



OPEN ACCESS

EDITED BY
Rudolf Lucas,
Augusta University, United States

REVIEWED BY
Surapaneni Krishna Mohan,
Panimalar Medical College Hospital and
Research Institute, India
Radha Chaube,
Banaras Hindu University, India

*CORRESPONDENCE
Yanlong Qiao
✉ m18302546688@163.com
Yanchun Hu
✉ hychun114@163.com

RECEIVED 03 May 2023
ACCEPTED 27 July 2023
PUBLISHED 17 August 2023

CITATION
Liao F, Yousif M, Huang R,
Qiao Y and Hu Y (2023) Network
pharmacology- and molecular docking-
based analyses of the antihypertensive
mechanism of *Ilex kudingcha*.
Front. Endocrinol. 14:1216086.
doi: 10.3389/fendo.2023.1216086

COPYRIGHT
© 2023 Liao, Yousif, Huang, Qiao and Hu.
This is an open-access article distributed
under the terms of the [Creative Commons
Attribution License \(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use,
distribution or reproduction in other
forums is permitted, provided the original
author(s) and the copyright owner(s) are
credited and that the original publication in
this journal is cited, in accordance with
accepted academic practice. No use,
distribution or reproduction is permitted
which does not comply with these terms.

Network pharmacology- and molecular docking-based analyses of the antihypertensive mechanism of *Ilex kudingcha*

Fei Liao^{1,2}, Muhammad Yousif¹, Ruya Huang¹, Yanlong Qiao^{2*} and Yanchun Hu^{1*}

¹Key Laboratory of Animal Disease and Human Health of Sichuan Province, College of Veterinary Medicine, Sichuan Agricultural University, Wenjiang, China, ²Department of Animal Husbandry and Fisheries, Guizhou Vocational College of Agriculture, Qingzhen, China

Herein, network pharmacology was used to identify the active components in *Ilex kudingcha* and common hypertension-related targets. Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) enrichment analyses were conducted, and molecular docking was performed to verify molecular dynamic simulations. Six active components in *Ilex kudingcha* were identified; furthermore, 123 target genes common to hypertension were identified. Topological analysis revealed the strongly associated proteins, with *RELA*, *AKT1*, *JUN*, *TP53*, *TNF*, and *MAPK1* being the predicted targets of the studied traditional Chinese medicine. In addition, GO enrichment analysis revealed significant enrichment of biological processes such as oxidative stress, epithelial cell proliferation, cellular response to chemical stress, response to xenobiotic stimulus, and wound healing. Furthermore, KEGG enrichment analysis revealed that the genes were particularly enriched in lipid and atherosclerosis, fluid shear stress and atherosclerosis, and other pathways. Molecular docking revealed that the key components in *Ilex kudingcha* exhibited good binding potential to the target genes *RELA*, *AKT1*, *JUN*, *TP53*, *TNF*, and *IL-6*. Our study results suggest that *Ilex kudingcha* plays a role in hypertension treatment by exerting hypolipidemic, anti-inflammatory, and antioxidant effects and inhibiting the transcription of atherosclerosis-related genes.

KEYWORDS

Ilex kudingcha, hypertension, network pharmacology, molecular target, molecular dynamic simulations

1 Introduction

Hypertension is the primary risk factor for mortality and the most frequent reason for renal, cardiovascular, and cerebrovascular disorders. Globally, approximately 1 billion individuals suffer from hypertension; it is directly responsible for 13% of all fatalities (1). In China, one in four individuals suffers from hypertension, with 40% suffering from severe hypertension. However, most individuals are unaware of their disease and receive inadequate care (2). The use of antihypertensive drugs is the most effective method for lowering blood pressure and preventing cardiovascular events; however, only 32.5% of patients with hypertension worldwide have controlled blood pressure (3). In general, blood pressure is controlled by using a combination of two or more antihypertensive drugs (4). In less developed areas, maintaining adequate blood pressure control is difficult because the medication is lowered (5). Epidemiological studies have reported that the prevalence of resistant hypertension is 10% among individuals with hypertension, with high cardiovascular risk in this patient cohort (1). Furthermore, despite conscientious clinical management, many adults with resistant hypertension fail to achieve their recommended blood pressure treatment targets on three antihypertensive medications or require more than four medications to achieve their targets (6).

Considering the significant role of diet in blood pressure homeostasis, the High Blood Pressure Clinical Practice Guidelines of the American College of Cardiology/American Heart Association recommend dietary strategies as a practical and acceptable approach to control blood pressure (7). Hippocrates mentioned that “Let food be thy medicine and medicine be thy food” (8). In China, *Ilex* species exhibit a broad geographic range, and some species have been used to develop daily herbal tea blends. *Ilex kudingcha*, a Chinese herbal tea, possesses many health-improving properties (9). It is a well-known traditional Chinese beverage in Southeast Asia. *Ilex kudingcha* contains saponins, polyphenols, and flavones and exerts anti-inflammatory (10), antioxidative (11), antiaging (12), anticancer (13, 14), antiobesity (15, 16), antihypertensive, and antidiabetic effects (17).

However, the ingredients and molecular mechanisms underlying its antihypertensive effects remain unelucidated. Network pharmacology combines system network analysis with pharmacology, bioinformatics, and other disciplines to demonstrate the multicomponent and multitarget drug treatment process from the perspective of genes (18). By building a network associated with “disease–phenotype–gene–drug,” the distribution, molecular function, and signaling pathways of traditional Chinese medicine (TCM) compounds may be investigated. At present, the basis and mechanism of the pharmacodynamic components in TCMs are frequently predicted using network pharmacology approaches (19). Therefore, the present study aimed to use network pharmacology and molecular docking to determine the molecular targets and processes involved in hypotension treatment.

2 Materials and methods

2.1 Screening of active ingredients and target genes

The parameters oral bioavailability (OB) of 30% and drug-like (DL) characteristic of 0.18 p were used in the Traditional Chinese Medicine Systems Pharmacology (TCMSP) platform (<http://lsp.nwu.edu.cn/tcmsp.php>) to screen the active ingredients in *Ilex kudingcha*. The DrugBank database (<https://www.drugbank.ca/>) and peer-reviewed literature were used to identify the expected targets of the tested compounds. Furthermore, UniProt (<https://www.uniprot.org/>) was used to standardize gene names and compare target information. The *Ilex kudingcha*–ingredient–target regulatory network was developed using Cytoscape 3.9.1 software.

2.2 Collection of hypertension-related targets

The hypertension-related targets with high relevance were chosen using GeneCards (<https://genecards.org>). The targets with relevance scores of ≥ 1 were chosen based on the keyword “hypertension.” The hypertension-related targets were identified after weight removal and consolidation.

2.3 Identification of intersection targets and development of the “drug–disease–target” regulatory network

The hypertension-related target genes and intersection genes of *Ilex kudingcha* were acquired using R software. Then, the *Ilex kudingcha*–disease–target regulatory network was developed using Cytoscape 3.9.1 software.

2.4 Protein–protein interaction network and topological analysis

The intersection targets of *Ilex kudingcha* and hypertension were imported into the STRING database (<https://string-db.org/cgi/input.pl>). The species was limited to humans and a minimum interaction score of 0.9 was used. The key targets were imported into the Cytoscape program. The interaction network diagram of the target proteins of the active ingredients in *Ilex kudingcha* and hypertension-related target proteins was developed by sorting based on degree value.

2.5 Gene Ontology and Kyoto Encyclopedia of Genes and Genomes enrichment analyses

R 4.2.2 with the “colorspace,” “stringi,” and “ggplot2” packages was installed. The “DOSE,” “clusterProfiler,” and “annotationHub” Bioconductor packages were used for Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) enrichment analyses. For GO enrichment analysis, the function “enrichGO” was employed. For KEGG enrichment analysis, the database org.Hs.eg.db and the “enrich-KEGG” tool were used. The KEGG database (<https://www.kegg.jp/>) was also used (doi: 10.18129/<http://b9.bioc.org.Hs.eg.db>). For the parameters of the two functions, species was set to “has,” and the filter values (P - and q -values) were set to 0.05. A bubble graph was prepared to visualize the top 10 enrichment findings, and Cytoscape 3.9.1 was used to develop the KEGG regulatory network.

2.6 Molecular docking

Three-dimensional (3D) protein conformations with a crystal resolution of <3 , as determined using X-ray crystal diffraction, were gathered by searching the Protein Data Bank (PDB) (<https://www.rcsb.org>) for the target genes implicated in the first eight protein–protein interaction (PPI) findings. The primary active ingredients in *Ilex kudingcha* were retrieved in a two-dimensional (2D) structure format from the PubChem website. The sdf files were converted into the pdb file format using Discovery Studio 2019 software. PyMOL 4.3.0 software was used to separate the original ligand and protein structures and to dehydrate and remove the organics. Furthermore, AutoDock Vina 1.1.2 software was used to process the proteins as follows: non-polar hydrogen was added, the Gasteiger charge was calculated, the AD4 type was assigned, and the flexible bonds of small molecules/ligands were set to be rotatable. Based on the original ligand coordinates, the docking box was adjusted to include all protein structures. Furthermore, the receptor protein was set to a semiflexible butt joint, and the Lamarckian genetic algorithm was selected. The docking results were obtained by running autogrid4 and autodock4; as a result, the binding energies were

revealed. PyMOL 4.3.0 and Discovery Studio 2019 software were used to analyze and visualize the forces in 3D and 2D.

2.7 Molecular dynamic simulation

To further elucidate the stability of protein–ligand binding, GROMACS 2023.36 software was used for molecular dynamic (MD) simulation. The AMBER force field was used to describe the proteins and ligand molecules. The water model was set to the SPC water model. The system temperature was set to a vacuum environment of 300 K, and the simulation time was 100 ns. First, the energy balance of the simulated system was determined by using the steepest gradient algorithm under the absolute vacuum environment. After energy balance, MD simulation was completed; thereafter, root mean square deviation (RMSD) analysis of the simulation results was conducted to analyze the relative binding stability between the two chains in the simulation process. Furthermore, root mean square fluctuation (RMSF), which demonstrates the structural adaptability of each protein residue, was used to analyze the flexibility and intensity of movement of the amino acid residues of the protein throughout the simulation. The radius of gyration (Rg) was used to characterize the compactness of the protein structure and changes in the looseness of the peptide chain during the simulation. VMD software was used to analyze the changes in the number of hydrogen bonds formed between the two chains over time as well as to observe whether the bond was stable from the interaction point of view.

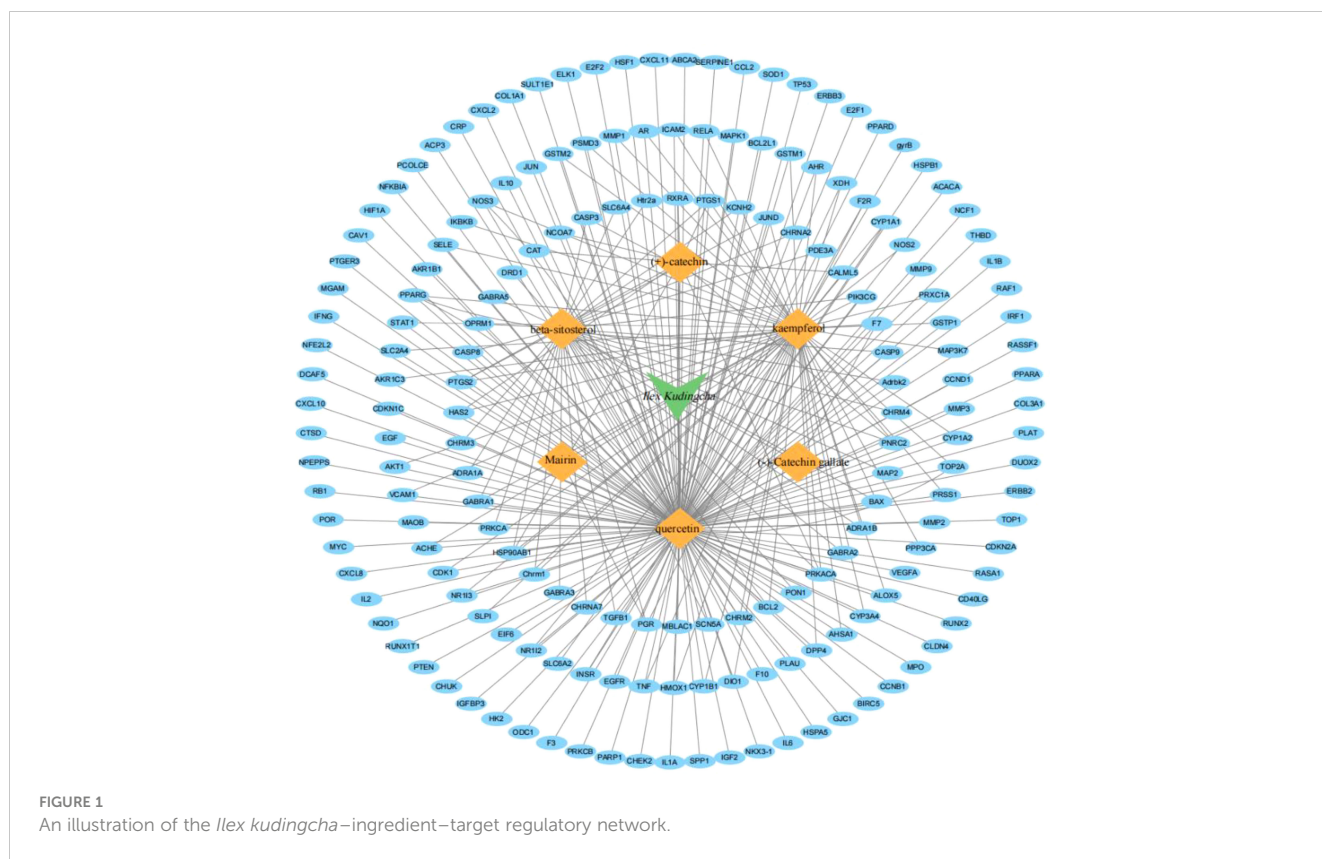
3 Results

3.1 Screening of active ingredients and target genes

The TCMSP database contains 94 active ingredients in *Ilex kudingcha*. They were screened using an OB of $\geq 30\%$ and a DL of 0.18. Subsequently, six active ingredients were obtained (Table 1). The targets were predicted using DrugBank and UniProt. Finally, 179 targets were obtained (153 for quercetin, 1 for mairin, 63 for kaempferol, 37 for beta-sitosterol, 11 for (+)-catechin, and 1 for (–)-catechin gallate). Figure 1 illustrates the *Ilex kudingcha*–ingredient–target regulatory network.

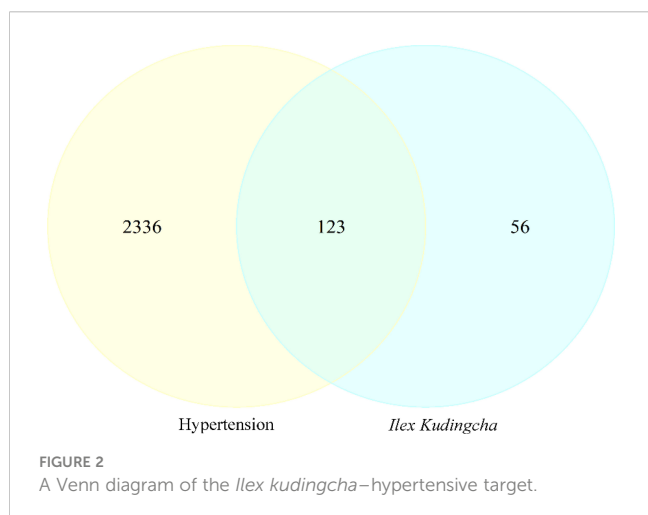
TABLE 1 The active ingredients of *Ilex kudingcha*.

Herbs	ID	Compound	OB (%)	DL
<i>Ilex kudingcha</i>	MOL000098	Quercetin	46.43	0.28
<i>Ilex kudingcha</i>	MOL000211	Mairin	55.38	0.78
<i>Ilex kudingcha</i>	MOL000422	Kaempferol	41.88	0.24
<i>Ilex kudingcha</i>	MOL000358	Beta-sitosterol	36.91	0.75
<i>Ilex kudingcha</i>	MOL000492	(+)-Catechin	54.83	0.24
<i>Ilex kudingcha</i>	MOL006504	(–)-Catechin gallate	53.57	0.75



3.2 Identification of intersection targets and development of the “drug–disease–target” regulatory network

A total of 2,459 hypertension-related target genes were obtained, with 123 intersection targets between *Ilex kudingcha* and hypertension (Figure 2). Figure 3 illustrates the *Ilex kudingcha*–ingredient–target–hypertension regulatory network. The active ingredients kaempferol and quercetin were associated with 42 and 111 target genes, respectively. Therefore, they were



classified as multitarget and multieffect compounds. The genes *PTGS1*, *PTGS2*, *PRKACA*, and *PPARG* were associated with the highest number of active components.

3.3 PPI network and topological analyses

The 123 intersection targets were imported into the STRING database to construct the PPI network; the species was limited to humans and a minimum interaction score of 0.9 was used. As shown in Figure 4, 123 protein nodes and 377 edges were obtained for the intersection genes. In the PPI network, the degree centrality of a node is simply the number of edges it has. The higher the degree, the more central the node is. The interaction network diagram of the target proteins of the active ingredients and hypertension-related target proteins of *Ilex kudingcha* was obtained by ranking the proteins based on their degree value. Finally, 105 core target genes were obtained. As demonstrated in Figure 5, the higher the degree value, the darker the color and the larger the circle. The main targets were *RELA*, *AKT1*, *JUN*, *TP53*, *TNF*, and *MAPK1*, with degree values of 58, 58, 58, 56, 50, and 50, respectively.

3.4 Analysis of GO function and KEGG enrichment of related targets

GO enrichment analysis revealed the gene functions at three levels: biological process (BP), cellular component (CC), and molecular

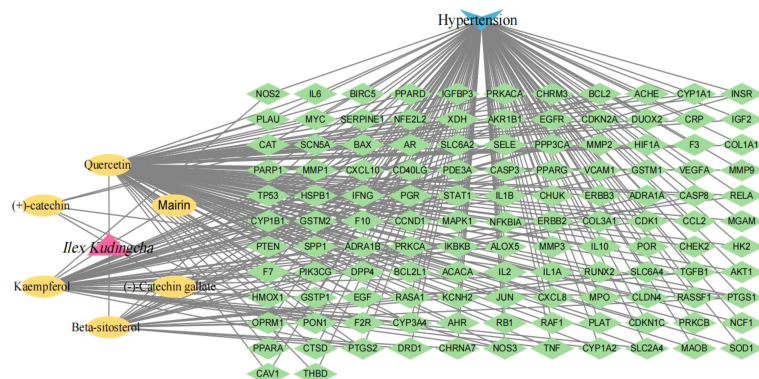


FIGURE 3 An illustration of the *Ilex kudingcha*–ingredient–target–hypertension regulatory network.

function (MF). BP was mainly associated with response to oxidative stress, epithelial cell proliferation, cellular response to chemical stress, response to xenobiotic stimulus, and wound healing. CC was mainly associated with the membrane raft, membrane microdomain, vesicle lumen, secretory granule lumen, cytoplasmic vesicle lumen, and plasma membrane raft. MF was mainly associated with DNA-binding transcription factor binding, RNA polymerase II-specific DNA-binding transcription factor binding,

signaling receptor activator activity, receptor–ligand activity, cytokine receptor binding, and cytokine activity (Figure 6).

KEGG enrichment analysis revealed that the antihypertensive mechanism of *Ilex kudingcha* was mainly concentrated in lipid and atherosclerosis, fluid shear stress and atherosclerosis, the AGE-RAGE signaling pathway in diabetic complications, human cytomegalovirus infection, the TNF signaling pathway, chemical carcinogenesis receptor

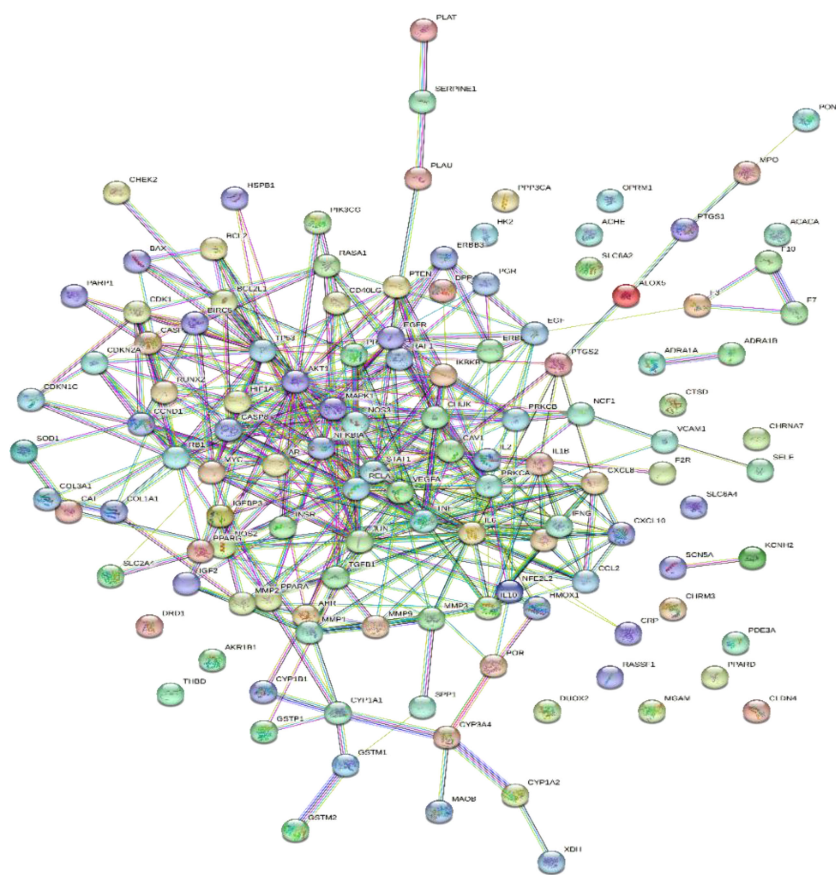
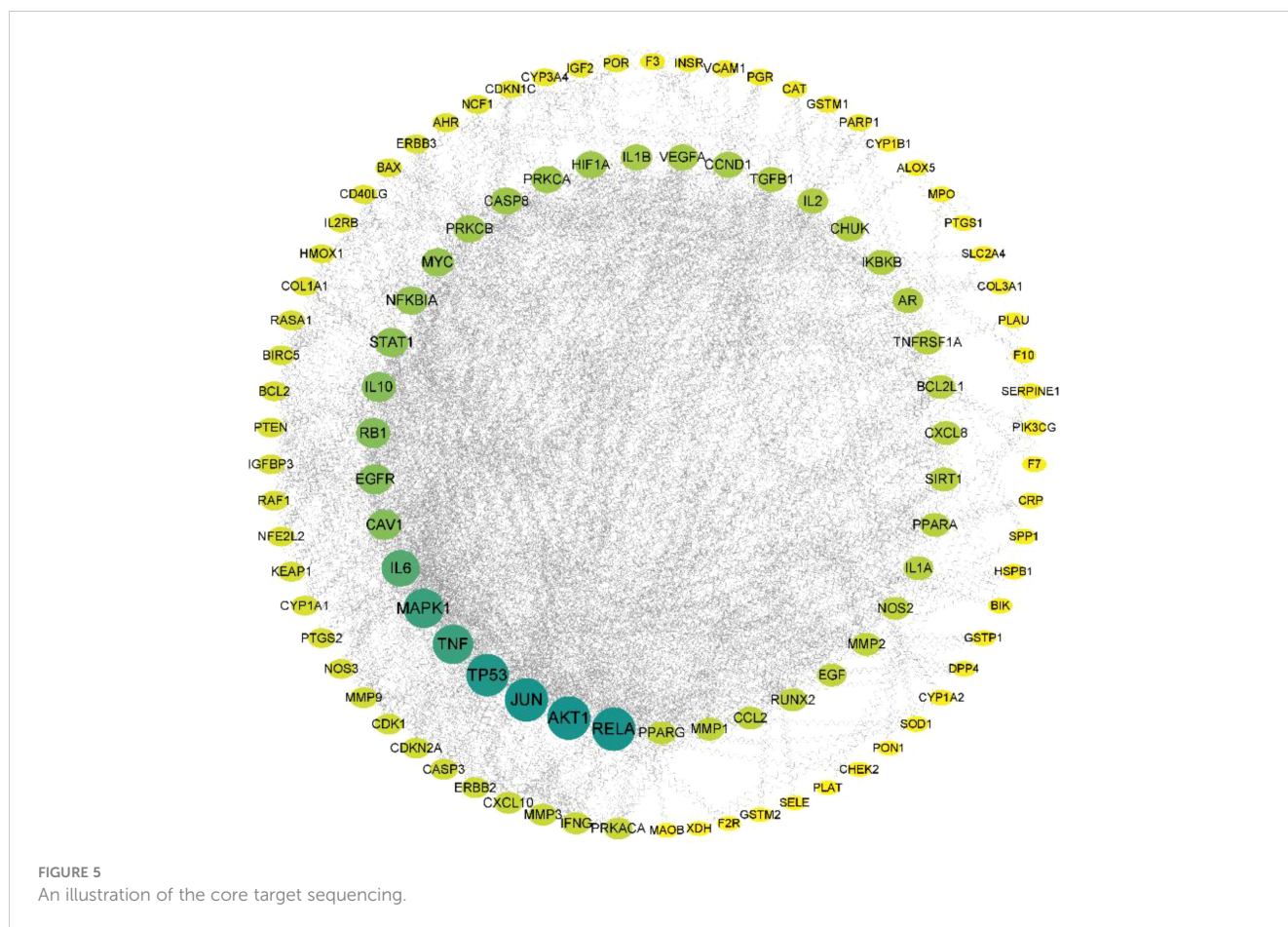


FIGURE 4 The protein–protein interaction network.



activation, hepatitis B, prostate cancer, and Chagas disease (Figure 7). The genes associated with the highest number of pathways were *AKT1*, *RELA*, and *MAPK1* (Table 2). Lipid and atherosclerosis may be the most important pathway for *Ilex kudingcha* to treat hypertension (Figure 8).

3.5 Molecular docking

The structures of the small molecules quercetin (PubChem CID: 5280343) and kaempferol (PubChem CID: 5280863) were downloaded from PubChem. Furthermore, the *AKT1*, *RELA*, *TNF*, *IL-6*, *JUN*, *MAPK1*, *RB1*, and *TP53* proteins with PDB IDs of 2uvm, 1nfi, 5uui, 1alu, 1jUN, 6g54, 1ad6, and 6ggc, respectively, were downloaded from PDB. The binding energies between quercetin and *AKT1*, *RELA*, *TNF*, *IL-6*, *JUN*, *MAPK1*, *RB1*, and *TP53* proteins were -6.1 kcal/mol, -7.4 kcal/mol, -8.8 kcal/mol, -7.0 kcal/mol, -5.7 kcal/mol, -8.4 kcal/mol, -7.4 kcal/mol, and -7.1 kcal/mol, respectively. Furthermore, the binding energies between kaempferol and *AKT1*, *RELA*, and *TNF* proteins were -6.1 kcal/mol, -7.4 kcal/mol, and -8.8 kcal/mol, respectively. The docking results were less than -5 kcal/mol. In general, if the binding energy of the ligand to the target protein is

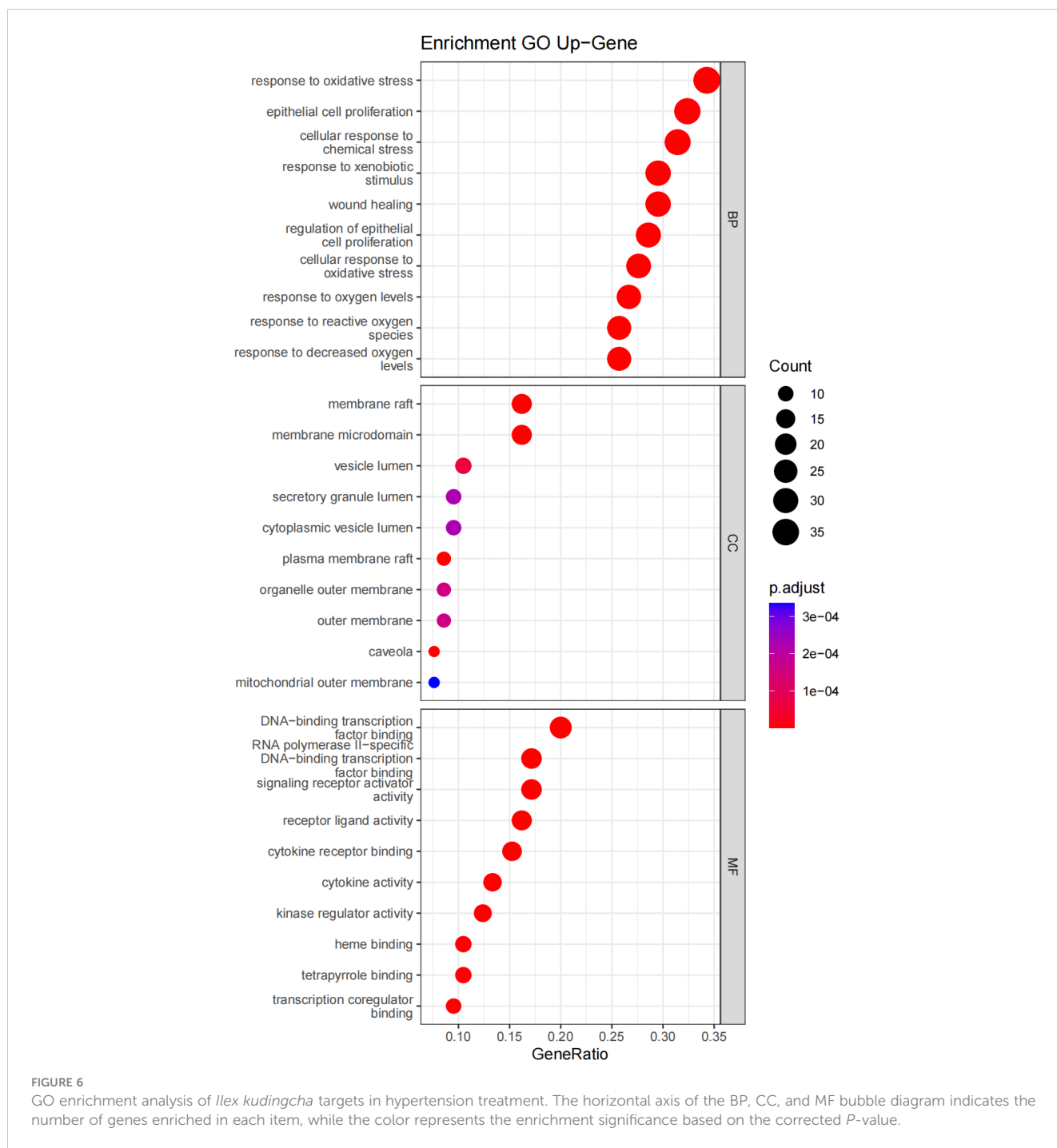
less than -5 , the binding between the ligand and receptor protein is stable. Table 3 displays the detailed results. Figure 9 comprehensively illustrates several regional molecular docking structures.

3.6 MD simulation

Molecular docking revealed that quercetin interacts with *TNF* primarily via the formation of hydrophobic forces and hydrogen bond forces. It binds to the amino acid residues VAL-17, PRO-20, and ARG-32 of the receptor protein via four hydrophobic bonds and to ALA-18, ARG-32, GLY-148, GLN-149, VAL-150, and amino acid residues of the receptor protein via seven hydrogen bonds (Figure 9D). To further elucidate the stability of protein–ligand binding, an MD simulation was performed.

3.6.1 RMSD

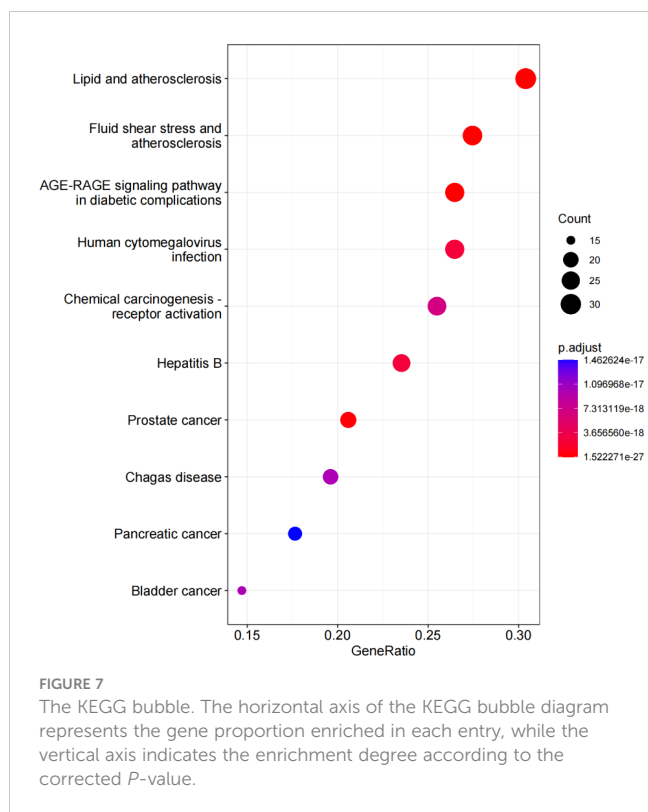
The RMSD curve revealed the RMSD of the structure during the simulation. The movement trajectory of the binding complex demonstrated that the amplitude of quercetin and *TNF* was small during the whole simulation process. Furthermore, the RMSD curve



of quercetin revealed that quercetin vibrated near 0–0.15 nm in the whole simulation process and near 0–0.1 nm in the first 30 ns (Figure 10A). Moreover, the RMSD curve of TNF demonstrated that the RMSD value kept vibrating at approximately 0.1–0.2 nm at 30 ns, followed by an upward trend of vibration to 0.2–0.3 nm and finally maintaining dynamic stability (Figure 10B). Taken together, the results suggest that the complex structure reached a stable conformation after 30 ns.

3.6.2 RMSF

RMSF analysis, which indicates the structural fitness of each protein residue, was performed in this study to analyze the flexibility and exercise intensity of amino acid residues in the protein throughout the simulation. After the amino acids of the TNF protein 5uui were completely sequenced and the discontinuous part was reordered, RMSF analysis was performed. The TNF protein showed weak jitter in simulation, with the amplitude



distribution ranging from 0.05 to 0.4 nm; the peak value appeared at Res9, Res97, and Res100; and the RMSF value was the largest. The reason was Res 9 was present at the N-terminal of the protein, and the flexibility of the two ends of the protein was relatively large (Figure 11). To summarize, the RMSF value at both ends of the protein was large, whereas that of the amino acid residues was generally small, indicating stable protein–ligand binding.

3.6.3 Rg

Rg can help characterize the compactness of a protein structure and a change in the peptide chain looseness of a protein during simulation. Here, the overall vibration amplitude of the TNF protein was small and remained relatively stable at 1.5–1.55 nm (Figure 12), indicating that the protein–ligand complex remained relatively stable during simulation.

3.6.4 Hydrogen bonds

The number of hydrogen bonds formed between the TNF protein and quercetin was analyzed with time during a 100-ns simulation. A maximum of three hydrogen bonds were formed at 100 ns, whereas no hydrogen bonds were formed near 90 ns. Mostly, hydrogen bonds were formed 1–2 (Figure 13).

4 Discussion

TCM is beneficial for treating complicated medical conditions owing to its holistic approach. However, multiple targets, components, and pathways associated with the action mechanism of TCM in disease treatment complicate its development (20). The properties of a Chinese medicinal formula and its mode of action, such as integrity, systematization, and comprehensiveness, are similar to those of network pharmacology (21). The study of the pharmacological mechanisms of Chinese medicines can be well-fitted into pharmacology equations (22).

In the present study, a network pharmacological analysis was performed on the active pharmaceutical components of *Ilex kudingcha* for hypertension. The active compounds quercetin and kaempferol showed the highest number of targets. In the Liu et al. study, the PPI network analysis results showed that the main antihypertensive components of *Ginkgo folium* included kaempferol, quercetin, and luteolin (23). Ye et al. found that the antihypertensive active ingredients of *Eucommia ulmoides* included quercetin, kaempferol, and rutin (24). Yang et al. reviewed Chinese herbal prescriptions for hypertension and found that components including quercetin, luteolin, linolenin, and kaempferol were closely associated with the regulatory targets of hypertension (25). Moreover, the present molecular docking results showed that the active components exhibited effective binding ability with most of the target genes. Quercetin is a typical natural flavonoid, which is a secondary metabolite found in plants and is thought to be one of the bioactive substances found in fruits and vegetables; it is beneficial in maintaining cardiovascular health (26). Quercetin is advantageous in decreasing blood pressure and inflammation at the highest effective dosage of 500 mg of the aglycone form (27). Kaempferol, another natural flavonoid, is an effective anti-inflammatory, antioxidant, and anticancer agent and is documented in treating

TABLE 2 The enrichment pathways corresponding to the intersection genes.

Term	Description	Count	Gene ID
has05417	Lipid and atherosclerosis	31	AKT1 BAX BCL2 BCL2L1 CASP3 CASP8 CD40LG CHUK CYP1A1 IKKB IL1B IL-6 CXCL8 JUN MMP1 MMP3 MMP9 NFE2L2 NFKBIA NOS3 PPARG PRKCA MAPK1 RELA CCL2 SELE TNF TNFRSF1A TP53 VCAM1 NCF1
hsa05418	Fluid shear stress and atherosclerosis	28	AKT1 BCL2 CAV1 CHUK GSTM1 GSTM2 GSTP1 HMOX1 IFNG IKKB IL1A IL1B JUN MMP2 MMP9 NFE2L2 NOS3 PLAT RELA CCL2 SELE TNF TNFRSF1A TP53 VCAM1 VEGFA KEAP1 NCF1
hashsa05163	Human cytomegalovirus infection	27	AKT1 BAX CCND1 CASP3 CASP8 CDKN2A CHUK EGFR IKKB IL1B IL-6 CXCL8 MYC NFKBIA PRKACA PRKCA PRKCB MAPK1 PTGS2 RAF1 RB1 RELA CCL2 TNF TNFRSF1A TP53 VhasA
hsa05207	Chemical carcinogenesis receptor activation	26	AHR AKT1 BIRC5 AR CCND1 BCL2 CYP1A1 CYP1A2 CYP1B1 CYP3A4 EGF EGFR GSTM1 GSTM2 JUN MYC PGR PPARA PRKACA PRKCA PRKCB MAPK1 RAF1 RB1 RELA VEGFA

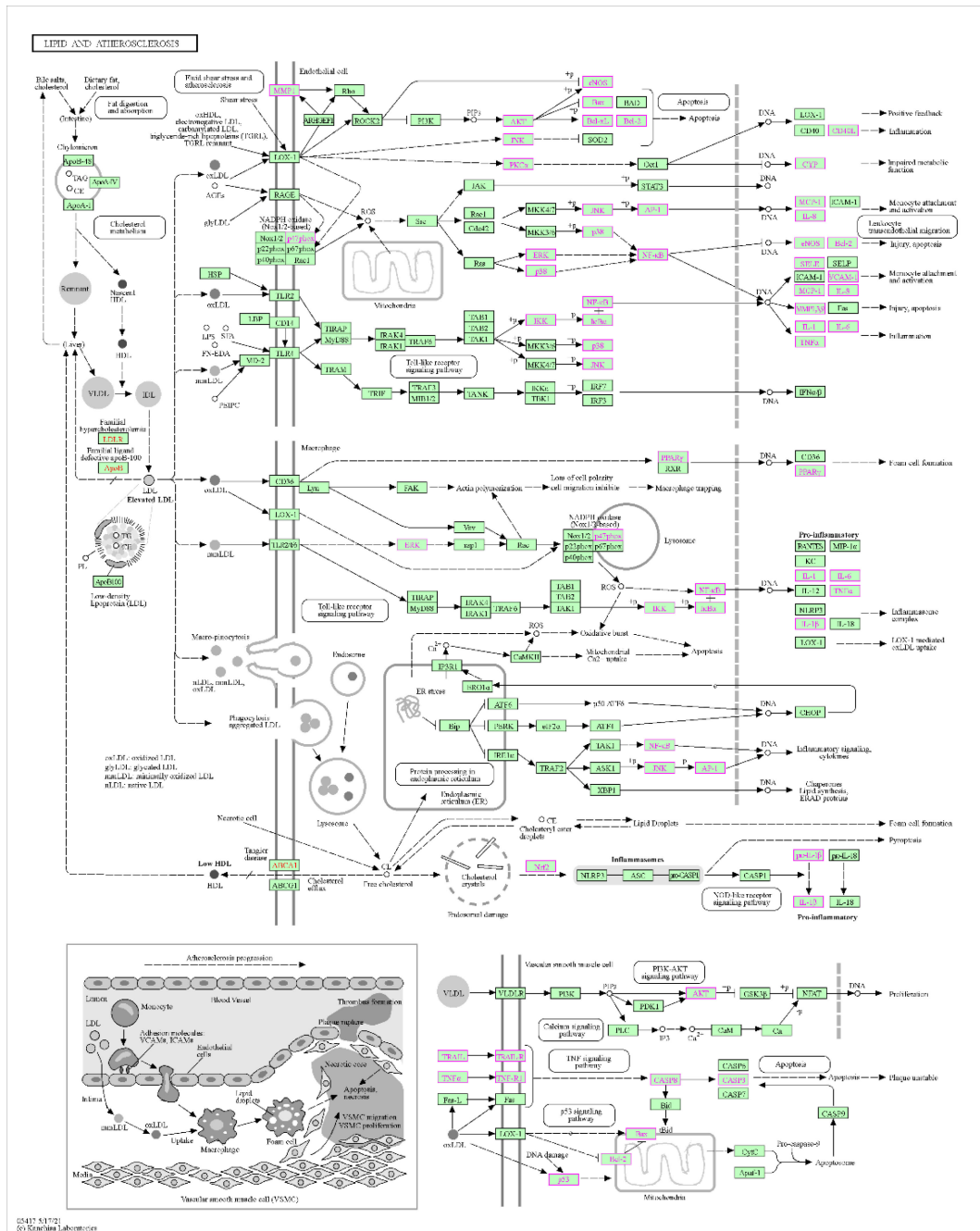


FIGURE 8 An important pathway of *Ilex kudingcha* treatment of hypertension disease—lipid and atherosclerosis.

several illnesses, including diabetes, obesity, and cancer (28). Quercetin and kaempferol exert the most significant hypolipidemic effects at a low concentration of 15 μM (29). Kaempferol exerts its effect in various ways, including decreasing the activity of human T cells and via phosphoinositide-3-kinase (PI3K)/AKT via human T-lymphotropic virus type 1 signaling pathways. The leukemia/lymphoma virus inhibits the production of several proteins that are the hallmarks of epithelial–mesenchymal

transition, such as Slug, N-cadherin, E-cadherin, and Snail and indicators of metastasis, including matrix metalloproteinase 2 (28).

The core genes were obtained based on the PPI network, and the main targets were *RELA*, *AKT1*, *JUN*, *TP53*, *TNF*, *MAPK1*, *IL-6*, *RBI*, *CAV1*, and *EGFR*, and their degree values were 58, 58, 58, 56, 50, 50, 44, 32, 32, and 32, respectively. *AKT1* is associated with hypertension, and *AKT1* mutations greatly increase the risk of hypertension. Furthermore, due to damage to *AKT1* signaling

TABLE 3 Binding energies of the *Ilex kudingcha* key components to the target gene molecules.

Compounds	Compound target	Docking score	Interaction							
			H-Bond interaction		Hydrophobic interaction		Π -Cation interaction		Π - Π stacking interaction	
			Distance (Å)	Amino acid	Distance (Å)	Amino acid	Distance (Å)	Amino acid	Distance (Å)	Amino acid
Quercetin	Quercetin- <i>AKT1</i> complex	-6.1	3.2	SER-2	3.6	GLN-113				
			3.8	SER-2						
			3.3	ASP-3						
			3.0	THR-105						
	Quercetin- <i>RELA</i> complex	-7.4	3.0	GLN-29	3.6	GLN-247	2.9	LYS-221		
			3.9	ARG-246	3.9	GLN-247				
			4.0	LYS-218	3.6	LYS-221				
			3.0	GLN-247	3.9	LYS-221				
			2.9	LYS-221	3.9	VAL-244				
	Quercetin- <i>TNF</i> complex	-8.8	3.9	ALA-18	3.5	VAL-17				
			3.1	ARG-32	3.8	VAL-17				
			3.2	GLY-148	3.5	PRO-20				
			3.9	GLN-149	3.6	ARG-32				
			3.1	VAL-150						
			3.1	VAL-150						
	Quercetin- <i>IL-6</i> complex	-7.0	2.8	ARG-30	4.0	LEU-33				
			3.9	ASP-34	4.0	LEU-33				
			2.9	ARG-30	3.8	GLN-175				
			4.1	ARG-182	3.9	LEU-178				
					4.0	ARG-179				
	Quercetin- <i>JUN</i> complex	-5.7	3.0	ARG-302	2.9	ASN-299				
			3.1	ARG-302	3.9	ASN-299				
			3.0	ASN-291	3.7	LEU-294				
			3.2	GLN-290	3.7	LEU-294				
			3.1	SER-292	3.5	ALA-295				
			3.1	ALA-298						
	Quercetin- <i>MAPK1</i> complex	-8.4	3.1	ASP-167	3.5	LYS-54				
			4.1	ALA-35	3.6	VAL-39				
2.9			MET-108	3.7	ILE-31					
4.0			LYS-114							
Quercetin- <i>RBI</i> complex	-7.4	3.1	SER-391	3.5	TYR-454					
		4.1	SER-391	3.6	ILE-388					
		2.7	GLU-458	2.9	ARG-455					
				3.4	ARG-455					
				4.0	ARG-455					

(Continued)

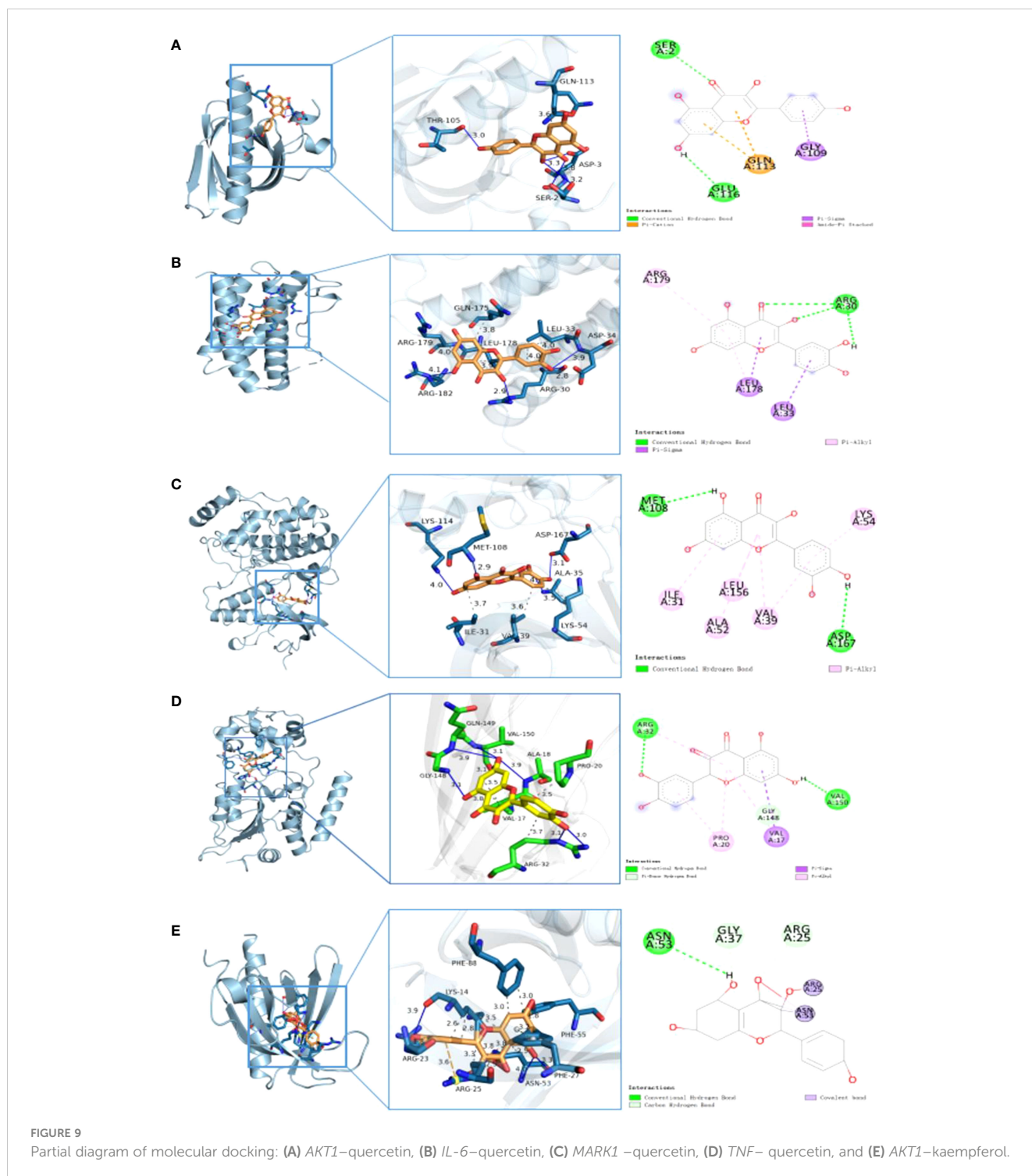
TABLE 3 Continued

Compounds	Compound target	Docking score	Interaction							
			H-Bond interaction		Hydrophobic interaction		Π -Cation interaction		Π - Π stacking interaction	
			Distance (Å)	Amino acid	Distance (Å)	Amino acid	Distance (Å)	Amino acid	Distance (Å)	Amino acid
	Quercetin-TP53 complex	-7.1	2.8	PRO-219	3.9	GLU-224				
			3.0	ASN-200	3.5	THR-230				
			4.0	HIS-233	3.0	GLU-221				
					4.0	GLU-221				
					3.6	VAL-218				
					3.6	ILE-232				
Kaempferol	Kaempferol-AKT1 complex	-6.1	3	PHE-88	2.5	ASN-53	3.6	ARG-25	3.2	PHE-27
			2.8	PHE-88	2.6	LYS-14				
			3.3	ASN-53	2.8	LYS-14				
			3.8	ASN-53	3.5	LYS-14				
			4.0	ASN-53	2.5	ARG-25				
			3.9	LYS-14	3.3	ARG-25				
			2	ARG-23	3.8	PHE-27				
	1.8	ARG-23								
	Kaempferol-RELA complex	-7.4	2.8	ARG-30	3.5	PHE-187				
			3	ARG-30	3.7	ALA-188				
			2.8	ASN-190	3.8	ASN-155				
	Kaempferol-TNF complex	-8.8	3	PRO-189						
			4	PHE-144	3.8	VAL-17				
					3.9	VAL-17				
				3.9	VAL-17					
				3.8	PRO-20					

pathways, insulin cannot be effectively transported to the vascular system by the action of endothelial nitric oxide synthase (*eNOS*), leading to obesity and insulin resistance, followed by artery dysfunction and hypertension (30, 31). *TNF* and *IL-6* are two proinflammatory cytokines associated with atherosclerosis, plaque development, and increased cardiovascular risk (32–34). In the present study, the core targets were involved in abnormal biological processes including inflammatory responses, lipid metabolism, vascular endothelial functions, and energy metabolism, which lead to hypertension.

GO enrichment was performed to identify disease–drug intersection genes. The findings revealed that the biological effects of the active ingredients of *Ilex kudingcha* were exerted via signaling cytokine receptor binding, cytokine activity, receptor–activator activity, and receptor–ligand activity

binding of transcription factors, which may have affected membrane rafts, membrane microdomains, cytoplasmic vesicle lumens, secretory granule lumens, plasma membrane rafts, and vesicle lumens, thus contributing to cellular responses to oxidative stress, epithelial cell proliferation, and biological processes including wound healing, reaction to xenobiotic stimulation, and chemical stress. The KEGG enrichment analysis indicated that the action mechanism of *Ilex kudingcha* in non-hypertensive atherosclerosis was primarily associated with lipids, fluid shear stress, and human cytomegalovirus infection, the *AGE–RAGE* signaling pathway in diabetes complications, and *TNF* activation of the chemical carcinogenesis signaling pathway. The lipid and atherosclerosis map (Figure 8) showed that the active ingredients of *Ilex kudingcha* acted on many targets and formed an interactive



relationship with atherosclerosis, fluid shear stress, and *MAPK*, *TNF*, and *PI3K/AKT* signaling pathways, all of which helped regulate oxidative stress and vascular hardness. *TNF* inhibits inflammatory responses while reducing blood pressure and excreting salt through the kidneys (35). The *PI3K/AKT* signaling pathway controls the cytoskeletal rearrangement and phenotypic transformation of arterial smooth muscle cells, affects the excitability of sympathetic nerves and the

function of vascular endothelial cells, and antagonizes angiotensin II (36, 37). Dyslipidemia and hypertension are considerable risk factors associated with the development of atherosclerotic cardiovascular illnesses (38) and induce various vascular events by promoting atherosclerosis occurrence and development as well as plaque formation (39–41). The pathophysiological interaction between hypertension and dyslipidemia, which includes oxidative stress,

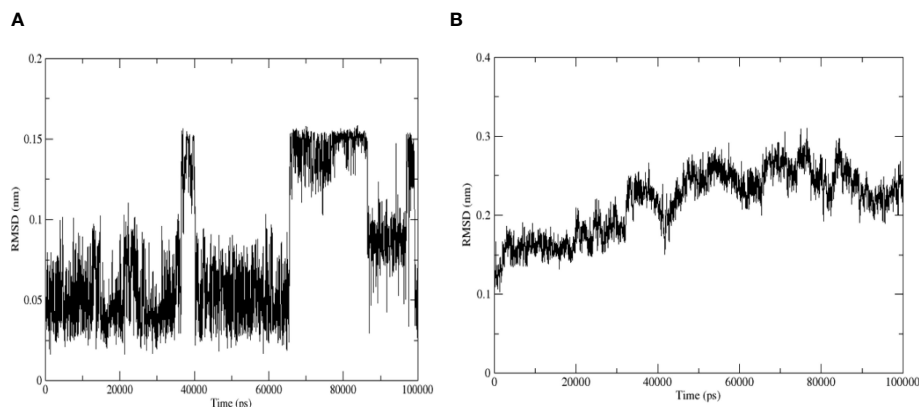


FIGURE 10
RMSD of quercetin and *TNF* protein 5uui. (A) Quercetin; (B) *TNF* protein 5uui.

proinflammatory activities, renin-angiotensin-aldosterone system activation, and endothelial dysfunction, has been supported by accumulating evidence (42). A considerable positive effect of lipid-lowering therapy has been observed in older patients with obesity and hypertension; the therapy markedly lowers obesity-related indicators including blood pressure, lipid levels, and glucose levels as well as considerably improves atherosclerosis-related symptoms without side effects (43). Several *in-vivo* experiments have shown that the *Ilex kudingcha* extract reduces atherosclerosis in apoE-deficient mice by decreasing cholesterol buildup in macrophages (44). Considering their effects on hemorheological properties, the total saponins in *Ilex kudingcha* may possess considerable therapeutic value for treating hypercholesterolemia and atherosclerosis (45). *Ilex kudingcha* exerts potent anti-inflammatory effects on LPS-induced inflammatory responses by inhibiting *NF-κB* and *MAPK* pathways (10). *Dioscorea opposita* Thunb., a common staple food in China, exerts antihypertensive effects by

inhibiting the endothelin-converting enzyme and antioxidant activity in 2K1C hypertensive rats (46). Furthermore, the binding activity of the protein and its ligand was explained by molecular docking in the present study. The results showed that quercetin and kaempferol bound well to the corresponding target proteins. Small molecules interacted with proteins mainly by forming hydrophobic and hydrogen bonds. The lowest binding energy of *TNF* with quercetin and kaempferol was -8.8 kcal/mol, followed by -8.4 kcal/mol of *MARKI* with quercetin. The docking result was less than -7.0 kcal/mol, indicating that the ligand and receptor exhibited strong binding activity (47). The MD simulation analysis results confirmed the stable binding of *TNF* and quercetin. MD simulation has been widely used in the biomedical field to study conformational transformations caused by protein mutations or ligand binding/debinding. It provides some findings that are difficult to obtain in traditional biochemical or pathological experiments, such as the detailed effects of mutations on protein structures and protein-protein/ligand

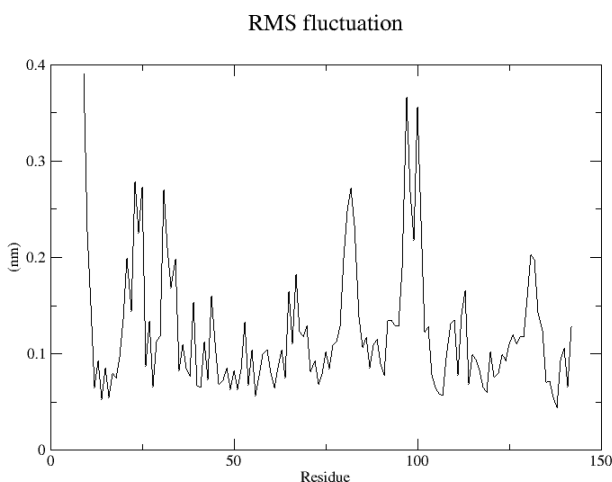


FIGURE 11
RMSF analysis of *TNF* protein.

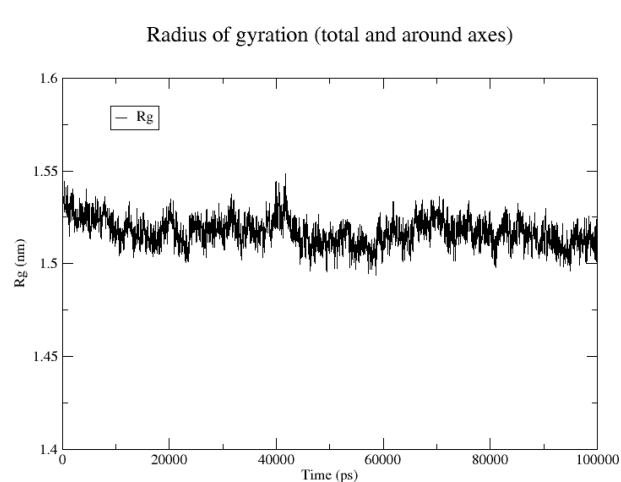
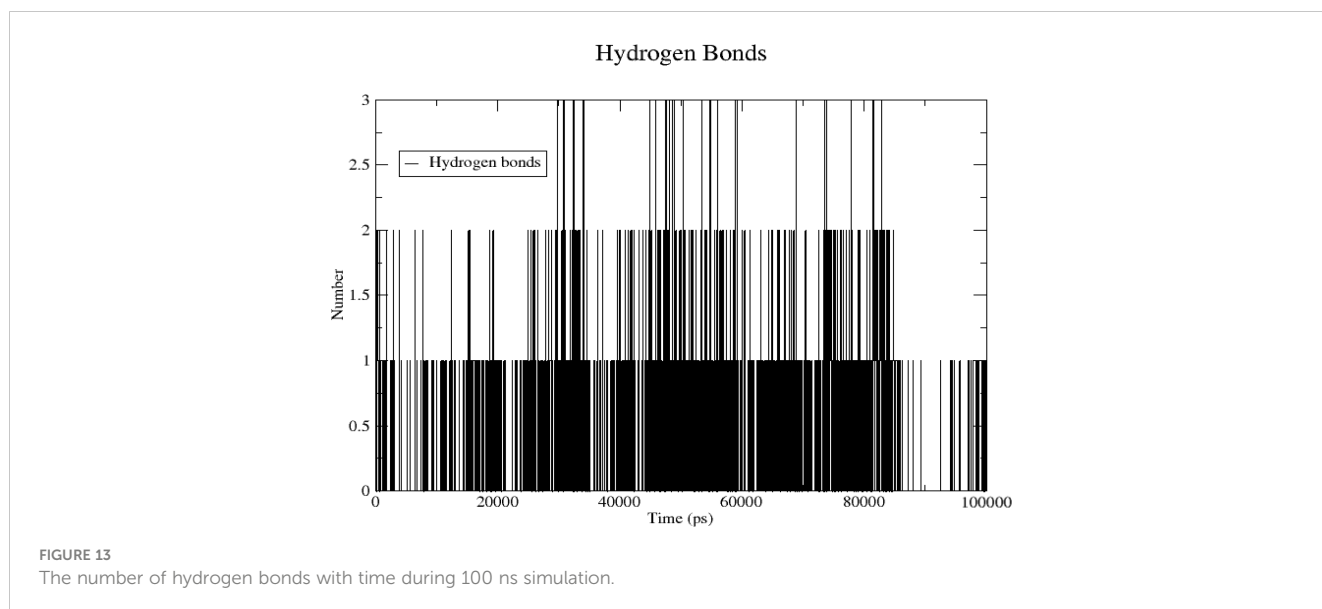


FIGURE 12
Rg analysis of *TNF* protein.



interactions at the atomic level (48). MD simulation can provide valuable information in deciphering the functional mechanisms of several biomolecules including proteins/peptides, overcoming existing sampling limitations in docking analysis (49).

5 Conclusion

Ilex kudingcha can play a role in lowering blood pressure via multiple components, targets, and pathways. This study provides insights into how the hypolipidemia-, antioxidation-, and atherosclerosis-related gene inhibitory effects of *Ilex kudingcha* are associated with its mode of action in treating hypertension. The action targets include *RELA*, *AKT1*, *TNF*, *IL-6*, and *MARK1*. Studies that are pertinent to these findings rely on data from already-existing databases and lack experimental validation. Therefore, further studies are required to verify the accuracy of the present findings *in vitro* and *in vivo*.

Data availability statement

The original contributions presented in the study are included in the article/supplementary material. Further inquiries can be directed to the corresponding authors.

Author contributions

FL wrote and revised the manuscript and constructed the tables and figures. MY and RH revised the manuscript. YQ and YH conceived the study and revised the manuscript. All authors have read and approved the final manuscript. Data authentication is not applicable.

Funding

This research was supported by the Qingzhen City Science and Technology Project Support Program (Grant No. 2023-25), Ecological Poultry Science and Technology Innovation Team of Higher Education Institutions in Guizhou Province (Grant No. 2023098), and Sichuan Province Science and Technology Support Program (Grant No. 2020YFS0337).

Acknowledgments

The authors would like to thank all the reviewers who participated in the review and MJEditor (www.mjeditor.com) for its linguistic assistance during the preparation of this manuscript.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

- Lamirault G, Artifoni M, Daniel M, Barber-Chamoux N, Nantes University Hospital Working Group On Hypertension. Resistant hypertension: novel insights. *Curr Hypertens Rev* (2020) 16(1):61–72. doi: 10.2174/157340211566619101111402
- Lu Y, Feng F, Lu JP, Li XY, Wang XC, Sun DQ, et al. Severe hypertension in China: results from the China PEACE million persons project. *J Hypertension* (2021) 39(3):461–70. doi: 10.1097/HJH.0000000000002675
- Chow CK, Teo KK, Rangarajan S, Islam S, Gupta R, Avezum A, et al. Prevalence, awareness, treatment, and control of hypertension in rural and urban communities in high-, middle-, and low-income countries. *Jama* (2013) 310(9):959–68. doi: 10.1001/jama.2013.184182
- Harrison MA, Marfo AFA, Opare-Addo MNA, Ankrah DNA, Acheampong F, Nelson F, et al. Anti-hypertensive medication access and affordability and their association with blood pressure control at a teaching hospital in Ghana. *Pan Afr Med J* (2021) 39:184. doi: 10.11604/pamj.2021.39.184.27977
- Attai MW, Khatib R, McKee M, Lear S, Dagenais G, Igumbor EU, et al. Availability and affordability of blood pressure-lowering medicines and the effect on blood pressure control in high-income, middle-income, and low-income countries: an analysis of the PURE study data. *Lancet Public Health* (2017) 2(9):e411–9. doi: 10.1016/S2468-2667(17)30141-X
- Carey RM, Calhoun DA, Bakris GL, Brook RD, Daugherty SL, Dennison-Himmelfarb CR, et al. Resistant hypertension: detection, evaluation, and management: A scientific statement from the American heart association. *Hypertension* (2018) 72(5):e53–90. doi: 10.1161/HYP.0000000000000884
- Dai S, Xiao X, Xu C, Jiao Y, Qin Z, Meng J, et al. Association of Dietary Approaches to Stop Hypertension diet and Mediterranean diet with blood pressure in less-developed ethnic minority regions. *Public Health Nutr* (2022) 25(12):1–29. doi: 10.1017/S1368890022000106
- Milner JA. Functional foods and health promotion 1. *J Nutr* (1999) 129(7):1395S–7S. doi: 10.1093/jn/129.7.1395S
- Wan P, Peng YJ, Chen GJ, Xie MH, Dai ZQ, Huang KY, et al. Dicafeoylquinic acids from *Ilex kudingcha* attenuate dextran sulfate sodium-induced colitis in C57BL/6 mice in association with the modulation of gut microbiota. *J Funct Foods* (2019) 61:103468. doi: 10.1016/j.jff.2019.103468
- Wan P, Xie MH, Chen GJ, Dai ZQ, Hu B, Zeng X, et al. Anti-inflammatory effects of dicafeoylquinic acids from *Ilex kudingcha* on lipopolysaccharide-treated RAW264.7 macrophages and potential mechanisms. *Food Chem Toxicol* (2019) 126:332–42. doi: 10.1016/j.fct.2019.01.011
- Sun Y, Hu Z, Zhang DL, Ye H. Extraction and antioxidant activities of polysaccharides from *Ilex kudingcha* made from *Ilex kudingcha* CJ tseng. *Glycobiology* (2009) 19(11):1307–8.
- Mu JF, Yang FP, Tan F, Zhou XR, Pan YN, Long XY, et al. Determination of polyphenols in *Ilex kudingcha* and insect tea (Leaves altered by animals) by ultra-high-performance liquid chromatography-triple quadrupole mass spectrometry (UHPLC-QQ-MS) and comparison of their anti-aging effects. *Front Pharmacol* (2021) 11. doi: 10.3389/fphar.2020.600219
- Zhu SL, Wei LY, Lin GX, Tong YL, Zhang JW, Jiang X, et al. Metabolic alterations induced by *Ilex kudingcha* lead to cancer cell apoptosis and metastasis inhibition. *Nutr Cancer-an Int J* (2020) 72(4):696–707. doi: 10.1080/01635581.2019.1645865
- Zhao X, Wang Q, Qian Y, Song JL. *Ilex kudingcha* CJ Tseng (*Ilex kudingcha*) has in vitro anticancer activities in MCF-7 human breast adenocarcinoma cells and exerts anti-metastatic effects in vivo. *Oncol Lett* (2013) 5(5):1744–8. doi: 10.3892/ol.2013.1253
- Che YY, Wang QH, Xiao RY, Zhang JY, Zhang YQ, Gu W, et al. Kudinoside-D, a triterpenoid saponin derived from *Ilex kudingcha* suppresses adipogenesis through modulation of the AMPK pathway in 3T3-L1 adipocytes. *Fitoterapia* (2018) 125:208–16. doi: 10.1016/j.fitote.2017.11.018
- Wu HL, Chen YL, Yu YY, Zang J, Wu YK, He Z. *Ilex latifolia* Thunb protects mice from HFD-induced body weight gain. *Sci Rep* (2017) 7(1):14660. doi: 10.1038/s41598-017-15292-x
- Song C, Xie C, Zhou Z, Yu S, Fang N. Antidiabetic effect of an active components group from *Ilex kudingcha* and its chemical composition. *Evidence-Based Complementary Altern Med* (2012) 2012:423690. doi: 10.1155/2012/423690
- Wu JS, Zhang FQ, Li ZZ, Jin WY, Shi Y. Integration strategy of network pharmacology in Traditional Chinese Medicine: a narrative review. *J Tradit Chin Med* (2022) 42(3):479–86. doi: 10.19852/j.cnki.jtcm.20220408.003
- Zhang R, Zhu X, Bai H, Ning K. Network pharmacology databases for traditional chinese medicine: review and assessment. *Front Pharmacol* (2019) 10(123). doi: 10.3389/fphar.2019.00123
- Li S, Zhang B. Traditional Chinese medicine network pharmacology: theory, methodology and application. *Chin J Nat Med* (2013) 11(2):110–20. doi: 10.1016/S1875-5364(13)60037-0
- Pan Z, Li M, Jin Z, Sun D, Zhang D, Hu B, et al. Research status of Chinese medicine formula based on network pharmacology. *Pharmacol Res - Modern Chin Med* (2022) 4:100132. doi: 10.1016/j.prmcm.2022.100132
- Zhao L, Zhang H, Li N, Chen J, Xu H, Wang Y, et al. Network pharmacology, a promising approach to reveal the pharmacology mechanism of Chinese medicine formula. *J Ethnopharmacol* (2023) 309:116306. doi: 10.1016/j.jep.2023.116306
- Cao Y, Xie L, Liu K, Liang Y, Dai X, Wang X, et al. The antihypertensive potential of flavonoids from Chinese Herbal Medicine: A review. *Pharmacol Res* (2021) 174:105919. doi: 10.1016/j.phrs.2021.105919
- Ye XT, Zhang BX, Wang HH, He SB, Zhang XH, Wang Y. Study on mechanism for anti-hypertension efficacy of *Eucommiae Cortex* through assistant analysis systems for acting mechanisms of traditional Chinese medicine. *Zhongguo Zhong Yao Za Zhi* (2015) 40(19):3718–22. doi: 10.4268/cjcm20151905
- Yang CR, Jiao HC, Li YL, Xue YT. Research progress on Traditional Chinese Medicine treatment of hypertension based on network pharmacology. *Chin Arch Traditional Med* (2021) 39(02):72–7. doi: 10.13193/j.issn.1673-7717.2021.02.020
- McCullough ML, Peterson JJ, Patel R, Jacques PF, Shah R, Dwyer JT, et al. Flavonoid intake and cardiovascular disease mortality in a prospective cohort of US adults. *Am J Clin Nutr* (2012) 95(2):454–64. doi: 10.3945/ajcn.111.016634
- Dabeek WM, Marra MV. Dietary quercetin and kaempferol: bioavailability and potential cardiovascular-related bioactivity in humans. *Nutrients* (2019) 11(10):2288. doi: 10.3390/nu11102288
- Imran M, Rauf A, Shah ZA, Saeed F, Imran A, Arshad MU, et al. Chemo-preventive and therapeutic effect of the dietary flavonoid kaempferol: A comprehensive review. *Phytother Res* (2019) 33(2):263–75. doi: 10.1002/ptr.6227
- Yusof HM, Sarah NML, Lam TW, Kassim MNI. Hypolipidemic effects of quercetin and kaempferol in human hepatocellular carcinoma (HepG2) cells. *Int Food Res J* (2018) 25(1):241–5.
- Cid-Soto MA, Martínez-Hernández A, García-Ortiz H, Córdova EJ, Barajas-Olmos F, Centeno-Cruz F, et al. Gene variants in AKT1, GSK3 and SOCS3 are differentially associated with metabolic traits in Mexican AmerIndians and Mestizos. *Gene* (2018) 679:160–71. doi: 10.1016/j.gene.2018.08.076
- Sankhe S, Manousakidi S, Antigny F, Arthur Ataam J, Bentebbal S, Ruchon Y, et al. T-type Ca(2+) channels elicit pro-proliferative and anti-apoptotic responses through impaired PP2A/Akt1 signaling in PSMCs from patients with pulmonary arterial hypertension. *Biochim Biophys Acta Mol Cell Res* (2017) 1864(10):1631–41. doi: 10.1016/j.bbamer.2017.06.018
- DeMarco VG, Aroor AR, Sowers JR. The pathophysiology of hypertension in patients with obesity. *Nat Rev Endocrinol* (2014) 10(6):364–76. doi: 10.1038/nrendo.2014.44
- Reiss AB, Siegart NM, De LJ. Interleukin-6 in atherosclerosis: atherogenic or atheroprotective? *Clin Lipidology* (2017) 12(1):14–23. doi: 10.1080/17584299.2017.1319787
- Naz S, Raza M, Akbar A, Zaidi AN, Khaliq S, Ikram T, et al. Serum interleukin-6 and lipid atherogenic index risk ratios as interpreters of cardiac disease in obese and non obese male patients of coronary artery disease. *Pakistan J Med Health Sci* (2021) 15(6):1203–5. doi: 10.53350/pjmhs211561203
- Zhu TT, Zhang WF, Yin YL, Liu YH, Song P, Xu J, et al. MicroRNA-140-5p targeting tumor necrosis factor- α prevents pulmonary arterial hypertension. *J Cell Physiol* (2019) 234(6):9535–50. doi: 10.1002/jcp.27642
- Fan Z, Li C, Qin C, Xie L, Wang X, Gao Z, et al. Role of the PI3K/AKT pathway in modulating cytoskeleton rearrangements and phenotype switching in rat pulmonary arterial vascular smooth muscle cells. *DNA Cell Biol* (2014) 33(1):12–9. doi: 10.1089/dna.2013.2022
- Geng Z, Ye C, Tong Y, Zhang F, Zhou YB, Xiong XQ. Exacerbated pressor and sympathoexcitatory effects of central Elabela in spontaneously hypertensive rats. *Am J Physiol Heart Circ Physiol* (2020) 318(1):H124–34. doi: 10.1152/ajpheart.00449.2019
- Xie HK, Zhuang Q, Mu JL, Sun JX, Wei PF, Zhao XH, et al. The relationship between lipid risk score and new-onset hypertension in a prospective cohort study. *Front Endocrinol* (2022) 13:916951. doi: 10.3389/fendo.2022.916951
- Guzel S, SC FB, Guzel EC, Kucukyalcin V, Kiziler AR, Cavusoglu C, et al. Midkine levels and its relationship with atherosclerotic risk factors in essential hypertensive patients. *Niger J Clin Pract* (2018) 21(7):894–900. doi: 10.4103/njcp.njcp_309_17
- Li M, Liu L, Song S, Shi A, Ma Y, Zhang S, et al. Effect of long-term lifestyle intervention on mild cognitive impairment in hypertensive occupational population in China. *Medicine* (2018) 97(34):e11975. doi: 10.1097/MD.00000000000011975
- Adam CA, Șalaru DL, Prisacariu C, Marcu DTM, Sascău RA, Stătescu C. Novel biomarkers of atherosclerotic vascular disease—Latest insights in the research field. *Int J Mol Sci* (2022) 23(9):4998.
- Ke C, Zhu X, Zhang Y, Shen Y. Metabolomic characterization of hypertension and dyslipidemia. *Metabolomics* (2018) 14(9):117. doi: 10.1007/s11306-018-1408-y
- Chao LX, Zhao YX, Lu FH, Wang GJ, Zhang H, Sun SW, et al. Effects of lipid-lowering therapy on blood pressure and arteriosclerosis in elderly patients with obese hypertension. *J Hypertension* (2018) 36:e144–5. doi: 10.1097/01.hjh.0000548584.81174.49
- Fujiwara Y, Okada S, Uryu K, Maru I, Komohara Y. The extract of *Ilex kudingcha* inhibits atherosclerosis in apoE-deficient mice by suppressing cholesterol accumulation in macrophages. *Bioscience Biotechnol Biochem* (2021) 85(10):2177–84. doi: 10.1093/bsbb/zbab140
- Zheng J, Wang X, Li H, Gu Y, Tu P, Wen Z. Improving abnormal hemorheological parameters in ApoE^{-/-} mice by *Ilex kudingcha* total saponins. *Clin Hemorheol Microcirculation* (2009) 42(1):29–36. doi: 10.3233/CH-2009-1183

46. Amat N, Amat R, Abdureyim S, Hoxur P, Osman Z, Mamut D, et al. Aqueous extract of *dioscorea opposita thunb.* normalizes the hypertension in 2K1C hypertensive rats. *BMC Complementary Altern Med* (2014) 14:36. doi: 10.1186/1472-6882-14-36
47. Bubiājiaer H, Li Y, Qin YD, Yao J, Cheng YF, Shen J. Prediction of antihypertensive active components and mechanism of *Caragana Sinca* based on network pharmacology and molecular docking. *J Xinjiang Med Univ* (2022) 45 (4):435–43. doi: 10.3639/j.issn.1009-5551.2022.04.015
48. Wu X, Xu LY, Li EM, Dong G. Application of molecular dynamics simulation in biomedicine. *Chem Biol Drug Des* (2022) 99(5):789–800. doi: 10.1111/cbdd.14038
49. Vidal-Limon A, Aguilar-Toalá JE, Liceaga AM. Integration of molecular docking analysis and molecular dynamics simulations for studying food proteins and bioactive peptides. *J Agric Food Chem* (2022) 70(4):934–43. doi: 10.1021/acs.jafc.1c06110