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Investigating the association between gut microbiome and aortic aneurysm diseases: a bidirectional two-sample Mendelian randomization analysis

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Objective: This study aims to investigate the associations between specific bacterial taxa of the gut microbiome and the development of aortic aneurysm diseases, utilizing Mendelian Randomization (MR) to explore these associations and overcome the confounding factors commonly present in observational studies.

Methods: Employing the largest available gut microbiome and aortic aneurysm Genome-Wide Association Study databases, including MiBioGen, Dutch Microbiome Project, FinnGen, UK Biobank, and Michigan Genomics Initiative, this study performs two-sample bidirectional MR analyses. Instrumental variables, linked to microbiome taxa at significant levels, were selected for identifying relationships with abdominal aortic aneurysms (AAA), thoracic aortic aneurysms (TAA), and aortic dissection (AD). Methods like inverse variance weighted, MR-PRESSO, MR-Egger, weighted median, simple mode, and mode-based estimate were used for MR analysis. Heterogeneity was assessed with the Cochran Q test. MR-Egger regression and MR-PRESSO addressed potential unbalanced horizontal pleiotropy.

Results: The analysis did not find any evidence of statistically significant associations between the gut microbiome and aortic aneurysm diseases after

Abbreviations: AAA, Abdominal Aortic Aneurysm; AD, Aortic Dissection; CHIP, The Cardiovascular Health Improvement Project; CI, Confidence Interval; DMP, Dutch Microbiome Project; ECM, Extracellular Matrix; FDR, False Discovery Rate; GM, Gut Microbiome; GWAS, Genome-Wide Association Study; IV, Instrumental Variable; IVW, Inverse Variance Weighted; LD, Linkage Disequilibrium; MGI, Michigan Genomics Initiative; MMP, Matrix Metalloproteinase; MR, Mendelian Randomization; NF-κB, Nuclear Factor kappa-light-chain-enhancer of activated B cells; OR, Odds Ratio; RCT, Randomized Controlled Trial; SNP, Single Nucleotide Polymorphism; TAA, Thoracic Aortic Aneurysm; TMAO, Trimethylamine N-oxide; UK Biobank, United Kingdom Biobank; VSMC, Vascular Smooth Muscle Cell.

adjusting for the false discovery rate (FDR). Specifically, while initial results suggested correlations between 19 taxa and AAA, 25 taxa and TAA, and 13 taxa with AD, these suggested associations did not hold statistical significance post-FDR correction. Therefore, the role of individual gut microbial taxa as independent factors in the development and progression of aortic aneurysm diseases remains inconclusive. This finding underscores the necessity for larger sample sizes and more comprehensive studies to further investigate these potential links.

Conclusion: The study emphasizes the complex relationship between the gut microbiome and aortic aneurysm diseases. Although no statistically significant associations were found after FDR correction, the findings provide valuable insights and highlight the importance of considering gut microbiota in aortic aneurysm diseases research. Understanding these interactions may eventually contribute to identifying new therapeutic and preventive strategies for aortic aneurysm diseases.

KEYWORDS

abdominal aortic aneurysm, thoracic aortic aneurysm, aortic dissection, gut microbiome, Mendelian randomization

1 Introduction

Aortic aneurysms and dissections are life-threatening conditions representing serious diseases of the aorta. Aortic aneurysms, including thoracic and abdominal variants, mainly occur due to chronic dilative changes caused by weakening of the aortic wall, with rupture of the aneurysms being a leading cause of death (Johnston et al., 1991). Aortic dissection (AD) occurs when there is an intimal tear that allows blood to flow between the layers of the aortic wall, splitting the intima longitudinally and creating a dissection flap that divides the true lumen from a newly formed false lumen (Isselbacher et al., 2022). Aortic aneurysm diseases are the 15th most common cause of death in individuals over 55 years of age and also impose significant economic burdens on patients' families and society (Mcclure et al., 2020; Bossone and Eagle, 2021). Currently, the etiology of aortic aneurysm diseases is not fully understood, making it essential to find effective ways to prevent and reduce the incidence of these conditions.

Historically, it was presumed that dietary components were solely absorbed and metabolized by human cells within the intestine. However, recent insights reveal that the gut microbiome (GM), comprising trillions of commensal organisms, acts as a metabolically active organ akin to an endocrine system. This complex microbial community significantly impacts host physiological functions, predominantly through the production of biologically active metabolites. It plays a crucial role in vitamin synthesis and the modulation of key mechanisms such as inflammation, immune response, and oxidative stress mitigation (Tang et al., 2019; Witkowski et al., 2020). Imbalances in the GM can influence the occurrence and development of various diseases, including aortic aneurysms and dissections.

Multicentric clinical studies have identified an association between imbalances in the GM and the development of aortic aneurysms and dissections (Nakayama et al., 2022; Ito et al., 2023; Jiang et al., 2023; Manabe et al., 2023). Experimental studies have shown that imbalances in the GM can affect the occurrence of aneurysms (Shinohara et al., 2022), and gut microbial metabolites such as Trimethylamine N-oxide (TMAO) can promote the occurrence of aneurysms (Benson et al., 2023), while metabolites like butyrate and Indole-3-aldehyde can inhibit the development of aortic aneurysms and dissections (Tian et al., 2022; Huang et al., 2023). These studies objectively demonstrate the significant role of the GM and its metabolic products in the disease progression of aortic aneurysms and dissections, with the pathological mechanisms primarily based on the direct damage to the aortic wall by gut microbes and the indirect effects of inflammation mediated by microbial imbalances. However, observational studies struggle to establish clear causal and reverse causal relationships, and although randomized controlled trials (RCTs) are considered the gold standard for establishing causality, their implementation can sometimes be hindered by ethical, practical, or financial obstacles.

Mendelian randomization (MR) analysis is an epidemiological design that can strengthen causal inference by using genetic variants as instrumental variables (IVs) for an exposure (Davies et al., 2018). The MR design, characterized by its immunity to confounding biases and reverse causality, owes its robustness to the random assortment of genetic alleles at conception and their insusceptibility to

modification by diseases (Pingault et al., 2018). Consequently, this study seeks to elucidate the potential causal associations between individual bacterial taxa and aortic aneurysm diseases through bidirectional MR analyses. This methodology has the potential to lay down a theoretical foundation and open up new avenues for the prevention and treatment of aortic aneurysm diseases.

2 Methods

2.1 Study design

Figure 1 illustrates the diagram of our study design, highlighting that the causal interpretation of MR estimates relies on three critical assumptions: (1) the IVs, which are single nucleotide polymorphisms (SNPs), must be associated with the exposures of interest; (2) the IVs must not be related to any confounders of the exposure-outcome relationship; (3) the association between the IVs and the outcome must be exclusively through the exposure of interest (Lawlor, 2016). This study conducts bidirectional Mendelian randomization analyses between the GM and aortic aneurysm diseases, using GM Genome-Wide Association Study (GWAS) data based on 16S rRNA sequencing methods for primary analysis and GM GWAS data based on Metagenomic Sequencing (MGS) methods for secondary analysis. Additionally, other GWAS datasets for abdominal aortic aneurysms (AAA) and thoracic aortic aneurysms (TAA) serve as validation sets for the positive findings identified.

2.2 Data sources

The characteristics of corresponding GWAS data sources are detailed in Table 1. Summary data on genes from the MiBioGen consortium, comprising participants of diverse ancestries from 24 cohorts totaling 18,340 individuals, represent the largest whole-genome meta-analysis to date. The genetic sequencing method was 16S rRNA, with 78% of participants being of European descent, comprising 211 taxonomic units across various classifications (Kurilshikov et al., 2021) (Supplementary Additional File 1; Order 1). We selected summary data from 7,738 individuals of European ancestry from the Dutch Microbiome Project (DMP) for the largest species-level GM dataset, sequenced using MGS, featuring 207 taxonomic units across different classifications (Lopera-Maya et al., 2022) (Supplementary Additional File 1; Order 2).

GWAS data for AAA, TAA, and AD were obtained from FinnGen Release 10, which maps genotype-phenotype correlations using Finnish biobank data. This includes genome and health data from 3,869 AAA, 3,880 TAA, 967 AD individuals and 381,977 controls. Diagnoses for the following conditions were based on hospital records and classified according to the respective International Classification of Diseases (ICD) codes: AAA: ICD-10: I71.3, I71.4, ICD-9: 4413A, 4414A, ICD-8: 44120; TAA: ICD-10: I71.01, I71.1, I71.2, Q25.43; AD: ICD-10: I71.00, I71.01, I71.09, ICD-9: 4410. Endpoint and control definitions are detailed on the FinnGen website (https://www.finngen.fi/en/researchers/clinicalendpoints). Validation datasets for AAA were sourced from the UK Biobank database, including 1,306 cases and 408,565 controls, and TAA datasets were obtained from The Cardiovascular Health



FIGURE 1

Schematic representation of the Mendelian randomization design. The traditional assumptions of Mendelian randomization are that the genetic instrumental variable is associated with the risk factor (assumption 1); the variants should not be associated with confounders (assumption 2); and that the variants should influence the outcome only through effects on the risk factor under investigation (assumption 3). SNP, single nucleotide polymorphism; LD, linkage disequilibrium; *P*, probability value; r^2 , coefficient of determination; MGI, the Michigan genomics initiative.

	Trait	Sample size	Ancestry	Dataset source	Year	Web source/PMID
Exposure	Gut microbiota	18 340 individuals	Mixed (78% European)	MiBioGen	2021	PMID: 33462485
	Gut microbiota	7 738 individuals	European	Dutch Microbiome Project	2022	PMID: 35115690
Outcome	Abdominal Aortic Aneurysm	3 869 cases and 381 977 controls	European	FinnGen	2023	https://www.finngen.fi/fi
	Thoracic Aortic Aneurysm	3 880 cases and 381 977 controls	European	FinnGen	2023	https://www.finngen.fi/fi
	Aortic Dissection	967 cases and 381 977 controls	European	FinnGen	2023	https://www.finngen.fi/fi
	Abdominal Aortic Aneurysm	1 306 cases and 408 565 controls	European	UK biobank	2023	https:// pan.ukbb.broadinstitute.org/
	Thoracic Aortic Aneurysm	1 351 cases and 18 295 controls	European	CHIP,MGI	2021	PMID: 34265237

CHIP, The Cardiovascular Health Improvement Project.

MGI, The Michigan Genomics Initiative.

Improvement Project (CHIP) and The Michigan Genomics Initiative (MGI), including 1,351 cases and 18,295 controls (Roychowdhury et al., 2021).

2.3 Instrumental variable

For MR analysis, it's crucial that the genetic variants used are representative of microbiome features. The selection criteria for IVs included: (1) SNPs associated with each genus at the locus-wide significance threshold ($P < 1.0 \times 10^{-5}$) were chosen as potential IVs (Sanna et al., 2019); (2) 1000 Genomes Project European samples data were referenced to calculate the linkage disequilibrium (LD) between SNPs, retaining only those with the lowest *P*-values and an $R^2 < 0.001$ (Sudmant et al., 2015); (3) SNPs with an F-statistic < 10 were excluded to avoid weak instrumental bias (Burgess and Thompson, 2011).

2.4 Genetic analyses to elucidate causality

Bidirectional MR analyses were initially conducted to explore the causal relationship between the GM and aortic aneurysm diseases. The main method for causal estimation was the Wald method when only one SNP tool was available. When multiple valid instrumental variables were available, the inverse-variance weighted method (IVW) was employed for maximum efficiency. The IVW method meta-analyzed SNP-specific Wald estimates using random effects to obtain the final estimate of the causal effect (Burgess et al., 2019), which was reported in beta (β) value with standard error for continuous outcomes and odds ratio (OR) with a 95% confidence interval (CI) for binary outcomes; *P*-values < 0.05 were considered nominally significant. Results were adjusted for the False Discovery Rate (FDR) to account for multiple testing.

2.5 Sensitivity analyses

Methods such as MR-Presso, MR-Egger, weighted median, simple mode, and Mode-based estimate method were utilized for sensitivity analysis (Bowden et al., 2015; Hemani et al., 2018). Heterogeneity was assessed using the Cochran Q test, with a Pvalue less than 0.05 indicating significant heterogeneity. A Q value substantially exceeding its degrees of freedom indicates evidence of heterogeneity and suggests the presence of invalid instruments (Greco et al., 2015). MR-Egger regression and MR-Presso were utilized to analyze potential unbalanced horizontal pleiotropy, combining the Wald ratio into a meta-regression with an intercept and slope parameter. This approach estimates the causal effect while adjusting for any directional pleiotropy (Bowden et al., 2015). All statistical analyses were performed using R platform (version 4.3.1). MR analyses were performed using the packages including TwoSampleMR (version 0.5.7) (Hemani et al., 2018), MRPRESSO (version 1.0) (Verbanck et al., 2018), vroom (version 1.6.5), dplyr (version 1.1.4), and others.

2.6 Ethical approval and consent to participate

This study was based on publicly available data. Each GWAS within the study received approval from the relevant Institutional Review Board, and informed consent was secured from all participants or their respective legal representatives.

3 Results

For the 211 GM taxa from the MiBioGen consortium, 2,774 SNPs were used as IVs. For the 207 GM taxa from the DMP, 1,961 SNPs served as IVs. From the FinnGen database, 65, 56, and 24 SNPs were

used as IVs for AAA, TAA, and AD, respectively. In the case of AAA data from the UK Biobank, 33 SNPs were utilized as IVs, and for TAA data from the MGI, 26 SNPs served as IVs. All SNPs included in our analysis had an F-statistic greater than 16, indicating that all are robust instruments (Supplementary Additional File 2).

None of the associations showed statistical significance after FDR correction (Supplementary Additional File 3). Specifically, while initial results suggested correlations between 19 taxa and AAA, 25 taxa and TAA, and 13 taxa with AD, these suggested associations did not hold statistical significance post-FDR correction (Figure 2, Supplementary Figures 1–3).All suggestive positive results are presented with visual information through "leave-one-out" sensitivity analysis, forest plots, scatter plots, and funnel plots (Supplementary Additional File 5).

outcomes	Exposures	OK(95%CI)		F
AAA	C_Lentisphaeria (16S)	0.81(0.68 - 0.97)		0.020
	C_Verrucomicrobiae (16S)	0.73(0.55 - 0.96)		0.026
	O_Verrucomicrobiales (16S)	0.73(0.55 - 0.96)		0.026
	O_Victivallales (16S)	0.81(0.68 - 0.97)		0.020
	F_Verrucomicrobiaceae (16S)	0.73(0.55 - 0.96)		0.026
	F_Family XIII (16S)	0.72(0.53 - 0.98)		0.036
	F_Lactobacillaceae (MGS)	0.89(0.81 - 0.98)	HER	0.015
	G_Catenibacterium (16S)	1.34(1.09 - 1.64)		 0.006
	G_Akkermansia (16S)	0.73(0.55 - 0.96)		0.026
	G_Oxalobacter (16S)	1.15(1.00 - 1.32)		 0.043
	G_Clostridium sensu stricto 1 (16S)	1.34(1.02 - 1.77)		 0.037
	G_Blautia (16S)	0.46(0.23 - 0.90)		0.023
	G_Anaerotruncus (16S)	0.73(0.55 - 0.97)		0.032
	G_Bilophila (16S)	1.33(1.06 - 1.68)		 0.014
	G_Pseudoflavonifractor (MGS)	0.80(0.67 - 0.94)		0.008
	G_Lactobacillus (MGS)	0.89(0.81 - 0.99)	Here a	0.025
	S_Bacteroides intestinalis (MGS)	1.43(1.17 - 1.75)		 <0.001
	S_Streptococcus parasanguinis (MGS)	0.80(0.69 - 0.92)	Here a	0.002
	S_Paraprevotella clara (MGS)	1.17(1.03 - 1.33)		 0.015
ТАА	P_Lentisphaerae (16S)	0.81(0.69 - 0.94)		0.006
	C_Lentisphaeria (16S)	0.84(0.71 - 0.99)		0.038
	C Actinobacteria (16S)	0.77(0.63 - 0.95)		0.013
	O_Bifidobacteriales (16S)	0.78(0.62 - 0.97)		0.023
	O Victivallale (16S)	0.84(0.71 - 0.99)		0.038
	F Bifidobacteriaceae (16S)	0.78(0.62 - 0.97)		0.023
	F Oxalobacteraceae (16S)	1.15(1.02 - 1.31)		 0.028
	F Lachnospiraceae (MGS)	0.78(0.63 - 0.97)		0.026
	F Lactobacillacea (MGS)	0.90(0.83 - 0.98)	HH	0.011
	F Micrococcaceae (MGS)	1.13(1.00 - 1.27)		0.045
	G Fusicatenibacter (16S)	1.39(1.10 - 1.74)		 0.005
	G Turicibacter (16S)	1.35(1.10 - 1.66)		 0.005
	G Ruminococcaceae UCG003 (16S)	0.80(0.64 - 0.99)	-	0.044
	G Lactobacillus (16S)	0.82(0.67 - 0.99)	-	0.041
	G Intestinibacter (16S)	0.76(0.63 - 0.92)	-	0.004
	G Ruminococcaceae UCG014 (16S)	1.25(1.00 - 1.55)		 0.049
	G Family XIII UCG001 (16S)	1.39(1.01 - 1.92)		 0.044
	G Bilophila (MGS)	1.29(1.06 - 1.58)		 0.012
	G Lactobacillus (MGS)	0.89(0.82 - 0.97)	HH	0.008
	G Rothia (MGS)	1.13(1.00 - 1.27)		0.045
	S Lachnospiraceae bacterium 5 1 63FAA (MGS	0.91(0.82 - 1.00)		0.043
	S Ruminococcus obeum (MGS)	1.19(1.04 - 1.38)		 0.015
	S Bacteroides stercoris (MGS)	1.27(1.06 - 1.53)		 0.011
	S Bilophila unclassified (MGS)	1.19(1.01 - 1.42)		 0.042
	S Subdoligranulum unclassified (MGS)	1.19(1.01 - 1.41)		 0.036
	S Parabacteroides merdae (MGS)	1.26(1.02 - 1.57)		 0.036
AD	P Lentisphaerae (16S)	0.60(0.42 - 0.87)		0.007
	C Lentisphaeria (16S)	0.72(0.52 - 1.00)	-	0.050
	C Actinobacteria (16S)	0.64(0.43 - 0.96)	-	0.030
	O Victivallales (16S)	0.72(0.52 - 1.00)	-	0.050
	E Oscillospiraceae (MGS)	0.61(0.38 - 0.98)	-	0.039
	G Collinsella (16S)	0.53(0.30 - 0.95)		0.034
	G Ruminococcus gnavus group (16S)	0.67(0.46 - 0.98)	-	0.040
	G Oscillibacter (MGS)	0.59(0.38 - 0.93)	-	0.022
	G. Odoribacter (MGS)	0.66(0.46 - 0.95)		0.025
	S Lachnospiraceae bacterium 8 1 57EAA (MCS	() 1 20(1 04 - 1 38)		 0.010
	S Coprococcus catus (MGS)	0.65(0.44 - 0.96)		0.032
	S. Clostridium lentum (MGS)	0.63(0.48 - 0.96)		0.001
		0.00(0.40 - 0.03)		5.001
	S Alistines shahii (MGS)	0 64(0 42 - 1 00)		0.048

FIGURE 2

The association between Gut Microbiota and Aortic Aneurysm Disease using the inverse variance weighted method. OR, odds ratio; CI, confidence interval; *P*, probability value; AAA, Abdominal Aortic Aneurysm; TAA, Thoracic Aortic Aneurysm; AD, Aortic Dissection; 16S rRNA, 16S ribosomal RNA; MGS, Metagenomic Species.

Further MR analysis on the AAA and TAA validation groups revealed that replicating suggestive associations with microbiota categories in the validation sets was challenging. We evaluated whether the direction of effect for relevant microbiota in the validation sets was consistent with that in the initial groups, based on their OR levels. Results demonstrated that nine microbiota categories maintained consistency with AAA and ten microbiota categories maintained consistency with TAA (Supplementary Figures 1–3).

Cochran's IVW Q test results showed no significant heterogeneity among these IVs (Supplementary Additional File 4: Order 1). Additionally, MR-PRESSO analysis indicated that there was pleiotropy in the relationship between Ruminococcaceae UCG003 and TAA (Supplementary Additional File 4: Order 3), but further analysis found no significant directional horizontal pleiotropy according to the MR-Egger regression intercept analysis (Supplementary Additional File 4: Order 2). Reverse MR analyses revealed bidirectional causal relationships between 17, 8, and 27 microbiota categories with AAA, TAA, and AD, respectively (Figure 3, Supplementary Additional File 3: Order 11-20), among which, Blautia showed a bidirectional causal relationship with AAA (Supplementary Additional File 3: Order 11).

4 Discussion

In this large-scale MR study, none of the identified associations between gut microbiota and aortic aneurysm diseases showed statistical significance after FDR correction. Despite this, the initial analysis combining information from two GM GWAS databases suggested potential associations between 19 taxa and AAA, 25 taxa and TAA, and 13 taxa and AD. Furthermore, the abundances of 16, 8, and 26 taxa were respectively affected by AAA, TAA, and AD.

In this MR study, we did not find significant associations between the GM and aortic aneurysm diseases after FDR correction, and several factors may contribute to this negative result. Aortic aneurysm diseases are relatively rare, leading to an insufficient sample size and thus inadequate statistical power to detect potential significant associations. As the sample size in GWAS databases related to aortic aneurysms increases in the future, the likelihood of identifying significant associations may also increase. The GM generally exerts its effects through alterations in microbial communities and microbially derived metabolites. These factors are influenced by the complex dynamics of microbial interactions and are particularly susceptible to modulation by diet, lifestyle, and environmental factors. For example, carnitine from red meat and phosphatidylcholine from egg yolks are metabolized by gut bacteria into trimethylamine, which is then converted into TMAO in the liver, a known contributor to various vascular diseases (Wang et al., 2011; Koeth et al., 2013). These confounding factors, along with the impact of host health on microbial function, may affect the pathogenic or protective roles of individual gut microbes as independent factors, resulting in low statistical significance in the analyses.

Despite the negative results after FDR correction, exploring the mechanisms behind the positive trends observed in the MR analysis of GM and aortic aneurysm diseases remains valuable. The pathogenesis of aortic aneurysmal diseases remains a subject of debate. Typical pathological changes include the degradation of the extracellular matrix, endothelial damage, inflammatory responses, and oxidative stress (Sakalihasan et al., 2005; Gao et al., 2023). GM and their secretions can influence the occurrence of aortic aneurysmal diseases through various mechanisms. As mentioned earlier, microbes can exert effects through their derived metabolites. Firstly, the TMAO pathway: TMAO increases PERK-mediated unfolded protein response, endoplasmic reticulum stress, and the expression of cleaved caspase-3, leading to increased apoptosis of vascular smooth muscle cells (VSMCs) (Benson et al., 2023). TMAO can also upregulate the expression of matrix metalloproteinase-2 (MMP-2) in aortic VSMCs, mediating extracellular matrix (ECM) degradation (Benson et al., 2023). Additionally, TMAO can induce vascular inflammation through various mechanisms, such as inducing mitochondrial reactive oxygen species production, activating the NLRP3 inflammasome, stimulating the NF-KB pathway, and releasing downstream pro-inflammatory cytokines (Liu and Dai, 2020). In this study, Akkermansia from the phylum Verrucomicrobia and Lactobacillus from the phylum Firmicutes were showed suggestive negatively associations with the disease, while Paraprevotella clara under the phylum Bacteroidetes, Bilophila from the phylum Proteobacteria, Ruminococcaceae UCG014 and Subdoligranulum from the family Ruminococcaceae, and Fusicatenibacter from the family Lachnospiraceae were showed suggestive positively associations with the disease. Research has shown corresponding abundance changes in these taxa in aortic aneurysms (Xie et al., 2020; Liu et al., 2022; Xiao et al., 2023) and they can influence disease development through the TMAO pathway (Liu et al., 2019; Fu et al., 2020; Liu et al., 2020; Franck et al., 2022; Luo et al., 2022; Gan et al., 2023; Jiang et al., 2023). Secondly, the butyrate pathway: butyrate can exert protective effects by maintaining the integrity of the gut barrier, inhibiting neutrophil infiltration and the formation of neutrophil extracellular traps, and downregulating the expression of MMP-2 and MMP-9 in the aortic wall, thereby reducing ECM elastin fiber degradation (Peng et al., 2009; Tian et al., 2022). Thirdly, the tryptophan pathway: Tryptophan, primarily secreted by Lactobacillus, and its metabolic product 3-IAId exert beneficial effects by inhibiting the phenotype transition of VSMCs from a contractile to a synthetic state. It also mitigates extracellular matrix degradation, attenuates macrophage infiltration, and suppresses the expression of inflammatory cytokines, collectively contributing to the attenuation of AD development (Huang et al., 2023). In this study, Lactobacillus from the phylum Firmicutes was showed suggestive negatively associations with AAA and TAA. Clinical studies have found that, compared to patients with atherosclerosis, AAA patients have a lower relative abundance of Firmicutes (Ji et al., 2021). Clinical trials have shown that taking Lactobacillus improved vascular endothelial function and decreased systemic inflammation in men (Malik et al., 2018). Additionally, GM can exert effects through direct colonization or by influencing the distribution of other microbiota. Bacteroides can directly invade and colonize in humans, promoting the occurrence of AAA diseases (Matsuoka et al., 2021). Akkermansia inhibited the formation of AAA by restoring gut microbiota diversity, altering the expression of peripheral immune factors, and the functions of E. coli, Clostridium, and Lactobacillus (He et al., 2022). Some taxa showing positive trends in this study have unclear mechanisms. For example,

Exposures	Outcomes	Beta(95%CI)		Р
AAA	C_Alphaproteobacteria(16S)	0.05(0.01 - 0.09)		0.010
	O_Rhodospirillales(16S)	0.06(0.02 - 0.10)		0.004
	O_Pasteurellales(MGS)	0.08(0.00 - 0.15)		0.046
	F_Rhodospirillaceae(16S)	0.06(0.02 - 0.10)		0.004
	F_Pasteurellaceae(MGS)	0.08(0.00 - 0.15)		0.046
	F_Veillonellaceae(MGS)	0.06(0.01 - 0.11)		0.029
	G_Desulfovibrio(16S)	-0.05(-0.090.01)		0.017
	G_Lachnospiraceae FCS020 group(16S)	-0.03(-0.060.00)		0.037
	G_Ruminococcaceae UCG011(16S)	-0.08(-0.140.02)		0.009
	G_Terrisporobacter(16S)	-0.05(-0.100.01)		0.022
	G_Blautia(16S)	-0.10(-0.200.01)		0.033
	G_Desulfovibrio(MGS)	0.10(0.03 - 0.17)		0.004
	G_Roseburia(MGS)	-0.04(-0.090.00)		0.034
	G Adlercreutzia(MGS)	-0.07(-0.130.01)		0.028
	S Haemophilus parainfluenzae(MGS)	0.08(0.00 - 0.15)		0.049
	S_Desulfovibrio_piger(MGS)	0.13(0.06 - 0.21)	· · · · · · · · · · · · · · · · · · ·	< 0.001
	S Adlercreutzia equolifaciens(MGS)	-0.07(-0.130.01)		0.029
ТАА	G Dialister(16S)	-0.05(-0.080.01)		0.018
	G Roseburia(16S)	0.03(0.00 - 0.05)		0.044
	G Prevotella7(16S)	-0 10(-0 170 03)		0.007
	G Sellimonas(16S)	-0.08(-0.160.01)		0.023
	G Desulfovibrio (MGS)	-0.06(-0.130.00)		0.025
	G Lachnospiraceae, noname(MGS)	-0.00(-0.130.00)		0.037
	S Roseburia unclassified/MGS)	-0.04(-0.060.00)		0.037
	S Lachnospiraceae bacterium 7 1 58544/4	-0.00(-0.140.02)		0.010
	O_Coriobacteriales/MCS)	0.06(0.02 - 0.10)		0.003
		0.00(0.02 - 0.10)		0.000
		-0.03(-0.000.00)		0.040
	F_Ruminococcaceae(165)	0.05(0.02 - 0.08)		0.001
	F_Family All(105)	0.04(0.01 - 0.07)		0.014
	F_Crimstensenenaceae(165)	0.08(0.00 - 0.16)		0.042
	F_bacteroidales_noname(MGS)	-0.05(-0.090.00)		0.032
	F_Conobacteriaceae(MGS)	0.06(0.02 - 0.10)		0.006
	G_Husicatenibacter(16S)	-0.04(-0.070.00)		0.047
	G_Anaeronium(16S)	-0.06(-0.110.01)		0.031
	G_Adiercreutzia(16S)	-0.06(-0.100.02)		0.005
	G_Ruminococcaceae UCG005(16S)	0.05(0.02 - 0.08)	1-8-1	0.002
	G_Christensenellaceae R 7 group(16S)	0.05(0.02 - 0.08)		0.001
	G_Senegalimassilia(16S)	0.06(0.01 - 0.10)		0.016
	G_Intestinibacter(16S)	-0.04(-0.080.01)		0.012
	G_Lachnospiraceae ND3007 group(16S)	-0.03(-0.070.00)		0.028
	G_Ruminococcaceae UCG014(16S)	0.06(0.02 - 0.09)		0.001
	G_Ruminococcaceae UCG011(16S)	-0.09(-0.160.03)		0.005
	G_Anaerotruncus(16S)	0.04(0.01 - 0.07)		0.010
	G_Ruminococcaceae UCG002(16S)	0.04(0.01 - 0.07)	H	0.003
	G_Oscillibacter(16S)	0.05(0.01 - 0.09)		0.018
	G_Bacteroidales_noname(MGS)	-0.05(-0.090.00)		0.032
	G_Collinsella(MGS)	0.07(0.02 - 0.11)		0.003
	S_Streptococcus_parasanguinis(MGS)	0.12(0.02 - 0.21)		• 0.019
	S_Paraprevotella_xylaniphila(MGS)	0.08(0.00 - 0.15)		0.038
	S_Bacteroidales_bacterium_ph8(MGS)	-0.05(-0.090.00)		0.031
	S_Bacteroidales_bacterium_ph8(MGS) S_Collinsella_aerofaciens(MGS)	-0.05(-0.090.00) 0.06(0.02 - 0.11)		0.031 0.009

FIGURE 3

The association between Aortic Aneurysm Disease and Gut Microbiota using the inverse variance weighted method. CI, confidence interval; *P*, probability value; AAA, Abdominal Aortic Aneurysm; TAA, Thoracic Aortic Aneurysm; AD, Aortic Dissection; 16S rRNA, 16S ribosomal RNA; MGS, Metagenomic Species.

Family XIII shows a suggestive negative associations with AAA, while Victivallales from Verrucomicrobia and Lentisphaeria from Lentisphaerae are suggestive protective factors for TAA and AD. Oxalobacter from the phylum Proteobacteria has been identified as having a suggestive association as a potential risk factor for aortic aneurysmal diseases; however, studies on the species characteristics of the aforementioned microbial taxa are scarce. Further research is needed to validate these results and elucidate the potential mechanisms involved. Additionally, there is a contradiction with previous studies; this study found that Parabacteroides merdae from the phylum Bacteroidetes has a suggestive association as a potential risk factor for TAA. In contrast, past research suggests this species may be beneficial for aneurysmal diseases, as it can reduce inflammation and cardiovascular damage, with no evidence proving it as a risk factor for aortic aneurysmal diseases (Qiao et al., 2022; Chen et al., 2023; He et al., 2023).

Through this study, several points warrant further consideration. We found reverse causal relationships between aortic aneurysm diseases and certain GM. The development of aortic aneurysms is typically accompanied by systemic inflammation and metabolic disturbances. These changes may alter the gut environment and barrier function, affecting the distribution of GM and potentially leading to perioperative gastrointestinal complications associated with aortic aneurysm diseases. Moreover, it is important to note that many diseases, including aortic aneurysms, often undergo a prolonged progression before diagnosis or onset. During this period, changes in the levels of some bacteria might not necessarily be harmful. On the contrary, these changes might represent a compensatory response by the microbiome to mitigate adverse reactions in the host. This possibility should be understood within a broader context. This study has analyzed and discussed the impact of gut bacteria on aortic aneurysm diseases. In fact, opportunistic pathogens in the oral microbiome also play a significant role in the progression of cardiovascular diseases. Clinically, various pathological oral bacteria have been detected in aortic aneurysm specimens (Kurihara et al., 2004; Marques et al., 2006; Nakano et al., 2008; Ding et al., 2014; Salhi et al., 2021). Infection with Porphyromonas gingivalis can promote the phenotypic transition of human aortic smooth muscle cells from a contractile to a synthetic state (Inaba et al., 2009). Periodontitis-induced inflammation and oxidative stress can lead to arterial stiffness, vascular dysfunction, and hypertension (Czesnikiewicz-Guzik et al., 2019; Munoz et al., 2020), and can activate the renin-angiotensin system, causing the release of Angiotensin II (Viafara-Garcia et al., 2019). These factors might contribute to the development of aortic aneurysms. Additionally, various non-bacterial microorganisms in the gut may also influence disease development, such as fungi and viruses in the gut. For instance, cytomegalovirus infection can promote aortic inflammation (Jablonska et al., 2021). Therefore, when exploring the relationship between human microbiota and aortic aneurysm diseases, it is essential to consider this broader, comprehensive perspective.

The strengths of our study include leveraging the largest available GM GWAS and metagenomic sequencing databases, enabling specieslevel microbiota classification. We utilized the FinnGen database, the most comprehensive for aortic aneurysm diseases, and confirmed our findings with data from the UK Biobank and MGI. Our MR analysis, which uses genetic variants as instrumental variables, reduces the influence of confounding factors. However, this study also has some limitations. Initially, the accuracy and reliability of MR analysis largely depend on the sample size and representativeness. Both the GM and aortic aneurysm disease GWAS data are relatively small in volume, which may reduce the statistical power of the analysis and the generalizability of the results, necessitating further expansion with updated data to strengthen the findings. Furthermore, despite employing several methods to assess and adjust for potential heterogeneity or pleiotropic effects, we cannot entirely exclude the impact of unknown heterogeneity or pleiotropy. This could confound the causal inference and necessitates further validation through different analytical approaches or additional datasets. Additionally, the study primarily includes data from individuals of European descent, which may not fully capture the genetic diversity and microbiome variations present in other populations. This limitation calls for caution when generalizing the findings to other ethnic groups. Lastly, to further confirm the mechanistic relationship between the gut microbiome and aortic aneurysm diseases, further experimental research is necessary to elucidate the biological pathways involved.

In conclusion, this study provides valuable insights into the potential role of the GM in cardiovascular health, particularly concerning aortic aneurysm diseases. It paves the way for new research directions and highlights the potential for microbiomebased interventions in mitigating the risk of these severe conditions.

Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article/Supplementary Material.

Ethics statement

Ethical approval was not required for the study involving humans in accordance with the local legislation and institutional requirements. Written informed consent to participate in this study was not required from the participants or the participants' legal guardians/next of kin in accordance with the national legislation and the institutional requirements.

Author contributions

YS: Conceptualization, Data curation, Investigation, Software, Visualization, Writing – original draft, Writing – review & editing, Formal analysis, Supervision. HD: Formal analysis, Writing – original draft, Data curation, Resources, Visualization. CS: Writing – original draft, Software. DD: Data curation, Formal analysis, Writing – review & editing. RG: Writing – review & editing, Resources. MV: Writing – review & editing, Validation. RK: Writing – review & editing, Conceptualization, Methodology, Supervision. NW: Writing – original draft.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fcimb.2024.1406845/full#supplementary-material

SUPPLEMENTARY ADDITIONAL FILE 1

Information on GWAS samples of gut microbiota used in this study

SUPPLEMENTARY ADDITIONAL FILE 2 Detailed Information of Instrumental Variables used in this study

SUPPLEMENTARY ADDITIONAL FILE 3

Mendelian randomization analysis results of gut microbiota and aortic aneurysm diseases

SUPPLEMENTARY ADDITIONAL FILE 4

Heterogeneity, level pleiotropy test, and the power of Mendelian randomization analysis

SUPPLEMENTARY ADDITIONAL FILE 5

The scatter plot effect size, leave-one-out analyses, and funnel plot for gut microbiota on gastrointestinal diseases in MR analysis.

SUPPLEMENTARY FIGURE 1

An overview of the Mendelian randomization study results on Gut Microbiota in relation to Abdominal Aortic Aneurysm. The analysis methods include inverse variance weighted, weighted median, and MR-PRESSO analysis. 16S rRNA = 16S ribosomal RNA; MGS = Metagenomic Species; SNP = single nucleotide polymorphism; IVW = inverse variance weighted; WM = weighted median; P = probability value

SUPPLEMENTARY FIGURE 2

An overview of the Mendelian randomization study results on Gut Microbiota in relation to Thoracic Aortic Aneurysm. The analysis methods include inverse variance weighted, weighted median, and MR-PRESSO analysis. 16S rRNA = 16S ribosomal RNA; MGS = Metagenomic Species; SNP = single nucleotide polymorphism; IVW = inverse variance weighted; WM = weighted median; *P* = probability value.

SUPPLEMENTARY FIGURE 3

An overview of the Mendelian randomization study results on Gut Microbiota in relation to Aortic Dissection. The analysis methods include inverse variance weighted, weighted median, and MR-PRESSO analysis. 16S rRNA = 16S ribosomal RNA; MGS = Metagenomic Species; SNP = single nucleotide polymorphism; IVW = inverse variance weighted; WM = weighted median; P = probability value.Abbreviation

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