

IGF-I influences everolimus activity in medullary thyroid carcinoma

Erica Gentilin^{1,2}, Carmelina Di Pasquale¹, Martina Rossi¹, Federico Tagliati¹, Teresa Gagliano¹, Roberta Rossi¹, Mariarosa Pelizzo³, Isabella Merante Boschin³, Ettore C. degli Uberti^{1,2} and Maria Chiara Zatelli^{1,2*}

¹ Section of Endocrinology and Internal Medicine, Department of Medical Sciences, University of Ferrara, Ferrara, Italy, ² Laboratorio in rete del Tecnopolo "Tecnologie delle Terapie Avanzate" (LTTA), University of Ferrara, Ferrara, Italy, ³ Department of Surgical, Oncological and Gastroenterological Sciences, University of Padova, Padova, Italy

OPEN ACCESS

Edited by:

Antonino Belfiore,
University Magna Graecia of
Catanzaro, Italy

Reviewed by:

Piero Ferolla,
Multidisciplinary Group for Diagnosis
and Treatment of Neuroendocrine
Tumors, Italy
Daniela Pasquali,
Seconda Università degli Studi di
Napoli, Italy

*Correspondence:

Maria Chiara Zatelli,
Section of Endocrinology and Internal
Medicine, Department of Medical
Sciences, University of Ferrara, Via
Savonarola 9, Ferrara 44100, Italy
ztlmch@unife.it

Specialty section:

This article was submitted to Cancer
Endocrinology, a section of the journal
Frontiers in Endocrinology

Received: 12 February 2015

Accepted: 10 April 2015

Published: 05 May 2015

Citation:

Gentilin E, Di Pasquale C, Rossi M,
Tagliati F, Gagliano T, Rossi R,
Pelizzo M, Merante Boschin I,
degli Uberti EC and Zatelli MC (2015)
IGF-I influences everolimus activity in
medullary thyroid carcinoma.
Front. Endocrinol. 6:63.
doi: 10.3389/fendo.2015.00063

Context: Medullary thyroid carcinoma (MTC) is a rare tumor originating from thyroid parafollicular C cells. It has been previously demonstrated that insulin-like growth factor I (IGF-I) protects MTC from the effects of antiproliferative drugs. Everolimus, an mTOR inhibitor, has shown potent antiproliferative effects in a human MTC cell line, TT, and in two human MTC primary cultures.

Objective: To verify whether IGF-I may influence the effects of everolimus in a group of human MTC primary cultures.

Design: We collected 18 MTCs that were dispersed in primary cultures, treated without or with 10 nM–1 μM everolimus and/or 50 nM IGF-I. Cell viability was evaluated after 48 h, and calcitonin (CT) secretion was assessed after a 6 h incubation. IGF-I receptor downstream signaling protein expression profile was also investigated.

Results: Everolimus significantly reduced cell viability in eight MTC [by ~20%; $P < 0.01$ vs. control; everolimus-responders (E-R) MTCs], while cell viability did not change in 10 MTCs [everolimus-non-responders (E-NR) MTCs]. In E-R MTCs, IGF-I blocked the antiproliferative effects of everolimus that did not affect CT secretion, but blocked the stimulatory effects of IGF-I on this parameter. IGF-I receptor downstream signaling proteins were expressed at higher levels in E-NR MTC as compared to E-R MTCs.

Conclusion: IGF-I protects a subset of MTC primary cultures from the antiproliferative effects of everolimus and stimulates CT secretion by an mTOR mediated pathway that, in turn, may represent a therapeutic target in the treatment of aggressive MTCs.

Keywords: IGF-I, medullary thyroid carcinoma, everolimus, mTOR, calcitonin

Introduction

Insulin-like growth factor I (IGF-I) system has been described in both rat (1) and human medullary thyroid carcinoma (MTC) (2), where it stimulates DNA synthesis (3). In a human MTC cell line, the TT cells, IGF-I increases cell proliferation, DNA synthesis, and the percentage of cells in S phase, all effects counteracted by co-incubation with either an IGF-I antibody or an IGF-I receptor (IGF-I R) antibody (4). In addition, it has been previously demonstrated that IGF-I protects TT cells from drug-induced cytotoxicity (5). Therefore, IGF-I may hamper the effects of antiproliferative drugs in MTC.

New medical approaches became lately available for MTC, thanks to the development of several agents that target tyrosine kinases. Clinical trials have reported variable results, since patients may develop drug resistance with consequent tumor progression (6). An alternative medical approach for neuroendocrine tumors, including MTC, is represented by mTOR inhibitors, such as everolimus. The latter displays antiproliferative effects on a wide spectrum of tumors including neuroendocrine tumors, both *in vitro* and *in vivo* (7–11). Moreover, it has been recently demonstrated that everolimus inhibits cell viability in a dose- and time-dependent fashion and reduces mTOR phosphorylation in a human MTC cell line and in two human MTC primary cultures (12). In addition, everolimus treatment of two patients with progressive metastatic MTC was associated with disease stabilization in one and disease progression in the other patient (13), indicating that mTOR pathway may effectively control MTC cell proliferation in a subset of patients. The variable effects of mTOR inhibitors may be ascribed to independent signaling mechanisms, activated by several growth factors, including IGF-I.

The aim of our study is therefore to verify whether IGF-I may influence the effects of everolimus in a group of human MTC primary cultures.

Materials and Methods

Materials

All reagents, if not otherwise specified, were purchased from Sigma-Aldrich (Milano, Italy). Everolimus was provided by Novartis Pharma (Basel, Switzerland).

Human MTCs

The samples derived from 18 patients diagnosed and operated on for MTC at the Section of Endocrinology and Internal Medicine of the University of Ferrara, and at the Department of Surgical, Oncological and Gastroenterological Sciences of the University of Padova. **Table 1** shows patients' characteristics and pre-operative hormonal values. All patients (six males and 12 females; age = 52.1 ± 3.9 years) underwent total thyroidectomy with central neck lymph node dissection and had histological and immunohistochemical diagnosis of MTC.

Tissue Collection and Primary Culture

Tissues were collected following the guidelines of the local committee on human research, and immediately minced in RPMI 1640 medium under sterile conditions. Primary cultures were then prepared as described previously (14, 15). Cells were then treated with test substances, with further evaluation of hormone secretion and cell viability. Informed consent of the patients was obtained for disclosing clinical investigation and performing the *in vitro* study.

Cell Viability

The effects of everolimus and IGF-I on MTC cell viability *in vitro* were assessed by ATPlite assay (Perkin-Elmer, Monza, Italy) on the Wallac Victor™ 1420 Multilabel Counter (Perkin-Elmer) as previously described (16). Cells were treated after 24 h with or without 10 nM–1 μ M everolimus and/or 50 nM IGF-I. Treatments

TABLE 1 | MTC patients clinical characteristics.

No.	Sex	Age	Plasma CT (pg/ml)	Stage	Inheritance
1	M	46	157	II	SP
2	M	40	1500	II	FMTc
3	M	47	940	II	MEN2A
4	M	39	1500	II	SP
5	F	35	700	II	SP
6	F	32	28	II	MEN2A
7	F	35	74	I	SP
8	F	33	19	I	FMTc
9	F	42	153	II	SP
10	M	52	207	II	SP
11	F	44	2350	II	SP
12	M	56	1258	II	SP
13	F	79	1500	III	SP
14	F	71	2578	III	SP
15	F	73	3848	IVc	SP
16	F	75	1500	III	SP
17	F	69	3405	III	SP
18	F	70	9227	IVb	SP

FMTc, familial medullary thyroid carcinoma; MEN2A, MEN2A RET mutation; SP, sporadic disease.

were renewed after the first 24 h of incubation. Cell viability was assessed after 48 h. Results were obtained by determining the mean value of six replicates.

Protein Expression Panel

Tissues were dissolved in cell lysis buffer (Bio-Rad, Milano, Italy) supplemented with cell lysis factor QG (Bio-Rad, Milano, Italy) and 2 mM phenylmethanesulfonyl fluoride. Protein concentration was measured by BCA Protein Assay Reagent Kit (Pierce, Rockford, IL, USA), as previously described (17). Bio-plex®/Luminex® Technology (Bioclarma Research and Molecular Diagnostics, Torino, Italy) was employed to assess total protein levels of IGF-I receptor (IGF-I R), AKT, p70S6K, p38MAPK, ERK1/2, and CREB in MTC tissue samples. The following phosphorylated forms were also investigated: p(Tyr1131) IGF-I R, p(Ser473) AKT, p(Thr421/Ser424) p70S6K, p(Thr180/Tyr182) p38MAPK, p(Thr202/Thr204, Thr185/Thr187) ERK1/2, and p(Ser133) CREB levels.

Calcitonin Assay

Calcitonin (CT) was measured in conditioned medium from primary cultured cells by an ELISA kit (DRG, Springfield, NJ, USA), after a 6 h treatment without or with the test substances. The intra- and inter-assay variation coefficients were 2.8–5.7% and 6.1–7.4%, respectively. The detection limit was 1.0 pg/ml. Assays were performed in duplicate.

Statistical Analysis

Fisher exact test was used to evaluate the association between clinical characteristics of the patients and MTC primary culture responsiveness to everolimus in terms of cell viability reduction.

Results are expressed as the mean \pm standard error of the mean (SEM). A Student's paired or unpaired *t* test was used to evaluate the individual differences between means.

P-values < 0.05 were considered significant.

Results

Effects of Everolimus on Cell Viability

To determine the effects of everolimus on cell viability of dispersed MTC cells, we assessed cell viability in MTC primary cultures after a 48 h treatment with 10 nM–1 μ M everolimus. According to the extent of cell viability reduction recorded after treatment with everolimus, the primary cultures were divided in two groups: the cultures responding to any everolimus concentration with a significant ($P < 0.05$) cell viability reduction were considered as “everolimus-responders” (E-R), while those displaying a non-significant cell viability variation were considered as “everolimus-non-responders” (E-NR). According to this criterion, cultures from eight MTC were considered as E-R and 10 as E-NR.

Evaluating the patients clinical charts, we found that mean CT plasma levels of patients which primary cultures belonged to the E-NR group were ~fourfold higher than the mean CT plasma levels of patients which primary cultures belonged to the E-R group ($P = 0.054$). In addition, disease stage and inheritance were differently distributed among patients which primary cultures belonged to the E-R and those which primary cultures belonged to the E-NR group. In particular, E-NR MTCs were significantly associated with a disease stage \geq III ($P < 0.02$) and with sporadic disease ($P < 0.03$).

In the E-R group (black bars), everolimus dose-dependently reduced cell viability, from –19% at 10 nM to –31% vs. control at 1 μ M ($P < 0.01$). In the E-NR MTCs (white bars), everolimus did not significantly modify cell viability (Figure 1). Further experiments were carried out only in the E-R group.

Protein Profiling

In a subset of MTC samples (including two E-R and four E-NR MTCs), we had a sufficient tissue amount that allowed us to investigate protein expression profile. We then assessed total and phosphorylated protein levels of IGF-I R, AKT, p70S6K, p38MAPK, ERK1/2, and CREB. We found that total IGF-I R, p70S6K,

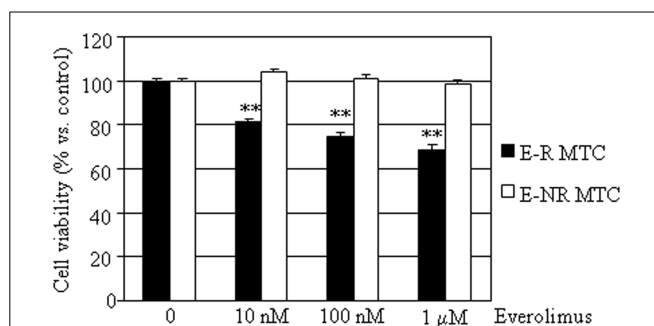


FIGURE 1 | Effects of everolimus on MTC primary culture cell viability. MTC primary cultures were incubated in 96-well plates for 48 h in culture medium supplemented with everolimus at increasing concentrations from 10 nM to 1 μ M, and control cells were treated with vehicle solution. Data from 18 MTC primary cultures were evaluated independently with six replicates each, and were expressed as the mean \pm SEM percent cell viability vs. control cells. ** $P < 0.01$ vs. control cells. As described in the Section “Results,” MTCs were divided according to cell viability inhibition after treatment with everolimus in E-R (eight samples, black bars) and E-NR (10 samples, white bars).

p38MAPK, ERK1/2, and CREB protein levels were higher in E-NR MTCs as compared to E-R MTCs (+153%, +259%, +68%, +53%, and +1735%, respectively). Statistical significance was observed for total CREB protein levels in E-NR MTCs vs. E-R MTCs ($P < 0.02$). On the contrary, total AKT protein levels were similar in the two groups (Figure 2A). In addition, phosphorylated IGF-I R, AKT, p70S6K, p38MAPK, and CREB protein levels were higher in E-NR MTCs as compared to E-R MTCs (+10%, +32%, +200%, +165%, and +426%, respectively). On the contrary, phosphorylated ERK1/2 protein levels were similar in the two groups (Figure 2B).

Effects of IGF-I on Cell Viability in Everolimus-Responder MTCs

To investigate whether everolimus inhibitory effects might involve growth factor activated pathways, cell viability was assessed in E-R MTC cultures treated with or without 50 nM IGF-I, in the presence or in the absence of 10 nM–1 μ M everolimus. IGF-I did not significantly affect cell viability, but counteracted the inhibitory effects of everolimus at all concentrations tested (Figure 3).

Effects of IGF-I on CT Secretion in Everolimus-Responder MTCs

To determine the effects of everolimus on CT secretion by dispersed MTC cells, we assessed CT concentrations in conditioned medium from the eight E-R MTC cultures. CT levels were not significantly influenced by treatment with everolimus at any concentration tested. On the other hand, IGF-I significantly induced CT secretion (+19%; $P < 0.01$ vs. control), an effect that was reduced by co-incubation with everolimus at all concentrations tested (Figure 4).

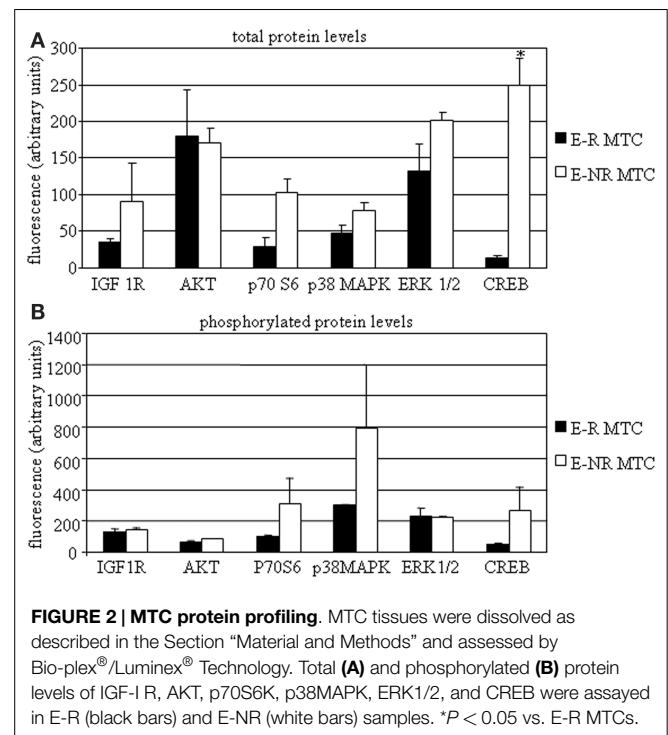


FIGURE 2 | MTC protein profiling. MTC tissues were dissolved as described in the Section “Material and Methods” and assessed by Bio-plex®/Luminex® Technology. Total (A) and phosphorylated (B) protein levels of IGF-I R, AKT, p70S6K, p38MAPK, ERK1/2, and CREB were assayed in E-R (black bars) and E-NR (white bars) samples. * $P < 0.05$ vs. E-R MTCs.

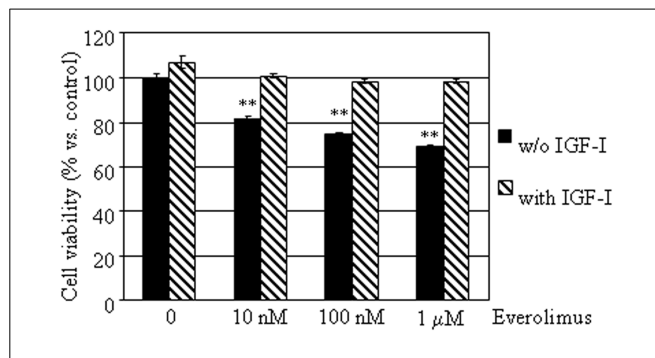


FIGURE 3 | Effects of IGF-I on MTC primary culture cell viability. MTC primary cultures were incubated in 96-well plates for 48 h in culture medium supplemented with everolimus at increasing concentrations from 10 nM to 1 μM, and control cells were treated with vehicle solution. Data from 18 MTC primary cultures were evaluated independently with six replicates each, and were expressed as the mean ± SEM percent cell viability vs. control cells. ** $P < 0.01$ vs. control cells. E-R MTC were incubated with (dashed bars) or without 50 nM IGF-I (black bars).

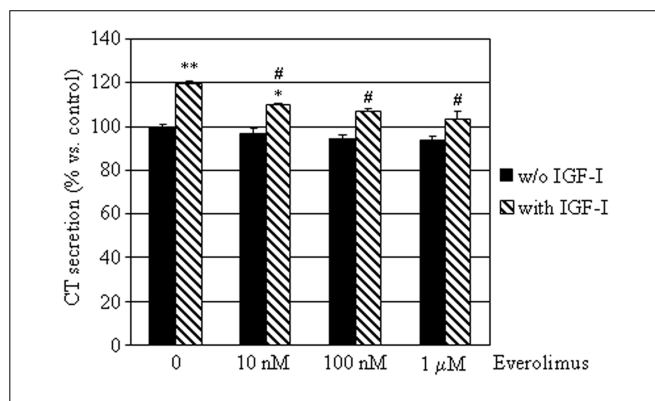


FIGURE 4 | Effects of everolimus and IGF-I on CT secretion by E-R MTCs. E-R MTC primary cultures were incubated in 96-well plates for 6 h in culture medium supplemented with indicated substances. Control cells were treated with vehicle solution. CT secretion by each primary culture was then measured by ELISA. Data from eight E-R MTC primary cultures were evaluated independently with two replicates each and were expressed as the mean ± SEM percent CT secretion vs. control cells. * $P < 0.05$ and ** $P < 0.01$ vs. control cells. # $P < 0.05$ vs. IGF-I treated cells.

Discussion

This study demonstrates that IGF-I influences the effects of everolimus in a sub-group of human MTC primary cultures, supporting the hypothesis that IGF-I may hamper the effects of antiproliferative drugs in MTC. In addition, our results indicate that mTOR inhibitors may be effective in restraining cell proliferation only in a subset of MTCs, in keeping with previously reported *in vivo* data (13, 18). Indeed, Lim et al. show that only one out of the nine MTC patients treated with everolimus displays a ~20% reduction in tumor bulk, while the other patients show either stable or progressive disease (18). These data support the hypothesis that other survival pathways, including those activated by *RET* mutations, are active in MTC and may hamper the effects of mTOR inhibitors. The latter may be cytostatic, since it has been demonstrated that everolimus treatment of pancreatic neuroendocrine tumor (pNET) and MTC cell lines inhibits cell growth by

increasing the G0/G1 phase of the cell cycle (12, 19). Moreover, it is widely demonstrated that mTOR inhibition has a significant antiproliferative effect on pNET cell lines (20) as well as on other tumor cells (8, 9).

Insulin-like growth factor I is confirmed as a protective growth factor toward C-cell survival, in keeping with previous evidence (21). Our data confirm that IGF-I does not stimulate cell proliferation of MTC primary cultures, but protects them from the effects of everolimus, suggesting that escape from therapy may occur in the presence of IGF-I or similar growth factors. This hypothesis is further strengthened by the finding that E-NR MTC display higher levels of IGF-I R and therefore may possibly be more sensitive to the protective effects of IGF-I. Indeed, we observed that IGF-I R displays higher phosphorylation levels in E-NR MTC as compared to E-R MTCs. This finding is in keeping with a greater activation of the IGF-I pathway, as also supported by higher phosphorylation levels (i.e., activation) of IGF-I downstream signaling proteins. Moreover, MTCs belonging to the E-NR group are significantly associated with sporadic and higher disease stage, indicating that this group represents a more aggressive disease.

In addition, we found that mTOR pathway is activated in MTC, as previously reported (22), especially in E-NR as compared to E-R MTCs. This finding indicates that mTOR signaling pathway over-activation is not invariably a marker of sensitivity to mTOR inhibitors in all endocrine and endocrine-related tumors (7, 9, 23). However, further studies are needed to identify a prognostic protein panel that may accurately predict responsiveness of MTCs to mTOR inhibitors on clinical grounds.

Our results confirm that IGF-I promotes CT secretion in a subset of MTC primary cultures, as previously demonstrated (21). In addition, we found that this effect is hampered by everolimus, indicating that IGF-I modulates CT secretion involving the mTOR pathway, which, in turn, may regulate CT secretion only in response to secretory stimuli, such as IGF-I. It has been previously shown that IGF-I stimulates hormone secretion in parathyroid cells by modulating calcium channels (24) that are present in MTC cells (25). Calcium-dependent signaling pathways are induced by IGF-I through CREB phosphorylation (26) that, in turn, regulates CT secretion (27). Higher levels of both total and phosphorylated CREB protein were found in E-NR MTCs, that also display higher IGF-I R levels and higher CT plasma levels, supporting the hypothesis that IGF-I system may be over-activated and may promote CT secretion in E-NR MTCs. These tumors, despite displaying aggressive characteristics, may anyway benefit from treatment with everolimus, since this drug may control CT secretion. Indeed, as shown in the available clinical studies, everolimus treatment is effective in significantly reducing circulating CT plasma levels also when no effect on tumor bulk was apparent (13, 18, 28).

Our results also indicate that everolimus treatment may not result in both tumor growth control and hormone secretion reduction in all MTCs, as also previously found for treatment with somatostatin analogs, that could exert antiproliferative effects despite the lack of antisecretory activity and vice versa (14). These finding suggest that proliferation and secretion are controlled by two different mechanisms in human MTC.

In conclusion, our data show that IGF-I protects MTC primary cultures from the effects of everolimus and stimulates CT

secretion by an mTOR mediated pathway that, in turn, may represent a therapeutic target in the treatment of aggressive MTCs. A synergistic effect of IGF-IR inhibitors and everolimus has already been hypothesized (29, 30). Indeed, mTOR inhibition increases Akt activity via the IGF-IR pathway, leading to the reduction of mTOR inhibitor effects (30). Therefore, IGF-IR signaling inhibition may prevent this positive feedback mechanism. The results of ongoing clinical trial employing both everolimus and the somatostatin receptor pan-agonist Pasireotide (SOM230 alone or in combination with RAD001 in patients with medullary thyroid cancer; NCT01625520) may help in clarifying this issue, also on the basis of the potent inhibitory effects of Pasireotide on IGF-I secretion.

Author Contributions

EG: wrote the manuscript. CDP: performed protein expression profile. MR: performed primary cultures. FT: performed

calcitonin secretion assay. TG: performed cell viability assay. RR: provided patients information. MP: provided patients information and surgical specimens. IMB: provided patients information and surgical specimens. EdU: supervised patients information and *in vitro* experiments. MCZ: supervised patients data and *in vitro* experiments, and wrote the manuscript.

Acknowledgments

We thank Novartis Pharma Italy for providing everolimus. **Funding:** This work was supported by grants from the Italian Ministry of Education, Research and University (FIRB RBAP11884M, RBAP1153LS, 2010TYCL9B_002), Fondazione Cassa di Risparmio di Ferrara, and Associazione Italiana per la Ricerca sul Cancro (AIRC) in collaboration with Laboratorio in rete del Tecnopolo “Tecnologie delle terapie avanzate” (LTTA) of the University of Ferrara, Ferrara.

References

- Höppener JW, Steenbergh PH, Slebos RJ, de Pagter-Holthuisen P, Roos BA, Jansen M, et al. Expression of insulin-like growth factor-I and -II genes in rat medullary thyroid carcinoma. *FEBS Lett* (1987) **215**:122–6. doi:10.1016/0014-5793(87)80125-4
- van der Laan BF, Freeman JL, Asa SL. Expression of growth factors and growth factor receptors in normal and tumorous human thyroid tissues. *Thyroid* (1995) **5**:67–73. doi:10.1089/thy.1995.5.67
- Okimura Y, Kitajima N, Uchiyama T, Yagi H, Abe H, Shakutsui S, et al. Insulin-like growth factor I (IGF-I) production and the presence of IGF-I receptors in rat medullary thyroid carcinoma cell line 6-23 (clone 6). *Biochem Biophys Res Commun* (1989) **161**:589–95. doi:10.1016/0006-291X(89)92640-5
- Yang KP, Samaan NA, Liang YF, Castillo SG. Role of insulin-like growth factor-I in the autocrine regulation of cell growth in TT human medullary thyroid carcinoma cells. *Henry Ford Hosp Med J* (1992) **40**:293–5.
- Mitsiades CS, McMillin D, Kotoula V, Poulaki V, McMullan C, Negri J, et al. Antitumor effects of the proteasome inhibitor bortezomib in medullary and anaplastic thyroid carcinoma cells in vitro. *J Clin Endocrinol Metab* (2006) **91**:4013–21. doi:10.1210/jc.2005-2472
- Wells SA Jr, Pacini F, Robinson BG, Santoro M. Multiple endocrine neoplasia type 2 and familial medullary thyroid carcinoma: an update. *J Clin Endocrinol Metab* (2013) **98**:3149–64. doi:10.1210/jc.2013-1204
- Gagliano T, Bellio M, Gentilin E, Molè D, Tagliati F, Schiavon M, et al. mTOR, p70S6K, AKT, and ERK1/2 levels predict sensitivity to mTOR and PI3K/mTOR inhibitors in human bronchial carcinoids. *Endocr Relat Cancer* (2013) **20**:463–75. doi:10.1530/ERC-13-0042
- Zatelli MC, Minoia M, Filieri C, Tagliati F, Buratto M, Ambrosio MR, et al. Effect of everolimus on cell viability in nonfunctioning pituitary adenomas. *J Clin Endocrinol Metab* (2010) **95**:968–76. doi:10.1210/jc.2009-1641
- Zatelli MC, Minoia M, Martini C, Tagliati F, Ambrosio MR, Schiavon M, et al. Everolimus as a new potential antiproliferative agent in aggressive human bronchial carcinoids. *Endocr Relat Cancer* (2010) **17**:719–29. doi:10.1677/ERC-10-0097
- Yao JC, Shah MH, Ito T, Bohas CL, Wolin EM, Van Cutsem E, et al. Everolimus for advanced pancreatic neuroendocrine tumors. *N Engl J Med* (2011) **364**:514–23. doi:10.1056/NEJMoa1009290
- Halperin DM, Kulke MH, Yao JC. A tale of two tumors: treating pancreatic and extrapancreatic neuroendocrine tumors. *Annu Rev Med* (2015) **66**:1–16. doi:10.1146/annurev-med-061813-012908
- Grozinsky-Glasberg S, Rubinfeld H, Nordenberg Y, Gorshtein A, Praiss M, Kendler E, et al. The rapamycin-derivative RAD001 (everolimus) inhibits cell viability and interacts with the Akt-mTOR-p70S6K pathway in human medullary thyroid carcinoma cells. *Mol Cell Endocrinol* (2010) **315**:87–94. doi:10.1016/j.mce.2009.09.027
- Faggiano A, Ramundo V, Dicitore A, Castiglioni S, Borghi MO, Severino R, et al. Everolimus is an active agent in medullary thyroid cancer: a clinical and in vitro study. *J Cell Mol Med* (2012) **16**:1563–72. doi:10.1111/j.1582-4934.2011.01438.x
- Zatelli MC, Piccin D, Tagliati F, Bottoni A, Luchin A, Vignali C, et al. Selective activation of somatostatin receptor subtypes differentially modulates secretion and viability in human medullary thyroid carcinoma primary cultures: potential clinical perspectives. *J Clin Endocrinol Metab* (2006) **91**:2218–24. doi:10.1210/jc.2006-0334
- Zatelli MC, Luchin A, Piccin D, Tagliati F, Bottoni A, Vignali C, et al. Cyclooxygenase-2 inhibitors reverse chemoresistance phenotype in medullary thyroid carcinoma by a permeability glycoprotein-mediated mechanism. *J Clin Endocrinol Metab* (2005) **90**:5754–60. doi:10.1210/jc.2005-1362
- Gentilin E, Tagliati F, Terzolo M, Zoli M, Lapparelli M, Minoia M, et al. Mitotane reduces human and mouse ACTH-secreting pituitary cell viability and function. *J Endocrinol* (2013) **218**:275–85. doi:10.1530/JOE-13-0210
- Gagliano T, Gentilin E, Benfina K, Di Pasquale C, Tassinari M, Falletta S, et al. Mitotane enhances doxorubicin cytotoxic activity by inhibiting P-gp in human adrenocortical carcinoma cells. *Endocrine* (2014) **47**:943–51. doi:10.1007/s12020-014-0374-z
- Lim SM, Chang H, Yoon MJ, Hong YK, Kim H, Chung WY, et al. A multicenter, phase II trial of everolimus in locally advanced or metastatic thyroid cancer of all histologic subtypes. *Ann Oncol* (2013) **24**:3089–94. doi:10.1093/annonc/mdt379
- Mulet-Margalef N, Capdevila J. Critical appraisal of the role of everolimus in advanced neuroendocrine tumors of pancreatic origin. *Gastrointest Cancer: Targets and Therapy* (2012) **2**:29–37. doi:10.2147/GICTT.S24826
- Moreno A, Akcakanat A, Munsell MF, Soni A, Yao JC, Meric-Bernstam F. Antitumor activity of rapamycin and octreotide as single agents or in combination in neuroendocrine tumors. *Endocr Relat Cancer* (2008) **15**:257–66. doi:10.1677/ERC-07-0202
- Molè D, Gentilin E, Gagliano T, Tagliati F, Bondanelli M, Pelizzo MR, et al. Protein kinase C: a putative new target for the control of human medullary thyroid carcinoma cell proliferation in vitro. *Endocrinology* (2012) **153**:2088–98. doi:10.1210/en.2011-1988
- Tamburrino A, Molinolo AA, Salerno P, Chernock RD, Raffeld M, Xi L, et al. Activation of the mTOR pathway in primary medullary thyroid carcinoma and lymph node metastases. *Clin Cancer Res* (2012) **18**:3532–40. doi:10.1158/1078-0432.CCR-11-2700
- Noh WC, Mondesire WH, Peng J, Jian W, Zhang H, Dong J, et al. Determinants of rapamycin sensitivity in breast cancer cells. *Clin Cancer Res* (2004) **10**:1013–23. doi:10.1158/1078-0432.CCR-03-0043
- Wong CK, Lai T, Holly JM, Wheeler MH, Stewart CE, Farndon JR. Insulin-like growth factors (IGF) I and II utilize different calcium signaling pathways in a primary human parathyroid cell culture model. *World J Surg* (2006) **30**:333–45. doi:10.1007/s00268-005-0339-8

25. Freichel M, Zink-Lorenz A, Holloschi A, Hafner M, Flockerzi V, Raue F. Expression of a calcium-sensing receptor in a human medullary thyroid carcinoma cell line and its contribution to calcitonin secretion. *Endocrinology* (1996) **137**:3842–8. doi:10.1210/en.137.9.3842
26. Zuloaga R, Fuentes EN, Molina A, Valdés JA. The cAMP response element binding protein (CREB) is activated by insulin-like growth factor-1 (IGF-1) and regulates myostatin gene expression in skeletal myoblast. *Biochem Biophys Res Commun* (2013) **440**:258–64. doi:10.1016/j.bbrc.2013.09.067
27. Zatelli MC, Tagliati F, Piccin D, Taylor JE, Culler MD, Bondanelli M, et al. Somatostatin receptor subtype 1-selective activation reduces cell growth and calcitonin secretion in a human medullary thyroid carcinoma cell line. *Biochem Biophys Res Commun* (2002) **297**:828–34. doi:10.1016/S0006-291X(02)02307-0
28. Druce M, Chung TT, Grozinsky-Glasberg S, Gross DJ, Grossman AB. Preliminary report of the use of everolimus in a patient with progressive medullary thyroid carcinoma. *Clin Endocrinol (Oxf)* (2012) **77**:154–5. doi:10.1111/j.1365-2265.2011.04296.x
29. Quek R, Wang Q, Morgan JA, Shapiro GI, Butrynski JE, Ramaiya N, et al. Combination mTOR and IGF-1R inhibition: phase I trial of everolimus and figitumumab in patients with advanced sarcomas and other solid tumors. *Clin Cancer Res* (2011) **17**:871–9. doi:10.1158/1078-0432.CCR-10-2621
30. Yim KL. Everolimus and mTOR inhibition in pancreatic neuroendocrine tumors. *Cancer Manag Res* (2012) **4**:207–14. doi:10.2147/CMAR.S25979

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2015 Gentilin, Di Pasquale, Rossi, Tagliati, Gagliano, Rossi, Pelizzo, Merante Boschin, degli Uberti and Zatelli. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.