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RECEIVED 14 November 2023

ACCEPTED 12 February 2024

PUBLISHED 04 March 2024

## CITATION

Zaczekiewicz M, Zimmermann O and  
Torzewski J (2024), A short review on CRP  
synthesis inhibition in cardiovascular disease.  
*Front. Drug Discov.* 4:1338535.  
doi: 10.3389/fddsv.2024.1338535

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# A short review on CRP synthesis inhibition in cardiovascular disease

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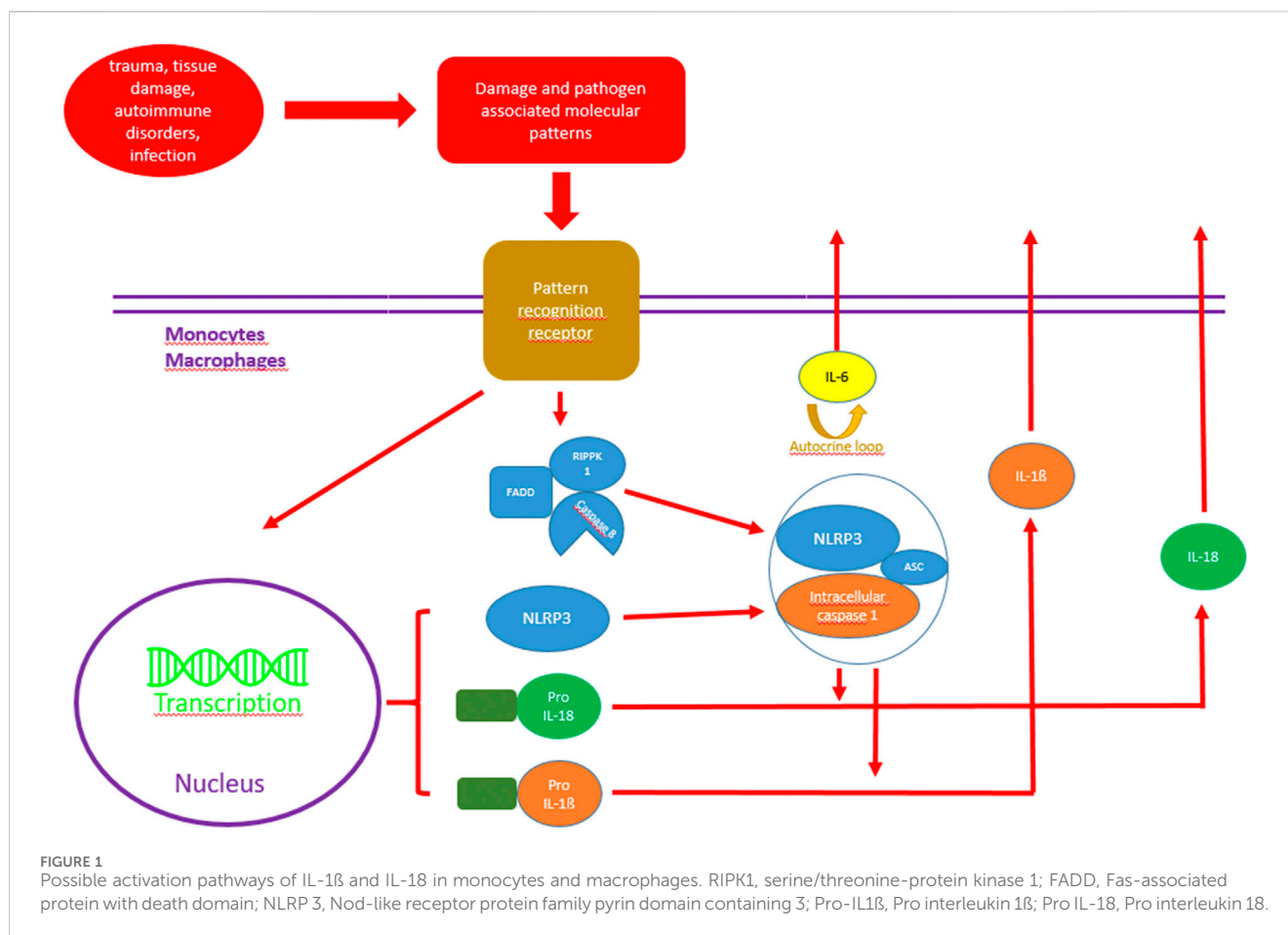
C-reactive Protein (CRP) is synthesized in the liver. Synthesis is stimulated via the IL-1 $\beta$ /IL6 pathway. CRP activates the complement system via C1q and macrophages via Fc $\gamma$  receptors. Since elevated CRP plasma levels are associated with increased cardiovascular risk, CRP may play a causal role in cardiovascular disease. One approach to transfer these observations into standard medical care would be to generate hepatic CRP synthesis inhibitors and use them in controlled clinical trials. Despite huge pharmacological efforts, the search for CRP synthesis inhibitors proved to be difficult. First, the antisense oligonucleotide RNA technology, although a promising idea, has not yet led to results feasible for clinical practice. Secondly, high throughput screening assays in search for hepatic CRP inhibitors were limited by the fact that primary human hepatocytes do not adequately grow *in vitro*. Use of genetically engineered hepatoma cells led to the observation that cardiac glycosides are capable of inhibiting CRP synthesis. Because of patent law considerations, however, pharmaceutical companies had limited interest in further pursuing this possible path. Upstream inhibition of IL-1 $\beta$  and IL-6 by antibodies has shown positive results in cardiovascular clinical trials, but because of side effects none of these antibodies has yet received FDA approval. In contrast, long-term colchicine treatment, though not being a CRP-specific approach, has recently been approved by the FDA. Taken together, there is no compelling evidence until today that hepatic CRP synthesis can specifically, effectively and safely be inhibited *in vivo* in human medicine. Currently, other avenues appear more promising. Here, we summarize contemporary approaches to inhibit CRP synthesis and potential goals for future clinical trials.

## KEYWORDS

cardiovascular disease, CRP synthesis, CRP inhibition, IL-6 downstream inhibition, IL-1 $\beta$  down stream inhibition

## 1 Introduction

C-reactive protein (CRP) is an acute phase protein consisting of five identical, non-covalently bound subunits and belongs to the pentraxin family (Gang et al., 2012; Du Clos, 2013). Each subunit contains a calcium-dependent phosphocholine binding site (Thompson et al., 1999). Following phosphocholine binding, a change in CRP conformation leads to C1q+ binding and, consequently, activation of the classical complement pathway (Roux et al., 1983; Volanakis, 2001; Du Clos, 2013). Unlike complement activation via antibodies, CRP-mediated complement activation is reported to result in almost complete consumption of C1, C4, C2 and (partially) C3 but only in a low activation level of terminal membrane attack complex mC5b-9 (Kaplan and Volanakis,



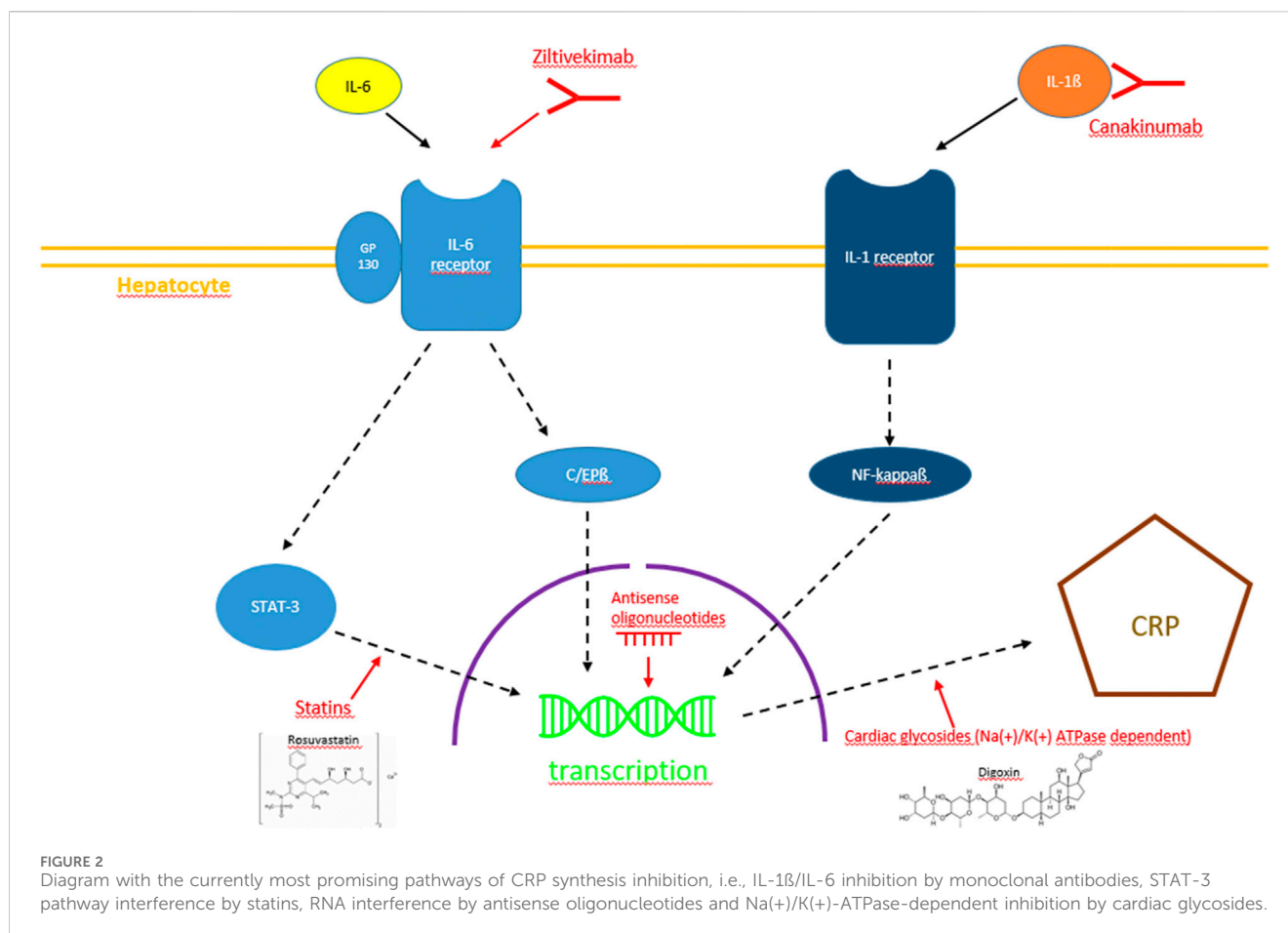
1974). CRP also interacts with Fc $\gamma$  receptors, especially Fc $\gamma$ RIIa, on monocytes and neutrophils leading to opsonisation, phagocytosis and cytokine release (Bharadwaj et al., 1999; Stein et al., 2000; Manolov et al., 2004; Lu et al., 2008). From an evolutionary genetic standpoint CRP is one of the oldest and most unspecific parts of the innate human immune system being a pattern recognition molecule binding to molecules exposed during cell death and expressed on the surfaces of pathogens (Black et al., 2004)

In humans, CRP is mainly synthesized in hepatic cells through activation at the transcriptional level by a variety of pathways. A low baseline production is induced by liver specific transcription factor NF-1 (Hurlimann et al., 1966). In case of sterile or infectious danger signals (damage-associated molecular patterns, pathogen-associated molecular patterns) caused by external triggers (trauma, tissue damage, autoimmune disorders, infection), innate immune cell pattern recognition receptors (PRRs) are activated (Latz et al., 2013). A great variety of different tissue damaging patterns induce inflammasome formation, the most prominent being nucleotide oligomerization domain-, leucine-rich repeat-, and pyrin domain containing protein (3NLRP3) inflammasome. Activated inflammasomes then act as a large multimolecular signaling platform and thereby induce maturation and release of cytokines, especially pro interleukin (IL)-1 $\beta$  and IL-18 (Próchnicki et al., 2016).

Pro IL-1 $\beta$  is mainly a product of tissue macrophages, dendritic cells and blood monocytes and needs intracellular caspase 1 (also activated by inflammasomes) in order to be transferred into the

active form IL-1 $\beta$  (Martinon et al., 2009; Dinarello and van der Meer, 2013). After binding to the membrane bound IL-1 receptor (IL-1R), IL-1 $\beta$  activates a cascade that results, among others, in intracellular activation of NF-kappaB, leading to cytokine and CRP induction. The same pathway and activation of NF-kappaB is also induced by IL-18 (Kaplanski, 2018). Figure 1 shows possible IL-1 $\beta$  activation pathways.

IL-6 can be found in and produced by different tissues and has many functions involving the regulation of the immune system and hematopoiesis as well as inflammation and acute phase response (Tanaka et al., 2014). IL-6 is mainly produced by monocytes and macrophages (Tanaka et al., 2016). It binds to the membrane bound IL-6 receptor (IL-6R), which interacts with membrane bound glycoprotein 130 and leads to an activation of NF-kappaB and the transcription factors STAT3 and C/EBP $\beta$ , all capable of upregulating CRP expression. Following a transcriptional complex formation of c-Fos, STAT3, and hepatocyte NF-1 alpha, which is essential for cytokine-driven C-reactive protein gene expression, this complex binds to the CRP promoter and upregulates CRP synthesis (Zhang et al., 1996; Agrawal et al., 2001; Singh et al., 2007; Nishikawa et al., 2008; Young et al., 2008). With the simultaneous presence of interleukin-1 $\beta$  (IL-1 $\beta$ ), hepatic CRP production is raised exponentially (Pepys and Hirschfield, 2003), potentially by triggering an autocrine IL-6 loop (Kramer et al., 2008). The latter displays the crucial role of IL-1 $\beta$  in the activation of the humoral part of the innate immune system (Grebe et al., 2018).



In clinical routine, CRP has, up to the present day, predominantly been quantified in inflammatory and infectious conditions and is widely used to monitor various autoimmune diseases such as rheumatoid arthritis (Du Clos, 2013). CRP levels are also known to correlate with mortality in septic patients with multiple organ failure (Lobo et al., 2003). Elevated CRP baseline levels are associated with an increased risk for cardiovascular events, predominantly represented by stable coronary artery disease, myocardial infarction and ischemic stroke, as first reported by Ridker et al. (1997). A large meta-analysis confirmed the correlation of elevated baseline CRP levels with a higher likelihood of relevant coronary disease (i.e., patients suffering from angina on exertion or myocardial infarction), ischemic stroke and mortality (Emerging Risk Factors Collaboration et al., 2010). In addition CRP has been shown to co-localize in atherosclerotic plaques in humans as well as mice (Reynolds and Vance, 1987; Torzewski et al., 1998; Zimmermann et al., 2014). Since these findings may imply a causal role of elevated CRP plasma levels in various diseases instead of CRP just being a marker, inhibiting CRP synthesis could be a potential therapeutic target in the setting of acute infectious and autoimmune diseases as well as in cardiovascular disease. Discussions are, however, still ongoing and controversial (Torzewski, 2005; Pepys, 2008; Ridker, 2019; Jimenez and Szalai, 2021).

One way to answer the question of CRP being a worthwhile therapeutic target would be to lower plasma levels via selective CRP synthesis inhibition in the liver in the setting of the above-mentioned diseases *in vivo*. Consequently, this article focuses on

contemporary approaches of CRP synthesis inhibition in the setting of cardiovascular disease. Substances inhibiting CRP synthesis and their potential intracellular targets are depicted in Figure 2.

## 2 Natural substances capable of lowering CRP plasma levels

A variety of natural substances such as rosmarinic acid, tart cherry juice and green tea have recently been reported to have anti-inflammatory effects with the capability of reducing CRP levels (Asbaghi et al., 2019; Chai et al., 2019; Yao et al., 2019; Asbaghi et al., 2020). The mechanisms remain largely unclear. Nonetheless, in the prevention of atherosclerosis and its sequelae such substances are worth further investigation, especially for primary prevention. In Table 1 we summarize the most natural promising substances with reported effects on CRP levels.

## 3 Reduction of CRP plasma levels via upstream inhibition of the IL-1β/IL-6/CRP pathway

The JUPITER trial (Ridker et al., 2008) showed a CRP lowering effect of 3-Hydroxy-3-Methylglutaryl-Coenzyme-A-Reductase-(HMG-CoA-Reductase) inhibitors (statins) and also a reduction

TABLE 1 Natural substances with anti-inflammatory and CRP lowering effects.

Substance	Effect	Dosing	Possible mechanism
rosmarinic acid	Pretreatment of smooth muscle cells with RA led to less inflammatory response including reduced CRP levels after exposing the cells to nicotinic acid	Effect was time and dose dependant	NLRP3 inflammasome inhibition in smooth muscle cells
tart cherry juice	Relevant reduction of plasma CRP levels, blood pressure and LDL-cholesterol	240 mL of tart cherry juice 2 x/day, 12 weeks	Mechanism unclear
green tea	Relevant Reduction of plasma CRP levels and total cholesterol levels	800 mg of green tea extract/day, > 8 weeks	Mechanism unclear

of cardiovascular event rates in patients with cholesterol levels of less than 130 mg/dL and CRP plasma levels of 2 mg/L or higher. These findings implied an anti-inflammatory effect of statins in addition to the reduction of cholesterol levels and underlined the importance of research on anti-inflammatory therapies in cardiovascular disease. Statins themselves have been part of the optimal medical treatment of cardiovascular disease since more than 40 years, nonetheless the mechanisms of statin-induced reduction of CRP plasma levels are not yet completely unraveled. Statins have shown to lower CRP levels in multiple ways. One reason being the reduction of LDL and oxidized LDL levels through 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase inhibition as well as the upregulation of HDL and its key component apolipoprotein A-I. Both mechanisms lead to an indirect reduction of CRP synthesis in smooth muscle cells and macrophages in atherosclerotic plaques (Calabro et al., 2003; Arévalo-Lorido, 2016). Another potentially important mechanism involves the inhibition of protein isoprenylation, which leads to an interference with a group of intracellular signaling involving guanosine triphosphate (GTP)-binding proteins as Rho, Rac and Ras (Schönbeck and Libby, 2004). Causing a reduction of RAC-1 geranylgeranylation leads to a lower level of IL-6 induced serine phosphorylation of the transcription factor STAT3 reducing CRP synthesis (Arnaud et al., 2005).

Consecutively, Ridker et al. designed a trial which examined whether the inhibition of inflammation by IL-1 $\beta$  targeting without influencing cholesterol levels at all would also have an effect on cardiovascular events. The CANTOS trial (Ridker et al., 2017) used Canakinumab, a human monoclonal antibody neutralizing IL-1 $\beta$ , for inhibition of the IL-1 $\beta$ /IL-6/CRP pathway. The CANTOS trial indeed showed a highly relevant risk reduction for cardiovascular events in individuals with an increased risk for cardiovascular disease. Canakinumab lowered CRP, fibrinogen and IL-6 levels effectively without influencing cholesterol levels. Side effects, however, included more severe infectious diseases in the treatment group compared to the control group and thus, Canakinumab in this indication, did not receive FDA approval. Furthermore, CANTOS did not answer the question of a causal CRP involvement in cardiovascular disease but proved the vital role of inflammation once more. The COLCOT trial (Tardif et al., 2019) confirmed these results by showing reduced cardiovascular event rates due to several anti-inflammatory effects of colchicine, most notably IL-1 and IL-8 inhibition (Leung et al., 2015), in secondary disease prevention. Based on the LoDoCo2 trial (Nidorf et al., 2020), low dose Colchicine has recently received FDA approval for the secondary prevention of cardiovascular disease. The latest study

investigating upstream CRP inhibition was the recently published RESCUE trial, a phase II study, which showed effective reduction of CRP levels through IL-6 inhibition via Ziltevikimab in a dose-dependent manner in patients at high cardiovascular risk and suffering from chronic kidney disease (Ridker et al., 2021). Based on these results, a large-scale randomized clinical trial has been initiated investigating the effect of Ziltevikimab on cardiovascular endpoints, results are pending. Table 2 summarizes the key results of the studies mentioned above.

Direct JAK inhibition to our knowledge has not yet been investigated in the context of cardiovascular disease. It has been shown though that JAK inhibitors are capable of lowering CRP levels (Pin et al., 2020). In clinical practice JAK inhibitors are mainly used to treat autoimmune disorders and strong data is available for patients with rheumatoid arthritis, which has led to JAK inhibitors being a part of the European and American guidelines treatment algorithms for patients with rheumatoid arthritis (Fraenkel et al., 2021; Smolen et al., 2023). In this context there have been evaluations of the cardiovascular safety of JAK inhibitors showing positive effects or no relevant effects on cardiovascular events, however these were metaanalyses (Charles-Schoeman et al., 2016; Charles-Schoeman et al., 2019; Xie et al., 2019).

## 4 CRP synthesis inhibition via antisense oligonucleotides

Antisense oligonucleotides (ASOs) are short single-stranded nucleic acids that selectively bind to a specific part of mRNA and can be used to modify gene expression. They have to be administered parenterally and processed to ensure cellular uptake (Bennett and Swayze, 2010). Szalai et al. successfully used ASOs to inhibit CRP expression and to lower CRP plasma levels in rats and in CRP transgenic mice. ASO treatment resulted in the reduction of atherosclerotic plaque burden compared to rats and transgenic mice which were not treated with ASOs (Szalai et al., 2014). Yu et al. showed effective reduction of CRP levels with ASOs in Watanabe heritable hyperlipidemic rabbits. In contrast to Szalai et al., this study could not demonstrate any difference in atherosclerotic plaque burden between ASO-treated and -non treated Watanabe heritable hyperlipidemic rabbits (Yu et al., 2014).

The only larger clinical trial showing that ASOs can effectively inhibit CRP synthesis in humans used endotoxin to induce an acute phase reaction. An effect on the release of other acute phase proteins

TABLE 2 Relevant clinical studies investigating reduction of CRP plasma levels.

Study	Inclusion criteria	Drug administration	Results
JUPITER (Ridker et al., 2008)	17,802 healthy adults with LDL-cholesterol level of <130 mg/dL, CRP level of >2 mg/dL	Randomization: 20 mg Rosuvastatin/day vs. Placebo	Median follow up 1.9 years 37% reduction of CRP plasma levels Significant reduction of death, MI and revascularization due to angina in Rosuvastatin group
CANTOS (Ridker et al., 2017)	10,061 patients with CRP level >2 mg/dL and previous myocardial infarction	50 mg, 150 mg or 300 mg Canakinumab administered subcutaneously every 3 months vs. Placebo	Significant reduction of combined efficacy end point (non-fatal myocardial infarction, stroke or cardiovascular death) in the 150 mg and 300 mg groups after 48 months Significantly higher incidence of fatal infection and sepsis
COLCOT (Tardif et al., 2019)	4,745 patients within 30 days after myocardial infarction	OMT plus 0.5 mg of Colchicine once a day versus OMT plus Placebo	Median follow up: 22.6 months Significant reduction of primary endpoint (composite of death from cardiovascular causes, resuscitated cardiac arrest, myocardial infarction, stroke, or urgent hospitalization for angina leading to coronary revascularization): 5.5% in Colchicine vs. 7.1% in Placebo group Significantly more infectious diseases and pneumonia in the Colchicine group
LoDoCo2 (Nidorf et al., 2020)	5,522 patients with chronic coronary disease	0.5 mg Colchicine once a day	Median follow 28.6 months Primary endpoint (cardiovascular death, spontaneous (nonprocedural) myocardial infarction, ischemic stroke, or ischemia-driven coronary revascularization) was significantly lower in Colchicine group: 6.8% (Colchicine) vs. 9.6% (Placebo)
RESCUE (Ridker et al., 2021)	264 patients with chronic kidney disease and CRP level > 2 mg/dL	Randomized 1:1:1:1: Placebo, Ziltivekimab 7.5 mg every 4 weeks, Ziltivekimab 15 mg every 4 weeks and Ziltivekimab 30 mg every 4 weeks	Significant reduction of hs-CRP plasma levels after 12 weeks
C-DOS (Zaczekiewicz et al., 2022)	60 patients with cardiac decompensation (NYHA III and IV) and no infection were assigned according to clinical needs and contraindications to OMT or OMT + Digoxin (1:1)	Digoxin and hs-CRP plasma levels were monitored in both groups on day 1, 3, 5, 7 and 21	After 21 days, borderline significant reduction of hs-CRP levels ( $p = 0.051$ ) in the Digoxin group compared to the OMT only group

was not seen (Novcek et al., 2014). In 2012, another group had already reported effective reduction of CRP levels using ASOs in 8 adults (Jones et al., 2012). Since then, however, no further studies or randomized controlled trials applying ASOs in patients suffering from cardiovascular disease have been published, especially no studies investigating clinical endpoints.

## 5 Direct drug-induced CRP synthesis inhibition

A number of drugs affect CRP plasma levels. Most of them, however, influence CRP levels indirectly (Prasad, 2006). The search for a substance directly inhibiting CRP synthesis proved to be difficult. The main reason was that, *in vitro*, primary human hepatocytes (although undoubtedly being the model closest to the human liver), do not grow adequately, and limited access to adequate tissue prevents their use in high throughput screening assays (Gómez-Lechón et al., 2003). Anyway, high throughput screening assays utilizing genetically engineered hepatoma cells

transfected with the human CRP promoter finally identified cardiac glycosides as potent inhibitors of *in vitro* CRP synthesis. It was indeed shown that cardiac glycosides inhibit the IL-1 $\beta$ -/IL-6 induced expression of acute phase proteins in human hepatoma cells as well as primary human hepatocytes via a Na(+)/K(+)-ATPase-dependent pathway, which has not been completely demasked (Kolkhof et al., 2010).

Up to the present day, however, there has been no larger study investigating CRP synthesis inhibition by cardiac glycosides in humans. This is partly due to the fact that cardiac glycosides have been used in medical therapy for more than two centuries (Withering, 1785) causing relevant patent law considerations by pharmaceutical companies. Furthermore, the well-known narrow therapeutic window as well as side effects of cardiac glycosides, especially AV-blockage and ventricular arrhythmia (Hauptman and Kelly, 1999; Adams et al., 2014), limit their use in long-term medication. The single-center C-reactive protein-Digoxin Observational Study (C-DOS) provides a first piece of evidence that cardiac glycosides may indeed inhibit CRP synthesis *in vivo* in humans (Zaczekiewicz et al., 2022).

TABLE 3 Current state and advantages/disadvantages of each of the CRP-lowering approaches.

Approach	Current state of investigation	Advantages	Disadvantages	Costs
IL1- $\beta$ /IL-6 Inhibition	Clinical trials published. No FDA approval yet	Highly effective reduction of CRP plasma levels	Increase of infections observed; parenteral administration necessary	High costs per patient
Statins (STAT-3 inhibition)	Well documented anti-inflammatory effects including CRP reduction	Oral administration	CRP inhibition is a side effect, other methods are more potent	High cost-effectiveness, low NNT
Colchicine	FDA approved for secondary prevention of cardiovascular disease	Oral administration	Colchicine has many anti-inflammatory effects, no direct CRP inhibition	Low costs per patient
ASOs (antisense oligonucleotides)	Small clinical trials only, large clinical trials still pending	Highly effective and selective CRP synthesis inhibition	Parenteral administration necessary. Multiple ASOs needed?	High costs per patient
CRP apheresis	Proof of concept in smaller trials, large trials still not published	Highly selective extracorporeal reduction of CRP plasma levels	Invasive method, only short-term CRP level reduction	High costs, may play a significant role in acute settings due to "simple", highly selective, highly effective and non-toxic approach
CRP-inhibition with cardiac glycosides	Only one small observational trial with borderline significance	Oral administration	Narrow therapeutic width with significant side effects	Low costs per patient
Inhibition of p-CRP to m-CRP dissociation	As of now no clinical trials	Highly effective approach to reduce inflammatory activity of CRP	No human trials yet	To be determined

## 6 Other contemporary ways of C-reactive protein inhibition/elimination

Thiele et al. observed that in human striated muscle, human atherosclerotic plaque, and infarcted myocardium (in both rat and human myocardium), monomeric CRP (mCRP) was colocalized with inflammatory cells rather than pentameric CRP (pCRP), which is mainly found in the blood serum. These findings suggested that inhibition of the dissociation of pCRP to mCRP may prove to be a vital approach in order to reduce CRP-induced inflammation (Badimon et al., 2018, Caprio et al., 2018, Filep et al., 2023). Thiele et al. showed in 2014 that inhibiting the phospholipase A2-dependent dissociation from pCRP to mCRP via stabilization of pCRP with 1,6-bis(phosphocholine)-hexane showed significantly less inflammatory tissue damage after inducing myocardial infarction in a rat model (Thiele et al., 2014). The most recent study dealing with this issue was published by Zeller et al., in 2023. The group designed the novel substance C10M derived from phosphocholine, which is expressed in activated cells and binds to the  $\beta$ -interface of pCRP. C10M was able to successfully inhibit pCRP dissociation into mCRP *in vitro* and *in vivo* in a mouse model with limb transplantation (Zeller et al., 2023). These promising results should encourage further studies following up on this approach.

A summary of CRP apheresis as a means of extracorporeal elimination of CRP has been published elsewhere (Ries et al. 2019; Torzewski et al., 2022).

## 7 Conclusion

In order to answer the question whether CRP plays a causal role in inflammatory processes, specific hepatic CRP synthesis inhibition may be helpful.

A promising approach seems the ASO technology. ASO technology is highly selective. Disadvantages of ASOs, however, include the need for parenteral application on the one hand and the expected high costs in case of a broader use on the other hand. No larger studies investigating the effect of CRP inhibition by ASOs on cardiovascular endpoints have been published yet.

Selective pharmacological inhibition of hepatic CRP synthesis, up to the present day, has not really been successful. Cardiac glycosides are the only substances that have been demonstrated to inhibit CRP synthesis in hepatocytes *in vitro* and lower CRP plasma levels *in vivo* in a small observational trial. Chemical modification of cardiac glycosides, pharmacological improvement and randomized controlled trials looking at cardiovascular endpoints would be necessary. This approach, however, is expensive and needs to overcome limitations like the toxicity and small therapeutic window of cardiac glycosides.

Most substantial progress has been made using upstream inhibitors, especially monoclonal IL-6 antibodies, with the first randomized controlled trial looking at cardiovascular endpoints. Potential disadvantages of this approach may be the need of parenteral application, the expected costs and the potential side effects of targeting a cytokine involved in immune regulation, hematopoiesis and acute phase response. Upstream inhibition with specific IL-6 inhibitors has already reached the phase of randomized controlled clinical trials. Long-term Colchicine application for secondary prevention of cardiovascular disease has recently received FDA approval. The latter is comparatively cheap and may potentially be suitable for daily clinical practice. We summarized the current state of investigations regarding the above mentioned approaches of CRP inhibition as well as the advantages and disadvantages in Table 3.

Future goals should include identifying patient populations benefiting the most from each of the specific approaches,

eventually artificial intelligence could be helpful in this process. These results should be combined with considerations regarding available resources to optimize resource allocation.

## Author contributions

MZ: Conceptualization, Writing—original draft. OZ: Writing—review and editing. JT: Conceptualization, Writing—review and editing.

## Funding

The author(s) declare that no financial support was received for the research, authorship, and/or publication of this article.

## References

- Adams, K. F., Jr, Ghali, J. K., Herbert Patterson, J., Stough, W. G., Butler, J., Bauman, J. L., et al. (2014). A perspective on re-evaluating digoxin's role in the current management of patients with chronic systolic heart failure: targeting serum concentration to reduce hospitalization and improve safety profile. *Eur. J. Heart Fail* 16 (5), 483–493. doi:10.1002/ehf.64
- Agrawal, A., Cha-Molstad, H., Samols, D., and Kushner, I. (2001). Transactivation of C-reactive protein by IL-6 requires synergistic interaction of CCAAT/enhancer binding protein beta (C/EBP beta) and Rel p50. *J. Immunol.* 166 (4), 2378–2384. doi:10.4049/jimmunol.166.4.2378
- Arévalo-Lorido, J. C. (2016). Clinical relevance for lowering C-reactive protein with statins. *Ann. Med.* 48 (7), 516–524. doi:10.1080/07853890.2016.1197413
- Arnaud, C., Burger, F., Steffens, S., Veillard, N. R., Nguyen, T. H., Trono, D., et al. (2005). Statins reduce interleukin-6-induced C-reactive protein in human hepatocytes: new evidence for direct antiinflammatory effects of statins. *Arterioscler. Thromb. Vasc. Biol.* 25 (6), 1231–1236. doi:10.1161/01.ATV.0000163840.63685.0c
- Asbahi, O., Fouladvand, F., Gonzalez, M. J., Aghamohammadi, V., Choghakhori, R., and Abbasnezhad, A. (2019). The effect of green tea on C-reactive protein and biomarkers of oxidative stress in patients with type 2 diabetes mellitus: a systematic review and meta-analysis. *Complement. Ther. Med.* 46, 210–216. doi:10.1016/j.ctim.2019.08.019
- Asbahi, O., Fouladvand, F., Moradi, S., Ashtary-Larky, D., Choghakhori, R., and Abbasnezhad, A. (2020). Effect of green tea extract on lipid profile in patients with type 2 diabetes mellitus: a systematic review and meta-analysis. *Diabetes Metab. Syndr.* 14 (4), 293–301. doi:10.1016/j.dsx.2020.03.018
- Badimon, L., Peña, E., Arderiu, G., Padró, T., Slevin, M., Vilahur, G., et al. (2018). C-reactive protein in atherothrombosis and angiogenesis. *Front. Immunol.* 9, 430. doi:10.3389/fimmu.2018.00430
- Bennett, C. F., and Swayze, E. E. (2010). RNA targeting therapeutics: molecular mechanisms of antisense oligonucleotides as a therapeutic platform. *Annu. Rev. Pharmacol. Toxicol.* 50, 259–293. doi:10.1146/annurev.pharmtox.010909.105654
- Bharadwaj, D., Stein, M. P., Volzer, M., Mold, C., and Du Clos, T. W. (1999). The major receptor for C-reactive protein on leukocytes is fcgamma receptor II. *J. Exp. Med.* 190 (4), 585–590. doi:10.1084/jem.190.4.585
- Black, S., Kushner, I., and Samols, D. (2004). C-Reactive protein. *J. Biol. Chem.* 279 (47), 48487–48490. doi:10.1074/jbc.R400025200
- Calabró, P., Willerson, J. T., and Yeh, E. T. (2003). Inflammatory cytokines stimulated C-reactive protein production by human coronary artery smooth muscle cells. *Circulation* 108 (16), 1930–1932. doi:10.1161/01.CIR.0000096055.62724.C5
- Caprio, V., Badimon, L., Di Napoli, M., Fang, W. H., Ferris, G. R., Guo, B., et al. (2018). pCRP-mCRP dissociation mechanisms as potential targets for the development of small-molecule anti-inflammatory chemotherapeutics. *Front. Immunol.* 9, 1089. doi:10.3389/fimmu.2018.01089
- Chai, S. C., Davis, K., Zhang, Z., Zha, L., and Kirschner, K. F. (2019). Effects of tart cherry juice on biomarkers of inflammation and oxidative stress in older adults. *Nutrients* 11 (2), 228. doi:10.3390/nu11020228
- Charles-Schoeman, C., Wicker, P., Gonzalez-Gay, M. A., Boy, M., Zuckerman, A., Soma, K., et al. (2016). Cardiovascular safety findings in patients with rheumatoid

## Conflict of interest

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The author(s) declared that they were an editorial board member of Frontiers, at the time of submission. This had no impact on the peer review process and the final decision.

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arthritis treated with tofacitinib, an oral janus kinase inhibitor. *Semin. Arthritis Rheum.* 46, 261–271. doi:10.1016/j.semarthrit.2016.05.014

Charles-Schoeman, C., DeMasi, R., Valdez, H., Soma, K., Hwang, L. J., Boy, M. G., et al. (2019). Risk factors for major adverse cardiovascular events in phase iii and long-term extension studies of tofacitinib in patients with rheumatoid arthritis. *Arthritis Rheumatol.* 71, 1450–1459. doi:10.1002/art.40911

Dinarello, C. A., and van der Meer, J. W. (2013). Treating inflammation by blocking interleukin-1 in humans. *Semin. Immunol.* 25 (6), 469–484. doi:10.1016/j.smim.2013.10.008

Du Clos, T. W. (2013). Pentraxins: structure, function, and role in inflammation. *ISRN Inflamm.* 2013, 379040. doi:10.1155/2013/379040

Emerging Risk Factors CollaborationKaptoge, S., Di Angelantonio, E., Lowe, G., Pepys, M. B., Thompson, S. G., et al. (2010). C-reactive protein concentration and risk of coronary heart disease, stroke, and mortality: an individual participant meta-analysis. *Lancet* 375 (9709), 132–140. doi:10.1016/S0140-6736(09)61717-7

Filep, J. G. (2023). Targeting conformational changes in C-reactive protein to inhibit pro-inflammatory actions. *EMBO Mol. Med.* 15 (1), e17003. Epub 2022 Dec 5. PMID: 36465053; PMCID: PMC9832832. doi:10.15252/emmm.202217003

Fraenkel, L., Bathon, J. M., England, B. R., St Clair, E. W., Arayssi, T., Carandang, K., et al. (2021). 2021 American college of rheumatology guideline for the treatment of rheumatoid arthritis. *Arthritis Care Res. Hob.* 73 (7), 924–939. doi:10.1002/acr.24596

Gang, T. B., Hammond, D. J., Jr, Singh, S. K., Ferguson, D. A., Jr, Mishra, V. K., and Agrawal, A. (2012). The phosphocholine-binding pocket on C-reactive protein is necessary for initial protection of mice against pneumococcal infection. *J. Biol. Chem.* 287 (51), 43116–43125. doi:10.1074/jbc.M112.427310

Gómez-Lechón, M. J., Donato, T., Ponsoda, X., and Castell, J. V. (2003). Human hepatic cell cultures: *in vitro* and *in vivo* drug metabolism. *Altern. Lab. Anim.* 31 (3), 257–265. doi:10.1177/026119290303100307

Grebe, A., Hoss, F., and Latz, E. (2018). NLRP3 inflammasome and the IL-1 pathway in atherosclerosis. *Circ. Res.* 122 (12), 1722–1740. doi:10.1161/CIRCRESAHA.118.311362

Hauptman, P. J., and Kelly, R. A. (1999). Digitalis. *Circulation.* 99 (9), 1265–1270. doi:10.1161/01.cir.99.9.1265

Hurlimann, J., Thorbecke, G. J., and Hochwald, G. M. (1966). The liver as the site of C-reactive protein formation. *J. Exp. Med.* 123 (2), 365–378. doi:10.1084/jem.123.2.365

Jimenez, R. V., and Szalai, A. J. (2021). Therapeutic lowering of C-reactive protein. *Front. Immunol.* 11, 619564. doi:10.3389/fimmu.2020.619564

Jones, N. R., Pegues, M. A., McCrory, M. A., Singleton, W., Bethune, C., Baker, B. F., et al. (2012). A selective inhibitor of human C-reactive protein translation is efficacious *in vitro* and in C-reactive protein transgenic mice and humans. *Mol. Ther. Nucleic Acids* 1 (11), e52. doi:10.1038/mtna.2012.44

Kaplan, M. H., and Volanakis, J. E. (1974). Interaction of C-reactive protein complexes with the complement system. *J. Immunol.* 112 (6), 2135–2147. PMID: 4151108. doi:10.4049/jimmunol.112.6.2135

Kaplanski, G. (2018). Interleukin-18: biological properties and role in disease pathogenesis. *Immunol. Rev.* 281 (1), 138–153. doi:10.1111/imr.12616

- Kolkhof, P., Geerts, A., Schäfer, S., and Torzewski, J. (2010). Cardiac glycosides potently inhibit C-reactive protein synthesis in human hepatocytes. *Biochem. Biophys. Res. Commun.* 394 (1), 233–239. doi:10.1016/j.bbrc.2010.02.177
- Kramer, F., Torzewski, J., Kamenz, J., Veit, K., Hombach, V., Dedio, J., et al. (2008). Interleukin-1 $\beta$  stimulates acute phase response and C-reactive protein synthesis by inducing an NF $\kappa$ B- and C/EBP $\beta$ -dependent autocrine interleukin-6 loop. *Mol. Immunol.* 45 (9), 2678–2689. doi:10.1016/j.molimm.2007.12.017
- Latz, E., Xiao, T. S., and Stutz, A. (2013). Activation and regulation of the inflammasomes. *Nat. Rev. Immunol.* 13 (6), 397–411. doi:10.1038/nri3452
- Leung, Y. Y., Yao Hui, L. L., and Kraus, V. B. (2015). Colchicine—Update on mechanisms of action and therapeutic uses. *Semin. Arthritis Rheum.* 45 (3), 341–350. doi:10.1016/j.semarthrit.2015.06.013
- Lobo, S. M., Lobo, F. R., Bota, D. P., Lopes-Ferreira, F., Soliman, H. M., Mélot, C., et al. (2003). C-reactive protein levels correlate with mortality and organ failure in critically ill patients. *Chest* 123 (6), 2043–2049. doi:10.1378/chest.123.6.2043
- Lu, J., Marnell, L. L., Marjon, K. D., Mold, C., Du Clos, T. W., and Sun, P. D. (2008). Structural recognition and functional activation of Fc $\gamma$ 2b $\mu$ R by innate pentraxins. *Nature* 456 (7224), 989–992. doi:10.1038/nature07468
- Manolov, D. E., Röcker, C., Hombach, V., Nienhaus, G. U., and Torzewski, J. (2004). Ultrasensitive confocal fluorescence microscopy of C-reactive protein interacting with Fc $\gamma$ 2b $\mu$ R. *Arterioscler. Thromb. Vasc. Biol.* 24 (12), 2372–2377. doi:10.1161/01.ATV.0000147407.17137.02
- Martinon, F., Mayor, A., and Tschopp, J. (2009). The inflammasomes: guardians of the body. *Annu. Rev. Immunol.* 27, 229–265. doi:10.1146/annurev.immunol.021908.132715
- Nidorf, S. M., Fiolet, A. T. L., Mosterd, A., Eikelboom, J. W., Schut, A., Opstal, T. S. J., et al. (2020). Colchicine in patients with chronic coronary disease. *N. Engl. J. Med.* 383 (19), 1838–1847. doi:10.1056/NEJMoa2021372
- Nishikawa, T., Hagihara, K., Serada, S., Isobe, T., Matsumura, A., Song, J., et al. (2008). Transcriptional complex formation of c-Fos, STAT3, and hepatocyte NF-1 $\alpha$  is essential for cytokine-driven C-reactive protein gene expression. *J. Immunol.* 180 (5), 3492–3501. doi:10.4049/jimmunol.180.5.3492
- Noveck, R., Stroes, E. S., Flaim, J. D., Baker, B. F., Hughes, S., Graham, M. J., et al. (2014). Effects of an antisense oligonucleotide inhibitor of C-reactive protein synthesis on the endotoxin challenge response in healthy human male volunteers. *J. Am. Heart Assoc.* 3 (4), e001084. doi:10.1161/JAHA.114.001084
- Pepys, M. B., and Hirschfield, G. M. (2003). C-reactive protein: a critical update. *J. Clin. Invest.* 111 (12), 1805–1812. doi:10.1172/JCI18921
- Pepys, M. B. (2008). C-reactive protein is neither a marker nor a mediator of atherosclerosis. *Nat. Clin. Pract. Nephrol.* 4 (5), 234–235. doi:10.1038/ncpneph0778
- Pin, A., Tesser, A., Pastore, S., Moressa, V., Valencic, E., Arbo, A., et al. (2020). Biological and clinical changes in a pediatric series treated with off-label JAK inhibitors. *Int. J. Mol. Sci.* 21 (20), 7767. doi:10.3390/ijms21207767
- Prasad, K. (2006). C-reactive protein (CRP)-lowering agents. *Cardiovasc Drug Rev.* 24 (1), 33–50. doi:10.1111/j.1527-3466.2006.00033.x
- Próchnicki, T., Mangan, M. S., and Latz, E. (2016). Recent insights into the molecular mechanisms of the NLRP3 inflammasome activation. *F1000Res* 5, F1000. doi:10.12688/f1000research.8614.1
- Reynolds, G. D., and Vance, R. P. (1987). C-reactive protein immunohistochemical localization in normal and atherosclerotic human aortas. *Arch. Pathol. Lab. Med.* 111 (3), 265–269.
- Ridker, P. M., Cushman, M., Stampfer, M. J., Tracy, R. P., and Hennekens, C. H. (1997). Inflammation, aspirin, and the risk of cardiovascular disease in apparently healthy men. *N. Engl. J. Med.* 336 (14), 973–979. doi:10.1056/NEJM199704033361401
- Ridker, P. M., Danielson, E., Fonseca, F. A., Genest, J., Gotto, A. M., Jr, Kastelein, J. J., et al. (2008). Rosuvastatin to prevent vascular events in men and women with elevated C-reactive protein. *N. Engl. J. Med.* 359 (21), 2195–2207. doi:10.1056/NEJMoa0807646
- Ridker, P. M., Everett, B. M., Thuren, T., MacFadyen, J. G., Chang, W. H., Ballantyne, C., et al. (2017). Antiinflammatory therapy with Canakinumab for atherosclerotic disease. *N. Engl. J. Med.* 377 (12), 1119–1131. doi:10.1056/NEJMoa1707914
- Ridker, P. M., Devalaraja, M., Baeres, F. M. M., Engelman, M. D. M., Hovingh, G. K., Ivkovic, M., et al. (2021). IL-6 inhibition with ziltivekimab in patients at high atherosclerotic risk (RESCUE): a double-blind, randomised, placebo-controlled, phase 2 trial. *Lancet* 397 (10289), 2060–2069. doi:10.1016/S0140-6736(21)00520-1
- Ridker, P. M. (2019). Anticytokine agents: targeting interleukin signaling pathways for the treatment of atherothrombosis. *Circ. Res.* 124 (3), 437–450. doi:10.1161/CIRCRESAHA.118.313129
- Ries, W., Heigl, F., Garlich, C., Sheriff, A., and Torzewski, J. (2019). Selective C-reactive protein-apheresis in patients. *Ther. Apher. Dial.* 23 (6), 570–574. doi:10.1111/1744-9987.12804
- Roux, K. H., Kilpatrick, J. M., Volanakis, J. E., and Kearney, J. F. (1983). Localization of the phosphocholine-binding sites on C-reactive protein by immunoelectron microscopy. *J. Immunol.* 131 (5), 2411–2415. doi:10.4049/jimmunol.131.5.2411
- Schönbeck, U., and Libby, P. (2004). Inflammation, immunity, and HMG-CoA reductase inhibitors: statins as antiinflammatory agents? *Circulation* 109 (2), I118–26. doi:10.1161/01.CIR.0000129505.34151.23
- Singh, P. P., Voleti, B., and Agrawal, A. (2007). A novel RBP-J kappa-dependent switch from C/EBP beta to C/EBP zeta at the C/EBP binding site on the C-reactive protein promoter. *J. Immunol.* 178 (11), 7302–7309. doi:10.4049/jimmunol.178.11.7302
- Smolen, J. S., Landewé, R. B. M., Bergstra, S. A., Kerschbaumer, A., Sepriano, A., Aletaha, D., et al. (2023). EULAR recommendations for the management of rheumatoid arthritis with synthetic and biological disease-modifying antirheumatic drugs: 2022 update. *Ann. Rheum. Dis.* 82 (1), 3–18. doi:10.1136/ard-2022-223356
- Stein, M. P., Mold, C., and Du Clos, T. W. (2000). C-reactive protein binding to murine leukocytes requires Fc gamma receptors. *J. Immunol.* 164 (3), 1514–1520. doi:10.4049/jimmunol.164.3.1514
- Szalai, A. J., McCrory, M. A., Xing, D., Hage, F. G., Miller, A., Oparil, S., et al. (2014). Inhibiting C-reactive protein for the treatment of cardiovascular disease: promising evidence from rodent models. *Mediat. Inflamm.* 2014, 353614. doi:10.1155/2014/353614
- Tanaka, T., Narazaki, M., and Kishimoto, T. (2014). IL-6 in inflammation, immunity, and disease. *Cold Spring Harb. Perspect. Biol.* 6 (10), a016295. doi:10.1101/cshperspect.a016295
- Tanaka, T., Narazaki, M., Masuda, K., and Kishimoto, T. (2016). Regulation of IL-6 in immunity and diseases. *Adv. Exp. Med. Biol.* 941, 79–88. doi:10.1007/978-94-024-0921-5\_4
- Tardif, J. C., Kouz, S., Waters, D. D., Bertrand, O. F., Diaz, R., Maggioni, A. P., et al. (2019). Efficacy and safety of low-dose colchicine after myocardial infarction. *N. Engl. J. Med.* 381 (26), 2497–2505. doi:10.1056/NEJMoa1912388
- Thiele, J. R., Habersberger, J., Braig, D., Schmidt, Y., Goerendt, K., Maurer, V., et al. (2014). Dissociation of pentameric to monomeric C-reactive protein localizes and aggravates inflammation: *in vivo* proof of a powerful proinflammatory mechanism and a new anti-inflammatory strategy. *Circulation* 130 (1), 35–50. doi:10.1161/CIRCULATIONAHA.113.007124
- Thompson, D., Pepys, M. B., and Wood, S. P. (1999). The physiological structure of human C-reactive protein and its complex with phosphocholine. *Structure* 7 (2), 169–77. doi:10.1016/S0969-2126(99)80023-9
- Torzewski, J., Torzewski, M., Bowyer, D. E., Fröhlich, M., Koenig, W., Waltenberger, J., et al. (1998). C-reactive protein frequently colocalizes with the terminal complement complex in the intima of early atherosclerotic lesions of human coronary arteries. *Arterioscler. Thromb. Vasc. Biol.* 18 (9), 1386–1392. doi:10.1161/01.atv.18.9.1386
- Torzewski, J., Brunner, P., Ries, W., Garlich, C. D., Kayser, S., Heigl, F., et al. (2022). Targeting C-reactive protein by selective apheresis in humans: pros and cons. *J. Clin. Med.* 11 (7), 1771. doi:10.3390/jcm11071771
- Torzewski, J. (2005). C-reactive protein and atherogenesis: new insights from established animal models. *Am. J. Pathol.* 167 (4), 923–925. doi:10.1016/S0002-9440(10)61182-0
- Volanakis, J. E. (2001). Human C-reactive protein: expression, structure, and function. *Mol. Immunol.* 38 (2-3), 189–197. PMID: 11532280. doi:10.1016/S0161-5890(01)00042-6
- Withering, W. (1785). *An account of the foxglove and some of its medical uses with practical remarks on dropsy and other diseases*. London, UK: GGJ and J Robinson.
- Xie, W., Huang, Y., Xiao, S., Sun, X., Fan, Y., and Zhang, Z. (2019). Impact of janus kinase inhibitors on risk of cardiovascular events in patients with rheumatoid arthritis: systematic review and meta-analysis of randomised controlled trials. *Ann. Rheum. Dis.* 78, 1048–1054. doi:10.1136/annrheumdis-2018-214846
- Yao, Y., Mao, J., Xu, S., Zhao, L., Long, L., Chen, L., et al. (2019). Rosmarinic acid inhibits nicotine induced C-reactive protein generation by inhibiting NLRP3 inflammasome activation in smooth muscle cells. *J. Cell Physiol.* 234 (2), 1758–1767. doi:10.1002/jcp.27046
- Young, D. P., Kushner, I., and Samols, D. (2008). Binding of C/EBP $\beta$  to the C-reactive protein (CRP) promoter in Hep3B cells is associated with transcription of CRP mRNA. *J. Immunol.* 181 (4), 2420–2427. doi:10.4049/jimmunol.181.4.2420
- Yu, Q., Liu, Z., Waqar, A. B., Ning, B., Yang, X., Shiomi, M., et al. (2014). Effects of antisense oligonucleotides against C-reactive protein on the development of atherosclerosis in WHHL rabbits. *Mediat. Inflamm.* 2014, 979132. doi:10.1155/2014/979132
- Zaczekiewicz, M., Kostenzer, K., Graf, M., Mayer, B., Zimmermann, O., and Torzewski, J. (2022). Cardiac glycosides lower C-reactive protein plasma levels in patients with decompensated heart failure: results from the single-center C-reactive protein-digoxin observational study (C-dos). *J. Clin. Med.* 11 (7), 1762. doi:10.3390/jcm11071762
- Zeller, J., Cheung Tung Shing, K. S., Nero, T. L., MacFadyen, J. D., Krippner, G., Bogner, B., et al. (2023). A novel phosphocholine-mimetic inhibits a pro-inflammatory conformational change in C-reactive protein. *EMBO Mol. Med.* 15, e16236. Epub 2022 Dec 5. PMID: 36468184. doi:10.15252/emmm.202216236
- Zhang, D., Sun, M., Samols, D., and Kushner, I. (1996). STAT3 participates in transcriptional activation of the C reactive protein gene by interleukin-6. *J. Biol. Chem.* 271 (16), 9503–9509. doi:10.1074/jbc.271.16.9503
- Zimmermann, O., Li, K., Zaczekiewicz, M., Graf, M., Liu, Z., and Torzewski, J. (2014). C-reactive protein in human atherosclerosis: facts and fiction. *Mediat. Inflamm.* 2014, 561428. doi:10.1155/2014/561428