Check for updates

OPEN ACCESS

EDITED BY Anca Dana Dobrian, Eastern Virginia Medical School, United States

REVIEWED BY Dina H. Kassem, Ain Shams University, Egypt Ingy Hashad, German University in Cairo, Egypt

*CORRESPONDENCE Le Feng M fengle1126@hotmail.com

SPECIALTY SECTION This article was submitted to Diabetes: Molecular Mechanisms, a section of the journal Frontiers in Endocrinology

RECEIVED 08 October 2022 ACCEPTED 13 December 2022 PUBLISHED 04 January 2023

CITATION

Chen T, Wang F, Wei J and Feng L (2023) Extracellular vesicles derived from different sources play various roles in diabetic retinopathy. *Front. Endocrinol.* 13:1064415. doi: 10.3389/fendo.2022.1064415

COPYRIGHT

© 2023 Chen, Wang, Wei and Feng. This is an open-access article distributed under the terms of the **Creative Commons Attribution License** (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Extracellular vesicles derived from different sources play various roles in diabetic retinopathy

Tingting Chen, Fang Wang, Jiayi Wei and Le Feng*

Department of Ophthalmology, Shanghai Tenth People's Hospital Affiliated to Tongji University School of Medicine, Shanghai, China

Extracellular vesicles (EVs) are present in almost all biological fluids and secreted by almost all cell types. A growing number of studies have revealed the potential roles of EVs in the diagnosis and treatment of the diabetic retinopathy (DR). Changes in the quantity and content of EVs may serve as biomarkers of cause or consequence of pathological status of DR, such as inflammation, neovascularization and epithelial-mesenchymal transition. In addition, as natural, safe and efficient drug carrier, EVs have been reported to play important roles in intercellular communication by acting for essential cell-specific information to target cells. In this review, we summarize the roles of EVs, secreted by various types of cells and participated in various biological processes, in the pathogenesis, diagnosis, and treatment of DR.

KEYWORDS

diabetic retinopathy, extracellular vesicles (EVs), exosomes, biomarker, treatment

1 Introduction

Diabetic retinopathy (DR) is one of the most common and serious complications of diabetes, which remains the most prevalent cause of visual impairment in the workingage adults, eventually resulting in irreversible vision loss. DR can be divided into nonproliferative DR (NPDR) and proliferative DR (PDR) according to the absence or presence of retinal neovascularization. Severe visual impairment and blindness may due to the late pathological events of PDR like vitreous hemorrhage, traction retinal detachment and neovascular glaucoma (1). It is worth mentioning that diabetic macular edema (DME) can appear in any stage of DR and has surpassed PDR as the leading cause of visual impairment in patients (2). At present, the treatment modalities for DR include laser photocoagulation, intraocular injections of anti-VEGF or steroids and vitrectomy (3–5). However, these can not completely prevent progression of the disease and reverse visual impairment. In recent years, the prevalence of DR has been increasing, but the early diagnosis and treatment of DR are limited. When patients

01

perceive changes in vision, the visual impairment is irreversible (6). Therefore, it is of great significance to improve the understanding of pathogenesis in DR, and looking for early diagnosis and treatment methods for preventing DR-related visual impairment.

Extracellular vesicles (EVs) which include ectosomes and exosomes, are membrane-derived vesicles that mediate intercellular communications (7). Ectosomes are formed by outward blebbing from the plasma membrane, which include microvesicles, microparticles, and large vesicles ranging from approximately 50 nm to 1 µm in diameter, while exosomes are endosomal origin ranging in size from 40 to 160 nm in diameter (8). EVs are extracellularly released from the majority of human cell types and have been isolated from most bodily fluids, such as vitreous fluid, blood, urine, saliva, breast milk, amniotic fluid, ascites, cerebrospinal fluid, bile and semen etc. (9, 10). The enrichment of tetraspanins such as CD6, CD81, CD9, CD82, CD53 and CD37 can be used as EVs markers (11, 12). EVs are known to contain proteins, lipids, and nucleic acids, which may change in different states of the extracellular environment or secreting cells. Of note, miRNA in nucleic acids are exported out of the cell and can influence gene expression in distant cells (9). Therefore structural and functional properties enable EVs with the characteristics of high specificity, high sensitivity and targeted transport, which are potentially valuable in diagnostic and therapeutic applications.

In recent years, many researches have shown that EVs also play important role in the diagnosis and treatment of DR. In this review, we have summarized EVs, from different sources like intraocular cell-derived EVs, mesenchymal stem cell (MSC)derived EVs and blood-derived EVs, work in different pathways in DR, so that we can provide new ideas for the study of pathogenesis in DR, and look for early diagnosis and treatment.

2 EVs that play roles in the pathogenesis/diagnosis/prognosis of DR

2.1 Intraocular cell-derived EVs

Endothelial cells (ECs), pericytes, astrocytes in the retina (RAC) and retinal pigment epithelial (RPE) cells are important components of inner and outer blood-retinal barrier (BRBs), and play important roles in maintaining the homeostasis of the retinal microenvironment (13). RAC is strictly related to retinal blood vessels and the main producer of VEGF in both normal and pathological angiogenesis (14). Normal pericytes provide vessel stability and inhibit the proliferation and migration of vascular ECs. The loss of pericytes lead not only to the vasodynamic changes in the early stage of DR, but also to the neovascularization in PDR (15). As DR progresses, the tight

junction complexes between RPE cells are disassemble and the pathological leakage occurs in paracellular space (13). When BRB is destroyed, retinal microglia and complement system can maintain retinal homeostasis by regulating the immune system (16). The proliferation and migration of RPE cells and the secretion of extracellular matrix molecules contribute to the formation of fibrotic membranes in the advanced stage of DR, PDR (17). The characteristics of EVs can generally reflect the phenotype and physiology of the parent cell. EVs of different retinal cells have different proteins or genetic information, which can change under pathophysiological conditions, and these minor changes can eventually lead to the retinopathy (18).

2.1.1 Retinal pigment epithelial cell-derived EVs

RPE-derived EVs are involved in the regulation of oxidative stress, inflammation, apoptosis and neovascularization during DR progression. In addition, endothelial-to-mesenchymal transition (EndoMT) lead to pathological fibrosis in PDR, and RPE-derived EVs play potential roles to inhibit EndoMT

Atienzar-Aroca, S et al. (19) found that EVs derived from healthy RPE cell inhibit blood vessel formation under physiological conditions, while those released from stressed ARPE-19 cells promote angiogenesis. This angiogenic effect might be because of the extra cargo of proteins and mRNA contained in the EVs and HUVEC cells influenced by them showed higher levels of VEGF receptors. The further research proposed that the abnormal vessel growth correlate with augmented VEGFR2-expressing EVs, which released from stressed ARPE-19 cells and directly associated with autophagy, causing ECs to migrate and angiogenesis (20). Maisto, R et al. (21) showed that VEGF-containing EVs, which released by ARPE-19 and promoted neovascularization in HUVECs, can be reversed by the melanocortin 5 receptor agonist (MCR5) activation. Ke, Y et al. (22) demonstrated that RPE-derived EVs under oxidative stress increased apoptosis, induced oxidative damage and inflammatory response through Apaf1/caspase-9 axis. Meanwhile, Gu, S et al. (23) proposed that EVs from ARPE-19 cells may transmit miR-202-5p through TGF/Smad pathway, which is an important mediator for intercellular crosstalk to suppress EndoMT and may be a potential treatment for PDR pathological fibrosis.

2.1.2 Pericyte-derived EVs

Pericyte loss is considered a hallmark of early DR, and the changes in pericyte-derived EVs in diabetic conditions may also be of potential value as early biomarkers of DR. Moreover, pericyte-derived Vs still play roles in the crosstalk between pericytes and ECs, which is essential for vessel stabilization and remodeling.

Study by Mazzeo, A et al. (24) showed that miR-126 expression is down-regulated pericyte-derived EVs in diabetic-like conditions, which involved in vessel destabilization in DR.

10.3389/fendo.2022.1064415

Unfortunately, they did not find PDGF and Ang-2 signalling pathways associated with this mechanism. In addition, Liu, C et al. (25) found that diabetes-related stress up-regulated the expression of mmu_circ_0000254 in pericytes and named it cPWWP2A. cPWWP2A directly regulates pericytes biology but indirectly regulates ECs biology *via* EVs carrying cPWWP2A, which inhibited miR-579 activity, increased expression of angiopoietin 1/occludin/SIRT1, and ultimately alleviates diabetes-induced retinal vascular dysfunction.

2.1.3 Retinal astrocytes-derived EVs

RAC-derived EVs are involved in the regulation of endothelial function. Study by Hajrasouliha, A. R et al. (26) showed that RAC-derived EVs inhibited retinal vessel leakage and choroidal neovascularization by targetting both macrophages and ECs. They reduced the number of infiltrating macrophages, directly antagonized the inflammatory and angiogenic factors, and inhibited the migration and vascular tubule forming of ECs. Zhu, L et al. (27) found that RAC-derived EVs under normal and oxidative stress conditions had different effects on the endothelial function, which can be reversed by the exosome inhibitor GW4869 or the autophagy inhibitor 3-methyladenine. This study indicated that oxidative stress can induced RAC autophagy and promote RACs to regulate ECs function by releasing EVs.

2.1.4 Retinal photoreceptors-derived EVs

Retinal photoreceptors (PRs) on the outer membrane of the retina are the most numerous and active cell in retina (28). It is certain that retinopathy is related to the thickness of the ratina and PRs (29). Maisto, R et al. (30) found that high glucose increased VEGF levels and decreased anti-angiogenic miR-20a-3p, miR-20a-5p, miR-106a-5p, and miR-20b expression either in PRs or in PR-derived EVs, RvD1 reverted the effects of glucose damage in photoreceptors and the pro-angiogenic potential of EVs, and ROS-induced NF-kB signaling was considered in relation to this mechanism. Therefore, inhibiting the changes of the outer membrane of the retina by regulating EVs may be a new treatment for the neovascularization of DR.

2.2 Extracocular cell-derived EVs

2.2.1 Blood-derived EVs

2.2.1.1 Serum/plasma-derived EVs

Biomarkers are very important indicators of abnormal biological conditions, and thus can be used as a powerful tool for early diagnosis of DR. Although biomarkers from ocular tissue are more reliable as pathological indicators, obtaining the sample of retina, vitreous fluid and aqueous humor is a high risk invasive procedure, whereas blood sample acquisition is relatively non-invasive, convenient and economical (31). Previous studies have confirmed that the biomarker function of blood-derived EVs could better identify DR at an early stage, and the changes in the quantity and content of EVs could also be used for assessing the severity of DR development.

By comparing the EVs and specific factors in serum or plasma of DR group, the diabetic group and the healthy control group, a large number of studies have found that the levels of cytokines and angiogenic factors in EVs of diabetic patients with retinopathy are significantly increased, which is correlated with the progression of diabetes (32). In addition, Mazzeo, A et al. (33) showed that the differences of miR-150-5p, miR-21-3p and miR-30b-5p in circulating EVs between diabetic patients and healthy subjects were significant, and decreased miR-150-5p and increased miR-21-3p, miR-30b-5p and HIF-1α may all together lead to vessel destabilization and angiogenesis. Zhang, Y et al. (34) found that the content of miR-26b-5p in serum-derived EVs of DR patients was up-regulated. Moreover, Xiao, J et al. (35) compared the proteomic profiles of plasma-derived small EVs from DR patients and normal subjects using iTRAQ-based quantitative proteomics. They found 90 proteins were significantly changed in DR, and confirmed that tumor necrosis factor-α-induced protein 8 (TNFAIP8) was upregulated in plasma-derived small EVs in the DR. All of these aforementioned molecules have been reported to be involved in endothelial dysfunction and angiogenesis, and may be consider as potential biomarkers for DR.

Other studies have found that plasma-derived EVs can be used as indicators of the stage of DR progression. Yu, B et al. (36) showed that miR-431-5p in plasma-derived EVs expression doubled in the PDR patients compared with the nonPDR patients and healthy controls, which indicate miR-431-5p could be used as a marker of disease entering the PDR stage. Tokarz, A et al. (37) suggested that the severity of DR was also statistically correlated with CCR5-positive plasma-derived large EVs.

Furthermore, several studies have shown that diabetic plasma-derived EVs were responsible for activation of the complement cascade. Huang, C et al. (38) found that the increased number of plasma-derived EVs in diabetic patients activated the complement system and upregulated of proinflammatory cytokines and chemokines, resulting in vascular injury. Through further research, they demonstrated that complement activation by IgG-laden EVs leads to membrane attack complex (MAC) deposition, promoting endothelium damage and DR progression (39).

2.2.1.2 Platelet-rich plasma-derived EVs

EVs derived from platelet-rich plasma (PRP) transferred active agents that mediate intercellular crosstalk through

phagocytosis, macropinocytosis, internalization, and endocytosis, affecting hemostasis and inflammatory responses (40). In addition, Zhang, W et al. (41) found that PRP-derived EVs mediated hyperglycemia-induced retinal endothelial injury through the TLR4/CXCL10 axis. Zhang, W et al. (42) demonstrated that DM-PRP-derived EVs activated YAP and enhanced müller cell proliferation and fibrosis through the PI3K/Akt pathway, thereby aggravating the PDR process and threatening the vision of patients.

2.2.2 Pancreatic β -cells-derived EVs

Recent studies have shown that miRNAs released by pancreatic β -cells-derived EVs under diabetic conditions have effects on self and recipient cells through cell-to-cell communication (43). Kamalden, T.A et al. (44) showed that miR-15a was produced in pancreatic β -cells and transported in EVs to distant microvascular beds. As diabetes progresses, more EVs containing miR-15a entered into the blood stream, which leads to the development of diabetic microvascular complications including DR. They further showed that miR-15a-enriched EVs released from β -cells could be taken up by müller cells, causing oxidant stress and apoptosis *via* PI3-kinase signaling pathway.

2.2.3 Lymphocyte-derived EVs

Yang, C et al. (45) found that miR-181a was selectively enriched in lymphocyte-derived EVs, and its overexpression could reduce EC viability and inhibit angiogenesis. They further confirmed *in vitro* and *in vivo* that miR-181a may play a role in reducing retinal neovascularization by interfering with the MAPK1/VEGF signaling pathway, which might provide a new therapeutic strategy for neovascularization (Figure 1).

3 EVs that play roles in treatment of DR

3.1 Mesenchymal stem cells-derived EVs

Mesenchymal stem cells (MSCs), which have the regeneration and differentiation ability and stability, can provide new therapeutic options for retinal diseases (46, 47). Moreover, it is gradually being confirmed that the paracrine trophic effect of MSCs can also be used in the treatment (48). EVs released from MSCs can not only provide the same effect as MSCs, but also avoid the side effects of cell transplantation therapy. Recent researches have demonstrated that MSC-derived EVs from different sources could repair DR-induced pathological changes of endothelial, müller and RPE cells, and played beneficial roles in the treatment of DR.

3.1.1 Adipose tissue MSC-derived EVs

Gu, C et al. (49) applied EVs collected from adipose tissue derived MSC (AD-MSC) to human retinal microvascular endothelial cells, müller cells and RPE cells. They found that AD-MSC-derived EVs could relieve inflammation and angiogenesis by shuttling miR-192, which targeted and negatively regulated ITGA1, thereby reducing diabetic retinal damage. Safwat, A et al. (50) found the decreased expression level of micRNA-222 in retinal tissue of diabetes caused by STZ were associated with severe retinal injury and hemorrhage in different layers of retina and they observed the retinal repair effect of AD-MSC-derived EVs in streptozotocin-induced DM rabbit model by different routes including intravenous injection, subconjunctival injection and intraocular injection. They showed that AD-MSC-derived EVs were able to repair DM-



The molecular mechanism of DR regulated by intraocular/extraocular cell-derived EVs plays roles in the pathogenesis/diagnosis/prognosis of DR.

induced retinal damage and hemorrhage by shuttling micRNA-222, which may occur through its role in regulation of vascular cell biology. And it is necessary to further explore the mechanism by which hyperglycemia caused decrease in expression level of micRNA-222.

3.1.2 Umbilical cord MSC-derived EVs

Xu, N.D et al. (51) showed that human umbilical cord mesenchymal stem cells (UC-MSC)-derived EVs could modulate the proliferation, apoptosis and migration of human RPE cells in hypoxia. Furthermore, Zhang, W et al. (52) found that UC-MSC-derived EVs overexpressing miR-126 could attenuate hyperglycemia-induced inflammatory response and endothelial injury by inhibiting the HMGB1 signaling pathway. Moreover, Li, W et al. (53) injected UC-MSCsderived EVs into the vitreous body of diabetic mouse model, and reveal that UC-MSCs-derived EVs decreased the levels of blood glucose and HbAlc, reduced the contents of inflammatory cytokines and VEGF, alleviated oxidative damage and inhibited retinal cell apoptosis in DR mice *via* shuttling miR-17-3p targeting STAT1.

3.1.3 Bone marrow MSC-derived EVs

Bone marrow contains the highest concentrations of adult stem cells, which are easily harvested and expanded in tissue culture and have the ability to repair damaged tissue, making them more promising for treating retinal diseases. Li, W et al. (54) made high glucose-treated müller cells co-cultured with bone marrow mesenchymal stem cells (BM-MSC)-derived EVs, and demonstrated that up-regulation of miR-486-3p induced by BM-MSC-derived EVs inhibited oxidative stress, inflammation and apoptosis *via* TLR4/NF-κB axis repression. Cao, X et al. (55) confirmed that BM-MSC-derived EVs could transfer SNHG7 to HRMECs and suppressed EndoMT and tube formation *via* miR- 34a-5p/XBP1 axis. Both studies indicated that BM-MSC-derived EVs played protective roles in DR (Figure 2).

4 Limitations of EVs in DR

EVs can be used not only as diagnostic biomarkers for DR, but also as novel targets for therapeutic intervention in DR. Nonetheless, EVs has not been quantitatively used to grade the severity of DR, and EVs' efficient and natural drug carrier role has not yet been used. The regulation of DR by EVs is very complex, and the protein and miRNA information carried by EVs still needs to be supplemented. The complete information database carried by EVs can provide directions for future researches of DR from pathogenesis to diagnosis and treatment.

5 Conclusion

The proteins and miRNAs contained in EVs can participate in the occurrence of DR pathological processes such as inflammation, oxidative stress, apoptosis, neovascularization and EndoMT through different mechanisms. Intraocular cell-derived EVs have two-sided effects according to the organism state. In physiological state, EVs maintain retinal homeostasis, while in pathological state, EVs induce oxidative stress and inflammation to damage retinal tissue, which can be alleviated by interfering with specific miRNAs or proteins in EVs. MSC-derived EVs can not only perfectly replicate the regeneration and differentiation abilities of MSCs, but also avoid their side effects. Therefore, EVs have the potential to be the natural material for the treatment of DR. Circulating EVs are relatively convenient and non-invasive to obtain, and the changes in their quantity and content of specific components in the early stage of DR are expected to be potential



biomarkers for early diagnosis. In addition, circulating EVs can also regulate the neovascularization and inflammatory response of DR, which are expected to play the roles of DR drug therapy carriers. In conclusion, the research on EVs in DR is still in its infancy, and has broad application prospects, which need to be further explored.

Author contributions

Conception or design: LF, FW. Search and organize literature: TC, JW. Drafting the work or revising: TC. Final approval of the manuscript: LF.

Funding

This work was supported by Shanghai Tenth People's Hospital Climbing Talent Program (No.2018SYPDRC029) and

References

1. Wong TY, Cheung CM, Larsen M, Sharma S, Simo R. Diabetic retinopathy. *Nat Rev Dis Primers* (2016) 2:16012. doi: 10.1038/nrdp.2016.12

2. Zhang X, Saaddine JB, Chou CF, Cotch MF, Cheng YJ, Geiss LS, et al. Prevalence of diabetic retinopathy in the united states, 2005-2008. *JAMA* (2010) 304(6):649-56. doi: 10.1001/jama.2010.1111

3. Rodriguez ML, Perez S, Mena-Molla S, Desco MC, Ortega AL. Oxidative stress and microvascular alterations in diabetic retinopathy: Future therapies. Oxid Med Cell Longev (2019) 2019:4940825. doi: 10.1155/2019/4940825

4. Tarr JM, Kaul K, Chopra M, Kohner EM, Chibber R. Pathophysiology of diabetic retinopathy. ISRN Ophthalmol (2013) 2013:343560. doi: 10.1155/2013/343560

5. Mitchell P, Wong TYDiabetic Macular Edema Treatment Guideline Working G. Management paradigms for diabetic macular edema. *Am J Ophthalmol* (2014) 157(3):505–13 e1-8. doi: 10.1016/j.ajo.2013.11.012

6. Wong TY, Sabanayagam C. Strategies to tackle the global burden of diabetic retinopathy: From epidemiology to artificial intelligence. *Ophthalmologica* (2020) 243(1):9–20. doi: 10.1159/000502387

7. Raposo G, Stoorvogel W. Extracellular vesicles: Exosomes, microvesicles, and friends. J Cell Biol (2013) 200(4):373–83. doi: 10.1083/jcb.201211138

8. Kalluri R, LeBleu VS. The biology, function, and biomedical applications of exosomes. *Science* (2020) 367(6478):eaau6977. doi: 10.1126/science.aau6977

9. Colombo M, Raposo G, Thery C. Biogenesis, secretion, and intercellular interactions of exosomes and other extracellular vesicles. *Annu Rev Cell Dev Biol* (2014) 30:255–89. doi: 10.1146/annurev-cellbio-101512-122326

10. Simons M, Raposo G. Exosomes-vesicular carriers for intercellular communication. *Curr Opin Cell Biol* (2009) 21(4):575-81. doi: 10.1016/j.ceb.2009.03.007

11. Zoller M. Tetraspanins: Push and pull in suppressing and promoting metastasis. Nat Rev Cancer (2009) 9(1):40-55. doi: 10.1038/nrc2543

12. Hemler ME. Tetraspanin proteins mediate cellular penetration, invasion, and fusion events and define a novel type of membrane microdomain. *Annu Rev Cell Dev Biol* (2003) 19:397–422. doi: 10.1146/annurev.cellbio.19.111301.153609

13. Jo DH, Yun JH, Cho CS, Kim JH, Kim JH, Cho CH. Interaction between microglia and retinal pigment epithelial cells determines the integrity of outer blood-retinal barrier in diabetic retinopathy. *Glia* (2019) 67(2):321–31. doi: 10.1002/glia.23542

14. Sorrentino FS, Allkabes M, Salsini G, Bonifazzi C, Perri P. The importance of glial cells in the homeostasis of the retinal microenvironment and their pivotal role in the course of diabetic retinopathy. *Life Sci* (2016) 162:54–9. doi: 10.1016/ j.lfs.2016.08.001

Shanghai Tenth People's Hospital Clinical Research Cultivation Project (No.YNCR2C011).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

15. Ejaz S, Chekarova I, Ejaz A, Sohail A, Lim CW. Importance of pericytes and mechanisms of pericyte loss during diabetes retinopathy. *Diabetes Obes Metab* (2008) 10(1):53–63. doi: 10.1111/j.1463-1326.2007.00795.x

16. Chen M, Luo C, Zhao J, Devarajan G, Xu H. Immune regulation in the aging retina. *Prog Retin Eye Res* (2019) 69:159–72. doi: 10.1016/j.preteyeres.2018.10.003

17. Qin D, Jiang YR, Meng Z. Gremlin in the vitreous of patients with proliferative diabetic retinopathy and the downregulation of gremlin in retinal pigment epithelial cells. *J Diabetes Res* (2020) 2020:9238742. doi: 10.1155/2020/9238742

18. van der Merwe Y, Steketee MB. Extracellular vesicles: Biomarkers, therapeutics, and vehicles in the visual system. *Curr Ophthalmol Rep* (2017) 5 (4):276–82. doi: 10.1007/s40135-017-0153-0

19. Atienzar-Aroca S, Flores-Bellver M, Serrano-Heras G, Martinez-Gil N, Barcia JM, Aparicio S, et al. Oxidative stress in retinal pigment epithelium cells increases exosome secretion and promotes angiogenesis in endothelial cells. *J Cell Mol Med* (2016) 20(8):1457–66. doi: 10.1111/jcmm.12834

20. Atienzar-Aroca S, Serrano-Heras G, Freire Valls A, Ruiz de Almodovar C, Muriach M, Barcia JM, et al. Role of retinal pigment epithelium-derived exosomes and autophagy in new blood vessel formation. *J Cell Mol Med* (2018) 22(11):5244–56. doi: 10.1111/jcmm.13730

21. Maisto R, Oltra M, Vidal-Gil L, Martinez-Gil N, Sancho-Pelluz J, Filippo CD, et al. Arpe-19-Derived vegf-containing exosomes promote neovascularization in huvec: The role of the melanocortin receptor 5. *Cell Cycle* (2019) 18(4):413–24. doi: 10.1080/15384101.2019.1568745

22. Ke Y, Fan X, Rui H, Xinjun R, Dejia W, Chuanzhen Z, et al. Exosomes derived from rpe cells under oxidative stress mediate inflammation and apoptosis of normal rpe cells through Apaf1/Caspase-9 axis. *J Cell Biochem* (2020). doi: 10.1002/jcb.29713

23. Gu S, Liu Y, Zou J, Wang W, Wei T, Wang X, et al. Retinal pigment epithelial cells secrete mir-202-5p-Containing exosomes to protect against proliferative diabetic retinopathy. *Exp Eye Res* (2020) 201:108271. doi: 10.1016/j.exer.2020.108271

24. Mazzeo A, Beltramo E, Iavello A, Carpanetto A, Porta M. Molecular mechanisms of extracellular vesicle-induced vessel destabilization in diabetic retinopathy. *Acta Diabetol* (2015) 52(6):1113–9. doi: 10.1007/s00592-015-0798-9

25. Liu C, Ge HM, Liu BH, Dong R, Shan K, Chen X, et al. Targeting pericyteendothelial cell crosstalk by circular rna-Cpwwp2a inhibition aggravates diabetesinduced microvascular dysfunction. *Proc Natl Acad Sci U.S.A.* (2019) 116 (15):7455–64. doi: 10.1073/pnas.1814874116 26. Hajrasouliha AR, Jiang G, Lu Q, Lu H, Kaplan HJ, Zhang HG, et al. Exosomes from retinal astrocytes contain antiangiogenic components that inhibit laser-induced choroidal neovascularization. *J Biol Chem* (2013) 288(39):28058–67. doi: 10.1074/jbc.M113.470765

27. Zhu L, Zang J, Liu B, Yu G, Hao L, Liu L, et al. Oxidative stress-induced rac autophagy can improve the huvec functions by releasing exosomes. *J Cell Physiol* (2020) 235(10):7392–409. doi: 10.1002/jcp.29641

28. Ames A3rd. Energy requirements of cns cells as related to their function and to their vulnerability to ischemia: A commentary based on studies on retina. *Can J Physiol Pharmacol* (1992) 70 Suppl:S158–64. doi: 10.1139/y92-257

29. Tavares Ferreira J, Proenca R, Alves M, Dias-Santos A, Santos BO, Cunha JP, et al. Retina and choroid of diabetic patients without observed retinal vascular changes: A longitudinal study. *Am J Ophthalmol* (2017) 176:15–25. doi: 10.1016/ j.ajo.2016.12.023

30. Maisto R, Trotta MC, Petrillo F, Izzo S, Cuomo G, Alfano R, et al. Resolvin D1 modulates the intracellular vegf-related mirnas of retinal photoreceptors challenged with high glucose. *Front Pharmacol* (2020) 11:235. doi: 10.3389/fphar.2020.00235

31. Kastelan S, Oreskovic I, Biscan F, Kastelan H, Gverovic Antunica A. Inflammatory and angiogenic biomarkers in diabetic retinopathy. *Biochem Med* (Zagreb) (2020) 30(3):30502. doi: 10.11613/BM.2020.030502

32. Tokarz A, Szuscik I, Kusnierz-Cabala B, Kapusta M, Konkolewska M, Zurakowski A, et al. Extracellular vesicles participate in the transport of cytokines and angiogenic factors in diabetic patients with ocular complications. *Folia Med Cracov* (2015) 55(4):35–48.

33. Mazzeo A, Lopatina T, Gai C, Trento M, Porta M, Beltramo E. Functional analysis of mir-21-3p, mir-30b-5p and mir-150-5p shuttled by extracellular vesicles from diabetic subjects reveals their association with diabetic retinopathy. *Exp Eye Res* (2019) 184:56–63. doi: 10.1016/j.exer.2019.04.015

34. Zhang Y, Wei J, Zhang L, Jiang G, Wang B, Jiang L. Extracellular vesiclederived mir-26b-5p is up-regulated in the serum of patients with diabetic retinopathy. *Comb Chem High Throughput Screen* (2022) 25(5):877–82. doi: 10.2174/1386207324666210216092917

35. Xiao J, Zhang H, Yang F, Xiao M, Zhou L, Yu R, et al. Proteomic analysis of plasma sevs reveals that Tnfaip8 is a new biomarker of cell proliferation in diabetic retinopathy. *J Proteome Res* (2021) 20(3):1770–82. doi: 10.1021/acs.jproteome.0c01048

36. Yu B, Xiao M, Yang F, Xiao J, Zhang H, Su L, et al. Microrna-431-5p encapsulated in serum extracellular vesicles as a biomarker for proliferative diabetic retinopathy. *Int J Biochem Cell Biol* (2021) 135:105975. doi: 10.1016/ibiocel.2021.105975

37. Tokarz A, Konkolewska M, Kusnierz-Cabala B, Maziarz B, Hanarz P, Zurakowski A, et al. Retinopathy severity correlates with rantes concentrations and ccr 5-positive microvesicles in diabetes. *Folia Med Cracov* (2019) 59(3):95–112. doi: 10.24425/fmc.2019.131139

38. Huang C, Fisher KP, Hammer SS, Navitskaya S, Blanchard GJ, Busik JV. Plasma exosomes contribute to microvascular damage in diabetic retinopathy by activating the classical complement pathway. *Diabetes* (2018) 67(8):1639–49. doi: 10.2337/db17-1587

39. Huang C, Fisher KP, Hammer SS, Busik JV. Extracellular vesicle-induced classical complement activation leads to retinal endothelial cell damage. *Via* Mac (2020) 21(5):1693. doi: 10.3390/ijms21051693

40. Xu N, Wang L, Guan J, Tang C, He N, Zhang W, et al. Wound healing effects of a curcuma zedoaria polysaccharide with platelet-rich plasma exosomes assembled on Chitosan/Silk hydrogel sponge in a diabetic rat model. *Int J Biol macromolecules* (2018) 117:102–7. doi: 10.1016/j.ijbiomac.2018.05.066

41. Zhang W, Dong X, Wang T, Kong Y. Exosomes derived from platelet-rich plasma mediate hyperglycemia-induced retinal endothelial injury *Via* targeting the Tlr4 signaling pathway. *Exp Eye Res* (2019) 189:107813. doi: 10.1016/j.exer.2019.107813

42. Zhang W, Jiang H, Kong Y. Exosomes derived from platelet-rich plasma activate yap and promote the fibrogenic activity of Muller cells *Via* the Pi3k/Akt pathway. *Exp Eye Res* (2020) 193:107973. doi: 10.1016/j.exer.2020.107973

43. Guay C, Menoud V, Rome S, Regazzi R. Horizontal transfer of exosomal micrornas transduce apoptotic signals between pancreatic beta-cells. *Cell Commun Signal* (2015) 13:17. doi: 10.1186/s12964-015-0097-7

44. Kamalden TA, Macgregor-Das AM, Kannan SM, Dunkerly-Eyring B, Khaliddin N, Xu Z, et al. Exosomal microrna-15a transfer from the pancreas augments diabetic complications by inducing oxidative stress. *Antioxid Redox Signal* (2017) 27(13):913–30. doi: 10.1089/ars.2016.6844

45. Yang C, Tahiri H, Cai C, Gu M, Gagnon C, Hardy P. Microrna-181a inhibits ocular neovascularization by interfering with vascular endothelial growth factor expression. *Cardiovasc Ther* (2018) 36(3):e12329. doi: 10.1111/1755-5922.12329

46. Park SS, Moisseiev E, Bauer G, Anderson JD, Grant MB, Zam A, et al. Advances in bone marrow stem cell therapy for retinal dysfunction. *Prog Retin Eye Res* (2017) 56:148–65. doi: 10.1016/j.preteyeres.2016.10.002

47. Wang Y, Fu B, Sun X, Li D, Huang Q, Zhao W, et al. Differentially expressed micrornas in bone marrow mesenchymal stem cell-derived microvesicles in young and older rats and their effect on tumor growth factor-Beta1-Mediated epithelial-mesenchymal transition in Hk2 cells. *Stem Cell Res Ther* (2015) 6:185. doi: 10.1186/s13287-015-0179-x

48. Gnecchi M, Zhang Z, Ni A, Dzau VJ. Paracrine mechanisms in adult stem cell signaling and therapy. *Circ Res* (2008) 103(11):1204–19. doi: 10.1161/CIRCRESAHA.108.176826

49. Gu C, Zhang H, Gao Y. Adipose mesenchymal stem cells-secreted extracellular vesicles containing microrna-192 delays diabetic retinopathy by targeting Itga1. *J Cell Physiol* (2021) 236(7):5036–51. doi: 10.1002/jcp.30213

50. Safwat A, Sabry D, Ragiae A, Amer E, Mahmoud RH, Shamardan RM. Adipose mesenchymal stem cells-derived exosomes attenuate retina degeneration of streptozotocin-induced diabetes in rabbits. *J Circ biomark* (2018) 7:1849454418807827. doi: 10.1177/1849454418807827

51. Xu ND, Huang LZ, Zhu L, Tang X, Li Y, Zhao MW. [Human umbilical mesenchymal stem cells-derived exosomes modulate the proliferation, apoptosis and migration of human retinal pigment epithelial cells in hypoxia]. *Zhonghua Yan Ke Za Zhi* (2019) 55(12):933–41. doi: 10.3760/cma.j.issn.0412-4081.2019.12.012

52. Zhang W, Wang Y, Kong Y. Exosomes derived from mesenchymal stem cells modulate mir-126 to ameliorate hyperglycemia-induced retinal inflammation *Via* targeting Hmgb1. *Invest Ophthalmol Vis Sci* (2019) 60(1):294–303. doi: 10.1167/iovs.18-25617

53. Li W, Jin LY, Cui YB, Xie N. Human umbilical cord mesenchymal stem cells-derived exosomal microrna-17-3p ameliorates inflammatory reaction and antioxidant injury of mice with diabetic retinopathy *Via* targeting Stat1. *Int Immunopharmacol* (2021) 90:107010. doi: 10.1016/j.intimp.2020.107010

54. Li W, Jin L, Cui Y, Nie A, Xie N, Liang G. Bone marrow mesenchymal stem cells-induced exosomal microrna-486-3p protects against diabetic retinopathy through Tlr4/Nf-kappab axis repression. *J Endocrinol Invest* (2021) 44(6):1193–207. doi: 10.1007/s40618-020-01405-3

55. Cao X, Xue LD, Di Y, Li T, Tian YJ, Song Y. Msc-derived exosomal lncrna Snhg7 suppresses endothelial-mesenchymal transition and tube formation in diabetic retinopathy *Via* mir-34a-5p/Xbp1 axis. *Life Sci* (2021) 272:119232. doi: 10.1016/j.lfs.2021.119232