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New insights into the genetic structure of the outbreak-prone bamboo grasshoppers

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Introduction: The genetic structure of species is shaped by natural (e.g., terrain, climate) and non-natural (e.g., human activities) factors. Geographical isolation and natural barriers are important causes of genetic structure formation of species. Here, we explored this issue in bamboo grasshopper, *Ceracris kiangsu*, which is an important pest that feeds on bamboo in East and Southeastern Asia.

Methods: Based on 186 newly sequenced and 286 previously sequenced mitochondrial *COI* fragments, and 8 nuclear microsatellite loci, we examined the genetic diversity and population genetic structure of *C. kiangsu*.

Results: The degree of genetic differentiation among populations was also high, and Mantel test showed that it was significantly correlated with geographical distance. Principal coordinate analysis and STRUCTURE results revealed two genetically different groups, a South China (S-China) Group and a Southeast Asia (SE-Asia) Group. Climate variables partly explained the population genetic structure. The demographic history and ABC showed that the S-China Group experienced population expansion, whereas the SE-Asia Group was consistently stable.

Discussion: Our study demonstrates an obvious population structure maintained in this migratory insect and reveals the potential effect of past climatic change, geographical isolation, and ecological factors on the evolution of their genetic structure.

KEYWORDS

genetic structure, migration, genetic differentiation, climate, bamboo grasshopper

1. Introduction

The genetic structure of a species is shaped by several natural and non-natural factors. Geological processes and recent human disturbances play a widespread role in driving the extensive population genetic structure of most existing species (Avise, 2000; Heller et al., 2015). Climate change, especially Holocene climate warming, has greatly changed biodiversity. Species had variable responses to Holocene climate fluctuations, thus altering their genetic structure, population expansion, and regional adaptation (e.g., Song et al., 2016, 2020; Liu et al., 2020).

Geographical isolation and natural barriers are also known to have had important influences on the formation of the genetic structure of species. For example, three divergent lineages of the strider *Metrocoris esakii* could have evolved from geographical isolation (Ye et al., 2020). However, some species are highly migratory, and populations are expanding with frequent seasonal outbreaks. The wide dispersal and frequent outbreaks

may overcome the limitations of natural isolation and barriers. A couple of studies found that the gene flow in outbreak populations moderately increases, resulting in serious homogenization (Chapuis et al., 2009, 2010). However, how the migratory capacity and outbreak nature of a species evolutionarily shape genetic structure remains controversial (Tao et al., 2012).

Assessing how ecological differences among contemporary populations affect the geographic structure of intraspecific genetic variation is of great significance for pest management and predicting the response of populations to climate change (Thomas, 1999; Chen et al., 2017, 2018). Here, we attempted to resolve this issue with the bamboo grasshopper, *Ceracris kiangsu*.

Ceracris kiangsu is an insect species that belongs to Orthoptera. It mainly feeds on bamboo leaves, and thus is an important bamboo forest pest (Xuan et al., 2009). Although the spectrum of plants on which they feed is very narrow, these grasshoppers possess a powerful flight capacity and are able to fly 7.8 km per day. Therefore, their populations have frequently expanded to new regions and crossed country borders. This insect is now thriving in East Asia and Southeast Asia, including China, Laos, and Vietnam. Just between 2014 and 2020, the bamboo grasshopper has had successive outbreaks in Laos, Vietnam, and many places in China (Fan et al., 2014). Thus, the bamboo grasshopper provides a good model to investigate the effects of dispersal and population outbreaks on the evolution of genetic structure among populations.

The wide-range dispersal of the bamboo grasshopper had caused a large amount of bamboo death in bamboo forests. However, because of their high flight capacity and migratory nature, chemical sprays presented limited control of this pest. Moreover, it is difficult to predict the occurrence and dispersal of the grasshopper populations for effective prevention and control because of our limited knowledge of the population structure, ancestral source, and dispersal route of the bamboo grasshopper. Previous studies found that the bamboo grasshoppers are monophyletic with low population differentiation and genetic diversity, and have no significant correlation with geographical distance (Fan et al., 2014; Liu et al., 2016). Microsatellite data showed that historical genetic exchange was frequent, and most of the genetic variation among individuals was within populations (Liu et al., 2016). However, these studies only involved limited sampling of the species' distribution. They investigated populations from 13 provinces in southern China; however, this species has a wide distribution and frequent population outbreaks outside of China, such as in the China–Indochina Peninsula. Specific population genetic structure and phylogeography of *C. kiangsu* may be shaped by the complex environments in their wide distribution range in eastern and southeast Asia. Furthermore, the origin and migration paths of the bamboo grasshopper also remain unknown.

Here, we attempted to resolve these issues by expanding the sampling sites to include the full distribution range of this species and using different genetic markers. The main objective of this research was to examine: (1) the genetic structure of the bamboo grasshopper, (2) the historical dynamics of population size, and (3) the driving forces of the genetic evolution of the bamboo grasshopper. The findings will elucidate the role of migration in shaping the genetic structure of a highly bamboo grasshopper, facilitate our understanding of the origin of this pest and help to predict their outbreaks.

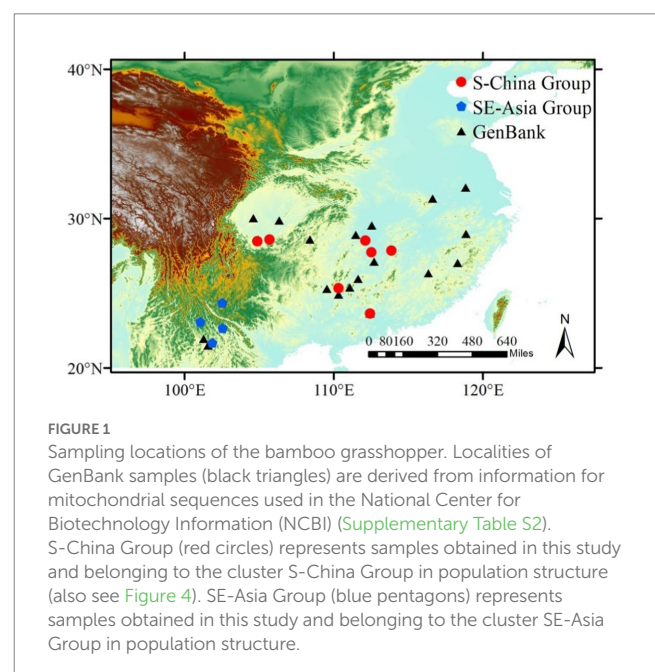
2. Materials and methods

2.1. Sampling and DNA extractions

In this study, 192 specimens from 10 geographic populations were collected. These 10 geographical populations were collected from 7 provinces in China, as well as Laos and Vietnam, with large-scale random sampling. Although there were two geographic populations in Yunnan Province, China, the results showed that they were not all sibling (Supplementary Table S1 in Supplementary material for specimen information and Figure 1). All specimens were stored in absolute ethanol and stored at -20°C . The whole genome DNA of the hind foot femoral joint muscle of the bamboo grasshopper was extracted by the “salting out method” (Sunnucks and Hales, 1996). The concentration and purity of the genomic DNA were detected with an ultra-micro spectrophotometer (Thermo Scientific Nano Drop 2000c, USA). Then, the DNA was diluted to 50–100 ng/ μL and stored in a freezer at -20°C .

2.2. PCR amplification and sequencing

The primers of the mitochondrial cytochrome oxidase subunit I (*COI*) gene were: 5'-TYTCAACAAAYCAYAARGATATTGG-3' and 5'-TAAACTTCWGGRTGWCCAAARAATCA-3' (Pan et al., 2006). PCR of mitochondrial mtDNA *COI* gene sequences was carried out on a ProFlex PCR instrument (Thermo Fisher Scientific, USA). The total reaction system was 15 μL , which included 1 μL genomic DNA, 0.6 μL upstream primer (10 μM), 0.6 μL downstream primer (10 μM), 7.5 μL Taq MasterMix (with 0.05 U/ μL Taq DNA Polymerase, 2 \times Taq PCR Buffer, 400 μM dNTP mix, and 3 mM MgCl_2) (CW Biotechnology Co., Beijing, China), and 5.3 μL sterilized double distilled water. The PCR procedure was as follows: pre-denaturation at 94°C for 2 min; denaturation at 94°C for 30 s, annealing at 54°C for 30 s, and extension at 72°C for 30 s for a total of 35 cycles; and final extension at 72°C for



2 min. The sequences were spliced using SeqMan ver. 7.1.0 software (DNASTar, Inc., USA). The amplified mtDNA *COI* gene sequences were routinely sequenced, and sequence identity was checked using Basic Local Alignment Search Tool (BLAST) on the National Center for Biotechnology Information (NCBI) website.¹

In microsatellite loci analysis, we screened the 12 pairs of previously reported microsatellite sequence primers (Xuan et al., 2009), and selected the 8 pairs with strong specificity (Supplementary Table S2). The total reaction system for microsatellite labeling was 15 μ L, which included 0.9 μ L genomic DNA, 0.3 μ L upstream primer with M13 (10 μ M), 1.2 μ L downstream primer (10 μ M), 1.2 μ L with FAM green fluorescent M13 primer (10 μ M), 8 μ L Es Taq Master Mix (with 0.05 U/ μ L Es Taq DNA Polymerase, 2 \times Taq PCR Buffer, 400 μ M dNTP mix, and 3 mM MgCl₂), and 3.4 μ L sterilized double distilled water. The PCR procedure was as follows: pre-denaturation at 94°C for 3 min; denaturation at 94°C for 30 s, annealing for 30 s (different primers have different annealing temperatures, see Supplementary Table S3 for details), and extension at 72°C for 30 s for a total of 35 cycles; denaturation at 94°C for 30 s, annealing at 53°C for 45 s, and extension at 72°C for 45 s for a total of 10 cycles; and final extension at 72°C for 10 min. The microsatellite primer products were sequenced by capillary electrophoresis (Sangon Biotech Co., China).

2.3. Mitochondrial *COI* analysis

Sequences of the 187 captured specimens (GenBank: OP341003-OP341189) and 286 GenBank sequences were obtained for genetic diversity statistics analysis (Supplementary Tables S1, S3). A nuclear mitochondrial pseudogene was discovered and eliminated it in the subsequent analysis. Haplotype diversity (Hd), nucleotide diversity (π), and the mean number of pairwise differences were calculated, not considering gaps, to estimate DNA polymorphism using DnaSP ver. 5.10.01 (Librado and Rozas, 2009). Analysis of molecular variance (AMOVA) was performed using Arlequin ver. 3.5 (Excoffier and Lischer, 2010) with 10,000 permutations based on populations and groups. A TCS network was constructed with the software Pop ART ver. 1.7 (Leigh and Bryant, 2015). Signatures of population demographic changes were tested for specimens in this study. First, Tajima's D and Fu's Fs statistics were calculated by Arlequin ver. 3.5, with the deviation from neutrality determined from 10,000 coalescent simulations. Second, mismatch distributions of pairwise sequences were calculated using DnaSP software.

We constructed a haplotype phylogenetic tree in BEAST ver. 1.10.4 (Suchard et al., 2018). The substitution model was selected using IQtree ver. 2.1.3 (Minh et al., 2020). Samples were drawn every 10,000 steps for 500 million steps under an uncorrelated lognormal relaxed clock model (Drummond and Rambaut, 2007) with a burn-in of 20%. *Ceracris nigricornis* and *C. versicolor* (GenBank accession numbers: JX840317.1, NC_025285.1) were selected as outgroups of *C. kiangsu* because they are phylogenetically sister species. *Ceracris nigricornis* differentiated from its sister species *C. kiangsu* 43.04 million years ago (Mya), and *C. versicolor* separated from *C. kiangsu* and *C. nigricornis*

44.81 Mya (Gao et al., 2018). The substitution rate was set to 1.77% per million years (Papadopoulou et al., 2010). All other parameters used were default values. To generate a tree with maximum credibility and 95% highest posterior density (HPD), trees were summarized by TreeAnnotator ver. 2.5.1 (Bouckaert et al., 2014). The resulting tree was visualized using FigTree ver. 1.4.2 (Rambaut, 2018).

To estimate the pattern of population size changes through time, Bayesian skyline plots (BSPs) implemented in BEAST were constructed. The outgroup taxa, which were for haplotype phylogenetic construction, were excluded from the time dating analysis. We ran BSP analyses separately for each group using the same data and settings as for the dating analyses with 'GTR+I+G' and 'HKY+I' as the substitution models selected by IQtree. The piecewise-linear skyline model and 10 groups were selected for Bayesian skyline coalescent tree priors (Drummond et al., 2005). All other parameters used were default values. Using effective sample size (ESS > 200) as an indicator, demographic plots were evaluated in Tracer ver. 1.5 (Rambaut et al., 2018) with a burn-in of 10%.

2.4. Microsatellite genotyping and statistical data analysis

2.4.1. Genetic diversity analysis

According to population and locus order, GenALEX ver. 6.503 (Peakall and Smouse, 2006) was used to convert data into format files required by analysis software in Excel 2010. GENEPOP ver. 4.7 (Rousset, 2008) detects Hardy-Weinberg equilibrium and the *p* value of each population. Genetic diversity of geographical populations and loci, such as the number of alleles (Na), number of effective alleles (Ne), observed heterozygosity (Ho), expected heterozygosity (He), inbreeding coefficient (F_{IS}), and polymorphism information content (PIC), were calculated with POPGENE ver. 1.3.1 (Yeh et al., 1999). POPGENE calculated the population F_{IS} , population differentiation coefficient (F_{ST}), and gene flow [$Nm = (1 - F_{ST}) / 4 F_{ST}$] of 8 microsatellite loci, and calculated the F_{ST} matrix among various populations of the bamboo grasshopper.

We estimated the geographical distance based on the longitude and latitude data of the sampling site and conducted a Mantel test based on GenALEX to infer the relationship between Slatkin's linearized F_{ST} and geographical distance; this allowed us to determine whether the observed genetic differentiation pattern conformed to the isolation by distance model. Principal coordinate analysis (PCoA) was carried out to show the genetic distance among *C. kiangsu* populations by GenALEX (Belkhir et al., 2004; Peakall and Smouse, 2006).

2.4.2. Structure analysis

The population structure of the 10 populations was constructed by Bayesian clustering analysis of STRUCTURE ver. 2.3.4 (Porrás-Hurtado et al., 2013). We used the mixed ancestral model without any population information to analyze and determine the number of groups; each *K* value was calculated 10 times, with 1,000,000 Markov chain Monte Carlo iterations and a burn-in of 100,000. Subsequently, the optimal value of group number *K* was determined with the online analysis website "structure harvest"². The software CLUMPP ver. 1.1

¹ <http://www.ncbi.nih.gov/BLAST/>

² http://taylor0.biology.ucla.edu/struct_harvest/

(Jakobsson and Rosenberg, 2007) was used to conduct model averaging of individual ancestry coefficients across 10 independent runs. Next, clusters were visualized using DISTRUCT ver. 1.1 (Rosenberg, 2004). According to the results of the STRUCTURE, we determined the genetic structure of the populations. Additionally, using Arlequin, based only on the variation of three levels among various groups, AMOVA was performed among various groups, within each group, and among individuals within each population for 10,000 permutations.

2.4.3. Population history by approximate Bayesian computation frame modeling

In order to explore the historical differentiation and possible historical dynamics of the populations, as implemented in DIYABC ver. 2.1.0, we performed ABC to explore the population history using the method based on the SSR sequence data (Cornuet et al., 2014). Based on the Bayesian clustering results, 10 populations are divided into two groups, namely, the southern group of China is N_1 , and the southeast group is N_2 . We analyzed its historical model (Figure 2B). We built a scenario to simulate the dynamics of population structure. All STRUCTURE clusters are considered as different groups, and 1,000,000 simulation data sets were run for the scenario. We analyzed the SSR dataset by using DIYABC v2.1.0 (Cornuet et al., 2014). In the model, it was the bifurcation time between the ancestral populations, and $N_1 + N_2$ was the effective population size of the common ancestor of the species.

2.5. Niche comparison

To quantify the niche comparisons between two groups identified by the genetic structure, 19 climatic variables (BIO1–19) at 10 sampling point records were extracted. Furthermore, we looked for differences in the S-China Group and SE-Asia Group. ArcGIS ver 10.2 software and climate layers downloaded from the global climate data³ were used to obtain the historical climate factor data.

Additionally, we used a multivariate approach of redundancy analysis (RDA) to estimate the extent to which the differences in microsatellite genotypes are explained by climate and geography and their collinear parts (spatial autocorrelation climate change). RDA is a constrained linear ranking method combining multiple linear regression and PCA (principal component analysis). In order to explain the genetic structure, we added two values of group variables: S-China Group and SE-Asia Group. Climate and group variables were classified as environmental conditions. Convert longitude and latitude coordinates into principal components of neighborhood matrix (PCNM) using the function PCNM in *vegan* R package (Ter Braak, 1986; R Core Team, 2014). In order to avoid high risk, we exclude variables with variance expansion coefficient (VIF) greater than 10. When all VIF values were lower than 10, three PCNM vectors and four climate variables remained including three temperature-related (BIO2, mean monthly temperature range; BIO10, mean temperature of warmest quarter; and two precipitation-related (BIO16, precipitation of wettest quarter; BIO19, precipitation of coldest

quarter) variables. Both full (climate and geography) and partial (climate) models of RDA were analyzed in the R package. The collinear ratio is calculated by subtracting the independent influence of climate and geography from the total variance explained in the complete RDA model (Cao et al., 2019).

3. Results

3.1. Genetic diversity

3.1.1. Mitochondrial DNA

We first examined genetic variability by sequencing a 567-bp fragment of the mtDNA *COI* gene in 473 bamboo grasshopper individuals that were sampled from 30 geographic populations in 14 provinces. A total of 122 haplotypes were identified. Genetic diversity indices, including H_d and π , are shown in Table 1. Average H_d and π were 0.744 ± 0.0191 and 0.1064 ± 0.00123 , respectively. The highest genetic diversity ($H_d = 0.983 \pm 0.028$, $\pi = 0.04215 \pm 0.01679$) was detected in the northeast population, Anhui (Table 1). In contrast, the genetic diversity was lowest in three of the southwest populations, i.e., Yunnan, Laos, and Vietnam.

3.1.2. Microsatellite DNA

We also analyzed genetic variation by genotyping 192 individuals collected from 10 geographic populations using eight microsatellite loci. These microsatellite loci showed significant deviation from Hardy–Weinberg equilibrium (Chi-Square test, $p < 0.05$). These loci were also highly polymorphic, with PIC values ranging from 0.471 to 0.864 (Supplementary Table S4). Thus, these microsatellite loci could provide rich genetic variation information and be used as effective genetic markers for population genetic diversity analysis.

We evaluated the genetic diversity of populations using six different parameters. The N_a among populations ranged from 4.750 to 17.375, the N_e from 3.700 to 9.089, and the PIC values from 0.563 to 0.853, which all revealed a diverse degree of polymorphism across the 10 populations (Table 2). The average H_o ranged from 0.331 to 0.750, with the lowest in the southwest population, Laos, and the highest in the southeast population, Guangdong. H_o was generally lower than H_e in all populations, indicating insufficient heterozygosity. F_{is} , the proportion of variance in the subpopulation contained in an individual, ranged from 0.046 (Guangdong) to 0.525 (Laos). The results illustrated that the four southwest populations, i.e., Yuxi, Puer, Laos, and Vietnam, had low genetic diversity and might have undergone frequent inbreeding, especially in Laos.

3.2. Phylogenetic analysis and TCS network

We reconstructed the phylogenetic tree using Bayesian Inference based on 128 haplotypes from the 473 mitochondrial *COI* gene sequences with gaps considered. The results showed that all haplotypes were split into two groups: a small-group branch including four haplotypes, Hap 54 (Yunnan), Hap 91, Hap 92 (Guangdong), and Hap 128 (Anhui), and a large-group branch including all the other haplotypes (Supplementary Figure S1). The differentiation time of different haplotypes was 31.95 Mya (HPD: 21.63–41.76 Mya), which

³ <https://www.worldclim.org/data/worldclim21.html>

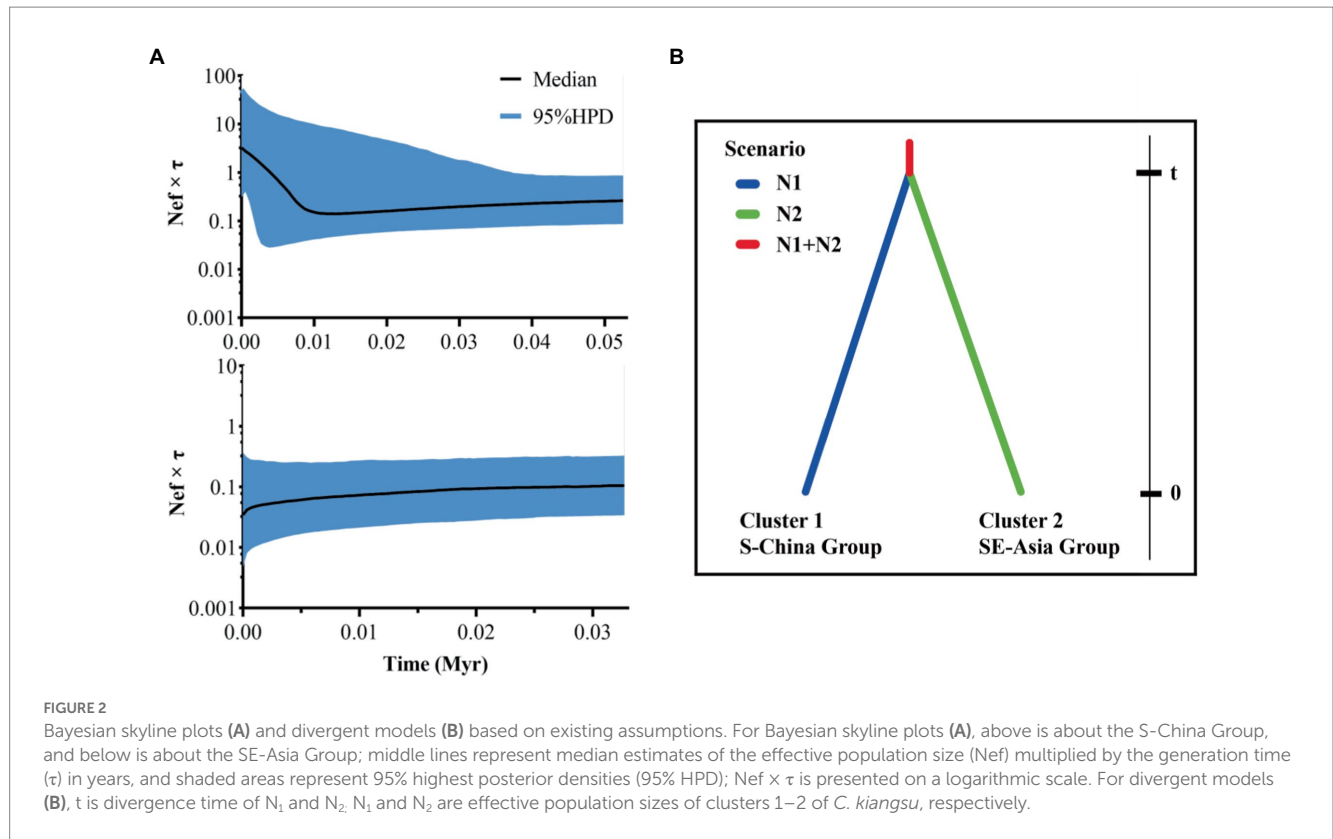


TABLE 1 Summary of genetic diversity based on mitochondrial DNA (mtDNA) in the bamboo grasshopper (*C. kiangsu*).

Code	N	Nh	Hd	π	Fu's Fs	Tajima's D
Guangdong	54	25	0.752 ± 0.065	0.01652 ± 0.00368	-3.997	-0.808
Guizhou	12	8	0.848 ± 0.104	0.02106 ± 0.00202	1.039	1.834
Guangxi	61	30	0.905 ± 0.026	0.01177 ± 0.00131	-11.901**	0.035
Hunnan	85	24	0.487 ± 0.069	0.00646 ± 0.00139	-9.579**	-1.105
Sichuan	37	7	0.345 ± 0.10	0.00081 ± 0.00027	-5.867***	-2.095**
Jiangxi	60	13	0.438 ± 0.081	0.0054 ± 0.00135	-2.131	-0.637
Jiangsu	20	5	0.442 ± 0.133	0.00126 ± 0.00047	-2.133*	-1.586*
Anhui	16	14	0.983 ± 0.028	0.04215 ± 0.01679	-1.154	-0.487
Chongqing	20	6	0.516 ± 0.132	0.00159 ± 0.00050	-2.743**	-1.570*
Fujian	12	6	0.682 ± 0.148	0.00225 ± 0.00068	-2.741**	-0.988
Zhejiang	14	9	0.912 ± 0.059	0.00573 ± 0.0009	-3.032*	-0.070
Yunnan	45	7	0.252 ± 0.086	0.01151 ± 0.00579	5.812	-2.411**
Laos	20	2	0.100 ± 0.088	0.00019 ± 0.00017	-0.879	-1.164
Vietnam	16	2	0.125 ± 0.106	0.00023 ± 0.00020	0	0
Mean	473	122	0.744 ± 0.0192	0.01053 ± 0.00122	-2.817	-2.346**

Samples sizes (N), number of haplotypes (Nh), haplotype diversity (Hd), nucleotide diversity (π), Fu's Fs and Tajima's D are showed. Asterisks represent level of statistical significance: * $p < 0.05$; ** $p < 0.05$.

was estimated by comparison with the two outgroups, i.e., Hap29 from its sister species *C. nigricornis* and Hap30 from *C. versicolor*.

We also performed haplotype network analysis based on 122 haplotypes with gaps excluded from analysis. The TCS network presented a star pattern, in which the most common haplotype (Hap 1) was located in the center, and other haplotypes were generally

connected to it in a star-like pattern (Figure 3). This pattern was consistent with the phylogenetic analysis (Supplementary Figure S1). Hap 1 was distributed in all populations except for the four southwest populations (Yuxi, Puer, Laos, and Vietnam). However, the second most common haplotype (Hap 3) was concentrated and only shared among the four southwest populations and the neighboring Guangxi

population; it did not exist in other populations. The two southern neighboring populations, i.e., the Guangxi and Guangdong populations, had the highest number of haplotypes, with 30 and 25 haplotypes, respectively. In addition, the Guangdong population possessed the most shared haplotypes (7) and the Guangxi population harbored the most unique haplotypes (24).

3.3. Genetic differentiation

To study genetic differentiation between populations based on the microsatellite data, we calculated F_{ST} values and gene flow (Nm) for population pairs. The degree of genetic differentiation between the six northeast populations, i.e., Jiangxi, Guangdong, Guizhou, Guangxi, Sichuan, and Hunan, was not significant, and most F_{ST} values were less than 0.05. The results demonstrated low genetic differentiation, and thus possible frequent gene exchange between these northeast

population pairs (Table 3). The results of genetic differentiation between the four southwest populations, i.e., Yuxi, Puer, Laos, and Vietnam, also revealed low genetic differentiation.

In contrast, there was a great degree of genetic differentiation between the population pairs of the six northeast populations and the four southwest populations ($F_{ST} > 0.05, p < 0.05$). F_{ST} values varied from 0.001 (Laos–Vietnam) to 0.236 (Guizhou–Laos, $p < 0.05$). Nm varied from 0.811 (Guizhou–Laos) to 219.048 (Laos–Vietnam). Moreover, the genetic differentiation between Guizhou and the four southeast populations was the largest ($F_{ST} > 0.15, p < 0.05$). The results indicated that there was much less gene exchange between Guizhou, the west central region, and the four southeast populations; however, there was frequent gene exchange between the two neighboring Southeast Asia regions, Laos and Vietnam.

3.4. Population genetic structure

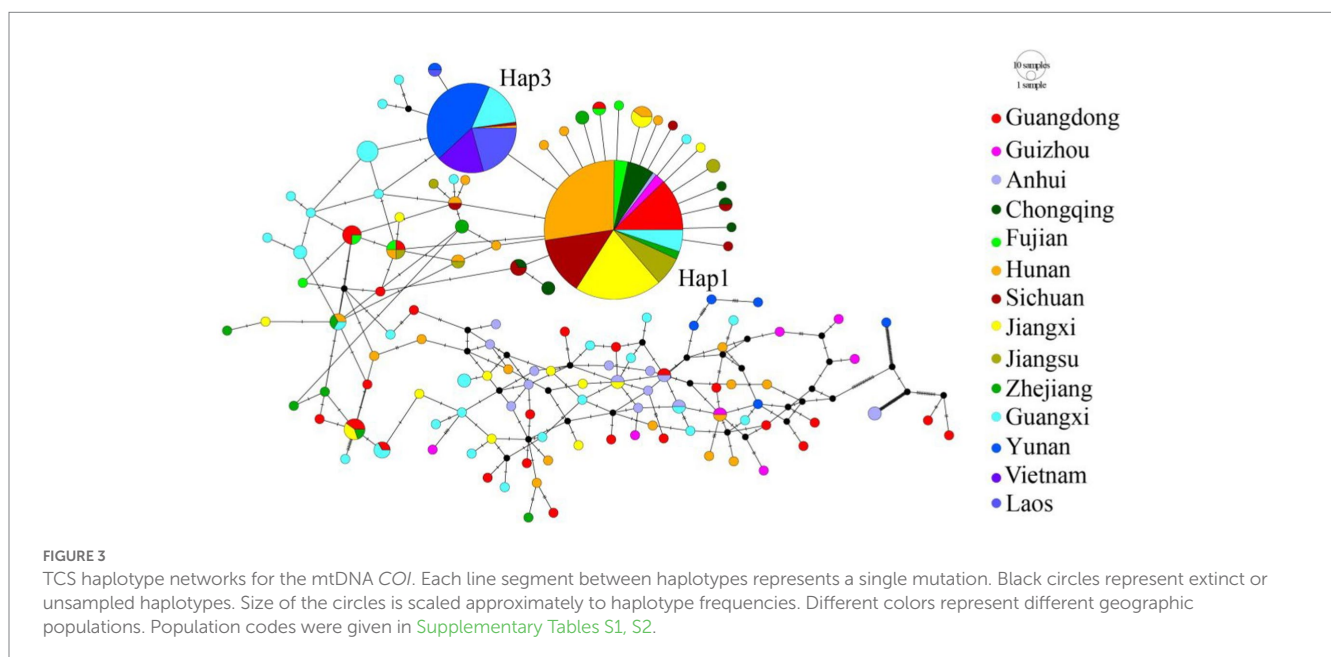
3.4.1. Microsatellite DNA

We then analyzed population genetic variation by PCoA using allele frequencies of the eight microsatellite markers. PCoA 1 explained 24.23% genetic variance and divided the 10 populations into two groups: the Southern China (S-China) Group, which included the six northeast populations, namely Jiangxi, Guizhou, Guangdong, Guangxi, Sichuan, and Hunan, and the Southeast Asia (SE-Asia) Group, which included the four southeast populations, namely Yuxi, Puer, Laos, and Vietnam (Figure 4A). This clustering pattern was consistent with the genetic differentiation results; that is, the genetic differentiation was low within the S-China Group and SE-Asia Group, but was large between the two groups.

We next analyzed the genetic structure of these 10 populations using Bayesian cluster analysis performed in STRUCTURE 2.3.4 (Figure 4B). The mean $LnP(D)$ values slowly increased starting from $K=2$, likely representing the most appropriate number of major clusters. The maximum ΔK was reached at $K=2$ (Supplementary Figure S2). Consistent with PCoA, two clusters were

TABLE 2 Genetic diversity on microsatellite data in the bamboo grasshopper.

Population code	Na	Ne	Ho	He	F_{IS}	PIC
Jiangxi	17.375	9.089	0.606	0.863	0.305	0.853
Guangdong	5.875	4.940	0.750	0.780	0.046	0.747
Guizhou	14.250	7.158	0.580	0.812	0.292	0.798
Guangxi	11.000	7.447	0.625	0.842	0.268	0.826
Sichuan	9.875	6.839	0.604	0.822	0.288	0.804
Hunan	10.750	6.768	0.606	0.813	0.232	0.796
Yuxi	8.250	4.533	0.396	0.607	0.439	0.587
Puer	9.000	4.626	0.417	0.628	0.401	0.611
Laos	7.750	3.700	0.331	0.58	0.525	0.563
Vietnam	7.500	3.923	0.404	0.606	0.384	0.585
Mean	10.163	5.902	0.532	0.735	0.318	0.717



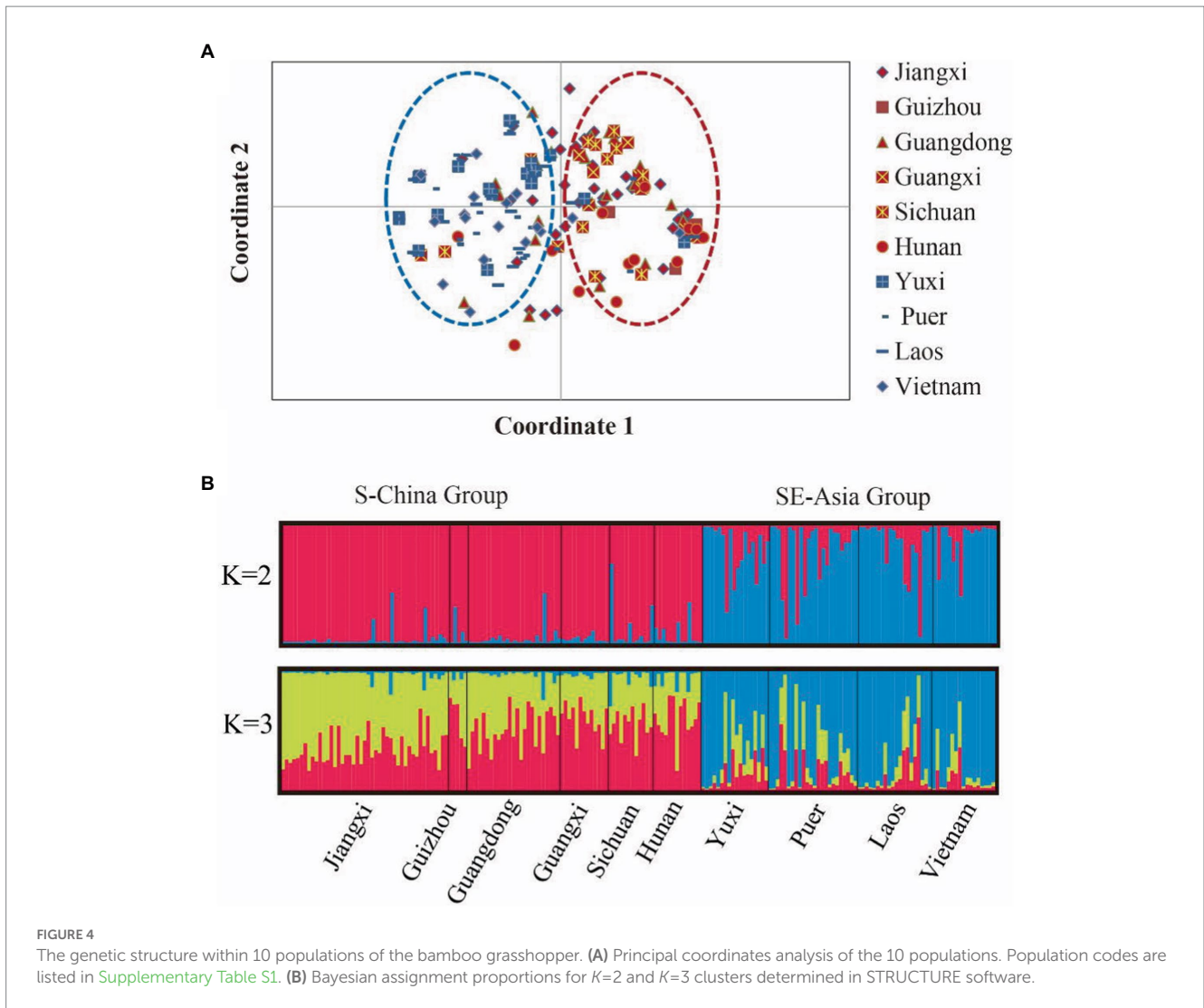


TABLE 3 Coefficient matrix representing genetic differentiation (F_{ST} , lower left) and gene flow (N_m , upper right) in the ten bamboo grasshopper populations.

Population code	Jiangxi	Guizhou	Guangdong	Guangxi	Sichuan	Hunan	Yuxi	Puer	Laos	Vietnam
Jiangxi	0	3.571	8.248	16.306	17.697	3.917	2.764	3.105	2.291	2.303
Guizhou	0.065*	0	3.438	5.620	5.292	4.136	0.982	0.971	0.811	0.900
Guangdong	0.029*	0.068*	0	14.560	11.526	3.271	3.400	2.360	2.590	2.780
Guangxi	0.015	0.043	0.017	0	25.208	8.679	1.926	2.110	1.691	1.808
Sichuan	0.014	0.045	0.021	0.010	0	5.564	2.980	3.210	2.308	2.453
Hunan	0.060*	0.057*	0.071*	0.028	0.043*	0	1.486	1.800	1.562	1.783
Yuxi	0.083*	0.203*	0.069*	0.115*	0.077*	0.144*	0	11.904	37.802	8.637
Puer	0.075*	0.205*	0.096*	0.106*	0.072*	0.122*	0.021	0	10.420	4.836
Laos	0.098*	0.236*	0.088*	0.129*	0.098*	0.138*	0.007	0.023	0	219.048
Vietnam	0.098*	0.217*	0.083*	0.121*	0.093*	0.123*	0.028	0.049*	0.001	0

Asterisks represent level of statistical significance: * $p < 0.05$. The gray indicate separate values between each population of the SE-Asia Group and each of S-China Group. F_{ST} greater than 0.2 are bolded, indicating large genetic differentiation between populations; F_{ST} equal to 0.001 is bolded, indicating minor genetic differentiation between populations.

identified among the samples, and the individuals in each cluster were similar to those in the corresponding PCoA. This further confirmed the accuracy of population genetic structure.

We also assessed population diversity and structure using AMOVA and F-statistics estimates based on microsatellite data. The global AMOVA indicated that the majority of genetic variance was

partitioned within individuals (92.92%), and the remaining genetic variation (7.08%) could be explained by the variation among populations (Supplementary Table S5). The hierarchical AMOVA revealed a similar population structure (overall $F_{ST}=0.031$; $p<0.001$), with 7.17% of the genetic variation distributed between two groups ($F_{CT}=0.072$; $p<0.01$) and less than 3% distributed among populations within the two groups ($F_{SC}=0.100$; $p<0.001$). The remaining 90% of the genetic variation was found within populations. By comparing AMOVA results, we found that most of the variation was within populations, and the rest existed between the two groups.

3.4.2. Mitochondrial DNA

We then analyzed the population genetic variation based on mitochondrial DNA data. The global AMOVA indicated that greater genetic variance was partitioned within individuals (80.81%), and the remaining genetic variation (19.19%) could be explained by the variation among populations (Supplementary Table S5). The hierarchical AMOVA of the bamboo grasshopper showed similar population structure (overall $F_{ST}=0.245$; $p<0.000$), with 9.61% of the genetic variation distributed between two groups ($F_{CT}=0.096$; $p<0.05$). Nearly 15% of the genetic variation was distributed among populations within the 2 groups ($F_{SC}=0.0164$; $p<0.001$). The remaining 75% of the genetic variation was found within populations. These results indicated significant genetic differences within the populations and shallow differentiation among populations.

There were some similarities and differences between mitochondrial DNA and microsatellite results. Similarly, most of the differences were concentrated within the populations. However, the microsatellite results showed that, except for the large differences within individuals, differences were concentrated among groups; alternatively, the mitochondrial results revealed that a greater degree of difference existed among populations.

3.5. Demographic history

We analyzed the population expansion history. The populations had significantly negative Tajima's D and Fu's F_s values (Table 1), indicating population size expansion during the bamboo grasshopper's evolutionary history. The unimodal mismatch distribution was strongly biased toward low divergence values, which demonstrated a recent demographic expansion from a small number of ancestors or a range expansion with high levels of migration between neighboring demes (Figure 2). The BSP of effective population size changed with time and remained stable for a long period, followed by a sharp rise 10 thousand years ago (Kya) in the S-China Group (Figure 2A). In contrast, the SE-Asia Group showed no population expansion in the Holocene (from 11.7 Kya to present) (Figure 2A). The BSP results indicated that the recently expanded ancestors of the bamboo grasshopper may have originated from the S-China Group.

Based on the detection of two clusters (N_1 and N_2) by STRUCTURE analysis (Figure 2B), ABC demographic analysis found that the population size of N_1 was about 5.5 times that of N_2 . The population size relationship was similar to the result of effective population size estimation of mitochondrial *COI* gene ($N_1 > N_2$). The divergence of the ancestral lineage forming clusters N_1 and N_2 occurred about 865 generations ago (97.5% HPD: 230–5,210, Table 4). The generation time of the bamboo grasshopper was calculated based

on their sexual maturity time of about 8 months (Liu et al., 2018). The results showed that the divergence between N_1 and N_2 clusters and the ancestral lineage occurred at about 1.30kya (97.5% HPD: 0.35–7.82 kya). The ABC framework demographic analysis shows that the divergence estimated by SSR was shorter than that estimated by mtDNA gene.

3.6. Correlation with climatic variables

We tested the correlation between genetic differentiation and geographical isolation in this bamboo grasshopper. Mantel test revealed that there was a significant positive correlation between genetic differentiation (F_{ST}) and geographical distance ($r=0.515$, $p<0.001$) (Figure 5C). However, four data points obviously deviated from the regression line. These data represented the differentiation between Guizhou and the four populations of SE-Asia Group (Figure 5C). After excluding these four data points, we then conducted correlation analysis again and found a stronger positive correlation ($r=0.607$, $p<0.001$). Geographical distance may not have been the main driving force of genetic differentiation between Guizhou and the SE-Asia Group populations.

We then tested the correlation effects of 19 climate variables and geography (mostly geographical position of latitude and longitude) on the genetic differentiation among the populations based on RDA. The influence of geography, climate and their interaction explained 19.7% of the total genetic variation. For the explained genetic variation, the independent effects of climatic conditions and geography accounted for 86.24 and 3.19%, respectively. When both climatic and geographical impacts are considered in the RDA analysis, three climatic variables (BIO2, mean monthly temperature range; BIO10, mean temperature of warmest quarter; BIO19, precipitation of coldest quarter), and one geographical variable (PCNM4) are highly correlated with genetic distance (Figure 5A). When geographical variables are limited in RDA analysis, mean monthly temperature range (BIO2), mean temperature of warmest quarter (BIO10), and precipitation of coldest quarter (BIO19) are highly correlated with genetic distance (Figure 5B).

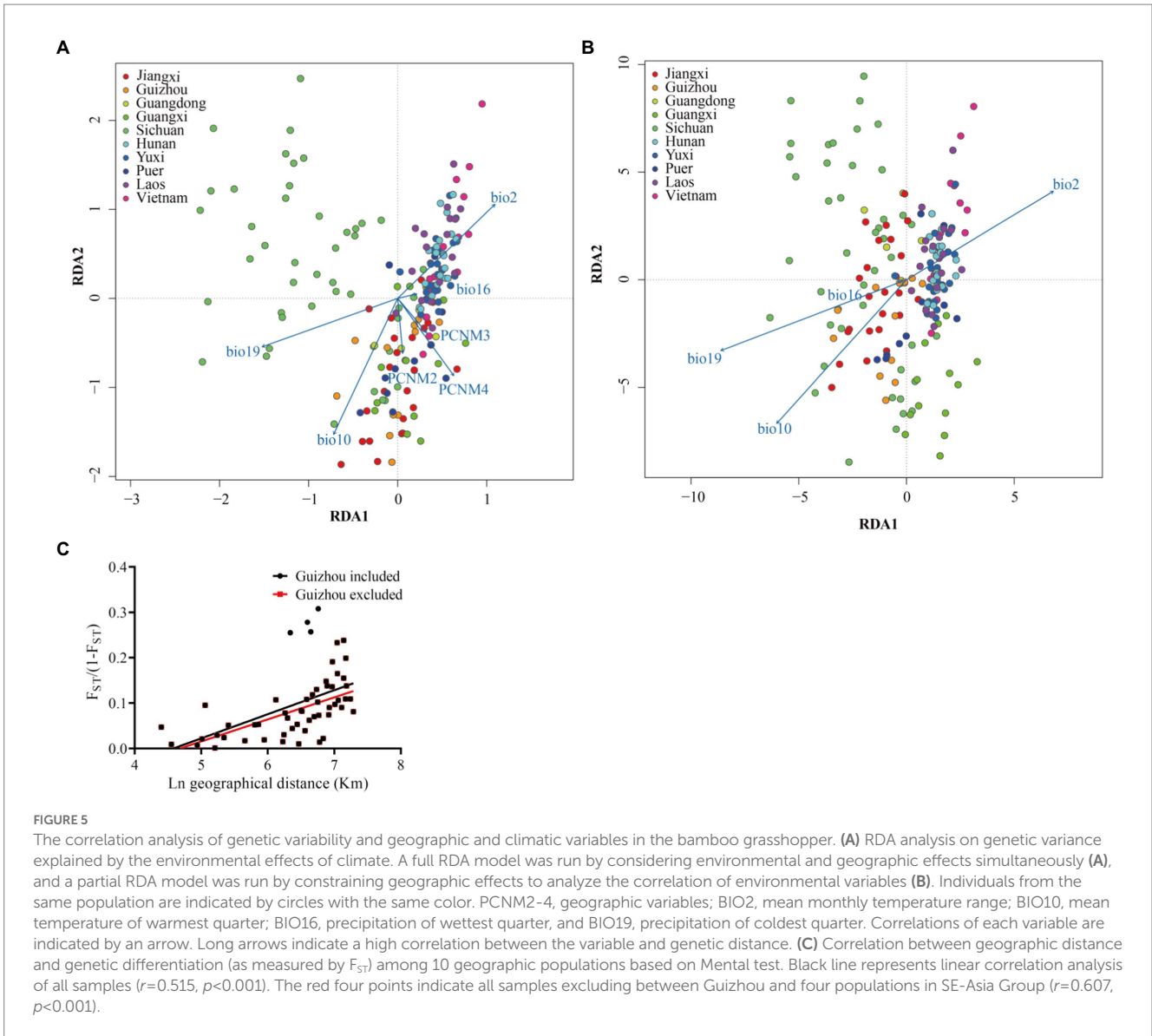
4. Discussion

The distribution range of a species is limited by many factors that, during evolutionary history, shape the genetic structure of the species. Here, we reported the potential genetic effects of some ecological, geographic, and climatic factors in a grasshopper species possessing a powerful long-distance migratory capacity. We used the mtDNA *COI* gene and microsatellite markers to explore the phylogenetic

TABLE 4 Posterior median, mean, mode, and range of 97.5% highest probability distribution (HPD) in lineage (cluster) merging model for *C. kiangsu* (time is in generations).

	2.5% HPD	Median	Mean	Mode	97.5% HPD
N_1	7.39×10^3	9.37×10^3	9.20×10^3	9.97×10^3	9.98×10^3
N_2	6.72×10^2	1.71×10^3	2.01×10^3	1.29×10^3	5.57×10^3
t	2.30×10^2	8.65×10^2	1.26×10^2	6.21×10^2	5.21×10^3

t is divergence time of N_1 and N_2 , N_1 and N_2 are effective population sizes of clusters 1–2 of *C. kiangsu*, respectively.



relationship and genetic variation (genetic diversity and genetic structure) among samples of the bamboo grasshopper from their wide distribution range. Mitochondrial data showed that the bamboo grasshopper was monophyletic, but had higher genetic diversity and more haplotypes than in previous studies (Ma et al., 2012; Pfeiler et al., 2017; Liu et al., 2019). Microsatellite data confirmed that these samples could be divided into two groups based on PCoA, Bayesian cluster analysis, and genetic differentiation analysis. The subsequent Mental test and RDA analysis indicated that geological barriers and climatic change effectively explained the genetic variation of this species.

4.1. Genetic diversity and phylogeny

The bamboo grasshopper had high haplotype diversity and nucleotide diversity based on the mtDNA *COI* gene, demonstrating a high level of genetic differentiation. Other migratory grasshoppers have a high level of genetic diversity, such as *Locusta migratoria* (Ma et al., 2012). However, the bamboo grasshopper had more haplotypes, especially unique haplotypes, compared with other migratory insects,

such as *L. migratoria*, *Danaus plexippus*, and *Aedes aegypti* (Ma et al., 2012; Pfeiler et al., 2017; Liu et al., 2019). Furthermore, the phylogenetic tree of haplotypes showed that most haplotypes of the bamboo grasshopper were clustered on one branch without obvious genetic structure, which was consistent with previous *C. kiangsu* results (Fan et al., 2014; Liu et al., 2016).

The distinct haplotype composition may be associated with the oligophagous traits of the bamboo grasshopper. There was a significant negative correlation between dietary breadth and genetic diversity at the population level. Populations with fewer mtDNA haplotypes had a wider range of food, whereas populations with higher haplotype diversity were more specialized (Singer and Parmesan, 2021). The distribution of this grasshopper largely overlaps with the distribution of bamboo (Li and Feng, 2019). The specialized host choice may promote haplotype differentiation. Furthermore, previous results have shown that oligophagous insects were phylogenetically conserved and belonged to monophyletic groups (Pekár et al., 2012; Gordon and Weirauch, 2016). Therefore, the oligophagous traits of the bamboo grasshopper may contribute to its high haplotype diversity.

4.2. Genetic differentiation and driving force

This study revealed significant genetic differentiation among populations that was positively correlated with geographical distance. Thus, the genetic variation among populations can be at least partly explained by geographical distance. Such close correlation between genetic differentiation and geographical distance has been observed in many other species, such as the whitefly *Bemisia tabaci* and the moths *Spodoptera exigua* and *Cydia pomonella* (Thaler et al., 2008; Díaz et al., 2015; Wang et al., 2020).

Geographic barriers could have also caused the genetic differentiation among some specific populations of the bamboo grasshoppers. There was a great degree of genetic differentiation between the Guizhou population (the west central region) and the SE-Asia Group. The terrain characteristics of Yunnan Province located between Guizhou and the SE-Asia Group include high-altitude terrain that is blocked by the famous Hengduan Mountains, and is high in the northwest and low in the southeast. This mountain barrier could also block the migration of the bamboo grasshopper and thus genetic exchange between populations in these local regions. It is not uncommon for geographic barriers to cause high genetic differentiation; for example, there is high genetic differentiation between giant pandas in the Daxiangling and Xiaoxiangling Mountains, and gene flow resistance of honeybees in valleys increases genetic differentiation (Zhu et al., 2010; Yu et al., 2019). Thus, we speculate that the genetic differentiation between Guizhou and the SE-Asia Group is affected by mountain isolation.

4.3. Distinct population structure and associated shaping factors

PCoA, STRUCTURE, and AMOVA results based on microsatellite data all supported a two-lineage structure for the 10 different geographic populations of the bamboo grasshopper. This finding is different from a previous report on this species (Liu et al., 2016). In fact, the two lineages are distinct in their genetic components and geographic origins. The S-China Group covers the six northeast populations, and the SE-Asia Group covers the four southeast populations (Figure 4B). This clustering pattern is consistent with the result of genetic differentiation, which was low within the S-China Group and SE-Asia Group, but was large between the two groups. These results indicated significant genetic differences within the populations and shallow differentiation among populations.

Interestingly, the RDA of 19 climate variables also revealed two different groups of these traits. The separation of the two groups was related to the change of difference in mean monthly temperature range (BIO2), mean temperature of warmest quarter (BIO10), and precipitation of coldest quarter (BIO19). Such climatic changes are related to the development and growth of the bamboo grasshoppers in their life cycle. Temperatures that are too high or too low damage the embryonic development of the bamboo grasshopper and reduce its hatching rate in winter (Zhao, 2009). The temperature variation and precipitation ranges are very important to the survival, development, and growth of the bamboo grasshopper. The population density was affected by ambient temperature (Liu et al., 2021). It was reported in other species that climatic variables play an important role in the formation of genetic structure (e.g., Ding et al., 2018). Climate

variables affected the genetic structure of *Aedes aegypti* (Mendonça et al., 2014; Gao et al., 2021), *Apis cerana japonica* (Nagamitsu et al., 2016), and *Trialeurodes vaporariorum* (Ovčárenko et al., 2014). Thus, temperature and precipitation are important factors affecting genetic structure. The current geographic pattern of genetic differentiation in the bamboo grasshopper populations may be the joint effect of geographic landscapes and ecological traits.

4.4. Demographic dynamics in the Holocene

Population expansion of insects with similar distributions in Eurasia mostly occurred in the Holocene (e.g., Ye et al., 2020). According to BSP analysis, the population size expansion for *C. kiangsu* occurred in the Holocene after the Last Glacial Maximum (LGM, 21 Kya). Similar Holocene expansion patterns in insects have been shown for *Chrysodeixis includens* (Silva et al., 2020), *Callosobruchus maculatus* (Kebe et al., 2017), *Peponapis pruinosa* (Lopez-Urbe et al., 2016), and *Homalictus fijiensis* (Dorey et al., 2021). All haplotypes were derived from the most common haplotype (Hap1), and the demographic history also showed that the current populations resulted from expansion from a few ancestors, supporting the same ancestor hypothesis. The Guangxi population had the most shared haplotypes and unique haplotypes in the TCS network. Therefore, Guangxi may have been the refuge from which the bamboo grasshopper expanded in the Holocene. In addition, changes during glacial–interglacial cycles produced large genetic changes in species. Correspondingly, the Earth's temperature rose sharply before and after the expansion of bamboo grasshoppers. We speculate that the population expansion was related to the rise of temperature. Similarly, many insects expanded their populations 10,000 years ago, such as *Aquarius paludum* (Ye et al., 2022). The population expansion during the glacial–interglacial cycles also confirms that the migration plays a significant role in shaping its current distribution pattern.

Ceracris kiangsu possesses a powerful of flight capacity, which is able to fly 7.8 km per day. Between 2014 and 2020, and the bamboo grasshopper has had successive outbreaks in Laos, Vietnam, and many places in China (Fan et al., 2014). The long-distance migratory behaviors of insects often work as a contrasting force that drives genetic differentiation, resulting in low genetic differentiation of many migratory insects, such as *Chlorops oryzae*, *Calliptamus italicus* and *Plutella xylostella* (Xu et al., 2019; Zhou et al., 2020; Chen et al., 2021). Frequent population outbreaks result in frequent gene exchange and a wide range of genetic homogenization among populations (Chapuis et al., 2009, 2010). The genetic variation results of microsatellite and mitochondrial *COI* gene showed that the population had low genetic differentiation. Therefore, we speculate that this is due to its strong migration ability.

What was the possible migration routes? Based on mtDNA data, the SE-Asia Group did not include the most extensive haplotypes (Hap1), but primarily had the divergent Hap3, which was also included in the Guangxi population. This result indicates that gene exchange between the Guangxi population and SE-Asia Group was frequent. Therefore, combining mitochondrial and microsatellite data, Guangxi was likely the refuge of *C. kiangsu* during the LGM, from which *C. kiangsu* migrated to other places and then formed the S-China Group. Therefore, we further speculate that the populations migrated from Guangxi to Yunnan, and finally to Laos and Vietnam in the SE-Asia Group (Figure 6).

Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article/[Supplementary material](#).

Author contributions

BC conceived the study. ZS, CL, and BC designed the experiments. SZ, BC, and CL collected samples. ZS conducted the experiments and analyses. ZS, CL, and BC wrote the manuscript. All authors contributed to the article and approved the submitted version.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fevo.2023.1062857/full#supplementary-material>

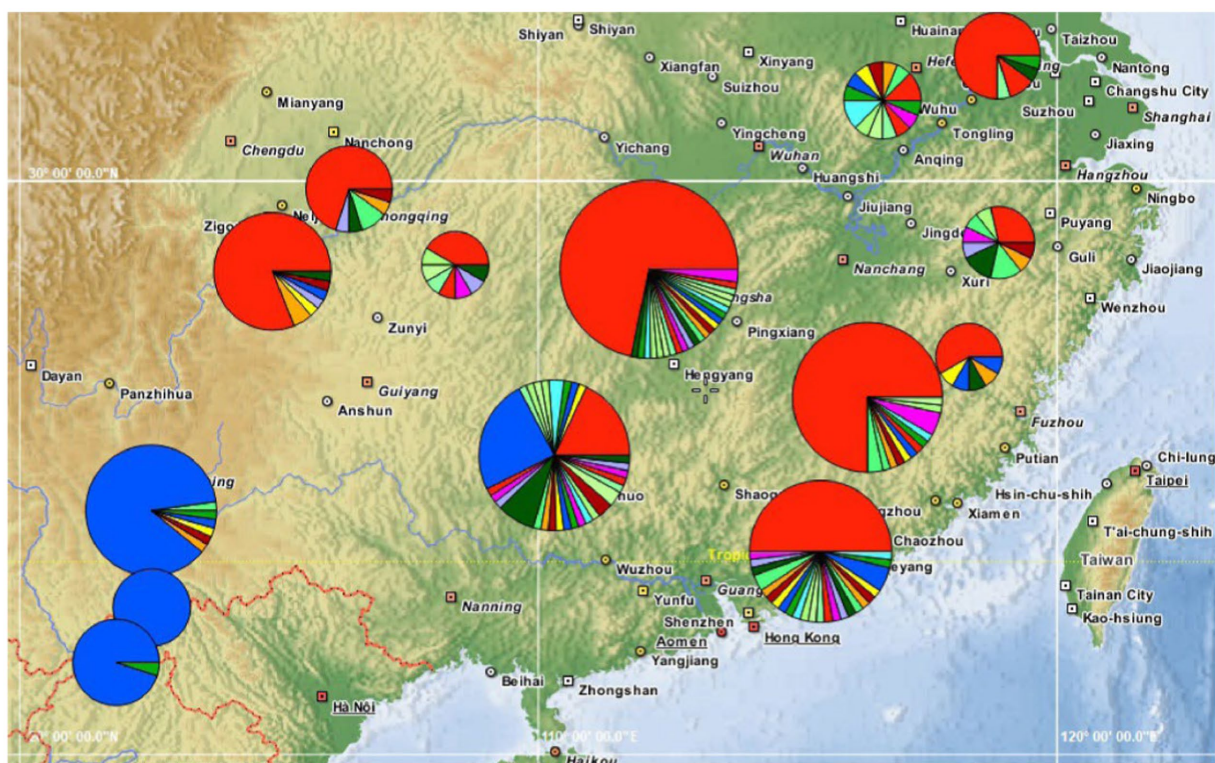


FIGURE 6
Phylogeography of the bamboo grasshopper. Pie charts show relative frequencies of *COI* haplotypes at each locality, and the colors are consistent with Figure 3.

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