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# The causal relationship between obesity and skin and soft tissue infections: A two-sample Mendelian randomization study

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**Objective:** Many observational studies have shown that obesity strongly affects skin and soft tissue infections (SSTIs). However, whether a causal genetic relationship exists between obesity and SSTIs is unclear.

**Methods:** A two-sample Mendelian randomization (MR) study was used to explore whether obesity is causally associated with SSTIs using a publicly released genome-wide association study (GWAS). An inverse-variance weighted (IVW) analysis was used as the primary analysis, and the results are reported as the odds ratios (ORs). Heterogeneity was tested using Cochran's Q test and the I<sup>2</sup> statistic, and horizontal pleiotropy was tested using the MR-Egger intercept and MR pleiotropy residual sum and outlier (MR-PRESSO).

**Results:** The results of the MR analysis showed a positive effect of BMI on SSTIs (OR 1.544, 95% CI 1.399-1.704,  $P = 5.86 \times 10^{-18}$ ). After adjusting for the effect of type 2 diabetes (T2D) and peripheral vascular disease (PVD), the positive effect still existed. Then, we further assessed the effect of BMI on different types of SSTIs. The results showed that BMI caused an increased risk of impetigo, cutaneous abscess, furuncle and carbuncle, cellulitis, pilonidal cyst, and other local infections of skin and subcutaneous tissues, except for acute lymphadenitis. However, the associations disappeared after adjusting for the effect of T2D and PVD, and the associations between BMI and impetigo or cellulitis disappeared. Finally, we assessed the effects of several obesity-related characteristics on SSTIs. Waist circumference, hip circumference, body fat percentage, and whole-body fat mass, excluding waist-to-hip ratio, had a causal effect on an increased risk of SSTIs. However, the associations disappeared after adjusting for the effect of BMI.

**Conclusion:** This study found that obesity had a positive causal effect on SSTIs. Reasonable weight control is a possible way to reduce the occurrence of SSTIs, especially in patients undergoing surgery.

#### KEYWORDS

obesity, skin and soft tissue infections, genome-wide association study, Mendelian randomization, causal relationship

## Introduction

Obesity is an excess of body fat that is detrimental to health and is often assessed clinically by the body mass index (BMI) (1). Obesity is a major public health problem, and in 2016, the World Health Organization (WHO) estimated that more than 1.9 billion people aged 18 years and older were overweight. More than 650 million of them are obese. In addition, more than 340 million children and adolescents aged 5-19 years are overweight or obese (2). Obesity reduces the health-related quality of life and longevity, and increases the risk of type 2 diabetes, coronary artery disease, gallbladder disease, and hyperlipidemia (3). Skin and soft tissue infections (SSTIs) are pathogenic bacteria that invade the epidermis, dermis, and subcutaneous tissues and induce a host response, and the diagnosis is based primarily on clinical features (4-6). The clinical features include sclerosis, erythema, fever, and pain or induration. Local manifestations may be accompanied by systemic signs and symptoms, such as fever, chills, and sometimes hemodynamic instability (5). SSTIs constitute the most common infectious disease in all age groups, and their incidence is increasing every year with the increase in surgical procedures, the use of immunosuppressive drugs, and cancer (7). SSTIs often require inpatient treatment and place a great burden on the health care system (8-10).

A cohort study involving 171,322 adults from 2011 to 2016 found that increased BMI was associated with an increased risk of cellulitis and hospitalization for cellulitis, and that obesity was an independent risk factor for cellulitis, after adjusting for confounders (11). Another case-control study noted that obese patients were 1.76 times more likely to have a surgical site infection than nonobese individuals (12). However, recently, it has been found that adipogenesis of the skin is an important source of resistance to infection and antimicrobial peptides (13, 14), which contradicts many observations regarding the increased risk of bacterial skin infections in obese individuals (15). Based on the results of these studies, the causal relationship between obesity and SSTIs is unclear as some confounding factors may have biased the results, such as the strong association between obesity and type 2 diabetes (T2D) and peripheral vascular disease (PVD) (5, 16). Therefore, the

causal relationship between obesity and SSTIs has not been established.

Genetic epidemiology is used to elucidate the determinants of disease because the inheritance of genetic variants is random and cannot be confounded by additional risk factors (17, 18). Mendelian randomization is a method used to assess whether a causal relationship exists between exposure factors and outcomes using genetic variants as instrumental variables that are equally, randomly, and independently distributed during the split (19-21). Therefore, we used two-sample MR to assess the causal relationship between study obesity and SSTIs.

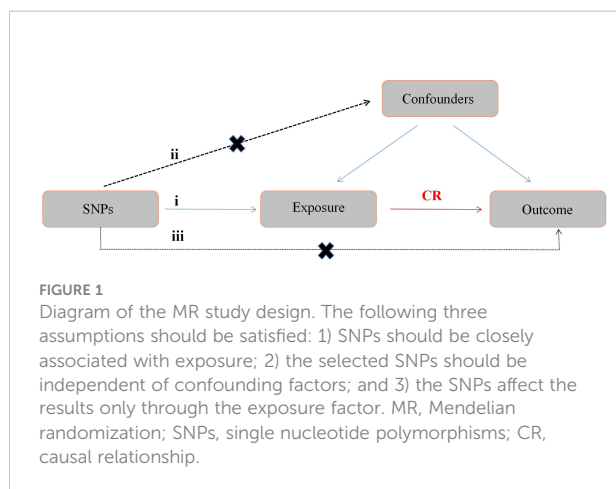
## Methods

### Study design

A two-sample MR design were used in this study, and genome-wide association study (GWAS) data were used to determine the causal relationship between obesity and SSTIs. Single nucleotide polymorphisms (SNPs) were instrumental variables for obesity. The BMI is often used to measure and assess obesity in the clinical practice (22). In addition, several other obesity-related characteristics were used in the assessment, including waist circumference (WC), hip circumference (HC), waist-to-hip ratio (WHR), body fat percentage (BF), and whole body fat mass (FM) (23-25). The main assumptions of this method include the following: 1) SNPs are associated with exposure factors, 2) SNPs are independent of confounding factors, and 3) SNPs affect outcomes only *via* exposure factors (Figure 1).

### Genome-wide association study summary data

The two-sample MR analysis in this study was performed based on data from a GWAS. BMI, obesity-related characteristics, and SSTIs (SSTIs as defined by the International Classification of Diseases- Tenth Revision code L00-L08) were obtained from the



MRC IEU OpenGWAS database (<https://gwas.mrcieu.ac.uk/>), which consists mainly of GWAS data that are currently publicly available for use in Mendelian randomization analyses (26, 27). All GWAS were tested using imputed genotype data from the UK Biobank study. All GWASs were tested using imputed genotype data from the UK Biobank study. The GWAS summary of BMI was based on 461460 individuals of European ancestry, and the GWAS summary statistics of SSTIs were based on 218792 individuals in the European population (10343 SSTI cases and 208449 controls).

We analyzed the causal relationship between BMI and the risk of SSTIs using univariate MR. Next, we reassessed the effect of BMI on SSTIs after adjusting for the effect of T2D and PVD on BMI. Then, we assessed the effect of SSTI on BMI. We performed a stratified analysis of the different types of SSTIs to further assess the effect of BMI on different types of SSTIs. Finally, we assessed the causal effects of several obesity-related characteristics. The details of the data sources are provided in [Supplementary Table S1](#).

## Instrumental variable selection

To select eligible genetic IVs that met the MR assumptions, we established a quality control technique. First, we selected independent SNPs that were strongly associated with the exposure factors with  $p$  values  $< 5 \times 10^{-8}$ . In assessing the effect of SSTIs on BMI, we expanded the  $p$  value to  $< 1 \times 10^{-5}$  to select suitable genetic instruments considering that few SNPs were associated with SSTIs at the  $p$  value  $< 5 \times 10^{-8}$  level. Then, to exclude SNPs with strong linkage disequilibrium, we performed an aggregation process ( $R^2 < 0.001$ ) (28). Finally, to ensure that the effect alleles belonged to the same allele, we adjusted the exposure and outcome datasets to eliminate ambiguous SNPs with inconsistent alleles and SNPs with intermediate allele frequencies (29).

We calculated the  $F$  statistic of each SNP as follows:  $F = R^2 \times (N - 2) / (1 - R^2)$  (30).  $R^2$  denotes the variance of exposure explained by each IV according to the calculation method used in Papadimitriou

et al. (30). IVs with  $F$ -statistic values less than 10 are considered weak instruments and were excluded from the MR analysis (31).

## Statistical analysis

The inverse variance weighted (IVW) method was used as the main method to analyze the causal relationship between obesity and SSTIs. The causal effect of each SNP on the outcome was assessed by calculating the Wald ratio of each SNP, and the inverse variance of the SNP was used as the weight in a meta-analysis to evaluate the combined causal effect. In addition, we used MR-Egger, weighted median, sample mode, and weighted mode to assess the causal relationship between BMI and SSTIs. MR-Egger has low statistical power; thus, the focus is more on direction and effect (32, 33). The weighted median provides a reliable Mendelian evaluation when 50% of the instrument variables are not valid (34). The odds ratio (OR) and 95% confidence interval (CI) were used to assess the relative risk.

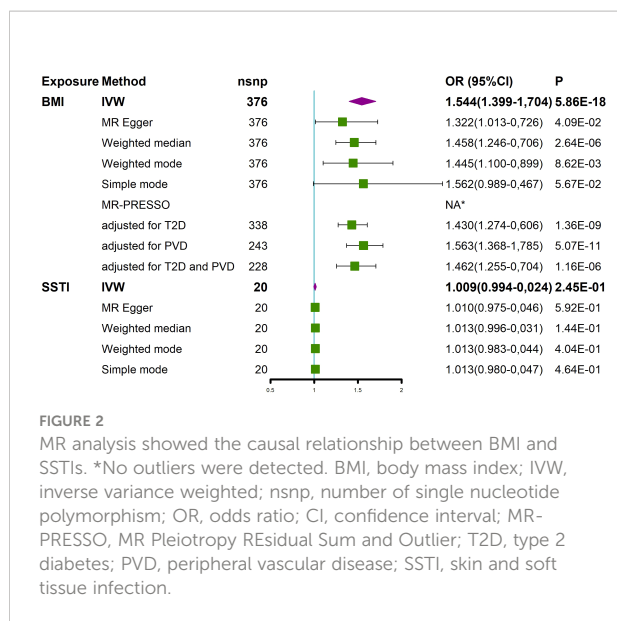
We used the Cochran's  $Q$  value and  $I^2$  statistic to assess heterogeneity among the SNPs and the MR-Egger method to test for horizontal pleiotropy. In addition, the MR pleiotropy residual sum and outlier (MR-PRESSO) method was used to detect outliers in the analysis and assess the adjusted causal effects after excluding the outliers (35). A sensitivity analysis was performed using the leave-one-out sensitivity test to assess the validity and stability of the MR results. We used MR-Steiger filtering to remove SNPs that implied reverse causal direction (36).

The data were analyzed using R (version 4.1.2) software with the R packages "two-sample MR" and "MRPRESSO".  $P < 0.05$  was considered statistically significant. The data used in this study were publicly available, and therefore, the study did not require ethical approval.

## Results

### Mendelian randomization to assess the causal relationship between obesity and SSTIs

The effect of BMI on the risk of an SSTI was first assessed using the univariate MR method (Figure 2 and [Supplementary Figure S1](#)). The results using the IVW method showed that with a 1-SD increase in the BMI level, the OR of SSTIs was 1.544 (95% CI 1.399-1.704,  $P = 5.86 \times 10^{-18}$ ). The MR-Egger method (OR=1.322, 95% CI 1.013-1.726,  $P = 4.09 \times 10^{-2}$ ) and weighted median (OR=1.458, 95% CI 1.246-1.706,  $P = 2.64 \times 10^{-6}$ ) also showed consistent results. In the above analysis, Cochran's  $Q$  and  $I^2$  tests did not detect heterogeneity, and the MR-Egger test did not detect horizontal pleiotropy ([Supplementary Figure S2](#) and [Supplementary Table S2](#)). The MR-PRESSO method did not detect outliers; thus, the association between BMI and SSTIs and



**FIGURE 2**  
MR analysis showed the causal relationship between BMI and SSTIs. \*No outliers were detected. BMI, body mass index; IVW, inverse variance weighted; nsnp, number of single nucleotide polymorphism; OR, odds ratio; CI, confidence interval; MR-PRESSO, MR Pleiotropy Residual Sum and Outlier; T2D, type 2 diabetes; PVD, peripheral vascular disease; SSTI, skin and soft tissue infection.

its significance remained unchanged (Figure 2). To assess whether these results were affected by a single SNP, we performed the leave-one-out sensitivity test, which showed that the causal effect of BMI on SSTIs did not significantly fluctuate in the absence of any single SNP (Supplementary Figure S3). We performed MR-Steiger filtering of the SNPs of BMI and did not find SNPs with reverse causality.

Next, we assessed the effect of an adjusted BMI on SSTIs after excluding T2D and PVD, which often coexist with obesity (Figure 2). After adjusting for T2D, BMI was still causally associated with an increased risk of SSTIs (OR=1.430, 95% CI 1.274-1.606, P=1.36×10<sup>-9</sup>). After adjusting for PVD, BMI was also causally associated with an increased risk of SSTIs (OR=1.563, 95% CI 1.368-1.785, P=5.07×10<sup>-11</sup>). Finally, after we adjusted for T2D and PVD, an increase in BMI was still positively associated with an increased risk of SSTIs (OR=1.462, 95% CI 1.255-1.704, P=1.16×10<sup>-6</sup>).

Finally, we also explored whether SSTIs had a causal effect on the BMI. The results of the IVW method showed no effect of SSTIs on the BMI. Due to the presence of heterogeneity (Supplementary Table S2), the results of the weighted median analysis were used according to Nazarzadeh et al. (37). The results of the weighted median method showed no effect of SSTIs on BMI (Figure 2).

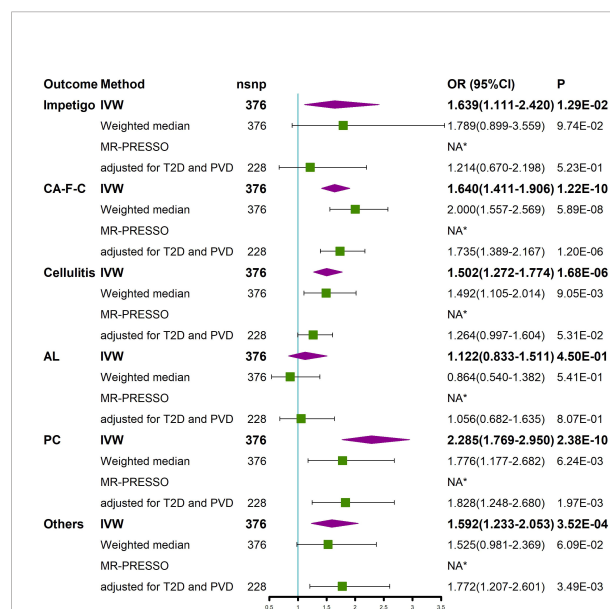
### Causal effects of BMI on different types of SSTIs

We performed a stratified analysis of different types of SSTIs (according to ICD-Tenth Revision) to further assess the effect of BMI on different types of SSTIs (Figure 3). SNPs associated with staphylococcal scalded skin syndrome (SSSS) were not available in the GWAS; thus, SSSS was not included in the type

stratification of SSTIs. The results of the univariate MR analysis showed that an increase in BMI caused an increased risk of impetigo, cutaneous abscess, furuncle and carbuncle (CA-F-C), cellulitis, pilonidal cyst (PC), and other local infections of skin and subcutaneous tissues, except for acute lymphadenitis (AL) (Figure 3). Next, we adjusted for the effects of T2D and PVD. The results of the multivariate MR study showed that after adjusting for both T2D and PVD, an increase in BMI resulted in an increased risk of CA-F-C, PC, and other local infections of skin and subcutaneous tissues (Figure 3). In addition, no outliers of BMI were detected by the MR-PRESSO method (Figure 3).

### Causal effect of obesity-related characteristics on SSTIs

We investigated whether obesity-related characteristics (WC, HC, WHR, BF, and FM) had a causal effect on SSTIs. The univariate MR analysis showed that the obesity-related characteristics of WC, HC, BF, and FM, excluding WHR, had a causal effect on the increased risk of SSTIs (Figure 4). The



**FIGURE 3**  
Univariable MR and multivariable MR analyses showed the causal relationship between BMI and different types of SSTIs. Univariable MR was performed using the IVW, weighted median, and MR-PRESSO methods. Multivariable MR was performed to assess the effect of BMI on different types of SSTIs after adjusting for T2D and PVD. \*No outliers were detected. BMI, body mass index; IVW, inverse variance weighted; nsnp, number of single nucleotide polymorphism; OR, odds ratio; CI, confidence interval; MR-PRESSO, MR Pleiotropy Residual Sum and Outlier; T2D, type 2 diabetes; PVD, peripheral vascular disease; CA-F-C, cutaneous abscess, furuncle and carbuncle; AL, acute lymphadenitis; PC, pilonidal cyst; Others, other local infections of skin and subcutaneous tissues; SSTIs, skin and soft tissue infections.



correlations were consistent in all sensitivity analyses, although the MR-Egger regression analysis detected pleiotropy in HC and BF. After removing outliers using the MR-PRESSO method, these factors remained significantly associated with SSTIs (Figure 4). Multivariate MR analyses adjusting for BMI were performed because these obesity-related characteristics were highly correlated with BMI. The results of the study showed that none of these obesity-related features were significantly associated with SSTIs after adjusting for BMI (Figure 4).

## Discussion

Many previous clinical studies have noted that obesity is a risk factor for a variety of diseases, including SSTIs. However, due to the limitations of previous studies, these findings may be influenced by confounding factors and reverse causality. Our study was based on MR to explore the relationship between obesity and SSTIs. The results of the study found a positive causal effect of obesity on an increased risk of SSTIs. This positive causality persisted after adjusting for the effects of T2D and PVD.

Our findings suggest that obesity increases the risk of SSTIs, which is similar to the results of previous studies (15). This finding may be related to the metabolic effects of obesity and

inadequate blood supply to adipose tissue (AT), which leads to poor host immune recruitment and thus causes skin defenses to be diminished (9, 38). Healthy AT acts as a defense barrier against microorganisms. Adipocytes are able to produce antimicrobial peptides (AMPs) in response to skin infections, thereby strengthening the innate immune defenses of the skin (39). Several findings confirm that host defense is mediated through adipocyte production of AMPs and that the inhibition of adipocytes reduces the expression of AMPs (13, 14). Recently, an animal study (40) found that obesity in mice leads to a loss of dermal preadipocytes (pADs) and inhibits the ability to initiate responsive adipose expression of antimicrobial peptides, and obesity leads to a loss of dermal fibroblasts (dFBs), which have the ability to polarize into mature adipocytes capable of expressing antimicrobial peptides, further promoting the development of infection. Obesity causes hypertrophy and proliferation in adipocytes, leading to a rapid increase in adipose tissue, and when angiogenesis cannot match the expanded AT, local hypoxia in AT causes an increase in pro-inflammatory leptin and a decrease in anti-inflammatory adiponectin (41–44). In addition, hypoxia progresses to induce endoplasmic reticulum stress, which combined with excess extracellular matrix (ECM) deposition, causes AT fibrosis (41, 45). All these changes promote the release of inflammatory factors and intensify the local inflammatory response, ultimately leading to adipocyte dysfunction and metabolic changes and causing decreased immunity and insulin resistance (41, 43–46). Furthermore, hypoxia in AT causes adipocytes to undergo anaerobic glycolysis, promoting the increased production and release of lactate, which further promotes the inflammatory pathway of macrophages, resulting in chronic inflammation (47, 48).

In chronic inflammation, immune cells are constantly stimulated by cytokines or chemokines and are activated, eventually causing immune cell depletion (49). In addition, chronic inflammation decreases the formation of memory T cells, reducing the ability of T cells to protect the host against infection (50). Recently, it has been found that long-term stimulation by chronic inflammation causes a significant decrease in the proliferative capacity of hematopoietic stem cells, the origin of immune cells, resulting in accumulative and irreversible functional damage and long-term suppressive effects on hematopoiesis, further contributing to immune cell failure and decreased immune function (51).

Obesity also affects the function of immune cells, which leads to decreased immune function and promotes the development of infections. Macrophages are key mediators of inflammation in adipose tissue and are the most abundant immune cells (52). In the obese state, macrophages mainly accumulate around adipocytes, forming crown-like structures and proliferating to clear dysfunctional and necrotic adipocytes, and inflammation cannot be cleared due to persistent obesity; then, inflammation changes from local inflammation to a systemic chronic inflammatory state

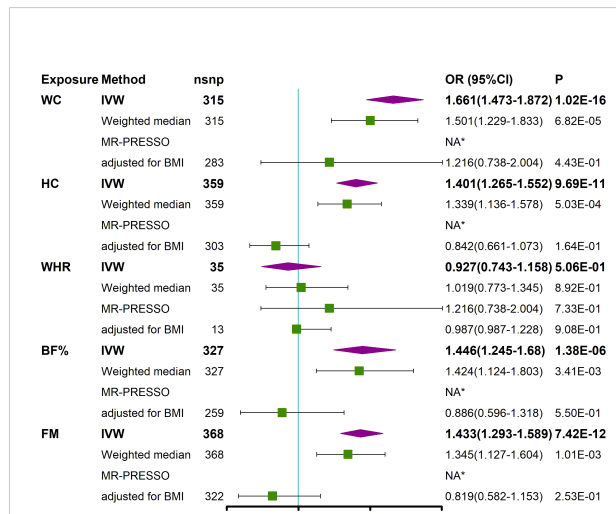


FIGURE 4

MR analyses showed a causal relationship between obesity-related characteristics and SSTIs. Univariable MR was performed using the IVW, weighted median, and MR-PRESSO methods. Multivariable MR was performed to assess the effect of obesity-related characteristics on SSTIs after adjusting for BMI. \*No outliers were detected. BMI, body mass index; IVW, inverse variance weighted; nsnp, number of single nucleotide polymorphism; OR, odds ratio; CI, confidence interval; MR-PRESSO, MR Pleiotropy RESidual Sum and Outlier; WC, waist circumference; HC, hip circumference; WHR, waist-to-hip ratio; BF, body fat percentage; FM, whole body fat mass; SSTIs, skin and soft tissue infections.

(41, 53). Other research has found that macrophages exhibit an anti-inflammatory M2 phenotype in thin mice, whereas in diet-induced obese (DIO) mice, macrophages switch from an anti-inflammatory M2 phenotype to a proinflammatory M1 phenotype, leading to chronic inflammation (54, 55). Additionally, these macrophages show decreased phagocytosis and a decreased ability to clear bacteria (44). In addition, obesity also inhibits T-lymphocyte receptor diversity, leading to abnormalities in the antigen presentation process, decreasing the efficiency of T-lymphocyte responses and increasing susceptibility to infection in obese individuals (44, 56). Neutrophils play a role in the inflammatory response induced by obesity, and neutrophils in AT produce chemokines and cytokines that promote macrophage infiltration, contribute to chronic low-grade inflammation, and induce insulin resistance (57). Furthermore, the imbalance of neutrophil elastase (NE) and its inhibitor  $\alpha$ 1-antitrypsin in obese individuals causes chronic inflammation and insulin resistance (58). Moreover, obesity causes the downregulation of neutrophil function, which reduces bacterial clearance (44, 59). Furthermore, the humoral immune response is defective in obese individuals, and B lymphocytes in obese individuals express a proinflammatory phenotype that reduces their ability to optimally respond to infection (44).

Obesity or high-fat diets modify the composition of the gut microbiota and increase intestinal permeability, leading to the passage of bacteria and bacterial products (e.g., lipopolysaccharides, LPS) across the intestinal barrier into the circulation, which can activate innate immune cells and immune signaling pathways and contribute to inflammation formation (2, 44, 60–63). In addition, a high-fat diet increases the proportion of gram-negative bacteria, further promoting the absorption of LPS across the intestinal barrier (60, 64). These factors all lead to a low-grade systemic inflammatory response (65–67). As obesity advances, the eventual mild low-grade inflammation becomes chronic inflammation that further causes systemic inflammation (68–70).

Overall, obesity leads to immune system dysregulation, a reduced cell-mediated immune response, and modified gut microbiota, representing possible reasons for the increased risk of infection associated with obesity (16, 38, 71–73). Similar to the findings reported by Conway et al. (74), the present study also showed that the genetic prediction of BMI was positively causally associated with an increased risk of SSTIs. Therefore, the risk of infection should be noted in obese patients, especially those who undergo surgery. Preoperatively, obese patients may be advised to lose weight to reduce systemic inflammation and the risk of infection (75).

We performed a stratified analysis of the types of SSTIs, and the findings showed that an increased BMI was positively associated with an increased risk of impetigo, CA-F-C, cellulitis, PC, and other local infections of skin and subcutaneous tissues. However, after adjusting for T2D and PVD, an increased BMI was not significantly associated with the development of cellulitis, which may be due to obesity-

induced elevated blood glucose due to insulin resistance and PVD to further mediate the development of cellulitis (76, 77). In addition, although we stratified the degree of severity and site of SSTIs, the GWAS lacked data of the different degrees or different sites of SSTIs; thus, an MR analysis with stratification of different degrees or sites of SSTIs is not available.

In addition, this study evaluated the relationship between several other obesity-related characteristics and the risk of SSTIs. Previous studies have found that obesity characteristics, such as WC and BF, are predictors of infection (78, 79). In the univariate MR analysis, we also found a positive association between waist circumference, hip circumference, body fat percentage and whole-body fat mass and the risk of SSTIs. However, this association may be mediated through the effect of BMI as we did not observe this significant effect after adjusting for BMI. Furthermore, the OR of HC, BF and FM on SSTIs was less than 1 after adjusting for BMI, although there was no significance, possibly implying that increased HC, BF, and FM are protective factors against SSTIs, similar to a recent cohort study that found a lower risk of death in people with a small WC and large HC (80). However, our findings are only based on the MR method, and large clinical studies are still needed to confirm whether such a relationship exists. In view of our findings, we recommend using BMI rather than other obesity characteristics in the risk assessment of SSTIs in the clinic.

To the best of our knowledge, this study is the first to use a two-sample MR analysis to assess the causal relationship between obesity and SSTIs. In addition, the genetic variants used as IVs were extracted from the largest GWAS. However, there are some limitations to this study. 1). The MR analysis conducted in this study was based on a European cohort, and whether there is a demographic effect on the results is unclear. 2). The genetic data of exposure and outcome were pooled data from a GWAS, and relevant data, such as disease prevalence and age specificity, are lacking. 3). This study focuses on the causal relationship between obesity and SSTIs, but whether a causal relationship exists between weight loss and SSTIs is unclear.

## Conclusion

This study shows that a positive causal relationship exists between a genetically predicted increase in BMI and an increased risk of SSTIs, further deepening our understanding of obesity causing SSTIs and providing guidance for prevention and treatment.

## Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and

accession number(s) can be found below: <https://gwas.mrcieu.ac.uk/datasets/>.

## Author contributions

HH and ML performed the study and wrote the manuscript. XW and HL revised the manuscript. JM wrote sections of the manuscript and performed the statistical analysis. GC designed the study. All authors contributed to the article and approved the submitted version.

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## Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## References

- González-Muniesa P, Martínez-González M-A, Hu FB, Després J-P, Matsuzawa Y, Loos RJJ, et al. Obesity. *Nat Rev Dis Primers* (2017) 3:17034. doi: 10.1038/nrdp.2017.34
- Uribe-Querol E, Rosales C. Neutrophils actively contribute to obesity-associated inflammation and pathological complications. *Cells* (2022) 11:1883. doi: 10.3390/cells11121883
- W D, Da A. The economic burden of obesity worldwide: a systematic review of the direct costs of obesity. *Obes Rev* (2011) 12:131–41. doi: 10.1111/j.1467-789X.2009.00712.x
- Ramakrishnan K, Salinas RC, Agudelo Higuera NI. Skin and soft tissue infections. *Am Fam Phys* (2015) 92:474–83.
- Eron LJ, Lipsky BA, Low DE, Nathwani D, Tice AD, Volturo GA. Expert panel on managing skin and soft tissue infections. managing skin and soft tissue infections: expert panel recommendations on key decision points. *J Antimicrob Chemother* (2003) 52 Suppl 1:i3–17. doi: 10.1093/jac/dkg466
- Fung HB, Chang JY, Kuczynski S. A practical guide to the treatment of complicated skin and soft tissue infections. *Drugs* (2003) 63:1459–80. doi: 10.2165/00003495-200363140-00003
- Suaya JA, Mera RM, Cassidy A, O'Hara P, Amrine-Madsen H, Burstin S, et al. Incidence and cost of hospitalizations associated with staphylococcus aureus skin

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## Supplementary material

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fendo.2022.996863/full#supplementary-material>

### SUPPLEMENTARY FIGURE 1

Plots of MR estimates of the causal relationship between SSTIs and BMI. (A) Scatter plot of SNPs showing the causal effect of BMI on SSTIs. The log odds ratio of risk is demonstrated, and five different methods were used. (B) Forest plot of SNPs associated with BMI on SSTIs. MR, Mendelian randomization; SSTI, skin and soft tissue infection; BMI, body mass index; SNP, single nucleotide polymorphism.

### SUPPLEMENTARY FIGURE 2

Funnel plot showed there were no significant heterogeneity among SNPs. SNPs, single nucleotide polymorphisms.

### SUPPLEMENTARY FIGURE 3

Leave-one-out analysis plots for BMI on SSTIs. MR, Mendelian randomization; SSTI, skin and soft tissue infection; BMI, body mass index.

### SUPPLEMENTARY TABLE 1

Data source information. BMI, body mass index; T2D, type 2 diabetes; SSTI, skin and soft tissue infection.

### SUPPLEMENTARY TABLE 2

Heterogeneity tests and directional horizontal pleiotropy test.

and soft tissue infections in the united states from 2001 through 2009. *BMC Infect Dis* (2014) 14:296. doi: 10.1186/1471-2334-14-296

8. Olaniyi R, Pozzi C, Grimaldi L, Bagnoli F. Staphylococcus aureus-associated skin and soft tissue infections: Anatomical localization, epidemiology, therapy and potential prophylaxis. *Curr Top Microbiol Immunol* (2017) 409:199–227. doi: 10.1007/82\_2016\_32

9. Mofarah AS, Al Mohajer M, Hurwitz BL, Armstrong DG. Skin and soft tissue infections. *Microbiol Spectr* (2016) 4:1–16. doi: 10.1128/microbiolspec.DMIH2-0014-2015

10. Corcione S, De Rosa FG. The optimal duration of treatment for skin and soft tissue infections and acute bacterial skin and skin structure infections. *Curr Opin Infect Dis* (2018) 31:155–62. doi: 10.1097/QCO.0000000000000440

11. Cheong HS, Chang Y, Joo E-J, Cho A, Ryu S. Metabolic obesity phenotypes and risk of cellulitis: A cohort study. *J Clin Med* (2019) 8:E953. doi: 10.3390/jcm8070953

12. Vilar-Compte D, Mohar A, Sandoval S, de la Rosa M, Gordillo P, Volkow P. Surgical site infections at the national cancer institute in Mexico: A case-control study. *Am J Infect Control* (2000) 28:14–20. doi: 10.1016/s0196-6553(00)90006-3

13. Zhang L, Guerrero-Juarez CF, Hata T, Bapat SP, Ramos R, Plikus MV, et al. Innate immunity. dermal adipocytes protect against invasive staphylococcus aureus skin infection. *Science* (2015) 347:67–71. doi: 10.1126/science.1260972

14. Zhang L-J, Chen SX, Guerrero-Juarez CF, Li F, Tong Y, Liang Y, et al. Age-related loss of innate immune antimicrobial function of dermal fat is mediated by transforming growth factor beta. *Immunity* (2019) 50:121–136.e5. doi: 10.1016/j.immuni.2018.11.003
15. Karppelin M, Siljander T, Vuopio-Varkila J, Kere J, Huhtala H, Vuento R, et al. Factors predisposing to acute and recurrent bacterial non-necrotizing cellulitis in hospitalized patients: a prospective case-control study. *Clin Microbiol Infect* (2010) 16:729–34. doi: 10.1111/j.1469-0691.2009.02906.x
16. Jalilvand AD, Villarreal M, Schubauer K, Rosales J, Gonzales-Gallo K, Wisler JR. Characterization of necrotizing soft tissue infections in obese and non-obese patients. *Surg Infect (Larchmt)* (2022) 23:475–82. doi: 10.1089/sur.2022.055
17. He B, Lyu Q, Yin L, Zhang M, Quan Z, Ou Y. Depression and osteoporosis: A mendelian randomization study. *Calcif Tissue Int* (2021) 109:675–84. doi: 10.1007/s00223-021-00886-5
18. Emdin CA, Khera AV, Kathiresan S. Mendelian randomization. *JAMA* (2017) 318:1925–6. doi: 10.1001/jama.2017.17219
19. Goto A, Yamaji T, Sawada N, Momozawa Y, Kamatani Y, Kubo M, et al. Diabetes and cancer risk: A mendelian randomization study. *Int J Cancer* (2020) 146:712–9. doi: 10.1002/ijc.32310
20. Vaucher J, Keating BJ, Lasserre AM, Gan W, Lyall DM, Ward J, et al. Cannabis use and risk of schizophrenia: a mendelian randomization study. *Mol Psychiatry* (2018) 23:1287–92. doi: 10.1038/mp.2016.252
21. Hartwig FP, Borges MC, Horta BL, Bowden J, Davey Smith G. Inflammatory biomarkers and risk of schizophrenia: A 2-sample mendelian randomization study. *JAMA Psychiatry* (2017) 74:1226–33. doi: 10.1001/jamapsychiatry.2017.3191
22. Yuan S, Gill D, Giovannucci EL, Larsson SC. Obesity, type 2 diabetes, lifestyle factors, and risk of gallstone disease: A mendelian randomization investigation. *Clin Gastroenterol Hepatol* (2022) 20:e529–37. doi: 10.1016/j.cgh.2020.12.034
23. Shah SM, Loney T, Dhaheri SA, Vatanparast H, Elbarazi I, Agarwal M, et al. Association between acculturation, obesity and cardiovascular risk factors among male south Asian migrants in the united Arab Emirates—a cross-sectional study. *BMC Public Health* (2015) 15:204. doi: 10.1186/s12889-015-1568-x
24. Zhang X, Theodoratou E, Li X, Farrington SM, Law PJ, Broderick P, et al. Genetically predicted physical activity levels are associated with lower colorectal cancer risk: a mendelian randomisation study. *Br J Cancer* (2021) 124:1330–8. doi: 10.1038/s41416-020-01236-2
25. Paterson EN, Maxwell AP, Kee F, Cruise S, Young IS, McGuinness B, et al. Association of renal impairment with cognitive dysfunction in the northern Ireland cohort for the longitudinal study of ageing (NICOLA). *Nephrol Dial Transplant* (2021) 36:1492–9. doi: 10.1093/ndt/gfab182
26. Hemani G, Zheng J, Elsworth B, Wade KH, Haberland V, Baird D, et al. The MR-base platform supports systematic causal inference across the human phenome. *Elife* (2018) 7:e34408. doi: 10.7554/eLife.34408
27. Liu Y-Q, Liu Y, Chen Z-Y, Li H, Xiao T. Rheumatoid arthritis and osteoporosis: a bi-directional mendelian randomization study. *Aging (Albany NY)* (2021) 13:14109–30. doi: 10.18632/aging.203029
28. 1000 Genomes Project Consortium, Auton A, Brooks LD, Durbin RM, Garrison EP, Kang HM, et al. A global reference for human genetic variation. *Nature* (2015) 526:68–74. doi: 10.1038/nature15393
29. Yin K-J, Huang J-X, Wang P, Yang X-K, Tao S-S, Li H-M, et al. No genetic causal association between periodontitis and arthritis: A bidirectional two-sample mendelian randomization analysis. *Front Immunol* (2022) 13:808832. doi: 10.3389/fimmu.2022.808832
30. Papadimitriou N, Dimou N, Tsilidis KK, Banbury B, Martin RM, Lewis SJ, et al. Physical activity and risks of breast and colorectal cancer: a mendelian randomisation analysis. *Nat Commun* (2020) 11:597. doi: 10.1038/s41467-020-14389-8
31. Burgess S, Thompson SGCRP CHD Genetics Collaboration. Avoiding bias from weak instruments in mendelian randomization studies. *Int J Epidemiol* (2011) 40:755–64. doi: 10.1093/ije/dyr036
32. Luo S, Au Yeung SL, Zuber V, Burgess S, Schooling CM. Impact of genetically predicted red blood cell traits on venous thromboembolism: Multivariable mendelian randomization study using UK biobank. *J Am Heart Assoc* (2020) 9:e016771. doi: 10.1161/JAHA.120.016771
33. Cai Y, Zhang G, Liang J, Jing Z, Zhang R, Lv L, et al. Causal relationships between osteoarthritis and senile central nerve system dysfunction: A bidirectional two-sample mendelian randomization study. *Front Aging Neurosci* (2021) 13:793023. doi: 10.3389/fnagi.2021.793023
34. Wu D, Xian W, Hong S, Liu B, Xiao H, Li Y. Graves' disease and rheumatoid arthritis: A bidirectional mendelian randomization study. *Front Endocrinol (Lausanne)* (2021) 12:702482. doi: 10.3389/fendo.2021.702482
35. Mi J, Liu Z. Obesity, type 2 diabetes, and the risk of carpal tunnel syndrome: A two-sample mendelian randomization study. *Front Genet* (2021) 12:688849. doi: 10.3389/fgene.2021.688849
36. Molenberg R, Thio CHL, Aalbers MW, Uyttenboogaart M, ISGC Intracranial Aneurysm Working Group\*, Larsson SC, et al. Sex hormones and risk of aneurysmal subarachnoid hemorrhage: A mendelian randomization study. *Stroke* (2022) 53:2870–5. doi: 10.1161/STROKEAHA.121.038035
37. Nazarzadeh M, Pinho-Gomes A-C, Bidel Z, Dehghan A, Canoy D, Hassaine A, et al. Plasma lipids and risk of aortic valve stenosis: a mendelian randomization study. *Eur Heart J* (2020) 41:3913–20. doi: 10.1093/eurheartj/ehaa070
38. Falagas ME, Athanasoulia AP, Peppas G, Karageorgopoulos DE. Effect of body mass index on the outcome of infections: a systematic review. *Obes Rev* (2009) 10:280–9. doi: 10.1111/j.1467-789X.2008.00546.x
39. Guerrero-Juarez CF, Plikus MV. Emerging nonmetabolic functions of skin fat. *Nat Rev Endocrinol* (2018) 14:163–73. doi: 10.1038/nrendo.2017.162
40. Zhang L-J, Guerrero-Juarez CF, Chen SX, Zhang X, Yin M, Li F, et al. Diet-induced obesity promotes infection by impairment of the innate antimicrobial defense function of dermal adipocyte progenitors. *Sci Transl Med* (2021) 13:eabb5280. doi: 10.1126/scitranslmed.abb5280
41. de Heredia FP, Gómez-Martínez S, Marcos A. Obesity, inflammation and the immune system. *Proc Nutr Soc* (2012) 71:332–8. doi: 10.1017/S0029665112000092
42. Bradley D, Xu A, Hsueh WA. Editorial: The immunomodulatory roles of adipocytes. *Front Immunol* (2021) 12:827281. doi: 10.3389/fimmu.2021.827281
43. Sun K, Kusminski CM, Scherer PE. Adipose tissue remodeling and obesity. *J Clin Invest* (2011) 121:2094–101. doi: 10.1172/JCI45887
44. De Bandt J-P, Monin C. Obesity, nutrients and the immune system in the era of COVID-19. *Nutrients* (2021) 13:610. doi: 10.3390/nu13020610
45. Sun K, Tordjman J, Clément K, Scherer PE. Fibrosis and adipose tissue dysfunction. *Cell Metab* (2013) 18:470–7. doi: 10.1016/j.cmet.2013.06.016
46. Marcelin G, Silveira ALM, Martins LB, Ferreira AV, Clément K. Deciphering the cellular interplays underlying obesity-induced adipose tissue fibrosis. *J Clin Invest* (2019) 129:4032–40. doi: 10.1172/JCI129192
47. Pérez de Heredia F, Wood IS, Trayhurn P. Hypoxia stimulates lactate release and modulates monocarboxylate transporter (MCT1, MCT2, and MCT4) expression in human adipocytes. *Pflugers Arch* (2010) 459:509–18. doi: 10.1007/s00424-009-0750-3
48. Samuvel DJ, Sundararaj KP, Nareika A, Lopes-Virella MF, Huang Y. Lactate boosts TLR4 signaling and NF-kappaB pathway-mediated gene transcription in macrophages via monocarboxylate transporters and MD-2 up-regulation. *J Immunol* (2009) 182:2476–84. doi: 10.4049/jimmunol.0802059
49. Aubert RD, Kamphorst AO, Sarkar S, Vezys V, Ha S-J, Barber DL, et al. Antigen-specific CD4 T-cell help rescues exhausted CD8 T cells during chronic viral infection. *Proc Natl Acad Sci U.S.A.* (2011) 108:21182–7. doi: 10.1073/pnas.1118450109
50. Stelekati E, Shin H, Doering TA, Dolfi DV, Ziegler CG, Beiting DP, et al. Bystander chronic infection negatively impacts development of CD8(+) T cell memory. *Immunity* (2014) 40:801–13. doi: 10.1016/j.immuni.2014.04.010
51. Bogeska R, Mikecin A-M, Kaschnig P, Fawaz M, Büchler-Schäff M, Le D, et al. Inflammatory exposure drives long-lived impairment of hematopoietic stem cell self-renewal activity and accelerated aging. *Cell Stem Cell* (2022) 29:1273–1284.e8. doi: 10.1016/j.stem.2022.06.012
52. Odegaard JI, Chawla A. Alternative macrophage activation and metabolism. *Annu Rev Pathol* (2011) 6:275–97. doi: 10.1146/annurev-pathol-011110-130138
53. Haase J, Weyer U, Immig K, Klötting N, Blüher M, Eilers J, et al. Local proliferation of macrophages in adipose tissue during obesity-induced inflammation. *Diabetologia* (2014) 57:562–71. doi: 10.1007/s00125-013-3139-y
54. Lumeng CN, Bodzin JL, Saltiel AR. Obesity induces a phenotypic switch in adipose tissue macrophage polarization. *J Clin Invest* (2007) 117:175–84. doi: 10.1172/JCI29881
55. Lumeng CN, Deyoung SM, Bodzin JL, Saltiel AR. Increased inflammatory properties of adipose tissue macrophages recruited during diet-induced obesity. *Diabetes* (2007) 56:16–23. doi: 10.2337/db06-1076
56. McLaughlin T, Ackerman SE, Shen L, Engleman E. Role of innate and adaptive immunity in obesity-associated metabolic disease. *J Clin Invest* (2017) 127:5–13. doi: 10.1172/JCI88876
57. Talukdar S, Oh DY, Bandyopadhyay G, Li D, Xu J, McNelis J, et al. Neutrophils mediate insulin resistance in mice fed a high-fat diet through secreted elastase. *Nat Med* (2012) 18:1407–12. doi: 10.1038/nm.2885
58. Mansuy-Aubert V, Zhou QL, Xie X, Gong Z, Huang J-Y, Khan AR, et al. Imbalance between neutrophil elastase and its inhibitor  $\alpha$ 1-antitrypsin in obesity alters insulin sensitivity, inflammation, and energy expenditure. *Cell Metab* (2013) 17:534–48. doi: 10.1016/j.cmet.2013.03.005



59. Mancuso P, Curtis JL, Weitzel AM, Griffin CA, Bouchard B, Freeman CM, et al. Diet-induced obesity in mice impairs host defense against klebsiella pneumonia in vivo and glucose transport and bactericidal functions in neutrophils in vitro. *Am J Physiol Lung Cell Mol Physiol* (2022) 322:L116–28. doi: 10.1152/ajplung.00008.2021
60. Cani PD, Amar J, Iglesias MA, Poggi M, Knauf C, Bastelica D, et al. Metabolic endotoxemia initiates obesity and insulin resistance. *Diabetes* (2007) 56:1761–72. doi: 10.2337/db06-1491
61. Tilg H, Zmora N, Adolph TE, Elinav E. The intestinal microbiota fuelling metabolic inflammation. *Nat Rev Immunol* (2020) 20:40–54. doi: 10.1038/s41577-019-0198-4
62. Wei Y-X, Zheng K-Y, Wang Y-G. Gut microbiota-derived metabolites as key mucosal barrier modulators in obesity. *World J Gastroenterol* (2021) 27:5555–65. doi: 10.3748/wjg.v27.i33.5555
63. Cani PD, Bibiloni R, Knauf C, Waget A, Neyrinck AM, Delzenne NM, et al. Changes in gut microbiota control metabolic endotoxemia-induced inflammation in high-fat diet-induced obesity and diabetes in mice. *Diabetes* (2008) 57:1470–81. doi: 10.2337/db07-1403
64. Amar J, Burcelin R, Ruidavets JB, Cani PD, Fauvel J, Alessi MC, et al. Energy intake is associated with endotoxemia in apparently healthy men. *Am J Clin Nutr* (2008) 87:1219–23. doi: 10.1093/ajcn/87.5.1219
65. Gérard P. Gut microbiota and obesity. *Cell Mol Life Sci* (2016) 73:147–62. doi: 10.1007/s00018-015-2061-5
66. Cox AJ, West NP, Cripps AW. Obesity, inflammation, and the gut microbiota. *Lancet Diabetes Endocrinol* (2015) 3:207–15. doi: 10.1016/S2213-8587(14)70134-2
67. Boulangé CL, Neves AL, Chilloux J, Nicholson JK, Dumas M-E. Impact of the gut microbiota on inflammation, obesity, and metabolic disease. *Genome Med* (2016) 8:42. doi: 10.1186/s13073-016-0303-2
68. Gregor MF, Hotamisligil GS. Inflammatory mechanisms in obesity. *Annu Rev Immunol* (2011) 29:415–45. doi: 10.1146/annurev-immunol-031210-101322
69. Illán-Gómez F, González-Ortega M, Orea-Soler I, Alcaraz-Tafalla MS, Aragón-Alonso A, Pascual-Díaz M, et al. Obesity and inflammation: change in adiponectin, c-reactive protein, tumour necrosis factor-alpha and interleukin-6 after bariatric surgery. *Obes Surg* (2012) 22:950–5. doi: 10.1007/s11695-012-0643-y
70. Shoelson SE, Herrero L, Naaz A. Obesity, inflammation, and insulin resistance. *Gastroenterology* (2007) 132:2169–80. doi: 10.1053/j.gastro.2007.03.059
71. Falagas ME, Kompoti M. Obesity and infection. *Lancet Infect Dis* (2006) 6:438–46. doi: 10.1016/S1473-3099(06)70523-0
72. Huttunen R, Syrjänen J. Obesity and the risk and outcome of infection. *Int J Obes (Lond)* (2013) 37:333–40. doi: 10.1038/ijo.2012.62
73. Hirt PA, Castillo DE, Yosipovitch G, Keri JE. Skin changes in the obese patient. *J Am Acad Dermatol* (2019) 81:1037–57. doi: 10.1016/j.jaad.2018.12.070
74. Conway EL, Sellick JA, Kurtzhals K, Mergenhagen KA. Obesity and heart failure as predictors of failure in outpatient skin and soft tissue infections. *Antimicrob Agents Chemother* (2017) 61:e02389–16. doi: 10.1128/AAC.02389-16
75. Sheu WH-H, Chang T-M, Lee W-J, Ou H-C, Wu C-M, Tseng L-N, et al. Effect of weight loss on proinflammatory state of mononuclear cells in obese women. *Obes (Silver Spring)* (2008) 16:1033–8. doi: 10.1038/oby.2008.37
76. Carey IM, Critchley JA, DeWilde S, Harris T, Hosking FJ, Cook DG. Risk of infection in type 1 and type 2 diabetes compared with the general population: A matched cohort study. *Diabetes Care* (2018) 41:513–21. doi: 10.2337/dc17-2131
77. Zacay G, Hershkovitz Sikron F, Heymann AD. Glycemic control and risk of cellulitis. *Diabetes Care* (2021) 44:367–72. doi: 10.2337/dc19-1393
78. Fernández-Real J-M, Ferri M-J, Vendrell J, Ricart W. Burden of infection and fat mass in healthy middle-aged men. *Obes (Silver Spring)* (2007) 15:245–52. doi: 10.1038/oby.2007.541
79. Chen Y-Y, Fang W-H, Wang C-C, Kao T-W, Chang Y-W, Yang H-F, et al. Increased body fat percentage in patients with hepatitis b and c virus infection. *PloS One* (2018) 13:e0200164. doi: 10.1371/journal.pone.0200164
80. Cameron AJ, Romaniuk H, Orellana L, Dallongeville J, Dobson AJ, Drygas W, et al. Combined influence of waist and hip circumference on risk of death in a Large cohort of European and Australian adults. *J Am Heart Assoc* (2020) 9:e015189. doi: 10.1161/JAHA.119.015189