Check for updates

OPEN ACCESS

EDITED BY Undurti Narasimha Das, UND Life Sciences LLC, United States

REVIEWED BY Yogesh A. Kulkarni, SVKM's Narsee Monjee Institute of Management Studies, India Ankit P. Laddha, University of Connecticut, United States Adriana Fodor, University of Medicine and Pharmacy Iuliu Hatieqanu, Romania

*CORRESPONDENCE Subrata Chakrabarti Subrata.Chakrabarti@lhsc.on.ca

[†]These authors have contributed equally to this work and share first authorship

SPECIALTY SECTION

This article was submitted to Diabetes: Molecular Mechanisms, a section of the journal Frontiers in Endocrinology

RECEIVED 21 September 2022 ACCEPTED 17 January 2023 PUBLISHED 27 January 2023

CITATION

Wang E, Wang H and Chakrabarti S (2023) Endothelial-to-mesenchymal transition: An underappreciated mediator of diabetic complications. *Front. Endocrinol.* 14:1050540. doi: 10.3389/fendo.2023.1050540

COPYRIGHT

© 2023 Wang, Wang and Chakrabarti. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Endothelial-tomesenchymal transition: An underappreciated mediator of diabetic complications

Eric Wang[†], Honglin Wang[†] and Subrata Chakrabarti^{*}

Department of Pathology and Laboratory Medicine, Schulich School of Medicine and Dentistry, Western University, London, ON, Canada

Diabetes and its complications represent a great burden on the global healthcare system. Diabetic complications are fundamentally diseases of the vasculature, with endothelial cells being the centerpiece of early hyperglycemia-induced changes. Endothelial-to-mesenchymal transition is a tightly regulated process that results in endothelial cells losing endothelial characteristics and developing mesenchymal traits. Although endothelial-to-mesenchymal transition has been found to occur within most of the major complications of diabetes, it has not been a major focus of study or a common target in the treatment or prevention of diabetic complications. In this review we summarize the importance of endothelial-to-mesenchymal transition in each major diabetic complication, examine specific mechanisms at play, and highlight potential mechanisms to prevent endothelial-to-mesenchymal transition in each of the major chronic complications of diabetes.

KEYWORDS

diabetes, diabetic complications, endothelial-to-mesenchymal transition, epigenetic regulation, vascular dysfunction

1 Introduction

Diabetes has long since reached epidemic levels around the globe and shows no signs of slowing down any time soon (1, 2). The current prevalence of diabetes has far surpassed projections made in the beginning of the century, and the number of people affected by diabetes is predicted to increase by nearly 50% by 2045 (3–5). Many people living with diabetes are affected by chronic diabetic complications, and with the increase in prevalence of diabetes, the burden of diabetic complications is sure to increase. Diabetes and its complications are significant causes of morbidity and mortality. Not only was diabetes directly responsible for 1.5 million deaths in 2019—making it the 9th leading cause of death—diabetes is also a major risk factor for other leading causes of death such as heart disease, stroke, and kidney failure (1, 4).

The pathology of diabetic complications begins with vascular endothelial cells and eventually snowball into dysfunction of organs (6-8). Besides glycemic control, current

approaches to management and treatment of these complications are focused on late-stage occurrences, and do not address the root problems (9–11). In an attempt to shift the focus of conversations surrounding diabetic complications away from later stage outcomes and toward the early events and root causes, we examine the importance of endothelial-to-mesenchymal transition (EndMT), an early occurrence in diabetic complications that may represent a common target for prevention or management of different complications of diabetes.

1.1 Diabetic complications

Diabetic complications are broadly separated into two categories, micro- and macrovascular (12–14). As their names imply, microvascular complications affect capillaries and small vessels, while macrovascular complications affect larger vessels. Major microvascular complications of diabetes include diabetic retinopathy, diabetic nephropathy, diabetic neuropathy, and diabetic cardiomyopathy (12–14). Both micro- and macrovascular diabetic complications are primarily the result of hyperglycemia. Excessive amounts of circulating glucose is taken up by vascular endothelial cells (ECs), causing metabolic derangements within the ECs and resulting in damage (12–14).

ECs are among the first to be damaged by hyperglycemia during diabetes. Because ECs express the insulin-independent glucose transporter, GLUT1, their rate of glucose uptake is proportional to the amount of glucose in circulation (12–15). Excessive uptake of glucose overloads the glycolytic pathway, leading to the shunting of glycolytic intermediates into other, more harmful routes (12, 13). Furthermore, elevated rates of oxidative respiration downstream of increased glycolysis causes enhanced production of reactive oxygen species (ROS) in the mitochondria, leading to oxidative damage (12, 13). Oxidative damage triggers the activation of anti-ROS responses, one side-effect of which is the inhibition a glycolytic enzyme—GAPDH—which further compounds the shunting of glycolytic intermediates into harmful pathways (12, 13).

Harmful pathways activated by hyperglycemia include the polyol, hexosamine, protein kinase C (PKC) and advanced glycation endproduct (AGE) pathways (12, 13). The polyol pathway reduces glucose into sorbitol at the expenditure of NADPH, an important cofactor in regenerating the antioxidant glutathione (12, 13, 16). Sorbitol can be oxidized into fructose, which can be fed back to the glycolytic pathway, but the depletion of NADPH increases susceptibility to damage by mitochondrial ROS (12, 13, 16). The hexosamine pathway shunts fructose-6-phosphate to generate uridine diphosphate N-acetyl glucosamine, which can covalently attach to transcription factors and disrupt gene expression (12, 13, 17). The PKC pathway involves PKC activation by diacylglycerol, converted from high levels of dihydroxyacetone phosphate, leading to widescale changes in gene expression (12, 13, 18). PKC activation in diabetes promotes angiogenic, fibrotic, and proinflammatory changes (12, 13, 18). Finally, the AGE pathway involves the non-enzymatic glycation of proteins by a variety of glucose-derived molecules, resulting in dysfunction of the glycated protein, and promoting inflammation by activating receptors of AGEs (RAGEs) (12, 13, 19).

The shunting of glucose into damaging pathways is a well-established part of hyperglycemia-induced endothelial dysfunction and occurs in large and small vessels alike. These pathways are activated early on during the pathogenesis of diabetic complications, but clinical manifestations of diabetic complications do not appear until much later. Thus, these occurrences alone do not paint the whole picture of early endothelial dysfunction and diabetic vascular complications.

1.2 Endothelial-to-mesenchymal transition

In response to specific stimuli, ECs can undergo a dramatic transformation known as EndMT. EndMT is a process whereby ECs, in response to specific internal and environmental triggers, transdifferentiate into mesenchymal like cells, losing their original endothelial characteristics and adopting mesenchymal phenotype (20-24). EndMT can be a physiological process and is a crucial occurrence in the embryonic development of heart valves (20-23). However, when aberrantly induced by environmental stressors such as hyperglycemia, EndMT gives rise to issues that contribute to dysfunction (21-24). The loss of endothelial junctional markers such as vascular endothelial cadherin (VE-CAD) and platelet endothelial cell adhesion molecule-1 (PECAM1) can lead to increased vascular permeability, resulting in increased infiltration of immune cells and unwanted exchange of fluids or factors between blood and the impacted tissue (25-27). While the gain of mesenchymal phenotype leads to increased production and deposition of extracellular matrix (ECM) proteins, contributing to sclerosis and fibrosis during later stages of disease (28-30).

EndMT, whether physiological or pathological, is a regulated process mediated by specific extracellular signals and intracellular changes. Transforming growth factor β (TGF- β) family of growth factors are the most common and most well-studied drivers of EndMT, thus the induction of EndMT can be broadly split into TGF- β and non-TGF- β pathways (22, 23, 31–33). The TGF- β pathway of EndMT itself can be subdivided into canonical and non-canonical pathways. The canonical pathway is mediated by Smad2/3, while the non-canonical pathway is Smad2/3-independent and can act through a variety of other signal transducers such as mitogen-activated protein kinase (MAPK), phosphatidylinositol 3-kinase (PI3K), and PKC-δ (22, 23, 31-35). Non-TGF-β pathways of EndMT include Notch, Wnt, endothelin-1 (ET-1), and inflammatory signaling (22, 23, 31-33). Though the exact mechanisms of these pathways are varied and have not been fully elucidated, most pathways ultimately converge through the regulation of transcription factors such as SNAI1 and TWIST to suppress the expression of EC markers and promote the expression of mesenchymal proteins (22, 23, 31-39) (Figure 1). All of these pathways have been implicated in diabetes-induced EndMT in one or more diabetic complication, though there has not been any studies simultaneously examining specific pathways across different diabetic complications (12, 24, 40-46). Once an EC has undergone EndMT, the EC-derived mesenchymal cell remains committed to the mesenchymal phenotype, even if the driving stimulus is no longer present.



FIGURE 1

Endothelial to mesenchymal transition. ECs can undergo EndMT in response to a variety of stimuli. EndMT is characterized by reduced expression of endothelial markers (green) and increased expression of mesenchymal markers (red). Common inducers of EndMT include NOTCH, WNT, TGF- β , ET-1, and inflammatory signaling. TGF- β is the best characterized and most common driver of EndMT. Canonical TGF- β signaling through SMAD2/3, non-canonical TGF- β signaling through other mechanisms such as MAPK, PI3K and PKC- δ , as well as other EndMT inducers such as NOTCH, WNT, and ET-1 generally converge through the regulation of key transcription factors such as SNAI and Twist in order to reduce endothelial protein expression and protein expression. Inflammatory cytokines contribute to EndMT by activating NF- κ B. Cells undergoing EndMT co-express endothelial and mesenchymal markers.

1.3 Epigenetic regulation and EndMT

The persistence of the mesenchymal phenotype in EC-derived mesenchymal cells is dependent on changes in epigenetic regulation. Epigenetic regulation describes heritable phenotypical changes absent changes in the genomic sequence. Epigenetic regulation encompasses several processes, including DNA methylation, histone modification, and non-coding RNA (ncRNA)-associated gene regulation (47-49). DNA methylation involves the covalent attachment of methyl groups to nucleotides, most commonly to the 5th carbon of the cytosine residue in a CpG pair, and generally results in transcriptional inhibition (47-50). Histone modifications involve covalent changes to the histone proteins that bind the DNA, leading to either activation or repression of the associated DNA regions (47-49, 51). Common histone modifications include methylation, acetylation, phosphorylation, SUMOylation, and citrullination; the specific effects of histone modifications typically depend on the type of modification, the amino acid residue that is modified, and the location of the modification (47-49, 51). ncRNA-associated gene regulation involves RNA molecules that are transcribed from DNA but not translated into proteins; ncRNAs can potentiate or disrupt gene expression both at and beyond the transcriptional level (47-49, 52). Long ncRNAs (lncRNAs, greater than 200 nucleotides in length) can form complex tertiary structures and interact with RNA-binding proteins, serving as scaffolds, guides, or decoys for protein complexes, and leading to activation or repression of target loci (53-55). microRNAs (miR, short ncRNAs roughly 22 nucleotides in length) regulate translational silencing of specific mRNAs through complementarity (56–58). Circular RNAs (circRNAs) are ncRNAs derived from mRNA transcripts but circularized *via* back-splicing. circRNAs can modulate the expression of their parental genes by a decoy for translational machinery or inhibitory miRNAs, they can also interact with other proteins in order to produce specific outcomes (59–63).

Epigenetic changes are involved at a variety of levels during EndMT, from modulating pro-EndMT signaling to enforcing longterm changes in endothelial and mesenchymal marker expression. Removal of DNA methylation has been shown to increase the expression of SNAI1 in epithelial cells (64). SNAI1 has been shown in a various cells to recruit histone deacetylases (HDACs) to target genes to suppress their expression (65, 66). HDAC9 has been found to mediate changes in endothelial and mesenchymal markers in EndMT, and targeted suppression of HDAC9 prevented EndMT in human coronary artery ECs (67). HDAC3α has been shown to promote TGF- β 2 secretion, which acts as an autocrine inducer of EndMT (68). ncRNAs H19 and miR-200b have been found to modulate TGF- β signaling in the induction of EndMT (40-42). Beyond individual effects of each of the epigenetic mechanisms, epigenetic regulators also have reciprocal regulatory effects on one another. Covalent modifications to the DNA or the histones can influence the transcription of ncRNAs (56, 69-73). lncRNAs can inhibit miRNA activity by acting as a molecular sponge (53-55), or influence DNA and histone modification by guiding or obstructing the protein complexes (53-55, 74). miRNAs can inhibit the translation of proteins involved in DNA and histone modifications (75–77); they can also inhibit lncRNAs by inducing degradation (78–80). circRNAs can regulate DNA/histone modifying machinery and sponge miRNAs (59–62, 81). This creates an intricate network of regulation that underlies the process of EndMT, the precise balance of which determines the endothelial or mesenchymal nature of the affected cells (Figure 2).

2 EndMT in diabetic complications

Hyperglycemia is a known driver of EndMT in diabetes. High glucose-mediated endothelial damage activates a variety of EndMTinducing pathways. High glucose promotes TGF- β signaling in ECs through the activation of the PKC pathway, which can trigger EndMT through both Smad-dependent and -independent pathways (12, 24, 40– 42). The PKC pathway also triggers ET-1 production, which promotes EndMT independently from TGF- β (12, 43, 82). Furthermore, inflammatory responses triggered by ROS, AGE-RAGE and other hyperglycemia-induced pathways also contribute to EndMT in diabetes (12, 22, 39). Additionally, hyperglycemia effects changes in epigenetic regulation, and can increase the susceptibility of ECs to EndMT. Specific mechanistic links between hyperglycemia and EndMT have been reported in different diabetic complications organs (44, 83– 87). Hyperglycemia and EndMT are closely intertwined in the early pathogenesis diabetic vascular complications, thus highlighting the importance of EndMT may provide new perspective and insight into the understanding of these complications.

2.1 Diabetic retinopathy and endothelialtomesenchymal transition

Diabetic retinopathy is the leading cause of blindness in workingaged adults. Diabetic retinopathy is broadly divided into two stages, non-proliferative (NPDR) and proliferative (PDR) (9, 88-90). NPDR begins with asymptomatic glucose-induced changes in the retinal vasculature which compound and worsen, leading to increased vascular permeability, capillary non-perfusion, microaneurysms, microhemorrhages, neuronal damage, and intraretinal microvascular abnormalities (9, 88-90). The non-proliferative changes disrupt the perfusion of the retina, causing overexpression of angiogenic factors, leading to the characteristic growth of abnormal vessels, i.e., neovascularization in PDR (9, 88-90). Vision loss in diabetic retinopathy typically involves one of the following mechanisms: diabetic macular edema (DME), or hemorrhage and tractional retinal detachment (TRD) (9, 88-90). DME can occur during NPDR or PDR and is caused by fluid buildup as a result of leaky blood vessels (9, 88-90). TRD more limited to PDR and is caused by fibrovascular scarring of the abnormal vessels generated during neovascularization (9, 88–90). EndMT occurs throughout NPDR and PDR, and may contribute to either mechanism of vision loss in diabetic retinopathy.



Epigenetic regulation of endothelial to mesenchymal transition. Induction and maintenance of EndMT is facilitated by changes in epigenetic regulation. Stimuli such as TGF-β, NOTCH, WNT, ET-1, inflammatory cytokines, which may result from hyperglycemia can effect changes in epigenetic regulation, resulting in persistently altered patterns of gene expression. Different epigenetic mechanisms also reciprocally regulate one another. DNA methylation, and histone modifications can influence the transcription of ncRNAs, while ncRNAs can regulate the proteins required for covalent modifications of DNA and histones. Different types of ncRNAs regulate also each other, with circRNAs and lncRNAs sponging miRNAs while miRNAs can target circRNAs and lncRNAs for degradation.

ECs in the retina play an important role in maintaining the bloodretinal barrier (BRB), a highly selective barrier that keeps the retinal environment separate from general circulation (91, 92). Losing endothelial characteristics through EndMT disrupts the barrier function of these retinal ECs and results in increased permeability. Furthermore, increased deposition of matrix proteins such as type I collagen, type IV collagen and fibronectin due to the gain of mesenchymal traits may lead to thickening of the basement membrane, which also contributes to increased vascular permeability in diabetic retinopathy (40, 93). Increased vascular permeability not only contributes to risk of DME, but also allows circulating factors such as inflammatory mediators to enter the retina, leading to inflammation and further damage (91, 92). Breakdown of the BRB is a well-documented part of diabetic retinopathy and has many contributors, but EndMT does appear to play a substantial role. Inducing EndMT in non-diabetic mice has been shown to result in similar retinal vascular leakage levels to diabetic mice (42). Apart from disrupting the barrier function of the BRB, EndMT also contributes to other harmful pathways within diabetic retinopathy. EC-derived mesenchymal-like cells contribute to the population of myofibroblasts that participate in fibrosis and mediate TRD during PDR (94).

EndMT in diabetic retinopathy has been verified to be mediated by a variety of pathways. High glucose induced the suppression of two IncRNAs, MEG3 and H19, which reportedly mediate EndMT in the retina through Smad-independent TGF- β signaling (42, 83). MEG3 inhibition was mediated by glucose-induced promoter hypermethylation, while the mechanism of H19 has not been reported (42, 83). High glucoseinduced suppression of miR-200b, facilitated EndMT in diabetic retinopathy through canonical TGF- β signaling (40). Experimentally induced overexpression of H19, MEG3 or miR-200b prevented EndMT by inhibiting MAPK, PI3K/AKT/mTOR, and Smad2 respectively, effectively cutting off TGF- β signal transduction (40, 42, 83). Other EndMT-inducing pathways are also at play in the diabetic retina. Notch2 is significantly upregulated in retinal ECs under high glucose conditions, and drove EndMT independently of TGF- β (44). Overexpression of miR-29a/b was able to suppress Notch2 and prevent EndMT (44). Differential expression of a variety of circRNAs has been reported in diabetic retinopathy and non-diabetic retinopathy animals, and circRNAs associated with EndMT-related processes were uniquely increased in animals with diabetic retinopathy (95).

2.2 Diabetic nephropathy and endothelialto-mesenchymal transition

Diabetic neuropathy is the leading cause of end-stage kidney disease and kidney failure and has a strong association with cardiovascular morbidity and mortality (10, 96). Diabetic neuropathy is characterized by failure of the renal filtration system and progressive proteinuria (10, 96). The severity of diabetic neuropathy is correlated with the concentration of albumin in the urine (10, 96). Glomerular basement membrane (GBM) thickening is one of the earliest signs of diabetic neuropathy; despite becoming thicker, the GBM becomes

disorganized and non-uniform, leading to passage of proteins from circulation into the filtrate (97). The passage of plasma proteins into the renal tubules, along with other high glucose-mediated events, such as mesangial expansion and generation of AGEs triggering inflammation and fibrotic changes within the glomeruli and renal tubules, resulting in glomerulosclerosis and tubulointerstitial fibrosis respectively (98–102). Inflammatory and fibrotic changes within the kidney impede kidney function, characterized by reduction of the glomerular filtration rate (GFR) (10, 97). The progressive decline in kidney function and GFR ultimately results in renal failure (10, 96, 97).

ECs of the kidney, somewhat similarly to those of the retina, have a role of maintaining a selective barrier-the glomerular filtration barrier (GFB) (103). EndMT negatively impacts the functional capabilities of the GFB by reducing endothelial junctional protein expression and increasing ECM protein deposition (85, 103, 104). Furthermore, EndMT in the glomerular ECs influences epithelial-tomesenchymal transition (EMT) in adjacent epithelial cells, otherwise known as podocytes, which are also important supporters of the GFB (105). Thus, glomerular EndMT is closely related with podocyte EMT, which contributes to podocyte loss and further breakdown of the GFB (106). Inhibition of EndMT in glomerular ECs has been shown to reduce hyperpermeability, and inhibition of EndMT in diabetic mice has been shown to reduce albuminuria (85, 104). Additionally, EndMT in the kidney, as is the case in other organs, is a significant source of myofibroblasts and a contributor to sclerosis and fibrosis (28, 107, 108). One study has found that up to 30% of myofibroblasts found in the renal interstitium were of endothelial origin (8).

EndMT in diabetic neuropathy, as it is in most cases, has been shown to involve TGF-\beta-mediated responses. Rho-associated kinase (ROCK) is an effector of TGF- β that is upregulated under hyperglycemic conditions and promotes EndMT in diabetic neuropathy (85, 104). Suppression of ROCK via upregulation of miR-497 was able to attenuate EndMT in glomerular ECs (104). Canonical TGF- β signaling is also involved; different researchers have found that inhibition of Smad3 prevented EndMT in diabetic neuropathy (109, 110). One group showed direct blockade of Smad3 using a Smad3 inhibitor prevented AGE-induced EndMT, while another group showed that induction of miR-29 using the drug linagliptin prevented EndMT by inhibiting Smad3 phosphorylation. Non-TGF-B pathways have also been reported to occur in diabetic neuropathy. The pattern recognition receptor NOD2 has been shown to promote EndMT in response to high glucose by bypassing TGF- β receptors and directly activating MAPK (111). The serine protease inhibitor α 2-antiplasmin similarly bypasses TGF- β and induces Smad2/3 activation via AGE-induced responses (112). The SET domain-containing protein 8 (SETD8) regulates EndMT in diabetic neuropathy by directly regulating SNAI1; upregulation of SETD8 suppressed SNAI1, and prevented EndMT (86). And signal transducer and activator of transcription 5A (STAT5A) has been reported to modulate latrophilin and seven transmembrane domain containing 1 (ELTD1) in order to regulate EndMT via a not well-characterized mechanism (113, 114). Lastly, on the chromatin level, methylation of histone 4 lysine 20 (H4K20me1) by lysine methyltransferase 5A has been found to inhibit EndMT, while trimethylation of histone 3 lysine 4 (H3K4me3) by Set1 has been found to induce EndMT (115-117).

2.3 Diabetic neuropathy and endothelial-tomesenchymal transition

Patients with diabetes are 15-30 times more likely to require a lower limb amputation than non-diabetics (118, 119). The associated loss of sensation in diabetic distal somatosensory neuropathy renders patients especially susceptible to foot ulcers and infected wounds which may result in amputations (120, 121). Approximately one half of people with diabetes will develop neuropathy throughout their life (122). Diabetic neuropathy manifests as both numbness and pain, first beginning in the distal extremities. One common occurrence is paradoxical numbness and elevated pain sensitivity (123). This numbness and pain lead to reduced mobility and results in patients being more susceptible to falls, further exacerbating the risks of disabling injury (124). Currently, diabetic neuropathy is only treated through glucose control and pain management, as there is no effective cure for most diabetic complications.

The pathogenesis of diabetic neuropathy was initially understood as having a neurological basis, however, recent understandings of diabetic neuropathy now better characterize its pathogenesis as being primarily vascular in nature (125-128). Vascular dysfunction is the initial and underlying cause of all diabetic complications, and this remains consistent in diabetic neuropathy. The vascular supply to peripheral nerves is limited and blood flow can be easily compromised, any damage to the vasculature surrounding peripheral nerves rendering peripheral nerves vulnerable to ischemia (129). While no research has yet been done on the prevalence of EndMT in diabetic neuropathy, microvessels of the neural vasculature show both thickened basement membranes and disrupted ECs, indicating the possibility for EndMT to have occurred (130). There is still much to be discovered in diabetic neuropathy and elucidating the potential role of EndMT in the peripheral neuronal vasculature are needed to fill the gaps in understanding.

2.4 Diabetic cardiomyopathy and endothelial-to-mesenchymal transition

Cardiovascular disease is the leading cause of death among people with diabetes as diabetes is commonly comorbid with various other cardiovascular diseases such as atherosclerosis, coronary artery disease and hypertension. However, independent of these comorbid risk factors, diabetic cardiomyopathy is defined as abnormal form and function of the heart driven solely by diabetes and high blood glucose, which exists independently of other cardiac risk factors (131-133). Although other mechanistic changes may also play specific roles, microvascular pathology remains the major contributor (131, 134, 135). Patients with type 1 diabetes who do not have hypertension or coronary artery disease suffer higher rates of cardiac dysfunction than non-diabetic cohorts (136). Studies have found that left ventricular dysfunction is associated with diabetes when controlled for coronary artery disease and other heart diseases (137, 138). Additionally, the development of and risk for heart failure is directly correlated with blood glucose levels, with each 1% increase in glycated hemoglobin being linked to increased risk of heart failure in T1DM and T2DM patients (139). The pathogenesis of diabetic cardiomyopathy is not clearly defined as the heart is affected in numerous ways in diabetes, however diabetic cardiomyopathy is generally characterized by left ventricular hypertrophy and a reduction in cardiac contractility and function.

Fibrosis is one characteristic of diabetic cardiomyopathy, it results in imbalanced extracellular matrix protein production, leading to cardiac remodeling and impaired cardiac function, as the adult heart lacks regenerative abilities, cardiac fibrosis is difficult to reverse (140). In cardiac fibrosis as with other fibrotic diseases, abhorrent stimulation of fibroblasts to activate produces an overabundance of extracellular matrix proteins, leading to interstitial fibrosis and a thickened basement membrane. EndMT contributes to the pool of activated cardiac myofibroblasts, lineage tracing has found that 20-35% of cardiac fibroblasts had an endothelial origin (141, 142). Hyperglycemia is a potent cause of EndMT as high levels of glucose damage ECs and result in signaling derangement, resulting in ECs differentiating into a mesenchymal phenotype (143). Oxidative stress triggered by hyperglycemia drives the differentiation of ECs through the TGF- β 1 and TGF- β 2 pathways (144). Oxidative stress induces TGF- β 1 and TGF- β 2 signaling, which results in a reduction of endothelial markers and an increase in fibrotic markers and ECM proteins (145, 146). In addition, TGF- β further contributes to the development of fibrosis through the promotion of if ALK5/Smad3/ NF-KB pathway as well as through aberrant activation of the Ras-GTPase pathway (141, 145-147). Receptor for advanced glycation end products (RAGE) is the receptor of advanced glycation end products (AGEs), both of which are increased in diabetes (148, 149). Knocking out RAGE is able to reduce the degree of EndMT and alleviated cardiac fibrosis in mice (149). As the transcriptional and post-transcriptional level, epigenetic alterations may play a major role in the mediation of EndMT.

EndMT in the heart was first discovered to be involved in the development of heart valves at an embryonic stage (150). Control of EndMT in the heart is epigenetically regulated and the persistent and heritable nature of epigenetic changes contributes to diabetic metabolic memory. Epigenetic modifications take the form of histone modifications, DNA methylation and through ncRNA mediation. During development, EndMT is terminated through the actions of HDAC3 (histone deacetylase 3), which results in the recruitment of EZH2 (enhancer of zeste homolog 2) to silence TGF- β 1 (151). Though no mechanistic link has been made between histone modifications and EndMT in the context of diabetes, HDAC3 has been found to be significantly increased in T2DM patients and HDAC3 mRNA levels were positively correlated to poor glycemic control (152). HDAC3 being both associated with the termination of EndMT while being upregulated in diabetes is an interesting avenue of future studies. DNA methylation is the presence of methyl groups on cytosine bases in CpG islands in DNA (50). Addition of methyl groups via DNA methyltransferases results in stable gene silencing (50). In various models of cardiac fibrosis, the gene RASAL1 (Ras protein activator like 1, a Ras-signaling inhibitor) has been shown to be hypermethylated (147, 153). Methylation of RASAL1 promoter results in increased Ras-GTP activity, resulting in EndMT (153). Noncoding RNA are a class of epigenetic molecules with novel importance. So far, various microRNA have been discovered to be important in the context of cardiac fibrosis. miR-126-3p was found to be downregulated in HUVECs undergoing EndMT and overexpression of miR-126-3p was about to maintain ECs in an

endothelial phenotype (142). Through regulation of the Wnt/ β catenin pathway, miR-222 is able to inhibit EndMT in mouse cardiac endothelial cells, and overexpression of miR-222 in diabetic mice reduces cardiac fibrosis (45). Transgenic mice that overexpressed miR-200b were found to have better cardiac function and reduced EndMT in cardiac tissue (41). miR-21, in contrast, was found to be upregulated in relation to EndMT, and inhibition of miR-21 improved cardiac function (46). Long noncoding RNA (lncRNA) have also been investigated as epigenetic regulators of EndMT. ANRIL regulates diabetic cardiomyopathy in concert with p300 and EZH2 of the PRC2 (polycomb repressive complex 2) complex and the hearts of diabetic ANRIL-knockout mice had reduced levels of ECM (154). LncRNA also interact with microRNA, for example, miR-9-5p interacts with the lncRNA ZFAS1 to mediate cardiac fibrosis in diabetic cardiomyopathy (155). Various other epigenetic modifications, including up- and down-regulated circRNAs have been observed in the diabetic heart which result in increased fibrosis, cardiac remodeling, and heart failure (95). Further investigations into the extent to which epigenetic modifications occur in EndMT in diabetic cardiomyopathy could lead to better and longer-lasting therapies for diabetic complications.

2.5 Endothelial-to-mesenchymal transition and macrovascular complications of diabetes

Atherosclerosis is the premier macrovascular complication of diabetes. It is thought to arise from chronic inflammation and injury to arterial walls, leading to accumulation of plaque which causes arteries to narrow and restricting blood flow (156). It is the cause of coronary artery disease, the most common heart disease in the US and Canada (157). Endothelial dysfunction lies at the heart of atherosclerosis. Hyperglycemic damage to the macrovasculature, as in the microvasculature, results in signaling derangement that leads to EndMT through the generation of reactive oxygen species and inflammatory cytokines (156, 158). Reactive oxygen species induce NF-KB signaling, triggering inflammation, promoting the accumulation of lipids and the formation of a fatty streak-the genesis of plaque (159). The mesenchymal cells derived from EndMT are critical in the progression of atherosclerosis. They secrete proinflammatory signaling molecules and produce and deposit ECM proteins that serve as scaffolding for the forming plaque (160). Recent search has delineated a substantial endothelial origin for mesenchymal cells of the arterial intima, with up to 30% in mice (156). Several pathways activated as a result of hyperglycemia all enhance TGF- β signaling, which directly induces the progression of EndMT (161). Signaling of TGF- β through ALK5 causes the activation of SMAD2/3 which results in the transcription of Snail, Slug and Twist, which contribute to the induction of EndMT and atherosclerosis development (87, 162-166). Various macrovascular complications are comorbid with diabetes. Individuals with T2DM are often also obese and suffer from hypertension. Hypertension, a common comorbidity of T2DM, further activates TGF- β signaling through Smad, exacerbating atherosclerosis.

Epigenetic modifications are also associated with atherosclerosis. PRC2 which is responsible for the repressive H3K27me3 histone modification has been found to be upregulated in the endothelium of blood vessels which are susceptible to atherosclerosis (167, 168). ECs isolated from human plaque show upregulated H3k27me3 when compared to ECs in regions without plaque (169). High levels of the histone modification H2K4me3 (histone 3 lysine 4 trimethylation) has been detected in the ECs of rat aortas exposed to hyperglycemia, which resulted in enriched expression of Notch and development of a mesenchymal-like phenotype (117). EZH2 has been found to be upregulated in atherosclerosis as well as in ECs treated with high glucose (168–170). miR-10a is regulated by NF- κ B signaling pathway and has been found to be downregulated in ECs in athero-susceptible regions compared to elsewhere, and low levels of miR-10a in the serum is associated with human atherosclerosis (171).

Although current studies have yet to concretely establish an epigenetic basis in EndMT in atherosclerosis, various epigenetic modifications that have been found in atherosclerosis are suggestive of EndMT pathways. For example, the previously mentioned miR-126 which is has been previously mentioned to be regulated in EndMT in cardiac fibrosis has also been found to be a regulator of the development of atherosclerosis in the coronary and aortic endothelium. MiR-126 inhibits VCAM-1 (vascular cell adhesion molecule 1), which in atherosclerosis interacts with inflammatory molecules to drive the formation of lesions (172). Atherosclerosis is a disease that exists outside of diabetes, and further research done on the potential role of EndMT may provide novel avenues for treatment of an incredibly prevalent disorder.

3 Molecules targeting EndMT in diabetic complications

A wide variety of pathways have been implicated in the pathogenesis of hyperglycemia-induced EndMT, but contributions of each pathway and the interplay between pathways in the regulation of EndMT is not fully understood. For example, inhibition of one of Notch, canonical TGF- β , or non-canonical TGF- β signaling alone can prevent EndMT in diabetic retinopathy, raising the question of if and how these pathways overlap and are co-regulated. Despite these gaps in understanding, there has been a wealth of research into the influences of various molecules on diabetes-induced EndMT, ranging from pharmaceutical agents which directly target EndMT-inducing pathways to miRNAs which act in an epigenetic manner. Most of these molecules have been identified in organ-specific research, however, as the molecules target common pathways, the findings may be applicable to diabetic complications in other organs as well.

In a case of having the cart before the horse, anti-diabetic drugs dapagliflozin, liraglutide, and linagliptin, which have long been used to aid in the regulation of blood glucose levels, have been shown to inhibit EndMT in of diabetic animals (110, 173, 174). The anti-EndMT effects of these drugs do not appear to be due to direct reduction of serum glucose levels, rather, dapagliflozin and liraglutide act by activating the AMP-activated protein kinase (AMPK), which attenuates intracellular TGF- β signaling (173–175), and linagliptin induces miR-29 to suppress Smad3 phosphorylation in TGF- β signaling. Further up the regulatory chain of hyperglycemia-induced EndMT, dietary supplements

resveratrol and eicosapentaenoic acid have been shown to prevent EndMT in retinal and glomerular ECs respectively, by inhibiting PKC, thereby preventing the induction TGF- β and ET-1 (12, 24, 82, 176, 177). Short interfering RNAs (siRNAs) targeting pro-EndMT genes and lncRNAs, or those that mimic anti-EndMT miRNAs highlighted throughout the previous sections may also be viable approaches to preventing EndMT (178). Experimental silencing of lncRNAs ZFAS1 and MALAT1, and experimental induction of miRNAs 9, 29, 126, 145, 200b, 222, and 497 have proven to be potent suppressors of hyperglycemia-induced EndMT through various pathways (41, 44, 45, 104, 142, 155, 179, 180). Alternatively, synthetic lncRNAs might also be an option to suppress glucose-induced EndMT (181). Experimental upregulation of lncRNAs inhibited by glucose, H19 and MEG3, have been shown to prevent EndMT in diabetic retinopathy (42, 83).

4 Conclusion

For the millions of people living with diabetes, diabetic complications are almost an inevitable cause of morbidity and mortality. Vascular dysfunction lies at the nexus of diabetic complications. The manifestations of diabetic complications vary throughout the body and their regulatory components are different as well. In this review, we have summarized many of the myriad pathways which converge to mediate endothelial-to-mesenchymal transition as well as the epigenetic regulations which maintain EndMT. Though the connection between diabetes and EndMT has been established in diabetic nephropathy, cardiomyopathy and retinopathy, the causational relationship has yet to be experimentally observed in peripheral neuropathy. However, much has yet to be discovered and novel mechanisms have yet to be explored for their therapeutic potential. Considering that diabetic complications have no current treatment aside from symptom management and glycemic control, the discovery of therapies that

References

1. Roglic GWorld Health Organization. *Global report on diabetes*. Geneva, Switzerland: World Health Organization (2016). 86 p.

2. Lin X, Xu Y, Pan X, Xu J, Ding Y, Sun X, et al. Global, regional, and national burden and trend of diabetes in 195 countries and territories: an analysis from 1990 to 2025. *Sci Rep* (2020) 10(1):14790. doi: 10.1038/s41598-020-71908-9

3. Wild S, Roglic G, Green A, Sicree R, King H. Global prevalence of diabetes: Estimates for the year 2000 and projections for 2030. *Diabetes Care* (2004) 27(5):1047–53. doi: 10.2337/diacare.27.5.1047

4. International Diabetes Federation. *IDF diabtes atlas. 10th ed.* (Brussels, Belgium: International Diabetes Federation) (2021).

5. Saeedi P, Petersohn I, Salpea P, Malanda B, Karuranga S, Unwin N, et al. Global and regional diabetes prevalence estimates for 2019 and projections for 2030 and 2045: Results from the international diabetes federation diabetes atlas, 9th edition. *Diabetes Res Clin Pract* (2019) 157:107843. doi: 10.1016/j.diabres.2019.107843

6. De Vriese AS, Verbeuren TJ, Van de Voorde J, Lameire NH, Vanhoutte PM. Endothelial dysfunction in diabetes. *Br J Pharmacol* (2000) 130(5):963–74. doi: 10.1038/sj.bjp.0703393

7. Knapp M, Tu X, Wu R. Vascular endothelial dysfunction, a major mediator in diabetic cardiomyopathy. *Acta Pharmacol Sin* (2019) 40(1):1–8. doi: 10.1038/s41401-018-0042-6

8. Li J, Qu X, Bertram JF. Endothelial-myofibroblast transition contributes to the early development of diabetic renal interstitial fibrosis in streptozotocin-induced diabetic mice. *Am J Pathol* (2009) 175(4):1380–8. doi: 10.2353/ajpath.2009.090096

may reverse or stop the progression of diabetic complications would make a huge impact on the lives of those suffering from the lifealtering morbidities resulting from diabetic complications.

Author contributions

EW and HW are equal contributors in writing the manuscript and the designing of figures. SC participated in topic design, manuscript editing and providing instructional support. All authors contributed to the article and approved the submitted version.

Funding

Funded in part by the Canadian Institutes of Health Research (grant number 173414). EW is also a recipient of the Deans Research Scholarship, Schulich School of Medicine and Dentistry.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

9. Duh EJ, Sun JK, Stitt AW. Diabetic retinopathy: current understanding, mechanisms, and treatment strategies. JCI Insight (2017) 2(14):e93751. doi: 10.1172/jci.insight.93751

10. Gross JL, de Azevedo MJ, Silveiro SP, Canani LH, Caramori ML, Zelmanovitz T. Diabetic nephropathy: Diagnosis, prevention, and treatment. *Diabetes Care* (2005) 28 (1):164–76. doi: 10.2337/diacare.28.1.164

11. Borghetti G, von Lewinski D, Eaton DM, Sourij H, Houser SR, Wallner M. Diabetic cardiomyopathy: Current and future therapies. beyond glycemic control. *Front Physiol* (2018) 9:1514. doi: 10.3389/fphys.2018.01514

12. Brownlee M. The pathobiology of diabetic complications: A unifying mechanism. *Diabetes* (2005) 54(6):1615–25. doi: 10.2337/diabetes.54.6.1615

13. Chan L, Terashima T, Fujimiya M, Kojima H. Chronic diabetic complications: The body's adaptive response to hyperglycemia gone awry? *Trans Am Clin Climatol Assoc* (2006) 117:341–52.

14. Fowler MJ. Microvascular and macrovascular complications of diabetes. *Clin Diabetes* (2011) 29(3):116–22. doi: 10.2337/diaclin.29.3.116

15. Artwohl M, Brunmair B, Fürnsinn C, Hölzenbein T, Rainer G, Freudenthaler A, et al. Insulin does not regulate glucose transport and metabolism in human endothelium. *Eur J Clin Invest* (2007) 37(8):643–50. doi: 10.1111/j.1365-2362.2007.01838.x

16. Yan L. Redox imbalance stress in diabetes mellitus: Role of the polyol pathway. *Anim Model Exp Med* (2018) 1(1):7–13. doi: 10.1002/ame2.12001

17. Schleicher ED, Weigert C. Role of the hexosamine biosynthetic pathway in diabetic nephropathy. *Kidney Int* (2000) 58:S13–8. doi: 10.1046/j.1523-1755.2000.07703.x

18. Koya D, King GL. Protein kinase c activation and the development of diabetic complications. *Diabetes* (1998) 47(6):859–66. doi: 10.2337/diabetes.47.6.859

19. Rhee SY, Kim YS. The role of advanced glycation end products in diabetic vascular complications. *Diabetes Metab J* (2018) 42(3):188–95. doi: 10.4093/dmj.2017.0105

20. Bischoff J. Endothelial-to-Mesenchymal transition. Circ Res (2019) 124(8):1163-5. doi: 10.1161/CIRCRESAHA.119.314813

21. Hong L, Du X, Li W, Mao Y, Sun L, Li X. EndMT: A promising and controversial field. *Eur J Cell Biol* (2018) 97(7):493–500. doi: 10.1016/j.ejcb.2018.07.005

22. Cho JG, Lee A, Chang W, Lee MS, Kim J. Endothelial to mesenchymal transition represents a key link in the interaction between inflammation and endothelial dysfunction. *Front Immunol* (2018) 9:294. doi: 10.3389/fimmu.2018.00294

23. Piera-Velazquez S, Jimenez SA. Endothelial to mesenchymal transition: Role in physiology and in the pathogenesis of human diseases. *Physiol Rev* (2019) 99(2):1281–324. doi: 10.1152/physrev.00021.2018

24. Yu CH, Suriguga, Gong M, Liu WJ, Cui NX, Wang Y, et al. High glucose induced endothelial to mesenchymal transition in human umbilical vein endothelial cell. *Exp Mol Pathol* (2017) 102(3):377–83. doi: 10.1016/j.yexmp.2017.03.007

25. Claesson-Welsh L. Vascular permeability-the essentials. Ups J Med Sci (2015) 120 (3):135-43. doi: 10.3109/03009734.2015.1064501

26. Alvandi Z, Bischoff J. Endothelial-mesenchymal transition in cardiovascular disease. *Arteriosclerosis Thrombosis Vasc Biol* (2021) 41(9):2357-69. doi: 10.1161/ATVBAHA.121.313788

27. Wautier JL, Wautier MP. Vascular permeability in diseases. *Int J Mol Sci* (2022) 23 (7):3645. doi: 10.3390/ijms23073645

28. Piera-Velazquez S, Li Z, Jimenez SA. Role of endothelial-mesenchymal transition (EndoMT) in the pathogenesis of fibrotic disorders. *Am J Pathol* (2011) 179(3):1074–80. doi: 10.1016/j.ajpath.2011.06.001

29. Jimenez SA. Role of endothelial to mesenchymal transition in the pathogenesis of the vascular alterations in systemic sclerosis. *ISRN Rheumatol* (2013) 2013:e835948. doi: 10.1155/2013/835948

30. Di Benedetto P, Ruscitti P, Berardicurti O, Vomero M, Navarini L, Dolo V, et al. Endothelial-to-mesenchymal transition in systemic sclerosis. *Clin Exp Immunol* (2021) 205(1):12–27. doi: 10.1111/cei.13599

31. Hulshoff MS, Xu X, Krenning G, Zeisberg EM. Epigenetic regulation of endothelial-to-Mesenchymal transition in chronic heart disease. *Arteriosclerosis Thrombosis Vasc Biol* (2018) 38(9):1986–96. doi: 10.1161/ATVBAHA.118.311276

32. Islam S, Boström KI, Di Carlo D, Simmons CA, Tintut Y, Yao Y, et al. The mechanobiology of endothelial-to-Mesenchymal transition in cardiovascular disease. *Front Physiol* (2021) 12:734215. doi: 10.3389/fphys.2021.734215

33. Medici D. Endothelial-mesenchymal transition in regenerative medicine. *Stem Cells Int* (2016) 2016:6962801. doi: 10.1155/2016/6962801

34. Medici D, Potenta S, Kalluri R. Transforming growth factor- β 2 promotes snailmediated endothelial-mesenchymal transition through convergence of smad-dependent and smad-independent signaling. *Biochem J* (2011) 437(3):515–20. doi: 10.1042/BJ20101500

35. Ma J, Sanchez-Duffhues G, Goumans MJ, ten Dijke P. TGF- β -Induced endothelial to mesenchymal transition in disease and tissue engineering. *Front Cell Dev Biol* (2020) 8:260. doi: 10.3389/fcell.2020.00260

36. Wermuth PJ, Li Z, Mendoza FA, Jimenez SA. Stimulation of transforming growth factor- β 1-Induced endothelial-To-Mesenchymal transition and tissue fibrosis by endothelin-1 (ET-1): A novel profibrotic effect of ET-1. *PloS One* (2016) 11(9): e0161988. doi: 10.1371/journal.pone.0161988

37. Mahmoud MM, Kim HR, Xing R, Hsiao S, Mammoto A, Chen J, et al. TWIST1 integrates endothelial responses to flow in vascular dysfunction and atherosclerosis. *Circ Res* (2016) 119(3):450–62. doi: 10.1161/CIRCRESAHA.116.308870

38. Li H, Zhao Q, Chang L, Wei C, Bei H, Yin Y, et al. LncRNA MALAT1 modulates ox-LDL induced EndMT through the wnt/ β -catenin signaling pathway. *Lipids Health Dis* (2019) 18(1):62. doi: 10.1186/s12944-019-1006-7

 Gee T, Farrar E, Wang Y, Wu B, Hsu K, Zhou B, et al. NFκB (Nuclear factor κ-Light-Chain enhancer of activated b cells) activity regulates Cell-Type-specific and context-specific susceptibility to calcification in the aortic valve. Arteriosclerosis Thrombosis Vasc Biol (2020) 40(3):638–55. doi: 10.1161/ATVBAHA.119.313248

40. Cao Y, Feng B, Chen S, Chu Y, Chakrabarti S. Mechanisms of endothelial to mesenchymal transition in the retina in diabetes. *Invest Ophthalmol Vis Sci* (2014) 55 (11):7321. doi: 10.1167/iovs.14-15167

41. Feng B, Cao Y, Chen S, Chu X, Chu Y, Chakrabarti S. miR-200b mediates endothelial-to-Mesenchymal transition in diabetic cardiomyopathy. *Diabetes* (2016) 65 (3):768–79. doi: 10.2337/db15-1033

42. Thomas AA, Biswas S, Feng B, Chen S, Gonder J, Chakrabarti S. IncRNA H19 prevents endothelial-mesenchymal transition in diabetic retinopathy. *Diabetologia* (2019) 62(3):517–30. doi: 10.1007/s00125-018-4797-6

43. Widyantoro B, Emoto N, Nakayama K, Anggrahini DW, Adiarto S, Iwasa N, et al. Endothelial cell-derived endothelin-1 promotes cardiac fibrosis in diabetic hearts through stimulation of endothelial-to-Mesenchymal transition. *Circulation* (2010) 121(22):2407– 18. doi: 10.1161/CIRCULATIONAHA.110.938217

44. Zhang J, Zeng Y, Chen J, Cai D, Chen C, Zhang S, et al. miR–29a/b cluster suppresses high glucose–induced endothelial–mesenchymal transition in human retinal microvascular endothelial cells by targeting Notch2. *Exp Ther Med* (2019) 17(4):3108–16. doi: 10.3892/etm.2019.7323

45. Wang Z, Wang Z, Gao L, Xiao L, Yao R, Du B, et al. miR-222 inhibits cardiac fibrosis in diabetic mice heart *via* regulating wnt/β-catenin-mediated endothelium to mesenchymal transition. *J Cell Physiol* (2020) 235(3):2149-60. doi: 10.1002/jcp.29119

46. Li Q, Yao Y, Shi S, Zhou M, Zhou Y, Wang M, et al. Inhibition of miR-21 alleviated cardiac perivascular fibrosis *via* repressing EndMT in T1DM. *J Cell Mol Med* (2020) 24 (1):910–20. doi: 10.1111/jcmm.14800

47. Jaenisch R, Bird A. Epigenetic regulation of gene expression: how the genome integrates intrinsic and environmental signals. *Nat Genet* (2003) 33(3):245-54. doi: 10.1038/ng1089

48. Gibney ER, Nolan CM. Epigenetics and gene expression. *Heredity* (2010) 105(1):4–13. doi: 10.1038/hdy.2010.54

49. Frías-Lasserre D, Villagra CA. The importance of ncRNAs as epigenetic mechanisms in phenotypic variation and organic evolution. *Front Microbiol* (2017) 8:2483. doi: 10.3389/fmicb.2017.02483

50. Moore LD, Le T, Fan G. DNA Methylation and its basic function. Neuropsychopharmacol (2013) 38(1):23–38. doi: 10.1038/npp.2012.112

51. Bannister AJ, Kouzarides T. Regulation of chromatin by histone modifications. *Cell Res* (2011) 21(3):381–95. doi: 10.1038/cr.2011.22

52. Kaikkonen MU, Lam MTY, Glass CK. Non-coding RNAs as regulators of gene expression and epigenetics. *Cardiovasc Res* (2011) 90(3):430–40. doi: 10.1093/cvr/cvr097

53. Statello L, Guo CJ, Chen LL, Huarte M. Gene regulation by long non-coding RNAs and its biological functions. *Nat Rev Mol Cell Biol* (2021) 22(2):96–118. doi: 10.1038/ s41580-020-00315-9

54. Zampetaki A, Albrecht A, Steinhofel K. Long non-coding RNA structure and function: Is there a link? *Front Physiol* (2018) 9:1201. doi: 10.3389/fphys.2018.01201

55. Kornienko AE, Guenzl PM, Barlow DP, Pauler FM. Gene regulation by the act of long non-coding RNA transcription. *BMC Biol* (2013) 11(1):59. doi: 10.1186/1741-7007-11-59

56. Chuang JC, Jones PA. Epigenetics and MicroRNAs. *Pediatr Res* (2007) 61(7):24–9. doi: 10.1203/pdr.0b013e3180457684

57. O'Brien J, Hayder H, Zayed Y, Peng C. Overview of MicroRNA biogenesis, mechanisms of actions, and circulation. *Front Endocrinol (Lausanne)* (2018) 9:402. doi: 10.3389/fendo.2018.00402

58. Miyoshi K, Okada TN, Siomi H, Siomi MC. Characterization of the miRNA-RISC loading complex and miRNA-RISC formed in the drosophila miRNA pathway. *RNA* (2009) 15(7):1282–91. doi: 10.1261/rna.1541209

59. Memczak S, Jens M, Elefsinioti A, Torti F, Krueger J, Rybak A, et al. Circular RNAs are a large class of animal RNAs with regulatory potency. *Nature* (2013) 495(7441):333–8. doi: 10.1038/nature11928

60. Hansen TB, Jensen TI, Clausen BH, Bramsen JB, Finsen B, Damgaard CK, et al. Natural RNA circles function as efficient microRNA sponges. *Nature* (2013) 495 (7441):384-8. doi: 10.1038/nature11993

61. Shao T, Pan Y-H, Xiong X-D. Circular RNA: an important player with multiple facets to regulate its parental gene expression. *Mol Ther Nucleic Acids* (2020) 23:369–76. doi: 10.1016/j.omtn.2020.11.008

62. Zang J, Lu D, Xu A. The interaction of circRNAs and RNA binding proteins: An important part of circRNA maintenance and function. *J Neurosci Res* (2020) 98(1):87–97. doi: 10.1002/jnr.24356

63. Yang Z, Huang C, Wen X, Liu W, Huang X, Li Y, et al. Circular RNA circ-FoxO3 attenuates blood-brain barrier damage by inducing autophagy during ischemia/ reperfusion. *Mol Ther* (2022) 30(3):1275–87. doi: 10.1016/j.ymthe.2021.11.004

64. Chen Y, Wang K, Qian CN, Leach R. DNA Methylation is associated with transcription of snail and slug genes. *Biochem Biophys Res Commun* (2013) 430 (3):1083–90. doi: 10.1016/j.bbrc.2012.12.034

65. Peinado H, Ballestar E, Esteller M, Cano A. Snail mediates e-cadherin repression by the recruitment of the Sin3A/Histone deacetylase 1 (HDAC1)/HDAC2 complex. *Mol Cell Biol* (2004) 24(1):306–19. doi: 10.1128/MCB.24.1.306-319.2004

66. Hong S, Derfoul A, Pereira-Mouries L, Hall DJ. A novel domain in histone deacetylase 1 and 2 mediates repression of cartilage-specific genes in human chondrocytes. *FASEB J* (2009) 23(10):3539–52. doi: 10.1096/fj.09-133215

67. Lecce L, Xu Y, V'Gangula B, Chandel N, Pothula V, Caudrillier A, et al. Histone deacetylase 9 promotes endothelial-mesenchymal transition and an unfavorable atherosclerotic plaque phenotype. *J Clin Invest* (2021) 131(15):e131178. doi: 10.1172/JCI131178

68. Zeng L, Wang G, Ummarino D, Margariti A, Xu Q, Xiao Q, et al. Histone deacetylase 3 unconventional splicing mediates endothelial-to-mesenchymal transition through transforming growth factor $\beta 2$. J Biol Chem (2013) 288(44):31853–66. doi: 10.1074/jbc.M113.463745

69. Wu Z, Liu X, Liu L, Deng H, Zhang J, Xu Q, et al. Regulation of lncRNA expression. Cell Mol Biol Lett (2014) 19(4):561–75. doi: 10.2478/s11658-014-0212-6

70. Li Z, Tan H, Yu H, Deng Z, Zhou X, Wang M. DNA Methylation and gene expression profiles characterize epigenetic regulation of lncRNAs in colon adenocarcinoma. J Cell Biochem (2020) 121(3):2406–15. doi: 10.1002/jcb.29463

71. Song P, Wu L, Guan W. Genome-wide identification and characterization of DNA methylation and long non-coding RNA expression in gastric cancer. *Front Genet* (2020) 11:91. doi: 10.3389/fgene.2020.00091

72. Aure MR, Fleischer T, Bjørklund S, Ankill J, Castro-Mondragon JA, Bathen TF, et al. Crosstalk between microRNA expression and DNA methylation drives the

hormone-dependent phenotype of breast cancer. Genome Med (2021) 13(1):72. doi: 10.1186/s13073-021-00880-4

73. He XX, Kuang SZ, Liao JZ, Xu CR, Chang Y, Wu YL, et al. The regulation of microRNA expression by DNA methylation in hepatocellular carcinoma. *Mol Biosyst* (2015) 11(2):532–9. doi: 10.1039/C4MB00563E

74. Hanly DJ, Esteller M, Berdasco M. Interplay between long non-coding RNAs and epigenetic machinery: emerging targets in cancer? *Philos Trans R Soc Lond B Biol Sci* (2018) 373(1748):20170074. doi: 10.1098/rstb.2017.0074

75. Roccaro AM, Sacco A, Jia X, Azab AK, Maiso P, Ngo HT, et al. microRNAdependent modulation of histone acetylation in waldenström macroglobulinemia. *Blood* (2010) 116(9):1506–14. doi: 10.1182/blood-2010-01-265686

76. Cao P, Deng Z, Wan M, Huang W, Cramer SD, Xu J, et al. MicroRNA-101 negatively regulates Ezh2 and its expression is modulated by androgen receptor and HIF-1 α /HIF-1 β . *Mol Cancer* (2010) 9(1):108. doi: 10.1186/1476-4598-9-108

77. Wang S, Wu W, Claret FX. Mutual regulation of microRNAs and DNA methylation in human cancers. *Epigenetics* (2017) 12(3):187–97. doi: 10.1080/15592294. 2016.1273308

78. Leucci E, Patella F, Waage J, Holmstrøm K, Lindow M, Porse B, et al. microRNA-9 targets the long non-coding RNA MALAT1 for degradation in the nucleus. *Sci Rep* (2013) 3(1):2535. doi: 10.1038/srep02535

79. Yoon JH, Abdelmohsen K, Kim J, Yang X, Martindale JL, Tominaga-Yamanaka K, et al. Scaffold function of long non-coding RNA HOTAIR in protein ubiquitination. *Nat Commun* (2013) 4(1):2939. doi: 10.1038/ncomms3939

80. Yoon JH, Abdelmohsen K, Gorospe M. Functional interactions among microRNAs and long noncoding RNAs. *Semin Cell Dev Biol* (2014) 34:9-14. doi: 10.1016/j.semcdb.2014.05.015

81. Ma X, Chen H, Li L, Yang F, Wu C, Tao K. CircGSK3B promotes RORA expression and suppresses gastric cancer progression through the prevention of EZH2 trans-inhibition. *J Exp Clin Cancer Res* (2021) 40(1):330. doi: 10.1186/s13046-021-02136-w

82. Ergul A. Endothelin-1 and diabetic complications: Focus on the vasculature. *Pharmacol Res* (2011) 63(6):477-82. doi: 10.1016/j.phrs.2011.01.012

83. He Y, Dan Y, Gao X, Huang L, Lv H, Chen J. DNMT1-mediated lncRNA MEG3 methylation accelerates endothelial-mesenchymal transition in diabetic retinopathy through the PI3K/Akt/mTOR signaling pathway. *Am J Physiology-Endocrinol Metab* (2021) 320(3):E598–608. doi: 10.1152/ajpendo.00089.2020

84. Yin Q, He M, Huang L, Zhang X, Zhan J, Hu J. IncRNA ZFAS1 promotes ox-LDL induced EndMT through miR-150-5p/Notch3 signaling axis. *Microvasc Res* (2021) 134:104118. doi: 10.1016/j.mvr.2020.104118

85. Peng H, Li Y, Wang C, Zhang J, Chen Y, Chen W, et al. ROCK1 induces endothelial-to-Mesenchymal transition in glomeruli to aggravate albuminuria in diabetic nephropathy. Sci Rep (2016) 6(1):20304. doi: 10.1038/srep20304

86. Li X, Lu L, Hou W, Wang F, Huang T, Meng Z, et al. The SETD8/ELK1/bach1 complex regulates hyperglycaemia-mediated EndMT in diabetic nephropathy. *J Trans Med* (2022) 20(1):147. doi: 10.1186/s12967-022-03352-4

87. Medici D, Potenta S, Kalluri R. Transforming growth factor- β 2 promotes snailmediated endothelial-mesenchymal transition through convergence of smad-dependent and smad-independent signalling. *Biochem J* (2011) 437(3):515–20. doi: 10.1042/ BJ20101500

88. Wang W, Lo A. Diabetic retinopathy: Pathophysiology and treatments. *IJMS* (2018) 19(6):1816. doi: 10.3390/ijms19061816

89. Lechner J, O'Leary OE, Stitt AW. The pathology associated with diabetic retinopathy. Vision Res (2017) 139:7-14. doi: 10.1016/j.visres.2017.04.003

90. Cheung N, Mitchell P, Wong TY. Diabetic retinopathy. Lancet (2010) 376 (9735):124–36. doi: 10.1016/S0140-6736(09)62124-3

91. Katamay R, Nussenblatt RB. Blood-retinal barrier, immune privilege, and autoimmunity. *Retina* (2013) 1:579–89. doi: 10.1016/B978-1-4557-0737-9.00027-8

92. Eshaq RS, Aldalati AMZ, Alexander JS, Harris NR. Diabetic retinopathy: Breaking the barrier. *Pathophysiology* (2017) 24(4):229–41. doi: 10.1016/j.pathophys.2017.07.001

93. Roy S, Kim D. Retinal capillary basement membrane thickening: Role in the pathogenesis of diabetic retinopathy. *Prog Retin Eye Res* (2021) 82:100903. doi: 10.1016/ j.preteyeres.2020.100903

94. Abu El-Asrar AM, De Hertogh G, van den Eynde K, Alam K, Van Raemdonck K, Opdenakker G, et al. Myofibroblasts in proliferative diabetic retinopathy can originate from infiltrating fibrocytes and through endothelial-to-mesenchymal transition (EndoMT). *Exp Eye Res* (2015) 132:179–89. doi: 10.1016/j.exer.2015.01.023

95. Patil NS, Feng B, Su Z, Castellani CA, Chakrabarti S. Circular RNA mediated gene regulation in chronic diabetic complications. *Sci Rep* (2021) 11(1):23766. doi: 10.1038/s41598-021-02980-y

96. Vinod PB. Pathophysiology of diabetic nephropathy. *Clin Queries: Nephrol* (2012) 1(2):121–6. doi: 10.1016/S2211-9477(12)70005-5

97. Jefferson JA, Shankland SJ, Pichler RH. Proteinuria in diabetic kidney disease: A mechanistic viewpoint. *Kidney Int* (2008) 74(1):22–36. doi: 10.1038/ki.2008.128

98. Qian Y, Feldman E, Pennathur S, Kretzler M, Brosius FC. Mechanisms of glomerulosclerosis in diabetic nephropathy. *Diabetes* (2008) 57(6):1439-45. doi: 10.2337/db08-0061

99. Abbate M, Zoja C, Remuzzi G. How does proteinuria cause progressive renal damage? JASN (2006) 17(11):2974–84. doi: 10.1681/ASN.2006040377

100. Yamagishi S, Matsui T. Advanced glycation end products, oxidative stress and diabetic nephropathy. *Oxid Med Cell Longev* (2010) 3(2):101-8. doi: 10.4161/ oxim.3.2.11148

101. Williams ME. Diabetic nephropathy: The proteinuria hypothesis. AJN (2005) 25 (2):77–94. doi: 10.1159/000084286

102. Eddy AA. Proteinuria and interstitial injury. Nephrol Dialysis Transplant (2004) 19(2):277-81. doi: 10.1093/ndt/gfg533

103. Sol M, Kamps JAAM, van den Born J, van den Heuvel MC, van der Vlag J, Krenning G, et al. Glomerular endothelial cells as instigators of glomerular sclerotic diseases. *Front Pharmacol* (2020) 11:573557. doi: 10.3389/fphar.2020.573557

104. Liu F, Zhang S, Xu R, Gao S, Yin J. Melatonin attenuates endothelial-to-Mesenchymal transition of glomerular endothelial cells *via* regulating miR-497/ROCK in diabetic nephropathy. *KBR* (2018) 43(5):1425–36. doi: 10.1159/000493380

105. Li J, Liu H, Srivastava SP, Hu Q, Gao R, Li S, et al. Endothelial FGFR1 (Fibroblast growth factor receptor 1) deficiency contributes differential fibrogenic effects in kidney and heart of diabetic mice. *Hypertension* (2020) 76(6):1935–44. doi: 10.1161/HYPERTENSIONAHA.120.15587

106. Ying Q, Wu G. Molecular mechanisms involved in podocyte EMT and concomitant diabetic kidney diseases: an update. *Ren Fail* (2017) 39(1):474-83. doi: 10.1080/0886022X.2017.1313164

107. Guan G, Xie J, Dai Y, Han H. TFPI2 suppresses the interaction of TGF- β 2 pathway regulators to promote endothelial-mesenchymal transition in diabetic nephropathy. *J Biol Chem* (2022) 298(3):101725. doi: 10.1016/j.jbc.2022.101725

108. Zeisberg EM, Potenta SE, Sugimoto H, Zeisberg M, Kalluri R. Fibroblasts in kidney fibrosis emerge *via* endothelial-to-Mesenchymal transition. *J Am Soc Nephrol.* (2008) 19(12):2282–7. doi: 10.1681/ASN.2008050513

109. Li J, Qu X, Yao J, Caruana G, Ricardo SD, Yamamoto Y, et al. Blockade of endothelial-mesenchymal transition by a Smad3 inhibitor delays the early development of streptozotocin-induced diabetic nephropathy. *Diabetes* (2010) 59(10):2612–24. doi: 10.2337/db09-1631

110. Kanasaki K, Shi S, Kanasaki M, He J, Nagai T, Nakamura Y, et al. Linagliptinmediated DPP-4 inhibition ameliorates kidney fibrosis in streptozotocin-induced diabetic mice by inhibiting endothelial-to-Mesenchymal transition in a therapeutic regimen. *Diabetes* (2014) 63(6):2120–31. doi: 10.2337/db13-1029

111. Shang J, Zhang Y, Jiang Y, Li Z, Duan Y, Wang L, et al. NOD2 promotes endothelial-to-mesenchymal transition of glomerular endothelial cells *via* MEK/ERK signaling pathway in diabetic nephropathy. *Biochem Biophys Res Commun* (2017) 484 (2):435–41. doi: 10.1016/j.bbrc.2017.01.155

112. Kanno Y, Hirota M, Matsuo O, Ozaki K. $\alpha 2$ -antiplasmin positively regulates endothelial-to-mesenchymal transition and fibrosis progression in diabetic nephropathy. Mol Biol Rep (2022) 49(1):205–15. doi: 10.1007/s11033-021-06859-z

113. Tian Y, Bi Z, Ge S, Ye B, Han W. STAT5A modulated EndMT *via* upregulation of ELTD1 expression in diabetic nephropathy. *Clin Exp Pharmacol Physiol* (2022) 49 (6):686–95. doi: 10.1111/1440-1681.13644

114. Sheldon H, Alexander J, Bridges E, Moreira L, Reilly S, Ang KH, et al. ELTD1 activation induces an endothelial-EMT transition to a myofibroblast phenotype. *Int J Mol Sci* (2021) 22(20):11293. doi: 10.3390/ijms222011293

115. Lu L, Li X, Zhong Z, Zhou W, Zhou D, Zhu M, et al. KMT5A downregulation participated in high glucose-mediated EndMT *via* upregulation of ENO1 expression in diabetic nephropathy. *Int J Biol Sci* (2021) 17(15):4093–107. doi: 10.7150/ijbs.62867

116. Lu L, Zhong Z, Gu J, Nan K, Zhu M, Miao C. ets1 associates with KMT5A to participate in high glucose-mediated EndMT *via* upregulation of PFN2 expression in diabetic nephropathy. *Mol Med* (2021) 27:74. doi: 10.1186/s10020-021-00339-7

117. Pandya Thakkar N, Pereira BMV, Katakia YT, Ramakrishnan SK, Thakar S, Sakhuja A, et al. Elevated H3K4me3 through MLL2-WDR82 upon hyperglycemia causes jagged ligand dependent notch activation to interplay with differentiation state of endothelial cells. *Front Cell Dev Biol* (2022) 10:839109. doi: 10.3389/fcell.2022.839109

118. Calle-Pascual AL, Redondo MJ, Ballesteros M, Martinez-Salinas MA, Diaz JA, De MP, et al. Nontraumatic lower extremity amputations in diabetic and non-diabetic subjects in Madrid, Spain. *Diabetes Metab* (1997) 23(6):519–23. doi: 10.2337/diacare.16.1.16

119. Margolis DJ, Malay DS, Hoffstad OJ, Leonard CE, MaCurdy T, de Nava KL, et al. Incidence of diabetic foot ulcer and lower extremity amputation among Medicare beneficiaries, 2006 to 2008: Data points 2. In: *Data points publication series*. Rockville (MD: Agency for Healthcare Research and Quality (US (2011). Available at: http://www. ncbi.nlm.nih.gov/books/NBK65149/.

120. Frykberg RG. Diabetic foot ulcers: Pathogenesis and management. *afp* (2002) 66 (9):1655–63.

121. Pecoraro RE, Reiber GE, Burgess EM. Pathways to diabetic limb amputation. basis for prevention. *Diabetes Care* (1990) 13(5):513–21. doi: 10.2337/diacare.13.5.513

122. Maser RE, Steenkiste AR, Dorman JS, Nielsen VK, Bass EB, Manjoo Q, et al. Epidemiological correlates of diabetic neuropathy. report from Pittsburgh epidemiology of diabetes complications study. *Diabetes* (1989) 38(11):1456–61. doi: 10.2337/diab.38.11.1456

123. Callaghan BC, Cheng H, Stables CL, Smith AL, Feldman EL. Diabetic neuropathy: Clinical manifestations and current treatments. *Lancet Neurol* (2012) 11(6):521–34. doi: 10.1016/S1474-4422(12)70065-0

124. Agrawal Y, Carey JP, Della Santina CC, Schubert MC, Minor LB. Diabetes, vestibular dysfunction, and falls: analyses from the national health and nutrition examination survey. *Otol Neurotol* (2010) 31(9):1445–50. doi: 10.1097/MAO.0b013e3181f2f035

125. Fagerberg SE. Studies on the pathogenesis of diabetic neuropathy. Acta Med Scandinavica (1957) 159(1):59-62. doi: 10.1111/j.0954-6820.1956.tb14308.x

126. Cameron NE, Eaton SEM, Cotter MA, Tesfaye S. Vascular factors and metabolic interactions in the pathogenesis of diabetic neuropathy. *Diabetologia* (2001) 44(11):1973–88. doi: 10.1007/s001250100001

127. Van Dam PS, Cotter MA, Bravenboer B, Cameron NE. Pathogenesis of diabetic neuropathy: Focus on neurovascular mechanisms. *Eur J Pharmacol* (2013) 719(1):180–6. doi: 10.1016/j.ejphar.2013.07.017

128. Otero-Siliceo E, Ruano-Calderón LA. Diabetic neuropathy: vascular disease. *Rev Neurol* (2003) 37(7):658–61.

129. Sima AA, Nathaniel V, Prashar A, Bril V, Greene DA. Endoneurial microvessels in human diabetic neuropathy. endothelial cell dysjunction and lack of treatment effect by aldose reductase inhibitor. *Diabetes* (1991) 40(9):1090–9. doi: 10.2337/diab.40.9.1090

130. Yagihashi S, Mizukami H, Sugimoto K. Mechanism of diabetic neuropathy: Where are we now and where to go? *J Diabetes Investig* (2011) 2(1):18–32. doi: 10.1111/j.2040-1124.2010.00070.x

131. Gamrat A, Surdacki MA, Chyrchel B, Surdacki A. Endothelial dysfunction: A contributor to adverse cardiovascular remodeling and heart failure development in type 2 diabetes beyond accelerated atherogenesis. *J Clin Med* (2020) 9(7):2090. doi: 10.3390/jcm9072090

132. Jia G, Whaley-Connell A, Sowers JR. Diabetic cardiomyopathy: a hyperglycaemia- and insulin-resistance-induced heart disease. *Diabetologia* (2018) 61 (1):21-8. doi: 10.1007/s00125-017-4390-4

133. Laakso M. Heart in diabetes: A microvascular disease. *Diabetes Care* (2011) 34 (Suppl 2):S145–9. doi: 10.2337/dc11-s209

134. Jia G, Hill MA, Sowers JR. Diabetic cardiomyopathy: An update of mechanisms contributing to this clinical entity. *Circ Res* (2018) 122(4):624–38. doi: 10.1161/ CIRCRESAHA.117.311586

135. Hinkel R, Howe A, Renner S, Ng J, Lee S, Klett K, et al. Diabetes mellitus-induced microvascular destabilization in the myocardium. *J Am Coll Cardiol* (2017) 69(2):131–43. doi: 10.1016/j.jacc.2016.10.058

136. Avogaro A, Azzolina D, Fadini GP, Baldi I. Incidence of heart failure in patients with type 1 diabetes: a systematic review of observational studies. *J Endocrinol Invest.* (2021) 44(4):745–53. doi: 10.1007/s40618-020-01368-5

137. Fang ZY, Prins JB, Marwick TH. Diabetic cardiomyopathy: Evidence, mechanisms, and therapeutic implications. *Endocrine Rev* (2004) 25(4):543-67. doi: 10.1210/er.2003-0012

138. Mizushige K, Yao L, Noma T, Kiyomoto H, Yu Y, Hosomi N, et al. Alteration in left ventricular diastolic filling and accumulation of myocardial collagen at insulinresistant prediabetic stage of a type II diabetic rat model. *Circulation* (2000) 101(8):899– 907. doi: 10.1161/01.CIR.101.8.899

139. Stratton IM, Adler AI, Neil HAW, Matthews DR, Manley SE, Cull CA, et al. Association of glycaemia with macrovascular and microvascular complications of type 2 diabetes (UKPDS 35): prospective observational study. *BMJ* (2000) 321(7258):405–12. doi: 10.1136/bmj.321.7258.405

140. Hinderer S, Schenke-Layland K. Cardiac fibrosis – a short review of causes and therapeutic strategies. *Advanced Drug Delivery Rev* (2019) 146:77–82. doi: 10.1016/j.addr.2019.05.011

141. Zeisberg EM, Tarnavski O, Zeisberg M, Dorfman AL, McMullen JR, Gustafsson E, et al. Endothelial-to-mesenchymal transition contributes to cardiac fibrosis. *Nat Med* (2007) 13(8):952–61. doi: 10.1038/nm1613

142. Jordan NP, Tingle SJ, Shuttleworth VG, Cooke K, Redgrave RE, Singh E, et al. MiR-126-3p is dynamically regulated in endothelial-to-Mesenchymal transition during fibrosis. *Int J Mol Sci* (2021) 22(16):8629. doi: 10.3390/ijms22168629

143. Ghosh AK, Quaggin SE, Vaughan DE. Molecular basis of organ fibrosis: Potential therapeutic approaches. *Exp Biol Med (Maywood)* (2013) 238(5):461–81. doi: 10.1177/1535370213489441

144. Zhang X, Ke PX, Yuan X, Zhang GP, Chen WL, Zhang GS. Forskolin protected against streptozotocin-induced diabetic cardiomyopathy *via* inhibition of oxidative stress and cardiac fibrosis in mice. *BioMed Res Int* (2021) 2021:8881843. doi: 10.1155/2021/8881843

145. Pardali E, Sanchez-Duffhues G, Gomez-Puerto MC, ten Dijke P. TGF-β-Induced endothelial-mesenchymal transition in fibrotic diseases. *Int J Mol Sci* (2017) 18(10):2157. doi: 10.3390/ijms18102157

146. Montorfano I, Becerra A, Cerro R, Echeverría C, Sáez E, Morales MG, et al. Oxidative stress mediates the conversion of endothelial cells into myofibroblasts *via* a TGF- β 1 and TGF- β 2-dependent pathway. *Lab Invest.* (2014) 94(10):1068–82. doi: 10.1038/labinvest.2014.100

147. Xu X, Tan X, Tampe B, Nyamsuren G, Liu X, Maier LS, et al. Epigenetic balance of aberrant Rasal1 promoter methylation and hydroxymethylation regulates cardiac fibrosis. *Cardiovasc Res* (2015) 105(3):279–91. doi: 10.1093/cvr/cvv015

148. Hou X, Hu Z, Xu H, Xu J, Zhang S, Zhong Y, et al. Advanced glycation endproducts trigger autophagy in cadiomyocyte via RAGE/PI3K/AKT/mTOR pathway. Cardiovasc Diabetol (2014) 13:78. doi: 10.1186/1475-2840-13-78

149. Zhang L, He J, Wang J, Liu J, Chen Z, Deng B, et al. Knockout RAGE alleviates cardiac fibrosis through repressing endothelial-to-mesenchymal transition (EndMT) mediated by autophagy. *Cell Death Dis* (2021) 12(5):1–13. doi: 10.1038/s41419-021-03750-4

150. Markwald RR, Fitzharris TP, Smith WNA. Structural analysis of endocardial cytodifferentiation. *Dev Biol* (1975) 42(1):160–80. doi: 10.1016/0012-1606(75)90321-8

151. Lewandowski SL, Janardhan HP, Trivedi CM. Histone deacetylase 3 coordinates deacetylase-independent epigenetic silencing of transforming growth factor-β1 (TGF-β1) to orchestrate second heart field development *. *J Biol Chem* (2015) 290(45):27067–89. doi: 10.1074/jbc.M115.684753

152. Sathishkumar C, Prabu P, Balakumar M, Lenin R, Prabhu D, Anjana RM, et al. Augmentation of histone deacetylase 3 (HDAC3) epigenetic signature at the interface of proinflammation and insulin resistance in patients with type 2 diabetes. *Clin Epigenet* (2016) 8:125. doi: 10.1186/s13148-016-0293-3

153. Xu X, Friehs I, Zhong Hu T, Melnychenko I, Tampe B, Alnour F, et al. Endocardial fibroelastosis is caused by aberrant endothelial to mesenchymal transition. *Circ Res* (2015) 116(5):857–66. doi: 10.1161/CIRCRESAHA.116.305629

154. Thomas AA, Feng B, Chakrabarti S. ANRIL regulates production of extracellular matrix proteins and vasoactive factors in diabetic complications. *Am J Physiology-Endocrinol Metab* (2018) 314(3):E191–200. doi: 10.1152/ajpendo.00268.2017

155. Feng B, Liu J, Wang E, Su Z, Chakrabarti S. Endothelial derived miRNA-9 mediated cardiac fibrosis in diabetes and its regulation by ZFAS1. *PloS One* (2022) 17(10): e0276076. doi: 10.1371/journal.pone.0276076

156. Chen PY, Qin L, Baeyens N, Li G, Afolabi T, Budatha M, et al. Endothelial-tomesenchymal transition drives atherosclerosis progression. *J Clin Invest.* (2015) 125 (12):4514–28. doi: 10.1172/JCI82719

157. Herrington W, Lacey B, Sherliker P, Armitage J, Lewington S. Epidemiology of atherosclerosis and the potential to reduce the global burden of atherothrombotic disease. *Circ Res* (2016) 118(4):535–46. doi: 10.1161/circresaha.115.307611

158. Giordo R, Ahmed YMA, Allam H, Abusnana S, Pappalardo L, Nasrallah GK, et al. EndMT regulation by small RNAs in diabetes-associated fibrotic conditions: Potential link with oxidative stress. *Front Cell Dev Biol* (2021) 9:683594. doi: 10.3389/fcell.2021.683594

159. Chang JC, Kou SJ, Lin WT, Liu CS. Regulatory role of mitochondria in oxidative stress and atherosclerosis. *World J Cardiol* (2010) 2(6):150–9. doi: 10.4330/wjc.v2.i6.150

160. Evrard SM, Lecce L, Michelis KC, Nomura-Kitabayashi A, Pandey G, Purushothaman KR, et al. Endothelial to mesenchymal transition is common in atherosclerotic lesions and is associated with plaque instability. *Nat Commun* (2016) 7 (1):11853. doi: 10.1038/ncomms11853

161. Chen PY, Schwartz MA, Simons M. Endothelial-to-Mesenchymal transition, vascular inflammation, and atherosclerosis. *Front Cardiovasc Med* (2020) 7. doi: 10.3389/ fcvm.2020.00053

162. Oh SP, Seki T, Goss KA, Imamura T, Yi Y, Donahoe PK, et al. Activin receptorlike kinase 1 modulates transforming growth factor- β 1 signaling in the regulation of angiogenesis. *Proc Natl Acad Sci* (2000) 97(6):2626–31. doi: 10.1073/pnas.97.6.2626

163. Goumans MJ, Valdimarsdottir G, Itoh S, Rosendahl A, Sideras P, ten Dijke P. Balancing the activation state of the endothelium *via* two distinct TGF- β type I receptors. *EMBO J* (2002) 21(7):1743–53. doi: 10.1093/emboj/21.7.1743

164. Kokudo T, Suzuki Y, Yoshimatsu Y, Yamazaki T, Watabe T, Miyazono K. Snail is required for TGF β -induced endothelial-mesenchymal transition of embryonic stem cell-derived endothelial cells. *J Cell Sci* (2008) 121(20):3317–24. doi: 10.1242/jcs.028282

165. Romano LA, Runyan RB. Slug is an essential target of TGFβ2 signaling in the developing chicken heart. *Dev Biol* (2000) 223(1):91–102. doi: 10.1006/dbio.2000.9750

166. Chakraborty S, Wirrig EE, Hinton RB, Merrill WH, Spicer DB, Yutzey KE. Twist1 promotes heart valve cell proliferation and extracellular matrix gene expression during development *in vivo* and is expressed in human diseased aortic valves. *Dev Biol* (2010) 347 (1):167–79. doi: 10.1016/j.ydbio.2010.08.021

167. Xu S, Xu Y, Yin M, Zhang S, Liu P, Koroleva M, et al. Flow-dependent epigenetic regulation of IGFBP5 expression by H3K27me3 contributes to endothelial antiinflammatory effects. *Theranostics* (2018) 8(11):3007–21. doi: 10.7150/thno.21966

168. Maleszewska M, Vanchin B, Harmsen MC, Krenning G. The decrease in histone methyltransferase EZH2 in response to fluid shear stress alters endothelial gene expression and promotes quiescence. *Angiogenesis* (2016) 19(1):9–24. doi: 10.1007/s10456-015-9485-2

169. Greißel A, Culmes M, Burgkart R, Zimmermann A, Eckstein HH, Zernecke A, et al. Histone acetylation and methylation significantly change with severity of atherosclerosis in human carotid plaques. *Cardiovasc Pathol* (2016) 25(2):79–86. doi: 10.1016/j.carpath.2015.11.001

170. Lv YC, Tang YY, Zhang P, Wan W, Yao F, He PP, et al. Histone methyltransferase enhancer of zeste homolog 2-mediated ABCA1 promoter DNA methylation contributes to the progression of atherosclerosis. *PloS One* (2016) 11(6):e0157265. doi: 10.1371/journal.pone.0157265

171. Fang Y, Shi C, Manduchi E, Civelek M, Davies PF. MicroRNA-10a regulation of proinflammatory phenotype in athero-susceptible endothelium in vivo and *in vitro*. *Proc Natl Acad Sci U.S.A.* (2010) 107(30):13450–5. doi: 10.1073/pnas.1002120107

172. Santovito D, Egea V, Bidzhekov K, Natarelli L, Mourão A, Blanchet X, et al. Noncanonical inhibition of caspase-3 by a nuclear microRNA confers endothelial protection by autophagy in atherosclerosis. *Sci Transl Med* (2020) 12(546):eaaz2294. doi: 10.1126/scitranslmed.aaz2294

173. Tsai TH, Lee CH, Cheng CI, Fang YN, Chung SY, Chen SM, et al. Liraglutide inhibits endothelial-to-Mesenchymal transition and attenuates neointima formation after endovascular injury in streptozotocin-induced diabetic mice. *Cells* (2019) 8(6):589. doi: 10.3390/cells8060589

174. Tian J, Zhang M, Suo M, Liu D, Wang X, Liu M, et al. Dapagliflozin alleviates cardiac fibrosis through suppressing EndMT and fibroblast activation via $AMPK\alpha/TGF$ -

 β/Smad signalling in type 2 diabetic rats. J Cell Mol Med (2021) 25(16):7642–59. doi: 10.1111/jcmm.16601

175. Lin H, Li N, He H, Ying Y, Sunkara S, Luo L, et al. AMPK inhibits the stimulatory effects of TGF-β on Smad2/3 activity, cell migration, and epithelial-to-Mesenchymal transition. *Mol Pharmacol* (2015) 88(6):1062–71. doi: 10.1124/mol.115.099549

176. Giordo R, Nasrallah GK, Posadino AM, Galimi F, Capobianco G, Eid AH, et al. Resveratrol-elicited PKC inhibition counteracts NOX-mediated endothelial to mesenchymal transition in human retinal endothelial cells exposed to high glucose. *Antioxidants (Basel)* (2021) 10(2):224. doi: 10.3390/antiox10020224

177. Yasuzawa T, Nakamura T, Ueshima S, Mima A. Protective effects of eicosapentaenoic acid on the glomerular endothelium *via* inhibition of EndMT in diabetes. *J Diabetes Res* (2021) 2021:2182225. doi: 10.1155/2021/2182225

178. Hu B, Zhong L, Weng Y, Peng L, Huang Y, Zhao Y, et al. Therapeutic siRNA: state of the art. *Sig Transduct Target Ther* (2020) 5(1):1–25. doi: 10.1038/s41392-020-0207-x

179. Liu B, Qiang L, Wang GD, Duan Q, Liu J. LncRNA MALAT1 facilities high glucose induced endothelial to mesenchymal transition and fibrosis *via* targeting miR-145/ZEB2 axis. *Eur Rev Med Pharmacol Sci* (2019) 23(8):3478–86. doi: 10.26355/ eurrev_201904_17713

180. Xiang Y, Zhang Y, Tang Y, Li Q. MALAT1 modulates TGF- β 1-Induced endothelial-to-Mesenchymal transition through downregulation of miR-145. Cell Physiol Biochem (2017) 42(1):357–72. doi: 10.1159/000477479

181. Mercer TR, Munro T, Mattick JS. The potential of long noncoding RNA therapies. *Trends Pharmacol Sci* (2022) 43(4):269-80. doi: 10.1016/j.tips.2022.01.008