Check for updates

OPEN ACCESS

EDITED BY Ajaz Bhat, Sidra Medicine, Qatar

REVIEWED BY Vijay Antharam, Methodist University, United States Javaid Ahmad Sheikh, Jamia Hamdard University, India

*CORRESPONDENCE Manzoor Ahmad Mir Imanzoor@kashmiruniversity.ac.in Abdul Haseeb Shah Imadulhaseeb@kashmiruniversity.ac.in

RECEIVED 16 January 2024 ACCEPTED 03 June 2024 PUBLISHED 05 July 2024

CITATION

Qadri H, Shah AH, Almilaibary A and Mir MA (2024) Microbiota, natural products, and human health: exploring interactions for therapeutic insights. *Front. Cell. Infect. Microbiol.* 14:1371312. doi: 10.3389/fcimb.2024.1371312

COPYRIGHT

© 2024 Qadri, Shah, Almilaibary and Mir. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Microbiota, natural products, and human health: exploring interactions for therapeutic insights

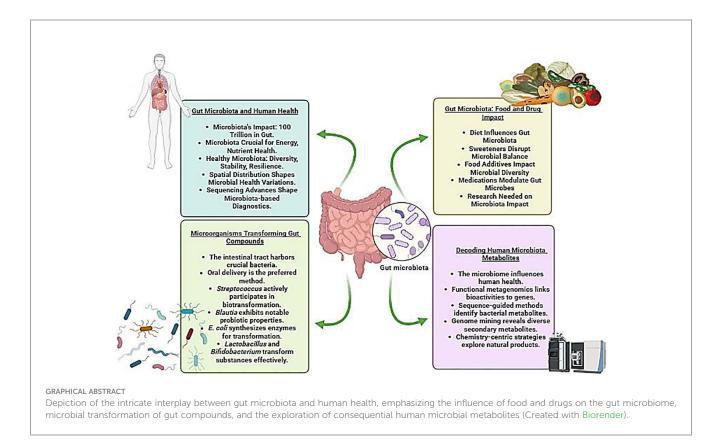
Hafsa Qadri¹, Abdul Haseeb Shah^{1*}, Abdullah Almilaibary^{1,2} and Manzoor Ahmad Mir^{1*}

¹Department of Bioresources, School of Biological Sciences, University of Kashmir, Srinagar, India, ²Department of Family and Community Medicine, Faculty of Medicine, Al Baha University, Al Bahah, Saudi Arabia

The symbiotic relationship between the human digestive system and its intricate microbiota is a captivating field of study that continues to unfold. Comprising predominantly anaerobic bacteria, this complex microbial ecosystem, teeming with trillions of organisms, plays a crucial role in various physiological processes. Beyond its primary function in breaking down indigestible dietary components, this microbial community significantly influences immune system modulation, central nervous system function, and disease prevention. Despite the strides made in microbiome research, the precise mechanisms underlying how bacterial effector functions impact mammalian and microbiome physiology remain elusive. Unlike the traditional DNA-RNA-protein paradigm, bacteria often communicate through small molecules, underscoring the imperative to identify compounds produced by human-associated bacteria. The gut microbiome emerges as a linchpin in the transformation of natural products, generating metabolites with distinct physiological functions. Unraveling these microbial transformations holds the key to understanding the pharmacological activities and metabolic mechanisms of natural products. Notably, the potential to leverage gut microorganisms for large-scale synthesis of bioactive compounds remains an underexplored frontier with promising implications. This review serves as a synthesis of current knowledge, shedding light on the dynamic interplay between natural products, bacteria, and human health. In doing so, it contributes to our evolving comprehension of microbiome dynamics, opening avenues for innovative applications in medicine and therapeutics. As we delve deeper into this intricate web of interactions, the prospect of harnessing the power of the gut microbiome for transformative medical interventions becomes increasingly tantalizing.

KEYWORDS

microbiota, gut microbiome, anaerobic bacteria, microbial transformation, natural products, human health

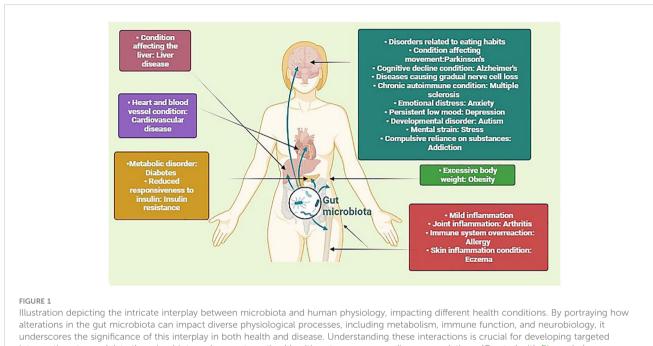


1 Introduction

The human microbiome encompasses the collective genetic material of microorganisms residing within us, including protozoa, archaea, eukaryotes, viruses, and primarily bacteria, which coexist symbiotically on and inside various regions of the human body. These inhabited sites include the mouth, reproductive organs, airways, skin, and digestive system (Lloyd-Price et al., 2016). Interestingly, the human digestive system harbors a thriving ecosystem of microbes collectively referred to as the microbiota, comprising an astonishing $\sim 10^{13}$ - 10^{14} entities. This intricate assembly, primarily composed of anaerobic bacteria, encompasses 500-1,000 distinct species with a genetic repertoire estimated to surpass the human genome by a factor of 150 (Kho and Lal, 2018; Marsh et al., 2023; Walker and Hoyles, 2023). Functioning as a metabolic entity finely attuned to human physiology, the microbiota operates as an organ, orchestrating processes that necessitate no evolutionary adaptations on our part. Among its pivotal functions is the processing of dietary components that would otherwise be indigestible like plant polysaccharides (Bäckhed et al., 2004). The human microbiome has emerged as a linchpin in complex physiological processes, influencing a spectrum ranging from immune system modulation to the central nervous system and brain development (Kau et al., 2011; Smith, 2015). Insights derived from mouse models underscore the indispensable part of a healthy bacterial environment in maintaining typical physiological processes, in contrast to imbalances, or dysbiosis, linked to an array of diseases including cancer, diabetes, obesity,

and colitis (Figure 1) (Garrett et al., 2007; Ridaura et al., 2013; Smith et al., 2013; Bongers et al., 2014). The burgeoning field of microbiome research is further underscored by a spike in clinical trials and venture capital investments, with over 1,200 clinical trials related to the "gut microbiome" registered in the National Institutes of Health clinical trials database (Gormley, 2016; Marchesi et al., 2016).

There is still a significant knowledge vacuum regarding the precise mechanisms by which bacterial functions, also known as effector functions, influence mammalian or microbiome physiology, despite growing evidence linking host-associated bacteria to both healthy development and disease in animal models as well as correlational findings in humans. The traditional framework of molecular biology, which follows the linear flow of genetic information from DNA to RNA to protein, may not fully capture the complexity of biological systems. In many cases, biological functions do not solely result in protein synthesis but also involve the generation and utilization of small molecules. This phenomenon is particularly evident in bacteria, where interactions with the environment heavily rely on low-molecular-weight compounds, including small molecules and natural products. While these small molecules are not directly encoded by DNA, they play crucial roles in mediating microbial functions and their interactions with the host organism. Therefore, to comprehensively understand the molecular mechanisms underlying the role of the human microbiome in both health and disease, it is essential to thoroughly identify and characterize the small compounds produced by human-associated bacteria (Milshteyn et al., 2018).



interventions to modulate the microbiota and promote optimal health outcomes across diverse populations. (Created with Biorender).

Gut microbes perform a pivotal function in decomposing and transforming natural products, generating a diverse array of metabolites and functional compounds having unique physiological functions that the host organism cannot synthesize on its own (Koppel et al., 2017; Xie et al., 2020). The process of microbial transformation in natural products encompasses various chemical reactions, including demethylation, hydrolysis, methylation, etc, which collectively regulate the form of the natural product substrates (Morgan et al., 2022; Rocchetti et al., 2022). Importantly, the impact of gut microbiota extends beyond mere structural modifications, significantly influencing the chemical landscape, pharmacological activities, and metabolic mechanisms of natural products. Surprisingly, the ability to harness gut microorganisms for the massive manufacture of active metabolites and compound synthesis remains largely unexplored. Delving into the study of these gut microorganisms, their metabolites, and the intricate reactions engaged in the dynamic interplay between gut microbiota and natural products holds immense importance. Unraveling these interactions is essential to both realizing the promise of natural products in several applications and comprehending the pharmacological processes at work (Zhao et al., 2022). This exploration opens avenues for further research into the utilization of gut microbes in the synthesis of bioactive compounds on a larger scale, shedding light on innovative approaches for the production and utilization of these compounds. Consequently, studying the multifaceted relationships between natural products and gut microbiota not only enhances our understanding of pharmacological mechanisms but also paves the way for harnessing the therapeutic potential of these interactions.

Our goal in this review study is to explore and synthesize the present state of information in this dynamic field, shedding light on the interactions between natural products, bacteria, and human health. Recent efforts have illuminated a fascinating facet of microbiome dynamics: the generation of natural products. This burgeoning area of research explores metabolites derived from the microbiome and seeks to unravel their part in human health (Wilson and Nicholson, 2017). In this review, we delve into these recent efforts, describing key approaches used to identify and characterize microbiome-derived natural products. This comprehensive exploration contributes to the evolving understanding of the intricate relationships between the microbiome, natural products, and human health, ultimately paving the way for innovative applications in medicine and therapeutics.

2 Microbiome

Microbial organisms commonly exist in communal arrangements and often form close associations with intricate beings like humans, forming mutualistic, parasitic, pathogenic, and other associations. The ensemble of these microorganisms is referred to as the microbiome or microbiota. While the term "microflora" has been used, it is considered a misnomer as "flora" traditionally represents the plant kingdom (Liang et al., 2018). Initially, the term "microbiome" denoted the group of microbes and the contents of their genomes, while "microbiota" described the microbial community within their hosts. However, the interchangeability of "microbiome" and "microbiota" has become prevalent (Ursell et al., 2012). The human body has a microbiome in every part of it, from the skin to the gut and even in places like the bloodstream that were thought to be sterile in the past (Proal et al., 2014). Numerous reports suggest the presence of over 10,000 microbial species occupying various human body parts (Blaser, 2006; Ley et al., 2006). Although

skin and vaginal sites exhibit comparatively minimal diversity of microbes, greater diversity is observed in sites such as the gut (Jiang et al., 2009). There can be a significant relationship between human diseases and the microbiome and vice versa. For instance, long-term lung conditions may change the lung microbiome's makeup, which may then impact host immunity and defense and worsen the conditions (O'Dwyer et al., 2016). Furthermore, research has revealed that the microbiome present in the blood or lungs is affected by the presence of infection (Jiang et al., 2009; Jostins et al., 2012; Kwan et al., 2013; Leung and Wu, 2015).

2.1 Gut microbiome

The gut microbiome, comprising the genetic material of microorganisms like fungi, protozoa, bacteria, etc, resides in the digestive tracts of humans and other animals (Table 1), including insects (Liang et al., 2018). Having evolved alongside its host for millennia, the human gut microbiome plays a crucial role in various essential activities, including nutrient absorption and digestion (Turnbaugh and Gordon, 2009; Hehemann et al., 2010), detox and body defense (Relman, 2012), host immune system development (Turnbaugh and Gordon, 2009), etc. The mammalian gut hosts a diverse array of microbes, with the majority belonging to the Firmicutes and Bacteroidetes (Ley et al., 2008). This pattern holds correct across different populations, such as Koreans (Nam et al., 2011), Africans (De Filippo et al., 2010; Schnorr et al., 2014), Europeans and Americans (Qin et al., 2010), and Danes though not in the case of Chinese (Wu G. et al., 2014). The diversity of microorganisms can carry distinct consequences for diseases in various populations. For instance, individuals with type 2 diabetes from European and Chinese backgrounds exhibit varying compositions of gut microbiomes, with the Chinese population showing greater species diversity (Qin et al., 2012). Nonetheless, understanding the significant differences among these populations, considering factors like age, environment, and genetics, requires further investigation (Tai et al., 2015).

The gut microbiome harbors millions of diverse microorganisms, each contributing to its metabolic diversity and adaptability, with some genes potentially acquired from environmental bacteria (Hehemann et al., 2010; Qin et al., 2010). Notably, three primary enterotypes [Enterotypes are defined as clusters or patterns of microbial community composition in the gut microbiome that are driven by various factors such as diet, geography, host genetics, and lifestyle. Rather than discrete and stable classifications, enterotypes are now viewed as dynamic and context-dependent configurations of the gut microbiota. These configurations may shift over time in response to environmental changes, host health status, and other factors, reflecting the complexity and flexibility of the gut microbial ecosystem (Costea et al., 2018)]-Bacteroides, Prevotella, and Ruminococcus-have been identified in the human gut across diverse populations (Arumugam et al., 2011). These enterotypes, observed in Europeans, Japanese, and Americans, demonstrate a consistent presence in mice and chimpanzees (Moeller et al., 2012; Hildebrand et al., 2013; Wang et al., 2014). While the composition is mainly shaped by the host's evolution, recent findings suggest that diet has a more significant impact on the metabolome than the microbiome. Discrepancies, especially regarding the prevalence of ecotypes like Ruminococcus, warrant further investigation considering factors such as sample size and variations in sampling methods (Arumugam et al., 2011; Wu et al., 2011; Huse et al., 2012; Zupancic et al., 2012; Gorvitovskaia et al., 2016; Liang et al., 2017). The gut microbiota comprises autochthonous microbes (Autochthonous refers to indigenous or native microbial species that are typically well-adapted to a specific environment, such as the gut) residing on the colonic mucosa epithelium and

TABLE 1 Primary categories of gut microbiota in both humans and animal models (Hillman et al., 2017).

S. No	Category	Human	Mouse	Rat
1.	Bacteria	Proteobacteria		
		Actinobacteria		
		Bacteroidetes	Bacteroidetes	Bacteroidetes
		Firmicutes	Firmicutes	Firmicutes
2.	Eukarya	Cladosporium	Zygomycota	Zygomycota
		Saccharomyces	Chytridiomycota	Chytridiomycota
		Candida	Ascomycota	Ascomycota
		Malassezia	Basidiomycota	Basidiomycota
3.	Archaea	Nitrososphaera		
		Methanobrevibacter	Methanobrevibacter	Methanobrevibacter
4.	Viruses	Adenoviridae		
		Polyomaviridae		
		Papillomaviridae		
		Herpesviridae	Variable	Variable

allochthonous microbes (Allochthonous refers to microbial species that are not native to a particular environment but are introduced from external sources) transiently passing through the lumen with digesta (Schnabl and Brenner, 2014). Distinguishing between these "residents" and "passengers" becomes crucial, as their roles are believed to differ significantly. The ratio of autochthonous to non-autochthonous microbes serves as a valuable indicator for assessing the progression of cirrhosis (Bajaj et al., 2014).

Diet and phylogeny are major contributors to the modification of the gut microbial community in various species, including mammals (Lev et al., 2008; Miyake et al., 2015). Genome-scale metabolic modeling reveals that changes in the host diet significantly alter the makeup of key human gut bacteria (eg. B. thetaiotaomicron) (Shoaie et al., 2013). Examples such as the impact of alcohol on intestinal microbiota emphasize the bidirectional relationship between diet and microbial composition, influencing host metabolism and related diseases (Mutlu et al., 2009; Chen et al., 2011; Yan et al., 2011; Mutlu et al., 2012; Queipo-Ortuño et al., 2012). Host genetics also play a crucial role in determining microbiome composition (Garrett et al., 2007; Maslowski et al., 2009; Benson et al., 2010; Brinkman et al., 2011; Lepage et al., 2011; Hashimoto et al., 2012; McKnite et al., 2012; Goodrich et al., 2014; Knights et al., 2014; Benson, 2015; Kubinak et al., 2015; Bonder et al., 2016a; Snijders et al., 2016; Igartua et al., 2017). The identification of shared susceptibility loci between inflammatory bowel disease and specific infectious organisms underscores the significance of exploring the interconnections among susceptibility, microbiome composition, and the development of the disease. This emphasizes the need to develop

effective protocols for disease prevention by understanding these relationships (Kane et al., 2011; Henao-Mejia et al., 2012; Jostins et al., 2012; Knights et al., 2014). In the gut, Gram-negative bacteria produce lipopolysaccharide (LPS), a microbial product transported with chylomicrons (Balagopal et al., 2008; Jiang et al., 2009; Hofer et al., 2010; Marchetti et al., 2013). LPS acts as a potent stimulator of innate immunity, with its levels serving as important indicators for survival in peritoneal dialysis patients (Kwan et al., 2013). Similarly, trimethylamine (TMA) and its oxidation product, trimethylamine N-oxide (TMAO), impact patient morbidity, highlighting the farreaching consequences of localized microbiome activity (Tang et al., 2017). These observations underscore the potential of using plasma levels of LPS and bacterial DNA as markers for both systemic inflammation and prognosis (Alexander and Rietschel, 2001).

3 Gut microbiota and human health

The GI tract of humans houses a vast community of microbes of approximately 100 trillion microorganisms, significantly impacting human health and disease (Figure 2). This intricate association between gut microbiota and fundamental biological mechanisms has been extensively studied, revealing its involvement in immune systems, metabolic processes, and nutrient absorption (Ley et al., 2006). The gut microbiota, comprising bacteria, yeasts, and viruses, plays a crucial role in energy and nutrient extraction through diverse metabolic genes, providing unique enzymes and biochemical pathways. Additionally, it contributes to the manufacture of essential substances like lipids, vitamins, and

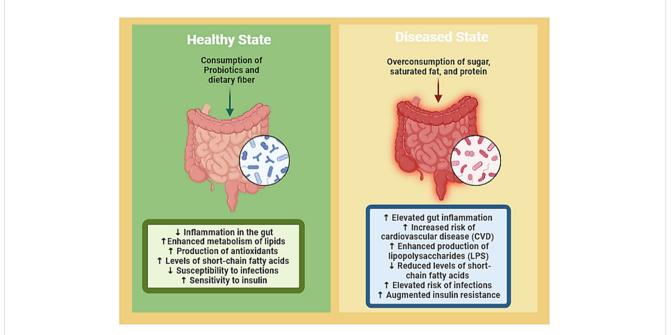


FIGURE 2

Visual representation delineates the intricate interplay between gut microbiota, nutrition, and the delicate balance between health and disease states. It illustrates how dietary factors can either promote a healthy gut microbiota composition, which contributes to overall well-being, or lead to dysbiosis, predisposing individuals to various diseases. Through highlighting this dynamic relationship, the figure underscores the potential of targeted dietary interventions to restore microbial balance and mitigate disease risk, thereby emphasizing the crucial role of nutrition in maintaining gut health and preventing the onset of illness. (Created with Biorender).

amino acids (Rowland et al., 2018). In terms of immunity, the human microbiota acts as a defense mechanism by producing antimicrobial substances and influencing intestinal mucosal growth as well as the immune system's development. When the gut microbiota is in a healthy state, it interacts symbiotically with the host and is stable and resilient. Defining a "healthy" gut microbiota involves considering factors such as high taxonomic diversity, microbial gene richness, and a stable core microbiota (Fan and Pedersen, 2021). Nevertheless, individual variations and dynamic changes influenced by aging, and contextual elements, including the use of medications, are noted.

Spatial distribution within the GI tract also contributes to microbial variation, with distinct microbial communities present in the small intestine and colon due to differences in transit time, bile concentration, flow rates, and pH (Flint et al., 2012). Agerelated variations indicate an increase in microbiota diversity from childhood to adulthood, followed by a decrease in older age, where shifts in microbial composition, including decreased Bifidobacterium and increased Clostridium and Proteobacteria, are observed (Rinninella et al., 2019). The impact of microbiota on human well-being extends beyond compositional studies. Recent advancements in high-throughput sequencing, microbiota interaction modeling, and simulation techniques have shifted focus toward understanding the causality of microbiota functions. This has significant implications for the advancement of microbiome-based diagnostics and customized medicine (Ley et al., 2006; Bouskra et al., 2008; Amabebe et al., 2020).

Therefore, the intricate interplay between gut microbiota and human health involves dynamic mechanisms influenced by age, spatial distribution within the GI tract, and environmental factors. As research progresses, a deeper understanding of microbiota's functions is essential to strengthen the advancement of personalized medicine and microbiome-based diagnostics.

3.1 The impact of food and drugs on the composition of the gut microbiota

The quantity of gut bacteria is greatly influenced by particular foods and dietary habits, which in turn affects health. High-intensity sweeteners, widely utilized as sugar substitutes, raise concerns based on animal studies. Despite regulatory agencies deeming them "generally recognized as safe," sucralose, aspartame, and saccharin disrupt gut microbiota balance and diversity. Rats exposed to sucralose for twelve weeks displayed elevated amounts of total aerobic bacteria, Bacteroides, and Clostridium, along with a significantly higher fecal pH (Abou-Donia et al., 2008). Mice exposed to sucralose for six months exhibited increased expression of pro-inflammatory genes and altered fecal metabolites (Bian et al., 2017). Common food additives that impact gut flora include emulsifiers, which are included in processed meals. Mice given carboxymethylcellulose and polysorbate-80 displayed decreased microbial diversity, decreased Bacteroidales and Verrucomicrobia, and enrichment of inflammation-promoting Proteobacteria (Chassaing et al., 2015). Concerns extend to certain restrictive diets. Studies on vegan diets revealed variations in the gut microbeproduced serum metabolites, but only slight alterations in the bacterial communities (Wu G. et al., 2014). Gluten-free diets, recommended for gluten sensitivity or celiac disease, altered gut microbiota profiles in healthy individuals, potentially affecting useful microbial species (Bonder et al., 2016b). The low FODMAP diet, beneficial for irritable bowel syndrome, led to significant microbiota and metabolome changes (McIntosh et al., 2016; Gibson, 2017; Halmos, 2017; Bennet et al., 2018). Medications also modulate the composition of gut flora. An important research identified drugs such as progesterone, rupatadine, TNF- α inhibitors, and osmotic laxatives, as major modulators (Falony et al., 2016). Proton pump inhibitors and antibiotics, extensively used in humans and livestock, influence gut microbes, potentially contributing to obesity (Jackson et al., 2015; Blaser, 2016).

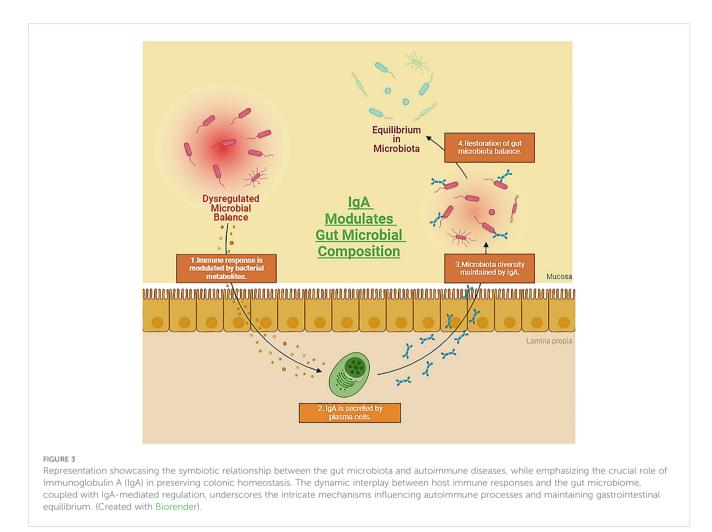
While clinical evidence is insufficient for clear recommendations, future studies on food additives, drugs, and dietary modifications should consider their effects on the gut microbiota. In medical contexts like cancer treatment and autoimmune disorders (Figure 3), where microbiota changes impact responses, understanding these dynamics becomes crucial (Alexander et al., 2017). Animal studies also highlight the role of specific gut microbes in transforming compounds for protective effects, emphasizing the interconnectedness of diet, gut health, and overall well-being (Spanogiannopoulos et al., 2016).

4 Crucial gut microorganisms involved in the transformation of natural compounds

Oral administration is the preferred mode of drug delivery, as evidenced by the 84% oral composition of the top 50 best-selling pharmaceuticals in the US and Europe (Vinarov et al., 2021). The impact of gut microbiota on the durability of orally administered natural products has garnered significant interest in recent years. The intestinal tract harbors a plethora of bacteria crucial for normal digestive function, with approximately 98% of gut microorganisms in healthy individuals falling into 4 phyla: *Proteobacteria*, *Actinobacteria*, *Firmicutes*, and *Bacteroidetes* (Manor et al., 2020; Ye et al., 2021; de Vos et al., 2022). Specific gut microorganisms, like *Escherichia coli*, *Streptococcus*, etc. actively take part in the biotransformation of natural products. The resulting metabolites contribute to enhanced intestinal absorption, playing a significant pharmacological role (Zhao et al., 2022).

4.1 Streptococcus

The *Streptococcus* species, characterized by gram-positive, circular, or ovoid cells organized in chains or pairs, is commonly found in the nasopharynx and human stool (Lannes-Costa et al., 2021). Meta-transcriptomic study reveals that *Streptococcus* predominantly expresses the phosphotransferase system, suggesting their primary role in utilizing the carbohydrates that are present in the small intestine (Zoetendal et al., 2012). *Streptococcus* LJ-22 expresses β -glucuronidase activity, converting



GL to 18β-glycyrrhetinic acid-3-O-β-D-glucuronic acid (GAMG), known for its anti-allergic properties. This compound has demonstrated anti-allergic potential against LPS-induced RAW264.7 cells (Guo et al., 2018). Furthermore, tannic acid degradation by Streptococcus gallolyticus subsp. Gallolyticus may contribute to colorectal cancer development by neutralizing its toxicity in the case of tumor cells (Oehmcke-Hecht et al., 2020). Streptococcus thermophilus GIM 1.321 demonstrates high β glucosidase production, facilitating the breakdown of fructus anthocyanins into beneficial compounds like ferulic acid, CAA (caffeic acid), and CHA (chlorogenic acid) (Cheng et al., 2016). Administering CAA and CHA from these processes has shown the potential to lower blood pressure and provide antioxidant effects (Agunloye et al., 2019). Streptococcus strains, acting as commensals, pathogens, or opportunistic pathogens in the gut, require further exploration regarding their impact on the well-being of humans. Comprehending the way Streptococcus metabolizes natural compounds may help regulate the gut microbiota and improve the effectiveness of treatments. In-depth in vivo research is crucial to find out if focusing on bacteria that break down tannic acid could help develop more potent treatments for colorectal cancer. Overall, unraveling the intricacies of Streptococcus metabolism holds promise for advancing our comprehension of the gut microbiota and optimizing treatment interventions (Zhao et al., 2022).

4.2 Blautia

Blautia species, prevalent in the anaerobic environments of mammalian guts and feces, exhibit notable probiotic properties in the biotransformation of natural products. Tremaroli and Bäckhed (2012) (Tremaroli and Bäckhed, 2012) highlight Blautia's involvement in flavonoid processing, encompassing demethylation, O- and C-deglycosylation, and C-ring cleavage catalyzed by enzymes like O-glycosidase and β -glucosidases (Braune et al., 2016). Notably, Blautia sp. MRG-PMF1 demonstrates hydrolytic prowess, conver ting compounds like 5,7-dimethoxyflavone into chrysin and 5,7,4trimethoxyflavone into apigenin. This strain also exhibits deglycosylation activity on various isoflavones, flavones, and flavonols, transforming them into their corresponding aglycones (Kim et al., 2014). In anaerobic environments, Blautia sp. MRG-PMF1 catalyzes curcumin to generate demethoxycurcumin, which has anti-inflammatory and anti-cancer characteristics and further metabolizes icariin into desmethylicaritin, which has estrogenic properties (Wu H. et al., 2016; Burapan et al., 2017; Hatamipour et al., 2019). Another strain, Blautia sp. AUH-JLD56, exclusively biotransforms arctiin or arctigenin into demethylated products with enhanced antioxidant potential (Liu et al., 2013). The creation of novel enzymes and bioactive metabolites is greatly encouraged by the increasing scholarly interest in Blautia's involvement in the

biotransformation and metabolism of herbal plants and functional foods (Meng et al., 2020).

4.3 E. coli

Escherichia coli, a facultative anaerobic, non-sporous, gramnegative bacterium, predominantly resides in vertebrate intestines, playing a crucial role in glycosidase production for the transformation of exogenous substances (Foster-Nyarko and Pallen, 2022). Notably, *E. coli* strain *HGU-3* synthesizes β glucuronidase, facilitating the conversion of baicalin to baicalein, which exhibits superior anti-oxidant and anti-inflammatory potential (Han et al., 2016; Li D. et al., 2019). Additionally, certain *E. coli* strains, like *DH10B*, demonstrate high curcuminconverting activity through the expression of NADPH-dependent curcumin/dihydrocurcumin reductase (CurA) (Hassaninasab et al., 2011; Tan et al., 2014). The resulting dihydrocurcumin (DHC) and tetrahydrocurcumin (THC) show promising therapeutic benefits in hepatic steatosis, surpassing the effects of curcumin (Chen et al., 2018; Yu et al., 2018).

Moreover, E. coli strains *Nu*, *MC*, and *WC-1* exhibit cinnamyl esterase properties, releasing hydroxycinnamic acids with *in-vitro* and *in-vivo* anticancer and antioxidant potential (Zhao et al., 2022). Understanding the genetic and biochemical aspects of *E. coli* opens avenues for synthesizing natural product derivatives with diverse health benefits. This knowledge is pivotal for exploring the potential of *E. coli* in producing compounds such as DHC (dihydrocaffeic acid) and THC (tetrahydrocurcumin), which effectively regulate triglyceride levels and demonstrate novel therapeutic advantages in hepatic steatosis. Overall, comprehending the capabilities of *E. coli* contributes to the development of natural product derivatives with diverse health benefits (Chen et al., 2018; Yu et al., 2018; Rodríguez-Daza et al., 2021; Candeliere et al., 2022).

4.4 Eubacterium

The genus *Eubacterium*, a gram-positive constituent of the human gut microbiota, plays a vital part in metabolizing various substances (Mukherjee et al., 2020). Notably, *E. ramulus*, a well-studied flavonoid-degrading bacterium in the human intestine, produces enzymes like chalcone isomerase and flavanone-/ flavanonol-cleaving reductase (Gall et al., 2014). These enzymes break down flavonoids into dihydrochalcone and its derivatives, known for their anti-inflammatory and antioxidant effects. *E. ramulus* strain wK1 further degrades flavonol quercetin and flavone luteolin, converting them into specific acids through reduction and ring fission (Zhao et al., 2022).

Another strain, *E. cellulosolvens ATCC 43171T*, is implicated in deglycosylating flavonoid O- and C-glucosides, exclusively catalyzing the deglycosylation of C-glucosides (Braune and Blaut, 2012; Braune et al., 2016). Additionally, *Eubacterium L-8* exhibits the ability to hydrolyze terpenoid glycyrrhizin into 18β -glycyrrhetinic acid (18β -GA), which, in turn, demonstrates anti-inflammatory effects in preventing airway allergic inflammation

(Liu et al., 2022). While these metabolic transformations highlight the potential health benefits of *Eubacterium* spp., it is emphasized that further *in vivo* studies are imperative to fully comprehend and harness the diverse advantages offered by this genus. This research could unlock opportunities to maximize the positive impact of *Eubacterium* strains on human health (Zhao et al., 2022).

4.5 Lactobacillus

The genus *Lactobacillus*, classified within the phylum *Firmicutes*, plays a crucial role in maintaining microbial balance and safeguarding gastrointestinal mucosal integrity (Dempsey and Corr, 2022). Certain *Lactobacillus* species are equipped with an array of metabolic enzymes, including α -rhamnosidases, tannase, and gallate decarboxylases, enabling them to transform exogenous substances (Reverón et al., 2017; Li B-C. et al., 2019; Ferreira-Lazarte et al., 2021).

L. rhamnosus NCTC 10302, possessing both β -glucosidase and α -rhamnosidase activities, demonstrates the ability to convert hesperetin-7-O-rutinoside and naringenin-7-O-rutinoside into their respective aglycones and 3-(phenyl) propionic acid through hydrolysis, ring fission, and dehydroxylation (Pereira-Caro et al., 2018). In a similar vein, *L. plantarum* expresses tannase, facilitating the hydrolysis of gallate and protocatechuate esters, ultimately producing gallic acid (Jiménez et al., 2014). Gallic acid, present in concentrations of 11.5–46 µg/ml, exhibits a protective part against LPS-induced inflammation and oxidative stress by suppressing the MAPK/NFF×B pathway and activating the Akt/AMPK/Nrf2 pathway (Tanaka et al., 2018).

Fang et al. witnessed the production of gallic acid and pyrogallol through the breakdown of gallotannins by gallotannin-metabolizing enzymes in *L. plantarum* WCFS1, suggesting potential prebiotic-probiotic assocaitions in preventing diet-induced metabolic diseases (Reverón et al., 2015; Fang et al., 2019). Additionally, *Lactobacillus* sp. *Niu-O16* reduces daidzein to dihydrodaidzein with daidzein reductase activity, and dihydrodaidzein, at concentrations of 2.5– 5μ M, supresses NF- κ B activation and MAPK phosphorylation, therefore enhancing osteoporosis (Wang et al., 2007; Heng et al., 2019; Kim et al., 2019).

L. casei, L. plantarum, and *L. acidophilus* significantly impact the deglycosylation of piceid to resveratrol, enhancing bioavailability and bioactivity (Basholli-Salihu et al., 2016). Furthermore, feruloyl esterases from *L. reuteri, L. helveticus*, and *L. fermentum* hydrolyze chlorogenic acid, releasing caffeic acid (Santos et al., 2018). These results underscore the potential of *Lactobacillus* in health-improving pharmaceuticals and food products, but more investigation is required to clarify the fundamental transformation mechanisms (Zhao et al., 2022).

4.6 Bifidobacterium

Bifidobacterium, a widely distributed genus within the Actinobacteria phylum, serves as one of the initial colonizers in the human gut microbiota (Satti et al., 2021; He et al., 2023). Key

species include *B. angulatum*, *B. breve*, *B. Catenulatum*, *B. adolescentis*, *B. longum* etc, collectively constituting less than 10% of the adult human microbiome yet playing a crucial role in host health (Turroni et al., 2008; Turroni et al., 2009; Hidalgo-Cantabrana et al., 2018). Some *Bifidobacterium* species express feruloyl esterase, enabling the generation of phenolic acids. For example, B. animalis's feruloyl esterase can hydrolyze chlorogenic acid into caffeic acid (CAA) (Raimondi et al., 2015). CAA (10–30 mg/kg) has demonstrated hepatoprotective effects in mice, preventing acetaminophen-induced acute liver injury by enhancing Nrf2 transcription (Raimondi et al., 2015; Pang et al., 2016). *Bifidobacterium's* involvement in the gut facilitates the metabolism of various compounds, including glycosides, saponins, and flavanones (Zhao et al., 2022).

Specific strains of *Bifidobacterium*, such as *B. longum R0175* and *B. longum SBT2928*, exhibit distinct metabolic activities. *B. longum R0175* facilitates the production of 3-(3'-hydroxyphenyl) propionic acid and 3-(phenyl) propionic acid from hesperidin by ringcleavage and demethylation (Pereira-Caro et al., 2018). *B. longum SBT2928* contributes to bile acid metabolism by hydrolyzing main human and animal bile salts, suggesting a potential role in reducing cholesterol levels *in vivo* (Tanaka et al., 2000). Furthermore, *B. breve ATCC 15700* demonstrates the production of β -glucosidase, which cleaves glycosides in ginsenoside Rd., generating deglycosylated ginsenoside compound K (Zhong et al., 2016; Zhang et al., 2019). Such metabolic properties position *Bifidobacterium* as a viable option for symbiotic establishment, utilizing its natural product synthesis capabilities for potential therapeutic applications (Zhao et al., 2022).

5 The role of biotransformation in uncovering the active substance

The intricate connection between many natural compounds' poor oral availability and significant pharmacological efficacy is being uncovered by studies on gut microbiota. The challenge lies in the complex structures of glycosides, hindering absorption by intestinal cells and limiting tissue-specific bio-accessibility. These substances undergo microbial enzymatic degradation, transforming into small molecule metabolites that exert diverse impacts on the host (Wardman et al., 2022).

Crucially, the therapeutic effects of natural compounds are significantly influenced by gut microorganisms. This is best illustrated by the conversion of ginsenosides into compound K (CK), which exhibits increased anti-inflammatory, anti-tumor, and lipid-reducing properties (Kim et al., 2013; Kim, 2018). Similarly, curcumin metabolites, influenced by the microbiota, exhibit antiinflammatory properties through pathways such as PPAR γ expression and NF- κ B inhibition (Tang et al., 2021; Lu et al., 2022). Urolithin A (UA), a gut microbe-derived compound, showcases neuroprotective and anti-inflammatory properties (Gong et al., 2019; Huang et al., 2022).

Furthermore, the intricate interplay of form, function, composition, etc within the community of gut microbes unveils

promising avenues for leveraging natural products. These compounds hold the potential to not only amplify therapeutic efficacy but also mitigate adverse effects, thereby offering a nuanced and refined approach to enhancing health outcomes. Gut microbes can modify the toxicity of certain compounds, such as reducing cardiotoxicity in digoxin metabolism (Kumar et al., 2018). However, harmful substances can also be synthesized by gut microorganisms, emphasizing the need for small molecule inhibitors to regulate specific transformations. Further research is necessary to determine how different doses of natural products affect gut microorganisms and metabolism because excessive consumption can upset the gut microbiota and cause negative effects (Lindell et al., 2022; Zhao et al., 2022).

6 Exploring human microbial metabolites

6.1 Screening to ecological insights

Diverse organisms, encompassing microorganisms, have long been recognized as prolific producers of secondary metabolites, also known as natural products (Law et al., 2020). Since the middle of the 20th century, the identification of these bioactive compounds, ranging from polyketides (PKs) to nonribosomal peptides (NRPs) and their hybrids, has been predominantly achieved through activity-based testing of cultivatable microorganisms, particularly Actinomycetes soil bacteria (Katz and Baltz, 2016). Over time, developments in molecular biology, DNA sequencing, and cultivation techniques have ushered in a new era, enabling the exploration of secondary metabolites from previously unculturable microbes (Lewis et al., 2010; Wilson and Piel, 2013; Sidebottom and Carlson, 2015; Browne et al., 2016). The impact of microbial natural products, as well as their semisynthetic derivatives, on human health, is profound, as they have emerged as vital sources for clinically relevant antibiotics, antifungals, immunosuppressants, anticancer agents, and other pharmaceuticals (Newman and Cragg, 2016). While the primary focus of natural product exploration has historically been medicinal usage, recent research has unveiled the intriguing ecological parts played by secondary metabolites in the organisms that produce them and their broader ecosystems, extending even to the human body (Crawford and Clardy, 2011; Cantley and Clardy, 2015; Wilson et al., 2017).

6.2 Secondary metabolites and health implications

The microbiome, consisting of a myriad of microorganisms that inhabit the human body (Figure 4), plays a crucial role in shaping human health and influencing various diseases. These microorganisms collectively harbor a gene pool that surpasses the human genome by a factor of 150 (Methé et al., 2012; Weinstock, 2012; Kho and Lal, 2018; Marsh et al., 2023; Walker and Hoyles, 2023). An analysis of data from the Human Microbiome Project (HMP), which includes genomic and metagenomic sequencing, has revealed that bacteria associated with humans possess the genetic machinery necessary to produce an extensive range of secondary metabolites (Donia et al., 2014). Despite this, the identities of the majority of such natural products remain undiscovered, and their functions are not fully comprehended. Investigating the ways through which these molecules impact interactions between hosts and microbes, as well as among different microbial species, could unveil factors that shape the impact of the microbiome on its host. This exploration holds the potential to yield targeted therapeutic options and may serve as a wellspring of innovative approaches to address human diseases. Few methods for uncovering natural products, and demonstrating their individual or collaborative use in deciphering the intricate metabolic interactions within the human microbiome have been presented (Wilson et al., 2017; Milshteyn et al., 2018).

6.2.1 Exploring biological functionality in metagenomic libraries through functional metagenomics

Microbial organisms exist widely in nature, thriving in diverse environmental conditions. Many of them cannot be cultured conventionally. Metagenomics allows the exploration of microbial communities directly from environmental samples, regardless of culturability. This method unveils species diversity and offers insights into their functions in natural settings. By cloning and expressing metagenomic DNA in a different host, function-based screenings can uncover novel proteins with industrial applications from previously inaccessible microorganisms. Functional metagenomics holds promise in industries like food and pharmaceuticals, facilitating the discovery of enzymes suitable for various processing conditions, novel bioactives including antimicrobial agents, and addressing concerns like development of antibiotic resistance (Coughlan et al., 2015).

Natural products have long played a pivotal role in the development of therapeutics for a variety of diseases. Traditionally, soil and marine environments have provided a rich reservoir from which diverse chemical scaffolds could be discovered. Recently, the human microbiome has been recognized as a promising niche from which secondary metabolites with therapeutic potential have begun to be isolated. The expansive history of identifying bacterial natural products in other environments is informing the approaches being brought to bear on the study of the human microbiota have also been addressed in many studies. These tools can lead to insights about microbemicrobe and host-microbe interactions and help generate biological hypotheses that may lead to developments of new therapeutic modalities (Milshteyn et al., 2018).

The functional metagenomic strategy seeks to explore bacterial metabolites through the systematic screening of metagenomic libraries for distinct bioactivities. Upon identifying a bioactive

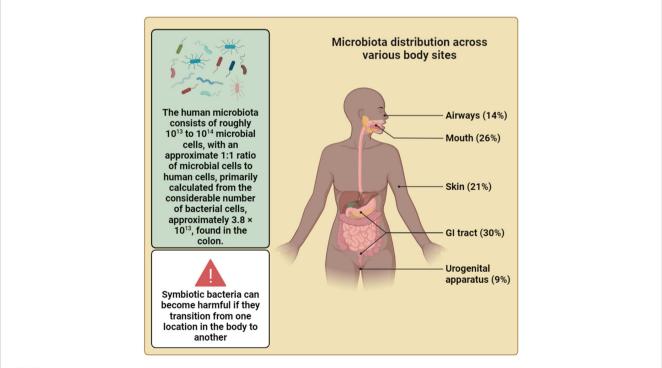


FIGURE 4

The intricate relationship between humans and their resident microbes is highlighted in this figure. The vast microbial community, comprising bacteria and other microbial organisms, plays a significant part in shaping human health and homeostasis, challenging traditional perceptions of the self as a singular entity. This depiction underscores the profound influence of the microbiota on human physiology and emphasizes the notion that, in the realm of the body's cellular landscape, we are not alone. Additionally, this depiction underscores the emerging recognition of the gut microbiota as a key orchestrator of systemic health, influencing processes beyond digestion and immunity, including metabolism etc. (Created with Biorender).

clone through functional screening, isolation of the clone and subsequent sequencing can pinpoint the gene or set of genes involved in producing the active metabolite. This direct association between the metabolite and its biosynthetic components is a key advantage of functional metagenomics. In the realm of the human microbiome, where a substantial portion of bacterial species have undergone full sequencing, this approach facilitates the simultaneous linking of bioactive metabolites with their functions inside the host and their producers within the Microbiome (Milshteyn et al., 2018).

Functional metagenomics can be applied in many different ways, but the most successful approaches that involve human microbiome metagenomic libraries have employed testing culture broth filtrates from metagenomic clones that are individually arrayed against assays using human cell reporter assays. In an early endeavor, Lakhdari and colleagues utilized a nuclear factorkB (NF-kB) activation screen to identify 171 clones from a 2,640clone human gut microbiome fosmid library, showing modulation of NF-kB reporter activity (Lakhdari et al., 2010). NF-kB, a rapidly inducible transcription factor with wide involvement in diverse cellular responses, is notably implicated in the immunoinflammatory response in the gut, rendering it a suitable indicator for detecting various microbe-host interactions (Zhang et al., 2017).

In groundbreaking research, Cohen et al. (2015) (Cohen et al., 2015) employed functional metagenomics to identify hostassociated bacterial effector genes, leading to the discovery of commendamide, a novel N-acyl amide. Commendamide was found to activate the GPR132/G2A receptor, implicated in immune cell functions. Subsequent research (Cohen et al., 2017) expanded the repertoire of N-acyl amides from the human microbiome, revealing structural similarities to endogenous GPCR ligands. Notably, certain bacterial GPR119 agonists mirrored the effects of endogenous ligands, regulating metabolic hormones and glucose homeostasis in mouse models. This suggests a potential for microbiome-biosynthetic gene therapy as a novel therapeutic modality. These studies highlight how functional metagenomics unveils microbiome-host interactions, identifies therapeutic targets, and explores microbiota-derived small molecules for innovative therapeutic strategies (Milshteyn et al., 2018).

6.2.2 Methods for extracting natural products through sequence-guided exploration of (Meta) genomes

Genome mining represents a computational approach to smallmolecule discovery, relying on algorithmic predictions to determine biosynthetic gene clusters (BGCs) within sequence data. Established algorithms, such as antiSMASH, NP.searcher, MultiGeneBlast, etc predominantly utilize conserved sequences from extensively studied BGC groups to pinpoint novel gene clusters within these families (Medema et al., 2013; Reddy et al., 2014; Medema and Fischbach, 2015). Recent developments, exemplified by ClusterFinder, aim to extend this capability to discover previously unknown BGC groups (Cimermancic et al., 2014). The detailed examination of the computational complexities associated with sequence-guided natural product identification has been extensively explored in other sources (Medema and Fischbach, 2015), our focus here centers on the application of sequence-based methods for bacterial metabolite identification within the human microbiome (Milshteyn et al., 2018).

Over the last decade, significant initiatives have been undertaken to sequence and annotate human microbiome bacteria, driven by a keen interest in understanding the microbiota's impact on human health. This has resulted in a remarkable wealth of sequence data unique to the human microbiome, encompassing over 3,000 full and partial reference genomes, along with numerous (meta)genomic shotgun sequencing datasets from both healthy and sick individuals (Aagaard et al., 2013; Donia et al., 2014). Utilizing this data, researchers are currently employing bioinformatics tools to uncover new secondary metabolite BGCs within the human microbiome. The overarching objective of these bioinformatics-driven discovery endeavors is to elucidate and functionally characterize the metabolites that are encoded by recently identified natural product BGCs (Milshteyn et al., 2018).

Utilizing the ClusterFinder algorithm, researchers identified over fourteen thousand biosynthetic gene clusters (BGCs) in the Human Microbiome Project (HMP) data, revealing the prevalence of various small molecule families, including thiopeptides (Cimermancic et al., 2014). Lactocillin, a novel thiopeptide antibiotic produced by the vaginal isolate Lactobacillus gasseri JV-V03, demonstrated potent activity against pathogenic bacteria (Mullane et al., 2015). Another innovative approach, involving the synthesis of bioinformatically predicted natural product-like structures termed syn-BNPs (Chu et al., 2016), led to the discovery of humimycins with broad-spectrum antibiotic activity. Genome mining also uncovered a family of 47 nonribosomal peptide synthetase (NRPS) BGCs present in over 90% of HMP stool samples, producing pyrazinones and dihydropyrazinones (Guo et al., 2017). Additionally, the exploration of mucin-utilizing bacteria and the identification of the fldAIBC cluster shed light on a specific bacterial metabolic pathway involving indoleacrylic acid, linking it to protective effects in a colitis model and proposing a connection between a healthy mucus layer, anti-inflammatory metabolites, and inflammatory bowel disease (IBD) (Wlodarska et al., 2017). These findings highlight the potential of mining the human microbiome for novel therapeutics and insights into hostmicrobe interactions (Milshteyn et al., 2018).

6.2.3 Exploring chemistry-centric strategies for uncovering natural products in the human microbiome

Many instances arise where genome-centric approaches are not the primary focus, highlighting the diverse avenues of research that extend beyond the realm of genomics. Essentially, both pathogenic and nonpathogenic bacteria have had their secondary metabolites studied for many years, predominantly through laboratory cultivation and analytical chemistry. The examination of individual bacteria in vitro has yielded a considerable array of compounds, with varied degrees of intricacy and biological significance, and these have been thoroughly reviewed in the latest research (Donia and Fischbach, 2015; Mousa et al., 2017). With the current intense scientific exploration of the human microbiome, there is a growing interest in delving into deeper biological contexts for in vitro-discovered secondary metabolites. This aspect has not always been a forefront consideration in traditional natural product discoveries. Increasingly, robust foundations of chemistry-centric discovery techniques are now employed to formulate intriguing biological hypotheses based on identified natural products. Strong underpinnings of chemistryfocused discovery methods are increasingly being employed to formulate intriguing biological theories rooted in discovered natural compounds (Milshteyn et al., 2018).

Utilizing sophisticated analytical methods, like liquid chromatography-mass spectrometry (LC-MS), has proven instrumental in dissecting bacterial metabolites and unraveling their functional roles. For instance, research in 2009 demonstrated that Lactobacillus plantarum, a beneficial probiotic, might reduce inflammation both in vivo and in vitro by regulating NF-kB (Petrof et al., 2009; Van Baarlen et al., 2009; Bansal et al., 2010). Zvanych et al.'s (2014) (Zvanych et al., 2014) study further applied principal component analysis to LC-MS data, identifying dipeptide molecules with pyroglutamic rings, like pyrophenylalanine and pyro-tryptophan, which, when introduced into mice, resulted into reduced splenic synthesis of IFN-g. Another notable approach involves the functional evaluation of secreted bacterial metabolites, as illustrated by the discovery of lugdunin, a novel antibiotic from nasal bacteria. Metabolomics platforms, especially those employing HRMS (High-Resolution Mass Spectrometry), have performed an essential part in examining the chemical fingerprint of the human microbiota, offering insights into diverse biological activities and potential biomarkers, as seen in studies comparing germ-free and colonized mice, cardiovascular health, and neurological diseases in mouse models (Table 2) (Wikoff et al., 2009; Hsiao et al., 2013; Brown and Hazen, 2017; Hou et al., 2022). While challenges persist in identifying unknown secondary metabolites, the evolving strategies for HRMS data collection and analysis provide optimism for unearthing therapeutic leads in the future (Peisl et al., 2018).

7 Conclusion

The exploration of the human microbiome and its intricate relationship with natural products has unveiled a captivating realm within the field of biology. The human digestive system, teeming

	Murine models	Area of Study	Importance
1.	Sterile mice populated with human microbial communities	Explore the nuanced relationship between host and microbiota in various systems, such as the gastrointestinal tract, cardiology, reproductive biology, lipid metabolism, and bone homeostasis. Investigate the dynamic interplay within each of these physiological contexts.	Create an environment devoid of any microorganisms and facilitate the introduction of particular microbiota, while simultaneously modifying the typical physiological parameters of the host.
2.	Chemically altered mice	Utilizing chemical agents to harm the epithelial cells of the gut or trigger an immune response within the mucosal lining.	A frequently employed method to instigate colitis in mice. May yield inconsistent outcomes due to differences in experimental design and environmental variables.
3.	Drug- induced mice	Antibiotics have the capacity to selectively reduce certain members of the microbiota, enabling the investigation of the bacteria's involvement in sustaining cellular functionality and signaling pathways post-development.	Relevant for any mouse genotype or condition. Could lead to the emergence of drug- resistant bacteria.
4.	Genetically engineered mice	Mimic the observable characteristics linked to genetic abnormalities in conditions like Inflammatory Bowel Disease (IBD).	Offers a potent tool for investigating the pathological processes of human illnesses. The presence of genes participating in multiple pathways could potentially impact the outcomes.

TABLE 2 Microbiota research: insights from murine models (Hou et al., 2022).

with a diverse microbiota, serves as a dynamic ecosystem comprising an astounding number of microorganisms. This microbial assembly, primarily constituted of anaerobic bacteria, orchestrates processes finely tuned to human physiology, functioning as an organ with a genetic repertoire surpassing the human genome. Among its pivotal roles is the breakdown of indigestible dietary components, highlighting its indispensable contribution to our overall well-being.

The impact of the microbiome extends beyond mere digestion, permeating into complex physiological processes, influencing immune system modulation, and perhaps aiding in the growth of the CNS and brain (Table 3) (Ghosh et al., 2021). Mouse models have underscored the vital role of a healthy bacterial environment in maintaining normal physiological processes, while dysbiosis has been associated with an array of diseases. Beyond dietary influences, gut microbiota is

TABLE 3	Compilation of microbial metabolites, gut microbiota, and					
their roles (Ghosh et al., 2021).						

	Microbial- derived metabolites	Assocaited Gut Flora/ microbiota	Role(s)
1.	Amino acids (like lysine etc)	S. aureus, E. coli, P. aeruginosa etc.	 SOS response: Triggering Biofilm formation: Modulation Deamination Regulating peptidoglycan synthesis
2.	Derivatives of indole (like indole-3- propionic acid, melatonin etc)	C. sporogenes, E. coli	 Regulation of the intestinal barrier. Control of endothelial dysfunction etc.
3.	Vitamins (like vitamin Kz2, B2 etc)	Bifidobacterium, S. aureus, etc.	 Involvement in redox cycling. Protection against pathogens. Facilitation of DNA replication, methylation, and repair. Synthesis of vitamins, nucleotides, and amino acids.
4.	Short-chain fatty acids (butyrate, valerate, acetate etc)	Bifidobacterium sp., Coprococcus, Clostridium, Bacteroidetes etc	 Regulation of host metabolic pathways via cell signaling. Modulation of the immune system. Sustaining energy balance. Enhanced glucose tolerance and insulin sensitivity. Regulation of osmotic balance.
5.	Bile acid metabolites (like cholic acid, deoxycholic acid, taurocholic acid etc)	Clostridium, Lactobacillus etc	 Activation of nuclear receptors and cellular signaling pathways in the host. Demonstrating antimicrobial properties. Regulation of lipid absorption.

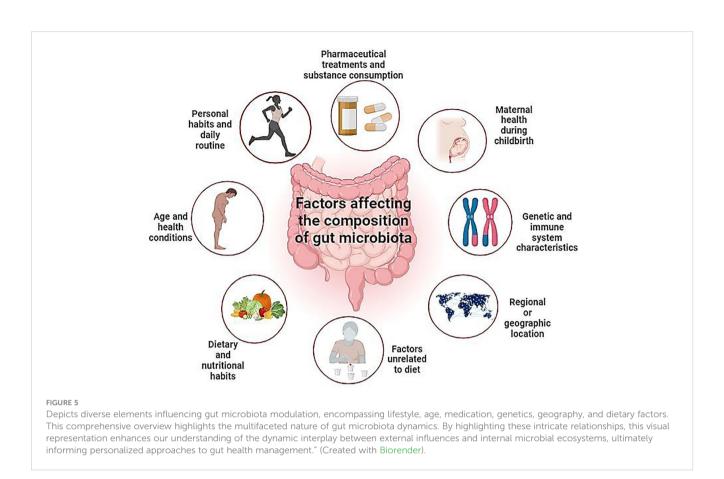
significantly affected by various factors (Figure 5) and exhibits rapid responses to system alterations due to the interplay between genetic and environmental factors. The burgeoning field of microbiome research is evidenced due to the increase in clinical studies and investments, emphasizing its potential to revolutionize healthcare. There is increasing evidence linking host-associated bacteria to both healthy development and disease, but there is still a significant knowledge vacuum regarding the precise mechanisms by which bacterial activities impact the physiology of mammals or microbiomes. The traditional molecular biology dogma falls short of elucidating the ultimate outcomes of information transfer within a biological system, particularly when it comes to small molecules and natural products.

Gut microorganisms perform a pivotal function in decomposing and transforming natural products, generating a diverse array of metabolites and functional compounds. The chemical transformations involved significantly regulate the form of natural product substrates. Importantly, the impact of gut microbiota extends beyond structural modifications, influencing the chemical landscape, pharmacological activities, and metabolic mechanisms of natural products. Intriguingly, the perspective of harnessing gut microorganisms for extensive manufacturing of active metabolites and compound synthesis remains largely unexplored. The study of gut microorganisms, their metabolites, and the intricate reactions concerned with the dynamic interplay between natural products and gut microbiota holds immense importance. Unraveling these interactions is not only crucial for understanding pharmacological mechanisms but also for unlocking the untapped potential of natural products in various applications.

8 Future perspectives

Looking ahead, the field stands at the precipice of unprecedented discoveries and applications. Delving into the study of gut microbes and their role in the synthesis of bioactive compounds on a larger scale presents an exciting avenue for future research. The untapped potential of harnessing the therapeutic benefits of these interactions holds promise for innovative approaches to medicine and therapeutics. The multifaceted relationships between natural products and gut microbiota open new doors for drug discovery, as the microbiome becomes a potential source of novel compounds with therapeutic applications. Future research should focus on elucidating the specific mechanisms by which these microbiome-derived naturally occurring compounds exert their effects on the well-being of humans. This involves a deeper understanding of the metabolic pathways involved, the pharmacological activities of these compounds, and their potential applications in treating various diseases.

Moreover, the development of technologies and methodologies for the systematic characterization of small molecules generated by human-associated microorganisms is crucial. Advancements in analytical techniques and high-throughput screening methods will facilitate the identification and isolation of novel microbiomederived natural products. Collaborations between microbiologists, pharmacologists, and clinicians will be essential in translating these discoveries into practical applications. The integration of microbiome-based therapies into personalized medicine holds the potential to revolutionize healthcare, offering tailored treatments based on an individual's unique microbiome profile. In a nutshell, the intersection of natural products, bacteria, and human health represents a frontier of scientific exploration with profound implications for medicine and therapeutics. As we navigate this



dynamic field, the collaborative efforts of researchers across disciplines will be instrumental in unlocking the full potential of microbiome-derived natural products and ushering in a new era of innovative healthcare solutions.

Author contributions

HQ: Conceptualization, Writing – original draft, Writing – review & editing. AHS: Writing – review & editing. AA: Funding acquisition, Writing – review & editing. MM: Conceptualization, Funding acquisition, Project administration, Supervision, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This work was financially supported by grants to MM by the Science and Engineering Research Board, Department of Science and Technology (SERB-DST) Govt. of India, New Delhi, vide Project Grant No: TAR/001213/2018.

Acknowledgments

HQ acknowledges fellowship from the Indian Council of Medical Research ICMR-SRF (2021–9917). The authors are thankful to the SERB-DST Govt. of India for financial support.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

Aagaard, K., Petrosino, J., Keitel, W., Watson, M., Katancik, J., Garcia, N., et al. (2013). The Human Microbiome Project strategy for comprehensive sampling of the human microbiome and why it matters. *FASEB J.* 27, 1012. doi: 10.1096/fj.12-220806

Abou-Donia, M. B., El-Masry, E. M., Abdel-Rahman, A. A., McLendon, R. E., and Schiffman, S. S. (2008). Environmental Health, Splenda alters gut microflora and increases intestinal p-glycoprotein and cytochrome p-450 in male rats. *J. Toxicol. Environ. Health A.* 71, 1415–1429. doi: 10.1080/15287390802328630

Agunloye, O. M., Oboh, G., Ademiluyi, A. O., Ademosun, A. O., Akindahunsi, A. A., Oyagbemi, A. A., et al. (2019). Cardio-protective and antioxidant properties of caffeic acid and chlorogenic acid: Mechanistic role of angiotensin converting enzyme, cholinesterase and arginase activities in cyclosporine induced hypertensive rats. *Biomed. Pharmacother.* 109, 450–458. doi: 10.1016/j.biopha.2018.10.044

Alexander, C., and Rietschel, E. T. (2001). Invited review: Bacterial lipopolysaccharides and innate immunity. J Endotoxin Res. 7, 167–202. doi: 10.1177/09680519010070030101

Alexander, J. L., Wilson, I. D., Teare, J., Marchesi, J. R., Nicholson, J. K., and Kinross, J. M. (2017). Gut microbiota modulation of chemotherapy efficacy and toxicity. *Nat. Rev. Gastroenterol. Hepatol.* 14, 356–365. doi: 10.1038/nrgastro.2017.20

Amabebe, E., Robert, F. O., Agbalalah, T., and Orubu, E. S. (2020). Microbial dysbiosis-induced obesity: role of gut microbiota in homoeostasis of energy metabolism. *Br. J. Nutr.* 123, 1127–1137. doi: 10.1017/S0007114520000380

Arumugam, M., Raes, J., Pelletier, E., Le Paslier, D., Yamada, T., Mende, D. R., et al. (2011). Enterotypes of the human gut microbiome. *Nature* 473, 174–180. doi: 10.1038/ nature09944

Bäckhed, F., Ding, H., Wang, T., Hooper, L. V., Koh, G. Y., Nagy, A., et al. (2004). The gut microbiota as an environmental factor that regulates fat storage. *Proc. Natl. Acad. Sci.* 101, 15718–15723. doi: 10.1073/pnas.0407076101

Bajaj, J. S., Heuman, D. M., Hylemon, P. B., Sanyal, A. J., White, M. B., Monteith, P., et al. (2014). Altered profile of human gut microbiome is associated with cirrhosis and its complications. *J. Hepatol.* 60, 940–947. doi: 10.1016/j.jhep.2013.12.019

Balagopal, A., Philp, F. H., Astemborski, J., Block, T. M., Mehta, A., Long, R., et al. (2008). Human immunodeficiency virus-related microbial translocation and progression of hepatitis C. *Gastroenterology* 135, 226–233. doi: 10.1053/j.gastro.2008.03.022

Bansal, T., Alaniz, R. C., Wood, T. K., and Jayaraman, A. (2010). The bacterial signal indole increases epithelial-cell tight-junction resistance and attenuates indicators of inflammation. *Proc. Natl. Acad. Sci.* 107, 228–233. doi: 10.1073/pnas.0906112107

Basholli-Salihu, M., Schuster, R., Mulla, D., Praznik, W., Viernstein, H., Mueller, M. J. B., et al. (2016). Bioconversion of piceid to resveratrol by selected probiotic cell extracts. *Bioprocess Biosyst. Eng.* 39, 1879–1885. doi: 10.1007/s00449-016-1662-1

Bennet, S. M., Böhn, L., Störsrud, S., Liljebo, T., Collin, L., Lindfors, P., et al. (2018). Multivariate modelling of faecal bacterial profiles of patients with IBS predicts responsiveness to a diet low in FODMAPs. *Gut* 67, 872–881. doi: 10.1136/gutjnl-2016-313128

Benson, A. K. (2015). Host genetic architecture and the landscape of microbiome composition: humans weigh in. *Genome Biol.* 16, 1–4. doi: 10.1186/s13059-015-0775-1

Benson, A. K., Kelly, S. A., Legge, R., Ma, F., Low, S. J., Kim, J., et al. (2010). Individuality in gut microbiota composition is a complex polygenic trait shaped by multiple environmental and host genetic factors. *Proc. Natl. Acad. Sci.* 107, 18933– 18938. doi: 10.1073/pnas.1007028107

Bian, X., Chi, L., Gao, B., Tu, P., Ru, H., and Lu, K. (2017). Gut microbiome response to sucralose and its potential role in inducing liver inflammation in mice. *Front, Physiol.* 8, 487. doi: 10.3389/fphys.2017.00487

Blaser, M. J. (2006). Who are we? Indigenous microbes and the ecology of human diseases. *EMBO reports* 7, 956–960. doi: 10.1038/sj.embor.7400812

Blaser, M. J. (2016). Antibiotic use and its consequences for the normal microbiome. *Science* 352, 544–545. doi: 10.1126/science.aad9358

Bonder, M. J., Kurilshikov, A., Tigchelaar, E. F., Mujagic, Z., Imhann, F., Vila, A. V., et al. (2016a). The effect of host genetics on the gut microbiome. *Nat. Genet.* 48, 1407–1412. doi: 10.1038/ng.3663

Bonder, M. J., Tigchelaar, E. F., Cai, X., Trynka, G., Cenit, M. C., Hrdlickova, B., et al. (2016b). The influence of a short-term gluten-free diet on the human gut microbiome. *Genome Med.* 8, 1–11. doi: 10.1186/s13073-016-0295-y

Bongers, G., Pacer, M. E., Geraldino, T. H., Chen, L., He, Z., Hashimoto, D., et al. (2014). Interplay of host microbiota, genetic perturbations, and inflammation promotes local development of intestinal neoplasms in mice. *J. Exp. Med.* 211, 457–472. doi: 10.1084/jem.20131587

Bouskra, D., Brézillon, C., Bérard, M., Werts, C., Varona, R., Boneca, I. G., et al. (2008). Lymphoid tissue genesis induced by commensals through NOD1 regulates intestinal homeostasis. *Nature* 456, 507–510. doi: 10.1038/nature07450

Braune, A., and Blaut, M. (2012). Intestinal bacterium Eubacterium cellulosolvens deglycosylates flavonoid C-and O-glucosides. *Appl. Environ. Microbiol.* 78, 8151–8153. doi: 10.1128/AEM.02115-12

Braune, A., Engst, W., and Blaut, M. (2016). Identification and functional expression of genes encoding flavonoid O-and C-glycosidases in intestinal bacteria. *Environ. Microbiol.* 18, 2117–2129. doi: 10.1111/1462-2920.12864

Brinkman, B., Hildebrand, F., Kubica, M., Goosens, D., Del Favero, J., Declercq, W., et al. (2011). Caspase deficiency alters the murine gut microbiome. *Cell Death Dis.* 2. doi: 10.1038/cddis.2011.101

Brown, J. M., and Hazen, S. L. (2017). Targeting of microbe-derived metabolites to improve human health: the next frontier for drug discovery. *J. Biol. Chem.* 292, 8560–8568. doi: 10.1074/jbc.R116.765388

Browne, H. P., Forster, S. C., Anonye, B. O., Kumar, N., Neville, B. A., Stares, M. D., et al. (2016). Culturing of 'unculturable'human microbiota reveals novel taxa and extensive sporulation. *Nature* 533, 543–546. doi: 10.1038/nature17645

Burapan, S., Kim, M., and Han, J. (2017). Curcuminoid demethylation as an alternative metabolism by human intestinal microbiota. *J. Agric. Food Chem.* 65, 3305–3310. doi: 10.1021/acs.jafc.7b00943

Candeliere, F., Raimondi, S., Ranieri, R., Musmeci, E., Zambon, A., Amaretti, A., et al. (2022). β -Glucuronidase pattern predicted from gut metagenomes indicates potentially diversified pharmacomicrobiomics. *Front. Microbiol.* 13, 826994. doi: 10.3389/fmicb.2022.826994

Cantley, A. M., and Clardy, J. (2015). Animals in a bacterial world: opportunities for chemical ecology. *Nat. Prod. Rep.* 32, 888-892. doi: 10.1039%2Fc4np00141a

Chassaing, B., Koren, O., Goodrich, J. K., Poole, A. C., Srinivasan, S., Ley, R. E., et al. (2015). Dietary emulsifiers impact the mouse gut microbiota promoting colitis and metabolic syndrome. *Nature* 519, 92–96. doi: 10.1038/nature14232

Chen, J.-W., Kong, Z.-L., Tsai, M.-L., Lo, C.-Y., Ho, C.-T., and Lai, C.-S. (2018). *Tetrahydrocurcumin ameliorates* free fatty acid-induced hepatic steatosis and improves insulin resistance in HepG2 cells. *J. Food Drug Anal.* 26, 1075–1085. doi: 10.1016/ j.jfda.2018.01.005

Chen, Y., Yang, F., Lu, H., Wang, B., Chen, Y., Lei, D., et al. (2011). Characterization of fecal microbial communities in patients with liver cirrhosis. *Hepatology* 54, 562–572. doi: 10.1002/hep.24423

Cheng, J.-R., Liu, X.-M., Chen, Z.-Y., Zhang, Y.-S., and Zhang, Y.-H. (2016). Mulberry anthocyanin biotransformation by intestinal probiotics. *Food Chem.* 213, 721–727. doi: 10.1016/j.foodchem.2016.07.032

Chu, J., Vila-Farres, X., Inoyama, D., Ternei, M., Cohen, L. J., Gordon, E. A., et al. (2016). Discovery of MRSA active antibiotics using primary sequence from the human microbiome. *Nat. Chem. Biol.* 12, 1004–1006. doi: 10.1038/nchembio.2207

Cimermancic, P., Medema, M. H., Claesen, J., Kurita, K., Brown, L. C. W., Mavrommatis, K., et al. (2014). Insights into secondary metabolism from a global analysis of prokaryotic biosynthetic gene clusters. *Cell* 158, 412–421. doi: 10.1016/j.cell.2014.06.034

Cohen, L. J., Esterhazy, D., Kim, S.-H., Lemetre, C., Aguilar, R. R., Gordon, E. A., et al. (2017). Commensal bacteria make GPCR ligands that mimic human signalling molecules. *Nature* 549, 48–53. doi: 10.1038/nature23874

Cohen, L. J., Kang, H.-S., Chu, J., Huang, Y.-H., Gordon, E. A., Reddy, B. V. B., et al. (2015). Functional metagenomic discovery of bacterial effectors in the human microbiome and isolation of commendamide, a GPCR G2A/132 agonist. *Proc. Natl. Acad. Sci.* 112, E4825–E4834. doi: 10.1073/pnas.1508737112

Costea, P. I., Hildebrand, F., Arumugam, M., Bäckhed, F., Blaser, M. J., Bushman, F. D., et al. (2018). Enterotypes in the landscape of gut microbial community composition. *Nat. Microbiol.* 3, 8–16. doi: 10.1038/s41564-017-0072-8

Coughlan, L. M., Cotter, P. D., Hill, C., and Alvarez-Ordóñez, A. (2015). Biotechnological applications of functional metagenomics in the food and pharmaceutical industries. *Front. Microbiol.* 6, 149339. doi: 10.3389/fmicb.2015.00672

Crawford, J. M., and Clardy, J. (2011). Bacterial symbionts and natural products. Chem. Commun. 47, 7559–7566. doi: 10.1039/c1cc11574j

De Filippo, C., Cavalieri, D., Di Paola, M., Ramazzotti, M., Poullet, J. B., Massart, S., et al. (2010). Impact of diet in shaping gut microbiota revealed by a comparative study in children from Europe and rural Africa. *Proc. Natl. Acad. Sci.* 107, 14691–14696. doi: 10.1073/pnas.1005963107

Dempsey, E., and Corr, S. C. (2022). Lactobacillus spp. for gastrointestinal health: Current and future perspectives. *Front. Immunol.* 13, 840245. doi: 10.3389/ fimmu.2022.840245

de Vos, W. M., Tilg, H., Van Hul, M., and Cani, P. D. (2022). Gut microbiome and health: mechanistic insights. *Gut* 71, 1020–1032. doi: 10.1136/gutjnl-2021-326789

Donia, M. S., Cimermancic, P., Schulze, C. J., Brown, L. C. W., Martin, J., Mitreva, M., et al. (2014). A systematic analysis of biosynthetic gene clusters in the human microbiome reveals a common family of antibiotics. *Cell* 158, 1402–1414. doi: 10.1016/j.cell.2014.08.032

Donia, M. S., and Fischbach, M. A. (2015). Small molecules from the human microbiota. *Science* 349, 1254766. doi: 10.1126/science.1254766

Falony, G., Joossens, M., Vieira-Silva, S., Wang, J., Darzi, Y., Faust, K., et al. (2016). Population-level analysis of gut microbiome variation. *Science* 352, 560–564. doi: 10.1126/science.aad3503

Fan, Y., and Pedersen, O. (2021). Gut microbiota in human metabolic health and disease. *Nat. Rev. Microbiol.* 19, 55–71. doi: 10.1038/s41579-020-0433-9

Fang, C., Kim, H., Yanagisawa, L., Bennett, W., Sirven, M. A., Alaniz, R. C., et al. (2019). Gallotannins and Lactobacillus plantarum WCFS1 mitigate high-fat dietinduced inflammation and induce biomarkers for thermogenesis in adipose tissue in gnotobiotic mice. *Mol. Nutr. Food Res.* 63, 1800937. doi: 10.1002/mnfr.201800937

Ferreira-Lazarte, A., Plaza-Vinuesa, L., de Las Rivas, B., Villamiel, M., Muñoz, R., and Moreno, F. J. (2021). Production of α -rhamnosidases from *Lactobacillus plantarum* WCFS1 and their role in deglycosylation of dietary flavonoids naringin and rutin. *Int. J. Biol. Macromol.* 193, 1093–1102. doi: 10.1016/j.ijbiomac.2021.11.053

Flint, H. J., Scott, K. P., Louis, P., and Duncan, S. H. (2012). The role of the gut microbiota in nutrition and health. *Nat. Rev. Gastroenterol. Hepatol.* 9, 577–589. doi: 10.1038/nrgastro.2012.156

Foster-Nyarko, E., and Pallen, M. J. (2022). The microbial ecology of Escherichia coli in the vertebrate gut. *FEMS Microbiol. Rev.* 46, fuac008. doi: 10.1093/femsre/fuac008

Gall, M., Thomsen, M., Peters, C., Pavlidis, I. V., Jonczyk, P., Grünert, P. P., et al. (2014). Enzymatic conversion of flavonoids using bacterial chalcone isomerase and enoate reductase. *Angew. Chem. Int. Ed.* 53, 1439–1442. doi: 10.1002/anie.201306952

Garrett, W. S., Lord, G. M., Punit, S., Lugo-Villarino, G., Mazmanian, S. K., Ito, S., et al. (2007). Communicable ulcerative colitis induced by T-bet deficiency in the innate immune system. *Cell* 131, 33–45. doi: 10.1016/j.cell.2007.08.017

Ghosh, S., Whitley, C. S., Haribabu, B., and Jala, V. R. (2021). Regulation of intestinal barrier function by microbial metabolites. *Cell. Mol. Gastroenterol. Hepatol.* 11, 1463–1482. doi: 10.1016/j.jcmgh.2021.02.007

Gibson, P. R. (2017). The evidence base for efficacy of the low FODMAP diet in irritable bowel syndrome: is it ready for prime time as a first-line therapy? J. Gastroenterol. Hepatol. 32, 32–35. doi: 10.1111/jgh.13693

Gong, Z., Huang, J., Xu, B., Ou, Z., Zhang, L., Lin, X., et al. (2019). Urolithin A attenuates memory impairment and neuroinflammation in APP/PS1 mice. J. Neuroinflammation 16, 1–13. doi: 10.1186/s12974-019-1450-3

Goodrich, J. K., Waters, J. L., Poole, A. C., Sutter, J. L., Koren, O., Blekhman, R., et al. (2014). Human genetics shape the gut microbiome. *Cell.* 159, 789–799. doi: 10.1016/j.cell.2014.09.053

Gormley, B. (2016). Microbiome companies attract big investments. *Wall Str. Lett.* 18, 2016.

Gorvitovskaia, A., Holmes, S. P., and Huse, S. M. (2016). Interpreting *Prevotella* and *Bacteroides* as biomarkers of diet and lifestyle. *Microbiome* 4, 1–12. doi: 10.1186/ s40168-016-0160-7

Guo, C.-J., Chang, F.-Y., Wyche, T. P., Backus, K. M., Acker, T. M., Funabashi, M., et al. (2017). Discovery of reactive microbiota-derived metabolites that inhibit host proteases. *Cell* 168, 517–526. e18. doi: 10.1016/j.cell.2016.12.021

Guo, L., Katiyo, W., Lu, L., Zhang, X., Wang, M., Yan, J., et al. (2018). Glycyrrhetic acid 3-O-mono- β -d-glucuronide (GAMG): an innovative high-potency sweetener with improved biological activities. *Compr. Rev. Food Sci. Food Saf.* 17, 905–919. doi: 10.1111/1541-4337.12353

Halmos, E. P. (2017). When the low FODMAP diet does not work. J. Gastroenterol. Hepatol. 32, 69–72. doi: 10.1111/jgh.13701

Han, D. H., Lee, Y., and Ahn, J.-H. (2016). Biological synthesis of baicalein derivatives using Escherichia coli. *J. Microbiol. Biotechnol.* 26, 1918–1923. doi: 10.4014/jmb.1605.05050

Hashimoto, T., Perlot, T., Rehman, A., Trichereau, J., Ishiguro, H., Paolino, M., et al. (2012). ACE2 links amino acid malnutrition to microbial ecology and intestinal inflammation. *Nature* 487, 477–481. doi: 10.1038/nature11228

Hassaninasab, A., Hashimoto, Y., Tomita-Yokotani, K., and Kobayashi, M. (2011). Discovery of the curcumin metabolic pathway involving a unique enzyme in an intestinal microorganism. *Proc. Natl. Acad. Sci.* 108, 6615–6620. doi: 10.1073/pnas.1016217108

Hatamipour, M., Ramezani, M., Tabassi, S. A. S., Johnston, T. P., and Sahebkar, A. (2019). Demethoxycurcumin: A naturally occurring curcumin analogue for treating non-cancerous diseases. *J. Cell. Physiol.* 234, 19320–19330. doi: 10.1002/jcp.28626

He, B.-L., Xiong, Y., Hu, T.-G., Zong, M.-H., and Wu, H. (2023). Bifidobacterium spp. as functional foods: A review of current status, challenges, and strategies. *Crit. Rev. Food Sci. Nutr.* 63, 8048–8065. doi: 10.1080/10408398.2022.2054934

Hehemann, J.-H., Correc, G., Barbeyron, T., Helbert, W., Czjzek, M., and Michel, G. (2010). Transfer of carbohydrate-active enzymes from marine bacteria to Japanese gut microbiota. *Nature* 464, 908–912. doi: 10.1038/nature08937

Henao-Mejia, J., Elinav, E., Jin, C., Hao, L., Mehal, W. Z., Strowig, T., et al. (2012). Inflammasome-mediated dysbiosis regulates progression of NAFLD and obesity. *Nature* 482, 179–185. doi: 10.1038/nature10809

Heng, Y., Kim, M. J., Yang, H. J., Kang, S., and Park, S. (2019). Lactobacillus intestinalis efficiently produces equol from daidzein and chungkookjang, short-term fermented soybeans. *Arch. Microbiol.* 201, 1009–1017. doi: 10.1007/s00203-019-01665-5

Hidalgo-Cantabrana, C., Delgado, S., Ruiz, L., Ruas-Madiedo, P., Sánchez, B., and Margolles, A. (2018). "Bifidobacteria and their health-promoting effects," in *Bugs as Drugs: Therapeutic Microbes for the Prevention and Treatment of Disease*, 73–98. doi: 10.1128/9781555819705.ch3

Hildebrand, F., Nguyen, T. L. A., Brinkman, B., Yunta, R. G., Cauwe, B., Vandenabeele, P., et al. (2013). Inflammation-associated enterotypes, host genotype, cage and inter-individual effects drive gut microbiota variation in common laboratory mice. *Genome Biol.* 14, 1–15. doi: 10.1186/gb-2013-14-1-r4

Hillman, E. T., Lu, H., Yao, T., and Nakatsu, C. H. (2017). Microbial ecology along the gastrointestinal tract. *Microbes Environ*. 32, 300–313. doi: 10.1264/jsme2.ME17017

Hofer, U., Schlaepfer, E., Baenziger, S., Nischang, M., Regenass, S., Schwendener, R., et al. (2010). Inadequate clearance of translocated bacterial products in HIV-infected humanized mice. *PLoS Pathog.* 6. doi: 10.1371/journal.ppat.1000867

Hou, K., Wu, Z.-X., Chen, X.-Y., Wang, J.-Q., Zhang, D., Xiao, C., et al. (2022). Microbiota in health and diseases. *Signal Transduct. Target. Ther.* 7, 135. doi: 10.1038/ s41392-022-00974-4

Hsiao, E. Y., McBride, S. W., Hsien, S., Sharon, G., Hyde, E. R., McCue, T., et al. (2013). Microbiota modulate behavioral and physiological abnormalities associated with neurodevelopmental disorders. *Cell* 155, 1451–1463. doi: 10.1016/j.cell.2013.11.024

Huang, W.-C., Liou, C.-J., Shen, S.-C., Hu, S., Chao, J. C., Hsiao, C.-Y., et al. (2022). Urolithin A inactivation of TLR3/TRIF signaling to block the NF-κB/STAT1 axis reduces inflammation and enhances antioxidant defense in poly (I: C)-induced RAW264. 7 cells. *Int. J. Mol. Sci.* 23, 4697. doi: 10.3390/ijms23094697

Huse, S. M., Ye, Y., Zhou, Y., and Fodor, A. A. (2012). A core human microbiome as viewed through 16S rRNA sequence clusters. *PloS one* 7. doi: 10.1371/journal.pone.0034242

Igartua, C., Davenport, E. R., Gilad, Y., Nicolae, D. L., Pinto, J., and Ober, C. (2017). Host genetic variation in mucosal immunity pathways influences the upper airway microbiome. *Microbiome* 5, 1–17. doi: 10.1186/s40168-016-0227-5

Jackson, M. A., Goodrich, J. K., Maxan, M.-E., Freedberg, D. E., Abrams, J. A., Poole, A. C., et al. (2016). Proton pump inhibitors alter the composition of the gut microbiota. *Gut* 65, 749–756. doi: 10.1136/gutjnl-2015-310861

Jiang, W., Lederman, M. M., Hunt, P., Sieg, S. F., Haley, K., Rodriguez, B., et al. (2009). Plasma levels of bacterial DNA correlate with immune activation and the magnitude of immune restoration in persons with antiretroviral-treated HIV infection. *J. Infect. Dis.* 199, 1177–1185. doi: 10.1086/597476

Jiménez, N., Esteban-Torres, M., Mancheño, J. M., de Las Rivas, B., and Muñoz, R. (2014). Tannin degradation by a novel tannase enzyme present in some *Lactobacillus plantarum* strains. *Appl. Environ. Microbiol.* 80, 2991–2997. doi: 10.1128/AEM.00324-14

Jostins, L., Ripke, S., Weersma, R. K., Duerr, R. H., McGovern, D. P., Hui, K. Y., et al. (2012). Host-microbe interactions have shaped the genetic architecture of inflammatory bowel disease. *Nature* 491, 119–124. doi: 10.1038/nature11582

Kane, M., Case, L. K., Kopaskie, K., Kozlova, A., MacDearmid, C., Chervonsky, A. V., et al. (2011). Successful transmission of a retrovirus depends on the commensal microbiota. *Science* 334, 245–249. doi: 10.1126/science.1210718

Katz, L., and Baltz, R. H. (2016). Natural product discovery: past, present, and future. J. Ind. Microbiol. Biotechnol. 43, 155–176. doi: 10.1007/s10295-015-1723-5

Kau, A. L., Ahern, P. P., Griffin, N. W., Goodman, A. L., and Gordon, J. I. (2011). Human nutrition, the gut microbiome and the immune system. *Nature* 474, 327–336. doi: 10.1038/nature10213

Kho, Z. Y., and Lal, S. K. (2018). The human gut microbiome-a potential controller of wellness and disease. *Front. Microbiol.* 9, 356589. doi: 10.3389/fmicb.2018.01835

Kim, D.-H. (2018). Gut microbiota-mediated pharmacokinetics of ginseng saponins. J. Ginseng Res. 42, 255–263. doi: 10.1016/j.jgr.2017.04.011

Kim, K.-A., Jung, I.-H., Park, S.-H., Ahn, Y.-T., Huh, C.-S., and Kim, D.-H. (2013). Comparative analysis of the gut microbiota in people with different levels of ginsenoside Rb1 degradation to compound K. *PloS one* 8. doi: 10.1371/ journal.pone.0062409

Kim, M., Kim, N., and Han, J. (2014). Metabolism of Kaempferia parviflora polymethoxyflavones by human intestinal bacterium Bautia sp. MRG-PMF1. J. Agric. Food Chem. 62, 12377–12383. doi: 10.1021/jf504074n

Kim, J.-S., Lee, H., Nirmala, F. S., Jung, C. H., Kim, M. J., Jang, Y.-J., et al. (2019). Dihydrodaidzein and 6-hydroxydaidzein mediate the fermentation-induced increase of antiosteoporotic effect of soybeans in ovariectomized mice. *FASEB J.* 33, 3252–3263. doi: 10.1096/fj.201800953R

Knights, D., Silverberg, M. S., Weersma, R. K., Gevers, D., Dijkstra, G., Huang, H., et al. (2014). Complex host genetics influence the microbiome in inflammatory bowel disease. *Genome Med.* 6, 1–11. doi: 10.1186/s13073-014-0107-1

Koppel, N., Maini Rekdal, V., and Balskus, E. P. (2017). Chemical transformation of xenobiotics by the human gut microbiota. *Science* 356. doi: 10.1126/science.aag2770

Kubinak, J. L., Stephens, W. Z., Soto, R., Petersen, C., Chiaro, T., Gogokhia, L., et al. (2015). MHC variation sculpts individualized microbial communities that control susceptibility to enteric infection. *Nat. Commun.* 6, 8642. doi: 10.1038/ncomms9642

Kumar, K., Jaiswal, S. K., Dhoke, G. V., Srivastava, G. N., Sharma, A. K., and Sharma, V. K. (2018). Mechanistic and structural insight into promiscuity based metabolism of cardiac drug digoxin by gut microbial enzyme. *J. Cell. Biochem.* 119, 5287–5296. doi: 10.1002/jcb.26638

Kwan, B. C.-H., Chow, K.-M., Leung, C.-B., Law, M.-C., Cheng, P. M.-S., Yu, V., et al. (2013). Circulating bacterial-derived DNA fragments as a marker of systemic inflammation in peritoneal dialysis. *Nephrol. Dial. Transplant.* 28, 2139–2145. doi: 10.1093/ndt/gft100

Lakhdari, O., Cultrone, A., Tap, J., Gloux, K., Bernard, F., Ehrlich, S. D., et al. (2010). Correction: functional metagenomics: A high throughput screening method to decipher microbiota-driven NF- κ B modulation in the human gut. *PloS one* 5, e13092. doi: 10.1371/annotation/4ea12169-7c97-497c-a45f-52203543065f

Lannes-Costa, P., De Oliveira, J., da Silva Santos, G., and Nagao, P. E. (2021). A current review of pathogenicity determinants of Streptococcus sp. J. Appl. Microbiol. 131, 1600–1620. doi: 10.1111/jam.15090

Law, J. W.-F., Law, L. N.-S., Letchumanan, V., Tan, L. T.-H., Wong, S. H., Chan, K.-G., et al. (2020). Anticancer drug discovery from microbial sources: The unique mangrove streptomycetes. *Molecules* 25, 5365. doi: 10.3390/molecules25225365

Lepage, P., Häsler, R., Spehlmann, M. E., Rehman, A., Zvirbliene, A., Begun, A., et al. (2011). Twin study indicates loss of interaction between microbiota and mucosa of patients with ulcerative colitis. *Gastroenterology* 141, 227–236. doi: 10.1053/j.gastro.2011.04.011

Leung, R. K.-K., and Wu, Y.-K. (2015). Circulating microbial RNA and health. Sci. Rep. 5, 16814. doi: 10.1038/srep16814

Lewis, K., Epstein, S., D'onofrio, A., and Ling, L. L. (2010). Uncultured microorganisms as a source of secondary metabolites. J. Antibiot. 63, 468-476. doi: 10.1038/ja.2010.87

Ley, R. E., Hamady, M., Lozupone, C., Turnbaugh, P. J., Ramey, R. R., Bircher, J. S., et al. (2008). Evolution of mammals and their gut microbes. *Science* 320, 1647–1651. doi: 10.1126/science.1155725

Ley, R. E., Turnbaugh, P. J., Klein, S., and Gordon, J. I. (2006). Human gut microbes associated with obesity. *Science* 444, 1022–1023. doi: 10.1038/4441022a

Li, D., Shi, G., Wang, J., Zhang, D., Pan, Y., Dou, H., et al. (2019). and Therapy, Baicalein ameliorates pristane-induced *lupus nephritis* via activating Nrf2/HO-1 in myeloid-derived suppressor cells. *Arthritis Res. Ther.* 21, 1–14. doi: 10.1186/s13075-019-1876-0

Li, B.-C., Zhang, T., Li, Y.-Q., and Ding, G.-B. (2019). Target discovery of novel α -L-rhamnosidases from human fecal metagenome and application for biotransformation of natural flavonoid glycosides. *Appl. Biochem. Biotechnol.* 189, 1245–1261. doi: 10.1007/s12010-019-03063-5

Liang, D., Leung, R. K.-K., Guan, W., and Au, W. W. (2018). Involvement of gut microbiome in human health and disease: brief overview, knowledge gaps and research opportunities. *Gut Pathog.* 10, 1–9. doi: 10.1186/s13099-018-0230-4

Liang, C., Tseng, H.-C., Chen, H.-M., Wang, W.-C., Chiu, C.-M., Chang, J.-Y., et al. (2017). Diversity and enterotype in gut bacterial community of adults in Taiwan. *BMC Genomics* 18, 1–11. doi: 10.1186/s12864-016-3261-6

Lindell, A. E., Zimmermann-Kogadeeva, M., and Patil, K. R. (2022). Multimodal interactions of drugs, natural compounds and pollutants with the gut microbiota. *Nat. Rev. Microbiol.* 20, 431–443. doi: 10.1038/s41579-022-00681-5

Liu, M.-Y., Li, M., Wang, X.-L., Liu, P., Hao, Q.-H., and Yu, X.-M. (2013). Study on human intestinal bacterium Blautia sp. AUH-JLD56 for the conversion of arctigenin to (-)-3'-desmethylarctigenin. J. Agric. Food Chem. 61, 12060–12065. doi: 10.1021/ jf403924c

Liu, J., Xu, Y., Yan, M., Yu, Y., and Guo, Y. (2022). 18β-Glycyrrhetinic acid suppresses allergic airway inflammation through NF-κB and Nrf2/HO-1 signaling pathways in asthma mice. *Sci. Rep.* 12, 3121. doi: 10.1038/s41598-022-06455-6

Lloyd-Price, J., Abu-Ali, G., and Huttenhower, C. (2016). The healthy human microbiome. *Genome Med.* 8, 1–11. doi: 10.1186/s13073-016-0307-y

Lu, B., Chen, X., Chen, H., Li, Q., Li, H., Xu, Y., et al. (2022). Demethoxycurcumin mitigates inflammatory responses in lumbar disc herniation via MAPK and NF- κ B pathways *in vivo* and *in vitro*. *Int. Immunopharmacol.* 108, 108914. doi: 10.1016/j.intimp.2022.108914

Manor, O., Dai, C. L., Kornilov, S. A., Smith, B., Price, N. D., Lovejoy, J. C., et al. (2020). Health and disease markers correlate with gut microbiome composition across thousands of people. *Nat. Commun.* 11, 5206. doi: 10.1038/s41467-020-18871-1

Marchesi, J. R., Adams, D. H., Fava, F., Hermes, G. D., Hirschfield, G. M., Hold, G., et al. (2016). The gut microbiota and host health: a new clinical frontier. *Gut* 65, 330–339. doi: 10.1136/gutjnl-2015-309990

Marchetti, G., Tincati, C., and Silvestri, G. (2013). Microbial translocation in the pathogenesis of HIV infection and AIDS. *Clin. Microbiol. Rev.* 26, 2–18. doi: 10.1128/CMR.00050-12

Marsh, J. W., Kirk, C., and Ley, R. E. (2023). Toward Microbiome Engineering: expanding the repertoire of genetically tractable members of the human gut Microbiome. *Annu. Rev. Microbiol.* 77, 427-449. doi: 10.1146/annurev-micro-032421-112304

Maslowski, K. M., Vieira, A. T., Ng, A., Kranich, J., Sierro, F., Yu, D., et al. (2009). Regulation of inflammatory responses by gut microbiota and chemoattractant receptor GPR43. *Nature* 461, 1282–1286. doi: 10.1038/nature08530

McIntosh, K., Reed, D. E., Schneider, T., Dang, F., Keshteli, A. H., De Palma, G., et al. (2017). FODMAPs alter symptoms and the metabolome of patients with IBS: a randomised controlled trial. *Gut* 66, 1241–1251. doi: 10.1136/gutjnl-2015-311339

McKnite, A. M., Perez-Munoz, M. E., Lu, L., Williams, E. G., Brewer, S., Andreux, P. A., et al. (2012). Murine gut microbiota is defined by host genetics and modulates variation of metabolic traits. *PloS one* 7. doi: 10.1371/journal.pone.0039191

Medema, M. H., and Fischbach, M. A. (2015). Computational approaches to natural product discovery. Nat. Chem. Biol. 11, 639-648. doi: 10.1038/nchembio.1884

Medema, M. H., Takano, E., and Breitling, R. (2013). Detecting sequence homology at the gene cluster level with MultiGeneBlast. *Mol. Biol. Evol.* 30, 1218–1223. doi: 10.1093/molbev/mst025

Meng, X., Zhang, G., Cao, H., Yu, D., Fang, X., de Vos, W. M., et al. (2020). Gut dysbacteriosis and intestinal disease: mechanism and treatment. *J. Appl. Microbiol.* 129, 787–805. doi: 10.1111/jam.14661

Methé, BA, Nelson, KE, Pop, M, Creasy, HH, Giglio, MG, Huttenhower, C, et al (2012). A framework for human microbiome research. *Nature* 486, 215–221. doi: 10.1038%2Fnature11209

Milshteyn, A., Colosimo, D. A., and Brady, S. F. (2018). Accessing bioactive natural products from the human microbiome. *Cell Host Microbe* 23, 725–736. doi: 10.1016/j.chom.2018.05.013

Miyake, S., Ngugi, D. K., and Stingl, U. (2015). Diet strongly influences the gut microbiota of surgeonfishes. *Mol. Ecol.* 24, 656–672. doi: 10.1111/mec.13050

Moeller, A. H., Degnan, P. H., Pusey, A. E., Wilson, M. L., Hahn, B. H., and Ochman, H. (2012). Chimpanzees and humans harbour compositionally similar gut enterotypes. *Nat. Commun.* 3, 1179. doi: 10.1038/ncomms2159

Morgan, E. W., Perdew, G. H., and Patterson, A. D. (2022). Multi-omics strategies for investigating the microbiome in toxicology research. *Toxicol. Sci.* 187, 189–213. doi: 10.1093/toxsci/kfac029

Mousa, W. K., Athar, B., Merwin, N. J., and Magarvey, N. A. (2017). Antibiotics and specialized metabolites from the human microbiota. *Nat. Prod. Rep.* 34, 1302–1331. doi: 10.1039/C7NP00021A

Mukherjee, A., Lordan, C., Ross, R. P., and Cotter, P. D. (2020). Gut microbes from the phylogenetically diverse genus Eubacterium and their various contributions to gut health. *Gut Microbes* 12, 1802866. doi: 10.1080/19490976.2020.1802866

Mullane, K., Lee, C., Bressler, A., Buitrago, M., Weiss, K., Dabovic, K., et al. (2015). Multicenter, randomized clinical trial to compare the safety and efficacy of LFF571 and vancomycin for Clostridium difficile infections. *Antimicrob. Agents Chemother.* 59, 1435–1440. doi: 10.1128/AAC.04251-14

Mutlu, E. A., Gillevet, P. M., Rangwala, H., Sikaroodi, M., Naqvi, A., Engen, P. A., et al. (2012). Colonic microbiome is altered in alcoholism. *Am. J. Physiol. Gastrointest. Liver Physiol.* 302, G966–G978. doi: 10.1152/ajpgi.00380.2011

Mutlu, E., Keshavarzian, A., Engen, P., Forsyth, C. B., Sikaroodi, M., and Gillevet, P. (2009). Intestinal dysbiosis: a possible mechanism of alcohol-induced endotoxemia and alcoholic steatohepatitis in rats. *Alcohol.: Clin. Exp. Res.* 33, 1836–1846. doi: 10.1111/j.1530-0277.2009.01022.x

Nam, Y.-D., Jung, M.-J., Roh, S. W., Kim, M.-S., and Bae, J.-W. (2011). Comparative analysis of Korean human gut microbiota by barcoded pyrosequencing. *PloS one* 6. doi: 10.1371/journal.pone.0022109

Newman, D. J., and Cragg, G. M. (2016). Natural products as sources of new drugs from 1981 to 2014. J. Nat. Prod. 79, 629-661. doi: 10.1021/acs.jnatprod.5b01055

O'Dwyer, D. N., Dickson, R. P., and Moore, B. B. (2016). The lung microbiome, immunity, and the pathogenesis of chronic lung disease. *J. Immun.* 196, 4839–4847. doi: 10.4049/jimmunol.1600279

Oehmcke-Hecht, S., Mandl, V., Naatz, L. T., Dühring, L., Köhler, J., Kreikemeyer, B., et al. (2020). Streptococcus gallolyticus abrogates anti-carcinogenic properties of tannic acid on low-passage colorectal carcinomas. *Sci. Rep.* 10, 4714. doi: 10.1038/s41598-020-61458-5

Pang, C., Zheng, Z., Shi, L., Sheng, Y., Wei, H., Wang, Z., et al. (2016). Caffeic acid prevents acetaminophen-induced liver injury by activating the Keap1-Nrf2 antioxidative defense system. *Free Radical Biol. Med.* 91, 236–246. doi: 10.1016/ j.freeradbiomed.2015.12.024

Peisl, L., Schymanski, E. L., and Wilmes, P. (2018). Dark matter in host-microbiome metabolomics: tackling the unknowns-a review. *Anal. Chim. Acta* 1037, 13–27. doi: 10.1016/j.aca.2017.12.034

Pereira-Caro, G., Fernández-Quirós, B., Ludwig, I. A., Pradas, I., Crozier, A., and Moreno-Rojas, J. M. (2018). Catabolism of citrus flavanones by the probiotics Bifidobacterium longum and Lactobacillus rhamnosus. *Eur. J. Nutr.* 57, 231–242. doi: 10.1007/s00394-016-1312-z

Petrof, E. O., Claud, E. C., Sun, J., Abramova, T., Guo, Y., Waypa, T. S., et al. (2009). Bacteria-free solution derived from Lactobacillus plantarum inhibits multiple NFkappaB pathways and inhibits proteasome function. *Inflamm. Bowel Dis.* 15, 1537– 1547. doi: 10.1002/ibd.20930

Proal, A. D., Albert, P. J., and Marshall, T. G. (2014). Inflammatory disease and the human microbiome. *Discov. Med.* 22, 257–265.

Qin, J., Li, R., Raes, J., Arumugam, M., Burgdorf, K. S., Manichanh, C., et al. (2010). A human gut microbial gene catalogue established by metagenomic sequencing. *Nature* 464, 59–65. doi: 10.1038/nature08821

Qin, J., Li, Y., Cai, Z., Li, S., Zhu, J., Zhang, F., et al. (2012). A metagenome-wide association study of gut microbiota in type 2 diabetes. *Nature* 490, 55–60. doi: 10.1038/ nature11450

Queipo-Ortuño, M. I., Boto-Ordóñez, M., Murri, M., Gomez-Zumaquero, J. M., Clemente-Postigo, M., Estruch, R., et al. (2012). Influence of red wine polyphenols and

ethanol on the gut microbiota ecology and biochemical biomarkers. Am. J. Clin. Nutr. 95, 1323–1334. doi: 10.3945/ajcn.111.027847

Raimondi, S., Anighoro, A., Quartieri, A., Amaretti, A., Tomás-Barberán, F. A., Rastelli, G., et al. (2015). Role of bifidobacteria in the hydrolysis of chlorogenic acid. *Microbiologyopen* 4, 41-52. doi: 10.1002/mbo3.219

Reddy, B. V. B., Milshteyn, A., Charlop-Powers, Z., and Brady, S. F. (2014). eSNaPD: a versatile, web-based bioinformatics platform for surveying and mining natural product biosynthetic diversity from metagenomes. *Chem. Biol.* 21, 1023–1033. doi: 10.1016/j.chembiol.2014.06.007

Relman, D. A. (2012). The human microbiome: ecosystem resilience and health. Nutr. Rev. 70, S2–S9. doi: 10.1111/j.1753-4887.2012.00489.x

Reverón, I., de las Rivas, B., Matesanz, R., Muñoz, R., and López de Felipe, F. (2015). Molecular adaptation of Lactobacillus plantarum WCFS1 to gallic acid revealed by genome-scale transcriptomic signature and physiological analysis. *Microb. Cell Fact.* 14, 1–12. doi: 10.1186/s12934-015-0345-y

Reverón, I., Jiménez, N., Curiel, J. A., Peñas, E., Lopez de Felipe, F., de Las Rivas, B., et al. (2017). Differential gene expression by Lactobacillus plantarum WCFS1 in response to phenolic compounds reveals new genes involved in tannin degradation. *Appl. Environ. Microbiol.* 83. doi: 10.1128/AEM.03387-16

Ridaura, V. K., Faith, J. J., Rey, F. E., Cheng, J., Duncan, A. E., Kau, A. L., et al. (2013). Gut microbiota from twins discordant for obesity modulate metabolism in mice. *Science* 341, 1241214. doi: 10.1126/science.1241214

Rinninella, E., Raoul, P., Cintoni, M., Franceschi, F., Miggiano, G. A. D., Gasbarrini, A., et al. (2019). What is the healthy gut microbiota composition? A changing ecosystem across age, environment, diet, and diseases. *Microorganisms* 7, 14. doi: 10.3390/microorganisms7010014

Rocchetti, G., Gregorio, R. P., Lorenzo, J. M., Barba, F. J., Oliveira, P. G., Prieto, M. A., et al. (2022). Safety, Functional implications of bound phenolic compounds and phenolics-food interaction: A review. *Compr. Rev. Food Sci. Food Saf.* 21, 811–842. doi: 10.1111/1541-4337.12921

Rodríguez-Daza, M. C., Pulido-Mateos, E. C., Lupien-Meilleur, J., Guyonnet, D., Desjardins, Y., and Roy, D. (2021). Polyphenol-mediated gut microbiota modulation: Toward prebiotics and further. *Front. Nutr.* 8, 689456. doi: 10.3389/fnut.2021.689456

Rowland, I., Gibson, G., Heinken, A., Scott, K., Swann, J., Thiele, I., et al. (2018). Gut microbiota functions: metabolism of nutrients and other food components. *Eur. J. Nutr.* 57, 1–24. doi: 10.1007/s00394-017-1445-8

Santos, E. A. A., Schieber, A., and Weber, F. (2018). Site-specific hydrolysis of chlorogenic acids by selected Lactobacillus species. *Int. Food Res.* 109, 426–432. doi: 10.1016/j.foodres.2018.04.052

Satti, M., Modesto, M., Endo, A., Kawashima, T., Mattarelli, P., and Arita, M. (2021). Host-diet effect on the metabolism of bifidobacterium. *Genes* 12, 609. doi: 10.3390/genes12040609

Schnabl, B., and Brenner, D. A. (2014). Interactions between the intestinal microbiome and liver diseases. *Gastroenterology* 146, 1513–1524. doi: 10.1053/j.gastro.2014.01.020

Schnorr, S. L., Candela, M., Rampelli, S., Centanni, M., Consolandi, C., Basaglia, G., et al. (2014). Gut microbiome of the Hadza hunter-gatherers. *Nat. Commun.* 5, 3654. doi: 10.1038/ncomms4654

Shoaie, S., Karlsson, F., Mardinoglu, A., Nookaew, I., Bordel, S., and Nielsen, J. (2013). Understanding the interactions between bacteria in the human gut through metabolic modeling. *Sci. Rep.* 3, 2532. doi: 10.1038/srep02532

Sidebottom, A. M., and Carlson, E. E. (2015). A reinvigorated era of bacterial secondary metabolite discovery. *Curr. Opin. Chem. Biol.* 24, 104–111. doi: 10.1016/j.cbpa.2014.10.014

Smith, P. A. (2015). The tantalizing links between gut microbes and the brain. *Nature* 526, 312–314. doi: 10.1038/526312a

Smith, M. I., Yatsunenko, T., Manary, M. J., Trehan, I., Mkakosya, R., Cheng, J., et al. (2013). Gut microbiomes of Malawian twin pairs discordant for kwashiorkor. *Science* 339, 548–554. doi: 10.1126/science.1229000

Snijders, A. M., Langley, S. A., Kim, Y.-M., Brislawn, C. J., Noecker, C., Zink, E. M., et al. (2016). Influence of early life exposure, host genetics and diet on the mouse gut microbiome and metabolome. *Nat. Microbiol.* 2, 1–8. doi: 10.1038/nmicrobiol.2016.221

Spanogiannopoulos, P., Bess, E. N., Carmody, R. N., and Turnbaugh, P. J. (2016). The microbial pharmacists within us: a metagenomic view of xenobiotic metabolism. *Nat. Rev. Microbiol.* 14, 273–287. doi: 10.1038/nrmicro.2016.17

Tai, N., Wong, F. S., and Wen, L. (2015). The role of gut microbiota in the development of type 1, type 2 diabetes mellitus and obesity. *Rev. Endocr. Metab. Disord.* 16, 55–65. doi: 10.1007/s11154-015-9309-0

Tan, S., Rupasinghe, T. W., Tull, D. L., Boughton, B., Oliver, C., McSweeny, C., et al. (2014). Degradation of curcuminoids by *in vitro* pure culture fermentation. *J. Agric. Food Chem.* 62, 11005–11015. doi: 10.1021/jf5031168

Tanaka, H., Hashiba, H., Kok, J., and Mierau, I. (2000). Bile salt hydrolase of Bifidobacterium longum—biochemical and genetic characterization. *Appl. Environ. Microbiol.* 66, 2502–2512. doi: 10.1128/AEM.66.6.2502-2512.2000

Tanaka, M., Kishimoto, Y., Sasaki, M., Sato, A., Kamiya, T., Kondo, K., et al. (2018). Terminalia bellirica (Gaertn.) Roxb. extract and gallic acid attenuate LPS-induced inflammation and oxidative stress via MAPK/NFrκB and Akt/AMPK/Nrf2 pathways. *Oxid. Med. Cell. Longev.* 2018, 9364364. doi: 10.1155/2018/9364364 Tang, W. W., Kitai, T., and Hazen, S. L. (2017). Gut microbiota in cardiovascular health and disease. Circ. Res. 120, 1183–1196. doi: 10.1161/CIRCRESAHA.117.309715

Tang, J., Tan, X., Huang, X., Zhang, J., Chen, L., Li, A., et al. (2021). Dual targeting of autophagy and NF- κ B pathway by PPAR γ contributes to the inhibitory effect of demethoxycurcumin on NLRP3 inflammasome priming. *Curr. Mol. Pharmacol.* 14, 914–921. doi: 10.2174/1874467214666210301121020

Tremaroli, V., and Bäckhed, F. (2012). Functional interactions between the gut microbiota and host metabolism. *Nature* 489, 242–249. doi: 10.1038/nature11552

Turnbaugh, P. J., and Gordon, J. I. (2009). The core gut microbiome, energy balance and obesity. J. Physiol. 587, 4153–4158. doi: 10.1113/jphysiol.2009.174136

Turroni, F., Marchesi, J. R., Foroni, E., Gueimonde, M., Shanahan, F., Margolles, A., et al. (2009). Microbiomic analysis of the bifidobacterial population in the human distal gut. *ISME J.* 3, 745–751. doi: 10.1038/ismej.2009.19

Turroni, F., Ribbera, A., Foroni, E., van Sinderen, D., and Ventura, M. (2008). Human gut microbiota and bifidobacteria: from composition to functionality. *Antonie Van Leeuwenhoek* 94, 35–50. doi: 10.1007/s10482-008-9232-4

Ursell, L. K., Metcalf, J. L., Parfrey, L. W., and Knight, R. (2012). Defining the human microbiome. *Nutr. Rev.* 70, S38–S44. doi: 10.1111/j.1753-4887.2012.00493.x

Van Baarlen, P., Troost, F. J., van Hemert, S., van der Meer, C., de Vos, W. M., de Groot, P. J., et al. (2009). Differential NF- κ B pathways induction by *Lactobacillus plantarum* in the duodenum of healthy humans correlating with immune tolerance. *Proc. Natl. Acad. Sci.* 106, 2371–2376. doi: 10.1073/pnas.0809919106

Vinarov, Z., Abrahamsson, B., Artursson, P., Batchelor, H., Berben, P., Bernkop-Schnürch, A., et al. (2021). Current challenges and future perspectives in oral absorption research: An opinion of the UNGAP network. *Adv. Drug Deliv. Rev.* 171, 289–331. doi: 10.1016/j.addr.2021.02.001

Walker, A. W., and Hoyles, L. (2023). Human microbiome myths and misconceptions. *Nat. Microbiol.* 8, 1392–1396. doi: 10.1038/s41564-023-01426-7

Wang, X.-L., Kim, H.-J., Kang, S.-I., Kim, S.-I., and Hur, H.-G. (2007). Production of phytoestrogen S-equol from daidzein in mixed culture of two anaerobic bacteria. *Arch. Microbiol.* 187, 155–160. doi: 10.1007/s00203-006-0183-8

Wang, J., Linnenbrink, M., Künzel, S., Fernandes, R., Nadeau, M.-J., Rosenstiel, P., et al. (2014). Dietary history contributes to enterotype-like clustering and functional metagenomic content in the intestinal microbiome of wild mice. *Proc. Natl. Acad. Sci.* 111, E2703–E2710. doi: 10.1073/pnas.1402342111

Wardman, J. F., Bains, R. K., Rahfeld, P., and Withers, S. G. (2022). Carbohydrateactive enzymes (CAZymes) in the gut microbiome. *Nat. Rev. Microbiol.* 20, 542–556. doi: 10.1038/s41579-022-00712-1

Weinstock, G. M. (2012). Genomic approaches to studying the human microbiota. Nature 489, 250–256. doi: 10.1038/nature11553

Wikoff, W. R., Anfora, A. T., Liu, J., Schultz, P. G., Lesley, S. A., Peters, E. C., et al. (2009). Metabolomics analysis reveals large effects of gut microflora on mammalian blood metabolites. *Proc. Natl. Acad. Sci.* 106, 3698–3703. doi: 10.1073/pnas.0812874106

Wilson, I. D., and Nicholson, J. K. (2017). Gut microbiome interactions with drug metabolism, efficacy, and toxicity. *Transl. Res.* 179, 204–222. doi: 10.1016/j.trsl.2016.08.002

Wilson, M. C., and Piel, J. (2013). Metagenomic approaches for exploiting uncultivated bacteria as a resource for novel biosynthetic enzymology. *Chem. Biol.* 20, 636–647. doi: 10.1016/j.chembiol.2013.04.011

Wilson, M. R., Zha, L., and Balskus, E. P. (2017). Natural product discovery from the human microbiome. J. Biol. Chem. 292, 8546–8552. doi: 10.1074/jbc.R116.762906

Wlodarska, M., Luo, C., Kolde, R., d'Hennezel, E., Annand, J. W., Heim, C. E., et al. (2017). Indoleacrylic acid produced by commensal peptostreptococcus species suppresses inflammation. *Cell Host Microbe* 22, 25–37. e6. doi: 10.1016/ j.chom.2017.06.007

Wu, G. D., Chen, J., Hoffmann, C., Bittinger, K., Chen, Y.-Y., Keilbaugh, S. A., et al. (2011). Linking long-term dietary patterns with gut microbial enterotypes. *Science* 334, 105–108. doi: 10.1126/science.1208344

Wu, G. D., Compher, C., Chen, E. Z., Smith, S. A., Shah, R. D., Bittinger, K., et al. (2016). Comparative metabolomics in vegans and omnivores reveal constraints on dietdependent gut microbiota metabolite production. *Gut.* 65, 63–72. doi: 10.1136/gutjnl-2014-308209

Wu, H., Kim, M., and Han, J. (2016). Icariin metabolism by human intestinal microflora. *Molecules* 21, 1158. doi: 10.3390/molecules21091158

Xie, Y., Hu, F., Xiang, D., Lu, H., Li, W., Zhao, A., et al. (2020). The metabolic effect of gut microbiota on drugs. *Drug Metab. Rev.* 52, 139–156. doi: 10.1080/03602532.2020.1718691

Yan, A. W., Fouts, D. E., Brandl, J., Stärkel, P., Torralba, M., Schott, E., et al. (2011). Enteric dysbiosis associated with a mouse model of alcoholic liver disease. *Hepatology* 53, 96–105. doi: 10.1002/hep.24018

Ye, M., Yu, J., Shi, X., Zhu, J., Gao, X., and Liu, W. (2021). and Nutrition, Polysaccharides catabolism by the human gut bacterium-Bacteroides thetaiotaomicron: Advances and perspectives. *Crit. Rev. Food Sci. Nutr.* 61, 3569– 3588. doi: 10.1080/10408398.2020.1803198

Yu, Q., Liu, Y., Wu, Y., and Chen, Y. (2018). Dihydrocurcumin ameliorates the lipid accumulation, oxidative stress and insulin resistance in oleic acid-induced L02 and

HepG2 cells. Biomed. Pharmacother. 103, 1327-1336. doi: 10.1016/ j.biopha.2018.04.143

Zhang, Q., Lenardo, M. J., and Baltimore, D. (2017). 30 years of NF-κB: a blossoming of relevance to human pathobiology. *Cell* 168, 37–57. doi: 10.1016/j.cell.2016.12.012

Zhang, R., Huang, X.-M., Yan, H.-J., Liu, X.-Y., Zhou, Q., Luo, Z.-Y., et al. (2019). Highly selective production of compound K from ginsenoside Rd by hydrolyzing glucose at C-3 glycoside using β -glucosidase of Bifidobacterium breve ATCC. J. Microbiol. Biotechnol. 15700, 410–418. doi: 10.4014/jmb.1808.08059

Zhao, Y., Zhong, X., Yan, J., Sun, C., Zhao, X., and Wang, X. (2022). Potential roles of gut microbes in biotransformation of natural products: An overview. *Front. Microbiol.* 13, 956378. doi: 10.3389/fmicb.2022.956378

Zhong, F.-L., Ma, R., Jiang, M., Dong, W.-W., Jiang, J., Wu, S., et al. (2016). Cloning and characterization of ginsenoside-hydrolyzing β -glucosidase from Lactobacillus

brevis that transforms ginsenosides Rb1 and F2 into ginsenoside Rd and compound K. J. Microbiol. Biotechnol. 26, 1661–1667. doi: 10.4014/jmb.1605.05052

Zoetendal, E. G., Raes, J., Van Den Bogert, B., Arumugam, M., Booijink, C. C., Troost, F. J., et al. (2012). The human small intestinal microbiota is driven by rapid uptake and conversion of simple carbohydrates. *ISME J.* 6, 1415–1426. doi: 10.1038/ ismej.2011.212

Zupancic, M. L., Cantarel, B. L., Liu, Z., Drabek, E. F., Ryan, K. A., Cirimotich, S., et al. (2012). Analysis of the gut microbiota in the old order Amish and its relation to the metabolic syndrome. *Plos one.* doi: 10.1371/journal.pone.0043052

Zvanych, R., Lukenda, N., Kim, J. J., Li, X., Petrof, E. O., Khan, W. I., et al. (2014). Small molecule immunomodulins from cultures of the human microbiome member *Lactobacillus plantarum. J. Antibiot.* 67, 85–88. doi: 10.1038/ja.2013.126