



Sensory Evolution and Ecology of Early Turtles Revealed by Digital Endocranial Reconstructions

Stephan Lautenschlager^{1†}, Gabriel S. Ferreira^{2,3,4†} and Ingmar Werneburg^{3,4,5*}

¹ School of Geography, Earth and Environmental Sciences, University of Birmingham, Birmingham, United Kingdom, ² Biology Department, Faculty of Philosophy, Science, and Letters at Ribeirão Preto, University of São Paulo, Ribeirão Preto, Brazil,

³ Senckenberg Center for Human Evolution and Palaeoenvironment (HEP), Eberhard Karls Universität, Tübingen, Germany,

⁴ Fachbereich Geowissenschaften der Eberhard Karls Universität, Tübingen, Germany, ⁵ Museum für Naturkunde, Leibniz-Institut für Evolutions- und Biodiversitätsforschung, Berlin, Germany

OPEN ACCESS

Edited by:

Corwin Sullivan,
Institute of Vertebrate Paleontology
and Paleoanthropology (CAS), China

Reviewed by:

Graciela Helena Piñeiro,
University of the Republic, Uruguay
Manuel F. G. Weinkauff,
Université de Genève, Switzerland
Márton Rabi,
Martin Luther University of
Halle-Wittenberg, Germany

*Correspondence:

Ingmar Werneburg
ingmar.werneburg@senckenberg.de

[†]These authors have contributed
equally to this work.

Specialty section:

This article was submitted to
Paleontology,
a section of the journal
Frontiers in Ecology and Evolution

Received: 24 October 2017

Accepted: 11 January 2018

Published: 05 February 2018

Citation:

Lautenschlager S, Ferreira GS and
Werneburg I (2018) Sensory Evolution
and Ecology of Early Turtles Revealed
by Digital Endocranial
Reconstructions.
Front. Ecol. Evol. 6:7.
doi: 10.3389/fevo.2018.00007

In the past few years, new fossil finds and novel methodological approaches have prompted intensive discussions about the phylogenetic affinities of turtles and rekindled the debate on their ecological origin, with very distinct scenarios, such as fossoriality and aquatic habitat occupation, proposed for the earliest stem-turtles. While research has focused largely on the origin of the anapsid skull and unique postcranial anatomy, little is known about the endocranial anatomy of turtles. Here, we provide 3D digital reconstructions and comparative descriptions of the brain, nasal cavity, neurovascular structures and endosseous labyrinth of *Proganochelys quenstedti*, one of the earliest stem-turtles, as well as other turtle taxa. Our results demonstrate that *P. quenstedti* retained a simple tube-like brain morphology with poorly differentiated regions and mediocre hearing and vision, but a well-developed olfactory sense. Endocast shape analysis indicates that an increase in size and regionalization of the brain took place in the course of turtle evolution, achieving an endocast diversity comparable to other amniote groups. Based on the new evidence presented herein, we further conclude that *P. quenstedti* was a highly terrestrial, but most likely not fossorial, taxon.

Keywords: neuroanatomy, sensory adaptation, 3D visualization, digital endocast, stem-turtles, turtle origin

INTRODUCTION

Turtles (Testudinata sensu Joyce et al., 2004) are a diverse group of reptiles with an unusual “bauplan” fundamentally different from that of other amniotes. Unique morphological characters, including the anapsid cranial configuration, which lacks temporal fenestrations, and the presence of a bony shell formed by a dorsal carapace and a ventral plastron have long obfuscated the phylogenetic affinities of turtles (Rieppel, 2007; Lyson et al., 2010). While most molecular studies have recovered turtles nested within diapsid reptiles and often as a sister-group to Archosauria (birds and crocodiles) (Hedges and Poling, 1999; Wang et al., 2013; Field et al., 2014), most studies based on comparative anatomy have placed turtles outside of Diapsida (Gauthier et al., 1988; Lee, 1997; Werneburg and Sánchez-Villagra, 2009; Neenan et al., 2013; Scheyer et al., 2017) or alternatively inside Lepidosauromorpha (deBraga and Rieppel, 1997; Rieppel and Reisz, 1999; Li et al., 2008; Liu et al., 2011). The scant fossil record of stem-turtles (i.e., non-Testudines Testudinata) has further obscured the evolutionary origin of this group. Recent discoveries of new species and reanalysis of existing specimens with novel methodological approaches (e.g., computed

tomography and digital visualization) have provided new data to the debate of turtle ancestry (Li et al., 2008; Bever et al., 2015; Schoch and Sues, 2015). These studies found support for the diapsid origin of turtles and produced potential evidence for closure of the temporal fenestrae early in their evolutionary history (Schoch and Sues, 2015; Werneburg, 2015; Lyson et al., 2016).

Regarding the environmental origin of the group, although all Triassic turtles were clearly terrestrial (Joyce, 2015), data provided by recently described taxa have painted an ambiguous picture regarding the paleoecological setting in which the Testudinata ancestors evolved. While the earliest known potential proto-turtle (i.e., non-Testudinata Pantestudines) *Eunotosaurus africanus* (ca. 260 Ma) has been found in terrestrial environments (Lyson et al., 2016), the somewhat younger *Pappochelys rosinae* (ca. 240 Ma) and *Odontochelys semitestacea* (ca. 220 Ma) were retrieved from lacustrine and deltaic deposits and were considered to have been semi-aquatic (Li et al., 2008; Rieppel, 2013; Schoch and Sues, 2015). In the last two taxa, the dorsoventrally flattened, expanded ribs, and thickened gastralia have been interpreted as adaptations for buoyancy control in an aquatic environment (Schoch and Sues, 2015). In contrast, the morphology of the ribs, as well as the more rigid body wall, powerful forelimbs and triangular skull, have been considered to represent adaptations to fossoriality in *E. africanus* (Lyson et al., 2016). On the other hand, the type localities of both *P. rosinae* and *O. semitestacea* have also yielded terrestrial taxa (Joyce, 2015; Schoch and Sues, 2017) and, in fact, terrestrial diapsid remains are dominant at the type locality of the former (Schoch and Sues, 2015). Additionally, Joyce (2015) argued in favor of *Odontochelys semitestacea* as a terrestrial proto-turtle, based on its phalangeal formula. Hence, terrestrial, fossorial and semi-aquatic habits have all been suggested for the early stages of turtle evolution, before the origin of the protective shell characteristic of the definitely terrestrial stem-turtles (Joyce and Gauthier, 2004; Scheyer and Sander, 2007; Joyce, 2015).

While research on early turtles has focused largely on the acquisition of the anapsid condition and the evolution of the postcranial anatomy employing comparative morphology, histology and genetics, little is known about the endocranial anatomy of stem-turtles (or indeed turtles in general). Using micro-computed tomography (μ CT) scanning and digital visualization, we here provide a reconstruction of the endocranial anatomy of *Proganochelys quenstedti*, one of the earliest testudinates from the Late Triassic of Germany. We further compare the reconstructed brain anatomy with different stem- and crown-turtles (Testudines) and other vertebrate taxa using endocast outline analysis to elucidate related anatomical and ecological aspects of turtle origins.

MATERIALS AND METHODS

For digital reconstruction of endocranial anatomy (brain, inner ear, neurovascular structures, nasal cavity) two specimens of *Proganochelys quenstedti* from the Late Triassic of Germany were studied: MB 1910.45.2 (Museum für Naturkunde Berlin) from

the Baerecke and Limpricht Quarry, Halberstadt (Jaekel, 1918), and SMNS 16980 (Staatliches Museum für Naturkunde Stuttgart) from the *Plateosaurus*-quarry in Trossingen (Gaffney, 1990). Both specimens consist of nearly complete and articulated cranial skeletons. MB 1910.45.2 shows substantial taphonomic artifacts in the form of anteroposterior shearing and some moderate mediolateral crushing and deformation. However, these artifacts only marginally affect the braincase and the digital reconstruction of the various endocranial structures (see Results for more details).

MB 1910.45.2 was CT scanned at the Leibniz-Institut für Zoo- und Wildtierforschung Berlin/Germany (IZW) using a Toshiba Aquilon ONE medical CT scanner. Scanning parameters were set at 225 kV and 300 μ A resulting in an image stack of 512 \times 512 \times 213 pixels and a voxel size of 2.0 mm per slice. The dataset was subsequently “upsampled” (1024 \times 1024 \times 426 pixels, 0.5 mm effective voxel size) by averaging the existing slice data. This process does not increase the actual resolution of the data, but provides more slices available for segmentation permitting clearer identification of features and resulting in smoother surface models.

SMNS 16980 was scanned at the Riedberg Campus of Goethe-Universität Frankfurt/Germany using a Phoenix Nanotom m scanner (Werneburg et al., 2015a). Due to its relatively large size, the specimen was scanned in three stages. The resulting image stacks were combined into a single stack with 3583 \times 4011 \times 5658 pixels and a voxel size of 0.025 mm per slice. The dataset was subsequently downsampled (870 \times 954 \times 1161 pixels, 0.1 mm voxel size) to permit further processing and segmentation.

Datasets for both specimens were imported into Avizo 8 (Visualization Science Group) for the segmentation of endocranial structures. Due to poor grayscale attenuation (in particular for SMNS 16980), segmentation was performed manually using the paintbrush and interpolation tools in the Avizo segmentation editor (both reconstructions performed by the first author for consistency following Balanoff et al., 2016). 3D surface models and volumes were created to visualize the endocranial components. In addition, surface models of the individual structures were downsampled to a degree that allowed for small file sizes but preserved all details, and were exported as separate OBJ-files for the creation of the interactive 3D-figures provided in the Supplementary Material as outlined in Lautenschlager (2014b) using Adobe 3D reviewer (Adobe Systems Inc.).

To provide a basis for comparisons, the endocranial anatomy of nine extant turtles and of one additional stem-turtle, *Naomichelys speciosa* (FMNH PR273), was reconstructed in the manner described above. FMNH PR273 was scanned at the Institut für Naturwissenschaftliche Archäologie at the Universität Tübingen at a resolution of 0.1 mm resulting in an image stack of 1068 \times 1382 \times 622 pixels. The following extant species were scanned at the Steinmann-Institut für Geologie, Mineralogie & Paläontologie/Rheinische Friedrich-Wilhelms-Universität Bonn/Germany and at the Museum für Naturkunde Berlin/Germany: *Podocnemis unifilis* (SMF 55470), *Chelodina reimanni* (ZMB Herpetologie 49659), *Emydura subglobosa* (PIMUZ lab# 2009.37), *Pelodiscus sinensis* (IW576-2),

Chelonia mydas (ZMB 37416MS), *Macrochelys temminckii* (TCGT, Teaching collection Geowissenschaften Towisse), *Emys orbicularis* (WGJ, 1987a), *Platysternon megacephalum* (SMF 69684), *Malacochersus tornieri* (SMF 58702) (see Supplemental Material for Collection abbreviations). Data derived from the reconstructions were further used for a shape analysis of brain morphology.

Due to the absence of unambiguous and consistently identifiable landmarks on the endocast across different amniote taxa, outline shape analysis was performed to quantify morphological differences. Although this approach uses only two-dimensional outlines (in contrast to three-dimensional landmarks), it allows quantification of shape data for geometries lacking homologous landmarks (Haines and Crampton, 2000). For shape analysis, a sagittal cross-section through the surface model of each brain (i.e., digital cast of the endocranial cavity) was produced in Avizo for each reconstruction. Contours of the two-dimensional cross-sections were imported into tpsDig2.16 (Rohlf, 2010), digitized and saved as 1000 x/y-coordinate pairs. All outline data were subsequently analyzed in PAST 3.17 (Hammer et al., 2001) using fast Fourier transformation (FFT) and principal components analysis (PCA) with the hangle module as outlined in Crampton and Haines (1996) and Lautenschlager (2014a). Outlines were smoothed ten times to eliminate pixel noise, and 23 Fourier harmonics were found to describe the outlines of all sampled taxa sufficiently (average Fourier power > 99%) (see also Supplementary Material). In addition to the reconstructed endocasts, further outlines of 52 taxa were collected from the literature (Hopson, 1979; Franzosa, 2004; Neenan and Scheyer, 2012; Bona and Paulina-Carabajal, 2013; Carabajal et al., 2013; George and Holliday, 2013; Herrera et al., 2013; Holloway et al., 2013; Lautenschlager and Butler, 2016; von Baczko and Desojo, 2016; Jirak and Janacek, 2017; Laaß et al., 2017; Paulina-Carabajal et al., 2017; Pierce et al., 2017; DigiMorph¹) for different turtle, archosauromorph, lepidosauromorph and other amniote taxa (for list of taxa see Table S1). These outlines were redrawn in Adobe Illustrator to ensure sufficient resolution for the digitization process. For PCA, each taxon was assigned to a phylogenetic and an ecological (marine, freshwater, terrestrial, fossorial) group. To test for significant differences between those groups, we also conducted a non-parametric MANOVA test (Anderson, 2001) using PC scores representing 95% of total variance transformed into an Euclidean distance matrix, replicated with 10,000 permutations and compared using Bonferroni correction for the *post-hoc* analyses.

RESULTS

Endocranial Anatomy

The reconstruction of MB 1910.45.2 (Figures 1A–D) provided most details of the endocranial anatomy, but exhibited some moderate medio-lateral deformation. In comparison, the reconstruction of SMNS 16980 (Figures 1E–H) showed no

obvious artifacts, but the poor grayscale contrast permitted only a few structures (i.e., brain, pituitary fossa and some cranial nerves) to be visualized. In combination, both specimens allowed for a detailed reconstruction of most endocranial components.

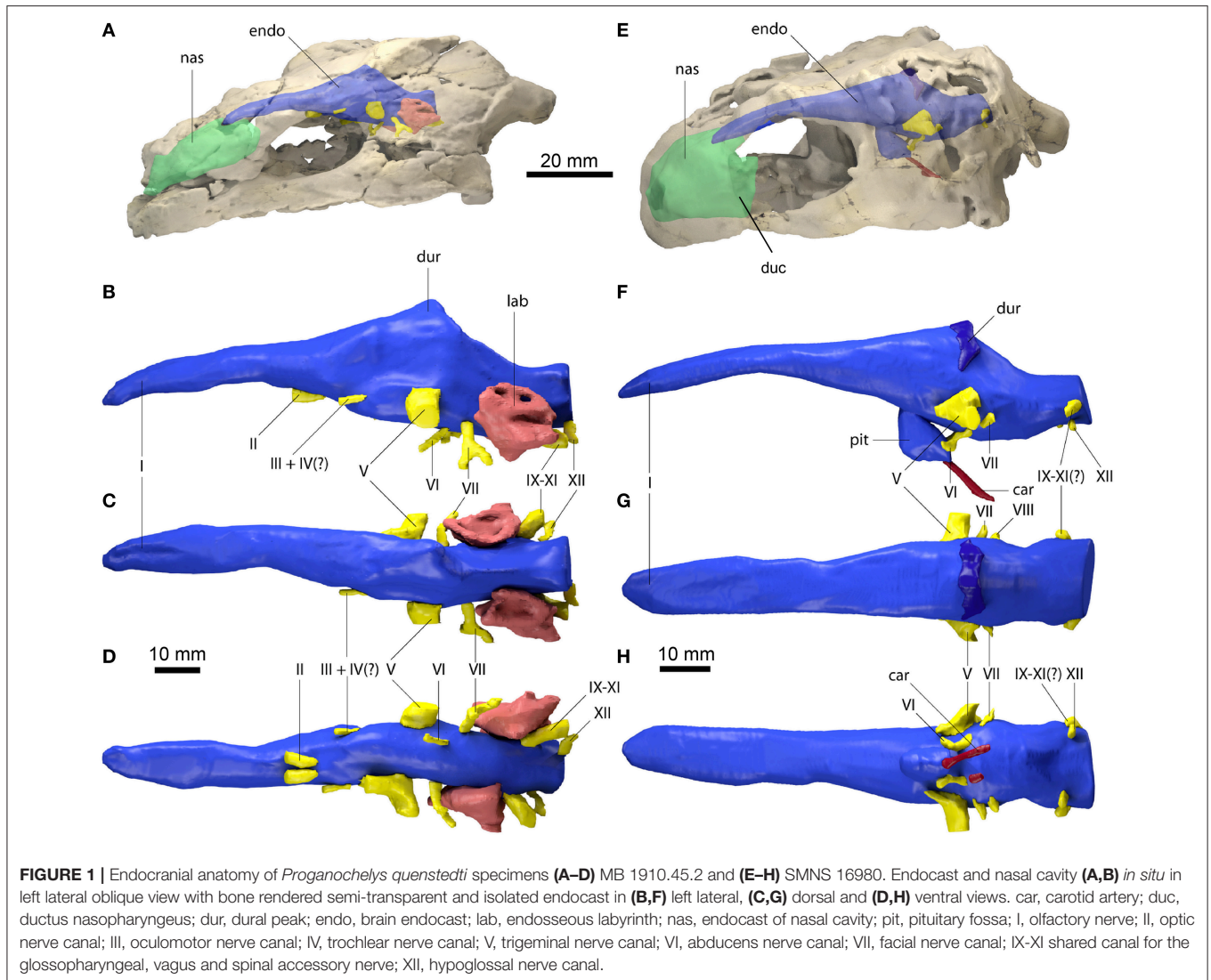
The brain endocast is anteroposteriorly elongate and straight in both specimens, with only moderate cephalic and pontine flexures (Figures 1B,F). The endocasts are tubular and mediolaterally narrow without prominent expansion or constriction of the fore-, mid- or hindbrain regions. The close similarity of these features in both specimens confirms that this morphology is natural and unlikely to be a result of taphonomic deformation. The olfactory nerve (CN I) contributes approximately a third to half of the full endocast's length, but a clear distinction between the base of the olfactory nerve and the cerebral hemispheres is not visible. The olfactory bulbs are only weakly reproduced by the ventral surfaces of the nasals. Cerebral hemispheres or distinct optic lobes are not visible in either specimen, suggesting that both structures were very small and/or that the venous sinus and the dura mater obscured the underlying morphology.

The midbrain region is confluent with the forebrain and only weakly demarcated. The only distinguishing feature is a dorsal expansion extending above the level of the olfactory nerve. This dural peak or cartilaginous rider (Zangerl, 1960; Gaffney and Zangerl, 1968; Paulina-Carabajal et al., 2017) is more prominently developed in MB 1910.45.2 (Figure 1B). In SMNS 16980, the dorsal expansion is shallower and somewhat separated from the main body of the midbrain by a bony margin, suggesting that this structure corresponds to the cartilaginous portion of the supraoccipital, which ends abruptly anteriorly in *Proganochelys quenstedti* (Gaffney, 1990). The pituitary fossa is visible in SMNS 16980 and forms a pendant pocket, projecting ventrally from the main body of the midbrain endocast.

The hindbrain region is anteroposteriorly short and not constricted mediolaterally between the endosseous labyrinths. Floccular lobes are not visible. Posteriorly, the hindbrain exits the braincase through the foramen magnum, which is oval and wider than high in SMNS 16980 and slightly higher than wide in MB 1910.45.2. The latter may be the result of the mediolateral compression of this specimen.

The nasal cavity is very enlarged when compared to the other sampled taxa (Figures 2, 3; Table 1). The strong lateral compression of MB 1910.45.2 may be responsible for the seemingly increased volume, and, hence, we consider the reconstruction of the nasal cavity in SMNS 16980 more reliable. Usually, three portions of the nasal cavity can be identified in turtles and other reptilians (Parsons, 1959, 1970; Halpern, 1992; Paulina-Carabajal et al., 2017): the vestibulum nasi, which connects the nasal chamber to the external nares; the ductus nasopharyngeus, connecting the nasal chamber to the choanae; and the cavum nasi proprium, the chamber itself, bounded anteriorly by the vestibulum, posteroventrally by the ductus, and posterodorsally by the olfactory nerve (CN I). The ductus nasopharyngeus can be distinguished from the rest of the nasal cavity in *P. quenstedti* as two ventrolateral projections (Figure 1). A proper duct (at least not bounded by bone) was not expected, since the choanae in *P. quenstedti* are very extensive, and

¹DigiMorph. *Digital Morphology - A Unique Biological Visualization Library*. Available online at: <http://www.digimorph.org/> (Accessed Aug 15, 2017)

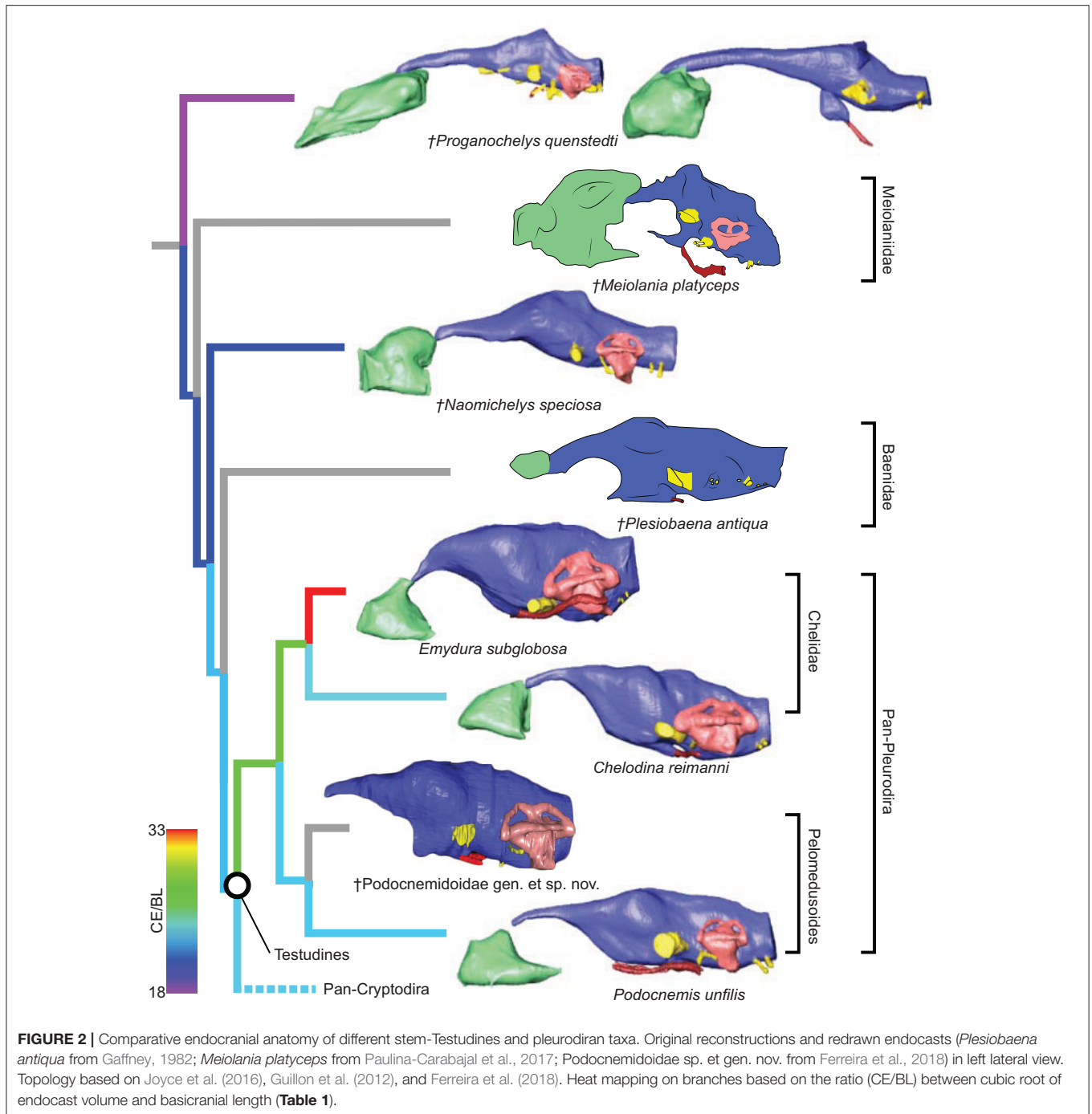


occupy almost the whole ventral surface of the nasal cavity. The vestibulum on the other hand is short, as in most other turtles (Paulina-Carabajal et al., 2017), connected to the large cavum nasi proprium, which constitutes most of the nasal cavity. The cavity as a whole is considerably broad and also high in comparison (Figures 2, 3) to several other taxa (Carabajal et al., 2013; Paulina-Carabajal et al., 2017).

The endosseous labyrinth is reconstructed only for MB 1910.45.2, as the grayscale attenuation did not allow differentiation of the bony housing in SMNS 16980. It is dorsoventrally compressed and compact. The anterior and posterior semicircular canals are small and anteroposteriorly longer than high and have low internal radii. The crus communis is also very low in comparison to other taxa (Carabajal et al., 2013; Mautner et al., 2017; Paulina-Carabajal et al., 2017; Ferreira et al., 2018) which results in an almost horizontal orientation of the anterior and posterior semicircular canals (Figure 4). The lateral semicircular canal barely extends laterally from the

vestibulum. The cochlear duct is expanded ventrally, but short. The canal of the fenestra ovalis is clearly visible projecting anterolaterally from the vestibulum.

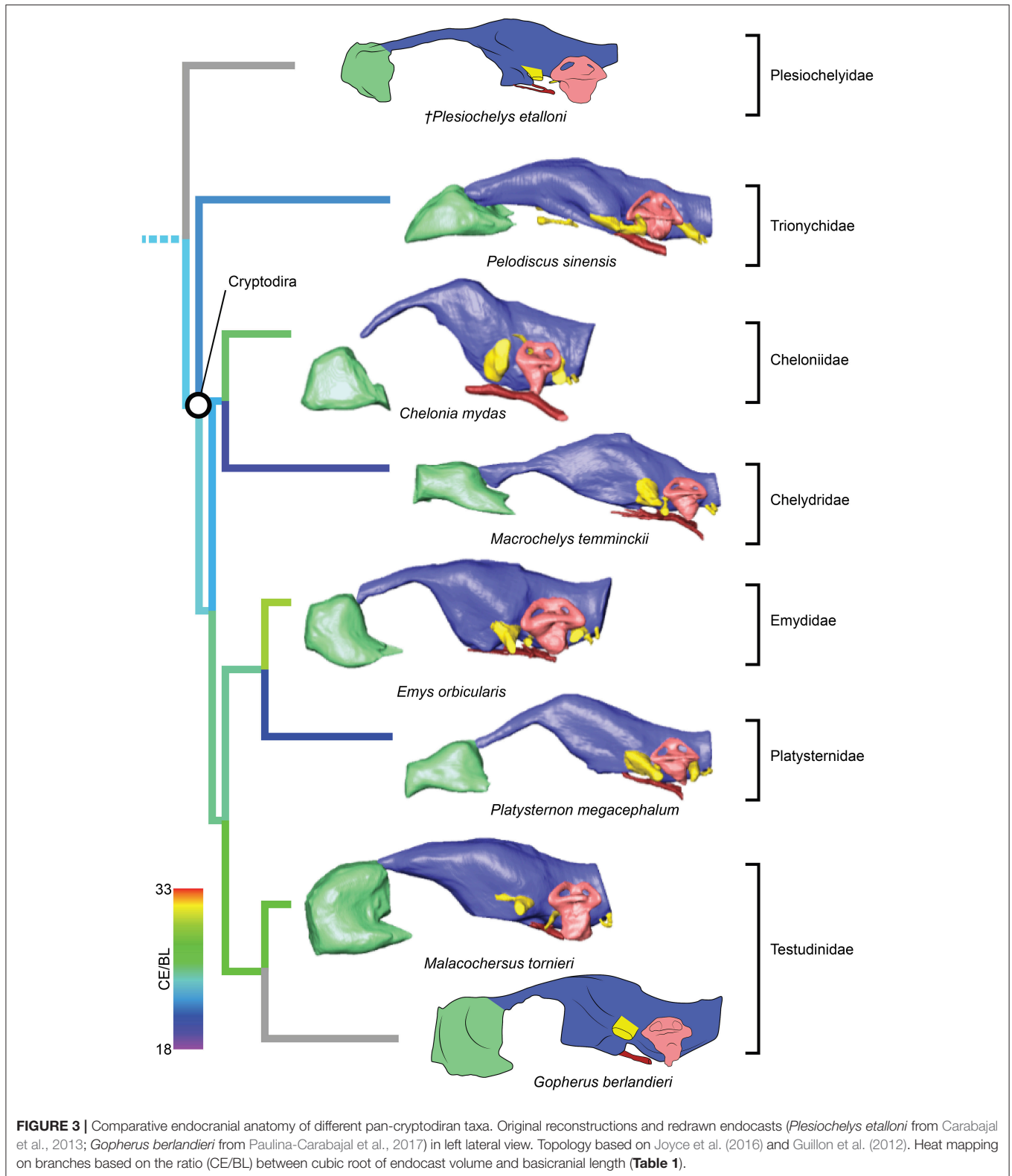
The proximal portion of the majority of cranial nerves could be reconstructed for MB 1910.45.2 (Figure 1), whereas only some of the larger nerve canals are visible in SMNS 16980. The optic nerves (CN II) exit the braincase through two large (3 mm in diameter each) foramina anteriorly and ventrally from the cerebral region of the endocranium in MB 1910.45.2. Posterior and lateral to CN II, the oculomotor (CN III) and possibly the trochlear nerve (CN IV) (Gaffney, 1990) originate ventrolaterally. In SMNS 16980, CN II–IV could not be reconstructed. The foramina through which those three cranial nerves (II–IV) exit the braincase are formed by the laterosphenoid (=“pleurosphenoid”) (Gaffney, 1990; Bhullar and Bever, 2009). This is the second *P. quenstedti* specimen with a preserved laterosphenoid, however the fact that this ossification is severely crushed leads us to refrain from commenting



further on its morphology. The trigeminal nerve (CN V) is large (ca. 6 mm in diameter) and exits the braincase laterally in both specimens through the prootic foramen. Based on both specimens (as well as other specimens described by Gaffney, 1990) we confirm that this foramen is surrounded exclusively by the prootic bone, contrary to Bhullar and Bever's (2009) interpretation that the laterosphenoid would form its anterior margin. A separation of the ophthalmic branch (CN V₁) is apparent on the right side in MB 1910.45.2, but this

could be a result of the high degree of distortion of this specimen.

The abducens nerve (CN VI), clearly visible in both specimens, originates from the ventral surface of the endocast. It pierces the basisphenoid through the foramen nervi abducens and enters laterally the pituitary fossa, which is bottomed by the sella turcica (Gaffney, 1990). Posterior to CN V, the facial nerve (CN VII) exits the braincase laterally through the prootic. In MB 1910.45.2, a distal branching of CN VII outside the braincase



wall is visible, also on the prootic bone. The vestibulocochlear nerves (CN VIII) could not be reconstructed in either specimen. The foramina for the CN VIII branches are usually very small

and may lie on cartilaginous structures (Gaffney, 1979), so they are not expected to leave unambiguous traces on fossilized skulls. The glossopharyngeal (CN IX), vagus (CN X) and

TABLE 1 | Measurements and ratios for sampled taxa.

Taxon	Specimen ID	SL [mm]	BL [mm]	EV [mm ³]	NV [mm ³]	N/E	CE/SL	CE/BL	OR [%]	ASC-PSC
<i>Proganochelys</i>	MB 1910.45.2	175	148.75	8,170.84	12,209.34	1.49	11.51	13.54	62.5	107°
<i>Proganochelys</i>	SMNS 16980	97	85.36	3,790.56	3,709.39	0.98	16.07	18.27	57.14	–
<i>Naomichelys</i>	FMNH PR273	117	103.50	9,805.97	4,077.77	0.42	18.29	20.68	15–19	79°
<i>Podocnemis</i>	SMF 55470	67	51.32	1,732.45	531.57	0.31	17.93	23.40	13.39	81°
<i>Chelodina</i>	ZMB H 49659	36	36.00	760.10	140.84	0.18	25.35	25.35	11.34	98°
<i>Chelonia</i>	ZMB 37416 MS	112	80.64	7,077.93	2,667.23	0.38	17.14	23.81	31.65	94°
<i>Macrochelys</i>	GPIT/RE/10801	120	105.88	9,583.53	3,568.33	0.37	17.70	20.06	38.18	88°
<i>Platysternon</i>	SMF 69684	60	46.43	898.13	314.33	0.35	16.08	20.78	28.23	82°
<i>Malacochersus</i>	SMF 58702	35	40.92	1,364.13	669.45	0.49	31.69	27.11	16.06	86°
<i>Emys</i>	WGJ 1987a	31	30.42	668.07	118.54	0.18	28.20	28.74	17.31	102°
<i>Emydura</i>	PIMUZ 2009.37	35	35.00	1,556.29	160.65	0.10	33.11	33.11	9.80	90°
<i>Pelodiscus</i>	IW576-2	59	39.48	707.23	444.09	0.63	15.10	22.57	20.25	79°

ASC-PSC, angle between anterior and posterior semicircular canals; BL, basicranial length; CE/BL, cubic root of endocast volume/basicranial length; CE/SL, cubic root of endocast volume/skull length; EV, endocast volume; NV, nasal cavity volume; N/E, nasal cavity/endocast volume ratio; OR, olfactory ratio; SL, skull length.

accessory nerves (CN XI) originate immediately posterior to the endosseous labyrinth and exit the braincase through the anterior jugular foramen in MB 1910.45.2. Although the sutures are not very clear, this foramen is thought to be formed by the exoccipital, basioccipital and opisthotic in *P. quenstedti* (Gaffney, 1990). In SMNS 16980, a large nerve canal originates in a more dorsolateral position (Figure 1). Due to the low resolution, it is unclear whether this canal represents the anterior jugular foramen or parts of the longitudinal sinus, though the latter is more likely. The hypoglossal nerve (CN XII) is transmitted through a single foramen on each side of the basioccipital (posterior to the jugular foramen) in both specimens.

Endocast Outline Analysis

The morphology of the endocast of *Proganochelys quenstedti* was compared to different turtles and other amniote taxa using shape analysis. The PCA results show that the first three PCs account for 71.7% (Table 2) of the brain endocast outline shape variation (Figures 5, 6). In no PC plot, there is a clear separation between either the phylogenetic or the ecological groups considered. However, the PERMANOVA tests support that Lepidosauromorpha differs significantly from Archosauromorpha ($p = 0.0006$) and from Testudinata ($p = 0.003$) although these tests find no significant differences between the ecological groups (Table 3). The outgroup *Diadectes* is recovered consistently in a position inside the morphospace occupied by other groups, whereas *P. quenstedti* is displaced from the occupied area in all plots; however, on the PC1 axis, *Kawingasaurus* is even more displaced in the positive direction (Figures 5, 6). *P. quenstedti* is distant from other turtles and the minimum spanning-tree (see Supplementary Material) places it closer to the lepidosauromorphs *Placodus* and *Chalarodon*, and to the archosauromorph *Pseudopalatus*, on the PC1/PC2, PC1/PC3, and PC2/PC3 plots, respectively. With regard to the ecological morphospaces, *P. quenstedti* is similarly found in a position outside all the groups, except on the PC1/PC3 plot, on

which it is inside the fossorial morphospace and very close to the terrestrial one (Figure 6).

DISCUSSION

Ancestral Condition for Testudinata

Even though more taxa have been assigned to the turtle stem-lineage recently (Li et al., 2008; Lyson et al., 2010; Schoch and Sues, 2015), *Proganochelys quenstedti* remains one of the most important stem-turtles, given its phylogenetic position as the earliest shelled turtle with a completely preserved skull (Parsons, 1959, 1970; Halpern, 1992; Joyce et al., 2016). Its endocast is a relatively simple structure when compared to that of crown-turtles (Carabajal et al., 2013; Mautner et al., 2017; Paulina-Carabajal et al., 2017; Ferreira et al., 2018). It has a tube-like shape, with only small pontine and cephalic flexures and poorly differentiated brain regions. As in other amniotes, the portion between the fore- and midbrain is the most voluminous, but this is achieved exclusively by an increase in height, since the endocast is nearly constant in width over its entire length (Figure 1). Another striking feature is the pendant pituitary fossa, which is very common in archosaurs (Witmer et al., 2008; Lautenschlager and Butler, 2016; Araújo et al., 2017; Pierce et al., 2017), but does not occur in extant turtles, in which the dorsum sellae and the sella turcica are aligned, positioning the pituitary fossa approximately at the same level as the posterior portions of the endocast (Figures 2, 3). Although the pituitary fossa of turtles can also house other smaller structures (e.g., internal carotid and abducens nerve) the size of the pituitary gland should be at least partially responsible for the larger size of the fossa in *P. quenstedti*. A similar condition was found for sauropod and theropod dinosaurs (Witmer et al., 2008), in which enlarged pituitary glands have been linked to larger body sizes (Edinger, 1942). While *P. quenstedti* reached a carapace length of at least 67 cm (based on MB 1910.45.2) (Gaffney, 1990), it was not one of the largest turtles, being much smaller than some extant turtles (e.g., up to 150–200 cm in *Chelonia mydas* and

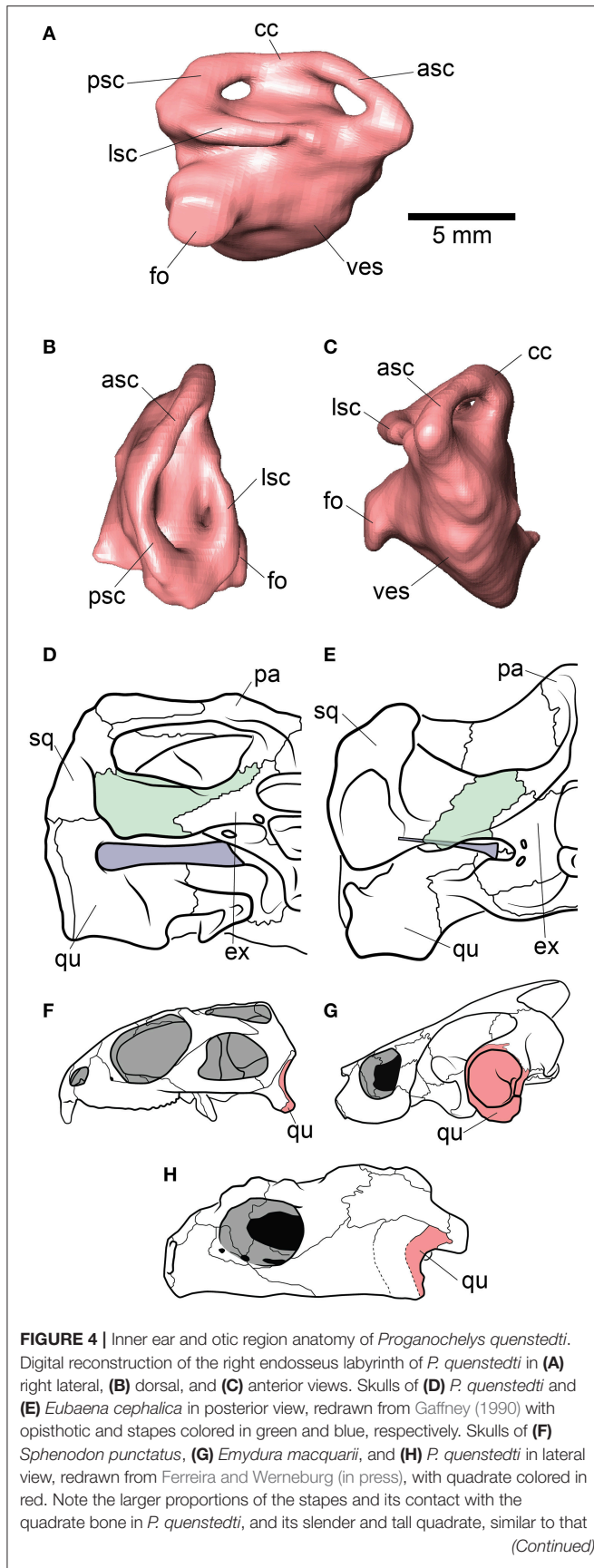


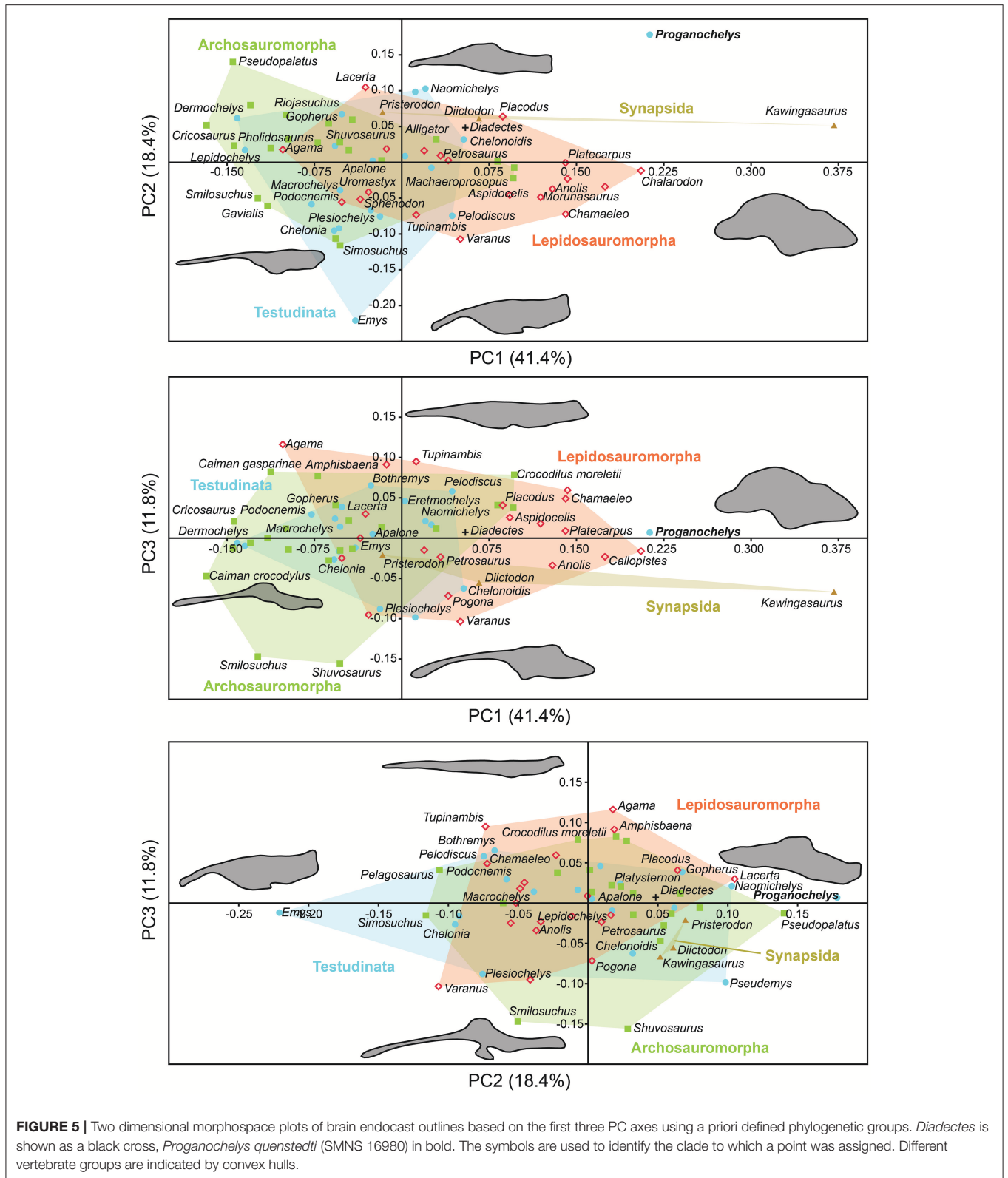
FIGURE 4 | of *S. punctatus*, and distinct from the round one of other turtles, which completely encloses the tympanic membrane. asc, anterior semicircular canal; cc, crus communis; ex, exoccipital; fo, fenestra ovalis; lsc, lateral semicircular canal; pa, parietal; psc, posterior semicircular canal; qu, quadrate; sq, squamosal; ves, vestibulum.

TABLE 2 | Summary of the results of the principal component analyses of the brain outlines of different specimens of turtles and other groups.

PC	Eigenvalue	% variance	% cumulative variance
1	0.01042	41.4	41.4
2	0.00464	18.5	59.9
3	0.00298	11.8	71.7
4	0.00174	6.9	78.7
5	0.00137	5.5	84.1
6	0.00093	3.7	87.8
7	0.00067	2.6	90.5
8	0.00042	1.7	92.1
9	0.00032	1.3	93.4
10	0.00028	1.1	94.5
11	0.00021	0.8	95.4

Pelochelys cantorii (Angielczyk et al., 2015) and meiolanids (Gaffney, 1996). Turtles included in our sample that are comparable in size to *P. quenstedti*, such as *Podocnemis unifilis* and *Macrochelys temminckii* (up to 68 and 66 cm of carapace length) (Angielczyk et al., 2015), and also *Chelonia mydas*, do not show a pendant pituitary fossa (Figures 2, 3). An alternative explanation is that it is not the pituitary fossa that is larger in *P. quenstedti*, but rather the brain that was comparatively smaller. Indeed, in our sample, this taxon has the lowest value for the ratio endocast volume/basicranial length (Table 1), supporting the hypothesis that the brain increased in size during turtle evolution.

The brain endocast in turtles does not seem to be consistent with general skull anatomy. Taxa with higher/lower and wider/thinner endocasts do not possess similar skull proportions, which seem more related to the size and shape of the adductor chamber and the associated supraoccipital and squamosal crests (Figure 7). Proportional changes observed in the adductor chamber throughout the turtle lineage rather reflect the distinct volume and size of the external jaw adductor musculature in different taxa (Claude et al., 2004; Foth and Joyce, 2016; Foth et al., 2017; Ferreira and Werneburg, in press). Also, the position of the exits of the cranial nerves change only slightly, even with profound changes in the arrangement of related structures such as the eyes and muscles. For example, in *P. quenstedti* the external jaw adductor musculature innervated by the trigeminal nerve (CN V₃) is vertically oriented and entirely positioned anteriorly to the quadrate (Ferreira and Werneburg, in press), while in crown-turtles it extends far posteriorly, following the enlargement of the supraoccipital and squamosal crests (Poglayen-Neuwall, 1953; Werneburg, 2011, 2013). However, the relative position of the exit of CN V remains roughly the same through turtle evolution



when compared to the remainder of the endocast and the surrounding bones (Figure 7). Hence, the actual change that occurs when the muscles expand posteriorly involves only growth

and reorientation of distal V₃-branches and not a repositioning of the trigeminal nerve foramen (Poglayen-Neuwall, 1953; Schumacher, 1973).

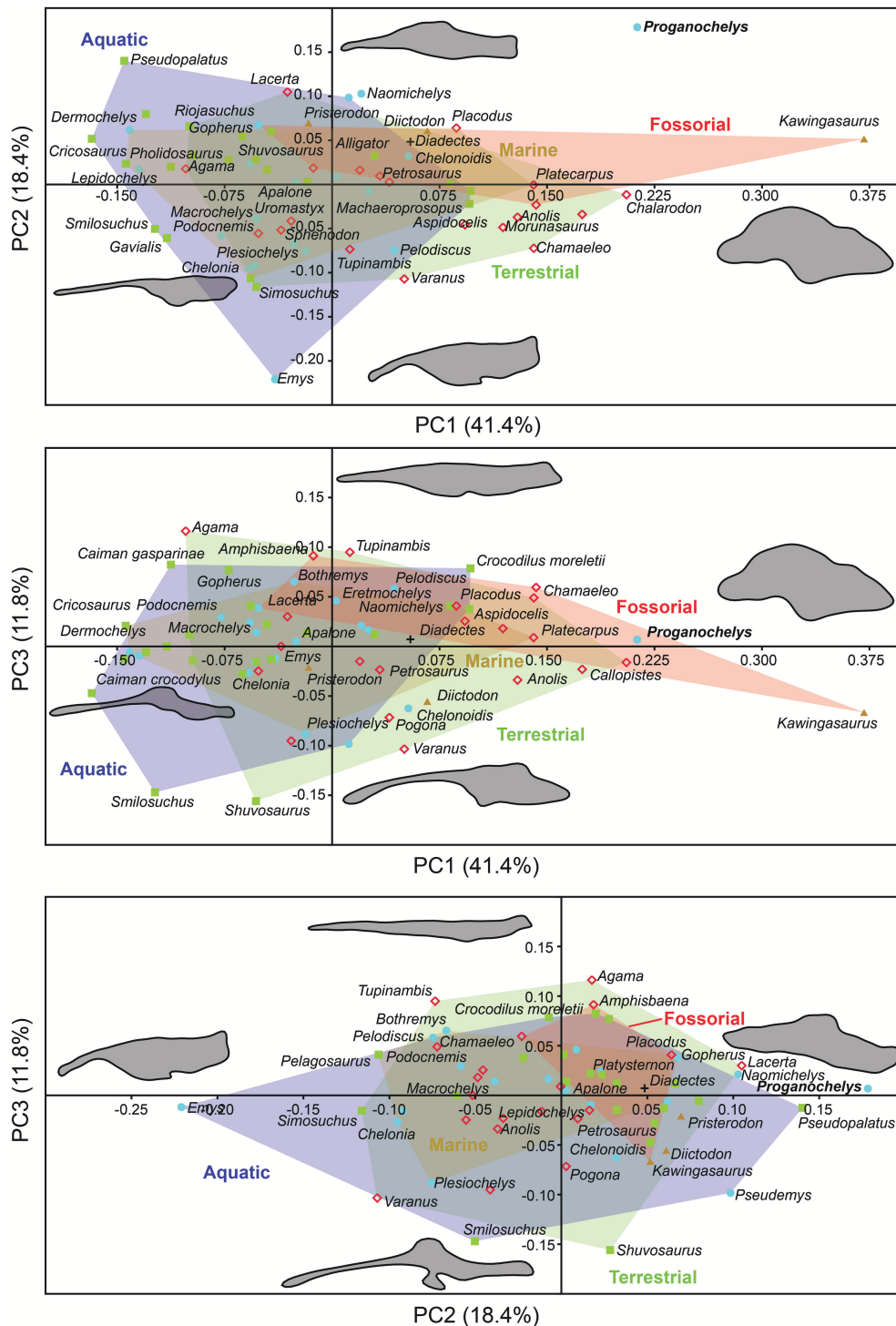


FIGURE 6 | Two dimensional morphospace plots of brain endocast outlines based on the first three PC axes using a priori defined ecological groups. *Diadectes* is shown as a black cross, *Proganochelys quenstedti* (SMNS 16980) in bold. The symbols are used to identify the clade to which a point was assigned. Different ecological groups are indicated by convex hulls.

Sensory Capabilities of *Proganochelys*

The endosseous labyrinth of *Proganochelys quenstedti* is slightly distinct from that of crown-turtles in being more compact

and robust, with short and thick semicircular canals and a low crus communis resulting in almost horizontally oriented canals (Figure 4). The anterior and posterior semicircular

TABLE 3 | Results of one-way PERMANOVA test (10000 permutations) with 95% of variance (PC1-PC11), excluding *Proganochelys* and *Diadectes* (for phylogenetic groups only).

PHYLOGENETIC GROUPS			
Permutation N	10000		
Total sum of squares (SQ)	1,901		
Within-group SQ	1,599		
F	3,653		
p	1.00E-01		
	Testudinata	Archosauromorpha	Lepidosauromorpha
Testudinata			
Archosauromorpha	0.8423		
Lepidosauromorpha	0.003	0.0005999	
Synapsida	0.327	0.1056	0.4488
ECOLOGICAL GROUPS			
Permutation N	10000		
Total SQ	77		
Within-group SQ	72		
F	1		
P	0.1656		
	Terrestrial	Aquatic	Marine
Terrestrial			
Aquatic	1		
Marine	0.252	1	
Fossorial	0.6485	1	0.8387

canals (ASC and PSC, respectively) are nearly at the same level as the lateral semicircular canal (LSC), whereas in other turtles the first two run dorsally in relation to the last (Carabajal et al., 2013; Mautner et al., 2017; Paulina-Carabajal et al., 2017; Ferreira et al., 2018). The angle between the ASC and LSC is also very wide (Table 1), with similar values to meiolaniids and tortoises (Paulina-Carabajal et al., 2017). This combination of features suggests that the semicircular canals of *P. quenstedti* were not very sensitive during movements within the sagittal (head moving up and down) and coronal planes (head tilt) (Brichta et al., 1988; Spoor et al., 2007; David et al., 2010). Instead, the LSC was likely more effective in stabilizing gaze during yaw movements (head moving left and right). Thus, the labyrinth anatomy of *P. quenstedti* indicates this species was slow and non-agile (Spoor et al., 2007; David et al., 2010), compatible with a highly terrestrial and possibly fossorial lifestyle. This is also tentatively indicated by its position in morphospace outside of, but close to, terrestrial and fossorial groupings in the shape analysis (Figure 6).

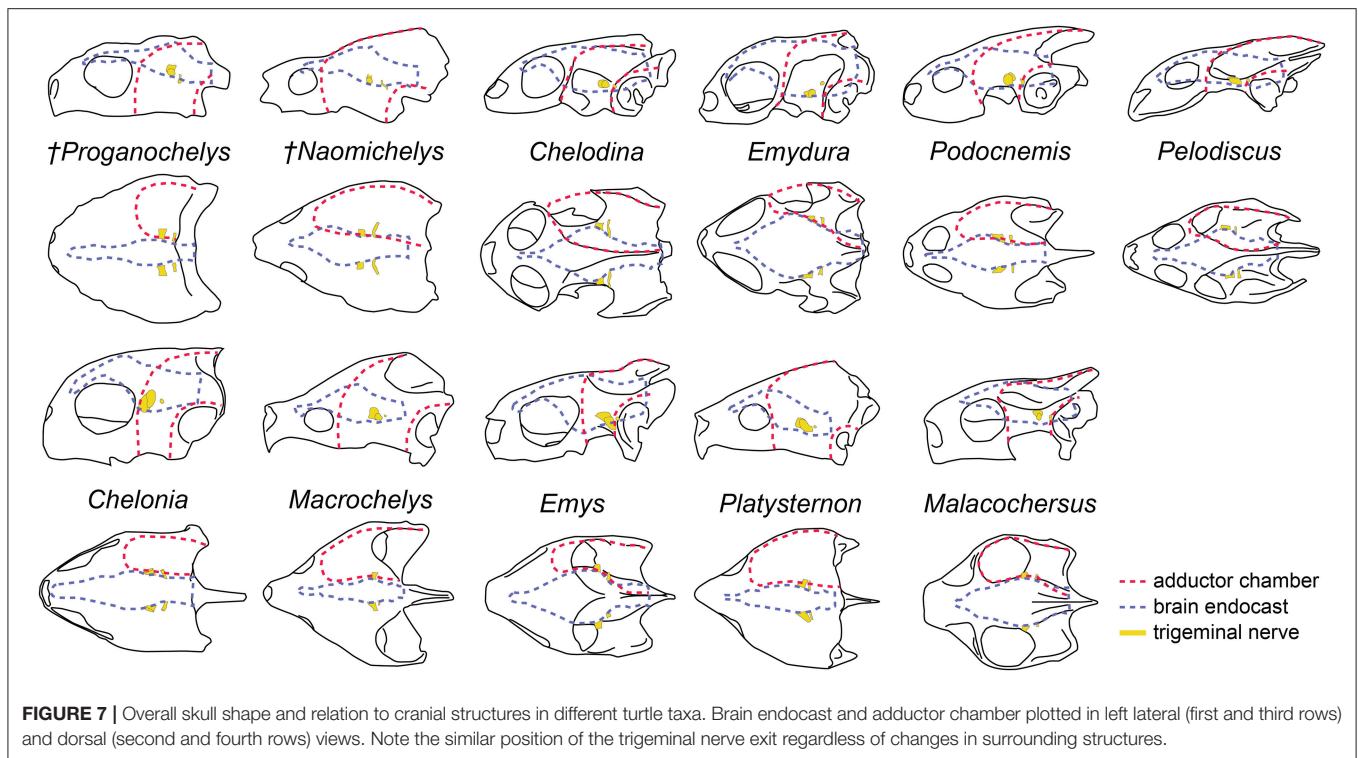
Although the cervical vertebrae of *P. quenstedti* were capable of a certain level of mobility (Werneburg et al., 2015a), its short neck coupled with the relatively low carapace, strong osteoderms on the dorsal neck surface and cervical ribs (Gaffney, 1990) imply restricted mobility along the same planes (sagittal and coronal) (Werneburg et al., 2015a,b) as indicated by its labyrinth morphology. Crown-turtles, however, evolved longer necks and

several taxa are capable of complex and, sometimes, very fast neck and head movements (Poglayen-Neuwall, 1953; Herrel et al., 2008; Werneburg et al., 2015a,b). This could be related to the apparent increase in size of the semicircular canals in crown-turtles (Spoor, 2003; Spoor et al., 2007) when compared to those of *P. quenstedti* (although when compared to more agile reptiles, all turtles possess short canals; Witmer et al., 2008).

Hearing was likely not well-developed in *P. quenstedti*, given the small overall size of the endosseous cochlear duct (Walsh et al., 2009) in comparison to other turtles. Even though its quadrate does not form the characteristic lateral round structure that encloses the cavum tympani in crown-turtles (Figures 4F–H), it possibly had a tympanic ear similar to those of extant squamates and cheloniids, in which the tympanum is supported by both bone and connective tissue (Henson, 1974; Gaffney, 1990). However, the stapes of *P. quenstedti* was much stouter than that of crown-turtles (Figures 4D,E), and possibly articulated with the quadrate (Gaffney, 1990), suggesting that it was not as effective as the thin vibratory element characteristic of extant amniotes with tympanic hearing, including modern turtles (Baird, 1970; Clack, 1997). As proposed by Clack (1997) for diapsids, elongation of the paraoccipital process of the opisthotic and its tight suturing to the squamosal, which occurred in the group including all testudines but *Proganochelys quenstedti* (Sterli et al., 2010), may have completely released the stapes from its ancestral structural function (connecting the quadrate to other elements of the braincase) during turtle evolution.

The nasal cavity of *P. quenstedti* represents at least 42.2% of the total endocast volume (Table 1), fitting in the volume spectrum of terrestrial turtle taxa, which ranges from 29 to 43% in tortoises and 58.5 to 64% in meiolaniids (Carabajal et al., 2013). Larger nasal cavities have been related to occupation of arid environments, thermoregulation, sound-production or higher olfactory capabilities (Parsons, 1959, 1970; Paulina-Carabajal et al., 2017). In *P. quenstedti*, the cavum nasi proprium represents most of the volume of the nasal cavity and extends far dorsally and posteriorly. Within the nasal cavity, sensory epithelium occurs only on the cavum walls (Parsons, 1970), and, as such, the cavum's relative size could be used as a proxy for inferences about olfactory capability in extinct reptiles. This connection, however, should be interpreted cautiously, due to the possible relation between cavum size and other functions, such as thermoregulation or vocalization (Bourke et al., 2014; Paulina-Carabajal et al., 2017).

The size and volume of the olfactory bulbs have been shown to be related to a greater reliance on the olfactory sense in mammals and birds (Bang, 1971; Bang and Wenzel, 1985; Healy and Guilford, 1990; Gittleman, 1991). In a series of studies the olfactory ratio (ratio between olfactory bulb and cerebral hemisphere maximum diameters; OR values) were used as a proxy to study olfactory acuity and capacity in theropod dinosaurs (including birds) and crocodylians (Zelenitsky et al., 2009, 2011). More recently this has also been applied to turtles (Paulina-Carabajal et al., 2017), showing that tortoises and meiolaniids (both terrestrial taxa) have the highest OR values (36–62 and 20–45%, respectively). Even though OR may not be an exact measure of olfactory acuity it is currently the



best available proxy and its use for a variety of reptilian taxa (Zelenitsky et al., 2009, 2011; Paulina-Carabajal et al., 2017) makes it a useful comparative metric. Here, we show that the OR is even higher in *P. quenstedti*, between 57 and 62% (Table 1), but in this case, these values may be also related to the less-developed cerebral hemispheres rather than to larger olfactory bulbs. Nevertheless, the large nasal cavity in association with the high OR values supports our hypothesis that olfaction was possibly the most developed sense in *P. quenstedti*.

Evolution of the Turtle Brain Endocast

In the shape analysis, *Proganochelys quenstedti* is not contained in the morphospace occupied by any of the considered phylogenetic groups (Figure 5). There is extensive overlap in the PCA plots, but, at the same time, the PERMANOVA test shows a separation between Lepidosauromorpha, Testudinata and Archosauromorpha (Table 3). These results suggest that all amniotes (excluding dinosaurs and mammals) share a similar plesiomorphic brain endocast morphology, but that those lineages evolved in different directions in the morphospace.

Comparing general ecological groups (freshwater, marine, terrestrial, and fossorial) provided similar results, with extensive overlap among the occupied morphospaces (Figure 6). *P. quenstedti* is contained in the morphospace occupied by the fossorial group on the PC1/PC3 plot, but it falls outside every group on the other plots. Additionally, the statistical tests do not support significant differences between any of the considered groups (Table 3). On the other hand, the minimum spanning trees (see Supplementary Material) show that even when inside the fossorial morphospace *P. quenstedti* is closest to *Placodus*,

a marine lepidosauromorph, and *Pseudopalatus*, an aquatic archosauromorph. A phylogenetic proximity to Sauropterygia (the lepidosauromorph lineage that includes *Placodus*) has been proposed previously (deBraga and Rieppel, 1997) and is associated with the hypothesis that turtles originated in marine environments (Joyce and Gauthier, 2004; Joyce, 2015). The proximity of *P. quenstedti* and *Placodus* in our PC1/PC2 plot (Figure 6) may recall this hypothesis, but the poor sampling of sauropterygians together with the extensive overlap between all groups (phylogenetic and ecological) cause us to refrain from considering this a robust interpretation.

The shape analysis presented here is the first attempt to explore the evolution of neuroanatomy in amniotes with a quantitative approach. Even though our results do not support inferences about lifestyles from neuroanatomical data, the significant separation between some of the considered phylogenetic groups (Figure 5, Table 3) seems promising. We can identify some caveats in our sample (e.g., few marine reptiles, synapsids and early amniotes) that can be easily overcome with the increasing use of computer tomography in paleontological and anatomical studies. Our approach using sagittal cross-section outlines could have also influenced the results, since there is a loss of information when the 3D endocast is simplified to a 2D outline.

More recently, Lyson et al. (2016) thoroughly analyzed the morphology of *Eunotosaurus africanus*, identifying some osteological correlates that led them to conclude that it was likely well-adapted for fossoriality. The authors also identified some of those correlates (e.g., large claws) in other proto- (e.g., *Odontochelys semitestacea*) and stem-turtles (*Proganochelys*

quenstedti and *Palaeochersis talampayensis*), concluding that “fossoriality played an important role in the early evolution of turtles” (Lyson et al., 2016). Although in the PC1/PC3 plot (Figure 6) *P. quenstedti* is contained in the fossorial morphospace, the minimum spanning tree (see Supplementary Material) shows it to be closest to the terrestrial non-fossorial taxon *Chalarodon* and the statistical analyses do not support any significant differences between the considered groups (Table 3). While the shape analyses do not shed light on this problem conclusively, other sources of data are more convincing. *Proganochelys quenstedti* fossils were found in continental deposits (Gaffney, 1990) and analyses of forelimb proportions (Joyce and Gauthier, 2004) and paleohistology (Scheyer and Sander, 2007) support it as a terrestrial turtle. The morphology of its endosseous labyrinth with short semicircular canals oriented at high angles to each other and the large *cavum nasi proprium* (Parsons, 1970; David et al., 2010; Paulina-Carabajal et al., 2017) agree with these previous studies, strongly supporting the interpretation that *P. quenstedti* was a well-adapted terrestrial turtle. However, since its vestibule is not particularly large, in contrast to the condition of truly fossorial taxa (Yi and Norell, 2015) or of the semi-fossorial tortoise *Gopherus* (Paulina-Carabajal et al., 2017), the present data suggest it was likely not a fossorial taxon. In *P. quenstedti*, the relatively enlarged vestibule in comparison to the other turtles in this study results from the relatively small semicircular canals. Thus, even if fossoriality had an important role during the early evolution of shell components (Lyson et al., 2016), our data suggests the complete turtle shell first appeared in a terrestrial taxon, with no evident link to fossoriality (at the Testudinata node).

If we assume that the relatively simple morphology of *P. quenstedti* closely resembles that of the testudinate ancestors, some trends can be inferred for the evolution of endocranial structures in turtles. An increase in overall encephalization, for example, with longer and more voluminous endocasts in relation to skull length is found already in the stem-turtle *Naomichelys speciosa* and continues in crown-turtles (Figures 2, 3). Some regions became more pronounced as well. In *N. speciosa*, meiolaniids (Paulina-Carabajal et al., 2017), *Plesiochelys etalloni* (Carabajal et al., 2013) and all other crown-turtles (Mautner et al., 2017; Ferreira et al., 2018) the cerebral hemispheres are clearly distinguishable from the remainder of the endocast and are wider in relation to skull and endocast length than in *P. quenstedti* (Figures 2, 3). The olfactory bulb can also be seen in the endocasts of some taxa, e.g., *Testudo graeca* and *Plesiochelys etalloni* (Carabajal et al., 2013; Paulina-Carabajal et al., 2017). However, this does not seem to be a general trend but rather one of the features that show noteworthy variations among crown-turtles, as are the degree of development of the cephalic and pontine inflexions and the sizes of the nasal cavity and the orbits. Considering that the brain of *P. quenstedti* was a simple tube-like structure with poorly differentiated regions, an increase in size and in regionalization of the brain took place later during the course of turtle evolution, similarly (although in a much lesser

degree) to the trend observed during bird evolution (Balanoff et al., 2013), and achieved an endocast diversity comparable to other groups of amniotes, such as lepidosaurs and archosaurs (excluding dinosaurs; Figures 5, 6). Indeed, extant turtles possess high brain weights in relation to body weight, comparable to that of crocodiles (Gürtürkün et al., 2016), but that was not the ancestral condition of the group based on our analyses. Given the recurrent results of phylogenetic analyses suggesting that turtles have parareptilian affinity (e.g., Laurin and Piñeiro, 2017), it is important to sample the endocast diversity in that clade and explore the similarities between turtles and all other reptilian lineages. The simpler brain structure together with the large nasal cavity and nearly horizontal and short semicircular canals of the inner ear supports a picture of *P. quenstedti* as a terrestrial but most likely not fossorial turtle, with likely mediocre hearing and vision, but a well-developed olfactory sense.

AUTHOR CONTRIBUTIONS

SL and IW: conceived and designed the study; SL: performed the three-dimensional reconstruction; IW: provided digital datasets; SL, GF, and IW: collected, analyzed and interpreted the data; SL and GF: created figures and supplementary data; SL, GF, and IW: contributed equally to the discussion, preparation and writing of the paper.

FUNDING

This research was funded by FAPESP (Fundação de Amparo à Pesquisa do Estado de São Paulo) grants 2016/03934-2 and 2014/2539-5 to GF and by SNF (Schweizerischer Nationalfonds zur Förderung der Wissenschaftlichen Forschung) advanced postdoc mobility grant P300PA_164720 to IW.

ACKNOWLEDGMENTS

We thank Daniela Schwarz for sharing the μ CT-scan of the Berlin specimen, and Rainer Schoch for the permission to μ CT the Stuttgart specimen of *Proganochelys*. We thank Walter G. Joyce, Bill Simpson and Virginie Volpato for access to the specimens and CT scan data of *Naomichelys* and *Emys*. Adrian Tröschler, Jan Prochel, Irina Ruf, and Kristin Mahlow are thanked for help with μ CT-scans of extant species. We thank Gabe S. Bever and Walter G. Joyce for useful comments on a previous version of the manuscript and the three reviewers that provided insightful comments and suggestions on the latest version of the manuscript.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fevo.2018.00007/full#supplementary-material>

REFERENCES

- Anderson, M. J. (2001). A new method for non-parametric multivariate analysis of variance. *Austral Ecol.* 26, 32–46. doi: 10.1111/j.1442-9993.2001.01070.pp.x
- Angielczyk, K. D., Burroughs, R. W., and Feldman, C. (2015). Do turtles follow the rules? Latitudinal gradients in species richness, body size, and geographic range area of the world's turtles. *J. Exp. Zool. B Mol. Dev. Evol.* 324, 270–294. doi: 10.1002/jez.b.22602
- Araújo, R., Fernandez, V., Polcyn, M. J., Fröbisch, J., and Martins, R. M. (2017). Aspects of gorgonopsian paleobiology and evolution: insights from the basicranium, occiput, osseous labyrinth, vasculature, and neuroanatomy. *Peer. J.* 5:e3119. doi: 10.7717/peerj.3119
- Baird, I. L. (1970). "The anatomy of the reptilian ear," in *Biology of the Reptilia*, eds C. Gans and T. S. Parsons (London: Academic Press), 193–275.
- Balanoff, A. M., Bever, G. S., Colbert, M. W., Clarke, J. A., Field, D. J., Gignac, P. M., et al. (2016). Best practices for digitally constructing endocranial casts: examples from birds and their dinosaurian relatives. *J. Anat.* 229, 173–190. doi: 10.1111/joa.12378
- Balanoff, A. M., Bever, G. S., Rowe, T. B., and Norell, M. A. (2013). Evolutionary origins of the avian brain. *Nature* 501, 93–96. doi: 10.1038/nature12424
- Bang, B. G. (1971). Functional anatomy of the olfactory system in 23 orders of birds. *Acta Anat.* 79, 1–76. doi: 10.1159/isbn.978-3-318-01866-0
- Bang, B. G., and Wenzel, B. M. (1985). "Nasal cavity and olfactory system," in *Form and function in birds*, eds A. S. King and J. McLelland (New York, NY: Academic Press), 195–225.
- Bever, G. S., Lyson, T. R., Field, D. J., and Bhullar, B.-A. S. (2015). Evolutionary origin of the turtle skull. *Nature* 525, 239–242. doi: 10.1038/nature14900
- Bhullar, B.-A. S., and Bever, G. S. (2009). An archosaur-like laterosphenoid in early turtles (*Reptilia: Pantestudines*). *Breviora* 518, 1–11. doi: 10.3099/0006-9698-518.1.1
- Bona, P., and Paulina-Carabajal, A. (2013). *Caiman gasparinae* sp. nov., a huge alligatorid (Caimaninae) from the late Miocene of Paraná, Argentina. *Alcheringa* 37, 1–12. doi: 10.1080/03115518.2013.785335
- Bourke, J. M., Ruger Porter, W. M., Ridgely, R. C., Lyson, T. R., Schachner, E. R., Bell, P. R., et al. (2014). Breathing life into dinosaurs: tackling challenges of soft-tissue restoration and nasal airflow in extinct species. *Anat. Rec.* 297, 2148–2186. doi: 10.1002/ar.23046
- Brichta, A., Acuña, D., and Peterson, E. (1988). Planar relations of semicircular canals in awake, resting turtles, *Pseudemys scripta*. *Brain Behav. Evol.* 32, 236–245. doi: 10.1159/000116551
- Carabajal, A. P., Sterli, J., Müller, J., and Hilger, A. (2013). Neuroanatomy of the marine Jurassic turtle *Plesiochelys etalloni* (Testudinata, Plesiochelyidae). *PLoS ONE* 8:e69264. doi: 10.1371/journal.pone.0069264
- Clack, J. (1997). The evolution of tetrapod ears and the fossil record. *Brain Behav. Evol.* 50, 198–212. doi: 10.1159/000113334
- Claude, J., Pritchard, P. C., Tong, H., Paradis, E., and Auffray, J.-C. (2004). Ecological correlates and evolutionary divergence in the skull of turtles: a geometric morphometric assessment. *Syst. Biol.* 53, 933–948. doi: 10.1080/10635150490889498
- Crampton, J. S., and Haines, A. J. (1996). *Users' Manual for Programs Hangle, Hmitch and Hcurve for the Fourier Shape Analysis of Two-Dimensional Outlines*, Vol. 96. Science Report, Institute of Geological and Nuclear Sciences.
- David, R., Droulez, J., Allain, R., Berthoz, A., Janvier, P., and Bennequin, D. (2010). Motion from the past. A new method to infer vestibular capacities of extinct species. *C. R. Palevol.* 9, 397–410. doi: 10.1016/j.crpv.2010.07.012
- Edinger, T. (1942). The pituitary body in giant animals fossil and living: a survey and a suggestion. *Q. Rev. Biol.* 17, 31–45. doi: 10.1086/394644
- Ferreira, G. S., Iori, F. V., Hermanson, G., and Langer, M. C. (2018). New turtle remains from the Late Cretaceous of Monte Alto-SP, Brazil, including cranial osteology, neuroanatomy and phylogenetic position of a new taxon. *Paläontol. Zeit.* doi: 10.1007/s12542-017-0397-x
- Ferreira, G. S., and Werneburg, I. (in press). "Evolution, diversity, and development of the craniocervical system in turtles with special reference to jaw musculature," in *Heads, Jaws and Muscles – Evolution, Development, Anatomical Diversity and Function*, eds J. Ziermann, R. R. Diaz Jr., and R. Diogo (Heidelberg: Springer).
- Field, D. J., Gauthier, J. A., King, B. L., Pisani, D., Lyson, T. R., and Peterson, K. J. (2014). Toward consilience in reptile phylogeny: miRNAs support an archosaur, not lepidosaur, affinity for turtles. *Evol. Dev.* 16, 189–196. doi: 10.1111/ede.12081
- Foth, C., and Joyce, W. G. (2016). Slow and steady: the evolution of cranial disparity in fossil and recent turtles. *P. Roy. Soc. Lond. B Bio.* 283m:20161881. doi: 10.1098/rspb.2016.1881
- Foth, C., Rabi, M., and Joyce, W. G. (2017). Skull shape variation in extant and extinct Testudinata and its relation to habitat and feeding ecology. *Acta Zool. Stockholm* 98, 310–325. doi: 10.1111/azo.12181
- Franzosa, J. W. (2004). *Evolution of the brain in Theropoda (Dinosauria)*. Ph.D. Dissertation, University of Texas, Austin, TX.
- Gaffney, E. S. (1979). Comparative cranial morphology of Recent and fossil turtles. *B. Am. Mus. Nat. Hist.* 164, 65–376.
- Gaffney, E. S. (1982). Cranial morphology of the baenid turtles. *Am. Mus. Novit.* 2737, 1–22.
- Gaffney, E. S. (1990). The comparative osteology of the Triassic turtle *Proganochelys*. *B. Am. Mus. Nat. Hist.* 194, 1–263.
- Gaffney, E. S. (1996). The postcranial morphology of *Meiolania platyceps* and a review of the Meiolaniidae. *B. Am. Mus. Nat. Hist.* 229, 1–166.
- Gaffney, E. S., and Zangerl, R. (1968). *A Revision of the Chelonian Genus Bothremys: (Pleurodira: Pelomedusidae)*. Chicago, IL: Field Museum of Natural History.
- Gauthier, J. A., Kluge, A. G., and Rowe, T. (1988). "The early evolution of the Amniota," in *The Phylogeny and Classification of the Tetrapods*, Vol. 1, ed M. J. Benton (Oxford: Oxford University Press) 103–155.
- George, I. D., and Holliday, C. M. (2013). Trigeminal nerve morphology in *Alligator mississippiensis* and its significance for crocodyliform facial sensation and evolution. *Anat. Rec.* 296, 670–680. doi: 10.1002/ar.22666
- Gittleman, J. L. (1991). Carnivore olfactory bulbs: allometry, phylogeny and ecology. *J. Zool.* 225, 253–272. doi: 10.1111/j.1469-7998.1991.tb03815.x
- Guillon, J.-M., Guéry, L., Hulin, V., and Girondot, M. (2012). A large phylogeny of turtles (Testudines) using molecular data. *Contrib. Zool.* 81, 147–158.
- Gürtürkün, O., Stacho, M., and Ströckens, F. (2016). "The brains of reptiles and birds," in *Evolution of Nervous Systems*, Vol. 1, ed J. H. Kaas (Oxford: Academic Press), 171–221.
- Haines, A. J., and Crampton, J. S. (2000). Improvements to the method of Fourier shape analysis as applied in morphometric studies. *Palaeontology* 43, 765–783. doi: 10.1111/1475-4983.00148
- Halpern, M. (1992). "Nasal chemical senses in reptiles," in *Biology of the Reptilia*, Vol. 18, eds C. Gans and D. Crews (Chicago, IL; London: University of Chicago Press), 423–523.
- Hammer, Ø., Harper, D. A. T., and Ryan, P. D. (2001). Past: paleontological statistics software package for education and data analysis. *Palaeontol. Electr.* 4, 1–9.
- Healy, S., and Guilford, T. (1990). Olfactory-bulb size and nocturnality in birds. *Evolution* 44, 339–346. doi: 10.1111/j.1558-5646.1990.tb05203.x
- Hedges, S. B., and Poling, L. L. (1999). A molecular phylogeny of reptiles. *Science* 283, 998–1001. doi: 10.1126/science.283.5404.998
- Henson Jr, O. (1974). "Comparative anatomy of the middle ear," in *Auditory System*, eds W. D. Keidel and W. D. Neff (Berlin: Springer Verlag), 39–110.
- Herrel, A., Van Damme, J., and Aerts, P. (2008). "Cervical anatomy and function in turtles," in *Biology of Turtles*, eds J. Wyneken, M. H. Godfrey, and V. Bels (New York, NY: CRC Press), 163–185.
- Herrera, Y., Fernández, M. S., and Gasparini, Z. (2013). The snout of *Cricosaurus araucanensis*: a case study in novel anatomy of the nasal region of metriorhynchids. *Lethaia* 46, 331–340. doi: 10.1111/let.12011
- Holloway, W. L., Claeson, K. M., and O'Keefe, F. R. (2013). A virtual phytosaur endocast and its implications for sensory system evolution in archosaurs. *J. Vertebr. Paleontol.* 33, 848–857. doi: 10.1080/02724634.2013.747532
- Hopson, J. A. (1979). "Palaeoneurology," in *Biology of the Reptilia*, Vol. 9, ed C. Gans (New York, NY: Academic Press), 39–146.
- Jaekel, O. (1918). Die Wirbeltierfunde aus dem Keuper von Halberstadt. Serie II. Testudinata. Teil 1. *Stegochelys dux*, n.g., n.sp. *Paläontol. Zeit.* 2, 88–214. doi: 10.1007/BF03160328
- Jirak, D., and Janacek, J. (2017). Volume of the crocodylian brain and endocast during ontogeny. *PLoS ONE* 12:e0178491. doi: 10.1371/journal.pone.0178491

- Joyce, W. G. (2015). The origin of turtles: a paleontological perspective. *J. Exp. Zool. B Mol. Dev. Evol.* 324, 181–193. doi: 10.1002/jez.b.22609
- Joyce, W. G., and Gauthier, J. A. (2004). Palaeoecology of Triassic stem turtles sheds new light on turtle origins. *P. Roy. Soc. Lond. B. Bio.* 271, 1–5. doi: 10.1098/rspb.2003.2523
- Joyce, W. G., Parham, J. F., and Gauthier, J. A. (2004). Developing a protocol for the conversion of rank-based taxon names to phylogenetically defined clade names, as exemplified by turtles. *J. Paleontol.* 78, 989–1013. doi: 10.1666/0022-3360(2004)078<0989:DAPFTC>2.0.CO;2
- Joyce, W. G., Rabi, M., Clark, J. M., and Xu, X. (2016). A toothed turtle from the Late Jurassic of China and the global biogeographic history of turtles. *BMC Evol. Biol.* 16:236. doi: 10.1186/s12862-016-0762-5
- Laaß, M., Schillingler, B., and Kaestner, A. (2017). What did the “unossified zone” of the non-mammalian therapsid braincase house? *J. Morphol.* 278, 1020–1032. doi: 10.1002/jmor.20583
- Laurin, M., and Piñeiro, G. H. (2017). A reassessment of the taxonomic position of mesosaurs, and a surprising phylogeny of early amniotes. *Front. Earth Sci.* 5:88. doi: 10.3389/feart.2017.00088
- Lautenschlager, S. (2014a). Morphological and functional diversity in therizinosaur claws and the implications for theropod claw evolution. *P. Roy. Soc. Lond. B. Bio.* 281:20140497. doi: 10.1098/rspb.2014.049
- Lautenschlager, S. (2014b). Palaeontology in the third dimension: a comprehensive guide for the integration of three-dimensional content in publications. *Paläontol. Zeit.* 88, 111–121. doi: 10.1007/s12542-013-0184-2
- Lautenschlager, S., and Butler, R. J. (2016). Neural and endocranial anatomy of Triassic phytosaurian reptiles and convergence with fossil and modern crocodylians. *PeerJ* 4:e2251. doi: 10.7717/peerj.2251
- Lee, M. S. (1997). Reptile relationships turn turtle. *Nature* 389, 245–245. doi: 10.1038/38422
- Li, C., Wu, X.-C., Rieppel, O., Wang, L.-T., and Zhao, L.-J. (2008). An ancestral turtle from the Late Triassic of southwestern China. *Nature* 456, 497–501. doi: 10.1038/nature07533
- Liu, J., Rieppel, O., Jiang, D.-Y., Aitchison, J. C., Motani, R., Zhang, Q.-Y., et al. (2011). A new pachypleurosaur (*Reptilia: Sauropterygia*) from the Lower Middle Triassic of Southwestern China and the phylogenetic relationships of Chinese pachypleurosaur. *J. Vertebr. Paleontol.* 85, 32–36. doi: 10.1666/09-131.1
- Lyson, T. R., Bever, G. S., Bhullar, B.-A. S., Joyce, W. G., and Gauthier, J. A. (2010). Transitional fossils and the origin of turtles. *Biol. Lett.* 6, 830–833. doi: 10.1098/rsbl.2010.0371
- Lyson, T. R., Rubidge, B. S., Scheyer, T. M., de Queiroz, K., Schachner, E. R., Smith, R. M. H., et al. (2016). Fossorial origin of the turtle shell. *Curr. Biol.* 26, 1887–1894. doi: 10.1016/j.cub.2016.05.020
- Mautner, A. K., Latimer, A. E., Fritz, U., and Scheyer, T. M. (2017). An updated description of the osteology of the pancake tortoise *Malacochersus tornieri* (*Testudines: Testudinidae*) with special focus on intraspecific variation. *J. Morphol.* 278, 321–333. doi: 10.1002/jmor.20640
- deBraga, M., and Rieppel, O. (1997). Reptile phylogeny and the interrelationships of turtles. *Zool. J. Linn. Soc. Lond.* 120, 281–354. doi: 10.1111/j.1096-3642.1997.tb01280.x
- Neenan, J. M., Klein, N., and Scheyer, T. M. (2013). European origin of placodont marine reptiles and the evolution of crushing dentition in Placodontia. *Nat. Commun.* 4, 1621. doi: 10.1038/ncomms2633
- Neenan, J. M., and Scheyer, T. M. (2012). The braincase and inner ear of *Placodus gigas* (*Sauropterygia, Placodontia*)—a new reconstruction based on micro-computed tomographic data. *J. Vertebr. Paleontol.* 32, 1350–1357. doi: 10.1080/02724634.2012.695241
- Parsons, T. S. (1959). Nasal anatomy and the phylogeny of reptiles. *Evolution* 13, 175–187. doi: 10.1111/j.1558-5646.1959.tb03003.x
- Parsons, T. S. (1970). “The Nose and Jacobson’s Organ,” in *Biology of the Reptilia*, Vol. 2B, ed C. Gans (New York, NY: Academic Press), 99–191.
- Paulina-Carabajal, A., Sterli, J., Georgi, J., Poropat, S. F., and Kear, B. P. (2017). Comparative neuroanatomy of extinct horned turtles (Meiolaniidae) and extant terrestrial turtles (*Testudinidae*), with comments on the palaeobiological implications of selected endocranial features. *Zool. J. Linn. Soc. Lond.* 180, 930–950. doi: 10.1093/zoolinnean/zlw024
- Pierce, S. E., Williams, M., and Benson, R. B. (2017). Virtual reconstruction of the endocranial anatomy of the early Jurassic marine crocodylomorph *Pelagosaurus typus* (*Thalattosuchia*). *PeerJ* 5:e3225. doi: 10.7717/peerj.3225
- Poglayen-Neuwall, I. (1953). Untersuchungen der Kiefermuskulatur und deren Innervation bei Schildkröten. *Acta Zool. Stockholm* 34, 241–292. doi: 10.1111/j.1463-6395.1953.tb00472.x
- Rieppel, O. (2007). “The Relationships of Turtles within Amniotes,” in *Biology of Turtles: from Structures to Strategies of Life*, eds J. Wyneken, M. H. Godfrey, and V. Bels (Boca Raton: CRC Press), 345–353.
- Rieppel, O. (2013). “The evolution of the turtle shell,” in *Morphology and Evolution of Turtles*, eds D. B. Brinkman, P. A. Holroyd, and J. D. Gardner (Heidelberg: Springer), 51–61.
- Rieppel, O., and Reisz, R. R. (1999). The origin and early evolution of turtles. *Annu. Rev. Ecol. Syst.* 30, 1–22. doi: 10.1146/annurev.ecolsys.30.1.1
- Rohlf, F. (2010). *TPSDig2, Version 2.16*. New York, NY: Stony Brook.
- Scheyer, T. M., Neenan, J. M., Bodogan, T., Furrer, H., Obrist, C., and Plamondon, M. (2017). A new, exceptionally preserved juvenile specimen of *Eusauropsargis dalsassoi* (*Diapsida*) and implications for Mesozoic marine diapsid phylogeny. *Sci. Rep.* 7:4406. doi: 10.1038/s41598-017-04514-x
- Scheyer, T. M., and Sander, P. M. (2007). Shell bone histology indicates terrestrial palaeoecology of basal turtles. *P. Roy. Soc. Lond. B. Bio.* 274, 1885–1893. doi: 10.1098/rspb.2007.0499
- Schoch, R. R., and Sues, H.-D. (2015). A Middle Triassic stem-turtle and the evolution of the turtle body plan. *Nature* 523, 584–587. doi: 10.1038/nature14472
- Schoch, R. R., and Sues, H.-D. (2017). Osteology of the Middle Triassic stem-turtle *Pappochelys rosinae* and the early evolution of the turtle skeleton. *J. Syst. Paleontol.* doi: 10.1080/14772019.2017.1354936. [Epub ahead of print].
- Schumacher, G. H. (1973). “The head muscles and hyolaryngeal skeleton of turtles and crocodylians,” in *Biology of the Reptilia*, eds C. Gans and T. S. Parsons (London; New York, NY: Academic Press), 101–200.
- Spoor, F. (2003). The semicircular canal system and locomotor behaviour, with special reference to hominin evolution. *Courier-Forschungsinstitut Senckenberg* 243, 93–104.
- Spoor, F., Garland, T., Krovitz, G., Ryan, T. M., Silcox, M. T., and Walker, A. (2007). The primate semicircular canal system and locomotion. *P. Acad. Nat. Sci. Phila.* 104, 10808–10812. doi: 10.1073/pnas.0704250104
- Sterli, J., Müller, J., Anquetin, J., and Hilger, A. (2010). The parabasisphenoid complex in Mesozoic turtles and the evolution of the testudinate basicranium. *Can. J. Earth Sci.* 47, 1337–1346. doi: 10.1139/E10-061
- von Baczko, M. B., and Desojo, J. B. (2016). Cranial Anatomy and Palaeoneurology of the Archosaur *Riojasuchus tenuisiceps* from the Los Colorados Formation, La Rioja, Argentina. *PLoSOne* 11:e0148575. doi: 10.1371/journal.pone.0148575
- Walsh, S. A., Barrett, P. M., Milner, A. C., Manley, G., and Witmer, L. M. (2009). Inner ear anatomy is a proxy for deducing auditory capability and behaviour in reptiles and birds. *Proc. Roy. Soc. Lond. B. Bio.* 276, 1355–1360. doi: 10.1098/rspb.2008.1390
- Wang, Z., Pascual-Anaya, J., Zadissa, A., Li, W., Niimura, Y., Huang, Z., et al. (2013). The draft genomes of soft-shell turtle and green sea turtle yield insights into the development and evolution of the turtle-specific body plan. *Nat. Genet.* 45, 701–706. doi: 10.1038/ng.2615
- Werneburg, I. (2011). The cranial musculature of turtles. *Palaeontol. Electron.* 14, 1–99.
- Werneburg, I. (2013). Jaw musculature during the dawn of turtle evolution. *Org. Divers. Evol.* 13, 225–254. doi: 10.1007/s13127-012-0103-5
- Werneburg, I. (2015). Neck motion in turtles and its relation to the shape of the temporal skull region. *C. R. Palevol* 14, 527–548. doi: 10.1016/j.crpv.2015.01.007
- Werneburg, I., Hinz, J. K., Gumpenberger, M., Volpato, V., Natchev, N., and Joyce, W. G. (2015a). Modeling neck mobility in fossil turtles. *J. Exp. Zool. B Mol. Dev. Evol.* 324, 230–243. doi: 10.1002/jez.b.22557
- Werneburg, I., and Sánchez-Villagra, M. R. (2009). Timing of organogenesis support basal position of turtles in the amniote tree of life. *BMC Evol. Biol.* 9:82. doi: 10.1186/1471-2148-9-82

- Werneburg, I., Wilson, L. A., Parr, W. C., and Joyce, W. G. (2015b). Evolution of neck vertebral shape and neck retraction at the transition to modern turtles: an integrated geometric morphometric approach. *Syst. Biol.* 64, 187–204. doi: 10.1093/sysbio/syu072
- Witmer, L. M., Ridgely, R. C., Dufeu, D. L., and Semones, M. C. (2008). “Using CT to Peer into the Past: 3D Visualization of the Brain and Ear Regions of Birds, Crocodiles, and Nonavian Dinosaurs,” in *Anatomical imaging*, eds H. Endo and R. Frey (Tokyo: Springer), 67–87.
- Yi, H., and Norell, M. A. (2015). The burrowing origin of modern snakes. *Sci. Adv.* 1:e1500743. doi: 10.1126/sciadv.1500743
- Zangerl, R. A. (1960). A new specimen of *Desmatochelys lowii* Williston; a primitive sea turtle from the Cretaceous of South Dakota. *Fieldiana Geol.* 14, 7–40.
- Zelenitsky, D. K., Therrien, F., and Kobayashi, Y. (2009). Olfactory acuity in theropods: palaeobiological and evolutionary implications. *Proc. R. Soc. B.* 276, 667–673. doi: 10.1098/rspb.2008.1075
- Zelenitsky, D. K., Therrien, F., Ridgely, R. C., McGee, A. R., and Witmer, L. W. (2011). Evolution of olfaction in non-avian theropod dinosaurs and birds. *Proc. R. Soc. B.* 278, 3625–3634. doi: 10.1098/rspb.2011.0238

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Lautenschlager, Ferreira and Werneburg. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.