Check for updates

### **OPEN ACCESS**

EDITED BY Guliz N. Guncu, Hacettepe University, Türkiye

REVIEWED BY Burcu Özdemir, Gazi University, Türkiye Dogukan Yilmaz, Sakarya University, Türkiye

\*correspondence Ying Gu

☑ ying.gu@stonybrookmedicine.edu RECEIVED 25 April 2024 ACCEPTED 01 July 2024

PUBLISHED 16 July 2024

### CITATION

Golub LM, Lee H-M, Bacigalupo J and Gu Y (2024) Host modulation therapy in periodontitis, diagnosis and treatment—status update. Front. Dent. Med 5:1423401.

doi: 10.3389/fdmed.2024.1423401

### COPYRIGHT

© 2024 Golub, Lee, Bacigalupo and Gu. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# Host modulation therapy in periodontitis, diagnosis and treatment—status update

Lorne M. Golub<sup>1</sup>, Hsi-Ming Lee<sup>1</sup>, Joseph Bacigalupo<sup>2</sup> and Ying Gu<sup>3\*</sup>

<sup>1</sup>Department of Oral Biology and Pathology, School of Dental Medicine, Stony Brook University, Stony Brook, NY, United States, <sup>2</sup>Private Practice, Hempstead, NY, United States, <sup>3</sup>Department of General Dentistry, School of Dental Medicine, Stony Brook University, Stony Brook, NY, United States

### KEYWORDS

host modulation therapy, periodontitis, diagnosis, treatment, inflammation

The microbial biofilm (dental plaque) has long been known to initiate and perpetuate the inflammatory response in periodontitis. Anaerobic micro-organisms in the biofilm (especially *Porphyromonas gingivalis, Tannerella Forsythia,* and *Prevotella intermedia*), and their release of endotoxin and other bacterial products, have long been investigated as causative agents of periodontal inflammation and connective tissue breakdown including alveolar bone loss (1-4).

However, a recent strategy to manage periodontitis, as an adjunct to controlling the microbial biofilm (involving daily optimal oral hygiene, and scaling and root planing (SRP) at regular intervals), has been called "Host-Modulation Therapy (HMT)". The concept of HMT was first introduced in the early 1980's (5); and later extensively discussed by Williams (6) and Golub in the 1990s (7). While periodontitis is initiated by specific anaerobic bacterial, the outcome of the disease is determined by the host response to the periodontal inflammation. As a result of these early discoveries, the treatment of periodontitis now includes two prongs: antimicrobial treatment (including SRP) and HMT which targets the host inflammatory responses, such as elevated levels of matrix metalloproteinases and inflammatory cytokines. One HMT approach involves the use of aspirin-like drugs or NSAIDs such as ibuprofen, flurbiprofen, etc (8).. However, the long-term use of these drugs poses risks for developing ulcers and other side-effects, and this approach is not discussed herein (8, 9).

More than 4-decades ago, Golub et al. (1983) (5) proposed, and then experimentally demonstrated, that tetracyclines, and by mechanisms unrelated to their use as broad-spectrum antibiotics, can effectively modulate the host response (10). The initial studies on experimental animals showed that minocycline can effectively reduce pathologically-excessive gingival collagenolytic activity and alveolar bone loss during diabetes in either conventional or germ-free rats (5), later studies showed more robust HMT efficacy with doxycycline (10).

Specifically, these investigators discovered that tetracyclines, unexpectedly, can inhibit the mammalian (host) cell production, and the extracellular activity, of host-generated matrix metalloproteinases (MMPs), specifically the collagenases, gelatinases, and macrophage metallo-elastase (10, 11). These proteolytic enzymes, produced by human and animal tissues, mediate the degradation and breakdown of the collagen fibers and other connective tissue constituents of the periodontium (proteoglycan ground substance, etc.) including the gingiva, the periodontal ligament, and the organic matrix of the alveolar bone (8).

This discovery ultimately led to the development of MMP-inhibitory host modulatory drugs for both dentistry (Periostat<sup>®</sup>) and medicine (Oracea<sup>®</sup>), both of which are government-approved (Food and Drug Administration; FDA) drugs requiring

prescriptions for clinical use. Periostat<sup>®</sup> can be prescribed for treating periodontitis, as an adjunct to scaling and root planing and oral hygiene instruction. (Of interest, a short communication titled "Managing Tissue Health Around Natural Teeth And Dental Implants: A Clinician's Experience" described the use of Periostat<sup>®</sup> in approximately 800 patients, which appeared to reduce the severity of periodontitis and peri-implantitis (12). Oracea<sup>®</sup> is prescribed for the treatment of the common inflammatory skin diseases, acne and rosacea with significant improvement in both diseases (10, 11, 13).

The rationale behind the development of these drugs was not obvious (i.e., efficacy of a NON-antibiotic dose of the antibiotic, doxycycline), and these regimens were demonstrated to be safe and effective in clinical studies (8). As a result, these drugs were patented, government (FDA)-approved (after extensive doubleblind, placebo-controlled clinical trials), and made commercially available to dental and medical practitioners (dermatologists have been the major prescribers). Another strategy in the development of tetracyclines as "Host-Modulators" was to chemically modify the tetracycline molecule by eliminating the side-chain responsible for its antibiotic activity, i.e., the dimethyl amino group on carbon-4 (10, 11, 13). This modification resulted in the development of a series of chemically-modified tetracyclines (CMTs), one of which, CMT-3 (6-demethy 6-deoxy 4-dedimethy amino tetracycline; Incyclinide), has been tested in patients with a type of cancer, Kaposi's sarcoma (14). In the preliminary studies, these patients were found to respond favorably, exhibiting a 44% reduction in angiogenic lesions in their skin. However, in clinical trials a significant side-effect of this novel orally-administered compound emerged, namely an increase in sunburn (14), which resulted in a cessation of further testing of this CMT. Currently, additional research continues with the goal of developing newer CMTs which: (i) lack antibiotic activity in order to prevent the side-effect of the emergence of antibiotic-resistant micro-organisms; (ii) are potent MMP-inhibitors; and (iii) exhibit few, if any, side-effects such as increased sensitivity to sunburn.

Recently, newer HMT agents have been developed and studied as adjunctive treatments for periodontitis. Curcumin, a natural spice used widely in India cuisine, has been shown to exhibit antiinflammatory properties. However, a significant deficit is its limited bioavailability. To overcome this problem, a novel group of chemically modified curcumins (CMCs) has been developed to increase its bioavailability and to enhance its anti-inflammatory activity (15). Research studies, involving in vitro enzyme assays, cell culture studies, and in vivo animal experiments, identified a lead compound, namely CMC 2.24, a phenylamino carbonyl curcumin. This novel compound, CMC 2.24, has demonstrated a significant effect on reducing pathologic elevated MMP levels, and reducing alveolar bone loss in animal models of periodontitis (16, 17). Moreover, CMC 2.24 can reduce the elevated MMP-9 levels (a biomarker for inflammatory periodontal and other diseases) in gingiva and plasma, before any visible clinical signs of periodontitis are observed (18). While investigating possible underlying mechanisms, in addition to its calcium and zinc binding  $\beta$ -diketone moiety that is similar to tetracyclines, CMC has shown a resolving-like activity to promote M2 macrophage polarization, therefore attenuating pathologic alveolar bone loss (19). These findings strongly suggest that CMCs can be developed as novel therapeutic HMT agents for treating periodontitis and other diseases, including arthritis, cardiovascular disease, and cancer.

As the concept of HMT has become widely accepted in the field, emerging HMTs have been studied as adjunctive therapies for periodontal diseases. In addition to CMTs and CMCs mentioned earlier, other potential host modulation therapies have been developed and studied by other groups. For example, anti-cytokine therapy utilizes monoclonal antibodies against inflammatory cytokines. Infliximab and Etanercept were developed as anti-TNF-a drugs and showed significant improvement in periodontal disease parameters (9). Studies have also demonstrated that probiotics can be a potential HMT in the management of periodontitis. Lactobacillus based probiotics revealed therapeutic effects in improving periodontal conditions (9, 20). Omega-3 polyunsaturated fatty acids and resolvins are another group of HMTs that have been studied extensively by Van Dyke et al. (21). These compounds do not inhibit acute inflammation, which is required for normal wound healing, but do prevent chronicity which would negatively affect wound healing (21). In animal and human clinical studies, Resolvins demonstrated great potential to reduce the severity of periodontitis (22, 23). As more innovations in HMT research emerge, a variety of potential HMT agents are being developed including stem cells (24), and complement inhibitors (25). Research has demonstrated that human periodontal stem cells can modulate responses (24), and bone marrow-derived inflammatory mesenchymal stem cells have shown great potential for periodontal regeneration in animal models (26). HMT targeting the complement C3 has been shown to reduce periodontal inflammation suggesting a therapeutic potential in treating periimplantitis (25).

With emerging development of additional host modulation agents, as adjunctive therapy to existing periodontal approaches, plus the new diagnostic concepts developed by Sorsa et al, ie, the aMMP-8 chairside test for gingival crevicular fluid and periimplantitis sulcular fluid (27), HMT can greatly contribute to long-term improved periodontal outcomes.

# Author contributions

LG: Writing – original draft, Writing – review & editing. H-ML: Writing – review & editing. JB: Writing – review & editing. YG: Writing – original draft, Writing – review & editing.

# Funding

The author(s) declare that no financial support was received for the research, authorship, and/or publication of this article. However, a number of pervious NIDCR-funded grants were received for earlier research.

# Conflict of interest

LG is listed as an inventor on several related patents (Periostat, Oracea, and CMC 2.24) and these have been fully assigned to his institution, Stony Brook University, The State University of New York (SUNY).

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## References

1. Socransky SS, Haffajee AD, Cugini MA, Smith C, Jr KR. Microbial complexes in subgingival plaque. *J Clin Periodontol.* (1998) 25(2):134–44. doi: 10.1111/j.1600-051X. 1998.tb02419.x

2. Gibbons RJ, Van Houte J. On the formation of dental plaques. J Perio. (1973) 44 (6):347–60.

3. Tanner AC, Haffer C, Bratthall GT, Visconti RA, Socransky SS. A study of the bacteria associated with advancing periodontitis in man. *J Clin Periodontol.* (1979) 6:278–307. doi: 10.1111/j.1600-051X.1979.tb01931.x

4. Slots J. Subgingival microflora and periodontal disease. J Clin Periodontol. (1979) 6(5):351-82. doi: 10.1111/j.1600-051X.1979.tb01935.x

5. Golub LM, Lee HM, Lehrer G, Nemiroff A, McNamara TF, Kaplan R, et al. Minocycline reduces gingival collagenolytic activity during diabetes. Preliminary observations and a proposed new mechanism of action. *J Periodontal Res.* (1983) 18 (5):516–26. doi: 10.1111/j.1600-0765.1983.tb00388.x

 Williams RC. Periodontal disease. N Engl J Med. (1990) 322(6):373. doi: 10.1056/ NEJM199002083220606

7. Golub LM, Suomalainen K, Sorsa T. Host modulation with tetracyclines and their chemically modified analogues. *Curr Opin Dent.* (1992) 2:80–90.

8. Golub LM, Lee HM. Periodontal therapeutics: current host-modulation agents and future directions. *Periodontol 2000.* (2020) 82(1):186–204. doi: 10.1111/prd.12315

9. Balta MG, Papathanasiou E, Blix IJ. And van dyke TE: host modulation and treatment of periodontal disease. *J Dent Res.* (2021) 100(8):798-809. doi: 10.1177/0022034521995157

10. Golub LM, Lee HM, Ryan ME, Giannobile WV, Payne J, Sorsa T. Tetracyclines inhibit connective tissue breakdown by multiple non-antimicrobial mechanisms. *Adv Dent Res.* (1998) 12(2):12–26. doi: 10.1177/08959374980120010501

11. Golub LM, Ramamurthy NS, McNamara TF, Greenwald RA, Rifkin BR. Tetracyclines inhibit connective tissue breakdown: new therapeutic implications for an old family of drugs. *Crit Rev Oral Biol Med.* (1991) 2(3):297–321. doi: 10.1177/10454411910020030201

12. Bacigalupo J, Golub LM, Sorsa T, Lee H-M, Gu Y, Walker SG. Managing tissue health around natural teeth and dental implants: a clinician's experience. *J Comm Med and Pub Health Rep.* (2024) 5(01). doi: 10.38207/JCMPHR/2024/JAN05010305

13. Sapadin AN, Fleischmajer R. Tetracyclines: nonantibiotic properties and their clinical implications. *J Am Acad Dermatol.* (2006) 54(2):258–65. doi: 10.1016/j.jaad. 2005.10.004

14. Dezube BJ, Krown SE, Lee JY, Bauer KS, Aboulafia DM. Randomized phase II trial of matrix metalloproteinase inhibitor COL-3 in AIDS-related Kaposi's sarcoma: an AIDS malignancy consortium study. *J Clin Oncol.* (2006) 24(9):1389–94. doi: 10. 1200/JCO.2005.04.2614

## Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

15. Zhang Y, Gu Y, Lee HM, Hambarjieva E, Vrankova K, Golub LM, et al. Design, synthesis and biological activity of new polyenolic inhibitors of matrix metalloproteinases: a focus on chemically-modified curcumins. *Curr Med Chem.* (2012) 19:4348–58. doi: 10.2174/092986712802884295

16. Wang H, Lee HM, Raja V, Iacono VJ, Scaduto J, Johnson F, et al. Enhanced efficacy of chemically modified curcumin in experimental periodontitis: systemic implications. *J Exp Pharmacol.* (2019) 11:1–14. doi: 10.2147/JEP.S171119

17. Deng J, Golub LM, Lee HM, Lin M, Bhatt H, Hong H, et al. Chemically-Modified curcumin 2.24: a novel systemic therapy for natural periodontitis in dogs. *J Exp Pharmacol.* (2020) 12:47–60. doi: 10.2147/JEP.S236792

18. Deng J, Golub LM, Lee HM, Bhatt H, Hong H, Johnson F, et al. A novel chemically-modified curcumin 2.24: short-term systemic treatment for natural periodontitis in dogs. *Front Dent. Med.* (2021) 2:609795. doi: 10.3389/fdmed.2021. 609795

19. Deng J, Golub LM, Lee HM, Bhatt H, Johnson F, Xu TM, et al. A novel modified-curcumin 2.24 resolves inflammation by promoting M2 macrophage polarization. *Sci Rep.* (2023) 13:15513. doi: 10.1038/s41598-023-42848-x

20. Hajishengallis G, Chavakis T. Lambris JD: current understanding of periodontal disease pathogenesis and targets for host-modulation therapy. *Periodontol 2000.* (2020) 84(1):14–34. doi: 10.1111/prd.12331

21. Van Dyke TE. Shifting the paradigm from inhibitors of inflammation to resolvers of inflammation in periodontitis. *J Periodontol.* (2020) 91(Suppl 1):S19–25. doi: 10.1002/JPER.20-0088

22. Van Dyke TE, Hasturk H, Kantarci A, Freire MO, Nguyen D, Dalli J, et al. Proresolving nanomedicines activate bone regeneration in periodontitis. *J Dent Res.* (2015) 94(1):148–56. doi: 10.1177/0022034514557331

23. Preshaw PM. Host modulation therapy with anti-inflammatory agents. Periodontol 2000. (2018) 76(1):131-49. doi: 10.1111/prd.12148

24. Cianci E, Recchiuti A, Trubiani O, Diomede F, Marchisio M, Miscia S, et al. Human periodontal stem cells release specialized proresolving mediators and carry immunomodulatory and prohealing properties regulated by lipoxins. *Stem Cells Transl Med.* (2016) 5(1):20–32. doi: 10.5966/sctm.2015-0163

25. Kajikawa T, Mastellos DC, Hasturk H, Kotsakis GA, Yancopoulou D, Lambris JD, et al. C3-targeted host-modulation approaches to oral inflammatory conditions. *Semin Immunol.* (2022) 59:101608. doi: 10.1016/j.smim.2022.101608

26. Kim SH, Seo BM, Choung PH, Lee YM. Adult stem cell therapy for periodontal disease. *Int J Stem Cells.* (2010) 3(1):16–21. doi: 10.15283/ijsc.2010.3.1.16

27. Sorsa T, Nwhator SO, Sakellari D, Grigoriadis A, Umeizudike KA, Brandt E, et al. aMMP-8 oral fluid PoC test in relation to oral and systemic diseases. *Front Oral Health.* (2022) 3:897115. doi: 10.3389/froh.2022.897115