical and 11-oxygenated androgens using liquid chromatography-tandem mass spectrometry (LC-MS/MS) enables the analysis of the comprehensive steroid profile in PCOS women (77). The physiological roles that 11-oxygenated androgens play in the female reproductive system need to be elucidated in future studies.

# 7 The effects of androgen excess in the uterus

A previous study has reported that the mRNA levels of insulin receptor IRS1/2 and GLUT1/2 increase in the endometria of patients with PCOS. Chronic DHT treatment affects the expression of genes involved in glucose metabolism and the cell cycle (95). High androgen levels, IR, and a disturbed HPO axis collectively contribute to anovulation and menstrual irregularities. Moreover, the persistent stimulation of estrogen and progesterone in the proliferative phase leads to endometrial overgrowth, impaired endometrial receptivity, and an increased risk of developing endometrial hyperplasia and endometrial cancer in women with PCOS. Obesity can exacerbate these risk factors. In mouse models, hyperandrogenic conditions enhanced the expression of adhesion molecules avß3-integrin and lysophosphatidic acid receptor 3 protein, leading to decreased endometrial thickness (96). Endometrial AR expression is overexpressed in patients with PCOS, thus activating TLR4-IRF-7-NFKB signaling and promoting the release of downstream pro-inflammatory cytokines IFN $\alpha$  and IFN $\gamma$ . Furthermore, the expression levels of ER $\alpha$  and AR increased with the onset of endometrial hyperplasia in women with PCOS (97).

# 8 The effects of androgen excess in other peripheral tissues

Only a small portion of A4 is released into the circulation and then metabolized into other forms of androgens in the periphery,



testosterone: 11-KT, 11-ketotestosterone.

such as the skin, causing dermatological symptoms including hirsutism (increased facial or body hair), acne, and female pattern alopecia (77). The severity of hirsutism can be evaluated using the modified Ferriman–Gallwey (m-FG) score. More recently, Stanczyk et al. identified an indirect DHT formation pathway in the genital skin of patients with PCOS, with the first step being the conversion from A4 to 5 $\alpha$ -androstane-3,17-dione (5 $\alpha$ -A) via 5 $\alpha$ -reductase. Subsequently, 5 $\alpha$ -A is metabolized into T by HSD17B5. The study has demonstrated that although serum T and A4 concentrations are significantly higher than 5 $\alpha$ -A and DHT, the genital tissue-toserum ratios indicate opposite results. The ratios in the 5 $\alpha$ -A and DHT groups are significantly higher than those in the T and A4 groups. Thus, circulating androgen levels do not completely reflect the local conversion and metabolism of androgen (98).

### 9 Androgen receptor

Classical androgen action is mediated by the nuclear AR. Additionally, androgens exert biological effects via non-genomic signaling involving the MAPK-ERK and AKT pathways. Androgen typically binds to its ARs, whereas DHEA can bind to AR and ER. Distinct differences in AR affinity were observed between different androgen subtypes. DHT has the highest binding affinity of androgens for ARs, followed by T, A4, and DHEA.

Mechanistically, in the absence of a ligand, AR is confined to the cytoplasm and binds to heat shock protein 90 (HSP90) and other co-chaperones. Upon ligand stimulation, HSP90 dissociates from AR, which then localizes to the nucleus and interacts with androgen response elements to initiate the transcription of downstream target genes. Previous studies have demonstrated that longer GGN repeats in the *AR* gene contribute to increased hyperandrogenic states in the Chinese PCOS population, whereas Turkish women with a shorter CAG repeat length had a higher risk for PCOS (99).

AR is widely expressed in female reproductive tissues and its expression is cell- and tissue-specific and fluctuates during the menstrual cycle. Under normal conditions, AR expression is highest in the stroma of the endometrium in the proliferative phase and decreases during the secretory phase until it becomes undetectable. AR expression is present in most stages of follicle development. It increases until the antral stages and then gradually decreases in GCs. Abnormal AR expression is detected in the ovary of patients with PCOS (100). High levels of phosphoglycerate kinase 1, as a binding partner of AR, stabilize ARs and inhibits them from being ubiquitinated through E3 ubiquitin ligase SKP2, thereby promoting AR nuclear translocation and later the expression of

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ovulation and glucose metabolism-related genes in the GCs of patients with PCOS, thus ultimately leading to anovulation (101). FHL2, a key regulator of follicular development, is also involved in the pathogenesis of anovulation. FHL2 overexpression induced by DHT treatment acts as a co-repressor of AR at the C/EBPβ promoter, thus suppressing C/EBPβ expression, which is a key ovulation-related gene. Furthermore, FHL2 directly inhibits ERK1/ 2 phosphorylation and downstream gene transcription (102). The inhibition of the ERK1/2 and MEK1/2 pathways is associated with increased androgen biosynthesis, independent of insulin activation (19).

Other studies have reported that AR expression in GCs is significantly decreased among patients with PCOS, which is negatively correlated with LH levels. However, this link disappeared when the number of antral follicles increased to 24 (103). Thus, downregulated ovarian ARs can be compensated by the elevation of androgen levels and increase in extra-ovarian ARs. Another possible explanation is that acute androgen stimulation promotes AR expression in animal models, whereas AR expression is reduced by chronic excess androgen exposure in patients with PCOS (104).

Chronic androgen-driven follicle arrest is caused by abnormal AR degradation, which downregulates RNF6 activity, induces K48linked polyubiquitination, and then leads to proteasome degradation of AR. Under normal conditions, K63-linked AR polyubiquitination promotes AR transcription activity and the downstream expression of GDF9, a key modulator of follicle growth (105). Consequently, investigating the function of AR and the amount of nuclear import is more important than the abundance of AR expression in patients with PCOS.

Two alternative splice variants (ASVs), Ins-AR and Del-AR, have been identified exclusively in the GCs of patients with PCOS and are closely associated with hyperandrogenism and ovarian dysfunction (106). The former is caused by the insertion of 69 base pairs between exons 2 and 3, whereas the latter lacks exon 3. In addition, patients who carry Ins-AR are positively correlated with high AMH levels; however, although AR mRNA is elevated in all groups, no significant difference in the abundance of AR mRNA was observed between the AR-SV and wild-type AR (WT-AR) groups (107). This could be explained by alterations in the AR DBD structure of the two isoforms, which then affect the transcription of genes regulating follicular growth and steroidogenesis. CYP17A1 and IGFBP7 expression was upregulated, whereas CYP19A1 expression was suppressed (106). Mechanically, one recent study has revealed that the decreased CYP19A1 expression is driven by the delayed AR nuclear translocation (108). HSP90 exhibits a greater affinity for Ins-AR and Del-AR than WT-AR, making the two AR variants more stable. The Del-AR variant exhibited the greatest binding affinity. Under DHT stimulation, importin a bound more tightly with WT-AR. However, its binding affinity with the Ins-AR and Del-AR isoforms declines dramatically, leading to decreased AR nuclear import. Of note, Ins-AR had the lowest affinity with importin  $\alpha$  in response to and rogen stimulation. This may explain why patients with the Del-AR variant exhibited a milder PCOS phenotype. Moreover, serine/arginine-rich splicing factors (SRSF), especially SRSF2 and SRSF3, which are regulated by miRNA183 and miRNA124, respectively, participate in the formation of ASVs (107). Xue et al. unveiled more specific molecular mechanisms of how PPT1 induces ovarian HA by disturbing AR transduction in patients with PCOS. In the presence of PPT1, HSP90 $\alpha$  is depalmitoylated at the sites of Cys598 and Cys599, and subsequently, damaged AR activation and function. This resulted in the inhibition of downstream *CYP19A1* gene transcription and contributed to the failure of androgen to estrogen conversion. With dipyridamole treatment, the low status of palmitoylated HSP90 $\alpha$  was restored, and deleterious effects caused by HA were counteracted (109).

Global or organ-specific AR knockout in DHT-induced mice completely or partially normalized the metabolic and reproductive abnormalities of PCOS, implying that PCOS symptoms are mediated by AR signaling. One study used TC-specific deletion of AR (ThARKO) mice models, only to identify no evident differences in steroid hormone profiles and fertility between the ThARKO group and control group, indicating that these effects are not mediated by AR expressed in TCs. When exposed to excess DHT, ThARKO mice exhibit milder reproductive phenotypes and similar hormone profiles compared with controls. Further studies have indicated that the elevated tissue inhibitor of metalloproteinase 1 (Timp1) is the key player in addressing reproductive dysfunctions. Thus, subfertility is mediated by AR expressed in TCs, although it is not involved in the regulation of sex hormone secretion in PCOS animal models (110). Intriguingly, most traits are still present in GC-specific loss of AR after DHT treatment, whereas anovulation and metabolic dysfunctions, including altered body composition and lipid profiles, are eliminated in neuron-specific AR knockout models (111). More specifically, ovulation is collectively controlled by leptin receptor (LepRb) neurons located in the ARN and the ventral premammillary nuclei in rodents. The study using PNA models with loss of AR in LepRb cells revealed no distinct improvement in metabolic functions and delayed sexual maturation compared with the control group. However, the disrupted estrous cycles were restored to a large degree (112). Muscle-specific AR knockout models displayed similar traits to controls after DHT exposure; however, some aberrant metabolic phenotypes ameliorated in adipose tissue-specific AR knockout mice, but not the reproductive features (113). The deletion of AR in the liver results in restored metabolic functions, such as impaired glucose homeostasis, whereas similar reproductive deficits remained in the control group after DHT exposure (114). Overall, these findings depict the key AR target sites outside of the ovary, including the brain, liver, and adipose tissues, and the crucial roles they play in the development of PCOS. Moreover, the subtype of androgens also matters. AR-knockout protects mice from reproductive and metabolic disorders induced by DHT administration. However, T treatment can still incur reproductive abnormalities. T may indirectly elicit its detrimental reproductive effects by further converting into E2, which can mediate its effects via the ER (115).

Of note, studying animal models mimicking PCOS-like symptoms, which allowed us to elucidate the mechanisms of this condition, had some drawbacks. The postnatal DHT exposure mouse displays typical traits of PCOS without elevated LH secretion. Specifically, despite increased AR expression, chronic androgen stimulation only exerts modest effects on the density of the GnRH spine as well as the number of neurons expressing ERa (116). This may be induced by downregulated kisspeptin activity, thereby decreasing control over GnRH neurons (117). Similarly, one recent study compared several kinds of PCOS animal models at different androgen exposure windows. Gene ontology enrichment analysis revealed that the hypothalamic differentially expressed genes are less enriched in the adult programming group (118). These contradictory results strongly suggest the importance of the timing of androgen exposure and androgen patterns. Although AR expression increases in neuropeptide Y (NPY) neurons, which control eating behaviors, the projections of NPY neurons to GnRH neurons did not change significantly in the PNA model (30). Therefore, different androgen actions may be regulated by different neural circuits.

# **10** Therapeutic strategies

Lifestyle modifications are first-line treatments for PCOS. In recent decades, promising strides have been made in the field of PCOS, and more therapeutic options to restore fertility and ameliorate endocrine and metabolic abnormalities are available. However, most of them are off-label, e.g., insulin-sensitizing agents, some nutritional supplements, and phytochemicals. This Review mainly focuses on alleviating the androgen excess in PCOS patients. Conventional treatments to relieve androgen-related symptoms include oral contraceptive pills and anti-androgen agents.

### 10.1 Oral contraceptive pills

Estrogen reduces the bioactivity of androgen by increasing SHBG contents. One study pointed out that the 6 months of OCP treatments worsened the IR state and hepatic lipid accumulation, which is closely associated with exacerbated inflammation (119, 120). A combined OCP, including estrogen and progestin components, provides greater benefits for HA, but increased thrombotic risk limits its use (121).

### 10.2 Anti-androgen agents

Common anti-androgen medications include flutamide, spironolactone, and finasteride (inhibitor of  $5\alpha$ -reductase). The safety and effects of low-dose flutamide, an androgen receptor antagonist, on alleviating hyperandrogenic symptoms have been well established in clinical practice. Spironolactone, through binding to the AR, has shown strong efficacy in improving hirsutism, acne, and sebum production. However, there are no positive effects on the aspects of BMI, LH, and FSH (122). Notably, contraception is required due to its malformation risk. Abiraterone acetate (AA) administration significantly lowers the 11-oxygenated androgens both in prostate cancer and women with classic 21hydroxylase deficiency due to its characteristic of inhibiting CYP17A1 activity. However, its role in reducing 11-oxygenated androgens in PCOS needs further investigation (123).

### 10.3 Phytochemicals

In recent years, there has been increased attention on the therapeutic potentials of phytochemicals in the management of PCOS, such as resveratrol, quercetin, proanthocyanidins, berberine, and soy isoflavones. These compounds are mainly derived from flavonoids, polyphenols, and alkaloids. In terms of molecular mechanism, their protection primarily focuses on regulating steroidogenic enzymes, improving IR, and reducing inflammation, apoptosis, and oxidative stress (124). Novel natural compounds like *Tinospora cordifolia* and *Garcinia cambogia* Desr have shown beneficial properties in improving hormonal parameters and restoring ovarian functions (125, 126). Of note, these anti-androgenic and therapeutic effects are mainly validated from the letrozole-induced or DHEA-induced rat models. More randomized controlled trials and further clinical studies investigating efficacy are needed.

### **10.4 Supplements**

A study has shown that myo-inositol treatment effectively lowers the LH/FSH ratio and increases the expression of aromatase and FSHR, whereas its isomer D-chiro-Ins (DCI) exhibits the inverse effects on ovarian steroidogenesis (127). Therefore, it has been reported that when given DCI and myoinositol at a ratio of 1:40, it can significantly mitigate the abnormalities of PCOS and improve insulin sensitivity (128). Evidence has supported the close relationship between vitamin D (VD) and female fertility, regulating IR, inflammatory response, and oxidative stress. It has been reported that there is a higher prevalence of VD deficiency in PCOS patients with HA. VD supplementation in women with PCOS is helpful in improving PCOS symptoms and related indices without adverse effects (129).

# 10.5 Nanotechnologies

Selenium nanoparticles (SeNPs) are promising tools with lower toxicity and enhanced bioavailability to target multiple diseases for their anti-inflammatory and anti-antioxidant properties (130). One study demonstrated that SeNPs reduce androgen production by downregulating the expression of key steroidogenic enzymes, including STAR, CYP11A1, CYP17A1, and HSD17B3, while increasing the expression of CYP19 $\alpha$ 1. Additionally, the block of AR expression was found (131). Fourteen days of low-dose SeNP supplementation normalizes the aberrant metabolic features of PCOS and inhibits the expression of inflammatory factors, including IL-6, TNF- $\alpha$ , and IL-1, via repairing the antioxidant KEAP1/NRF2 cascades (132). The combination with metformin largely alleviates IR and metabolic abnormalities by upregulating PI3K-AKT signaling (133).

### 10.6 Target circadian rhythm

In recent years, promising therapies modifying circadian rhythm to alleviate HA have been proposed. Long-term exposure to light would result in increased FSH and estradiol levels. Interestingly, prolonged darkness would lead to HA and IR in PCOS rat models. Downregulated melatonin receptor 1A disturbs AR expression and downstream CYP19A1 transcription (134). Another study revealed that the dysregulation of some core clock gene expression is closely associated with HA. CYP17A1 has been identified as the direct target of the brain and muscle ARNT-like 1 (BMAL1) gene in peripheral blood mononuclear cells (PBMCs), and low expression of BMAL1 increased the activity of  $5\alpha$ reductase, catalyzing more T into DHT (135). In the liver, BMAL1 inhibits GLUT4 expression, whereas period (PER1/2) interferes with AR and the production of hepatic IGFBP4 and SHBG, ultimately causing IR and androgen excess (136). Moreover, androgen directly disrupts the hepatic timing system, ultimately impairing liver metabolism through methylation modification of H3K27me3 (137). Additionally, BMAL1 is considered an activator of luteinizing hormone receptor (LHCGR) transcription, which promotes LH surge and ovulation (138). Clinical trials have shown that the administration of melatonin at 5 mg for 12 weeks significantly lowers the levels of T and biomarkers of inflammation and alleviates hirsutism in women with PCOS (139).

### 10.7 Target neuroendocrine dysfunction

New therapies targeting the abnormal GnRH neuron circuits have been developed. Neurokinin 3 receptor (NK3R) antagonist (MLE4901) mitigates GnRH pulsatility, androgen excess, and metabolic abnormalities in a DHT-induced PCOS mouse model, with no effect on reproductive traits (140). Notably, antagonizing the local ovarian NK3R systems also alleviates the PCOS-like symptoms. In terms of mechanism, treatment with NK3R antagonist restored mitochondrial function and decreased oxidative stress in KGN cells (141). In addition, as mentioned before, NPY is found to participate in the modulation of reproduction and metabolism in rats. Hypothalamic NPY enhances GnRH-stimulated LH secretion and inhibits kisspeptin. It has been shown that NPY Y5 receptor antagonists significantly suppress weight gain and food intake (142). Further research and clinical trials are required before this neuropeptide can be used to treat the metabolic and reproductive dysfunctions observed in PCOS.

### 10.8 Mesenchymal stem cell therapy

Recently, mesenchymal stem cell (MSC) therapies have been seen as promising and safe tools to reverse some PCOS traits, including mitigating inflammation and restoring fertility. After the injection of human bone marrow MSCs into the ovary, androgensynthesizing gene expression, including *CYP17A1* and *DENND1A*, was significantly downregulated, while *FSHR* and *CYP19A1* gene expression was upregulated. These effects were partly mediated by secretome components IL-10 and BMP-2 (143, 144). One study further uncovered several key protein components from hMSCs regulating androgen production (145).

### 11 Conclusion and future directions

HA is a key feature of PCOS. The utilization of selective or complete AR knockout animal models has revealed that HA induces PCOS traits via AR and highlights the extra ovarian target sites of AR action, including the brain, adipose tissue, and liver, of which the brain is the core site.

The current therapeutic approaches aimed at alleviating hyperandrogenic manifestations may not be applicable to patients with phenotype D, based on the Rotterdam classification. Conventionally, biochemical androgen measurements primarily focus on total T or T levels and the free androgen index. The application of LC-MS/MS enables the simultaneous measurement of several kinds of steroid hormones in women with PCOS. Recently, A4 has been identified as a sensitive marker for identifying women with hyperandrogenic PCOS. Clinical hyperandrogenism is defined as the presence of hirsutism recorded as an m-FG score of  $\geq 8$ , which can become unreliable when applied to different ethnicities. Therefore, these traditional perspectives have been challenged. Apart from the previous findings that ovarian androgen is a dominant source of androgen excess in women with PCOS, adrenal-derived androgen, particularly 11-oxygenated androgen, also contributes to the circulating androgen pools.

HA in PCOS is characterized by an overall elevation of serum androgen levels via the involvement of classic, 11-oxygenated, and backdoor pathways (146). Therefore, future research should aim to identify more precise detection methods covering the entire androgen profile and quantity of androgen, as well as variations in the menstrual cycle, circadian rhythms, age, BMI, and samples from different collection sites (saliva, urine, and blood), which may all potentially influence the measurement of androgen levels (89). Moreover, serum androgen levels are not completely equivalent to the local conversion of androgen. Different bioactivities and binding affinities of androgens to AR are also important. In conclusion, the current understanding of the regulatory role of androgens in the development of PCOS remains limited; therefore, continuous efforts to determine the exact mechanism of this condition are required.

## Author contributions

KW: Conceptualization, Investigation, Methodology, Writing – original draft, Writing – review & editing. YL: Supervision, Writing – review & editing. YC: Methodology, Writing – review & editing.

# Funding

The author(s) declare that no financial support was received for the research, authorship, and/or publication of this article.

### Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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