



Spatial and Temporal Scales Matter When Assessing the Species and Genetic Diversity of Springtails (Collembola) in Antarctica

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Seven species of springtail (Collembola) are present in Victoria Land, Antarctica and all have now been sequenced at the DNA barcoding region of the mitochondrial cytochrome c oxidase subunit I gene (COI). Here, we review these sequence data ($n = 930$) from the GenBank and Barcode of Life Datasystems (BOLD) online databases and provide additional, previously unpublished sequences ($n = 392$) to assess the geographic distribution of COI variants across all species. Four species (*Kaylathalia klovstadi*, *Cryptopygus cisantarcticus*, *Friesea grisea*, and *Cryptopygus terranovus*) are restricted to northern Victoria Land and three (*Antarcticinella monoculata*, *Cryptopygus nivicolus*, and *Gomphiocephalus hodgsoni*) are found only in southern Victoria Land, the two biogeographic zones which are separated by the vicinity of the Drygalski Ice Tongue. We found highly divergent lineages within all seven species (range 1.7–14.7%) corresponding to different geographic locations. Levels of genetic divergence for the southern Victoria Land species *G. hodgsoni*, the most widespread species (~27,000 km²), ranged from 5.9 to 7.3% divergence at sites located within 30 km, but separated by glaciers. We also found that the spatial patterns of genetic divergence differed between species. For example, levels of divergence were much higher for *C. terranovus* (>10%) than for *F. grisea* (<0.2%) that had been collected from the same sites in northern Victoria Land. Glaciers have been suggested to be major barriers to dispersal and two species (*C. cisantarcticus* and *F. grisea*) showed highly divergent (>5%) populations and over 87% of the total genetic variation (based on AMOVA) on either side of a single, 16 km width glacier. Collectively, these data provide evidence for limited dispersal opportunities among populations of springtails due to geological and glaciological barriers (e.g., glaciers and ice tongues). Some locations harbored highly genetically divergent populations and these areas are highlighted from a conservation perspective such as avoidance of human-mediated transport between sites. We conclude that species-specific spatial and temporal scales need to be considered when addressing ecological and physiological questions as well as conservation strategies for Antarctic Collembola.

Keywords: Antarctica, biogeography, collembola, dispersal, mitochondrial DNA barcodes, population genetic structure, species diversity, springtails

INTRODUCTION

Due to the extreme environmental conditions that characterize Antarctica (Convey, 2013), as well as the geographical, and island-like, isolation of suitable terrestrial habitats (Bergstrom and Chown, 1999), long-range dispersal events for Antarctic Collembola (springtails) are rare and they usually rely on liquid water for transport via flotation (Hawes et al., 2008; McGaughan et al., 2011a; Carapelli et al., 2017). It is likely that springtails are currently unable to disperse among the three Antarctic Conservation Biogeographic Regions (ACBRs; Terauds and Lee, 2016) in the Ross Sea Sector of Antarctica: northern Victoria Land (NVL), southern Victoria Land (SVL), and the Transantarctic Mountains (TAM), owing to various physical obstacles in the marine and terrestrial realms (e.g., the Drygalski Ice Tongue; **Figure 1**). Within each of these three ACBRs, available habitat is patchy and local microhabitats are likely to be important for long-term persistence of populations (Sinclair and Sjørnsen, 2001). These small “island” populations can thus accumulate genetic mutations resulting in distinct genetic patterns across the landscape. Previous studies on the diversity patterns of lichen communities within the Antarctic have suggested that distributions are likely to have been driven by geological or glaciological features rather than environmental gradients that are associated with latitudinal or longitudinal distances (Adams et al., 2006; Peat et al., 2007; Green et al., 2011; Colesie et al., 2014). Similarly, the distributions of Antarctic springtail species may not follow a pattern of decreasing species diversity with increasing latitude (e.g., Caruso et al., 2009). The spatial scales at which sampling is undertaken are, therefore, important as genetic differences between populations do not necessarily increase proportionately with distance. However, until now a broader synthesis of genetic data for multiple species across larger spatial scales has not been undertaken. This is unfortunate as such an analysis could help to assess the relative role of geographic barriers in structuring populations of Antarctic springtails.

Terrestrial invertebrates were first discovered in the Ross Sea sector of Antarctica during the early 1900s (e.g., Carpenter, 1902, 1908; Willem, 1902; Gregory, 1909; Macnamara, 1919). Further entomological research was undertaken during the 1960s and 1970s and provided morphological descriptions and distributional ranges as well as general ecological observations and physiological studies, particularly for the most widespread species in this area—*Gomphiocephalus hodgsoni* (Gressitt and Leech, 1961; Salmon, 1962; Gressitt et al., 1963; Janetschek, 1963, 1967, 1970). More recently, molecular approaches including allozyme analyses (e.g., Frati et al., 1996; Stevens and Hogg, 2003) and DNA sequencing of mitochondrial cytochrome *c* oxidase subunit II (e.g., Frati and Dell’Ampio, 2000; Frati et al., 2001; McGaughan et al., 2010) and cytochrome *c* oxidase subunit I (COI) gene regions (e.g., Nolan et al., 2006; Stevens and Hogg, 2006; McGaughan et al., 2010) have been undertaken for Antarctic springtails. These data have enhanced understanding of their evolutionary histories and allowed testing of hypotheses, such as endemism of the fauna, proposed by earlier researchers (e.g., Wise, 1967). Further, they have provided insights into

the long-term persistence of endemic taxa, their survival in refugia and subsequent expansion during interglacial periods (e.g., Convey et al., 2008; McGaughan et al., 2011b; Beet et al., 2016; Carapelli et al., 2017). In turn, this has facilitated predictions as to how populations may respond to future changes in habitat availability and environmental conditions (e.g., Chown and Convey, 2007; Lee et al., 2017). While these studies either focus on a single species or have made comparisons between two species, none has assessed the full range of COI sequences available for all known species in the area.

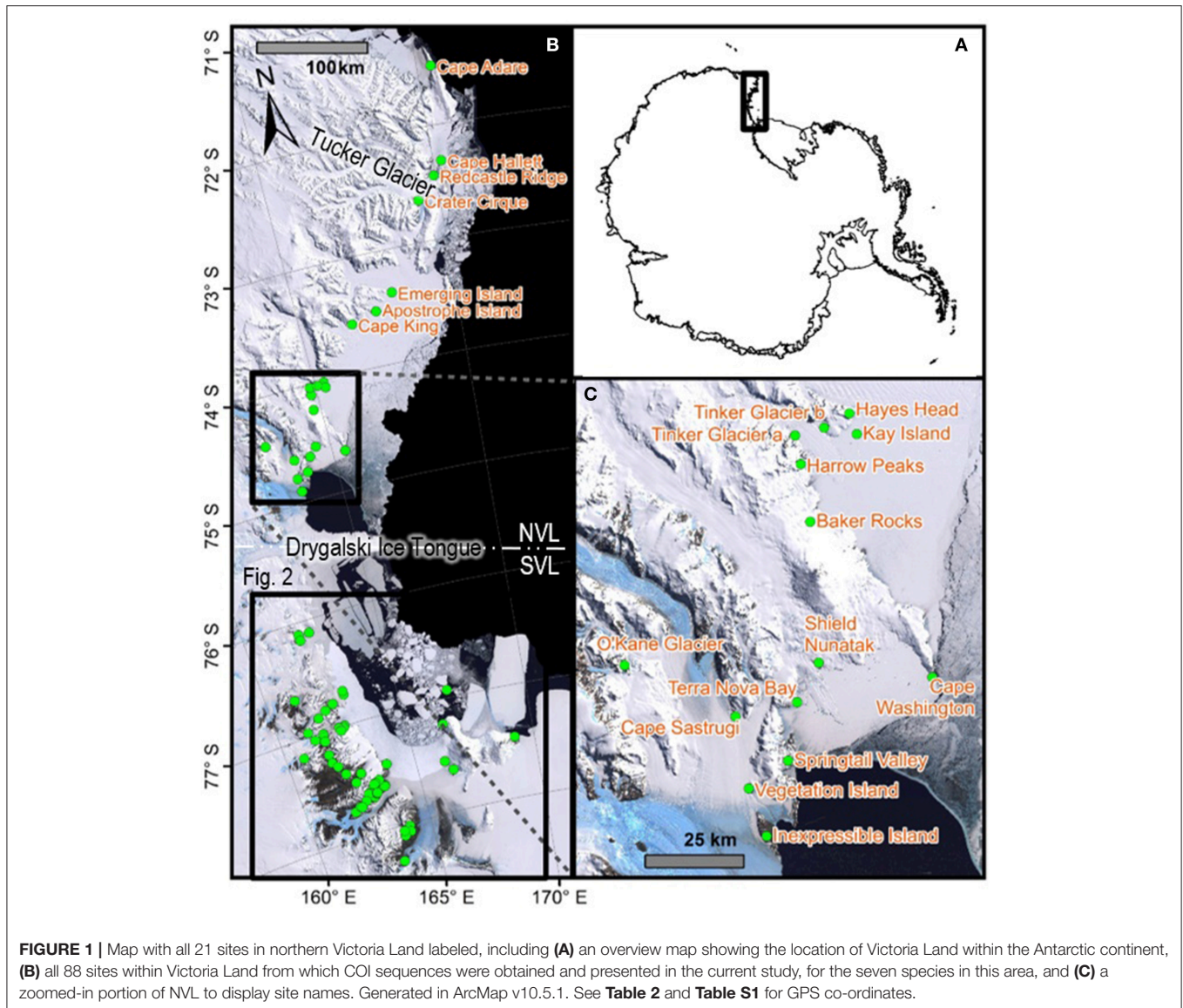
Based on early studies of springtail distribution using morphological taxonomy, the Ross Sea sector (longitude 160–175°E) was subdivided into five areas (Wise, 1967), although more recent data have confirmed that some of the described species distributions within these five areas were incorrect (e.g., Ross Island contains only *G. hodgsoni*). Studies focused on specific taxa (e.g., *Cryptopygus terranovus*) have suggested additional landscape divisions based on genetic evidence for geographical isolation (Stevens et al., 2006b; Hawes et al., 2010; Carapelli et al., 2017). However, other species present in the same area do not necessarily exhibit the same divergence patterns (Caruso et al., 2009). For example, in an area of NVL where *Friesea grisea* and *C. terranovus* have overlapping ranges, the two species have very different levels of genetic divergence; <0.2% for *F. grisea* (Torricelli et al., 2010b) and >10% for *C. terranovus* (Carapelli et al., 2017).

Our current study compiled and consolidated all available COI sequences and collection information for each of the seven species found in Victoria Land; *Cryptopygus cisantarcticus*, *C. terranovus*, *Friesea grisea*, and *Kaylathalia klovestadi* in NVL, and *Antarcticinella monoculata*, *C. nivicolus*, and *Gomphiocephalus hodgsoni* in SVL (**Table 1**). We provide a comprehensive list of collection sites for each haplotype to assess spatial variability. We also examined whether spatially isolated populations would be distinct at the COI gene region, and if so, whether landscape features (such as particularly large glaciers and ice tongues) may provide contemporary barriers to dispersal. Where relevant, we highlight instances of possible cryptic speciation as indicated by highly distinct genetic lineages, as well as identify priority sites from a conservation perspective.

METHODS

Existing Sequence Data

All publicly available sequences were downloaded from the BOLD and GenBank databases ($n = 930$ sequences), covering the seven springtail species present in Victoria Land. Supporting information was also downloaded from BOLD where possible, although in many instances these data were absent, particularly for sequences that had been obtained directly from GenBank. Where possible, we endeavored to manually retrieve collection details from original research articles. To maximize sequence length of previously-analyzed specimens, original trace files (where possible) were imported into Geneious v11.1.5 (<https://www.geneious.com>) for complete re-analysis. Forward and reverse sequence reads were assembled, any discrepancies were edited and then consensus sequences



were extracted. For cases where no trace files were available, final sequences that had been uploaded to GenBank or BOLD were used.

New Sequence Data

A total of 392 previously unpublished sequences have been included in the current study (**Tables 2–4** and **Table S2**). Of these, 56 were obtained from individuals collected from NVL in January 2004 (C. Beard and R. Seppelt), January 2015 (C. Cary) and November 2017 (I. Hogg). For previously unpublished sequences of *G. hodgsoni*, specimens were collected from sites in the vicinities of Mackay Glacier ($n = 60$), Taylor Valley ($n = 21$) and the southern Dry Valleys ($n = 249$) from 2009 to 2016 (I. Hogg, G. Collins, C. Beet and N. Demetras). An additional six specimens of *C. nivicolus* were collected from Mount Seuss in 2008 (I. Hogg), and Mount Seuss and Tiger Island in 2015 (G. Collins, C. Beet, I. Hogg and D. Cowan). For a full list of sites

where specimens were collected, see **Figures 1, 2**; **Tables 2, 3**, and **Tables S1–S9**.

These previously unpublished sequences were from specimens collected in pitfall traps (described in McGaughan et al., 2011a), using modified aspirators (Stevens and Hogg, 2002), or isolated by flotation from soil samples (Freckman and Virginia, 1993). In all cases, individual specimens were immediately preserved in 100% ethanol for later DNA extraction. DNA extractions, PCR amplifications and COI sequencing were carried out at the University of Waikato following procedures outlined in Collins and Hogg (2015) or at the Canadian Centre for DNA Barcoding following established protocols (see <http://ccdb.ca/resources/>).

Data Analyses

A separate alignment was made for each species using ClustalW within Geneious. After forward (LCO or LepF1) and reverse (HCO or LepR1) primer regions (26 bp each) were removed,

TABLE 1 | The seven springtail species present in the two Antarctic Conservation Biogeographic Regions (ACBRs) in Victoria Land (NVL and SVL), within the Ross Sea sector of Antarctica.

ACBR	Species code	Order	Family	Genus species
Northern Victoria Land (NVL)	KKLO	Entomobryomorpha	Isotomidae	<i>Kaylathalia klovstadi</i> (Carpenter, 1902)
	CCIS	Entomobryomorpha	Isotomidae	<i>Cryptopygus cisantarcticus</i> (Wise, 1967)
	FGRI	Entomobryomorpha	Neanuridae	<i>Friesea grisea</i> (Schaeffer, 1891)
	CTER	Entomobryomorpha	Isotomidae	<i>Cryptopygus terranovus</i> (Wise, 1967)
Southern Victoria Land (SVL)	AMON	Entomobryomorpha	Isotomidae	<i>Antarcticinella monoculata</i> (Salmon, 1965)
	CNIV	Entomobryomorpha	Isotomidae	<i>Cryptopygus nivicolus</i> (Salmon, 1965)
	GHOD	Poduromorpha	Hypogastruridae	<i>Gomphiocephalus hodgsoni</i> (Carpenter, 1908)

Refer to Sinclair and Stevens (2006) for additional taxonomic and distributional detail, and Register of Antarctic Species (RAS; raw.biodiversity.aq) for history of name changes for each species.

TABLE 2 | Collection sites in northern Victoria Land, grouped by region, including the number of COI sequences at each site for the four species in this area; *Cryptopygus cisantarcticus* (CCIS), *Cryptopygus terranovus* (CTER), *Friesea grisea* (FGRI), and *Kaylathalia klovstadi* (KKLO).

Region	Site name	Site code	Latitude	Longitude	CCIS	CTER	FGRI	KKLO
N Tucker Gl.	Cape Adare	CAD	-71.48170	170.38813				8 (8)
	Cape Hallett	CHA	-72.31986	170.23404	14 (10)		20 (10)	24 (19)
	Redcastle Ridge	RCR	-72.44110	169.94400	1 (1)			3 (3)
S Tucker Gl.	Crater Cirque	CCI	-72.63333	169.36667	10		6	
	Emerging Island	EMI	-73.38333	168.03333		10		
	Apostrophe Island	API	-73.51667	167.43333		10		
	Cape King	CAK	-73.58333	166.61667		10	7	
S Aviator Gl.	Baker Rocks	BAR	-74.23333	164.75000		10		
	Cape Washington	CAW	-74.65000	165.41667		10		
	Harrow Peaks	HPE	-74.10000	164.80000			5	
	Hayes Head	HHE	-74.01667	165.30000			5	
	Kay Island	KAI	-74.06667	165.31667		10	10	
	Tinker Glacier a	TGA	-74.03333	164.81667			7	
	Tinker Glacier b	TGB	-74.03333	165.06667		10	6	
S Campbell Gl.	Cape Sastrugi	CAS	-74.61667	163.68333		10		
	Inexpressible Island	INI	-74.90000	163.65000		10		
	O'Kane Glacier	OKG	-74.43070	162.89700	1 (1)			
	Shield Nunatak	SHN	-74.55000	164.50000		10		
	Terra Nova Bay/Jang Bogo Station	TNB	-74.62390	164.22890		53 (4)		
	Springtail Valley	SVA	-74.74725	164.01435		9		
	Vegetation Island	VEI	-74.78333	163.61667		10		

The number of sequences newly generated in the current study are shown in parentheses. See **Table S1** for additional details, such as correlation of site codes used in previous studies.

each species' alignment was individually assessed and trimmed accordingly to maximize alignment length but also retain maximum sequence coverage for each species. Final alignments varied in length, from 422 to 586 bp. Based on these final trimmed alignments, unique haplotypes for each species were then manually assigned codes in order of the date they were sequenced (e.g., GHOD-001 for *Gomphiocephalus hodgsoni*, haplotype 1). Future haplotypes can therefore be added sequentially to this dataset.

The numbers of duplicate sequences for each haplotype were manually calculated based on the number of replicates within our trimmed alignments, as well as from original tables and

figures provided in published papers for cases where only unique sequences were originally deposited online (Nolan et al., 2006; Demetras, 2010; McGaughan et al., 2010). This was particularly problematic for *G. hodgsoni*, as the numbers of each haplotype from each specific location could only be determined by a process of elimination through cross-referencing all available tables, figures and supplementary data. Unfortunately, collection information is absent for 46 *G. hodgsoni* specimens (see **Table S9**), outlined as follows. The precise collection location is uncertain for the two full mitochondrial genomes that have been previously assembled for *G. hodgsoni* (GenBank accessions AY191995 and NC005438; Nardi et al., 2003) as well as the three

TABLE 3 | List of collection sites in southern Victoria Land, grouped by region, including the number of COI sequences at each site for the three species in this area; *Antarcticinella monoculata* (AMON), *Cryptopygus nivicolus* (CNIV) and *Gomphiocephalus hodgsoni* (GHOD).

Region	Site name	Latitude	Longitude	AMON	CNIV	GHOD
N Mawson Gl.	Cliff Nunatak	-76.11000	162.01400	12		
	Mt Murray	-76.16100	162.01600	11		
	Tripp Island	-76.11667	162.41667			8
Mackay Gl. region	Benson Glacier a	-76.82200	162.10700	1		21 (4)
	Benson Glacier b	-76.87000	161.75400			3
	Botany Bay	-77.00000	162.65000			15
	Cape Geology	-77.01438	162.60143			12
	Cape Ross	-76.73333	162.96667			5
	Depot Island	-76.70000	162.96667			5
	Flatiron	-77.00500	162.40800			28 (6)
	Mount England	-77.03333	162.46667		2	11
	Mount Gran	-76.96600	161.17900		7	3
	Mount Seuss a	-77.03333	161.73333		12 (4)	45
	Mount Seuss b	-77.01500	161.75000		3	64 (49)
	Pegtop Mountain	-77.04600	161.36200	2		
	Sperm Bluff	-77.08333	161.71667			8
	Springtail Point	-77.16700	160.71000	7	11	
	Tiger Island	-76.78400	162.45200		3(1)	14(2)
	Towle Glacier	-76.65500	161.09300		2	6(1)
Northern Dry Valleys	Clark Glacier	-77.41667	162.11667			4
	Lake Brownworth	-77.45000	162.71667			6
	Marble Point	-77.43537	163.82615			3
	Mautrino Peak	-77.51667	162.41667			6
	Saint John's Range a	-77.28000	161.73100			23
	Saint John's Range b	-77.20800	161.70000			5
	Saint John's Range c	-77.28500	161.72600			4
	Saint John's Range d	-77.33333	161.90000			7
Taylor Valley	Borns Glacier	-77.76665	162.01999			16
	Canada Glacier	-77.60965	163.00913			2
	Commonwealth Glacier a	-77.61238	163.40228			1
	Commonwealth Glacier b	-77.61667	163.40000			3
	Delta Stream	-77.64295	163.13297			8 (4)
	Goldman Glacier a	-77.68665	162.87223			1
	Goldman Glacier b	-77.68590	162.92163			1
	Goldman Glacier c	-77.68707	162.94993			6
	Goldman Glacier d	-77.68870	162.97068			1
	Howard Glacier	-77.66225	163.09725			25 (8)
	Lake Chad a	-77.64285	162.77487			1
	Lake Chad b	-77.64567	162.76300			1
	Lake Fryxell a	-77.62665	163.11502			1
	Lake Fryxell b	-77.63220	163.21262			1
	Lake Fryxell c	-77.61998	163.18502			2
	Lake Fryxell d	-77.63333	163.21667			27
	Mount Barnes a	-77.60715	163.50088			3
	Mount Barnes b	-77.61283	163.52695			1
	Mount Barnes c	-77.61930	163.49298			1
	Mount Barnes d	-77.62112	163.48913			1
Mount Cerberus	-77.70000	162.58333			2	
Mount Coleman	-77.55142	163.34123			6	
Mount Falconer a	-77.58107	163.10958			1	

(Continued)

TABLE 3 | Continued

Region	Site name	Latitude	Longitude	AMON	CNIV	GHOD
Southern Dry Valleys	Mount Falconer b	-77.56892	163.15578			1
	Mount Falconer c	-77.57725	163.09210			1
	Spaulding Pond a	-77.65861	163.10808			65 (3)
	Spaulding Pond b	-77.65789	163.12044			35 (1)
	Spaulding Pond c	-77.65889	163.12539			56 (1)
	Taylor Valley 389	-77.73400	162.28100			2 (2)
	Upper Taylor Valley	-77.74100	162.34200			2 (2)
	Garwood Valley	-78.01965	164.05673			74 (61)
	Lake Penny	-78.30992	163.40788			4
	Marshall Valley	-78.06988	164.03370			8 (8)
	Miers Valley	-78.09603	163.75900			173 (164)
Ross Island area	Shangri La	-78.05425	163.77730			16 (16)
	Beaufort Island	-76.93177	166.91347			12
	Cape Bird	-77.22110	166.44683			12
	Cape Crozier	-77.46325	169.19687			11
	Cape Evans	-77.63388	166.44252			4
	Cape Royds	-77.54600	166.16298			10

The number of sequences newly generated in the current study are shown in parentheses. See **Table S1** for additional details, such as correlation of site codes used in previous studies.

TABLE 4 | Sequence details for the seven species of Collembola present in NVL and SVL, showing alignment lengths used in the current study (which vary for each species).

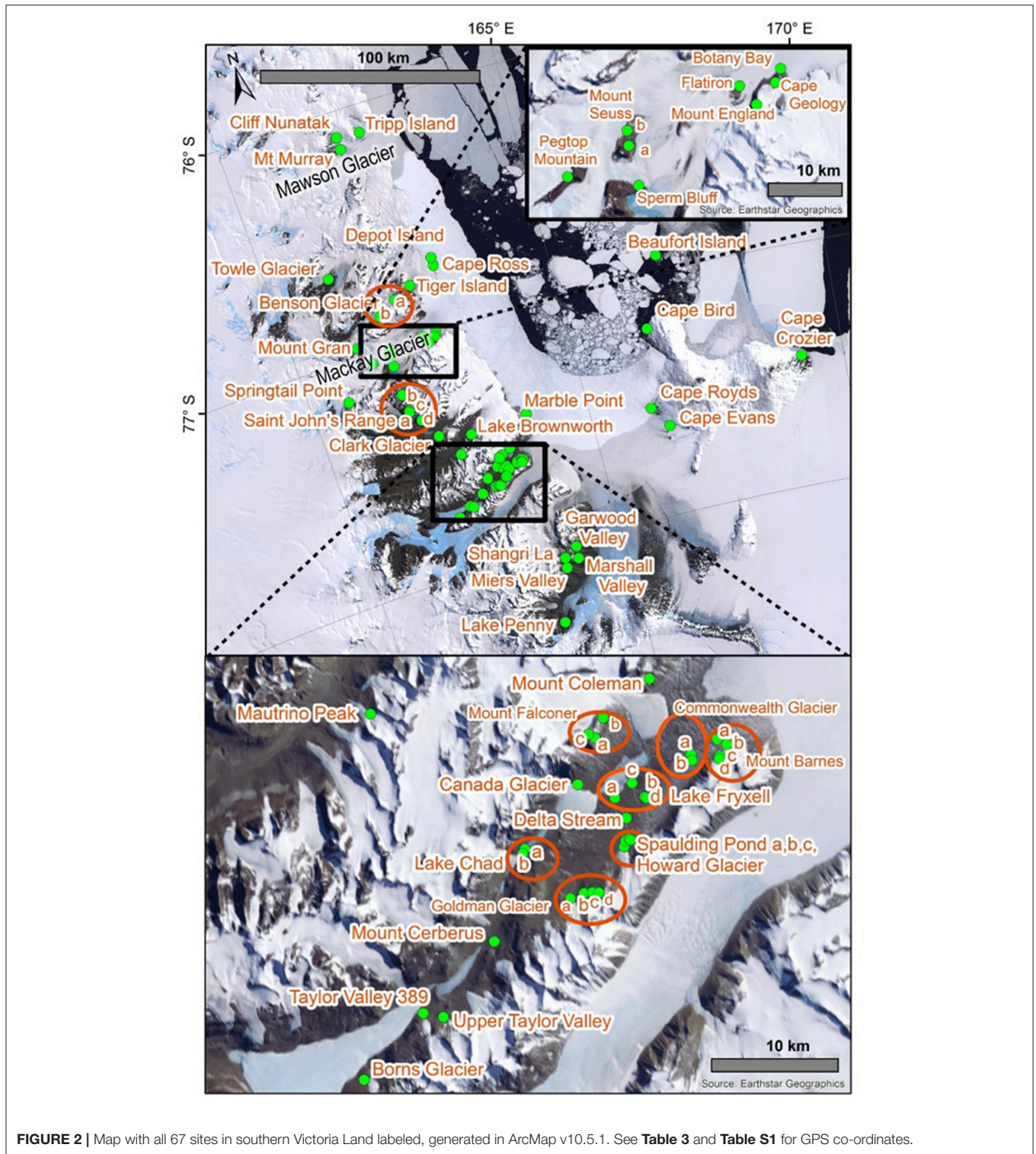
ACBR	Species code	Alignment length (bp)	<i>n</i> (new)	<i>h</i> (new)	% A-T	Intraspecific divergence (maximum)	<i>n</i> divergent lineages (mean)
NVL (<i>n</i> = 299)	KKLO	582	35 (30)	19 (15)	59.0%	2.7%	2 (1.7%)
	CCIS	567	25 (11)	14 (9)	62.6%	6.4%	3 (5.8%)
	FGRI	478	66 (10)	11 (1)	64.5%	9.8%	2 (9.2%)
	CTER	577	173 (5)	68 (1)	63.1%	15.4%	4 (6.9–14.7%)
SVL (<i>n</i> = 1023)	AMON	530	33 (0)	5 (0)	58.3%	10.4%	2 (10.1%)
	CNIV	586	40 (6)	16 (0)	61.7%	4.6%	3 (3.3–4.3%)
	GHOD	422	950 (330)	88 (12)	64.6%	7.3%	3 (5.9–7.3%)
TOTAL			1322 (392)	222 (38)			

Of the number of sequences (*n*) within each alignment, the number of sequences new to this study are shown in parentheses. The number of unique haplotypes are also shown (*h*), along with relative proportion of A and T nucleotides (% A–T) and maximum divergences (uncorrected *p*-distance) for each species. We also highlight the potential for multiple divergent lineages within each of the species, with mean divergences (uncorrected *p*-distance) provided in parentheses.

sequences from Greenslade et al. (2011). GenBank accession numbers for the five sequences new to McGaughan et al. (2008) could not be linked with their respective site data, and 41 specimens from McGaughan et al. (2010) which include those from McGaughan et al. (2008) were also lacking site data. For specimens from Demetras (2010) where collection data were not resolved to precise site, we have used the broader “valley-level” locations (i.e., Garwood, Marshall and Miers Valleys, and Shangri La) in our analyses. Overall, sequences from a total of 88 sites were assessed in this study (**Figures 1, 2; Tables 2, 3 and Table S1**). For specimens with collection data absent, sequences were included in the initial alignment for phylogenetic tree construction and haplotype assignments, and were then removed for subsequent biogeographic analyses.

Each of the individual alignments for the seven species were then assembled into one master alignment which was trimmed to 422 bp and a Maximum Likelihood tree was generated in MEGA v7.0.26 (Kumar et al., 2016) based on the sequence model GTR+I+G (Guindon and Gascuel, 2003; Darriba et al., 2012; JModelTest2), including 1,000 bootstrap replicates. All sequence divergence values included in the current study were also generated in MEGA based on uncorrected *p*-distances.

Haplotype pie charts (**Figures 4–7**) were generated in R v3.5.1 utilizing the packages “mapdata” (Becker et al., 2016) and “mapplots” (Gerritsen, 2014) based on the individual species alignments which were each trimmed to different lengths, depending on the sequences. To retain haplotype-level resolution, phylogenetic tree excerpts



included in these figures were based on the individual species alignments (422–586 bp).

To determine potential barriers to dispersal, Analysis of Molecular Variance (AMOVA) analyses were performed separately for each species in R v3.5.1 utilizing the package

“poppr” with 16,000 permutations (Kamvar et al., 2014, 2015). For this, species alignments that had each been trimmed to between 422 and 586 bp (**Table 4**) were used, and haplotypes were grouped according to their occurrence at sites within particular regions (listed in **Tables 2, 3**) to test the amount of genetic

variability occurring within and between regions. As specimens of *Cryptopygus nivicolus* were all found within the Mackay Glacier region, we performed the AMOVA for this species with the sites Towle Glacier and Springtail Point designated as separate regions.

RESULTS AND DISCUSSION

We compiled all available COI sequences for the seven springtail species in Victoria Land. Sequence coverage ranged from 25 to 950 individuals per species (total $n = 1,322$), with between 5 and 88 haplotypes identified for each species (Figure 3; Table 4). The final alignments for each species varied in length (422–586 bp), largely due to variation in sequence quality, and we treated each species separately for biogeographic analyses to maximize variability within the datasets, although a final alignment trimmed to 422 bp was used to construct the phylogenetic tree (Figure 3). Relative proportions of AT-richness ranged from 58.3 to 64.6% for each species (Table 4), and no insertions or deletions were found. Maximum intraspecific sequence divergences ranged from 2.7 to 15.4% (uncorrected p-distance) and for all seven species, high divergence values (1.7–14.7%) were found among individuals from different geographic locations (Table 4), supported by AMOVAs.

Northern Victoria Land Taxa

Northern Victoria Land is the northern-most Antarctic Conservation Biogeographic Region (ACBR) of the Ross Sea sector and extends from Cape Adare (71.3°S) to the Drygalski Ice Tongue (75.3°S) (Figure 1). We recovered a total of 299 COI sequences from the four springtail species that are present in this area; *Kaylathalia klovstadi* (KKLO; $n = 35$ sequences), *Cryptopygus cisantarcticus* (CCIS; $n = 25$), *Friesea grisea* (FGRI; $n = 66$), and *Cryptopygus terranovus* (CTER; $n = 173$). Each species was collected from a different range of sampling locations and also showed contrasting population genetic structures. For example, two genetically distinct lineages (clades C and D) of *C. terranovus* from sites in the central and northern zones as designated by previous studies (e.g., Fanciulli et al., 2001; Hawes et al., 2010; Carapelli et al., 2017), were highly divergent (>10%) whereas *F. grisea* specimens from the same area all had identical sequences (Figures 4, 5). Similarly, sequences of *K. klovstadi* were only 0.7% divergent between Cape Hallett and Redcastle Ridge, whereas *C. cisantarcticus* specimens differed by 4.3% between the same two collection sites. The Tucker Glacier is a significant barrier to dispersal for Antarctic springtails in the area, with highly distinct (>5%) populations present on either side of this glacier and no shared haplotypes between populations, consistent across species.

Kaylathalia klovstadi (Carpenter, 1902)

The first collections of this species (originally described as *Isotoma klovstadi*) were obtained from Geikie Ridge during the British Antarctic Expedition (1898–1901), after which it was not collected again until 1965 from Ridley Beach at Cape Adare (Wise, 1971). The first five COI sequences obtained for *K.*

klovstadi were from individuals collected at Cape Hallett and published in a study that reclassified the genus from *Isotoma* to *Desoria* (Stevens et al., 2006a). The same five sequences were further used to again reclassify the genus to *Kaylathalia* (Stevens and D'Haese, 2016).

We now include a further 30 previously unpublished COI sequences from three locations (including Cape Hallett), revealing 15 new haplotypes (KKLO-005 to KKLO-019; Tables 2, 4; Tables S2, S3). By expanding the sequence coverage and including two additional sites, we are now able to demonstrate geographical isolation of distinct genetic lineages (2%) for *K. klovstadi* (Figure 4B). When haplotypes in the combined dataset were grouped by region, an AMOVA showed that 75% of the sequence variation occurred between the northern Cape Adare group ($n = 8$), and the southern Cape Hallett and Redcastle Ridge group ($n = 27$) (Table 5; $p < 0.01$). No haplotypes were shared between sites, although Redcastle Ridge was represented by only 3 sequences which were similar to those from Cape Hallett (0.7% divergence).

Based on a study of 40 COII gene sequences, Frati et al. (2001) suggested that the Tucker Glacier was as a major barrier to dispersal for *K. klovstadi*. Currently, no COI sequences were available for *K. klovstadi* from the southern side of the Tucker Glacier and this would benefit from further attention, particularly as strong population genetic structure for the COI gene was observed for *C. cisantarcticus* and *F. grisea* on opposing sides of this glacier (see sections *Cryptopygus cisantarcticus* Wise, 1967 and *Friesea grisea* Schaeffer, 1891).

Cryptopygus cisantarcticus (Wise, 1967)

The first four COI sequences of *C. cisantarcticus* were from specimens collected at Cape Hallett and included in Stevens et al. (2006b), who suggested this species has been isolated for 18–11 MY based on comparisons with other Southern Hemisphere springtail species. An additional 10 sequences were obtained from Crater Cirque and presented as an outgroup in Carapelli et al. (2017).

Here, we added a further 10 sequences from Cape Hallett and a single sequence from Redcastle Ridge, representing nine new haplotypes (CCIS-006 to CCIS-014), and provide comparison among all COI sequences currently available for *C. cisantarcticus* (Tables 2, 4; Tables S2, S4). When haplotypes in the combined dataset were grouped by location, an AMOVA showed that 87% of the sequence variation occurred across the Tucker Glacier, between the northern Cape Hallett and Redcastle Ridge group ($n = 15$) and the southern Crater Cirque population ($n = 10$) (Table 5; $p < 0.01$).

Population genetic structure differed for *C. cisantarcticus* as compared to other species in the area where their sequence distributions overlapped (Figure 4). Sites at Redcastle Ridge and Cape Hallett are in relatively close proximity (<16 km), and divergence between these two locations was very low for *K. klovstadi* (0.7%) while much higher for *C. cisantarcticus* (4.3%). Furthermore, mean divergence for *C. cisantarcticus* across the Tucker Glacier was 5.8%, whereas divergence was higher for *F. grisea*, at >9%.

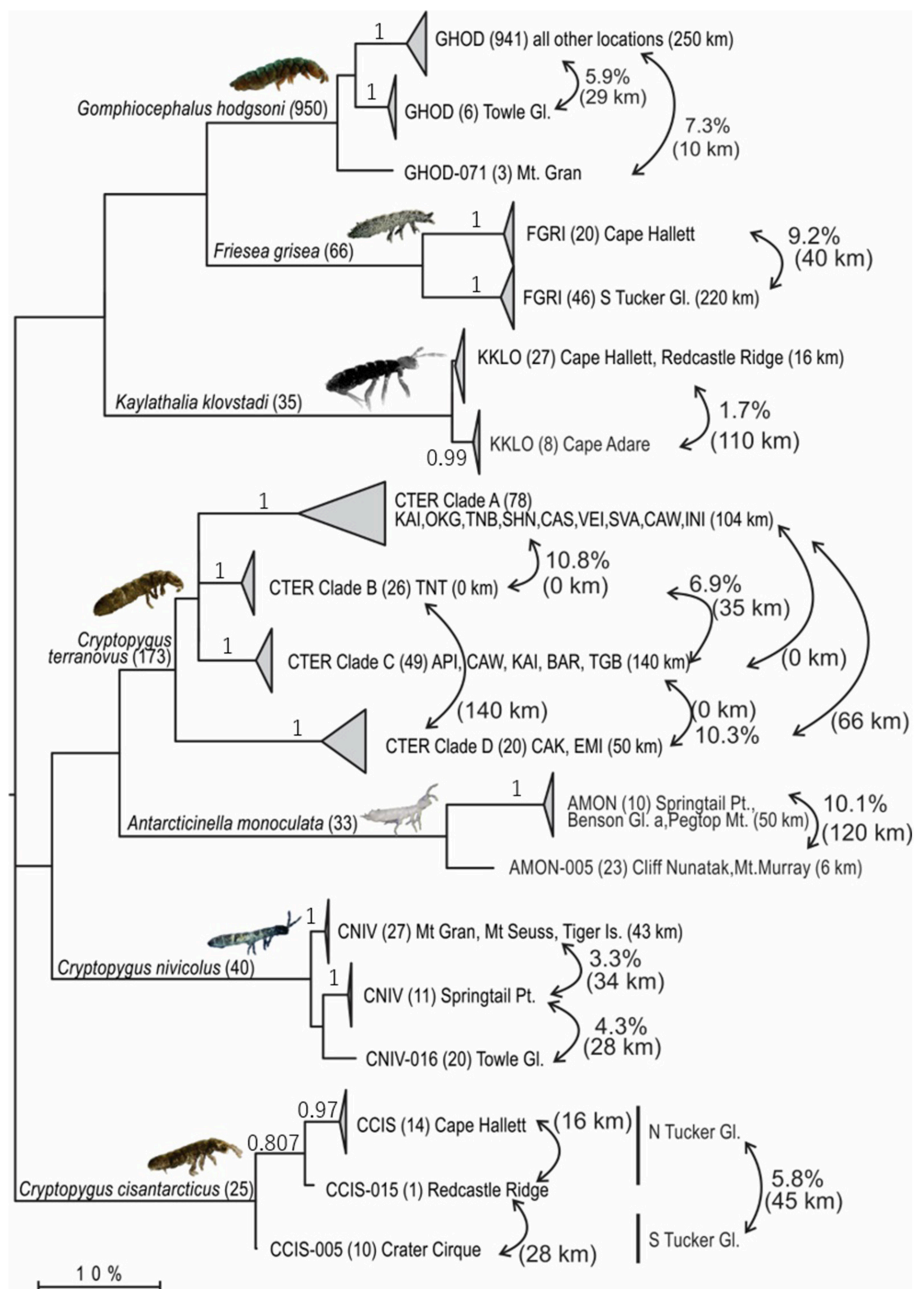


FIGURE 3 | Maximum Likelihood tree (GTR + Γ + I; 1,000 bootstrap replicates) based on the trimmed alignment (422 bp) for all 1,322 COI sequences available for the seven springtail species in Victoria Land. Clades are collapsed proportionately, where each collapsed clade represents a potential cryptic species (>1.7% divergence), and number of sequences are shown in parentheses. Geographic distances within and between clades are provided, along with p-distances between clades, to highlight geographical isolation of well-supported clades (bootstrap values provided on branches, where relevant).

Friesea grisea (Schaeffer, 1891)

This species has been considered “pan-Antarctic” until very recently; specimens of *Friesea grisea* from the Antarctic Peninsula were found to be morphologically distinct to those in Eastern Antarctica (Greenslade, 2018) in agreement with the absence of haplotype sharing between the two locations (>15% divergence; Torricelli et al., 2010b). A total of 55 COI sequences for *F. grisea*

have been published (Torricelli et al., 2010b). In addition, the full mitochondrial genome has previously been assembled for *F. grisea* (GenBank accession KR180288) collected from Kay Island (Torricelli et al., 2010a) and we have included the COI gene region from this sequence in our analyses.

We added an additional 10 unpublished sequences from Cape Hallett (situated to the north of Tucker Glacier;

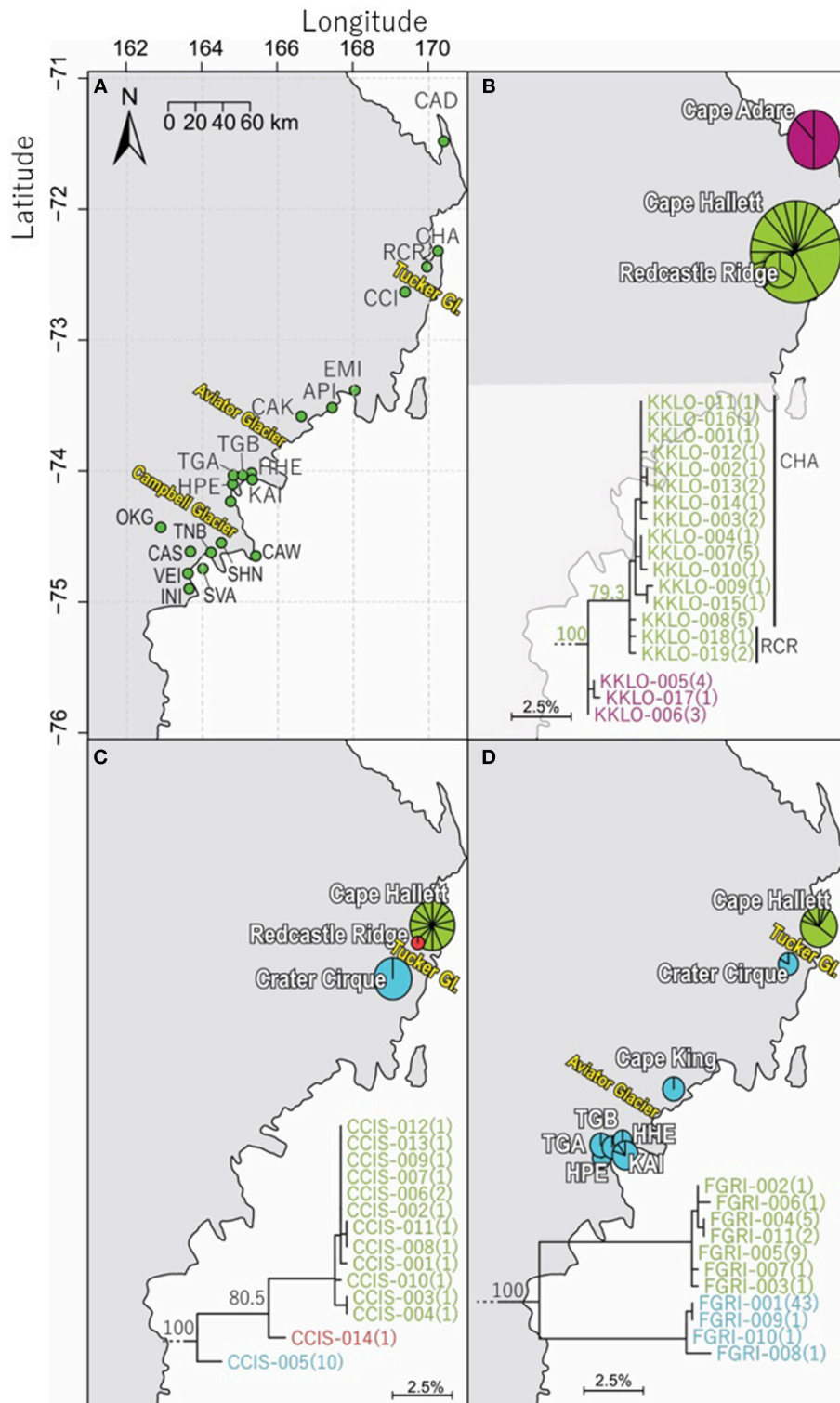


FIGURE 4 | Geographical distribution of COI sequences for three of the four springtail species within northern Victoria Land, with **(A)** an overview of collection sites and major glacial barriers, **(B)** geographical distribution of the 19 haplotypes of *Kaylathalia klovstadi* (KKLO; $n = 35$; 582 bp), **(C)** geographical distribution of the 14 haplotypes of *Cryptopygus cisantarcticus* (CCIS; $n = 25$; 567 bp), and **(D)** geographical distribution of the 11 haplotypes of *Friesea grisea* (FGRI; $n = 66$; 478 bp). Pie charts are proportionate to the number of sequences, centered at collection site co-ordinates. Tucker Glacier is a major barrier of dispersal for CCIS and FGRI, and no sequences of KKLO have been obtained from south of the Tucker Glacier.

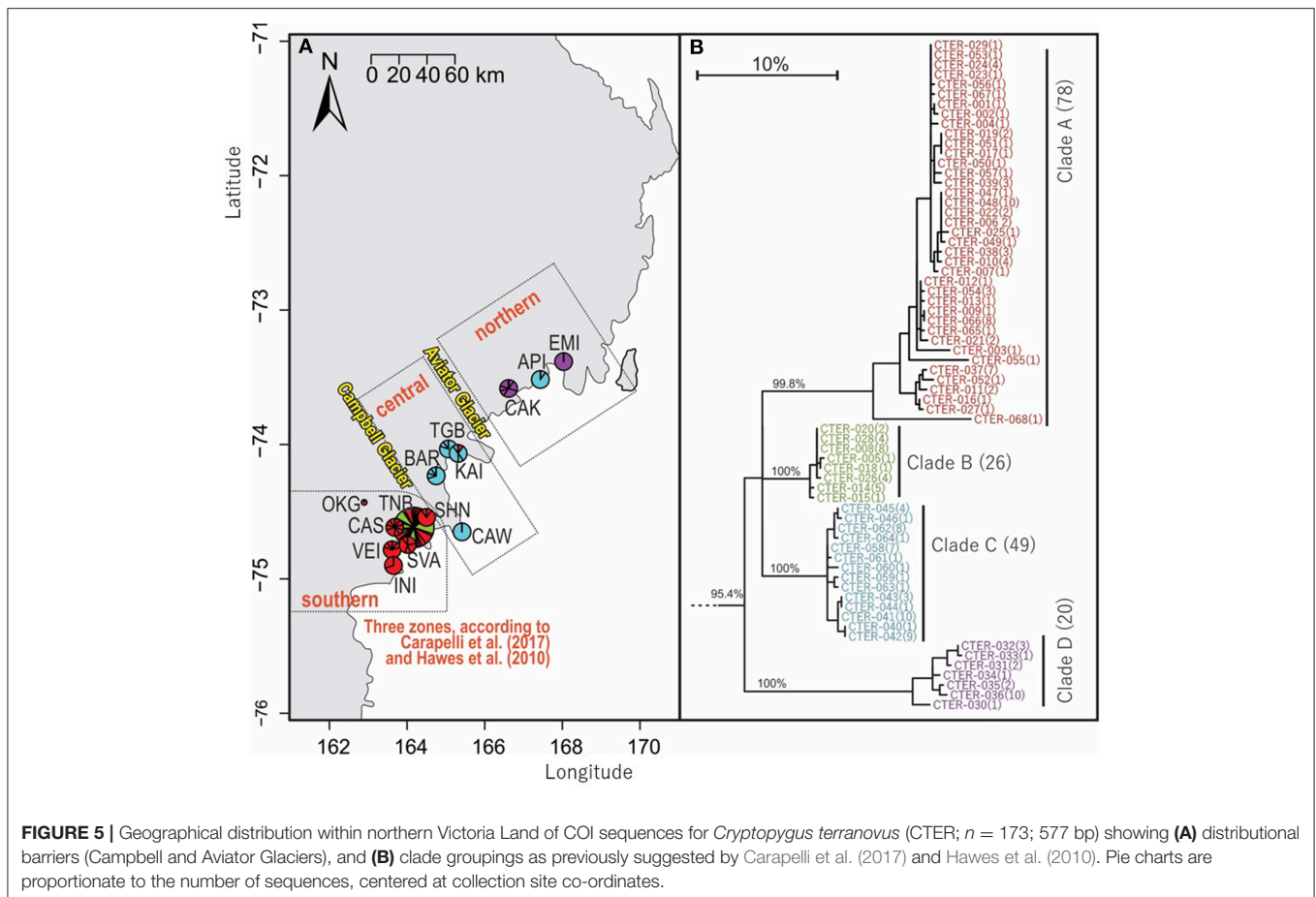


Figure 4), including one new haplotype, for a total of 20 sequences from this site (**Table 2**; **Tables S2, S5**). When haplotypes in the combined dataset were grouped by location, an AMOVA showed that 98.3% of the sequence variation occurred across the Tucker Glacier (**Table 5**; $p < 0.01$), between the northern Cape Hallett population ($n = 20$) and the southern group ($n = 46$) comprised of seven sites across approximately 220 km of coastal terrain (**Figure 4D**). Individuals of *F. grisea* along that entire coastal area south of the Tucker Glacier were within 0.2% divergence, while the population at Cape Hallett was highly differentiated (9.2% divergence), further supporting a lack of dispersal across the Tucker Glacier, in agreement with our findings for *C. cisantarcticus*.

Cryptopygus terranovus (Wise, 1967)

An analysis of allozymes for *Cryptopygus terranovus* (previously known as *Gressittacantha terranova*; Greenslade, 2015) showed northern, central and southern zones of genetic differentiation within the vicinity of Terra Nova Bay that were separated by the Aviator and Campbell Glaciers (**Figure 5A**; Fanciulli et al., 2001). It was also suggested that the population at Apostrophe Island (API; northern zone) was comprised of individuals that had more recently migrated from the central zone. The first COI sequences

($n = 4$) for this species were collected from near the Mario Zuchelli station (C. Beard, pers. comm.) and were reported in Stevens et al. (2006b). Hawes et al. (2010) provided 54 sequences and 25 new haplotypes from fine-scale (~15 km²) sampling in the southern zone (Terra Nova Bay). Broader-scale sampling by Carapelli et al. (2017) provided a further 114 sequences from 11 sites, including 38 new haplotypes. Each of these three genetic studies provided additional evidence to support the division of *C. terranovus* into the three zones as first suggested by Fanciulli et al. (2001) and our AMOVA analyses found that 59.8% of the sequence variation occurred between the three zones (**Table 5**; $p < 0.01$).

We provide a further five sequences (no new haplotypes) from near Jang Bogo Station in Terra Nova Bay, and O’Kane Glacier (**Table 2**; **Tables S2, S6**). We analyzed a total of 173 COI sequences for *C. terranovus* and identified 68 unique haplotypes from the 577 bp alignment. Sequences were grouped into the four distinct clades that have been previously identified for this species (Hawes et al., 2010; Carapelli et al., 2017). Overall, each clade contained between 20 and 78 sequences, mean sequence divergences within each clade ranged from 0.48 to 2.09%, and mean sequence divergences between clades ranged from 6.9 to 14.7% (**Figure 5**; **Table 4**; **Table S6**).

TABLE 5 | Analysis of molecular variance (AMOVA) (Excoffier et al., 1992) results for the seven species of Collembola in Victoria Land, as implemented in R v3.5.1 using the package “poppr” (Kamvar et al., 2014, 2015).

Species	Source of variation	df	Sum of squares	Variance components	Percentage of variation	p
<i>Kaylathalia klovstadi</i>	Among regions	1	58.47	4.61	75.02	<0.001
	Within regions	33	50.68	1.54	24.98	
<i>Cryptopygus cisantarcticus</i>	Among regions	1	185.33	15.25	87.00	<0.001
	Within regions	23	52.41	2.28	13.00	
<i>Friesea grisea</i>	Among regions	1	565.35	20.27	98.27	<0.001
	Within regions	64	22.88	0.36	1.73	
<i>Cryptopygus terranovus</i>	Among regions	2	1136.31	11.53	59.80	<0.001
	Within regions	170	1317.57	7.75	40.20	
<i>Antarcticinella monoculata</i>	Among regions	1	369.12	26.46	98.75	<0.001
	Within regions	31	10.40	0.34	1.25	
<i>Cryptopygus nivicolus</i>	Among regions	2	42.26	1.89	35.03	<0.001
	Within regions	37	129.69	3.51	64.97	
<i>Gomphiocephalus hodgsoni</i>	Among regions	4	409.03	0.63	39.86	<0.001
	Within regions	899	859.23	0.96	60.14	

Sites were grouped into regions according to **Tables 2, 3**, while Towle Glacier and Springtail Point were each designated as separate regions for *C. nivicolus*. Statistical significance of variance components were tested with 16,000 permutations.

Southern Victoria Land Taxa

This region extends from the Drygalski Ice Tongue to the north and the Koettlitz Glacier to the south, and includes the Mackay Glacier region as well as the Victoria Land Dry Valleys including Victoria, Wright, Taylor, Garwood, Marshall, and Miers Valleys (**Figures 1, 2**). We analyzed a total of 1,023 COI sequences from the three springtail species that are currently known from this region; *Antarcticinella monoculata* (AMON; $n = 33$), *Cryptopygus nivicolus* (CNIV; $n = 40$), and *Gomphiocephalus hodgsoni* (GHOD; $n = 950$). All three species are range-restricted, with genetic differentiation found among different geographic locations. However, patterns of population structure vary among the three species. For example, individuals of *G. hodgsoni* at Mount Gran ($n = 3$) showed sequence divergences of 7.6% (Bennett et al., 2016), while individuals of *C. nivicolus* from Mount Gran ($n = 7$) were similar to those from other nearby sites such as Mount Seuss and Mount England (Beet et al., 2016). No sites were found with all three species present. Both *C. nivicolus* and *A. monoculata* were found at Springtail Point, while *C. nivicolus* and *G. hodgsoni* were both found at 5 sites (Towle Glacier, Tiger Island, and Mounts Seuss, Gran and England).

Antarcticinella monoculata (Salmon, 1965)

In total, 33 COI sequences were available for *A. monoculata* (Beet et al., 2016; Bennett et al., 2016) and all specimens were collected from within a 250 km² range of the Mackay Glacier (**Figure 6A**; **Table 3**; **Table S7**). Sequences from the two northern-most sites, Mount Murray and Cliff Nunatak, were all the same haplotype (AMON-005) and were divergent from the other four haplotypes that were found at the more southerly sites (10.1% mean p-distance). An AMOVA revealed that 98.7% of the genetic variation occurred between these two geographically isolated populations (**Table 5**; $p < 0.01$).

Cryptopygus nivicolus (Salmon, 1965)

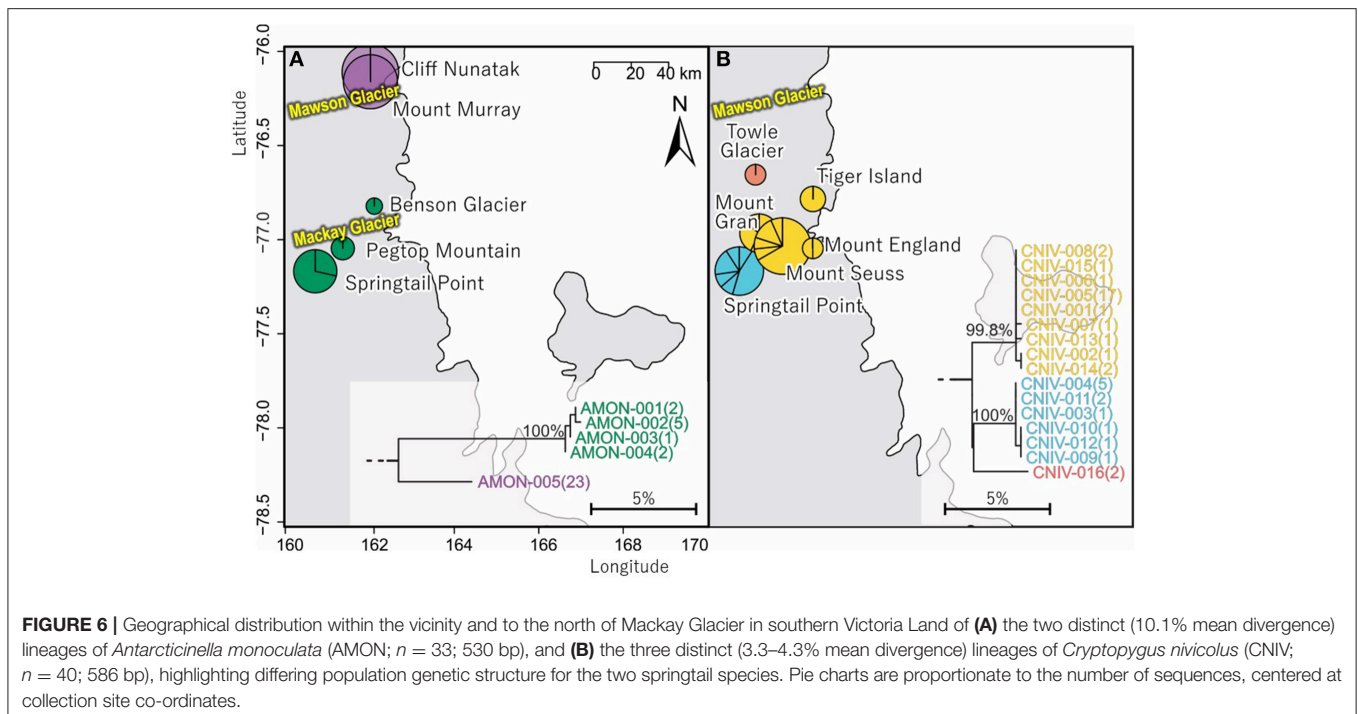
The first COI sequences ($n = 2$) for this species were obtained from Mount England (Stevens et al., 2006b). More recently, Bennett et al. (2016) and Beet et al. (2016) contributed another 32 sequences from an additional 5 sites, showing genetic divergence among habitats.

A further 6 sequences were included as part of our study (**Tables 3, 4**; **Tables S2, S7**). In total, we identified 16 haplotypes from the 40 COI sequences for this species, clustering into three distinct groups (3.3–4.3% mean divergence) corresponding to different geographic locations (**Figure 6B**). Springtail Point and the Towle Glacier site each had unique *C. nivicolus* sequences that were not found at any other site. The remaining nine haplotypes ($n = 27$ sequences) occurred across four sites in the vicinity of the Mackay Glacier (Mounts Gran, Seuss, England and Tiger Island; **Figure 6B**), suggesting individuals may have recently dispersed among these sites (within 40 km of each other). Possible explanations for connectivity among these sites include dispersal via meltwater streams or in marine, nearshore environments when sea ice is absent along coastal boundaries. An AMOVA found that when sites at Springtail Point and Towle Glacier were designated as separate regions, only 35% of the genetic variation occurred between each of the three regions (**Table 5**; $p < 0.01$). However, genetically divergent populations of *C. nivicolus* were found at Springtail Point and the Towle Glacier site (**Figure 6B**), highlighting the importance of conservation strategies to prevent human-mediated transport of genetic variants between these currently isolated populations.

Gomphiocephalus hodgsoni (Carpenter, 1908)

Previous Studies

The first COI sequences for *G. hodgsoni* were contributed by a study that also examined allozymes and was focused on phylogeography (Stevens and Hogg, 2003). COI sequences were



obtained for 45 individuals from 21 localities, and 14 unique haplotypes were identified (A–N). Among their findings it was suggested that two sympatric populations were present in Taylor Valley (1.5% divergence), and that recent transport (possibly by birds or humans) had occurred between Ross Island and Granite Harbor as similar haplotypes were shared between these locations. Sequences from Stevens and Hogg (2003) were included in their subsequent study (Stevens and Hogg, 2006), where genetically divergent populations were identified at Beaufort Island and in Taylor Valley, when compared to the remaining sites on Ross Island ($n = 5$ sites) and the continental mainland ($n = 11$ sites, including Taylor Valley). However, the Beaufort Island haplotype (our haplotype GHOD-010) has since been found at additional sites on the continent (see **Table S9**), while the unique Taylor Valley haplotype (our haplotype GHOD-011) remains restricted to Taylor Valley and belongs to Group Y in Collins and Hogg (2015) and Nolan et al. (2006).

A following study of *G. hodgsoni* (Nolan et al., 2006) targeted the two sympatric groups in Taylor Valley that had previously been identified by Stevens and Hogg (2003). The eight Taylor Valley sequences from Stevens and Hogg (2003) were also incorporated into their dataset, and 10 unique haplotypes (A–J) were identified from the combined dataset ($n = 48$). The study concluded that the two sympatric phylogroups (haplotypes A–F = group X; G–J = group Y) probably diverged less than 1 million years ago, around the time when ancient Lake Washburn flooded the Valley (40 kya). Ancestors of the two populations probably survived in isolated refugia at either end of the Valley (group Y inland and group X in higher elevation coastal areas), recolonizing the Valley and reconnecting following the retreat of Lake Washburn.

An additional five COI sequences and two new haplotypes were recovered from GenBank (McGaughan et al., 2008). Unfortunately, we were unable to determine which sequences corresponded to haplotype codes referred to in the manuscript. As these sequences were included in the subsequent study of McGaughan et al. (2010), they have also been included in our study. The McGaughan et al. (2008) study included sequences from Stevens and Hogg (2003) and identified 20 haplotypes (G1–G20) from a total dataset of 96 sequences from the wider McMurdo Dry Valleys and Ross Island area. Divergences of *G. hodgsoni* COI sequences were up to 2.1%, suggesting the populations had diverged within the last million years (based on molecular clock calibration of 1.5–2.3%/my e.g., Brower, 1994; Juan et al., 1996; Quek et al., 2004). Further, haplotype diversity tended to be higher at more inland sites and particularly at higher altitudes, suggesting this was a result of persistence in refugial habitats. McGaughan et al. (2008) also suggested that Ross Island individuals were derived from a founder population that originated from the Dry Valleys.

McGaughan et al. (2010) compared patterns of sequence divergences of *G. hodgsoni* to those of *Cryptopygus antarcticus* from the Antarctic Peninsula and provided COI and COII sequences for both taxa. Existing *G. hodgsoni* COI sequences from Stevens and Hogg (2003) and McGaughan et al. (2008) were also included, and 45 haplotypes (H1–H45) were identified from their final alignment of 289 individuals (471 bp). In reviewing these sequences, we were able to identify exact site information for all but 31 specimens (see **Table S9**). Key findings from McGaughan et al. (2010) included limited haplotype sharing between local sites within both the Peninsula and continental areas, and that survival in refugial habitats was

likely to have occurred on a Pleistocene timescale. However, the Peninsula contained more rare haplotypes and it was suggested that differences in population structure were a result of landscape differences and colonization events between the two localities. Greenslade et al. (2011) provided three additional COI sequences (658 bp) for the existing haplotype H43 from McGaughan et al. (2010) (our haplotype GHOD-046) in a study which confirmed *Gomphiocephalus* as a distinct genus.

A further 90 *G. hodgsoni* COI sequences were contributed by Demetras (2010) from the southern Dry Valleys area (Marshall, Garwood and Miers Valleys) with limited diversity found (10 haplotypes TABS Gh1–Gh10; <0.8% divergence). Based on a haplotype network analysis, it was suggested that all 10 haplotypes were derived from a single lineage (300–500 kya). Possible reasons for the low diversity in *G. hodgsoni* from the southern Dry Valleys include more recent recolonization of this area or bottleneck events and differences in landscapes as compared to the larger Dry Valleys (e.g., Taylor) where there is much higher COI diversity. As the data from Demetras (2010) were previously unpublished, we have now uploaded them to BOLD and added them to our analyses.

Collins and Hogg (2015) provided a further 151 *G. hodgsoni* COI sequences as part of a two-hourly time-series of pitfall trap collections from Spaulding Pond in Taylor Valley. This study targeted the two main “X” and “Y” haplotype groupings previously reported by Nolan et al. (2006), known to occur in sympatry at Spaulding Pond (Gh1–Gh12 = Y; Gh13–Gh19 = X). More individuals were found from group Y ($n = 120$) relative to group X ($n = 31$). As group Y is thought to have recolonized from inland “colder” refugial sites, this study inferred that the site was dominated by intrinsically cold-adapted individuals, and that the relative proportions of X and Y individuals could change with a warming climate. Furthermore, activity of individuals from the X and Y haplotype lineages during each two-hourly pitfall trap collection was more closely linked to air temperature than any of the other measured environmental variables. This highlights the potential for temporal shifts in the genetic structure of populations as a consequence of environmental changes.

An additional 67 sequences (658 bp; 16 haplotypes, 8 new) were contributed by Bennett et al. (2016), along with sequences from *A. monoculata* and *C. nivicolus*. This study was focused on past isolation events using a molecular clock analysis. They concluded that isolation was likely to have occurred during the last 4 million years and that glaciation events since that time have further contributed to the high COI diversity within both *G. hodgsoni* and *C. nivicolus*. In particular, specimens of *G. hodgsoni* from Mount Gran (our haplotype GHOD-071) were found to be highly divergent (>7%) from the remaining sampled dataset. In a study focused on genetic diversity of Collembola from sites in the vicinity of Mackay Glacier and further north, Beet et al. (2016) contributed 66 new sequences (527 bp; 5 new haplotypes). Sequences from Bennett et al. (2016) were also included in Beet et al. (2016) analyses and, based on the combined dataset, it was hypothesized that populations may have been isolated for 3–5 million years, consistent with a collapse and reformation of the West Antarctic Ice Sheet.

Our Findings

As the most widespread and well-studied springtail in the Ross Sea region, the dataset of COI sequences for *G. hodgsoni* ($n = 950$) was larger than that of any other species and represented specimens from 67 sites (Tables 3, 4; Tables S8, S9). Overall, 620 sequences were available from nine published studies as outlined above, and a further 330 previously unpublished sequences have now been added here (Table S2). The majority of sequences were <2.6% divergent, while distinct populations were present at Towle Glacier ($n = 6$; 5.9% divergence) and Mount Gran ($n = 3$; 7.3% divergence) which are located further inland (Figure 7). Similarly, genetically divergent populations of *C. nivicolus* were present at the inland sites Towle Glacier and Springtail Point, suggesting that greater connectivity occurs among coastal sites. Overall, an AMOVA revealed that the majority (>60%) of haplotype diversity occurred within regions (Table 5; $p < 0.01$), particularly as the highly diverged individuals from sites Towle Glacier and Mount Gran were sequenced at low numbers.

Tripp Island is approximately 70 km from other sites and located to the north of Mawson glacier which appeared to be a dispersal barrier for *A. monoculata* (>10% divergence). However, individuals of *G. hodgsoni* from Tripp Island were not genetically divergent (<0.5%) relative to other locations. In contrast, individuals of *G. hodgsoni* at Mount Gran <10 km from other sites (Figure 7) were highly genetically divergent (>7%). These findings highlight that geographical barriers to dispersal such as glaciers, and lack of water transport between locations, which is unlikely for sites further inland, have a stronger influence on the population genetic structures of Antarctic springtails than distance alone.

CONCLUDING DISCUSSION

In this study we examined the available COI sequence data for all seven of the Collembola species that occur within Victoria Land (71° to 78.5°S). All species harbored distinct lineages (1.4–14.7% mean sequence divergences) that were isolated by geological or glaciological features, rather than by distance alone. Such levels of COI diversity are suggestive of long-term isolation and lack of dispersal among locations.

In several cases, the distribution of genetic variants among sites differed for the different species. Indeed, differing distributional patterns were previously reported in a large-scale study of the three species *C. terranovus*, *F. grisea*, and *G. hodgsoni* (Caruso et al., 2009). Here, we showed that sequence divergence for *F. grisea* (0.2%) and *C. terranovus* (10.8%) differed greatly among the same sites in northern Victoria Land. As a possible explanation, *F. grisea* may have more recently dispersed throughout this area whereas *C. terranovus* may have remained isolated by the Aviator and Campbell Glaciers (e.g., Fanciulli et al., 2001; Carapelli et al., 2017). In southern Victoria Land (SVL), Springtail Point harbored a distinct lineage (>3.3% divergence) of *C. nivicolus* whereas *A. monoculata* from Springtail Point was highly similar (<0.9% divergence) to individuals from nearby sites.

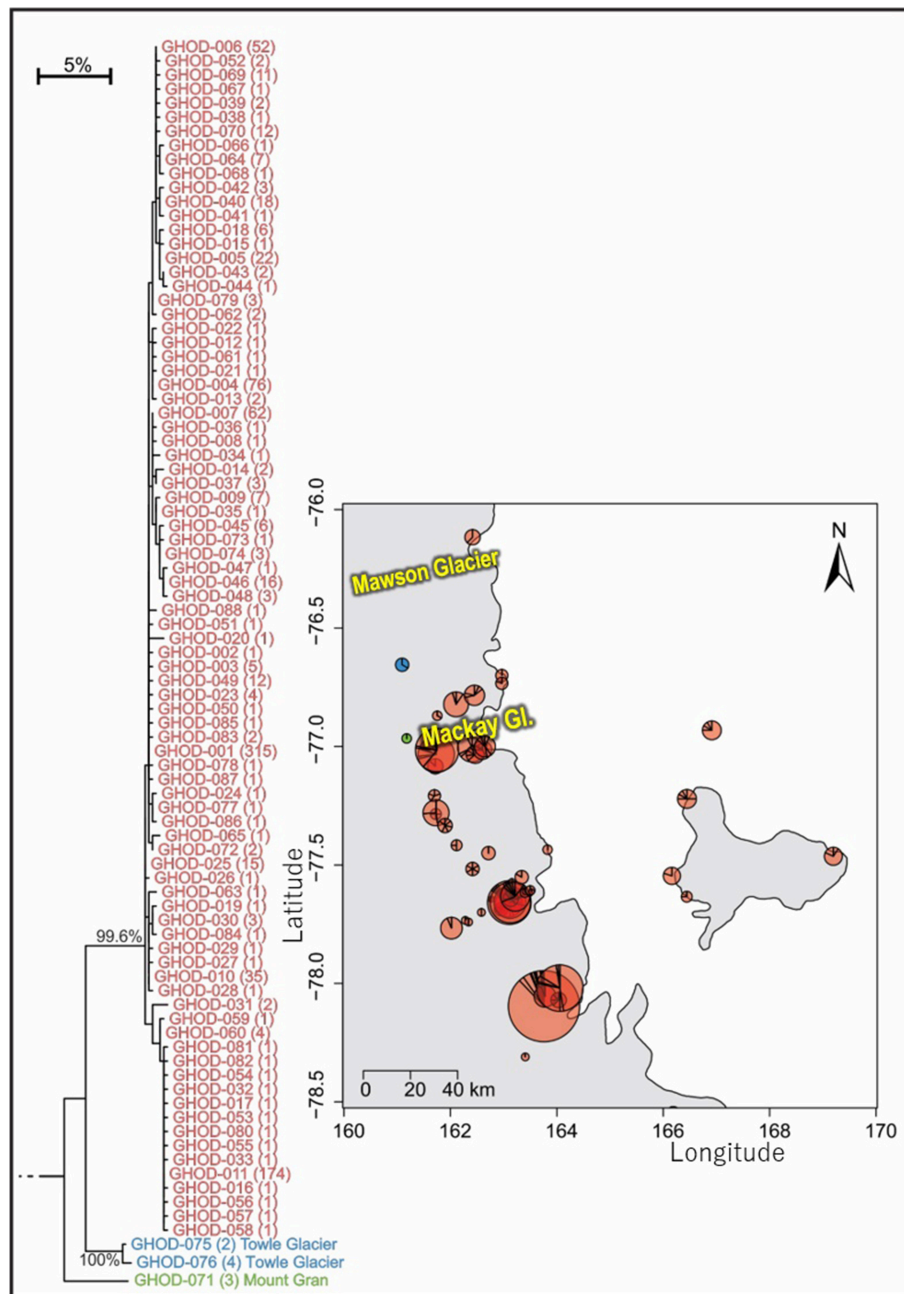


FIGURE 7 | Geographical distribution of the three distinct (5.9–7.3% mean divergence) lineages of *Gomphiocephalus hodgsoni* within southern Victoria Land for specimens where collection data were available (GHOD; $n = 904$; 422 bp), showing widespread distribution (500 km reach) of the main group of haplotypes ($n = 895$; 2.6% mean divergence) and distinct populations at both Mount Gran and Towle Glacier sites, further inland. Pie charts are proportionate to the number of sequences, centered at collection site co-ordinates. See **Tables S8, S9** for further haplotype and collection details for each specimen.

Similarly, the Mount Gran population of *G. hodgsoni* was highly distinct (7.3% divergence) whereas sequences of *C. nivicolus* from Mount Gran were identical to those from other sites. Populations to the north of Mawson Glacier were genetically distinct for *A. monoculata* (>10% sequence divergence; Cliff Nunatak and Mount Murray) while the *G. hodgsoni* population (Tripp Island) was genetically similar (0.5%), even though

these sites are geographically isolated from other sites by >70 km (**Figures 2, 6, 7**).

Given the high levels of genetic divergence among populations for all seven springtail species (1.7–14.7%), we highlight the potential that these distinct lineages could be cryptic species. Cryptic diversity has been suggested for *C. terranovus* in NVL (Hawes et al., 2010; Carapelli et al., 2017) and *G. hodgsoni*,

C. nivicolus, and *A. monoculata* in SVL (Beet et al., 2016). Suggestions of cryptic diversity for the springtail *F. grisea* between NVL and the Antarctic Peninsula (Torricelli et al., 2010a,b) have now been validated through morphological differences and redescription of the species (Greenslade, 2018). However, the potential for cryptic variability among Victoria Land specimens of *F. grisea* has not previously been suggested, although the Cape Hallett population has been identified as genetically distinct, possibly resulting from pre-Pleistocene isolation (Torricelli et al., 2010b).

The Tucker Glacier in NVL provides an example of a small-scale (<16 km) barrier to springtail dispersal as genetically distinct (>5% sequence divergence) populations of both *C. cisanantarcticus* and *F. grisea* have been found either side of the glacier. Previous studies using the COII gene have also demonstrated this for *K. klovstadi* (Frati et al., 2001; Stevens et al., 2007), suggesting the potential for microspeciation processes occurring at these sites. In SVL, we show that the phylogeographic patterns differed for each of the three species, and we highlight Springtail Point, Towle Glacier, Mount Gran and sites to the north of Mawson Glacier as possible sites of conservation focus, as these locations harbored genetically divergent (3.3–10.1%) populations. The vicinity of the Drygalski Ice Tongue continues to provide a current barrier to dispersal for springtail taxa between NVL and SVL and we found no evidence of gene flow or species sharing between these regions.

Future studies focused on longer COI sequences (or indeed, additional genetic markers), are likely to reveal further genetic diversity for springtails in Victoria Land. The potential for temporal changes in genetic diversity as a consequence of environmental changes such as climate warming has also been highlighted. Next generation sequencing and metabarcoding approaches for genomic monitoring will be important for future studies, particularly to detect spatial and temporal shifts in genetic diversity. With a warming climate, future glacier melt will provide additional opportunities for the dispersal of springtails (via flotation) while also exposing new habitats for colonization.

The Antarctic Conservation Biogeographic Regions (Terauds et al., 2012; Terauds and Lee, 2016) provide a geographical framework for assessing broad-scale biological diversity. However, we suggest that knowledge of local-scale patterns of genetic diversity will be critical for addressing ecological and physiological questions, such as those pertaining to dispersal, and responses to climate changes. The understanding of relevant temporal and spatial scales as well as the current distribution of genetic diversity is essential to identify current barriers to dispersal as well as sites that harbor unique genetic resources and, therefore, areas of conservation value.

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DATA AVAILABILITY

All previously unpublished sequences and associated data are available in the dataset “DSANTSP18” on BOLD. GenBank accession numbers and BOLD IDs are provided in supplementary data sheets for all sequences included in this manuscript. Original trace files can be found associated with specimens in the BOLD dataset, where available. For any additional information, contact GC: gec9@students.waikato.ac.nz/.

AUTHOR CONTRIBUTIONS

GC and IH conceived the study. IH collected new specimens from NVL, and processed these with GC. GC and IH collected and processed additional specimens from SVL. All alignments and analyses were performed by GC, with assistance from AB using R. The manuscript was prepared by GC, with contributions from all co-authors.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fevo.2019.00076/full#supplementary-material>

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The reviewer, BV, declared past collaborations with one of the authors, PC, to the handling Editor.

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