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Low back pain (LBP) is a disabling condition with no available cure, severely affecting patients' quality of life. Intervertebral disc degeneration (IVDD) is the leading cause of chronic low back pain (CLBP). IVDD is a common and recurrent condition in spine surgery. Disc degeneration is closely associated with intervertebral disc inflammation. The intervertebral disc is an avascular tissue in the human body. Transitioning from hematopoietic bone marrow to bone marrow fat may initiate an inflammatory response as we age, resulting in bone marrow lesions in vertebrae. In addition, the development of LBP is closely associated with spinal stability imbalance. An excellent functional state of paraspinal muscles (PSMs) plays a vital role in maintaining spinal stability. Studies have shown that the diminished function of PSMs is mainly associated with increased fat content, but whether the fat content of PSMs is related to the degree of disc degeneration is still under study. Given the vital role of PSMs lesions in CLBP, it is crucial to elucidate the interaction between PSMs changes and CLBP. Therefore, this article reviews the advances in the relationship and the underlying mechanisms between IVDD and PSMs fatty infiltration in patients with CLBP.

KEYWORDS

low back pain, intervertebral disc degeneration, paraspinal muscles, fatty infiltration, Modic changes, inflammation

Introduction

Low back pain (LBP) is a disabling condition with no available cure, often caused by a sedentary lifestyle and reduced exercise ([1\)](#page-6-0), severely affecting patients' quality of life. Chronic low back pain (CLBP) accounts for approximately 23% of LBP [\(2\)](#page-6-0). Intervertebral disc degeneration (IVDD) is the leading cause of CLBP ([3](#page-6-0)). IVDD is a chronic, multifactorial and irreversible process that severely compromises spinal stability and

disc shock absorption [\(3](#page-6-0)). Early biochemical changes in IVDD include loss of proteoglycans and water, while late morphological changes include reduced disc height, nucleus pulposus herniation, and annular tears [\(4\)](#page-6-0). The paraspinal muscles (PSMs) are fundamental determinants of the structural stability and function of the lumbar spine ([5](#page-6-0)). There is a potential mechanism of action between defects in the vertebral endplate and decreased muscle mass of the PSMs during the development of disc degeneration. Previous studies have shown that CLPB induced myoelectric activity and muscle remodeling (e.g., muscle atrophy, fatty infiltration, and altered fiber type) [\(6](#page-6-0)–[10\)](#page-6-0). At the L4/L5 level, fatty infiltration in PSMs is more severe when damage to the adjacent spinal endplates ([11](#page-6-0)). Thus the formation of IVDD is not an isolated process but a chain reaction that includes vertebral endplate changes and fatty infiltration in the PSMs [\(12\)](#page-6-0). Given the vital role of PSMs lesions in CLBP, it is crucial to elucidate the interaction between PSMs changes and CLBP. Therefore, this article reviews the advances in the relationship and the underlying mechanisms between IVDD and PSMs fatty infiltration in patients with CLBP.

Fatty infiltration in PSMs

The PSMs are the general term for the muscles surrounding the spine, which include the psoas, multifidus, and erector spinae. An excellent functional state of the PSMs is essential for maintaining the spine structure. A decrease in the function of PSMs can alter the original biomechanical relationships and increase the load on the disc, thus causing IVDD. Conversely, IVDD can also cause PSMs to compensate, leading to an imbalance in loads of PSMs and producing atrophy. Studies have shown that the atrophy of the PSMs is highly correlated with the degree of IVDD. Muscle degeneration is characterized by atrophy of muscle fibers, fiber bundles, and fat infiltration ([13](#page-6-0), [14\)](#page-6-0). Muscle atrophy and fat replacement are thought to be the main features of PSMs remodeling in patients with CLBP, and fat infiltration may exacerbate CLBP. It is, therefore, crucial to elucidate the relationship between fatty infiltration of PSMs and IVDD.

With the development of medical imaging modalities, the metrics for assessing the atrophy of PSMs have gradually become diverse. Earlier, the degree of atrophy of PSMs was mainly determined by measuring the cross-sectional area (CSA) of PSMs using computed tomography (CT) or real-time ultrasound. In 1994, Goutallier et al. [\(15](#page-6-0)) proposed a semiquantitative assessment of fatty infiltration in PSMs based on CT images, which opened new doors to exploring the mechanisms of IVDD. Using CT images, researchers found that fatty infiltration in PSMs was associated with small joint degeneration, lumbar spondylolisthesis, and narrowing of the vertebral space ([16](#page-6-0), [17](#page-6-0)). The degree of fatty infiltration in PSMs is significantly increased in patients with higher degrees of small

joint degeneration [\(16](#page-6-0)–[19\)](#page-6-0). With the advent of high-resolution magnetic resonance imaging (MRI), MRI techniques have become the primary technique for assessing the atrophy of PSMs. Earlier magnetic resonance techniques used axial T2 weighted scans more often. In recent years, MRI has been able to distinguish well between muscle and adipose tissue by threshold segmentation techniques to assess the degree of atrophy of PSMs better. Eksi et al. ([20](#page-6-0)) proposed a new scoring system that included fatty infiltration in PSMs, Modic changes (MCs), and IVDD. Patients with more intense LBP had a more degenerative spine ([20](#page-6-0)). However, this scoring system did not detail the role fat infiltration of PSMs played in LPB.

In 2015, Teichtahl et al. ([21\)](#page-6-0) used the iterative decomposition of water and fat with echo asymmetry and least square estimation-iron quantification (IDEAL-IQ) technique to quantify the fat content of PSMs and to assess the correlation between the fat content of PSMs and IVDD. They found that the fat content of PSMs was associated with reduced disc height. In addition, in 2016, the team found IVDD in all lumbar spine segments was associated with high-fat content in PSMs [\(22\)](#page-6-0). A more recent study analyzed the correlation between fatty infiltration in different PSMs and IVDD in more detail using the Pfirmann classification [\(23\)](#page-6-0) to assess the degree of IVDD. The study showed a strong positive correlation between Pfirmann classification and fat infiltration in the multifidus muscle (Rho=0.57, p<0.001) and a moderate positive correlation with fat infiltration in the erector spinae (Rho=0.49, p<0.001) and psoas major (Rho=0.31, p<0.001) [\(24\)](#page-6-0).

Multifidus

The multifidus is a general term for a group of PSMs that are shorter in cross-section but run almost the entire length of the spine and are, therefore, more susceptible to pathological changes. The multifidus is lateral to the spinous process, covering the corresponding vertebral plate, and is more closely related to the vertebral plate and spinous process than the erector spinae. Sun et al. ([25\)](#page-6-0) found that atrophy of the multifidus was significantly and positively correlated with IVDD at the L3/L4 disc level compared to the other PSMs. The exact mechanism of muscle degeneration is unclear. Disuse and denervation are two main mechanisms often mentioned [\(26](#page-6-0)). Liu et al. [\(27](#page-6-0)) proposed two hypothetical models by studying 264 subjects. One was that degeneration of the multifidus caused lumbar instability, which exacerbated upper lumbar disc degeneration. The other was that a herniated lumbar disc compressed the nerve roots of the corresponding segment, resulting in post-denervated multifidus atrophy. Hodges et al. [\(28\)](#page-6-0) and Goubert et al. ([29](#page-6-0)) suggested the reduction in multifidus activity due to pain as the leading cause of wasting muscle atrophy. Studies have shown that fatty infiltration in PSMs is strongly associated with high-intensity pain or disability

and structural abnormalities of the lumbar spine [\(30\)](#page-6-0). However, a study of patients with high-intensity pain and disability, which excluded the effect of physical activity level by adjusting for bias in the results, demonstrated that fatty infiltration in multifidus was an independent influence factor on the degree of disc degeneration [\(22](#page-6-0)). The facts about the fatty infiltration in multifidus during disc degeneration are clear. But the opposite conclusion is shown in studies targeting the muscle's CSA. Faur et al. [\(26\)](#page-6-0) reported that multifidus degeneration occurs mainly in the cross-section of MRI scans. However, the more common view is that muscle CSA does not correlate with IVDD ([24](#page-6-0), [31](#page-6-0)). To improve the bias caused by muscle CSA in individual body size, Urrutia et al. [\(32\)](#page-6-0) calculated the relative CSA (RCSA) by dividing the CSA of the L3 vertebrae by the muscle CSA and showed a stronger correlation between fat signal fraction and IVDD. In addition, muscle symmetry became a perspective that was looked at. It has been suggested that a 10% or more significant asymmetry in the multifidus' CSA be used to indicate potential spinal abnormalities [\(33\)](#page-6-0). However, it was found that more than 10% of men with no history of LBP also had asymmetry of PSMs ([34\)](#page-6-0) and that asymmetry of muscle CSA was not associated with lumbar disc herniation ([24](#page-6-0)). The Atrophy of PSMs is seen mainly in the inner side and deep layers of cross-sectional scans on MRI of the lumbar spine ([26](#page-6-0)). Thus, measuring the CSA of PSMs and the ratio of functional CSA to CSA to assess the degree of fatty infiltration in PSMs can be biased by individual measurement differences. In addition, it may also produce inconsistent results for different lumbar spine segments. Some investigators have suggested that fatty infiltration in PSMs correlates more strongly with pathological changes in the intervertebral disc than muscle CSA ([35](#page-6-0)).

Sarcopenia is defined as systemic muscle mass loss and a decline in physical performance ([36](#page-7-0)), of which back muscle atrophy or fat infiltration may be a component ([37](#page-7-0)). A study has shown that systemic muscle mass loss substantially impacts back muscle atrophy and fatty infiltration more than disc degeneration [\(37\)](#page-7-0). In other words, the effect of age and gender on systemic muscle mass can further affect back muscle atrophy and fat infiltration. Another study using CT techniques to analyze trunk muscles showed that, in addition to the multifidus, fatty infiltration in the gluteus maximus and transversus abdominis muscles was also associated with IVDD ([17\)](#page-6-0). Still, the exact mechanisms involved need to be further explored.

Significant results have been obtained in cross-sectional studies, and experimental animal studies under univariate control are essential. By quantitative MRI, Huang et al. ([38](#page-7-0)) assessed fat infiltration in PSMs of patients with discogenic LBP and rats in a novel discogenic LBP model. It was found that fatty infiltration was present in the PSMs of both LBP patients and rats and that there was a causal relationship between fatty infiltration and IVDD ([38\)](#page-7-0). Another study showed that dogs with higher IVDD grades had less fat infiltration in psoas and

multifidus than those with lower mean IVDD degrees [\(39](#page-7-0)). From this, the authors speculated that the presence or severity of IVDD was not uniquely associated with fat infiltration in these muscles. A study using a porcine model showed that disc and nerve root injury might lead to a CSA reduction of the multifidus and its fatty infiltration [\(28\)](#page-6-0). However, there was no atrophy of the multifidus following disc injury in sheep [\(40](#page-7-0)). This evidence challenges whether IVDD affects the characteristics of PSMs, but we are more skeptical that their relationship may not be purely causal. Özcan-Ekşi et al. ([30](#page-6-0)) found that fatty infiltration in multifidus increased the probability of severe LBP fourfold. Patients with severe lumbar disc herniation were likelier to have increased fatty infiltration of the multifidus and erector spinae muscles [\(12\)](#page-6-0). Therefore, further investigation is needed to determine whether lack of muscle strength and poor control due to fatty infiltration in the multifidus is the cause of LPB or vice versa.

Erector spinae

Although the role of the erector spinae in the spine's biomechanics is uncertain, its primary function is to be responsible for the flexion movement of the spine and, together with the multifidus, to maintain the stability of the lumbar spine. A cross-sectional study with Japanese subjects showed that fatty infiltration in PSMs correlated with age, and fatty infiltration of the upper lumbar erector spinae was significantly associated with LBP [\(41](#page-7-0)). In a separate study, the proton density fat fraction (PDFF) of multifidus and erector spinae at both L4/5 and L5/S1 levels was explored by MR techniques and analyzed the correlation with IVDD. The results showed a significant correlation between the PDFF of the PSMs and the degree of IVDD ([42](#page-7-0)). This correlation also confirms that the two are mutually reinforcing processes, i.e., disc degeneration can also lead to further atrophy of the erector spinae by destabilizing the spine. However, a study on erector spinae in adolescents showed that the more intense the patient's LBP, the less fatty infiltration in erector spinae ([43](#page-7-0)). The investigators suggest that this may be an automatic compensatory mechanism for the lumbar spine during the development of LBP in adolescents and children ([43\)](#page-7-0).

Psoas major

The psoas major is an essential flexor muscle of the spine and is the primary connection between the trunk and lower limbs. It contributes to the extension and general stability of the lumbar spine ([44](#page-7-0)). Animal studies point to significant differences in the psoas major in comparing different degrees of disc degeneration ([39](#page-7-0)). But it has also been shown that fatty infiltration in erector spinae and multifidus was significantly

associated with IVDD, whereas in psoas major was not significantly associated with IVDD ([12\)](#page-6-0). A study explicitly analyzing degenerative changes in the psoas major and lumbar spine showed that degenerative changes in the lumbar spine, including MCs, do not alter the activity of the psoas major ([45](#page-7-0)). The CSA of the psoas major at the L3/L4 and L4/L5 discs is even greater in patients with LBP compared to the healthy population. The result is inconsistent with the results of previous animal experiments. In addition, Parkkola et al. ([46](#page-7-0)) found that patients with CLBP had smaller psoas major by comparison with volunteers.

In contrast, Danneels et al. [\(47\)](#page-7-0) showed no difference in CSA of the psoas major between patients with CLBP and healthy controls. Considering that the study by Danneels et al. chose subjects who did not undergo surgery, the author speculates that the difference in results may be related to increased activity of the psoas major during treatment such as surgery. It has been suggested that gender, age, and degree of disc degeneration are independently associated with the PSMs' fat signal fraction (FSF) ([32](#page-6-0)). However, only gender and age affect the FSF of the psoas major, and the degree of disc degeneration does not alter the degree of fat infiltration in the psoas major ([32\)](#page-6-0). A study conducted to overcome gender bias concluded that the psoas major becomes more active in female patients with pain to stabilize the lumbar spine due to significant fat infiltration in the multifidus as a compensatory mechanism [\(15\)](#page-6-0). Although gender is an essential factor influencing PSMs infiltration [\(42\)](#page-7-0), this does not affect the validity of the conclusions of the above study.

Vertebral endplate changes

MCs refer to MRI signal intensity changes in the spinal endplate and subendplate bone. The characteristics of MCs were systematically described by Modic in 1988, who concluded that MCs are caused by disc degeneration and that their pathological evolution is characterized by disc degeneration \rightarrow weakening or loss of endplate protection \rightarrow edema of the adjacent cancellous bone \rightarrow fatty infiltration of the vertebral body \rightarrow fibrosis and calcification [\(48\)](#page-7-0). They described three different Modic types (I, II, and III). Since then, mixed Modic lesions (I/II and II/III) have also been identified, which indirectly suggests that all Modic lesions can progress from one type to another ([48](#page-7-0)–[50\)](#page-7-0). Based on the results of previous studies, types I and II are the most common types of the lumbar spine, with the most common distribution at the L4-L5 or L5-S1 levels ([48](#page-7-0), [51\)](#page-7-0). Studies have concluded that Modic type II changes are less associated with LBP [\(51](#page-7-0)–[54](#page-7-0)). The current studies confirm that Modic type II changes are more common than type I changes ([48](#page-7-0), [49,](#page-7-0) [51,](#page-7-0) [55](#page-7-0)– [57](#page-7-0)) yet remain rare in individuals without degenerative lumbar disc disease [\(51,](#page-7-0) [58](#page-7-0), [59\)](#page-7-0).

MCs have previously been reported to occur mainly in the lower lumbar segments (L4-L5 and L5-S1) [\(60,](#page-7-0) [61](#page-7-0)). In a recent

study, Ekşi et al. [\(43\)](#page-7-0) found that MCs were predominantly seen at the L1-L4 level rather than the L4-S1 level and were more common in patients with severe IVDD than in those with mild to moderate IVDD. When analyzing this association on a levelby-level basis, the authors found that severe IVDD was significantly associated with MCs at the L1-L2 and L3-L4 disc levels [\(43](#page-7-0)). And multifidus' fatty infiltration in the L3-L4 and L4- L5 segments increased the risk of MCs in all lumbar parts by 8.3 fold and 9.1-fold, respectively ([43](#page-7-0)). An MRI study showed that fatty infiltration in PSMs was associated with reduced disc height and MCs [\(31](#page-6-0)). In addition, Patients with Modic type I or I/II changes had more fatty degeneration in the lumbar PSMs [\(62\)](#page-7-0). However, there is still considerable debate as to whether MCs precede lipoatrophy or occur after back pain.

Molecular mechanisms of fat infiltration in PSMs

The lumbar discs and the PSMs are not only adjacent but also interconnected at the molecular and metabolic levels. IVDD is characterized by a progressive decrease in the proteoglycan and water content of the nucleus pulposus and a loss of resistance to compressive loads [\(63](#page-7-0)). The above mechanism is one of many, so we have sorted out the possible underlying mechanisms.

Inflammation

Early views suggested that fatty infiltration compromised the mass of the PSMs because the adipose tissue was noncontractible ([64](#page-7-0), [65\)](#page-7-0). There are currently many hypotheses for the mechanism of the relationship between fatty infiltration of the PSMs and spinal disorders, such as loss of nerve ([28\)](#page-6-0), chronic disuse [\(66](#page-7-0)), and inflammation ([67](#page-7-0)). Inflammation, in particular, has been extensively studied. An experiment modeled in rats demonstrated that fatty infiltration in PSMs was closely associated with inflammation [\(38\)](#page-7-0). Inflammation contributes to the development of pain [\(68\)](#page-7-0) and may contribute to MCs [\(69\)](#page-7-0). Increased reactive oxygen species (ROS) production has been reported to be associated with the differentiation of preadipocytes to adipocytes and the accumulation of adipose tissue [\(70\)](#page-7-0). Thus, effectively mitigating cellular oxidative stress in an inflammatory environment would also block ROS-induced adipogenesis ([71\)](#page-7-0). In a study by James et al. [\(72](#page-7-0)), muscle and fat specimens were collected intraoperatively from patients with herniated discs, and gene expression was detected using a quantitative polymerase chain reaction, dividing the patients into a high-fat infiltration group and a low-fat infiltration group. The results showed high tumor necrosis factor (TNF) expression in the multifidus of subjects in the high-fat infiltration group. Another study addressing the mechanism showed that the expression levels of interleukin (IL)-1 β , IL-6, IL-8, nitric oxide

synthase-2 (NOS-2), and transforming growth factor (TGF)- β did not differ in severe IVDD compared to mild IVDD ([24](#page-6-0)). The expression of TNF in lumbar disc tissue was significantly higher in the severe degeneration group than in the mild degeneration group ([24\)](#page-6-0). During the inflammatory process, TNF possesses intense pro-inflammatory activity and is closely associated with various pathological processes in IVDD [\(73\)](#page-7-0). Some researchers have speculated that TNF may not only be a product of adipose tissue but also regulate adipogenesis [\(72\)](#page-7-0).

Fibroblasts and preadipocytes are found in the connective tissue surrounding muscle fibers and can differentiate in response to inflammation. Adipocytes also increase following sympathetic degeneration, which is likely to occur following nerve injury. On the other hand, the dramatic increase in deoxyribonucleic acid synthesis following injury leads to the secretion of pro-inflammatory cytokines, stimulating fibroblasts, preadipocytes, and muscle precursor cells, ultimately leading to adipocyte proliferation.

Histological analysis shows that patients with LBP primarily display degeneration of the multifidus muscle, which occurs in relation to elevated inflammation, fiber size, and the ratio of fat to connective tissue ([74\)](#page-7-0). In addition, it was found that degenerating muscles were predominantly composed of type I fibers with less vascularity [\(74](#page-7-0)). Although there was no concurrent sign of atrophy at the individual fiber level, inflammatory cell density and vascular density changed in different muscle groups. In particular, inflammatory cells were significantly increased in normal skeletal muscle cells in the subgroup with 10%-50% fat infiltration, which suggests that regeneration and degeneration were out of balance in that condition [\(74](#page-7-0)).

Obesity

Obesity is a pro-inflammatory state that releases cytokines such as TNF- α and IL-6. It is commonly believed that obesity is closely associated with MCs. Albert et al. suggest that it is not obesity but rather its resulting overweight that plays a vital role in the development of MCs [\(50](#page-7-0)). Two possible mechanisms explain this effect: 1) When the disc is stressed, matrix synthesis and proteoglycan content are reduced. The load-bearing capacity then gradually decreases. 2) IVDD or disc herniation can increase the shear forces on the vertebral endplates due to loss of the nucleus pulposus. The increased axial and torsional stresses may result in microfractures of the vertebral endplate.

LBP has been reported to be significantly associated with body mass index (BMI) ([75\)](#page-7-0). However, it has also been suggested that BMI is not associated with fatty infiltration in PSMs [\(24\)](#page-6-0). A study of fatty infiltration in PSMs showed no difference in pain scores between obese and non-obese patients [\(76\)](#page-7-0). Still, obese patients had more severe disc degeneration in the lower lumbar spine, possibly due to the increased load on the vertebral body caused by obesity [\(76\)](#page-7-0). Subcutaneous fat tissue thickness (SFTT) is a new

radiological index for assessing body fat percentage ([77](#page-7-0)). Recent studies have shown that SFTT at L1-L2 level was superior to BMI in predicting severe IVDD and MCs [\(77](#page-7-0), [78\)](#page-7-0). A zoological study showed that a high-calorie diet did not cause disc degeneration in the vertebrae of mice ([79](#page-8-0)). However, advanced glycation end products (AGEs) can lead to IVDD [\(80,](#page-8-0) [81\)](#page-8-0). The receptor for advanced glycation end-products (RAGE) deletion inhibits systemic pro-inflammatory cytokine activity. D'Erminio et al. ([79\)](#page-8-0) used the RAGE knockout (RAGE-KO) model to control inflammation. They found that the effect of RAGE-KO in improving IVDD was limited and gender-related, suggesting that obesity and other sources of inflammation leading to a biomechanical overload of the lumbar spine may also have an impact ([79\)](#page-8-0). Another study showed that diabetes, rather than obesity, reduced the glycosaminoglycan and water content of the discs, and IVDD was associated with increased vertebral endplate thickness, reduced endplate porosity, and increased levels of AGEs ([81\)](#page-8-0). Due to their reduced glycosaminoglycan and water content and higher AGEs levels, the discs from diabetic rats became stiffer and had less alteration during compression [\(81](#page-8-0)). These findings suggest that endplate sclerosis, increased oxidative stress, and AGE/RAGE-mediated interactions may explain the high incidence of IVDD in patients with type 2 diabetes [\(81](#page-8-0)). Cell culture studies have shown increased palmitic acid-induced apoptosis in nucleus pulposus cells and activation of caspases 3, 7, 9, and poly (ADP-ribose) polymerase (PARP) mainly through the mitogen-activated protein kinases (MAPK) pathway, particularly the extracellular-signal-regulated kinases (ERK) pathway [\(82](#page-8-0)). Most obese patients have abnormally high blood lipid levels, and hypertriglyceridemia can induce IVDD independent of age and BMI [\(82](#page-8-0)). The results do not exclude the possibility of additional direct mechanical influences in the process of disc degeneration in humans [\(82](#page-8-0)).

Conversion of hematopoietic bone marrow to fatty bone marrow

The intervertebral disc is an avascular tissue in the human body. Its nutritional supply depends on the transport of capillaries from the adjacent vertebrae. The study of Krug et al. showed that the conversion of hematopoietic bone marrow to fatty bone marrow impairs the supply of adequate nutrients to the disc cells and thus may accelerate disc degeneration ([83](#page-8-0)). The MRI quantitative analysis confirmed that in the early stages of IVDD, IVDD and bone marrow fat interacted to some extent, with the severity of lumbar disc degeneration increasing with the adjacent vertebral fatty conversion ([84](#page-8-0)). The relationship was particularly evident in the L4/5 lumbar segment [\(84\)](#page-8-0). Focal fat conversion in normal hematopoietic red bone marrow may impede the transport of nutrients from the bone marrow to the end plate ([85\)](#page-8-0). IVDD is usually accompanied by osteoporosis, suggesting that the development of osteoporosis and IVDD may be a concomitant

process ([86](#page-8-0), [87\)](#page-8-0). Adipocytes and osteoblasts are derived from bone marrow mesenchymal stem cells (BMSCs). In BMSCs, there is a balance between osteogenesis and lipogenesis. If this balance is disturbed, it leads to a physiological disturbance, i.e., an increase in adipocytes in the bone marrow and decreased bone formation ([79\)](#page-8-0). Focal fatty degeneration of the bone marrow near the disc endplates can lead to disc degeneration by impeding the transport and metabolic exchange of nutrients essential to the disc. In addition, adipocyte growth and inflammatory edema compress the blood vessels in the confined bone cavity, further reducing blood flow ([49,](#page-7-0) [88\)](#page-8-0).

Adipokines

Adipose tissue releases pro-inflammatory cytokines that have a potential role in various tissue pathologies. Cytokines such as leptin, adiponectin, and TNF produced by adipocytes have been shown to be associated with obesity and osteoarthritis [\(89\)](#page-8-0).

Leptin regulates adipose tissue metabolism and inflammation ([90\)](#page-8-0) and can lead to adipocyte hypertrophy [\(91](#page-8-0)). Leptin and TNF are components of a positive feedback loop that promotes adipocyte hypertrophy ([90\)](#page-8-0). This cascade response could explain the rapid deterioration of adipose infiltration over time. Segar et al. [\(92](#page-8-0)) found that leptin acting alone or in concert with TNF- α , IL-1 β , or IL-6 in the nucleus pulposus significantly increased nitric oxide (NO) production and promoted inflammatory cytokines and matrix metalloproteinases (MMP). These processes further initiate the degradation of disc cells and the inflammatory cascade response, thereby accelerating the degenerative process [\(92\)](#page-8-0). Meanwhile, a study by Han et al. ([93\)](#page-8-0) confirmed that leptin expression was associated with the calcification of the cartilage endplates.

Adiponectin, mainly produced by lipids, is downregulated in patients with disc degeneration [\(94](#page-8-0)). Adiponectin may play an antiinflammatory role in maintaining the homeostasis of the degenerating disc environment by down-regulating TNF- α production by degenerating nucleus pulposus cells ([94](#page-8-0)). And adiponectin can reduce TNF- α and IL-6 significantly upregulated by IL-1 β stimulation in nucleus pulposus cells and annulus fibrosus cells ([95\)](#page-8-0). James et al. [\(72](#page-7-0)) found increased expression of lipocalin and NOS-2 in epidural fat. And high leptin and low arginase 1 expressions were found in the intramuscular and subcutaneous adipose tissues ([72\)](#page-7-0). They speculated that disc disease is associated with a dysregulation of the local inflammatory condition ([72](#page-7-0)).

Resistin is commonly involved in intra-articular angiogenesis and the inflammatory milieu [\(96,](#page-8-0) [97\)](#page-8-0). Resistin expression is upregulated in degenerating disc tissue. In nucleus pulposus cells, it binds to Toll-like receptor 4 via the p38-MAPK and NF-KB signaling pathways, leading to inflammation [\(98](#page-8-0)), further leading to metabolic disturbances in nucleus pulposus cells, and accelerated disc degeneration processes [\(99](#page-8-0)).

Visfatin is secreted by visceral adipocytes and is involved in immunity, stress, and inflammation processes. In degenerated

disc tissue, visfatin expression levels were progressively upregulated as degeneration progressed ([100\)](#page-8-0). In the nucleus pulposus cells, increased visfatin expression was associated with an upregulation of degradation-related proteins ([100](#page-8-0)). In contrast, the knockdown of visfatin expression or the use of inhibitors showed a decrease in cellular autophagy and a downregulation of autophagy-related protein expression ([100\)](#page-8-0). Similarly, a study modeled in rats to simulate severe IVDD and performed pathway analysis indicated that inhibition of visfatin protected the nucleus pulposus from degeneration and that focusing on epidural lipids and visfatin would be a potential therapeutic target to control the inflammation associated with IVDD [\(101](#page-8-0)).

Conclusion

IVDD is the leading cause of CLBP. IVDD is a chronic, multifactorial, irreversible process that severely compromises spinal stability and disc shock absorption. The PSMs are fundamental determinants of the structural stability and function of the lumbar spine. Studies have confirmed that fatty infiltration in PSMs plays a crucial role in IVDD. Inflammation, obesity, conversion of hematopoietic bone marrow to fatty bone marrow, and adipokines may be potential mechanisms for fat infiltration in PSMs. However, the quantitative methods and determination criteria for fat infiltration in PSMs and the vertebral plate need to be further studied. The biochemical and molecular mechanisms of fat infiltration in IVDD remain to be further investigated. The communication between the two at the molecular level still needs to be confirmed, especially concerning the potential signaling pathways in adipocytokines in IVDD.

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All authors listed have made a substantial, direct, and intellectual contribution to the work and approved it for publication.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

1. Chen SM, Liu MF, Cook J, Bass S, Lo SK. Sedentary lifestyle as a risk factor for low back pain: a systematic review. Int Arch Occup Environ Health (2009) 82 (7):797–806. doi: [10.1007/s00420-009-0410-0](https://doi.org/10.1007/s00420-009-0410-0)

2. Maher C, Underwood M, Buchbinder R. Non-specific low back pain. Lancet (2017) 389(10070):736–47. doi: [10.1016/S0140-6736\(16\)30970-9](https://doi.org/10.1016/S0140-6736(16)30970-9)

3. Vergroesen PP, Kingma I, Emanuel KS, Hoogendoorn RJ, Welting TJ, van Royen BJ, et al. Mechanics and biology in intervertebral disc degeneration: a vicious circle. Osteoarthritis Cartilage (2015) 23(7):1057–70. doi: [10.1016/](https://doi.org/10.1016/j.joca.2015.03.028) [j.joca.2015.03.028](https://doi.org/10.1016/j.joca.2015.03.028)

4. Modic MT, Ross JS. Lumbar degenerative disk disease. Radiology (2007) 245 (1):43–61. doi: [10.1148/radiol.2451051706](https://doi.org/10.1148/radiol.2451051706)

5. Hodges PW, Danneels L. Changes in structure and function of the back muscles in low back pain: different time points, observations, and mechanisms. J Orthop Sports Phys Ther (2019) 49(6):464–76. doi: [10.2519/jospt.2019.8827](https://doi.org/10.2519/jospt.2019.8827)

6. Geisser ME, Ranavaya M, Haig AJ, Roth RS, Zucker R, Ambroz C, et al. A meta-analytic review of surface electromyography among persons with low back pain and normal, healthy controls. *J Pain* (2005) 6(11):711-26. doi: [10.1016/](https://doi.org/10.1016/j.jpain.2005.06.008) [j.jpain.2005.06.008](https://doi.org/10.1016/j.jpain.2005.06.008)

7. Solomonow M, Hatipkarasulu S, Zhou BH, Baratta RV, Aghazadeh F. Biomechanics and electromyography of a common idiopathic low back disorder. Spine (2003) 28(12):1235-48. doi: [10.1097/01.BRS.0000065568.47818.B9](https://doi.org/10.1097/01.BRS.0000065568.47818.B9)

8. Ranger TA, Cicuttini FM, Jensen TS, Heritier S, Urquhart DM. Paraspinal muscle cross-sectional area predicts low back disability but not pain intensity. Spine J (2019) 19(5):862–8. doi: [10.1016/j.spinee.2018.12.004](https://doi.org/10.1016/j.spinee.2018.12.004)

9. Agten A, Stevens S, Verbrugghe J, Timmermans A, Vandenabeele F. Biopsy samples from the erector spinae of persons with nonspecific chronic low back pain display a decrease in glycolytic muscle fibers. Spine J (2020) 20(2):199–206. doi: [10.1016/j.spinee.2019.09.023](https://doi.org/10.1016/j.spinee.2019.09.023)

10. Kalichman L, Carmeli E, Been E. The association between imaging parameters of the paraspinal muscles, spinal degeneration, and low back pain. BioMed Res Int (2017) 2017:2562957. doi: [10.1155/2017/2562957](https://doi.org/10.1155/2017/2562957)

11. Bailey JF, Fields AJ, Ballatori A, Cohen D, Jain D, Coughlin D, et al. The relationship between endplate pathology and patient-reported symptoms for chronic low back pain depends on lumbar paraspinal muscle quality. Spine (2019) 44(14):1010–7. doi: [10.1097/BRS.0000000000003035](https://doi.org/10.1097/BRS.0000000000003035)

12. Özcan-Ekşi EE, Ekşi MŞ, Akçal MA. Severe lumbar intervertebral disc degeneration is associated with modic changes and fatty infiltration in the paraspinal muscles at all lumbar levels, except for L1-L2: a cross-sectional analysis of 50 symptomatic women and 50 age-matched symptomatic men. World Neurosurg (2019) 122:e1069–77. doi: [10.1016/j.wneu.2018.10.229](https://doi.org/10.1016/j.wneu.2018.10.229)

13. Goldspink DF. The influence of immobilization and stretch on protein turnover of rat skeletal muscle. J Physiol (1977) 264(1):267–82. doi: [10.1113/](https://doi.org/10.1113/jphysiol.1977.sp011667) [jphysiol.1977.sp011667](https://doi.org/10.1113/jphysiol.1977.sp011667)

14. Safran O, Derwin KA, Powell K, Iannotti JP. Changes in rotator cuff muscle volume, fat content, and passive mechanics after chronic detachment in a canine model. J Bone Joint Surg Am (2005) 87(12):2662–70. doi: [10.2106/JBJS.D.02421](https://doi.org/10.2106/JBJS.D.02421)

15. Goutallier D, Postel JM, Bernageau J, Lavau L, Voisin MC. Fatty muscle degeneration in cuff ruptures. pre- and postoperative evaluation by CT scan. *Clin*
Orthop Relat Res (1994) 304):78–83.

16. Kalichman L, Klindukhov A, Li L, Linov L. Indices of paraspinal muscles degeneration: reliability and association with facet joint osteoarthritis: feasibility study. Clin Spine Surg (2016) 29(9):465–70. doi: [10.1097/BSD.0b013e31828be943](https://doi.org/10.1097/BSD.0b013e31828be943)

17. Sebro R, O'Brien L, Torriani M, Bredella MA. Assessment of trunk muscle density using CT and its association with degenerative disc and facet joint disease
of the lumbar spine. Sk*eletal Radiol* (2016) 45(9):1221–6. doi: [10.1007/s00256-016-](https://doi.org/10.1007/s00256-016-2405-8) [2405-8](https://doi.org/10.1007/s00256-016-2405-8)

18. Kalichman L, Hodges P, Li L, Guermazi A, Hunter DJ. Changes in paraspinal muscles and their association with low back pain and spinal degeneration: CT study. Eur Spine J (2010) 9(7):1136–44. doi: [10.1007/s00586-](https://doi.org/10.1007/s00586-009-1257-5) [009-1257-5](https://doi.org/10.1007/s00586-009-1257-5)

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19. Kalichman L, Kim DH, Li L, Guermazi A, Hunter DJ. Computed tomography-evaluated features of spinal degeneration: prevalence, intercorrelation, and association with self-reported low back pain. Spine J (2010) 10(3):200–8. doi: [10.1016/j.spinee.2009.10.018](https://doi.org/10.1016/j.spinee.2009.10.018)

20. Ekşi MŞ, Özcan-Ekşi EE, Orhun Ö, Turgut VU, Pamir MN. Proposal for a new scoring system for spinal degeneration: Mo-Fi-Disc. Clin Neurol Neurosurg (2020) 198:106120. doi: [10.1016/j.clineuro.2020.106120](https://doi.org/10.1016/j.clineuro.2020.106120)

21. Teichtahl AJ, Urquhart DM, Wang Y, Wluka AE, O'Sullivan R, Jones G, et al. Physical inactivity is associated with narrower lumbar intervertebral discs, high fat content of paraspinal muscles and low back pain and disability. Arthritis Res Ther (2015) 17(1):114. doi: [10.1186/s13075-015-0629-y](https://doi.org/10.1186/s13075-015-0629-y)

22. Teichtahl AJ, Urquhart DM, Wang Y, Wluka AE, O'Sullivan R, Jones G, et al. Lumbar disc degeneration is associated with modic change and high paraspinal fat content - a 3.0T magnetic resonance imaging study. BMC Musculoskelet Disord (2016) 17(1):439. doi: [10.1186/s12891-016-1297-z](https://doi.org/10.1186/s12891-016-1297-z)

23. Pfirrmann CW, Metzdorf A, Zanetti M, Hodler J, Boos N. Magnetic resonance classification of lumbar intervertebral disc degeneration. Spine (2001) 26(17):1873–8. doi: [10.1097/00007632-200109010-00011](https://doi.org/10.1097/00007632-200109010-00011)

24. Shi L, Yan B, Jiao Y, Chen Z, Zheng Y, Lin Y, et al. Correlation between the fatty infiltration of paraspinal muscles and disc degeneration and the underlying mechanism. BMC Musculoskelet Disord (2022) 23(1):509. doi: [10.1186/s12891-](https://doi.org/10.1186/s12891-022-05466-8) [022-05466-8](https://doi.org/10.1186/s12891-022-05466-8)

25. Sun D, Liu P, Cheng J, Ma Z, Liu J, Qin T. Correlation between intervertebral disc degeneration, paraspinal muscle atrophy, and lumbar facet joints degeneration in patients with lumbar disc herniation. BMC Musculoskelet Disord (2017) 18(1):167. doi: [10.1186/s12891-017-1522-4](https://doi.org/10.1186/s12891-017-1522-4)

26. Faur C, Patrascu JM, Haragus H, Anglitoiu B. Correlation between multifidus fatty atrophy and lumbar disc degeneration in low back pain. BMC Musculoskelet Disord (2019) 20(1):414. doi: [10.1186/s12891-019-2786-7](https://doi.org/10.1186/s12891-019-2786-7)

27. Liu C, Xue J, Liu J, Ma G, Moro A, Liang T, et al. Is there a correlation between upper lumbar disc herniation and multifidus muscle degeneration? a retrospective study of MRI morphology. BMC Musculoskelet Disord (2021) 22 (1):92. doi: [10.1186/s12891-021-03970-x](https://doi.org/10.1186/s12891-021-03970-x)

28. Hodges P, Holm AK, Hansson T, Holm S. Rapid atrophy of the lumbar multifidus follows experimental disc or nerve root injury. Spine (Phila Pa 1976) (2006) 31(25):2926–33. doi: [10.1097/01.brs.0000248453.51165.0b](https://doi.org/10.1097/01.brs.0000248453.51165.0b)

29. Goubert D, De Pauw R, Meeus M, Willems T, Cagnie B, Schouppe S, et al. Lumbar muscle structure and function in chronic versus recurrent low back pain: a cross-sectional study. Spine J (2017) 17(9):1285–96. doi: [10.1016/j.spinee.2017.04.025](https://doi.org/10.1016/j.spinee.2017.04.025)

30. Özcan-Ekşi EE, Ekşi MŞ, Turgut VU, Canbolat Ç, Pamir MN. Reciprocal relationship between multifidus and psoas at L4-L5 level in women with low back pain. Br J Neurosurg (2021) 35(2):220–8. doi: [10.1080/02688697.2020.1783434](https://doi.org/10.1080/02688697.2020.1783434)

31. Teichtahl AJ, Urquhart DM, Wang Y, Wluka AE, Wijethilake P, O'Sullivan R, et al. Fat infiltration of paraspinal muscles is associated with low back pain, disability, and structural abnormalities in community-based adults. Spine J (2015) 15(7):1593–601. doi: [10.1016/j.spinee.2015.03.039](https://doi.org/10.1016/j.spinee.2015.03.039)

32. Urrutia J, Besa P, Lobos D, Campos M, Arrieta C, Andia M, et al. Lumbar paraspinal muscle fat infiltration is independently associated with sex, age, and inter-vertebral disc degeneration in symptomatic patients. Skeletal Radiol (2018) 47 (7):955–61. doi: [10.1007/s00256-018-2880-1](https://doi.org/10.1007/s00256-018-2880-1)

33. Hides J, Gilmore C, Stanton W, Bohlscheid E. Multifidus size and symmetry among chronic LBP and healthy asymptomatic subjects. Man Ther (2008) 13 (1):43–9. doi: [10.1016/j.math.2006.07.017](https://doi.org/10.1016/j.math.2006.07.017)

34. Niemeläinen R, Briand MM, Battié MC. Substantial asymmetry in paraspinal muscle cross-sectional area in healthy adults questions its value as a marker of low back pain and pathology. Spine (Phila Pa 1976) (2011) 36(25):2152– 7. doi: [10.1097/BRS.0b013e318204b05a](https://doi.org/10.1097/BRS.0b013e318204b05a)

35. Fortin M, Lazáry À, Varga PP, McCall I, Battié MC. Paraspinal muscle asymmetry and fat infiltration in patients with symptomatic disc herniation. Eur
Spine J (2016) 25(5):1452–9. doi: [10.1007/s00586-016-4503-7](https://doi.org/10.1007/s00586-016-4503-7)

36. Cruz-Jentoft AJ, Bahat G, Bauer J, Boirie Y, Bruyère O, Cederholm T, et al. Sarcopenia: revised European consensus on definition and diagnosis. Age Ageing (2019) 48(1):16–31. doi: [10.1093/ageing/afy169](https://doi.org/10.1093/ageing/afy169)

37. Lee D, Kang M. Correlation between psoas muscle index and degeneration of spinal back muscle in patients with back pain. Healthcare (Basel) (2021) 9 (9):1189. doi: [10.3390/healthcare9091189](https://doi.org/10.3390/healthcare9091189)

38. Huang Y, Wang L, Luo B, Yang K, Zeng X, Chen J, et al. Associations of lumber disc degeneration with paraspinal muscles myosteatosis in discogenic low back pain. Front Endocrinol (Lausanne) (2022) 13:891088. doi: [10.3389/](https://doi.org/10.3389/fendo.2022.891088) [fendo.2022.891088](https://doi.org/10.3389/fendo.2022.891088)

39. Lerer A, Nykamp SG, Harriss AB, Gibson TW, Koch TG, Brown SH. MRI-Based relationships between spine pathology, intervertebral disc degeneration, and muscle fatty infiltration in chondrodystrophic and non-chondrodystrophic dogs. Spine J (2015) 15(11):2433-9. doi: [10.1016/j.spinee.2015.08.014](https://doi.org/10.1016/j.spinee.2015.08.014)

40. Hodges PW, James G, Blomster L, Hall L, Schmid A, Shu C, et al. Multifidus muscle changes after back injury are characterized by structural remodeling of muscle, adipose and connective tissue, but not muscle atrophy: molecular and morphological evidence. Spine (2015) 40(14):1057–71. doi: [10.1097/](https://doi.org/10.1097/BRS.0000000000000972) [BRS.0000000000000972](https://doi.org/10.1097/BRS.0000000000000972)

41. Sasaki T, Yoshimura N, Hashizume H, Yamada H, Oka H, Matsudaira K, et al. MRI-Defined paraspinal muscle morphology in Japanese population: the wakayama spine study. PloS One (2017) 12(11):e0187765. doi: [10.1371/](https://doi.org/10.1371/journal.pone.0187765) [journal.pone.0187765](https://doi.org/10.1371/journal.pone.0187765)

42. Huang Y, Wang L, Zeng X, Chen J, Zhang Z, Jiang Y, et al. Association of paraspinal muscle CSA and PDFF measurements with lumbar intervertebral disk degeneration in patients with chronic low back pain. Front Endocrinol (Lausanne) (2022) 13:792819. doi: [10.3389/fendo.2022.792819](https://doi.org/10.3389/fendo.2022.792819)

43. Ekşi MŞ, Özcan-Ekşi EE, Özmen BB, Turgut VU, Huet SE, Dinç T, et al. Lumbar intervertebral disc degeneration, end-plates and paraspinal muscle changes in children and adolescents with low-back pain. *J Pediatr Orthop B* (2022) 31(1):93–102. doi: [10.1097/BPB.0000000000000833](https://doi.org/10.1097/BPB.0000000000000833)

44. Nachemson A. The possible importance of the psoas muscle for stabilization of the lumbar spine. Acta Orthop Scand (1968) 39(1):47–57. doi: [10.3109/](https://doi.org/10.3109/17453676808989438) [17453676808989438](https://doi.org/10.3109/17453676808989438)

45. Arbanas J, Pavlovic I, Marijancic V, Vlahovic H, Starcevic-Klasan G, Peharec S, et al. MRI Features of the psoas major muscle in patients with low back pain. Eur Spine J (2013) 22(9):1965–71. doi: [10.1007/s00586-013-2749-x](https://doi.org/10.1007/s00586-013-2749-x)

46. Parkkola R, Rytökoski U, Kormano M. Magnetic resonance imaging of the discs and trunk muscles in patients with chronic low back pain and healthy control subjects. Spine (Phila Pa 1976) (1993) 18(7):830–6. doi: [10.1097/00007632-199306000-00004](https://doi.org/10.1097/00007632-199306000-00004)

47. Danneels LA, Vanderstraeten GG, Cambier DC, Witvrouw EE, De Cuyper HJ. CT imaging of trunk muscles in chronic low back pain patients and healthy control subjects. Eur Spine J (2000) 9(4):266–72. doi: [10.1007/s005860000190](https://doi.org/10.1007/s005860000190)

48. Modic MT, Steinberg PM, Ross JS, Masaryk TJ, Carter JR. Degenerative disk disease: assessment of changes in vertebral body marrow with MR imaging. Radiology (1988) 166(1 Pt 1):193–9. doi: [10.1148/radiology.166.1.3336678](https://doi.org/10.1148/radiology.166.1.3336678)

49. de Roos A, Kressel H, Spritzer C, Dalinka M. MR imaging of marrow changes adjacent to end plates in degenerative lumbar disk disease. AJR Am J Roentgenol (1987) 149(3):531–4. doi: [10.2214/ajr.149.3.531](https://doi.org/10.2214/ajr.149.3.531)

50. Albert HB, Kjaer P, Jensen TS, Sorensen JS, Bendix T, Manniche C. Modic changes, possible causes and relation to low back pain. Med Hypotheses (2008) 70 (2):361–8. doi: [10.1016/j.mehy.2007.05.014](https://doi.org/10.1016/j.mehy.2007.05.014)

51. Kuisma M, Karppinen J, Niinimäki J, Ojala R, Haapea M, Heliövaara M, et al. Modic changes in endplates of lumbar vertebral bodies: prevalence and association with low back and sciatic pain among middle-aged male workers. Spine
(2007) 32(10):1116–22. doi: [10.1097/01.brs.0000261561.12944.ff](https://doi.org/10.1097/01.brs.0000261561.12944.ff)

52. Toyone T, Takahashi K, Kitahara H, Yamagata M, Murakami M, Moriya H. Vertebral bone-marrow changes in degenerative lumbar disc disease. an MRI study of 74 patients with low back pain. J Bone Joint Surg Br (1994) 76(5):757–64.

53. Mitra D, Cassar-Pullicino VN, McCall IW. Longitudinal study of vertebral type-1 end-plate changes on MR of the lumbar spine. Eur Radiol (2004) 14 (9):1574–81. doi: [10.1007/s00330-004-2314-4](https://doi.org/10.1007/s00330-004-2314-4)

54. Albert HB, Manniche C. Modic changes following lumbar disc herniation. Eur Spine J (2007) 16(7):977–82. doi: [10.1007/s00586-007-0336-8](https://doi.org/10.1007/s00586-007-0336-8)

55. Modic MT, Masaryk TJ, Ross JS, Carter JR. Imaging of degenerative disk disease. Radiology (1988) 168(1):177–86. doi: [10.1148/radiology.168.1.3289089](https://doi.org/10.1148/radiology.168.1.3289089)

56. Schmid G, Witteler A, Willburger R, Kuhnen C, Jergas M, Koester O. Lumbar disk herniation: correlation of histologic findings with marrow signal intensity changes in vertebral endplates at MR imaging. Radiology (2004) 231 (2):352–8. doi: [10.1148/radiol.2312021708](https://doi.org/10.1148/radiol.2312021708)

57. Karchevsky M, Schweitzer ME, Carrino JA, Zoga A, Montgomery D, Parker L. Reactive endplate marrow changes: a systematic morphologic and epidemiologic evaluation. Skeletal Radiol (2005) 34(3):125–9. doi: [10.1007/s00256-004-0886-3](https://doi.org/10.1007/s00256-004-0886-3)

58. Weishaupt D, Zanetti M, Hodler J, Boos N. MR imaging of the lumbar spine: prevalence of intervertebral disk extrusion and sequestration, nerve root compression, end plate abnormalities, and osteoarthritis of the facet joints in asymptomatic volunteers. Radiology (1998) 209(3):661–6. doi: [10.1148/](https://doi.org/10.1148/radiology.209.3.9844656) [radiology.209.3.9844656](https://doi.org/10.1148/radiology.209.3.9844656)

59. Kjaer P, Korsholm L, Bendix T, Sorensen JS, Leboeuf-Yde C. Modic changes and their associations with clinical findings. Eur Spine J (2006) 15(9):1312–9. doi: [10.1007/s00586-006-0185-x](https://doi.org/10.1007/s00586-006-0185-x)

60. Tarukado K, Ono T, Tono O, Tanaka H, Ikuta K, Harimaya K, et al. Does modic change progresss with age? Spine (2017) 42(23):1805–9. doi: [10.1097/](https://doi.org/10.1097/BRS.0000000000002254) [BRS.0000000000002254](https://doi.org/10.1097/BRS.0000000000002254)

61. Mok FP, Samartzis D, Karppinen J, Fong DY, Luk KD, Cheung KM. Modic changes of the lumbar spine: prevalence, risk factors, and association with disc degeneration and low back pain in a large-scale population-based cohort. Spine J (2016) 16(1):32–41. doi: [10.1016/j.spinee.2015.09.060](https://doi.org/10.1016/j.spinee.2015.09.060)

62. Atci IB, Yilmaz H, Samanci MY, Atci AG, Karagoz Y. The prevalence of lumbar paraspinal muscle fatty degeneration in patients with modic type I and I/II end plate changes. Asian Spine J (2020) 14(2):185–91. doi: [10.31616/asj.2018.0333](https://doi.org/10.31616/asj.2018.0333)

63. Smith LJ, Nerurkar NL, Choi KS, Harfe BD, Elliott DM. Degeneration and regeneration of the intervertebral disc: lessons from development. Dis Model Mech (2011) 4(1):31–41. doi: [10.1242/dmm.006403](https://doi.org/10.1242/dmm.006403)

64. MacDonald D, Moseley GL, Hodges PW. People with recurrent low back pain respond differently to trunk loading despite remission from symptoms. Spine (2010) 35(7):818–24. doi: [10.1097/BRS.0b013e3181bc98f1](https://doi.org/10.1097/BRS.0b013e3181bc98f1)

65. Macdonald DA, Dawson AP, Hodges PW. Behavior of the lumbar multifidus during lower extremity movements in people with recurrent low back pain during symptom remission. J Orthop Sports Phys Ther (2011) 41(3):155–64. doi: [10.2519/jospt.2011.3410](https://doi.org/10.2519/jospt.2011.3410)

66. Pagano AF, Brioche T, Arc-Chagnaud C, Demangel R, Chopard A, Py G. Short-term disuse promotes fatty acid infiltration into skeletal muscle. J Cachexia Sarcopenia Muscle (2018) 9(2):335–47. doi: [10.1002/jcsm.12259](https://doi.org/10.1002/jcsm.12259)

67. James G, Sluka KA, Blomster L, Hall L, Schmid AB, Shu CC, et al. Macrophage polarization contributes to local inflammation and structural change in the multifidus muscle after intervertebral disc injury. Eur Spine J (2018) 27(8):1744–56. doi: [10.1007/s00586-018-5652-7](https://doi.org/10.1007/s00586-018-5652-7)

68. Ray L, Lipton RB, Zimmerman ME, Katz MJ, Derby CA. Mechanisms of association between obesity and chronic pain in the elderly. Pain (2011) 152(1):53– 9. doi: [10.1016/j.pain.2010.08.043](https://doi.org/10.1016/j.pain.2010.08.043)

69. Ekşi MŞ, Kara M, Özcan-Ekşi EE, Aytar MH, Güngör A, Özgen S, et al. Is diabetes mellitus a risk factor for modic changes?: a novel model to understand the association between intervertebral disc degeneration and end-plate changes. J Orthop Sci (2020) 25(4):571–5. doi: [10.1016/j.jos.2019.09.005](https://doi.org/10.1016/j.jos.2019.09.005)

70. Furukawa S, Fujita T, Shimabukuro M, Iwaki M, Yamada Y, Nakajima Y, et al. Increased oxidative stress in obesity and its impact on metabolic syndrome. J Clin Invest (2004) 114(12):1752–61. doi: [10.1172/JCI21625](https://doi.org/10.1172/JCI21625)

71. Xiao L, Aoshima H, Saitoh Y, Miwa N. The effect of squalane-dissolved fullerene-C60 on adipogenesis-accompanied oxidative stress and macrophage activation in a preadipocyte-monocyte co-culture system. Biomaterials (2010) 31 (23):5976–85. doi: [10.1016/j.biomaterials.2010.04.032](https://doi.org/10.1016/j.biomaterials.2010.04.032)

72. James G, Chen X, Diwan A, Hodges PW. Fat infiltration in the multifidus muscle is related to inflammatory cytokine expression in the muscle and epidural adipose tissue in individuals undergoing surgery for intervertebral disc herniation. Eur Spine J (2021) 30(4):837–45. doi: [10.1007/s00586-020-06514-4](https://doi.org/10.1007/s00586-020-06514-4)

73. Wang Y, Che M, Xin J, Zheng Z, Li J, Zhang S. The role of IL-1 β and TNF- α in intervertebral disc degeneration. BioMed Pharmacother (2020) 131:110660. doi: [10.1016/j.biopha.2020.110660](https://doi.org/10.1016/j.biopha.2020.110660)

74. Shahidi B, Hubbard JC, Gibbons MC, Ruoss S, Zlomislic V, Allen RT, et al. Lumbar multifidus muscle degenerates in individuals with chronic degenerative lumbar spine pathology. J Orthop Res (2017) 35(12):2700–6. doi: [10.1002/jor.23597](https://doi.org/10.1002/jor.23597)

75. Goode A, Cook C, Brown C, Isaacs R, Roman M, Richardson W. Differences in comorbidities on low back pain and low back related leg pain. Pain Pract (2011) 11(1):42–7. doi: [10.1111/j.1533-2500.2010.00391.x](https://doi.org/10.1111/j.1533-2500.2010.00391.x)

76. Özcan-Ekşi EE, Turgut VU, Küçüksüleymanoğlu D, Ekşi MŞ. Obesity could be associated with poor paraspinal muscle quality at upper lumbar levels and
degenerated spine at lower lumbar levels: Is this a domino effect? J Clin Neurosci (2021) 94:120–7 doi: [10.1016/j.jocn.2021.10.005](https://doi.org/10.1016/j.jocn.2021.10.005)

77. Özcan-Ekşi EE, Kara M, Berikol G, Orhun Ö, Turgut VU, Ekşi MŞ. A new radiological index for the assessment of higher body fat status and lumbar spine degeneration. Skeletal Radiol (2022) 51(6):1261–71 doi: [10.1007/s00256-021-](https://doi.org/10.1007/s00256-021-03957-8) [03957-8](https://doi.org/10.1007/s00256-021-03957-8)

78. Berikol G, Ekşi MŞ, Aydın L, Börekci A, Özcan-Ekşi EE. Subcutaneous fat index: a reliable tool for lumbar spine studies. Eur Radiol (2022) 32(9):6504–13. doi: [10.1007/s00330-022-08775-7](https://doi.org/10.1007/s00330-022-08775-7)

79. D'Erminio DN, Krishnamoorthy D, Lai A, Hoy RC, Natelson DM, Poeran J, et al. High fat diet causes inferior vertebral structure and function without disc degeneration in RAGE-KO mice. J Orthop Res (2022) 40(7):1672–86. doi: [10.1002/jor.25191](https://doi.org/10.1002/jor.25191)

80. Illien-Jünger S, Palacio-Mancheno P, Kindschuh WF, Chen X, Sroga GE, Vashishth D, et al. Dietary advanced glycation end products have sex- and agedependent effects on vertebral bone microstructure and mechanical function in mice. J Bone Miner Res (2018) 33(3):437–48. doi: [10.1002/jbmr.3321](https://doi.org/10.1002/jbmr.3321)

81. Fields AJ, Berg-Johansen B, Metz LN, Miller S, La B, Liebenberg EC, et al. Alterations in intervertebral disc composition, matrix homeostasis and biomechanical behavior in the UCD-T2DM rat model of type 2 diabetes. J Orthop Res (2015) 33(5):738–46. doi: [10.1002/jor.22807](https://doi.org/10.1002/jor.22807)

82. Zhang X, Chen J, Huang B, Wang J, Shan Z, Liu J, et al. Obesity mediates apoptosis and extracellular matrix metabolic imbalances via MAPK pathway activation in intervertebral disk degeneration. Front Physiol (2019) 10:1284. doi: [10.3389/fphys.2019.01284](https://doi.org/10.3389/fphys.2019.01284)

83. Krug R, Joseph GB, Han M, Fields A, Cheung J, Mundada M, et al. Associations between vertebral body fat fraction and intervertebral disc biochemical composition as assessed by quantitative MRI. *J Magn Reson Imaging*
(2019) 50(4):1219–26. doi: [10.1002/jmri.26675](https://doi.org/10.1002/jmri.26675)

84. Ji Y, Hong W, Liu M, Liang Y, Deng Y, Ma L. Intervertebral disc degeneration associated with vertebral marrow fat, assessed using quantitative magnetic resonance imaging. Skeletal Radiol (2020) 49(11):1753–63. doi: [10.1007/](https://doi.org/10.1007/s00256-020-03419-7) [s00256-020-03419-7](https://doi.org/10.1007/s00256-020-03419-7)

85. Urban JP, Smith S, Fairbank JC. Nutrition of the intervertebral disc. Spine (2004) 29(23):2700–9. doi: [10.1097/01.brs.0000146499.97948.52](https://doi.org/10.1097/01.brs.0000146499.97948.52)

86. Castaño-Betancourt MC, Oei L, Rivadeneira F, de Schepper EI, Hofman A, Bierma-Zeinstra S, et al. Association of lumbar disc degeneration with osteoporotic fractures; the Rotterdam study and meta-analysis from systematic review. Bone (2013) 57(1):284–9. doi: [10.1016/j.bone.2013.08.004](https://doi.org/10.1016/j.bone.2013.08.004)

87. Fabreguet I, Fechtenbaum J, Briot K, Paternotte S, Roux C. Lumbar disc degeneration in osteoporotic men: prevalence and assessment of the relation with presence of vertebral fracture. J Rheumatol (2013) 40(7):1183–90. doi: [10.3899/](https://doi.org/10.3899/jrheum.120769) irheum.120769

88. Wang GJ, Sweet DE, Reger SI, Thompson RC. Fat-cell changes as a mechanism of avascular necrosis of the femoral head in cortisone-treated rabbits. *J Bone Joint Surg* Am (1977) 59(6):729–35. doi: [10.2106/00004623-197759060-00003](https://doi.org/10.2106/00004623-197759060-00003)

89. de Boer TN, van Spil WE, Huisman AM, Polak AA, Bijlsma JW, Lafeber FP, et al. Serum adipokines in osteoarthritis; comparison with controls and relationship with local parameters of synovial inflammation and cartilage damage. Osteoarthritis Cartilage (2012) 20(8):846–53. doi: [10.1016/j.joca.2012.05.002](https://doi.org/10.1016/j.joca.2012.05.002)

90. La Cava A. Leptin in inflammation and autoimmunity. Cytokine (2017) 98:51–8. doi: [10.1016/j.cyto.2016.10.011](https://doi.org/10.1016/j.cyto.2016.10.011)

91. Skurk T, Alberti-Huber C, Herder C, Hauner H. Relationship between adipocyte size and adipokine expression and secretion. J Clin Endocrinol Metab (2007) 92(3):1023–33. doi: [10.1210/jc.2006-1055](https://doi.org/10.1210/jc.2006-1055)

92. Segar AH, Fairbank JCT, Urban J. Leptin and the intervertebral disc: a biochemical link exists between obesity, intervertebral disc degeneration and low back pain-an in vitro study in a bovine model. Eur Spine J (2019) 28(2):214–23. doi: [10.1007/s00586-018-5778-7](https://doi.org/10.1007/s00586-018-5778-7)

93. Han YC, Ma B, Guo S, Yang M, Li LJ, Wang SJ, et al. Leptin regulates disc cartilage endplate degeneration and ossification through activation of the MAPK-ERK signalling pathway in vivo and in vitro. J Cell Mol Med (2018) 22(4):2098–109. doi: [10.1111/jcmm.13398](https://doi.org/10.1111/jcmm.13398)

94. Yuan B, Huang L, Yan M, Zhang S, Zhang Y, Jin B, et al. Adiponectin downregulates TNF- α expression in degenerated intervertebral discs. Spine (2018) 43(7):E381–9. doi: [10.1097/BRS.0000000000002364](https://doi.org/10.1097/BRS.0000000000002364)

95. Terashima Y, Kakutani K, Yurube T, Takada T, Maeno K, Hirata H, et al. Expression of adiponectin receptors in human and rat intervertebral disc cells and changes in receptor expression during disc degeneration using a rat tail temporary static compression model. J Orthop Surg Res (2016) 11(1):147. doi: [10.1186/s13018-](https://doi.org/10.1186/s13018-016-0481-z) $0.16 - 0.481 - 2$

96. Jamaluddin MS, Weakley SM, Yao Q, Chen C. Resistin: functional roles and therapeutic considerations for cardiovascular disease. Br J Pharmacol (2012) 165 (3):622–32. doi: [10.1111/j.1476-5381.2011.01369.x](https://doi.org/10.1111/j.1476-5381.2011.01369.x)

97. Fang WQ, Zhang Q, Peng YB, Chen M, Lin XP, Wu JH, et al. Resistin level is positively correlated with thrombotic complications in southern Chinese metabolic syndrome patients. J Endocrinol Invest (2011) 34(2):e36–42. doi: [10.1007/](https://doi.org/10.1007/BF03347059) [BF03347059](https://doi.org/10.1007/BF03347059)

98. Li Z, Wang X, Pan H, Yang H, Li X, Zhang K, et al. Resistin promotes CCL4 expression through toll-like receptor-4 and activation of the p38-MAPK and NFkB signaling pathways: implications for intervertebral disc degeneration. Osteoarthritis Cartilage (2017) 25(2):341–50. doi: [10.1016/j.joca.2016.10.002](https://doi.org/10.1016/j.joca.2016.10.002)

99. Liu C, Yang H, Gao F, Li X, An Y, Wang J, et al. Resistin promotes intervertebral disc degeneration by upregulation of ADAMTS-5 through p38 MAPK signaling pathway. Spine (2016) 41(18):1414–20. doi: [10.1097/BRS.000](https://doi.org/10.1097/BRS.0000000000001556) [0000000001556](https://doi.org/10.1097/BRS.0000000000001556)

100. Shi C, Wu H, Du D, Im HJ, Zhang Y, Hu B, et al. Nicotinamide
phosphoribosyltransferase inhibitor APO866 prevents IL-1β-induced human nucleus pulposus cell degeneration via autophagy. Cell Physiol Biochem (2018) 49(6):2463–82. doi: [10.1159/000493843](https://doi.org/10.1159/000493843)

101. Cui H, Du X, Liu C, Chen S, Cui H, Liu H, et al. Visfatin promotes intervertebral disc degeneration by inducing IL-6 expression through the ERK/ JNK/p38 signalling pathways. Adipocyte (2021) 10(1):201–15. doi: [10.1080/](https://doi.org/10.1080/21623945.2021.1910155) [21623945.2021.1910155](https://doi.org/10.1080/21623945.2021.1910155)