



Thyroid hormone and seasonal rhythmicity

Hugues Dardente^{1,2,3,4*}, David G. Hazlerigg⁵ and Francis J. P. Ebling⁶

¹ Physiologie de la Reproduction et des Comportements, INRA, UMR085, Nouzilly, France

² CNRS, UMR7247, Nouzilly, France

³ Université François Rabelais de Tours, Tours, France

⁴ Institut français du cheval et de l'équitation, Nouzilly, France

⁵ Department of Arctic and Marine Biology, University of Tromsø, Tromsø, Norway

⁶ School of Life Sciences, University of Nottingham, Nottingham, UK

Edited by:

Frédéric Flamant, Ecole Normale Supérieure de Lyon, France

Reviewed by:

Valerie Simonneaux, Centre National de la Recherche Scientifique, France
Sulay Tovar, University of Cologne, Germany

*Correspondence:

Hugues Dardente, INRA, UMR85
Physiologie de la Reproduction et des Comportements, CNRS, UMR7247,
Université François Rabelais de Tours,
IFCE, F-37380 Nouzilly, France
e-mail: hdardente@tours.inra.fr

Living organisms show seasonality in a wide array of functions such as reproduction, fattening, hibernation, and migration. At temperate latitudes, changes in photoperiod maintain the alignment of annual rhythms with predictable changes in the environment. The appropriate physiological response to changing photoperiod in mammals requires retinal detection of light and pineal secretion of melatonin, but extraretinal detection of light occurs in birds. A common mechanism across all vertebrates is that these photoperiod-regulated systems alter hypothalamic thyroid hormone (TH) conversion. Here, we review the evidence that a circadian clock within the pars tuberalis of the adenohypophysis links photoperiod decoding to local changes of TH signaling within the medio-basal hypothalamus (MBH) through a conserved thyrotropin/deiodinase axis. We also focus on recent findings which indicate that, beyond the photoperiodic control of its conversion, TH might also be involved in longer-term timing processes of seasonal programs. Finally, we examine the potential implication of kisspeptin and RFRP3, two RF-amide peptides expressed within the MBH, in seasonal rhythmicity.

Keywords: seasonality, reproduction, pars tuberalis, melatonin rhythm, kisspeptins, RF-amide, GnRH neurons

INTRODUCTION

Seasonality is a critical property of most organisms. At temperate latitudes, photoperiod is the main synchronizer of seasonal functions. Photoperiodism defines the use of the annual cycle of day and night length to coordinate functions such as reproduction, fattening, hibernation, and migration with predictable changes in the environment, for example in food availability or climatic conditions. Seasonal changes in physiology and behavior typically are innately timed long-term processes, requiring weeks or months to wax and wane. Therefore, additional to photoperiodic readout mechanisms, living creatures have evolved endogenous long-term timing devices, which allow them to anticipate forthcoming seasonal changes. In the most extreme cases, cycles of about 365 days recur for years in animals kept under constant photoperiods; such so-called circannual rhythms exist in a variety of birds and longer-lived mammals.

Species with relatively short life spans such as voles and hamsters usually do not display circannual rhythms, but their seasonal cycles also comprise an endogenously generated part, which corresponds to the overwintering period and allows timely emergence from the burrow and reproductive recrudescence in early spring. Endogenous long-term timing is commonplace in vertebrates but its mechanistic basis remains mysterious [for reviews, see Ref. (1–6)]. Here we review findings, essentially in birds and mammals, which clarify the mechanisms of photoperiodic readout and provide a rationale for the seasonal control of thyroid hormone (TH) metabolism within the hypothalamus.

PHOTOPERIODISM: MELATONIN AND THE PARS TUBERALIS

The crucial role of melatonin in mammalian photoperiodism has been established in many species including hamsters, ferrets, and sheep (7–9). Within the pineal, melatonin is produced and released during the night and therefore constitutes an internal neurochemical representation of photoperiod. Timed melatonin-infusion experiments established that duration is the key parameter of the melatonin pattern that triggers the photoperiodic response [for review, see Ref. (10)]. In order to map central binding sites, autoradiography with 2-iodo-melatonin was used in a wide range of mammals (11). Surprisingly, across all species the highest density of melatonin-binding sites was found in the *pars tuberalis* (PT), a region of the pituitary stalk apposed to the median eminence. The suprachiasmatic nuclei (SCN) also showed moderate labeling in most species while many brain nuclei showed weak to moderate labeling, with very little species overlap [for reviews, see Ref. (12, 13)]. The presence of melatonin receptors within the SCN was consistent with the effects of melatonin on daily timing in mammals (14). Conversely, since the PT was the only neuroendocrine structure labeled in the highly photoperiodic ferret, a role in seasonality was anticipated (15). However, melatonin-binding sites were also disclosed within the PT of species, which are not overtly photoperiodic such as mouse, rat, and human.

Melatonin-binding studies also led to the recognition that the binding site(s) for melatonin was a classical GPCR, with picomolar affinity for its ligand. In mammals, two high-affinity melatonin receptors (MT1 and MT2) were cloned (16, 17). Subsequent studies showed that MT1 is the predominant subtype, both necessary

and sufficient to mediate the photoperiodic effect of melatonin (18–22). The number of central sites expressing melatonin receptors as revealed by *in situ* hybridization was comparatively more restricted – mostly the PT and the SCN – than that observed with melatonin-binding studies. This may reflect the difference in sensitivity of the techniques and/or the existence of a low-affinity melatonin-binding site. The latter would be physiologically irrelevant, and probably corresponds to quinone reductase 2 rather than a true melatonin receptor (23).

MELATONIN-DEPENDENT TSH RELEASE IN THE PARS TUBERALIS

The PT is the most rostral part of the adenohypophysis. Many reviews detailing the ontogeny, morphology, and immunohistochemical characteristics of the PT are available (24–28). The PT was once considered an “undifferentiated embryological remnant of the hypophysis” whose “only function is to provide mechanical support role for the hypothalamo-hypophyseal portal vessels” [see Ref. (29)]. However, its location and anatomical features pleaded in favor of a specific role: the PT extends along the ventral aspect of the median eminence, surrounds the pituitary stalk in its most caudal part, and is in contact with nerve endings of the median eminence and capillaries of the pituitary primary plexus.

The PT is phylogenetically conserved in tetrapods, but is generally absent in fish (30), and consists of endocrine cells, which exhibit early secretory activity compared to the pars distalis (PD). Three different cell types occur in the PT: (i) follicular cells; (ii) gonadotropes, which constitute ~10% of the endocrine PT cells, have dense-core granules and occur mostly in the caudal PT (known as the *zona tuberalis*); (iii) PT-specific cells, which are virtually agranular thyrotropes and constitute ~90% of endocrine PT cells. The PT gonadotropes appear identical to those in the PD, while shape and ultrastructure of PT-specific thyrotropes differ strikingly from those in the PD (24, 25). These thyrotropes were therefore suspected to be a peculiar pituitary endocrine cell type, possibly producing a novel glycoprotein [“tuberalin,” Ref. (31)]. These cells exhibit early secretory activity compared to PD endocrine cells (32). This depends upon the induction of *Tsh β* transcription by a transcription factor consequently called TEF [Thyrotroph Embryonic Factor; Ref. (33)].

Based on ultrastructure and immunohistochemistry, these PT-specific thyrotropes were predicted to be melatonin-responsive, a prediction which has since been validated (34, 35). TSH immunoreactivity within these cells displays dramatic melatonin-dependent photoperiodic changes, with high and low levels under long (LP) and short photoperiod (SP), respectively (36, 37). Finally, the TSH produced by these PT-specific thyrotropes may be identical to that produced by the PD, but the transcriptional control of the *Tsh β* gene in the two populations differs since PT thyrotropes do not express receptors for either TRH or TH (38). Hence, *Tsh β* expression by PT-specific thyrotropes is disconnected from the classical hypothalamic–pituitary–thyroid axis; instead it depends upon melatonin.

However, considering the Harris dogma of a descending flow of information from the hypothalamus to the pituitary, a role for PT-derived TSH was not forthcoming. Rather, it was assumed that, should the PT play a role in seasonality, it would most probably be

to release tuberalin(s) in the pituitary portal plexus, which would then target the PD. This might be the case for the seasonal control of the lactotropic axis, even though the mechanism is unclear (39). This aspect will not be considered further here as it has been discussed elsewhere (28, 40–42).

THYROID HORMONE SIGNALING IN SEASONAL CYCLES

AN OVERVIEW

The pioneering work of Benoit on ducks in the 1930s revealed that the thyroid gland is mandatory for seasonal transitions in reproductive states, a finding which applies to a wide range of vertebrates [reviewed by Nicholls et al. (43); Hazlerigg and Loudon (44); Yoshimura (45)]. Thyroidectomy prevents the cessation of breeding in starlings (46), quail (47), and sheep [Ref. (43, 48, 49); for review, see Ref. (50)]. In rams, thyroidectomy during the non-breeding season almost immediately reactivates the gonadotropic axis (51). Therefore, TH appeared to transmit the message of long-day lengths. Microimplants releasing small amount of TH were then surgically placed within the brain of the ewe (52, 53), which revealed that TH acts centrally, and most likely within the medio-basal hypothalamus (MBH), to impact seasonal reproduction. Studies in Siberian hamsters using a similar microimplants approach further showed that other seasonal axes are also controlled by central actions of T3: providing T3 directly within the MBH overrides the SP-induced inactivation of the gonadotropic axis (54) and triggers premature gonadal recrudescence in SP-exposed animals. T3 implants also override SP-induced seasonal inappetence, weight loss, and expression of torpor [Ref. (55); see **Figure 1**]. Similar outcomes are found when T3 is provided by daily subcutaneous injections to SP-exposed hamsters (56). In contrast to these effects on reproduction and energy metabolism, T3 implants do not impact the lactotropic axis, consistent with a distinct mechanism of control (57, 58) while not incompatible with a common melatonin target tissue as discussed later.

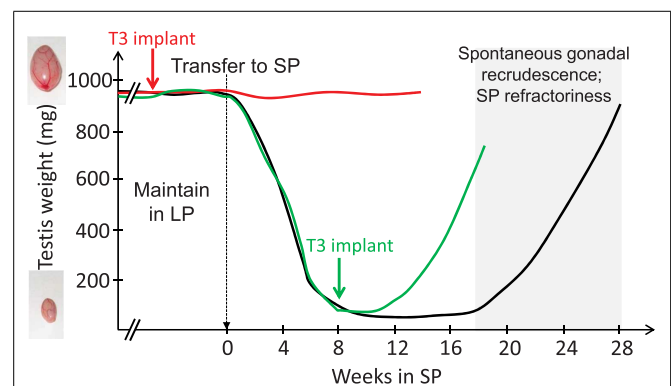


FIGURE 1 | T3 implants prevent SP-induced inactivation of the gonadal axis (red line) and reactivate the gonadal axis in SP-adapted Siberian hamsters (green line; after data from Barrett et al. (54) and Murphy et al. (55)). Siberian hamsters (black line) kept in LP remain indefinitely sexually active (broken lines) unless they are transferred to SP; gonads then progressively regress (testes depicted here, but data are similar for female reproductive organs). However, prolonged SP exposure leads to a spontaneous recrudescence of the gonads, which reflects SP-refractoriness.

These observations added to the well-documented role of TH in key transitions between life cycles, such as metamorphosis in amphibians and developmental growth and differentiation of the mammalian brain (59, 60). In adults, TH also has key roles in the control of metabolism and thermoregulation, two processes intertwined with the seasonal reproductive cycle. The seasonal program encompasses profound and coordinated changes in behavioral, reproductive, and metabolic states (61). The finding that TH regulates the basal metabolic rate is not new, but the recognition that it reflects a central action within the MBH is very recent (62, 63). Indeed, T3 injection within the MBH suffices to promote food intake and weight gain in rats (64). Interestingly, this effect is mimicked by LP, which triggers weight gain in many species, including sheep (65). Such a process bears critical adaptive value, best exemplified in species that hibernate (e.g., groundhog) or undergo daily torpor (e.g., Siberian hamster), which have evolved a strategy to build up abdominal fat depots during spring/summer to survive the harsh winter season (1, 61). Photoperiodic cues and the metabolic status interact in many seasonal breeders, including sheep (66, 67), goats (68, 69), and horses (70). In all these species, feeding modulates the duration of the breeding season and/or depth of the anestrus. Therefore, TH integrates and coordinates physiological changes, which are integral to the seasonal program.

LOCAL CONTROL OF TH METABOLISM WITHIN THE MBH

Although cold exposure activates thyroid activity, under constant ambient temperature conditions, TH concentrations do not display marked or consistent seasonal fluctuations in the plasma or cerebro-spinal fluid. Rather, fine temporal and local control of TH action is achieved through opposite actions of specific enzymes known as deiodinases (71–73). Deiodinase 2 (DIO2) converts the relatively inactive T4 into the active T3 while deiodinase 3 (DIO3) inactivates T4 by converting it into rT3, and also degrades T3 into T2. Very precise control of T3 concentrations is further achieved through reciprocal control of the expression and activity of these two enzymes by their ligand: a hypothyroid state up-regulates DIO2 and down-regulates DIO3, and vice-versa (72, 74, 75).

The central expression of *Dio2* is restricted to a few structures. The pineal gland is one of them (76), but the strongest expression occurs in astrocytes and tanycytes lining the third ventricle and median eminence (77, 78). These tanycytes also express two major TH transporters, MCT8 and OATP1c1 (79–81), and MCT8 is expressed at higher levels under SP than LP in the Siberian hamster (82, 83). Tanycytes are a heterogeneous and complex population of ependymal cells, which constitute a gateway between the CSF and the MBH and median eminence (84). In a pioneering study, Yoshimura and colleagues (85) showed that both *Dio2* and *Dio3* are expressed within tanycytes of the quail MBH. Crucially, the expression of these two enzymes displays opposite regulation by photoperiod: *Dio2* is highly expressed under LP while *Dio3* is highly expressed under SP. This predicted a local increase of T3 content within the MBH under LP, which was validated by radioimmunoassay (85). The opposite regulation of *Dio2* and *Dio3* by photoperiod has since been described in sparrows and Siberian and Syrian hamsters (86–89). Importantly, the expression of *Dio2* is down-regulated by melatonin, independently of sex steroids (88, 90). Melatonin is also required to trigger *Dio3* expression under

SP in Siberian hamster (54). Collectively, these data provided an enzymatic means through which local T3 levels in the MBH could increase under LP.

CLOSING THE LOOP: TSH OUTPUT FROM THE PT GOVERNS T3 REGULATION WITHIN THE MBH

The PT seemed well located to mediate photoperiodic switches in *Dio2–Dio3* usage. To decipher the mechanism of the photoperiodic response, Yoshimura and colleagues (91) set out an ambitious experimental set-up: hypothalamic blocks containing the MBH and PT/median eminence from quails submitted to a long-day transfer, known to activate the gonadotropic axis within 24 h, were used for hybridization on a chicken gene chip. This revealed that the expression of two genes, *Tsh β* and *Eya3*, is rapidly triggered by the transfer from SP to LP. A second wave of transcriptional changes was also observed for a handful of genes including *Dio2* and *Dio3*, which displayed acute and simultaneous induction and repression, respectively. Crucially, expression of the cognate TSH receptor (TSHR) was found in tanycytes, which express the deiodinases, providing the link between TSH output from the PT and T3 regulation within the MBH. The pathway was uncovered using an acute intracerebroventricular injection of TSH to SP-exposed quails, which induced *Dio2* expression and led to gonadal recrudescence.

In a contemporaneous study in sheep, Hanon et al. (92) suggested this mechanism to be ancestral, since their data were similar in many respects: higher *Tsh β* expression within the PT under LP than SP (see **Figure 3A**), expression of the TSHR within tanycytes and PT/median eminence, higher *Dio2* expression under LP than SP (see **Figure 3A**), and TSH-dependent induction of *Dio2* both *in vitro* and *in vivo*. The latter finding was not unexpected, since TSHR signals through a Gs protein, and *Dio2* is a cAMP-responsive gene (93). In contrast, the MT1 receptor couples to a Gi protein and the interplay between TSHR and MT1 signaling within the PT may be part of the photoperiod decoding mechanism, at least in sheep (92, 94, 95). Under LP, the PT therefore functions as an “indirect T3-generator,” disconnected from both TRH and T3 feedback (see above).

Since these studies in quail and sheep, a similar TSH/deiodinases/T3 retrograde pathway (from the pituitary back to the hypothalamus, **Figure 2**) has been described not only in other photoperiodic species such as the European hamster (96), the Syrian hamster (97), the Siberian hamster (89), the common vole (98), but also in photoresponsive juvenile Fisher 344 rats (99) and in a melatonin-producing but non-photoperiodic CBA/N

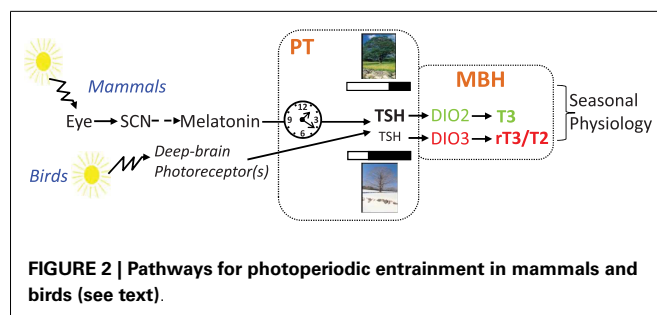


FIGURE 2 | Pathways for photoperiodic entrainment in mammals and birds (see text).

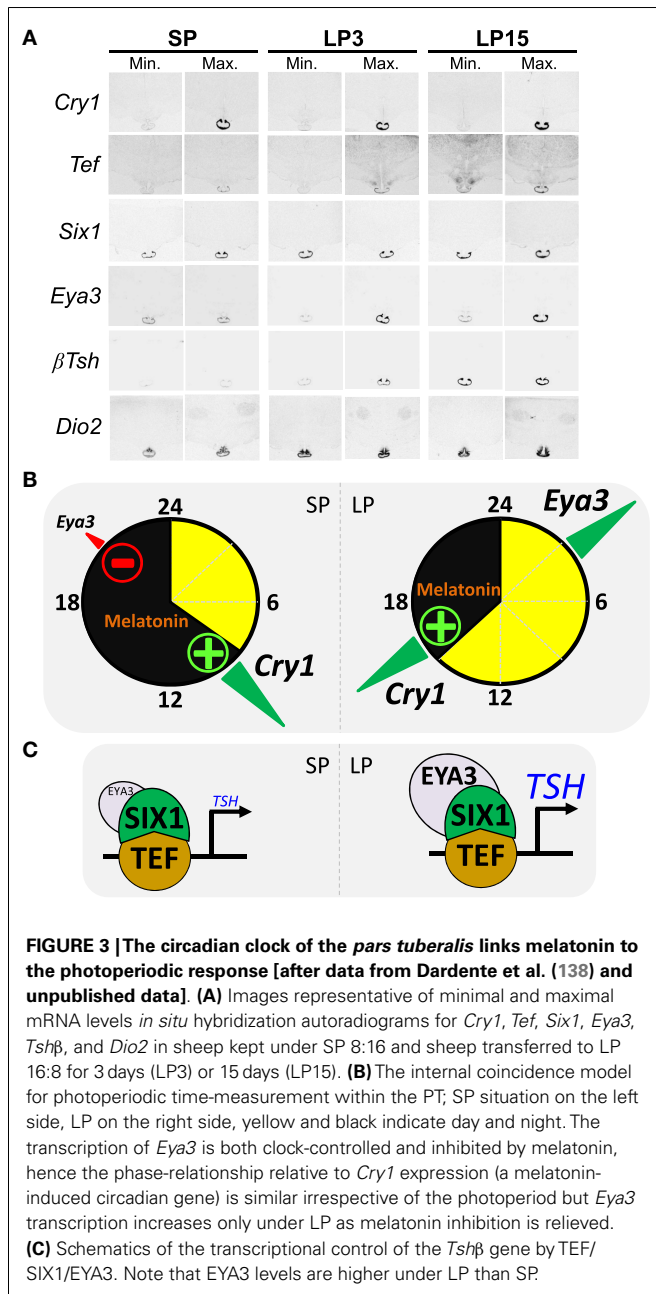


FIGURE 3 | The circadian clock of the *pars tuberalis* links melatonin to the photoperiodic response [after data from Dardente et al. (138) and unpublished data]. (A) Images representative of minimal and maximal mRNA levels *in situ* hybridization autoradiograms for *Cry1*, *Tef*, *Six1*, *Eya3*, *Tsh β* , and *Dio2* in sheep kept under SP 8:16 and sheep transferred to LP 16:8 for 3 days (LP3) or 15 days (LP15). (B) The internal coincidence model for photoperiodic time-measurement within the PT; SP situation on the left side, LP on the right side, yellow and black indicate day and night. The transcription of *Eya3* is both clock-controlled and inhibited by melatonin, hence the phase-relationship relative to *Cry1* expression (a melatonin-induced circadian gene) is similar irrespective of the photoperiod but *Eya3* transcription increases only under LP as melatonin inhibition is relieved. (C) Schematics of the transcriptional control of the *Tsh β* gene by TEF/SIX1/EYA3. Note that EYA3 levels are higher under LP than SP.

mouse strain (22, 100, 101). Photoperiodic variations in *Dio2* expression were however not observed in the non-photoperiodic Wistar rat (88). The use of murine knock-out strains confirmed that the MT1 melatonin receptor (22) and TSHR (100) are mandatory for the LP induction of *Dio2* expression within tanycytes. Whether this pathway is present in all vertebrates remains to be determined (102).

As mentioned before, fish species investigated thus far do not have a distinct PT, but in masu salmon a TSH/DIO2 axis implicating the *saccus vasculosus*, located below the hypothalamus and caudally to the pituitary gland, has been proposed (103). However, the *saccus vasculosus* is absent in several species of fish such as the pike (104), which is nonetheless photoperiodic (105).

The other few studies on this matter in fish have yielded varied outcomes (106, 107). Regarding birds, studies in tits (108) and starlings (109) did not lend clear support to the model, but aspects of the experimental set-up prevent any conclusion to be drawn. For example, the studies of starlings were carried out in outdoor aviaries, so effects of fluctuating temperature on the peripheral thyroid axis may have obscured the photoperiodic regulation of DIO2 and DIO3 centrally. Finally, we are not aware of any study on this matter in either reptiles or amphibians.

ENCODING AND DECODING THE PHOTOPERIODIC MESSAGE UPSTREAM OF THE PT

Birds and mammals possess a similar mechanism to respond to photoperiod, but they perceive the photoperiodic message in different ways. In mammals, light is exclusively perceived by the retina, with a key role for ganglion cells expressing the photopigment melanopsin [for review, see Ref. (110)]. This information is relayed to the circadian clock of the SCN, which governs melatonin production by the pineal gland through a multi-synaptic sympathetic pathway. Melatonin is the mandatory messenger of photoperiod in mammals. In striking contrast, removing the eyes and suppressing melatonin by pinealectomy does not disrupt photoperiodism in birds [for reviews, see Ref. (44, 45, 111–113)]. In birds, light goes through the skull and acts directly upon hypothalamic deep-brain photoreceptors to control seasonal reproduction (Figure 2). Several photopigments expressed by different cell types, all located within the MBH and projecting to the PT/median eminence, are plausible candidates: VA-opsin (114), neuropsin [Opn5, Ref. (115, 116)], and melanopsin [Opn4, Ref. (117)]. The neurotransmitter(s) and/or neuropeptide(s) used by these cells, and how they impinge on PT thyrotropes, remain to be elucidated.

WITHIN THE PT: FROM THE CIRCADIAN CLOCK TO THE SEASONAL OUTPUT

Photoperiodic species such as quail (118) and Siberian and Syrian hamsters (119, 120) measure photoperiod length with remarkable accuracy. In these three species, reproduction switches off when the photoperiod is shorter than 12.5 h. The narrow photoperiod range over which physiological changes occur is one of the lines of evidence implicating some sort of daily timing device. The concept that circadian clock(s), clocks with a period of about 24 h, control seasonal timing is indeed not novel (120, 121).

The genetic and molecular bases and organization of circadian clocks have been recently identified (122–124). These clocks are not only located within the SCN, but are present in virtually every tissue and cell where they impact “local” physiology. The PT is no exception as it expresses a full set of clock genes and displays persistent circadian rhythmicity *in vitro* [Ref. (125–127); for review, see Ref. (28)]. The SCN and peripheral clocks share fundamental characteristics: they are cell-autonomous and self-sustained. However, individual cellular clocks within peripheral tissues rapidly become desynchronized and exhibit phase drifting in the absence of regular resetting by cues emanating, directly or indirectly, from the SCN. These cues include inputs from the autonomic nervous system, temperature cycles, and humoral factors such as glucocorticoids and melatonin.

The PT can be defined as a melatonin-dependent circadian oscillator (28, 50). Resetting of the PT clock by melatonin requires acute induction of *Cry1* expression [Ref. (128, 129), see **Figure 3A**]; *CRY1* being a key repressor of the circadian clock (130–132). The acute induction of *Cry1* expression involves EGR1-like factors (133) and the transcription factor NPAS4 (134, 135). In sheep, *Cry1* expression remains tightly linked to the onset of melatonin secretion and by implication night onset, irrespective of the duration of the day length [Ref. (136), see **Figure 3B**]. Interestingly, light given during the night induces *Cry1* expression within the quail PT (137), which suggests a phylogenetically conserved role for *Cry1* in the photoperiodic resetting of the PT clock.

How do we connect melatonin resetting of the PT clock with differential photoperiodic output of TSH and seasonal reproduction? The expression of the transcriptional co-activator *EYA3* within the ovine PT displays large photoperiodic changes in both phase and amplitude [Ref. (39, 138); see **Figure 3A**]. Interestingly, *Eya3* was the other gene (besides *Tsh β*) immediately induced in the quail PT during the first long-day release experiment (91). We therefore investigated the transcriptional control of *Eya3* and searched for a link between inductions of both genes. The expression of *Eya3* is clock-controlled, through conserved DNA binding motifs within its promoter, and therefore phase-locked to that of the circadian clock [Ref. (138), see **Figure 3B**]. Because of this, expression peaks during the night under SP but during the day under LP. The amplitude of the peak is higher under LP than SP because melatonin suppresses *Eya3* expression, a suppression which can only occur in SP-exposed animals [Ref. (138); see **Figure 3B**]. Finally, *in vitro* data showed that induction of *Tsh β* expression is triggered by the circadian-controlled transcription factor TEF (33), which then recruits the co-activators SIX1 and *EYA3*. This leads to a marked increase in transcription under LP due to higher levels of *EYA3* [Ref. (102, 138), **Figures 3B,C**]. A critical role for SIX1/*EYA3*, but not TEF, in the photoperiodic control of *Tsh β* transcription in the mouse PT has been proposed (139, 140).

IS T3 OUTPUT SUFFICIENT TO ELICIT THE FULL SPECTRUM OF SEASONAL CHANGES?

The data reviewed so far are consistent with a crucial role for the TSH output of the PT in driving seasonal changes in T3 availability within the MBH. However, swings in TSH/T3 may not be sufficient to elicit all seasonal changes. As mentioned before, since control of the lactotropic axis does not depend on T3 [for review, see Ref. (50)], complementary mechanisms are indeed expected. Neuromedin U (89, 141), histamine, and VGF secretion (82, 142, 143) may mediate seasonal effects on body weight and metabolism since their synthesis and cognate receptors display expression patterns and seasonal changes reminiscent of those seen for TSH/TSHR. However, since TSH infusion in SP-adapted Siberian hamster restores hypothalamic expression of somatostatin and body weight to LP levels (144), Neuromedin U, histamine, or VGF may be dispensable.

Retinoic acid signaling is also likely to be involved as retinoic acid receptors, transporters, and associated binding proteins display prominent photoperiodic regulation in the ependymal cell layer and posterior arcuate nucleus of Siberian hamsters and

juvenile Fischer F344 rats (142, 145, 146). Interestingly, the retinoic X receptor (RXR) can heterodimerize with either the TH receptors (THR α /THR β) or the retinoic acid related receptor (RAR). The target genes and downstream pathways governed by THR and RAR diverge, and therefore the photoperiodic regulation of RXR/RAR may fine-tune the seasonal adaptation of the metabolic status. From a more general standpoint, the notion that tancytes coordinate a host of seasonal neuroendocrine cycles including reproduction, metabolism, and hibernation is emerging rapidly [Ref. (147, 148); for reviews, see Ref. (61, 149, 150)].

PHOTOPERIODIC TIMING AND THE CIRCANNUAL CLOCK: T3 AS A UNIFYING COMPONENT?

As mentioned earlier, whether species are classified as photoperiodic (e.g., Siberian and Syrian hamsters) or circannual (e.g., sheep), part of the seasonal cycle is generated endogenously. Hamsters and sheep maintained under constant SP do spontaneously revert to the opposite physiological state after several months. This phenomenon, referred to as “SP refractoriness,” is typical of an interval timer/hourglass (5, 151). In contrast, sheep but not Siberian or Syrian hamsters, also display refractoriness to LP. Whether this species difference reflects fundamentally divergent underlying mechanisms is questionable. Indeed, Follett and Nicholls (47) proposed years ago that “it may well be that essentially identical physiological mechanisms underlie the photoperiodic responses of a wide range of vertebrates and that very minor modifications of these can cause surprisingly large (though superficial) changes in the overt responses of the animal in terms of reproduction.” These authors devised a model, based on differences in threshold sensitivity, which rationalizes the LP refractoriness process (see **Figure 4**). There are indeed similarities between the photoperiodic control of the seasonal program in photoperiodic and circannual species (43, 50, 152). Siberian or Syrian hamsters and sheep might therefore exemplify “variations on a theme” rather than fundamentally different models.

Because TH is involved in many long-term life cycles events, it seems plausible that photoperiod-induced changes in T3 levels may also trigger more profound long-term changes, culminating weeks to months later. In particular, TH-induced plasticity and

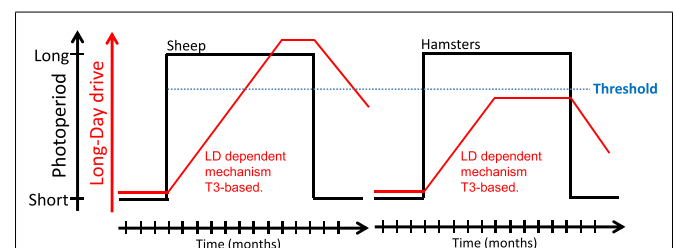


FIGURE 4 | A model for long-day refractoriness [adapted from Figure 2 in Ref. (152)]. In sheep (left panel) and hamsters (right panel), exposure to long days (black line) leads to the development of a mechanism of unknown nature, most likely T3-dependent (red line). In sheep, the long-day drive eventually exceeds a “threshold” (blue dotted line); the animal then becomes refractory to long days and spontaneously reverts to an SP phenotype. In hamsters, the long-day drive never exceeds the threshold and the animal displays the LP phenotype indefinitely; exposure to SP is mandatory to get the SP physiological state.

cell-cycle related events have long time constants, which appear compatible with seasonal cycles (6, 153). Recent data in sheep demonstrate a photoperiodic gating of cell division within the PT and ependymal cells of the 3V and are consistent with this scheme (154–156). Nevertheless, whether photoperiodic gating of cell division depends on TH and/or is involved in seasonal transitions remains to be established.

To address a potential role for TH turn-over beyond the photoperiodic response, we investigated the expression of *Tsh β* and *Dio2/Dio3* within the MBH of sheep under distinct physiological states: LP, LP refractory (LPR) obtained after prolonged LP exposure, SP and SP refractory (SPR) obtained after prolonged SP exposure (157). The expressions of *Tsh β* and *Dio2* were diminished in LPR compared to LP animals but remained low in SP and SPR animals. The expression of *Dio3* was high in SP but very low in all other photoperiodic conditions, most notably under SPR; so the expression of *Dio3* under SP is transient (see Figure 5A).

Therefore, a diminished TSH output may cause the LPR state, while development of the SPR state would be disconnected from it. This would be consistent with the hourglass properties of the SPR mentioned before. However, changes in *Dio2/Dio3* may reflect an indirect effect of photoperiod: within the MBH, the local hyperthyroid state triggered by persistent LP exposure would eventually cross a certain threshold, thereby triggering *Dio3* induction and *Dio2* down-regulation (72, 74, 75). Following this, T3 levels would

be cleared by DIO3, ultimately leading to the demise of *Dio3* expression; LP exposure would then somehow be required to induce *Dio2* once more and prime a new cycle. This LP requirement to prime the seasonal sequence may explain why circannual cycles in sheep are most obvious under constant LP (50).

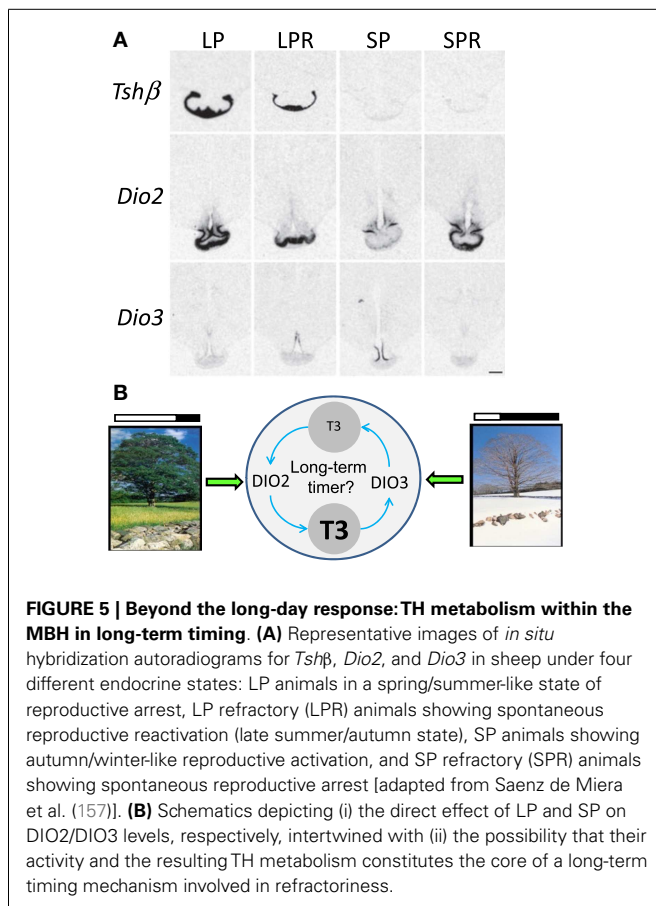
Interestingly, Syrian hamsters in SPR state do not exhibit spontaneous reactivation of *Dio2* expression (88) while Siberian hamsters do (83). Furthermore, Siberian hamsters express *Dio3* upon transfer from LP to SP but its expression is not sustained through time (54, 83, 87), similar to what occurs in sheep (157). Therefore, transient *Dio3* expression under SP appears as a common feature and may explain why *Dio3* expression has not been observed in Syrian hamster (54). Even though the relative variations of *Dio2* and *Dio3* differ between species (e.g., Siberian vs. Syrian hamster) the central tenet remains the same: T3 levels are higher within the MBH under LP compared to SP (158). Furthermore, TH metabolism within the MBH may not only intervene in the photoperiodic response but may also be integral to longer-term timing processes such as circannual rhythms (see Figure 5B).

CONCLUSION

At this stage several outstanding questions remain: first, since the same TSH/deiodinase/T3 pathway is triggered by LP not only in long-day breeders (e.g., hamsters and quail) but also in short-day breeders (e.g., sheep) and non-photoperiodic species (e.g., mouse), how do we get opposite responses, or no response at all, of the hypothalamic–pituitary–gonadal axis? This is particularly intriguing because the increased intra-hypothalamic availability of TH is uniformly linked to an anabolic state across seasonal species. Second, through which mechanisms do local changes of T3 within the MBH ultimately impinge on gonadotropin-releasing hormone neurons? Pertinent to this second question, the MBH hosts two cell populations expressing RF-amide peptides which have attracted particular attention: neurons of the arcuate nucleus, which express *Kiss1* and neurons of the VMH/DMH, which express the *Rfrp* precursor. The concept that these RF-amide peptides are involved in seasonal breeding has been the topic of several excellent reviews (50, 158–161) and we will therefore only briefly review the most recent and salient findings.

Kisspeptin is a very potent GnRH secretagogue and governs most aspects of reproduction in mammals including sexual differentiation, steroid-dependent gonadotropin release, puberty onset, and the control of fertility by metabolic cues (162, 163). Interestingly, the annual onset of fertility in photoperiodic species had been compared to a reoccurrence of puberty, and common underlying processes were anticipated (2, 164, 165). Kisspeptin therefore appeared a prime candidate for the integration of photoperiodic and metabolic cues across the seasonal program, a prediction which has now received strong support (159–161).

In contrast to kisspeptin, the exact role(s) of peptides derived from the *Rfrp* precursor, RFRP1 and RFRP3, remain(s) unclear (160). RFRP3 may modulate feeding and various stress responses (166, 167). In the context of breeding, RFRP3 inhibits GnRH in sheep [Ref. (168), but see Ref. (169)] but inhibits or activates GnRH in Syrian and Siberian hamsters, depending on the photoperiod (170, 171). The *Rfrp* gene is orthologous to avian *GnIH*, which gives rise to gonadotropin inhibitory hormone (GnIH),



a peptide with well-characterized inhibitory effects upon the gonadotropic axis in birds (172). Interestingly, there is no avian ortholog of the *Kiss1* (or *Kiss2*) gene (173), which implies that the concept of a balance between KISS1 and RFRP3 in governing GnRH secretion in mammals (174) does not apply to birds.

Both *Kiss1* and *Rfrp* expression display marked photoperiodic, melatonin-dependent, changes in mammals (159, 160, 175). Even though melatonin receptors have been localized to several hypothalamic nuclei it seems likely that the photoperiodic control over *Kiss1* and *Rfrp* is indirect [see above, Ref. (50, 145, 161)]. In this context, a role for PT-derived TSH appeared plausible. In a landmark study, Klosien et al. (144) showed that intracerebroventricular delivery of TSH in Siberian and Syrian hamsters induces *Dio2* expression within ependymal cells, restores expression of *Kiss1* and *Rfrp* to their LP levels and, most importantly, triggers reactivation of the gonadal axis. Furthermore, Henson et al. (176) showed that T3 injections to SP-adapted Siberian hamsters reactivated the gonadotropic axis, thereby confirming prior data (see Section “An Overview” and **Figure 1**), but also led to LP-like levels of RF-amide peptides within the MBH.

Therefore, even though a theoretical possibility exists that another TSH-dependent – but T3-independent pathway – leads to seasonal changes of the reproductive axis, the most parsimonious model is one in which T3 action on RF-amide neurons link the photoperiodic production of TSH within the PT to the seasonal control of GnRH secretion.

ACKNOWLEDGMENTS

Hugues Dardente is supported by a Marie Curie Career Integration Grant from the FP7-People-2012 program. Hugues Dardente wishes to thank Paul Pévet (INCI, Strasbourg, France) for continuing support and exciting discussions and Olivier Kah (Rennes, France) for sharing ideas about photoperiodism in fish.

REFERENCES

- Davis DE. Hibernation and circannual rhythms of food consumption in marmots and ground squirrels. *Q Rev Biol* (1976) **51**:477–514. doi:10.1086/409594
- Lincoln GA, Short RV. Seasonal breeding: nature's contraceptive. *Recent Prog Horm Res* (1980) **36**:1–52.
- Goldman BD, Darrow JM. The pineal gland and mammalian photoperiodism. *Neuroendocrinology* (1983) **37**:386–96. doi:10.1159/000123579
- Karsch FJ, Bittman EL, Foster DL, Goodman RL, Legan SJ, Robinson JE. Neuroendocrine basis of seasonal reproduction. *Recent Prog Horm Res* (1984) **40**:185–232.
- Zucker I. Circannual rhythms. In: Takahashi JS, Turek FW, Moore RY, editors. *Handbook of Behavioural Neurobiology, Circadian Clocks*. (Vol. 12), New York: Kluwer/Plenum (2001). p. 511–28.
- Lincoln GA, Hazlerigg DG. Mammalian circannual pacemakers. *Soc Reprod Fertil Suppl* (2010) **67**:171–86.
- Reiter RJ. Evidence for refractoriness of the pituitary-gonadal axis to the pineal gland in golden hamsters and its possible implications in annual reproductive rhythms. *Anat Rec* (1972) **173**:365–71. doi:10.1002/ar.1091730311
- Carter DS, Herbert J, Stacey PM. Modulation of gonadal activity by timed injections of melatonin in pinealectomized or intact ferrets kept under two photoperiods. *J Endocrinol* (1982) **93**:211–22. doi:10.1677/joe.0.0930211
- Bittman EL, Dempsey RJ, Karsch FJ. Pineal melatonin secretion drives the reproductive response to daylength in the ewe. *Endocrinology* (1983) **113**:2276–83. doi:10.1210/endo-113-6-2276
- Bartness TJ, Powers JB, Hastings MH, Bittman EL, Goldman BD. The timed infusion paradigm for melatonin delivery: what has it taught us about the melatonin signal, its reception, and the photoperiodic control of seasonal responses? *J Pineal Res* (1993) **15**:161–90. doi:10.1111/j.1600-079X.1993.tb00903.x
- Vanecek J, Pavlik A, Illnerova H. Hypothalamic melatonin receptor sites revealed by autoradiography. *Brain Res* (1987) **435**:359–62. doi:10.1016/0006-8993(87)91625-8
- Morgan PJ, Barrett P, Howell HE, Helliwell R. Melatonin receptors: localization, molecular pharmacology and physiological significance. *Neurochem Int* (1994) **24**:101–46. doi:10.1016/0197-0186(94)90100-7
- von Gall C, Stehle JH, Weaver DR. Mammalian melatonin receptors: molecular biology and signal transduction. *Cell Tissue Res* (2002) **309**:151–62. doi:10.1007/s00441-002-0581-4
- Pévet P, Challet E. Melatonin: both master clock output and internal time-giver in the circadian clocks network. *J Physiol Paris* (2011) **105**:170–82. doi:10.1016/j.jphysparis.2011.07.001
- Weaver DR, Reppert SM. Melatonin receptors are present in the ferret pars tuberalis and pars distalis, but not in brain. *Endocrinology* (1990) **127**:2607–9. doi:10.1210/endo-127-5-2607
- Reppert SM, Weaver DR, Ebisawa T. Cloning and characterization of a mammalian melatonin receptor that mediates reproductive and circadian responses. *Neuron* (1994) **13**:1177–85. doi:10.1016/0896-6273(94)90055-8
- Reppert SM, Godson C, Mahle CD, Weaver DR, Slaugenhaupt SA, Gusella JF. Molecular characterization of a second melatonin receptor expressed in human retina and brain: the Mel1b melatonin receptor. *Proc Natl Acad Sci U S A* (1995) **92**:8734–8. doi:10.1073/pnas.92.19.8734
- Weaver DR, Liu C, Reppert SM. Nature's knockout: the Mel1b receptor is not necessary for reproductive and circadian responses to melatonin in Siberian hamsters. *Mol Endocrinol* (1996) **10**:1478–87. doi:10.1210/me.10.11.1478
- Liu C, Weaver DR, Jin X, Shearman LP, Pieschl RL, Gribkoff VK, et al. Molecular dissection of two distinct actions of melatonin on the suprachiasmatic circadian clock. *Neuron* (1997) **19**:91–102. doi:10.1016/S0896-6273(00)80350-5
- Jin X, von Gall C, Pieschl RL, Gribkoff VK, Stehle JH, Reppert SM, et al. Targeted disruption of the mouse Mel(1b) melatonin receptor. *Mol Cell Biol* (2003) **23**:1054–60. doi:10.1128/MCB.23.3.1054-1060.2003
- Cogé F, Guenin SP, Fery I, Migaud M, Devavry S, Slugocki C, et al. The end of a myth: cloning and characterization of the ovine melatonin MT(2) receptor. *Br J Pharmacol* (2009) **158**:1248–62. doi:10.1111/j.1476-5381.2009.00453.x
- Yasuo S, Yoshimura T, Ebihara S, Korff HW. Melatonin transmits photoperiodic signals through the MT1 melatonin receptor. *J Neurosci* (2009) **29**:2885–9. doi:10.1523/JNEUROSCI.0145-09.2009
- Nosjean O, Ferro M, Coge F, Beauverger P, Henlin JM, Lefoulon F, et al. Identification of the melatonin-binding site MT3 as the quinone reductase 2. *J Biol Chem* (2000) **275**:31311–7. doi:10.1074/jbc.M005141200
- Wittkowski WH, Schulze-Bonhage AH, Bockers TM. The pars tuberalis of the hypophysis: a modulator of the pars distalis? *Acta Endocrinol (Copenh)* (1992) **126**:285–90.
- Wittkowski W, Bockmann J, Kreutz MR, Bockers TM. Cell and molecular biology of the pars tuberalis of the pituitary. *Int Rev Cytol* (1999) **185**:157–94. doi:10.1016/S0074-7696(08)60151-5
- Morgan PJ. The pars tuberalis: the missing link in the photoperiodic regulation of prolactin secretion? *J Neuroendocrinol* (2000) **12**:287–95. doi:10.1046/j.1365-2826.2000.00459.x
- Hazlerigg DG. What is the role of melatonin within the anterior pituitary? *J Endocrinol* (2001) **170**:493–501. doi:10.1677/joe.0.1700493
- Dardente H. Does a melatonin-dependent circadian oscillator in the pars tuberalis drive prolactin seasonal rhythmicity? *J Neuroendocrinol* (2007) **19**:657–66. doi:10.1111/j.1365-2826.2007.01564.x
- Gross DS. The mammalian hypophyseal pars tuberalis: a comparative immunocytochemical study. *Gen Comp Endocrinol* (1984) **56**:283–98. doi:10.1016/0016-6480(84)90043-1
- Fitzgerald KT. The structure and function of the pars tuberalis of the vertebrate adenohypophysis. *Gen Comp Endocrinol* (1979) **37**:383–99. doi:10.1016/0016-6480(79)90012-1
- Stoeckel ME, Hindelang C, Klein MJ, Poissonnier M, Felix JM. Expression of the alpha-subunit of glycoprotein hormones in the pars tuberalis-specific glandular cells in rat, mouse and guinea-pig. *Cell Tissue Res* (1994) **278**:617–24. doi:10.1007/s004410050253
- Stoeckel ME, Hindelang-Gertner C, Porte A. Embryonic development and secretory differentiation in the pars tuberalis of the mouse hypophysis. *Cell Tissue Res* (1979) **198**:465–76. doi:10.1007/BF00234191
- Drolet DW, Scully KM, Simmons DM, Wegner M, Chu KT, Swanson LW, et al. TEF, a transcription factor expressed specifically in the anterior pituitary during

- embryogenesis, defines a new class of leucine zipper proteins. *Genes Dev* (1991) **5**:1739–53. doi:10.1101/gad.5.10.1739
34. Klosen P, Bienvenu C, Demarteau O, Dardente H, Guerrero H, Pevet P, et al. The mt1 melatonin receptor and RORbeta receptor are co-localized in specific TSH-immunoreactive cells in the pars tuberalis of the rat pituitary. *J Histochem Cytochem* (2002) **50**:1647–57. doi:10.1177/002215540205001209
 35. Dardente H, Klosen P, Pevet P, Masson-Pevet M. MT1 melatonin receptor mRNA expressing cells in the pars tuberalis of the European hamster: effect of photoperiod. *J Neuroendocrinol* (2003) **15**:778–86. doi:10.1046/j.1365-2826.2003.01060.x
 36. Wittkowski W, Hewing M, Hoffmann K, Bergmann M, Fechner J. Influence of photoperiod on the ultrastructure of the hypophyseal pars tuberalis of the Djungarian hamster, *Phodopus sungorus*. *Cell Tissue Res* (1984) **238**:213–6. doi:10.1007/BF00215166
 37. Wittkowski W, Bergmann M, Hoffmann K, Pera F. Photoperiod-dependent changes in TSH-like immunoreactivity of cells in the hypophyseal pars tuberalis of the Djungarian hamster, *Phodopus sungorus*. *Cell Tissue Res* (1988) **251**:183–7. doi:10.1007/BF00215463
 38. Bockmann J, Bockers TM, Winter C, Wittkowski W, Winterhoff H, Deufel T, et al. Thyrotropin expression in hypophyseal pars tuberalis-specific cells is 3,5,3'-triiodothyronine, thyrotropin-releasing hormone, and pit-1 independent. *Endocrinology* (1997) **138**:1019–28. doi:10.1210/en.138.3.1019
 39. Dupré SM, Miedzinska K, Duval CV, Yu L, Goodman RL, Lincoln GA, et al. Identification of Eya3 and TAC1 as long-day signals in the sheep pituitary. *Curr Biol* (2010) **20**:829–35. doi:10.1016/j.cub.2010.02.066
 40. Lincoln GA, Clarke IJ, Hut RA, Hazlerigg DG. Characterizing a mammalian circannual pacemaker. *Science* (2006) **314**:1941–4. doi:10.1126/science.1132009
 41. Johnston JD. Photoperiodic regulation of prolactin secretion: changes in intra-pituitary signalling and lactotroph heterogeneity. *J Endocrinol* (2004) **180**:351–6. doi:10.1677/joe.0.1800351
 42. Dupré SM. Encoding and decoding photoperiod in the mammalian pars tuberalis. *Neuroendocrinology* (2011) **94**:101–12. doi:10.1159/000328971
 43. Nicholls TJ, Follett BK, Goldsmith AR, Pearson H. Possible homologies between photorefractoriness in sheep and birds: the effect of thyroidectomy on the length of the ewe's breeding season. *Reprod Nutr Dev* (1988) **28**:375–85. doi:10.1051/rnd:19880304
 44. Hazlerigg D, Loudon A. New insights into ancient seasonal life timers. *Curr Biol* (2008) **18**:R795–804. doi:10.1016/j.cub.2008.07.040
 45. Yoshimura T. Neuroendocrine mechanism of seasonal reproduction in birds and mammals. *Anim Sci J* (2010) **81**:403–10. doi:10.1111/j.1740-0929.2010.00777.x
 46. Wieselthier AS, Van Tienhoven A. The effect of thyroidectomy on testicular size and on the photorefractory period in the starling (*Sturnus vulgaris* L.). *J Exp Zool* (1972) **179**:331–8. doi:10.1002/jez.1401790306
 47. Follett BK, Nicholls TJ. Influences of thyroidectomy and thyroxine replacement on photoperiodically controlled reproduction in quail. *J Endocrinol* (1985) **107**:211–21. doi:10.1677/joe.0.1070211
 48. Webster JR, Moenter SM, Barrell GK, Lehman MN, Karsch FJ. Role of the thyroid gland in seasonal reproduction. III. Thyroidectomy blocks seasonal suppression of gonadotropin-releasing hormone secretion in sheep. *Endocrinology* (1991) **129**:1635–43. doi:10.1210/endo-129-3-1635
 49. Moenter SM, Woodfill CJ, Karsch FJ. Role of the thyroid gland in seasonal reproduction: thyroidectomy blocks seasonal suppression of reproductive neuroendocrine activity in ewes. *Endocrinology* (1991) **128**:1337–44. doi:10.1210/endo-128-3-1337
 50. Dardente H. Melatonin-dependent timing of seasonal reproduction by the pars tuberalis: pivotal roles for long daylengths and thyroid hormones. *J Neuroendocrinol* (2012) **24**:249–66. doi:10.1111/j.1365-2826.2011.02250.x
 51. Parkinson TJ, Follett BK. Effect of thyroidectomy upon seasonality in rams. *J Reprod Fertil* (1994) **101**:51–8. doi:10.1530/jrf.0.1010051
 52. Vigié C, Battaglia DF, Krasa HB, Thrun LA, Karsch FJ. Thyroid hormones act primarily within the brain to promote the seasonal inhibition of luteinizing hormone secretion in the ewe. *Endocrinology* (1999) **140**:1111–7. doi:10.1210/en.140.3.1111
 53. Anderson GM, Hardy SL, Valente M, Billings HJ, Connors JM, Goodman RL. Evidence that thyroid hormones act in the ventromedial preoptic area and the premammillary region of the brain to allow the termination of the breeding season in the ewe. *Endocrinology* (2003) **144**:2892–901. doi:10.1210/en.2003-0322
 54. Barrett P, Ebling FJ, Schuhler S, Wilson D, Ross AW, Warner A, et al. Hypothalamic thyroid hormone catabolism acts as a gatekeeper for the seasonal control of body weight and reproduction. *Endocrinology* (2007) **148**:3608–17. doi:10.1210/en.2007-0316
 55. Murphy M, Jethwa PH, Warner A, Barrett P, Nilaweera KN, Brameld JM, et al. Effects of manipulating hypothalamic triiodothyronine concentrations on seasonal body weight and torpor cycles in Siberian hamsters. *Endocrinology* (2012) **153**:101–12. doi:10.1210/en.2011-1249
 56. Freeman DA, Teubner BJ, Smith CD, Prendergast BJ. Exogenous T3 mimics long day lengths in Siberian hamsters. *Am J Physiol Regul Integr Comp Physiol* (2007) **292**:R2368–72. doi:10.1152/ajpregu.00713.2006
 57. Duncan MJ, Goldman BD, Di Pinto MN, Stetson MH. Testicular function and pelage color have different critical daylengths in the Djungarian hamster, *Phodopus sungorus sungorus*. *Endocrinology* (1985) **116**:424–30. doi:10.1210/endo-116-1-424
 58. Maywood ES, Hastings MH. Lesions of the iodomelatonin-binding sites of the mediobasal hypothalamus spare the lactotropic, but block the gonadotropic response of male Syrian hamsters to short photoperiod and to melatonin. *Endocrinology* (1995) **136**:144–53. doi:10.1210/en.136.1.144
 59. Galton VA. The roles of the iodothyronine deiodinases in mammalian development. *Thyroid* (2005) **15**:823–34. doi:10.1089/thy.2005.15.823
 60. Williams GR. Neurodevelopmental and neurophysiological actions of thyroid hormone. *J Neuroendocrinol* (2008) **20**:784–94. doi:10.1111/j.1365-2826.2008.01733.x
 61. Ebling FJ, Barrett P. The regulation of seasonal changes in food intake and body weight. *J Neuroendocrinol* (2008) **20**:827–33. doi:10.1111/j.1365-2826.2008.01721.x
 62. Lopez M, Varela L, Vazquez MJ, Rodriguez-Cuenca S, Gonzalez CR, Velagapudi VR, et al. Hypothalamic AMPK and fatty acid metabolism mediate thyroid regulation of energy balance. *Nat Med* (2010) **16**:1001–8. doi:10.1038/nm.2207
 63. Lopez M, Alvarez CV, Nogueiras R, Dieguez C. Energy balance regulation by thyroid hormones at central level. *Trends Mol Med* (2013) **19**:418–27. doi:10.1016/j.molmed.2013.04.004
 64. Kong WM, Martin NM, Smith KL, Gardiner JV, Connoley IP, Stephens DA, et al. Triiodothyronine stimulates food intake via the hypothalamic ventromedial nucleus independent of changes in energy expenditure. *Endocrinology* (2004) **145**:5252–8. doi:10.1210/en.2004-0545
 65. Lincoln GA, Rhind SM, Pompolo S, Clarke IJ. Hypothalamic control of photoperiod-induced cycles in food intake, body weight, and metabolic hormones in rams. *Am J Physiol Regul Integr Comp Physiol* (2001) **281**:R76–90.
 66. Hulet CV, Shupe WL, Ross T, Richards W. Effects of nutritional environment and ram effect on breeding season in range sheep. *Theriogenology* (1986) **25**:317–23. doi:10.1016/0093-691X(86)90067-1
 67. Menassol JB, Collet A, Chesneau D, Malpoux B, Scaramuzzi RJ. The interaction between photoperiod and nutrition and its effects on seasonal rhythms of reproduction in the ewe. *Biol Reprod* (2012) **86**:52. doi:10.1095/biolreprod.111.092817
 68. Walkden-Brown SW, Restall BJ, Norton BW, Scaramuzzi RJ, Martin GB. Effect of nutrition on seasonal patterns of LH, FSH and testosterone concentration, testicular mass, sebaceous gland volume and odour in Australian cashmere goats. *J Reprod Fertil* (1994) **102**:351–60.
 69. Zarazaga LA, Celi I, Guzman JL, Malpoux B. The role of nutrition in the regulation of luteinizing hormone secretion by the opioidergic, dopaminergic, and serotonergic systems in female Mediterranean goats. *Biol Reprod* (2011) **84**:447–54. doi:10.1095/biolreprod.110.086520
 70. Salazar-Ortiz J, Camous S, Briant C, Lardic L, Chesneau D, Guillaume D. Effects of nutritional cues on the duration of the winter anovulatory phase and on associated hormone levels in adult female Welsh pony horses (*Equus caballus*). *Reprod Biol Endocrinol* (2011) **9**:130. doi:10.1186/1477-7827-9-130
 71. Bianco AC, Salvatore D, Gereben B, Berry MJ, Larsen PR. Biochemistry, cellular and molecular biology, and physiological roles of the iodothyronine seleno-deiodinases. *Endocr Rev* (2002) **23**:38–89. doi:10.1210/er.23.1.38
 72. Lechan RM, Fekete C. Role of thyroid hormone deiodination in the hypothalamus. *Thyroid* (2005) **15**:883–97. doi:10.1089/thy.2005.15.883

73. St Germain DL, Galton VA, Hernandez A. Minireview: defining the roles of the iodothyronine deiodinases: current concepts and challenges. *Endocrinology* (2009) **150**:1097–107. doi:10.1210/en.2008-1588
74. Tu HM, Legradi G, Bartha T, Salvatore D, Lechan RM, Larsen PR. Regional expression of the type 3 iodothyronine deiodinase messenger ribonucleic acid in the rat central nervous system and its regulation by thyroid hormone. *Endocrinology* (1999) **140**:784–90. doi:10.1210/en.140.2.784
75. Bianco AC, Larsen PR. Cellular and structural biology of the deiodinases. *Thyroid* (2005) **15**:777–86. doi:10.1089/thy.2005.15.777
76. Kalsbeek A, Buijs RM, van Schaik R, Kaptein E, Visser TJ, Doulabi BZ, et al. Daily variations in type II iodothyronine deiodinase activity in the rat brain as controlled by the biological clock. *Endocrinology* (2005) **146**:1418–27. doi:10.1210/en.2004-0763
77. Tu HM, Kim SW, Salvatore D, Bartha T, Legradi G, Larsen PR, et al. Regional distribution of type 2 thyroxine deiodinase messenger ribonucleic acid in rat hypothalamus and pituitary and its regulation by thyroid hormone. *Endocrinology* (1997) **138**:3359–68. doi:10.1210/endo.138.8.5318
78. Guadano-Ferraz A, Obregon MJ, St Germain DL, Bernal J. The type 2 iodothyronine deiodinase is expressed primarily in glial cells in the neonatal rat brain. *Proc Natl Acad Sci U S A* (1997) **94**:10391–6. doi:10.1073/pnas.94.19.10391
79. Nakao N, Takagi T, Iigo M, Tsukamoto T, Yasuo S, Masuda T, et al. Possible involvement of organic anion transporting polypeptide 1c1 in the photoperiodic response of gonads in birds. *Endocrinology* (2006) **147**:1067–73. doi:10.1210/en.2005-1090
80. Ceballos A, Belinchon MM, Sanchez-Mendoza E, Grijota-Martinez C, Dumitrescu AM, Refetoff S, et al. Importance of monocarboxylate transporter 8 for the blood-brain barrier-dependent availability of 3,5,3'-triiodo-L-thyronine. *Endocrinology* (2009) **150**:2491–6. doi:10.1210/en.2008-1616
81. Visser WE, Friesema EC, Visser TJ. Minireview: thyroid hormone transporters: the knowns and the unknowns. *Mol Endocrinol* (2011) **25**:1–14. doi:10.1210/me.2010-0095
82. Herwig A, Wilson D, Logie TJ, Boelen A, Morgan PJ, Mercer JG, et al. Photoperiod and acute energy deficits interact on components of the thyroid hormone system in hypothalamic tanycytes of the Siberian hamster. *Am J Physiol Regul Integr Comp Physiol* (2009) **296**:R1307–15. doi:10.1152/ajpregu.90755.2008
83. Herwig A, de Vries EM, Bolborea M, Wilson D, Mercer JG, Ebling FJ, et al. Hypothalamic ventricular ependymal thyroid hormone deiodinases are an important element of circannual timing in the Siberian hamster (*Phodopus sungorus*). *PLoS One* (2013) **8**:e62003. doi:10.1371/journal.pone.0062003
84. Guerra M, Blazquez JL, Peruzzo B, Pelaez B, Rodriguez S, Toranzo D, et al. Cell organization of the rat pars tuberalis. Evidence for open communication between pars tuberalis cells, cerebrospinal fluid and tanycytes. *Cell Tissue Res* (2010) **339**:359–81. doi:10.1007/s00441-009-0885-8
85. Yoshimura T, Yasuo S, Watanabe M, Iigo M, Yamamura T, Hirunagi K, et al. Light-induced hormone conversion of T4 to T3 regulates photoperiodic response of gonads in birds. *Nature* (2003) **426**:178–81. doi:10.1038/nature02117
86. Watanabe M, Yasuo S, Watanabe T, Yamamura T, Nakao N, Ebihara S, et al. Photoperiodic regulation of type 2 deiodinase gene in Djungarian hamster: possible homologies between avian and mammalian photoperiodic regulation of reproduction. *Endocrinology* (2004) **145**:1546–9. doi:10.1210/en.2003-1593
87. Watanabe T, Yamamura T, Watanabe M, Yasuo S, Nakao N, Dawson A, et al. Hypothalamic expression of thyroid hormone-activating and -inactivating enzyme genes in relation to photorefractoriness in birds and mammals. *Am J Physiol Regul Integr Comp Physiol* (2007) **292**:R568–72. doi:10.1152/ajpregu.00521.2006
88. Revel FG, Saboureaux M, Pevet P, Mikkelsen JD, Simonneaux V. Melatonin regulates type 2 deiodinase gene expression in the Syrian hamster. *Endocrinology* (2006) **147**:4680–7. doi:10.1210/en.2006-0606
89. Helfer G, Ross AW, Morgan PJ. Neuromedin U partly mimics thyroid stimulating hormone and triggers Wnt/beta-catenin signalling in the photoperiodic response of F344 rats. *J Neuroendocrinol* (2013) **25**:1264–72. doi:10.1111/jne.12116
90. Yasuo S, Yoshimura T, Ebihara S, Korf HW. Temporal dynamics of type 2 deiodinase expression after melatonin injections in Syrian hamsters. *Endocrinology* (2007) **148**:4385–92. doi:10.1210/en.2007-0497
91. Nakao N, Ono H, Yamamura T, Anraku T, Takagi T, Higashi K, et al. Thyrotrophin in the pars tuberalis triggers photoperiodic response. *Nature* (2008) **452**:317–22. doi:10.1038/nature06738
92. Hanon EA, Lincoln GA, Fustin JM, Dardente H, Masson-Pevet M, Morgan PJ, et al. Ancestral TSH mechanism signals summer in a photoperiodic mammal. *Curr Biol* (2008) **18**:1147–52. doi:10.1016/j.cub.2008.06.076
93. Gereben B, Salvatore D. Pretranslational regulation of type 2 deiodinase. *Thyroid* (2005) **15**:855–64. doi:10.1089/thy.2005.15.855
94. Dupré SM, Dardente H, Birnie MJ, Loudon AS, Lincoln GA, Hazlerigg DG. Evidence for RGS4 modulation of melatonin and thyrotrophin signalling pathways in the pars tuberalis. *J Neuroendocrinol* (2011) **23**:725–32. doi:10.1111/j.1365-2826.2011.02168.x
95. Ebenhoeh O, Hazlerigg D. Modelling a molecular calendar: the seasonal photoperiodic response in mammals. *Chaos Solitons Fractals* (2013) **50**:39–47. doi:10.1016/j.chaos.2012.11.007
96. Hanon EA, Routledge K, Dardente H, Masson-Pevet M, Morgan PJ, Hazlerigg DG. Effect of photoperiod on the thyroid-stimulating hormone neuroendocrine system in the European hamster (*Cricetus cricetus*). *J Neuroendocrinol* (2010) **22**:51–5. doi:10.1111/j.1365-2826.2009.01937.x
97. Yasuo S, Yoshimura T, Ebihara S, Korf HW. Photoperiodic control of TSH-beta expression in the mammalian pars tuberalis has different impacts on the induction and suppression of the hypothalamo-hypophyseal gonadal axis. *J Neuroendocrinol* (2010) **22**:43–50. doi:10.1111/j.1365-2826.2009.01936.x
98. Krol E, Douglas A, Dardente H, Birnie MJ, Vinne V, Eijer WG, et al. Strong pituitary and hypothalamic responses to photoperiod but not to 6-methoxy-2-benzoxazolinone in female common voles (*Microtus arvalis*). *Gen Comp Endocrinol* (2012) **179**:289–95. doi:10.1016/j.ygcen.2012.09.004
99. Ross AW, Helfer G, Russell L, Darras VM, Morgan PJ. Thyroid hormone signalling genes are regulated by photoperiod in the hypothalamus of F344 rats. *PLoS One* (2011) **6**:e21351. doi:10.1371/journal.pone.0021351
100. Ono H, Hoshino Y, Yasuo S, Watanabe M, Nakane Y, Murai A, et al. Involvement of thyrotrophin in photoperiodic signal transduction in mice. *Proc Natl Acad Sci U S A* (2008) **105**:18238–42. doi:10.1073/pnas.0808952105
101. Unfried C, Ansari N, Yasuo S, Korf HW, von Gall C. Impact of melatonin and molecular clockwork components on the expression of thyrotrophin beta-chain (Tshb) and the TSH receptor in the mouse pars tuberalis. *Endocrinology* (2009) **150**:4653–62. doi:10.1210/en.2009-0609
102. Hazlerigg D. The evolutionary physiology of photoperiodism in vertebrates. *Prog Brain Res* (2012) **199**:413–22. doi:10.1016/B978-0-444-59427-3.00023-X
103. Nakane Y, Ikegami K, Iigo M, Ono H, Takeda K, Takahashi D, et al. The saccus vasculosus of fish is a sensor of seasonal changes in day length. *Nat Commun* (2013) **4**:2108. doi:10.1038/ncomms3108
104. Necrasov O, Adascalitei E. Contribution à l'étude du sac vasculaire des poissons osseux. *Trav Mus Nat His Nat Gr Antipa* (1968) **8**:465–83.
105. Frost WE, Kipling C. A study of reproduction, early life, weight-length relationship and growth of pike, *Esox lucius* L., in Windermere. *J Anim Ecol* (1967) **36**:651–93. doi:10.2307/2820
106. Kitano J, Lema SC, Luckenbach JA, Mori S, Kawagishi Y, Kusakabe M, et al. Adaptive divergence in the thyroid hormone signaling pathway in the stickleback radiation. *Curr Biol* (2010) **20**:2124–30. doi:10.1016/j.cub.2010.10.050
107. O'Brien CS, Bourdo R, Bradshaw WE, Holzapfel CM, Cresko WA. Conservation of the photoperiodic neuroendocrine axis among vertebrates: evidence from the teleost fish, *Gasterosteus aculeatus*. *Gen Comp Endocrinol* (2012) **178**:19–27. doi:10.1016/j.ygcen.2012.03.010
108. Perfito N, Jeong SY, Silverin B, Calisi RM, Bentley GE, Hau M. Anticipating spring: wild populations of great tits (*Parus major*) differ in expression of key genes for photoperiodic time measurement. *PLoS One* (2012) **7**:e34997. doi:10.1371/journal.pone.0034997
109. Bentley GE, Tucker S, Chou H, Hau M, Perfito N. Testicular growth and regression are not correlated with Dio2 expression in a wild male songbird, *Sturnus vulgaris*, exposed to natural changes in photoperiod. *Endocrinology* (2013) **154**:1813–9. doi:10.1210/en.2013-1093
110. Lucas RJ. Mammalian inner retinal photoreception. *Curr Biol* (2013) **23**:R125–33. doi:10.1016/j.cub.2012.12.029
111. Wyse C, Hazlerigg D. Seasonal biology: avian photoreception goes deep. *Curr Biol* (2009) **19**:R685–7. doi:10.1016/j.cub.2009.07.036

112. Ikegami K, Yoshimura T. Seasonal time measurement during reproduction. *J Reprod Dev* (2013) **59**:327–33. doi:10.1262/jrd.2013-035
113. Kosonsiriluk S, Mauro LJ, Chaiworakul V, Chaiseha Y, El Halawani ME. Photoreceptive oscillators within neurons of the premammillary nucleus (PMM) and seasonal reproduction in temperate zone birds. *Gen Comp Endocrinol* (2013) **190**:149–55. doi:10.1016/j.ygcen.2013.02.015
114. Halford S, Pires SS, Turton M, Zheng L, Gonzalez-Menendez I, Davies WL, et al. VA opsin-based photoreceptors in the hypothalamus of birds. *Curr Biol* (2009) **19**:1396–402. doi:10.1016/j.cub.2009.06.066
115. Nakane Y, Ikegami K, Ono H, Yamamoto N, Yoshida S, Hirunagi K, et al. A mammalian neural tissue opsin (opsin 5) is a deep brain photoreceptor in birds. *Proc Natl Acad Sci U S A* (2010) **107**:15264–8. doi:10.1073/pnas.1006393107
116. Yamashita T, Ohuchi H, Tomonari S, Ikeda K, Sakai K, Shichida Y. Opn5 is a UV-sensitive bistable pigment that couples with Gi subtype of G protein. *Proc Natl Acad Sci U S A* (2010) **107**:22084–9. doi:10.1073/pnas.1012498107
117. Kang SW, Leclerc B, Kosonsiriluk S, Mauro LJ, Iwasawa A, El Halawani ME. Melanopsin expression in dopamine-melatonin neurons of the premammillary nucleus of the hypothalamus and seasonal reproduction in birds. *Neuroscience* (2010) **170**:200–13. doi:10.1016/j.neuroscience.2010.06.082
118. Follett BK, Maung SL. Rate of testicular maturation, in relation to gonadotropin and testosterone levels, in quail exposed to various artificial photoperiods and to natural daylengths. *J Endocrinol* (1978) **78**:267–80. doi:10.1677/joe.0.0780267
119. Hoffmann K. The critical photoperiod in the Djungarian hamster *Phodopus sungorus*. In: Aschoff J, Daan S, Groos G, editors. *Vertebrate Circadian Systems*. Heidelberg: Springer-Verlag (1982). p. 297–304.
120. Elliott JA. Circadian rhythms and photoperiodic time measurement in mammals. *Fed Proc* (1976) **35**:2339–46.
121. Pittendrigh CS. Circadian surfaces and the diversity of possible roles of circadian organization in photoperiodic induction. *Proc Natl Acad Sci U S A* (1972) **69**:2734–7. doi:10.1073/pnas.69.9.2734
122. Dardente H, Cermakian N. Molecular circadian rhythms in central and peripheral clocks in mammals. *Chronobiol Int* (2007) **24**:195–213. doi:10.1080/07420520701283693
123. Dibner C, Schibler U, Albrecht U. The mammalian circadian timing system: organization and coordination of central and peripheral clocks. *Annu Rev Physiol* (2010) **72**:517–49. doi:10.1146/annurev-physiol-021909-135821
124. Mohawk JA, Green CB, Takahashi JS. Central and peripheral circadian clocks in mammals. *Annu Rev Neurosci* (2012) **35**:445–62. doi:10.1146/annurev-neuro-060909-153128
125. Messenger S, Ross AW, Barrett P, Morgan PJ. Decoding photoperiodic time through Per1 and ICER gene amplitude. *Proc Natl Acad Sci U S A* (1999) **96**:9938–43. doi:10.1073/pnas.96.17.9938
126. Dupré SM, Burt DW, Talbot R, Downing A, Mouzaki D, Waddington D, et al. Identification of melatonin-regulated genes in the ovine pituitary pars tuberalis, a target site for seasonal hormone control. *Endocrinology* (2008) **149**:5527–39. doi:10.1210/en.2008-0834
127. Guilding C, Hughes AT, Brown TM, Namvar S, Piggins HD. A riot of rhythms: neuronal and glial circadian oscillators in the mediobasal hypothalamus. *Mol Brain* (2009) **2**:28. doi:10.1186/1756-6606-2-28
128. Dardente H, Menet JS, Poiriel VJ, Streicher D, Gauer F, Vivien-Roels B, et al. Melatonin induces Cry1 expression in the pars tuberalis of the rat. *Brain Res Mol Brain Res* (2003) **114**:101–6. doi:10.1016/S0169-328X(03)00134-7
129. Hazlerigg DG, Andersson H, Johnston JD, Lincoln G. Molecular characterization of the long-day response in the Soay sheep, a seasonal mammal. *Curr Biol* (2004) **14**:334–9. doi:10.1016/j.cub.2004.01.057
130. Dardente H, Fortier EE, Martineau V, Cermakian N. Cryptochromes impair phosphorylation of transcriptional activators in the clock: a general mechanism for circadian repression. *Biochem J* (2007) **402**:525–36. doi:10.1042/BJ20060827
131. Ukai-Tadenuma M, Yamada RG, Xu H, Ripperger JA, Liu AC, Ueda HR. Delay in feedback repression by cryptochrome 1 is required for circadian clock function. *Cell* (2011) **144**:268–81. doi:10.1016/j.cell.2010.12.019
132. Maywood ES, Drynan L, Chesham JE, Edwards MD, Dardente H, Fustin JM, et al. Analysis of core circadian feedback loop in suprachiasmatic nucleus of mCry1-luc transgenic reporter mouse. *Proc Natl Acad Sci U S A* (2013) **110**:9547–52. doi:10.1073/pnas.1220894110
133. Fustin JM, Dardente H, Wagner GC, Carter DA, Johnston JD, Lincoln GA, et al. Egr1 involvement in evening gene regulation by melatonin. *FASEB J* (2009) **23**:764–73. doi:10.1096/fj.08-121467
134. Unfried C, Burbach G, Korf HW, von Gall C. Melatonin receptor 1-dependent gene expression in the mouse pars tuberalis as revealed by cDNA microarray analysis and in situ hybridization. *J Pineal Res* (2010) **48**:148–56. doi:10.1111/j.1600-079X.2009.00738.x
135. West A, Dupré SM, Yu L, Paton IR, Miedzinska K, McNeilly AS, et al. Npas4 is activated by melatonin, and drives the clock gene Cry1 in the ovine pars tuberalis. *Mol Endocrinol* (2013) **27**:979–89. doi:10.1210/me.2012-1366
136. Lincoln G, Messenger S, Andersson H, Hazlerigg D. Temporal expression of seven clock genes in the suprachiasmatic nucleus and the pars tuberalis of the sheep: evidence for an internal coincidence timer. *Proc Natl Acad Sci U S A* (2002) **99**:13890–5. doi:10.1073/pnas.212517599
137. Yasuo S, Watanabe M, Tsukada A, Takagi T, Iigo M, Shimada K, et al. Photoinducible phase-specific light induction of Cry1 gene in the pars tuberalis of Japanese quail. *Endocrinology* (2004) **145**:1612–6. doi:10.1210/en.2003-1285
138. Dardente H, Wyse CA, Birnie MJ, Dupré SM, Loudon AS, Lincoln GA, et al. A molecular switch for photoperiod responsiveness in mammals. *Curr Biol* (2010) **20**:2193–8. doi:10.1016/j.cub.2010.10.048
139. Masumoto KH, Ukai-Tadenuma M, Kasukawa T, Nagano M, Uno KD, Tsujino K, et al. Acute induction of Eya3 by late-night light stimulation triggers TSHbeta expression in photoperiodism. *Curr Biol* (2010) **20**:2199–206. doi:10.1016/j.cub.2010.11.038
140. Tsujino K, Narumi R, Masumoto KH, Susaki EA, Shinohara Y, Abe T, et al. Establishment of TSH beta real-time monitoring system in mammalian photoperiodism. *Genes Cells* (2013) **18**:575–88. doi:10.1111/gtc.12063
141. Helfer G, Ross AW, Russell L, Thomson LM, Shearer KD, Goodman TH, et al. Photoperiod regulates vitamin A and Wnt/beta-catenin signaling in F344 rats. *Endocrinology* (2012) **153**:815–24. doi:10.1210/en.2011-1792
142. Ross AW, Bell LM, Littlewood PA, Mercer JG, Barrett P, Morgan PJ. Temporal changes in gene expression in the arcuate nucleus precede seasonal responses in adiposity and reproduction. *Endocrinology* (2005) **146**:1940–7. doi:10.1210/en.2004-1538
143. Barrett P, Ross AW, Balik A, Littlewood PA, Mercer JG, Moar KM, et al. Photoperiodic regulation of histamine H3 receptor and VGF messenger ribonucleic acid in the arcuate nucleus of the Siberian hamster. *Endocrinology* (2005) **146**:1930–9. doi:10.1210/en.2004-1452
144. Klosen P, Seibert ME, Rasri K, Laran-Chich MP, Simonneaux V. TSH restores a summer phenotype in photoinhibited mammals via the RF-amides RFRP3 and kisspeptin. *FASEB J* (2013) **27**:2677–86. doi:10.1096/fj.13-229559
145. Morgan PJ, Hazlerigg DG. Photoperiodic signalling through the melatonin receptor turns full circle. *J Neuroendocrinol* (2008) **20**:820–6. doi:10.1111/j.1365-2826.2008.01724.x
146. Shearer KD, Goodman TH, Ross AW, Reilly L, Morgan PJ, McCaffery PJ. Photoperiodic regulation of retinoic acid signaling in the hypothalamus. *J Neurochem* (2010) **112**:246–57. doi:10.1111/j.1471-4159.2009.06455.x
147. Bechtold DA, Sidibe A, Saer BR, Li J, Hand LE, Ivanova EA, et al. A role for the melatonin-related receptor GPR50 in leptin signaling, adaptive thermogenesis, and torpor. *Curr Biol* (2012) **22**:70–7. doi:10.1016/j.cub.2011.11.043
148. Hand LE, Saer BR, Hui ST, Jinnah HA, Steinlechner S, Loudon AS, et al. Induction of the metabolic regulator Txnip in fasting-induced and natural torpor. *Endocrinology* (2013) **154**:2081–91. doi:10.1210/en.2012-2051
149. Scherbarth F, Steinlechner S. Endocrine mechanisms of seasonal adaptation in small mammals: from early results to present understanding. *J Comp Physiol B* (2010) **180**:935–52. doi:10.1007/s00360-010-0498-2
150. Bolborea M, Dale N. Hypothalamic tancytes: potential roles in the control of feeding and energy balance. *Trends Neurosci* (2013) **36**:91–100. doi:10.1016/j.tins.2012.12.008
151. Paul MJ, Zucker I, Schwartz WJ. Tracking the seasons: the internal calendars of vertebrates. *Philos Trans R Soc Lond B Biol Sci* (2008) **363**:341–61. doi:10.1098/rstb.2007.2143
152. Follett BK, Nicholls TJ. Photorefractoriness in Japanese quail: possible involvement of the thyroid gland. *J Exp Zool* (1984) **232**:573–80. doi:10.1002/jez.1402320325
153. Hazlerigg DG, Lincoln GA. Hypothesis: cyclical histogenesis is the basis of circannual timing. *J Biol Rhythms* (2011) **26**:471–85. doi:10.1177/0748730411420812

154. Migaud M, Batailler M, Pillon D, Franceschini I, Malpoux B. Seasonal changes in cell proliferation in the adult sheep brain and pars tuberalis. *J Biol Rhythms* (2011) **26**:486–96. doi:10.1177/0748730411420062
155. Lee DA, Blackshaw S. Functional implications of hypothalamic neurogenesis in the adult mammalian brain. *Int J Dev Neurosci* (2012) **30**:615–21. doi:10.1016/j.ijdevneu.2012.07.003
156. Hazlerigg DG, Wyse CA, Dardente H, Hanon EA, Lincoln GA. Photoperiodic variation in CD45-positive cells and cell proliferation in the mediobasal hypothalamus of the Soay sheep. *Chronobiol Int* (2013) **30**:548–58. doi:10.3109/07420528.2012.754450
157. Saenz de Miera C, Hanon EA, Dardente H, Birnie M, Simonneaux V, Lincoln GA, et al. Circannual variation in thyroid hormone deiodinases in a short-day breeder. *J Neuroendocrinol* (2013) **25**:412–21. doi:10.1111/jne.12013
158. Murphy M, Ebling FJ. The role of hypothalamic tri-iodothyronine availability in seasonal regulation of energy balance and body weight. *J Thyroid Res* (2011) **2011**:387562. doi:10.4061/2011/387562
159. Simonneaux V, Bur I, Ancel C, Ansel L, Klosen P. A kiss for daily and seasonal reproduction. *Prog Brain Res* (2012) **199**:423–37. doi:10.1016/B978-0-444-59427-3.00024-1
160. Clarke IJ, Caraty A. Kisspeptin and seasonality of reproduction. *Adv Exp Med Biol* (2013) **784**:411–30. doi:10.1007/978-1-4614-6199-9_19
161. Beltramo M, Dardente H, Cayla X, Caraty A. Cellular mechanisms and integrative timing of neuroendocrine control of GnRH secretion by kisspeptin. *Mol Cell Endocrinol* (2014) **382**:387–99. doi:10.1016/j.mce.2013.10.015
162. Oakley AE, Clifton DK, Steiner RA. Kisspeptin signaling in the brain. *Endocr Rev* (2009) **30**:713–43. doi:10.1210/er.2009-0005
163. Hill JW, Alreja M, Elias CF. From precocious puberty to infertility: metabolic control of the reproductive function. *Front Endocrinol (Lausanne)* (2013) **4**:43. doi:10.3389/fendo.2013.00043
164. Ebling FJ, Foster DL. Pineal melatonin rhythms and the timing of puberty in mammals. *Experientia* (1989) **45**:946–54. doi:10.1007/BF01953052
165. Ebling FJ. Photoperiodic regulation of puberty in seasonal species. *Mol Cell Endocrinol* (2010) **324**:95–101. doi:10.1016/j.mce.2010.03.018
166. Clarke IJ, Smith JT, Henry BA, Oldfield BJ, Stefanidis A, Millar RP, et al. Gonadotropin-inhibitory hormone is a hypothalamic peptide that provides a molecular switch between reproduction and feeding. *Neuroendocrinology* (2012) **95**:305–16. doi:10.1159/000332822
167. Kirby ED, Geraghty AC, Ubuka T, Bentley GE, Kaufer D. Stress increases putative gonadotropin inhibitory hormone and decreases luteinizing hormone in male rats. *Proc Natl Acad Sci U S A* (2009) **106**:11324–9. doi:10.1073/pnas.0901176106
168. Clarke IJ, Sari IP, Qi Y, Smith JT, Parkinson HC, Ubuka T, et al. Potent action of RFamide-related peptide-3 on pituitary gonadotropes indicative of a hypophysiotropic role in the negative regulation of gonadotropin secretion. *Endocrinology* (2008) **149**:5811–21. doi:10.1210/en.2008-0575
169. Caraty A, Blomenrohr M, Vogel GM, Lomet D, Briant C, Beltramo M. RF9 powerfully stimulates gonadotropin secretion in the ewe: evidence for a seasonal threshold of sensitivity. *J Neuroendocrinol* (2012) **24**:725–36. doi:10.1111/j.1365-2826.2012.02283.x
170. Ancel C, Bentsen AH, Seberr ME, Tena-Sempere M, Mikkelsen JD, Simonneaux V. Stimulatory effect of RFRP-3 on the gonadotropic axis in the male Syrian hamster: the exception proves the rule. *Endocrinology* (2012) **153**:1352–63. doi:10.1210/en.2011-1622
171. Ubuka T, Inoue K, Fukuda Y, Mizuno T, Ukena K, Kriegsfeld LJ, et al. Identification, expression, and physiological functions of Siberian hamster gonadotropin-inhibitory hormone. *Endocrinology* (2012) **153**:373–85. doi:10.1210/en.2011-1110
172. Tsutsui K, Ubuka T, Bentley GE, Kriegsfeld LJ. Review: regulatory mechanisms of gonadotropin-inhibitory hormone (GnIH) synthesis and release in photoperiodic animals. *Front Neurosci* (2013) **7**:60. doi:10.3389/fnins.2013.00060
173. Osugi T, Ohtaki N, Sunakawa Y, Son YL, Ohkubo M, Iigo M, et al. Molecular evolution of kiss2 genes and peptides in vertebrates. *Endocrinology* (2013) **154**:4270–80. doi:10.1210/en.2012-2267
174. Kriegsfeld LJ. Driving reproduction: RFamide peptides behind the wheel. *Horm Behav* (2006) **50**:655–66. doi:10.1016/j.yhbeh.2006.06.004
175. Simonneaux V, Ansel L, Revel FG, Klosen P, Pevet P, Mikkelsen JD. Kisspeptin and the seasonal control of reproduction in hamsters. *Peptides* (2009) **30**:146–53. doi:10.1016/j.peptides.2008.06.006
176. Henson JR, Carter SN, Freeman DA. Exogenous T(3) elicits long day-like alterations in testis size and the RFamides kisspeptin and gonadotropin-inhibitory hormone in short-day Siberian hamsters. *J Biol Rhythms* (2013) **28**:193–200. doi:10.1177/0748730413487974

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 18 December 2013; paper pending published: 03 February 2014; accepted: 10 February 2014; published online: 26 February 2014.

Citation: Dardente H, Hazlerigg DG and Ebling FJP (2014) Thyroid hormone and seasonal rhythmicity. *Front. Endocrinol.* 5:19. doi: 10.3389/fendo.2014.00019

This article was submitted to *Thyroid Endocrinology*, a section of the journal *Frontiers in Endocrinology*.

Copyright © 2014 Dardente, Hazlerigg and Ebling. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.