

OPEN ACCESS

EDITED BY Steven Gill, University of Rochester, United States

REVIEWED BY Timothy Richard Hughes, Cardiff University, United Kingdom

George Grant, Independent researcher, Aberdeen, United Kingdom Tianming Yao, Purdue University, United States

*CORRESPONDENCE Hea-Jong Chung Michung84@kbsi.re.kr Seong-Tshool Hong seonghong@jbnu.ac.kr

[†]These authors have contributed equally to this work

RECEIVED 08 September 2023 ACCEPTED 12 August 2024 PUBLISHED 08 October 2024

CITATION

Islam MM, Mahbub NU, Hong S-T and Chung H-J (2024) Gut bacteria: an etiological agent in human pathological conditions. *Front. Cell. Infect. Microbiol.* 14:1291148. doi: 10.3389/fcimb.2024.1291148

COPYRIGHT

© 2024 Islam, Mahbub, Hong and Chung. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Gut bacteria: an etiological agent in human pathological conditions

Md Minarul Islam^{1†}, Nasir Uddin Mahbub^{1†}, Seong-Tshool Hong^{1*} and Hea-Jong Chung^{2*}

¹Department of Biomedical Sciences and Institute for Medical Science, Jeonbuk National University Medical School, Jeonju, Republic of Korea, ²Gwangju Center, Korea Basic Science Institute, Gwangju, Republic of Korea

Through complex interactions with the host's immune and physiological systems, gut bacteria play a critical role as etiological agents in a variety of human diseases, having an impact that extends beyond their mere presence and affects the onset, progression, and severity of the disease. Gaining a comprehensive understanding of these microbial interactions is crucial to improving our understanding of disease pathogenesis and creating tailored treatment methods. Correcting microbial imbalances may open new avenues for disease prevention and treatment approaches, according to preliminary data. The gut microbiota exerts an integral part in the pathogenesis of numerous health conditions, including metabolic, neurological, renal, cardiovascular, and gastrointestinal problems as well as COVID-19, according to recent studies. The crucial significance of the microbiome in disease pathogenesis is highlighted by this role, which is comparable to that of hereditary variables. This review investigates the etiological contributions of the gut microbiome to human diseases, its interactions with the host, and the development of prospective therapeutic approaches. To fully harness the benefits of gut microbiome dynamics for improving human health, future research should address existing methodological challenges and deepen our knowledge of microbial interactions.

KEYWORDS

gut microbiota, probiotic, prebiotic, metabolites, gut-brain axis

1 Introduction

Trillions of microorganisms, including bacteria, fungi, archaea, protozoa, and viruses, reside within the human body, collectively representing a genetic repertoire that encompasses approximately 100-150 times more genes than the host genome (Thomas et al., 2017; Lkhagva et al., 2021b; Afzaal et al., 2022). Alterations in the microbiome's composition within an individual can profoundly affect overall health, with modifications in microbial populations potentially leading to considerable changes in health outcomes

(Voigt et al., 2015; Gacesa et al., 2022). Their composition is modulated by factors such as mode of birth, feeding system, dietary habits, lifestyle, exercise, drug use, and host genetics (Chang and Kao, 2019; Hasan and Yang, 2019; Lkhagva et al., 2021a; Nandwana et al., 2022; Kok et al., 2023).

The microbiota colonizes the human gut immediately after birth (Milani et al., 2017), though some research has also reported that the microbiome may colonize the human gut during the prenatal period (Perez-Munoz et al., 2017; Xiao and Zhao, 2023). The mode of delivery is a critical determinant in this process. In vaginal deliveries, the infant's gut is initially colonized by the maternal gut and vaginal microbiota. Conversely, infants delivered via cesarean section are predominantly exposed to skin and environmental microbes from the hospital setting (Dunn et al., 2017). Facultative anaerobes, such as Escherichia coli, Staphylococcus, and Streptococcus, first colonize the infant gut following a vaginal delivery and, for a few days, create an anaerobic environment that allows the survival of strict anaerobes, such as Bacteroides and Bifidobacterium spp. (Pantoja-Feliciano et al., 2013). Cesarean-section infants are enriched with opportunistic pathogens, including Enterococcus, Enterobacter, and Klebsiella species, and show disturbed transmission of Bacteroides and Bifidobacterium spp. Breastfeeding can partially restore delayed gut microbiota establishment (Shao et al., 2019; Guo et al., 2020). The mode of early feeding (breastfeeding vs. formula feeding) plays a major role in shaping the gut microbiota during the formative years. Bezirtzoglou et al. (Bezirtzoglou et al., 2011) showed that Bifidobacterium species were more than two times higher in breastfeeding infants than in formula-fed infants. Over time, the diversity of the gut microbiota is modulated by a range of factors, including age, sex, dietary intake, pharmacological treatments, physical activity, geographical location, and occupational environment.

The gut microbiome is integral to the maturation of the host immune system, the metabolism of pharmaceuticals, the process of digestion, cognitive function, the neutralization of toxins, the synthesis of essential vitamins, and the establishment of a conducive environment for commensal microorganisms. These functions collectively contribute to pathogen defense and the prevention of severe diseases (Abt and Pamer, 2014; Li et al., 2016a; Mohajeri et al., 2018; Nguyen et al., 2019; Sasso et al., 2023). Probiotic bacteria, often termed "beneficial" or "commensal" microorganisms, are essential for sustaining a balanced and healthful microbial milieu within the digestive system by suppressing pathogenic or "harmful" microbes and regulating the pH of the gut. Dietary fibers and resistant starches, which are not metabolized by endogenous digestive enzymes, are fermented by the gut microbiota to produce short-chain fatty acids (SCFAs) such as acetate, butyrate, and propionate within the colon. SCFAs, particularly butyrate, are vital for maintaining the gut microbiome's equilibrium through various localized effects, including the preservation of intestinal barrier integrity, enhancement of mucus production, attenuation of inflammation, modulation of immune responses, and potential reduction of the detrimental impacts associated with pathogenic microorganisms (Vinolo et al., 2011; Corrêa-Oliveira et al., 2016; Fang et al., 2021; Vinelli et al., 2022). Non-digestible food supplements, called prebiotics, can selectively stimulate the growth of beneficial microbes, such as *Bifidobacterium* and *Lactobacillus* species (Rastall and Gibson, 2015). Prebiotics contribute to the maintenance of a balanced microbial ecosystem in the gut by supplying essential nutrients to beneficial microorganisms. This support enhances digestive function, optimizes nutrient absorption, and bolsters immune function, while also mitigating the risk of gastrointestinal disorders (Guarino et al., 2020; You et al., 2022).

The disruption of the gut microbiota ecosystem, referred to as dysbiosis, alters the normal gut flora and precipitates the onset of diverse pathologies. This disruption leads to an overgrowth of pathogenic bacteria and a decrease in beneficial microbes, which compromises gut barrier integrity and triggers pro-inflammatory responses and dysregulated immune function. Consequently, dysbiosis is implicated in a wide array of diseases, including neurodegenerative disorders, cardiovascular diseases, metabolic syndromes, gastrointestinal disorders, COVID-19, and colorectal carcinoma, as confirmed by both human and animal studies (Degruttola et al., 2016; Hou et al., 2022). In this review, we encapsulate the most recent research on the impact of the gut bacteria on human diseases, focusing on its underlying mechanisms and regulatory aspects. Moreover, we explicate the potential mechanisms of microbial metabolism and its derivatives that facilitate disease development and progression, offering insights into potential targets for future preventive and therapeutic strategies.

2 The impact of the gut bacteria on human health maintenance

Gut microbiota profoundly affects human health by impacting immune system activity, metabolic functions, and the balance of microbial communities essential for physiological stability. The gut microbiota modulates the host's innate and adaptive immune responses through their components and metabolic products. Metabolites produced by the gut microbiota are crucial in maintaining the fundamental functions of the host. Conversely, disruption in the production of these metabolites can contribute to the pathogenesis of various diseases. Short-chain fatty acids (SCFAs) are microbial metabolites that facilitate the differentiation of T cells into effector T cells and regulatory T cells (Park et al., 2015). The peptides B7 and B12, secreted by Bifidobacterium longum and Bacteroides fragilis, respectively, play a role in modulating intestinal cytokine production in patients suffering from inflammatory bowel disease (IBD) (Fernández-Tomé et al., 2019). In an unhealthy infant, the gut fosters a pronounced TH1 profile and a pro-inflammatory immune response, leading to the secretion of IL-12 and interferon (IFN)gamma. This, in turn, causes tissue damage and disrupts normal immune functions (Tamburini et al., 2016). Moreover, the gut microbiome is crucial for the digestion of dietary fiber, as human digestive enzymes are incapable of fermenting non-starch polysaccharides (Holscher, 2017). The short-chain fatty acid butyrate, generated from the microbial fermentation of dietary fiber,

Abbreviations: BFT, *Bacteroides fragilis* toxin; CAD, coronary artery disease; CD, Crohn's disease; CKD, chronic kidney disease; CRC, colorectal cancer; GF, germfree; GI, gastrointestinal tract; IBD, inflammatory bowel disease; IBS, irritable bowel syndrome; PAGln, phenylacetylglutamine; SCFA, short-chain fatty acid; TMAO, trimethylamine N-oxide.

is essential for epithelial cell metabolism and functionality. It also promotes the wound-healing response and fortifies epithelial barrier functions in patients with inflammatory bowel disease (IBD) (Wang et al., 2020). Furthermore, it contributes to the regulation of antiinflammatory responses and shows promise for therapeutic applications in diseases such as intestinal neoplasia and inflammatory bowel disease (IBD) (Salvi and Cowles, 2021). Bifidobacterial strains impede the infection caused by Shiga toxinproducing E. coli O157:H7, leading to increased acetate synthesis from carbohydrates (Fukuda et al., 2011). A study involving germ-free (GF) rats demonstrated impaired intestinal transit and diminished small intestinal contractions compared to conventional rats (Husebye et al., 2001). The enteric glial cell network is pivotal in regulating the function of the intestinal epithelial barrier and modulating the intestinal immune response (Inlender et al., 2021). The gut microbiota plays a critical role in the postnatal development of enteric glial cell networks within the intestinal mucosa and influences the colonization of glial cells in the lamina propria of adult mice (Kabouridis et al., 2015). Gut bacteria such as Bifidobacterium dentium and Akkermansia muciniphila contribute to a healthier gut environment by promoting intestinal mucus production and enhancing goblet cell function, without causing significant degradation of mucin (Engevik et al., 2019; Kim et al., 2021). Additionally, the human gut microbiome participates in the chemical transformation of industrial chemicals and pharmaceuticals into metabolites, which can influence their bioavailability, bioactivity, and toxicity (Koppel et al., 2017). Chemotherapeutic agents, such as doxorubicin, may induce severe gastrointestinal complications, including enteric mucositis and dysbiosis, and the intestinal microbiome has the capacity to modulate these adverse effects (Blaustein et al., 2021). Specifically, the intestinal bacterium Raoultella planticola can detoxify the anticancer drug doxorubicin under anaerobic conditions through glycosylation, thereby enhancing host survival. Similarly, E. coli and Klebsiella pneumoniae strains facilitate the same biotransformation process via analogous biosynthetic pathways, reducing toxicity in the model eukaryote Caenorhabditis elegans by generating metabolites (Yan et al., 2018). Furthermore, the intestinal bacterium Eggerthella lenta can deactivate the cardiac drug digoxin, converting it to dihydrodigoxin (Haiser et al., 2013). These findings underscore the critical role of the gut microbiome in modulating drug metabolism and its potential implications for therapeutic efficacy and safety. The gut bacteria play a pivotal role in the maintenance of human health by influencing a range of physiological processes, including immune system regulation, nutrient metabolism, and disease prevention. Their intricate interactions with the host contribute to overall wellbeing and highlight the importance of maintaining a balanced microbiome for optimal health outcomes.

3 The role of gut bacteria in disease pathogenesis

The human gut bacteria are essential to multiple facets of host physiology and health, with their dysregulation being strongly associated with numerous diseases. The gut microbiome significantly affects gastrointestinal conditions such as inflammatory bowel disease (IBD), irritable bowel syndrome (IBS), chronic constipation, and colorectal cancer (CRC), as well as influencing a spectrum of systemic disorders, including metabolic and neurodegenerative diseases (Figure 1).

3.1 Gastrointestinal disorders and complications associated with gut bacteria

3.1.1 Inflammatory bowel disease

Inflammatory bowel disease (IBD), a chronic inflammation of the GI tract, is an umbrella term used to incorporate ulcerative colitis and Crohn's disease (CD) (Thompson-Chagoyan et al., 2005). The development of IBD is complex and poorly understood due to its multifactorial nature, which involves genetic factors, immune dysfunction, imbalances in gut microbiota, environmental influences, and various aspects of intestinal health (Yuan et al., 2023) (Figure 2). The intestinal microbiota plays a crucial role in IBD, as it involves a compromised immune tolerance that results in an excessive immune reaction to gut bacteria. IBD patients frequently exhibit a modified composition of the gut microbiome (Zuo and Ng, 2018; Zhang et al., 2022). Patients with IBD often show a decrease in gut microbiota diversity, particularly in Firmicutes. This reduction can have adverse effects on mucosal integrity and butyrate production, potentially leading to inflammation and impacting cytokine production in the colon (Fernandes and Andreyev, 2022). In patients with CD, anaerobic gram-positive coccoid rods and gram-negative rods are more prevalent than in their healthy counterparts (Van De Merwe et al., 1988); Bacteroides and Bifidobacteria are poorly associated, whereas Enterobacteriaceae are richest in CD (Seksik et al., 2003). The abundance of beneficial bacteria is significantly decreased in patients with IBD. Kang et al. (Kang et al., 2010) found that anti-inflammatory bacteria Faecalibacterium prausnitzii and beneficial Ruminococcus species are 5-10 times less abundant in patients with inflammatory bowel disease (IBD) compared to the control group. Additionally, pathobionts, which are typically commensal microbes that can become harmful under certain conditions, are often found in higher numbers in individuals with inflammatory bowel disease (IBD) (Nagao-Kitamoto and Kamada, 2017). Gammaproteobacteria become the most abundant when Firmicutes are reduced, and evidence indicates that pathogenic E. coli strains accumulate in the intestinal mucosa of patients with IBD, especially in CD (Kang et al., 2010; Lapaquette et al., 2010; Mirsepasi-Lauridsen et al., 2019). Enterotoxigenic Bacteroides fragilis colonizes patients with IBD, where B. fragilis toxin (BFT) cleaves E-cadherin, reduces mucosal barrier function, and increases epithelial cell proliferation and proinflammatory cytokine IL-8 production (Sears, 2009). A recent study conducted on a population revealed a significant link between the use of antibiotics and the likelihood of developing inflammatory bowel disease (IBD), which includes Crohn's disease and ulcerative colitis. This increased risk was observed across various age groups,



especially in cases where the cumulative duration of antibiotic use exceeded 30 days (Aniwan et al., 2018). Clostridium leptum group bacteria, including Faecalibacterium prausnitzii, were significantly reduced in IBD, shortening the production of SCFAs (Kabeerdoss et al., 2013; Lopez-Siles et al., 2016). Fecal microbiome transplantation might be a potential strategy to fight against IBD. A combination therapy of fecal microbiome with an antiinflammatory diet helped patients with mild to moderate ulcerative colitis feel better and improved their digestive tracts (Kedia et al., 2022). A pilot study investigated the efficacy of multi-session fecal microbiota transplantation (FMT) in managing active ulcerative colitis (UC). Participants were administered 200 mL of FMT from healthy donors through either colonoscopy or gastroscopy, with evaluations conducted at baseline, week 7, and six months after the intervention. Clinical improvements, including reductions in inflammatory markers, were observed, alongside significant alterations in gut microbiota composition. These findings suggest that multi-session FMT effectively reconstitutes gut microbiota and induces remission in UC patients (Mańkowska-Wierzbicka et al., 2020). Probiotic bacteria, especially engineered probiotics such as E. coli, Lactobacillus paracasei, Bifidobacterium longum, Lactococcus lactis, and Bacteroides ovatus, are effective therapeutic strategies in the treatment of IBD (Pesce et al., 2022). The administration of intracolonic synbiotic treatment,

consisting of *Bifidobacterium animalis subsp. lactis* and xyloglucan, demonstrated significant improvements in mucosal healing and alleviation of colonic symptoms among patients with severe ulcerative colitis (Bozkurt and Kara, 2020).

3.1.2 Irritable bowel syndrome

Irritable bowel syndrome (IBS) is a group of symptoms characterized by abdominal discomfort and irregular bowel movements (Saha, 2014). The pathophysiology of IBS is multifactorial (Surdea-Blaga et al., 2012), including disturbed gut motility (Shaidullov et al., 2021), visceral sensitivity (Li et al., 2020a), neural dysfunction of the gut-brain axis (Suganya and Koo, 2020), autonomous nervous system dysfunction (Mazur et al., 2012), and psychological factors (Van Tilburg et al., 2013), which are implicated in disease progression. Fecal SCFAs are biomarkers for diagnosing IBS (Farup et al., 2016) and are produced by the microbial fermentation of ingestible polysaccharides and proteins, through which commensal microbiota communicate with the host (Rasmussen et al., 1988; Natarajan and Pluznick, 2014). A study of twenty-five subjects with IBS and 25 controls aimed to diagnose IBS by measurement of fecal SCFA; the propionic acid-butyric acid ratio (mmol/l) showed the best diagnostic properties, with a sensitivity of 92% and a specificity of 72% at a cut-off value >0.015 mmol/L (Farup et al., 2016). In a recent study, 490 individuals with IBS and 122



colorectal cancer. (Created with Biorender.com).

individuals without IBS were analyzed using metagenomic sequencing. In IBS, alpha diversity was significantly lower, rich in gram-negative bacteria, including Shigella species, and contained disrupted metabolic pathways associated with short-chain fatty acid and vitamin synthesis, whereas Eubacterium rectale and Faecalibacterium prausnitzii were relatively sparse in individuals with IBS (Phan et al., 2021). A systematic review and meta-analysis aimed to evaluate the alterations in the intestinal microbiota in IBS using qPCR to quantify bacterial groups. Additionally, there was a significant decrease in Lactobacillus, Bifidobacterium and Faecalibacterium prausnitzii in IBS compared with the corresponding populations in healthy controls (Liu et al., 2017). Another study reported an increased abundance of Dorea, Ruminococcus, and Clostridium spp. and a reduction in Bacteroides, Bifidobacterium, and Faecalibacterium in patients with IBS (Rajilic-Stojanovic et al., 2011). Probiotics and prebiotics are commonly used to treat the symptoms of irritable bowel syndrome (IBS). In a clinical trial, two probiotic strains, Lactobacillus acidophilus DDS-1 and Bifidobacterium lactis UABla-12, were found to improve abdominal pain in individuals with IBS. These probiotics also help regulate bowel movements, potentially relieving diarrhea or constipation, which are common symptoms of IBS (Martoni et al., 2020). Another study showed that supplementation of Bacillus coagulans MTCC 5856, along with standard care, was safe and effective in treating diarrhea-predominant IBS in patients, suggesting it as a potential

treatment option (Majeed et al., 2016). Furthermore, several other probiotics, including *Lactobacillus bulgaricus*, *Lactobacillus paracasei*, *Lactobacillus reuteri*, *Lactobacillus plantarum*, *Pediococcus acidilactici*, *Streptococcus thermophilus*, *Bifidobacterium infantis*, *Bifidobacterium bifidum*, *Lactobacillus brevis*, *Bifidobacterium longum*, and *Saccharomyces boulardii*, have shown positive effects on IBS symptoms (Chlebicz-Wojcik and Slizewska, 2021). The prebiotic galactooligosaccharide promotes the growth of beneficial gut *Bifidobacterium* in IBS patients, leading to a reduction in symptoms such as flatulence, abdominal pain, and discomfort (Silk et al., 2009). Psyllium fiber has also been found to reduce abdominal pain and inflammation in IBS patients (Shulman et al., 2017; Garg, 2021).

3.1.3 Celiac disease

Celiac disease is a critical autoimmune pathology that affects the small intestine. Although gluten is the main trigger, the gut microbiome also has a notable effect on the disease's progression (Marasco et al., 2016) (Figure 2). The gut bacteria from the phyla Firmicutes and Actinobacteria, mostly *Lactobacillus, Streptococcus, Staphylococcus, Bifidobacterium*, and *Clostridium*, is involved in gluten metabolism (Caminero et al., 2014). A previous study showed that rod-shaped bacteria were predominantly associated with the intestinal mucosa of children with active and inactive celiac disease, compared with the bacterial flora seen in the

controls (Forsberg et al., 2004). In their culture-dependent method, Collado et al. (Collado et al., 2007) reported that the presence of *Bacteroides, Clostridium*, and *Staphylococcus* was significantly higher in fecal samples from patients with coeliac disease than in healthy subjects. The gluten-free diet is a commonly recommended treatment for patients with celiac disease. However, incorporating probiotics and prebiotics into this diet can potentially restore the gut microbiome, leading to improved gluten breakdown in the gut (Olivares et al., 2014; Drabińska et al., 2020). Consequently, this can reduce inflammation, enhance gut health, and decrease the production of cytokines and antibodies that contribute to issues in celiac disease. As a result, patients may experience fewer symptoms and an overall better quality of life (De Sousa Moraes et al., 2014).

3.1.4 Colorectal cancer

The gut microbiota significantly influences the pathogenesis of colorectal cancer (CRC) through its effects on microbial composition, metabolic activity, and interactions with the host (Figure 2). Gut microbial dysbiosis has been reported in patients with CRC, where there is a lower abundance of commensal microbiota, especially SCFA-producing bacteria, and a higher prevalence of proinflammatory pathogenic microbes (Sanchez-Alcoholado et al., 2020). Colonizing GF mice with the microbiota of patients with CRC and their healthy counterparts, CRC fecal-receiving mice develop epithelial hyperplasia and DNA methylation in the intestine (Sobhani et al., 2019). Bacteroides fragilis is involved in CRC pathogenesis with the production of Bacteroides fragilis toxins (BFT) and biofilm (Cheng et al., 2020). Research has highlighted that the bft gene, responsible for encoding BFT toxins, and biofilm formation are essential virulence determinants contributing to colorectal cancer (CRC) pathogenicity. The prevalence of BFT toxin-producing and biofilm-forming strains of Bacteroides fragilis is significant among patients with colorectal cancer (Jasemi et al., 2020). However, it has been found that cell-free supernatants from Clostridium butyricum can inhibit the growth of B. fragilis, prevent biofilm production, and potentially serve as a biotherapeutic agent against CRC (Shin et al., 2020). Studies using quantitative PCR and 16s rRNA gene sequencing methods have shown an increased presence of Fusobacterium species and a reduction in the Bacteroidetes and Firmicutes phyla in colorectal carcinoma (Kostic et al., 2012). Research indicated that the supplementation of six viable strains from Lactobacillus and Bifidobacterium strains significantly lowered levels of proinflammatory cytokines, including TNF-α, IL-6, IL-10, IL-12, IL-17A, IL-17C, and IL-22. Furthermore, this approach was beneficial in reducing the incidence of post-surgical complications in individuals with colorectal cancer (CRC) (Zaharuddin et al., 2019). Another research revealed that a prebiotic formulation containing fructooligosaccharide, xylooligosaccharide, polydextrose, and resistant dextrin had profound impacts on immune-related markers both prior to and following surgical procedures in colorectal cancer (CRC) patients and the administration of these prebiotics led to significant modifications in the populations of commensal bacteria and opportunistic pathogens within the patient cohort (Xie et al., 2019).

3.2 Metabolic disorders associated with gut microbiota

3.2.1 Obesity

Comprehending the intricate interactions between the gut microbiome and obesity is crucial for devising effective strategies to prevent and manage this escalating health concern (Figure 3). Emerging evidence suggests an intrinsic link between microbial dysbiosis and obesity (Joseph et al., 2020). Obesity and obesityassociated complications are consequences of alterations in the composition and function of the gut microbiota (Shin and Cho, 2020). Resveratrol intake enhances glucose metabolism and maintains homeostasis, indicating its potential as a therapeutic intervention for obesity. Treating obese mice with resveratrol reduces gut dysbiosis, increases the Bacteroidetes-to-Firmicutes ratio, inhibits the growth of Enterococcus faecalis, and promotes the growth of Lactobacillus and Bifidobacterium (Qiao et al., 2014). Studies with human subjects reported a significant reduction of the Firmicutes/ Bacteroidetes ratios in people with obesity compared with those in healthy control (Verdam et al., 2013; Kasai et al., 2015; Duan et al., 2021; Zhai et al., 2022a). In individuals with obesity, beneficial microbiota such as Bifidobacterium, Faecalibacterium, and Ruminococcaceae are significantly depleted, whereas Bacillus and potential opportunistic pathogens such as Fusobacterium, Escherichia, and Shigella increased (Gao et al., 2018). Mechanistically, the microbiota of individuals with obesity is rich in indigestible polysaccharide-degrading enzymes that produce increased levels of acetate and butyrate (Martinez-Cuesta et al., 2021). Evidence indicates that acetate is linked to obesity, with elevated acetate levels generated by the gut microbiota activating the parasympathetic nervous system, promoting insulin secretion, and leading to hyperphagia and obesity in rodents (Perry et al., 2016). Dietary strategies, such as the administration of probiotics, prebiotics, synbiotics, and fecal microbiota transplants, may facilitate microbial reconstitution and assist in controlling weight gain and associated health conditions. The bacterium Akkermansia muciniphila is recognized for its antiobesity effects. It is well-tolerated and safe for consumption, and has been shown to enhance insulin sensitivity, lower insulin levels, and decrease cholesterol, underscoring its potential role in weight management among individuals with overweight (Depommier et al., 2019). Lactobacillus plantarum LMT1-48, for example, can reduce body weight and abdominal fat by regulating lipogenic genes in adipose tissue and the liver (Choi et al., 2020). Oral intake of Lactobacillus fermentum strain 4B1 has been found to reduce body weight, adipose tissue weight, and adipose cell size, similar to the effects of the drug orlistat (Balolong et al., 2017). Lactobacillus mali APS1 has been shown to restore the gut microbiome, regulate metabolism and appetite, resulting in weight loss, reduced body fat, liver weight, fat accumulation in the mesenteric adipose depot, and improved hepatic steatosis compared to a diet lacking this probiotic strain (Lin et al., 2016; Chen et al., 2018). Chung et al. (Chung et al., 2016) investigated the efficacy of Lactobacillus reuteri JBD301 against obesity and found that, like orlistat, Lactobacillus JBD301 absorbs free fatty acids and excretes them in the feces, leading to significant weight loss in both



Phenylacetylglutamine. (Created with Biorender.com).

mice and humans. Lactobacillus gasseri SBT2055 has multiple benefits for the host, including reduced TAG absorption, accelerated energy expenditure, improved glucose tolerance, increased butyrate production, reduced inflammation, inhibited body weight gain, and decreased fat accumulation. These effects may be linked to the amelioration of adipose tissue inflammation and reduced expression of lipogenic genes in the liver (Miyoshi et al., 2014; Shirouchi et al., 2016). Bifidobacterium longum BORI and Lactobacillus paracasei CH88, along with fermented ginseng, have demonstrated their ability to reduce various obesity-related markers, including weight gain, lipid deposition, adipocyte size, inflammation, fasting blood glucose levels, and total cholesterol excretion in mouse models and human intervention studies (Kang et al., 2018; Schellekens et al., 2021). Arabinoxylans, a prebiotic derived from rice bran and wheat sources, have the potential to reduce obesity by regulating lipid metabolism, reducing inflammation through the manipulation of gut microbiota, and promoting the production of beneficial short-chain fatty acids. This highlights their potential as prebiotic agents for managing obesity (Neyrinck et al., 2011; Luo et al., 2022).

3.2.2 Diabetes

The gut microbiota also affects diabetes, with disruptions in glucose homeostasis leading to alterations in the host's gut microbial community, which subsequently contributes to the development of type 2 diabetes mellitus (T2D) and associated comorbidities (Sharma and Tripathi, 2019). An altered gut microbiome deteriorates intestinal barrier function and hosts metabolic and signaling pathways, which

results in the progression of T2D (Riedel et al., 2021) (Figure 3). Differences in the gut microbiome between patients with T2D and healthy controls have been reported in 16S rDNA amplicon sequencing studies, where butyrate-producing Faecalibacterium, Bifidobacterium, and Akkermansia were significantly reduced in patients with diabetes. The abundance of Dorea was significantly increased in individuals with T2D (Li et al., 2020b). A previous study demonstrated that the proportions of the phylum Firmicutes and class Clostridia were significantly reduced in patients with diabetes compared with those in the non-diabetic control group. At the same time, there was an increased presence of Bacteroidetes and Proteobacteria. In addition, the Bacteroidetes to Firmicutes ratio was significantly and positively correlated with reduced glucose tolerance (Larsen et al., 2010). Studies on mice have shown that supplementation with Bifidobacterium genus supplements can improve glucose tolerance, insulin secretion, and reduce inflammation (Cani et al., 2007). Metformin and berberine have also been found to be effective in reducing diabetic complications in mice by increasing the presence of SCFA-producing bacteria (such as Butyricimonas, Coprococcus, and Ruminococcus), while reducing opportunistic pathogens (such as Prevotella and Proteus), body weight, blood glucose levels, and intestinal inflammation (Zhang et al., 2019). Lactobacillus strains, such as Lactobacillus reuteri 263 and a synbiotic combination of mangiferin and Lactobacillus reuteri 1-12, have shown promise in improving insulin resistance, hepatic steatosis, and blood glucose levels in rats fed a high fructose diet (Hsieh et al., 2013; Meng et al., 2023). Other lactobacillus strains, including Lactobacillus

rhamnosus BSL, Lactobacillus rhamnosus R23, Lactobacillus plantarum HAC01, Lactobacillus fermentum TKSN041, Lactobacillus gasseri SBT2055, Lactobacillus sakei Probio65, Lactobacillus plantarum Probio-093, Lactobacillus plantarum ZJUFB2, Lactobacillus fermentum MF423, Lactobacillus salivarius AP-32, and Lactobacillus reuteri GL-104, have also shown potential as therapeutic agents for treating type 2 diabetes (Niibo et al., 2019; Farida et al., 2020; Hsieh et al., 2020; Lee et al., 2021; Zhong et al., 2021a; Zhou et al., 2021). In animal models, the probiotic strain Bifidobacterium animalis 01 has been found to improve glucose metabolism and exhibit hepatoprotective effects, while Bifidobacterium animalis ssp. lactis GCL2505 (BlaG) reduces symptoms of metabolic syndrome, including visceral fat reduction and improved glucose tolerance, through modulation of the gut microbiota and increased acetate levels (Aoki et al., 2017; Zhang et al., 2020). Akkermansia muciniphila has been shown to improve glucose tolerance and insulin sensitivity (Zhang et al., 2021). Clostridium butyricum has beneficial effects in reducing high blood sugar levels (hyperglycemia) and promoting the secretion of glucagon-like peptide-1 and insulin (Jia et al., 2017). Anaerobutyricum soehngenii has been found to improve insulin sensitivity by stimulating intestinal GLP-1 production (Koopen et al., 2022). Additionally, supplementation with oligofructose-enriched inulin, a prebiotic substance, has been shown to significantly improve glycemic control and reduce inflammatory markers in women with T2D (Dehghan et al., 2014). Emerging therapeutic strategies, including probiotic supplementation, prebiotic intake, and microbial modulation, demonstrate potential in improving glucose metabolism, insulin sensitivity, and reducing diabetic complications. These findings underscore the gut microbiota as a promising target for novel therapeutic approaches in managing and preventing T2D.

3.3 The impact of gut bacteria on cardiovascular disorders

Comprehending the interplay between gut dysbiosis and cardiovascular disease offers significant insights into innovative therapeutic and preventive strategies that target cardiovascular risk mitigation through microbiota modulation. Homeostatic processes mediated by host-microbial interactions govern physiological balance and may activate various pathways, thereby contributing to the progression of cardiovascular risk factors (Troseid et al., 2020). Trimethylamine N-oxide (TMAO) and phenylacetylglutamine (PAGln) are metabolites derived from gut microbiota that are linked to cardiovascular disease. Elevated concentrations of TMAO, which is produced by gut bacteria from dietary substrates such as choline, phosphatidylcholine, and L-carnitine, are associated with an increased risk of cardiovascular diseases, including congenital heart disease and atherosclerosis (Chen et al., 2020; Zhen et al., 2023) (Figure 3). TMAO induces the production of reactive oxygen species, which induces inflammatory reactions and inhibits the reverse cholesterol transport pathway, resulting in atherosclerosis (Zhu et al., 2020). Phenylacetylglutamine (PAGln) promotes platelet activation and enhances the likelihood of thrombosis by engaging G protein-coupled adrenergic receptors on the platelet surface. This interaction results in receptor activation by PAGIn, which causes excessive platelet stimulation, rendering them hyperreactive and contributing to accelerated platelet aggregation and an increased risk of thrombosis (Nemet et al., 2020; Yu et al., 2021). A study suggests the effects of several strains of Lactobacillus on atherosclerosis in ApoE-/- mice. They found that this probiotic strain significantly reduced atherosclerotic lesion area, lowered serum lipid levels, and decreased inflammatory markers. The researchers attributed these effects to the probiotic's ability to modulate gut microbiota composition and enhance intestinal barrier function (Zhai et al., 2022b). A study by Li et al., (Li et al., 2016b) explored the effects of Akkermansia muciniphila on atherosclerosis in ApoE-/- mice. They found that A. muciniphila supplementation reduced atherosclerotic plaque formation, decreased inflammatory markers, and improved gut barrier function. This study highlights the potential of specific probiotic strains in targeting atherosclerosis. Short-chain fatty acids (SCFAs) play a crucial role in regulating anti-inflammatory responses, lipid metabolism, and gluconeogenesis. Bacteria that generate butyrate are instrumental in impeding the advancement of atherosclerosis. Moreover, butyrate demonstrates diverse pharmacological effects, including the promotion of microbial homeostasis, reinforcement of intestinal barrier function, and exertion of anti-inflammatory activities (Amiri et al., 2021). The reduced amount of beneficial or commensal bacteria, including Faecalibacterium prausnitzii and Bacteroides fragilis, are observed in patients with coronary artery disease (CAD) and type 2 diabetes and increased number of opportunistic pathogens, such as Enterobacteriaceae, Streptococcus, and Desulfovibrio, have observed in CAD patients without type 2 diabetes. Moreover, patients with CAD-DM2 possessed significantly elevated levels of zonullin and TMAO, the pro-inflammatory cytokine IL-1B, and lower levels of IL-10 (Sanchez-Alcoholado et al., 2017). Gozd-Barszczewska et al. (Gozd-Barszczewska et al., 2017) reported the dominance of Firmicutes and Bacteroidetes in middle-aged men in eastern Poland, with improper levels of total cholesterol and LDL-C rich in Prevotella and low in Clostridium and Faecalibacterium. A Chinese report on patients with atherosclerosis showed a lower abundance of Bacteroides and Prevotella, which are rich in Streptococcus and Escherichia (Jie et al., 2017). The opportunistic pathogen Collinsella is more prevalent in patients with symptomatic atherosclerosis. In contrast, Roseburia and Eubacterium are enriched in healthy controls (Karlsson et al., 2012).

Fecal microbiota transplantation (FMT) has demonstrated efficacy in reestablishing microbial equilibrium within the gut and alleviating myocarditis, thereby presenting a potential innovative therapeutic modality for its management (Hu et al., 2019). Modulating the gut microbiota via fecal microbiota transplantation (FMT) has shown a pronounced effect on atherosclerosis in murine models. Transplantation of control microbiota into atherosclerosisprone mice resulted in a reduction in the advancement of atherosclerotic lesions. In contrast, the transfer of atherosclerosisprone microbiota into control mice exacerbated lesion progression. These findings indicate that restoring microbial homeostasis in the gut may serve as a viable therapeutic strategy for atherosclerosis (Kim et al., 2022). A study found that clearing gut microbiota with antibiotics and transplanting healthy fecal microbiota could



These disturbances adversely affect brain health, contributing to cognitive disorders, mood disorders, and inflammation affecting the central nervous system. Specifically, in Alzheimer's disease, the production of amyloid plaques and neurofibrillary tangles is exacerbated by inflammatory processes and the disruption of gut-brain signaling. In Parkinson's disease, altered gut microbiota can increase alpha-synuclein aggregation and inflammation, contributing to the degeneration of dopaminergic neurons. GABA, Gamma Amino Butyric Acid; SCFA, Short Chain Fatty acids; ENS, Enteric Nervous System; BA, Bile Acids; LPS, Lipopolysaccharides; TMAO, Trimethylamine N-oxide. (Created with Biorender.com).

alleviate cardiac fibrosis. This indicates that probiotics, specifically Clostridium butyricum and Bifidobacterium pseudolongum, as well as metabolite interventions, could offer new strategies for treating cardiovascular disease (Wang et al., 2022). In obese mice with obstructive sleep apnea (OSA) induced by a high-fat high-fructose diet and intermittent hypoxia (IH), both Lactobacillus rhamnosus GG (LGG) and LGG cell-free supernatant (LGGs) effectively protected against heart dysfunction, cardiac remodeling, and inflammation. This protection is potentially achieved through the up-regulation of antioxidant pathways mediated by nuclear factor erythroid 2-related factor 2 (Nrf2) (Xu et al., 2019). A six-week regimen of probiotic supplementation in individuals with type 2 diabetes mellitus (T2DM) resulted in significant improvements in cardiovascular disease-related parameters, such as blood pressure and atherogenic indices. Additionally, the Framingham risk score suggested potential benefits in mitigating the risk of future cardiovascular events within this cohort. Nevertheless, further empirical research is necessary to validate these findings (Ahmadian et al., 2022).

3.4 Gut bacteria influence on central nervous system pathologies

3.4.1 Anxiety and depression

The bidirectional relationship between the gastrointestinal (GI) and central nervous systems underscores their close connection. The investigation into the gut microbiota's role in the gut-brain axis has sparked significant scientific interest, with research focusing on how this axis affects neurodegenerative disorders, including anxiety and depression. Recent studies have explored the potential links between the gut microbiota and these mental health conditions (Figure 4). Studies utilizing animal models have indicated that alterations in gut microbiota composition, induced by stress, can influence host behavior and interfere with normal behavioral patterns (Geng et al., 2019; Xu et al., 2020). Compared to conventional specific pathogen-free (SPF) mice, adult germ-free (GF) mice demonstrate reduced anxiety-like behavior in the elevated plus maze, exhibit elevated expression of brainderived neurotrophic factor (BDNF), and show decreased expression of

10.3389/fcimb.2024.1291148

serotonin (5-hydroxytryptamine) receptors in the brain (Neufeld et al., 2011b; Chen et al., 2013). A subsequent study reported that the offspring of germ-free (GF) mice, which exhibited reduced stress, were colonized with specific pathogen-free (SPF) feces to introduce a normal gut microbiota, followed by a reassessment of anxiety-like behavior. The reconstitution of the gut microbiota did not normalize the behavioral phenotype; anxiolytic behavior persisted in GF mice colonized with SPF microbiota. This indicates that interactions between the gut and the brain are crucial for the development of stress-related systems in the central nervous system (Neufeld et al., 2011a). Increasing evidence highlights the potential of gut bacteria-focused therapies, including fecal microbiota transplantation (FMT), probiotics, prebiotics, and synbiotics, in mitigating anxiety and depression. Studies involving rats have shown that FMT from healthy donors can improve depressive-like symptoms by restoring the gut microbiota, decreasing intestinal inflammation, and reinforcing gut barrier integrity (Rao et al., 2021; Hu et al., 2022). It also progressively alleviated alcohol-induced anxiety and depression in mice (Xu et al., 2018). Fecal microbiome used as adjunctive therapy significantly improved depressive symptoms in patients with depression disorder 4 weeks after transplantation (Doll et al., 2022). In patients with irritable bowel syndrome with diarrhea, fecal microbiota transplantation has been shown to substantially alleviate symptoms of anxiety and depression (Lin et al., 2021a).

Mice fed the probiotic *Lactobacillus rhamnosus* showed decreased depression and anxiety-like behavioral traits (Bravo et al., 2011). *Lactobacillus reuteri NK33* and *Bifidobacterium adolescentis* NK98 alleviate stress, depression, anxiety, and sleep disturbances through the suppression of inflammation, improvement of the gut ecosystem and increased brain-derived neurotrophic factor expression (Jang et al., 2019). Daily supplementation with *Lactobacillus plantarum P8* for 12 weeks notably reduces depression, anxiety, and pro-inflammatory cytokines, while enhancing memory and cognitive traits, particularly in women. This suggests that *Lactobacillus plantarum P8* may be an effective intervention for diminishing these symptoms in stressed adults (Lew et al., 2019).

Akkermansia muciniphila and Clostridium butyricum were shown to have antidepressant effects in mice with chronic stress by modifying gut microbiota (Ding et al., 2021). They also increased serotonin, brainderived neurotrophic factor, and glucagon-like peptide-1 (Sun et al., 2018). *Bifidobacterium breve* CCFM1025 and *Bifidobacterium breve* A-1 are two promising psychobiotics. The former reduces depression and related GI disorders by modulating the gut microbiota and tryptophan metabolism, while the latter improves anxiety and depressive symptoms in patients with schizophrenia through altered gut microbiome (Okubo et al., 2019; Tian et al., 2022).

In a randomized controlled trial involving 80 students facing examination stress, a multi-strain probiotic capsule intake with glutamine (*Bacillus coagulans* Unique IS2, *Lactobacillus rhamnosus UBLR58*, *Bifidobacterium lactis UBBLa70*, *Lactobacillus plantarum UBLP40* (each of 2x109 CFU); *Bifidobacterium breve UBBr01*, *Bifidobacterium infantis* UBBI01 (each of 1x10⁹ CFU) was found to substantially reduce experienced stress, depression, anxiety, and cortisol levels compared to the placebo group, with no adverse effects (Venkataraman et al., 2021). Another triple-blind, placebo-controlled study investigated the impacts of a multispecies probiotic strain containing Bifidobacterium bifidum W23, Bifidobacterium lactis W52, Lactobacillus acidophilus W37, Lactobacillus brevis W63, Lactobacillus casei W56, Lactobacillus salivarius W24, and Lactococcus lactis W19 and Lactococcus lactis W58 supplement on cognitive reactivity to sad moods in non-depressed individuals. The study found that the probiotic group experienced a significant reduction in cognitive reactivity to sad mood, particularly in rumination and aggressive thoughts (Steenbergen et al., 2015). Apart from probiotics, prebiotics such as fructo-oligosaccharides and galacto-oligosaccharides have been shown to exert antidepressant effects in mice by altering gut microbiota and enhancing short-chain fatty acid (SCFA) production (Burokas et al., 2017).

3.4.2 Autism spectrum disorder, Parkinson's disease, schizophrenia, Alzheimer's disease, and prion disease

Gastrointestinal dysfunction is commonly reported in Parkinson's disease (PD), schizophrenia [SCZ], autism spectrum disorder (ASD) Alzheimer's Disease (AD), prion disease, and promotes the onset of these diseases (Pfeiffer, 1998; Davies et al., 2006; Kang et al., 2014; Patrono et al., 2021; Sohrabi et al., 2022; Taniya et al., 2022) (Figure 4). Several studies have reported alterations in the gut microbial community in patients with PD/ SCZ/ASD compared with the gut microbiota in healthy controls. However, there is little harmony in the results on which groups of microbiomes reduce or increase, and there are even conflicting results (Strati et al., 2017; Zheng et al., 2019; Gorecki et al., 2019; Afroz et al., 2021; Ha et al., 2021; Li et al., 2021b).

In PD Clostridium cluster IV, Akkermansia, Bifidobacterium, and Lactobacillus are more abundant, and Faecalibacterium spp., Coprococcus spp., Blautia spp., Prevotella spp., and Prevotellaceae are significantly reduced in the PD group compared with the corresponding populations in the control group (Gerhardt and Mohajeri, 2018; Bullich et al., 2019). In an alpha-synucleinoverexpressing mouse model, the gut microbiota was required for motor deficits, microglia activation, and alpha-synuclein pathology. Specific microbial metabolites administered orally to GF mice promote neuroinflammation and motor symptoms. Physical impairments were observed in alpha-synuclein-overexpressing mice colonized with microbiota from patients with PD compared with microbiota transplants from healthy human donors (Sampson et al., 2016). Fecal microbiota transplantation (FMT) may exert a positive impact on both motor and non-motor symptoms in individuals with Parkinson's disease. Over a 12-week treatment period, FMT was associated with increased gut microbiome diversity, reduced constipation, and reported subjective improvements in both motor and non-motor symptoms (Xue et al., 2020; Kuai et al., 2021; Segal et al., 2021; Dupont et al., 2023). In addition, a meta-analysis of nine randomized controlled trials with 663 subjects found that oral probiotic intake substantially improved motor symptoms, gastrointestinal symptoms, anxiety, depression, and reduced laxative use and increased glutathione levels in Parkinson's disease patients (Chu et al., 2023). Probiotic Bifidobacterium animalis subsp. lactis Probio-M8 with conventional drug enhanced sleep patterns, reduced anxiety and GI disorders, and positively changes intestinal microbes and metabolic pathways

(Sun et al., 2022a). A mixture of multispecies probiotics containing *Lactobacillus acidophilus, Bifidobacterium bifidum, Lactobacillus reuteri*, and *Lactobacillus fermentum* improved motor behavior, cognitive function, reduced oxidative stress, and neuronal damage in rat model of Parkinson's disease (Alipour Nosrani et al., 2021).

Aberrant metabolite production by the gut microbiota can modulate immune responses and alter the gut microbiome profile in individuals with autism spectrum disorder (ASD) (Oh and Cheon, 2020). One study reported lower levels of acetic acid and butyrate and an elevated level of valeric acid in patients with ASD. In addition, the butyrate-producing taxa Ruminococcaceae, Eubacterium, Lachnospiraceae, and Erysipelotrichaceae decreased, and the abundance of valeric acid-associated bacteria (Acidobacteria) increased among subjects with autism (Liu et al., 2019). Another study reported that mothers on a long-term high fat diet produced a threat of ASD-like behavior in babies compared with chow-fed mothers, and the gut microbiota differed, with a significant decrease in Lactobacillus reuteri observed (Buffington et al., 2016). Fecal microbiota transplantation (FMT) and probiotic supplementation have demonstrated potential in mitigating the disorders associated with autism spectrum disorder (ASD). In a clinical trial, FMT treatment was observed to enhance gastrointestinal (GI) symptoms and behavioral symptoms in children with ASD by modulating gut microbiota composition and altering serum neurotransmitter levels. This treatment reportedly reduced the abundance of Eubacterium coprostanoligenes (Li et al., 2021a). Additionally, the probiotic bacterium Limosilactobacillus reuteri was found to improve social behavior in a mouse model of neurodegenerative disorders. This improvement was achieved through the vagus nerve, oxytocin, and biopterin pathways which restored synaptic plasticity in the ventral tegmental area (Sgritta et al., 2019; Dooling et al., 2022). A three-month administration of a multispecies probiotic, which includes Lactobacillus acidophilus, Lactobacillus rhamnosus, and Bifidobacteria longum, has demonstrated the ability to enhance beneficial gut bacteria, decrease body weight, and significantly improve symptoms associated with autism and gastrointestinal disorders (Shaaban et al., 2018). In a case study, a 12-year-old boy with autism spectrum disorder (ASD) underwent treatment using a multi strain probiotic mixture consisting of 10 probiotics, including Bifidobacterium, Lactobacillus, and Streptococcus genera. The results were astonishing, as the boy experienced significant improvements in his autistic core symptoms after just four months of probiotic treatment (Grossi et al., 2016).

The gut microbiome of individuals with schizophrenia (SCZ) exhibits significant divergence from that of healthy controls or individuals with metabolic syndrome. The SCZ gut is enriched with *Flavonifractor plautii*, *Collinsella aerofaciens, Bilophila wadsworthia*, and *Sellimonas intestinalis*. At the same time, there is a paucity of *Faecalibacterium prausnitzii*, *Ruminococcus lactaris*, *Ruminococcus bicirculans*, and *Veillonella rogosae*. Thirion et al. (Thirion et al., 2023) conducted a study to examine the gut microbiome in individuals with schizophrenia (SCZ). The researchers discovered notable differences in the gut bacteria of SCZ patients compared to both the healthy control group and the metabolic syndrome group. Notably, they identified a connection between the biosynthesis of tyrosine by gut bacteria and cognitive function in individuals with SCZ. Antipsychotic agents, including Haloperidol, fluphenazine,

chlorpromazine, quetiapine, risperidone, and aripiprazole, are commonly prescribed for the treatment of schizophrenia. However, it is important to note that these medications can have certain side effects related to movement, such as tremors and dystonia. Prolonged administration of these pharmacological agents may result in the onset of uncontrollable orofacial muscle movements (Kang and Kim, 2011; Stroup and Gray, 2018). Conversely, a 12-week intervention involving 60 patients with chronic schizophrenia utilized a probiotic formulation consisting of Lactobacillus acidophilus, Bifidobacterium bifidum, Lactobacillus reuteri, and Lactobacillus fermentum, along with vitamin D. The findings revealed substantial improvements in both overall and composite Positive and Negative Syndrome Scale (PANSS) scores, signifying a reduction in clinical symptoms (Ghaderi et al., 2019). A randomized, double-blind, placebo-controlled trial was conducted to examine the effects of probiotic supplementation on individuals diagnosed with schizophrenia. The trial utilized a combination of Lactobacillus rhamnosus strain GG and Bifidobacterium animalis subsp. lactis strain Bb12. The findings indicated that the supplementation did not yield a significant impact on the overall severity of symptoms experienced by these patients. However, it was observed that the supplementation was linked to a decreased likelihood of developing severe bowel difficulties, a prevalent somatic symptom within this population (Dickerson et al., 2014). In addition, complex polyphenols are broken down by the gut microbiota into smaller molecules, which has several health effects including changing the composition of the gut flora. Additionally, polyphenols have important roles in memory, learning, and cognitive processes by shielding neurons from damage and inflammation (Filosa et al., 2018). There is no concrete proof linking the pathophysiology and patho-biochemistry of schizophrenia in humans to a disruption in the intake of polyphenols from food. However, some polyphenols, such as quercetin, have been effectively added to clozapine, an antipsychotic drug used to treat schizophrenia (Schwartz, 2016). Alzheimer's disease is a progressive neurodegenerative disorder characterized by cognitive decline, memory loss, and behavioral changes. It is marked by the accumulation of amyloid- β plaques and neurofibrillary tangles in the brain (Peddinti et al., 2024). The gut-brain axis involves various pathways, including neural, endocrine, and immune mechanisms, through which the gut microbiota can influence brain function and potentially contribute to the development of AD (Ullah et al., 2023). Gut dysbiosis, an imbalance in the gut microbiota composition, can lead to increased intestinal permeability and the release of pro-inflammatory molecules (Di Vincenzo et al., 2024). These molecules can cross the blood-brain barrier, triggering neuroinflammation and potentially contributing to AD pathogenesis (Peddinti et al., 2024). Some studies have suggested that certain gut bacteria can produce amyloid proteins, which may contribute to the accumulation of amyloid- β in the brain, a hallmark of AD (Chen et al., 2022). Dysbiosis in the gut microbiota can lead to increased oxidative stress, which has been implicated in the pathogenesis of AD (Chen et al., 2022). A study by Li et al. (Li et al., 2019a) found that individuals with mild cognitive impairment and AD had similar alterations in their gut microbiota composition compared to healthy controls. Vogt et al., 2017) observed alterations in the gut microbiome of AD patients,



suggesting a potential role of gut microbiota in AD pathogenesis. Nagpal et al. (Nagpal et al., 2019) found that a modified Mediterranean ketogenic diet modulated the gut microbiome and short-chain fatty acids in association with Alzheimer's disease markers in subjects with mild cognitive impairment. This suggests that dietary interventions targeting the gut microbiota may have potential in managing AD. A study by Saji et al. (Saji et al., 2019) revealed a relationship between gut microbiome composition and mild cognitive impairment in patients without dementia, indicating that gut microbiota alterations may precede the development of AD. Saiyasit et al. (Saiyasit et al., 2020). demonstrated that gut dysbiosis develops before metabolic disturbance and cognitive decline in highfat diet-induced obsec conditions. This suggests that gut microbiota alterations may be an early event in the pathogenesis of AD.

Prion diseases, on the other hand, are rare but fatal neurodegenerative disorders caused by the misfolding of prion proteins, leading to brain damage and various neurological symptoms (Soto and Satani, 2011). Some studies have proposed that gut microbiota may influence the misfolding of prion proteins, a key process in the development of prion diseases (Mahbub et al., 2024). Gut microbiota plays a crucial role in maintaining intestinal barrier integrity. Disruption of this barrier could potentially facilitate the entry of prions into the body (Mahbub et al., 2024).

The gut microbiota's influence on the immune system may affect the body's response to prion proteins and their accumulation (Mahbub et al., 2024). Quigley et al. (Quigley, 2017) reviewed the role of the microbiota-brain-gut axis in neurodegenerative diseases, including prion diseases. The review highlighted the potential influence of gut microbiota on protein misfolding and neuroinflammation, which are key processes in prion diseases. Although not specifically focused on prion diseases, research by Cox and Weiner (2018) on microbiota signaling pathways in neurologic diseases provides a framework for understanding how gut microbiota might influence prion disease progression (Cox and Weiner, 2018). The relationship between gut microbiota and neurodegenerative disorders, particularly Alzheimer's disease and prion diseases, represents a promising area of research with potential implications for disease prevention and treatment. The gut-brain axis emerges as a critical pathway through which gut microbiota may influence the development and progression of these disorders. This growing body of evidence linking gut microbiota to neurodegenerative disorders opens new avenues for understanding and potentially treating these complex diseases. As our knowledge in this field expands, it may lead to novel therapeutic approaches and preventive strategies, offering hope for those affected by Alzheimer's disease, prion diseases, and other neurodegenerative disorders.

3.5 Mechanistic insight of gut bacteria on renal pathology

Renal disorders, encompassing a range of pathological conditions affecting kidney function, are intricately linked with alterations in the gut microbiota. Recent studies have elucidated several mechanisms by which gut microbiota dysbiosis influences renal health. The microbial fermentation of proteins and amino acids in the gut leads to the generation of excessive toxic metabolites, such as ammonia, amines, thiols, phenols, and indoles, while concurrently diminishing the production of shortchain fatty acids (SCFAs). In the context of chronic kidney disease (CKD), the heightened levels of uremic toxins promote intestinal dysbiosis. This dysbiotic condition undermines the integrity of the intestinal epithelial barrier, thereby aggravating renal damage (Ramezani et al., 2016). A study on patients with CKD showed an increased abundance of Lactobacillus, Clostridium IV, Paraprevotella, Clostridium sensu stricto, Desulfovibrio, and Alloprevotella in the fecal samples of patients with CKD, with a decreased presence of Akkermansia and Parasutterella compared with the corresponding populations in healthy control subjects (Li et al., 2019b). Low microbial diversity and higher levels of proinflammatory cytokines were observed in deceased patients with CKD compared with those in survivors. SCFA-producing bacteria Succinivibrio and Anaerostipes were considerably lower in nonsurvivors (Lin et al., 2021b) (Figure 5).

With the deepening insight into the microbiome, contemporary research endeavors aim to exploit its potential for enhancing health outcomes in patients with chronic kidney disease (CKD). The most frequently utilized probiotics are *Bifidobacterium* and *Lactobacillus* species. *Bifidobacteria* are recognized for their capacity to ameliorate

epithelial damage, restore epithelial functionality, and synthesize short-chain fatty acids. Additionally, they play a pivotal role in the biosynthesis of vitamins and facilitate the proliferation of Lactobacillus. Both Lactobacillus and Bifidobacterium have immune-enhancing and gut mucosal barrier-stabilizing effects (Sanekommu et al., 2023). Bifobacterium bifidum A218, Bifidobacterium catenulatum A302, Bifidobacterium longum A101, and Lactobacillus plantarum combined probiotic administration in peritoneal dialysis patients; serum TNF-α, IL-5, IL-6, and endotoxins significantly decreased, while levels of serum IL-10 significantly increased compared with the corresponding parameters in the placebo group (Shemin et al., 2001). A probiotic mixture consisting of L. acidophilus KB27, B. longum KB31, and S. thermophilus KB19, when administered orally, has been found to effectively reduce blood urea nitrogen (BUN) levels. This probiotic formulation not only improves the quality of life but is also well-tolerated. These findings support the potential of probiotics in extracting uremic toxins from the intestines (Ranganathan et al., 2010; Vitetta et al., 2019). Elevated levels of Indoxyl sulfate, a uremic toxin, are often associated with kidney dysfunction. Furthermore, a study has shown that oral administration of Bifidobacterium longum in a gastro-resistant seamless capsule can effectively reduce serum levels of indoxyl sulfate in hemodialysis patients by rectifying intestinal microflora (Takayama et al., 2003; Taki et al., 2005). Sporosarcina pasteurii has emerged as a promising candidate for an "enteric dialysis" application. It has exhibited the capacity to catabolize uremic solutes within the gastrointestinal tract, withstand gastric conditions, and efficiently remove urea in vitro. Furthermore, it has been shown to positively influence fermentation processes within the intestinal microbiota. Preliminary findings in nephrectomized rodent models suggest its potential efficacy in alleviating uremic toxicity associated with chronic kidney disease (Ranganathan et al., 2006). Plasma p-cresol, a compound detectable in the bloodstream, is implicated in renal dysfunction and classified as a uremic toxin when present at elevated concentrations. This association poses significant health risks for individuals with chronic kidney disease. A potential therapeutic approach involves synbiotic treatment, which combines probiotics such as Lactobacillus casei strain Shirota and Bifidobacterium breve strain Yakult with prebiotics like galactooligosaccharides. This intervention has demonstrated efficacy in reducing serum p-cresol levels and may mitigate the adverse effects of p-cresol, including its role as a protein-bound uremic toxin contributing to constipation (Nakabayashi et al., 2011). Furthermore, the use of prebiotic gum acacia has shown promising results in altering microbial composition, replenishing depleted butyrate levels, and displaying anti-inflammatory and antioxidant properties. These findings suggest that it could play a beneficial role in the treatment of CKD (Lakshmanan et al., 2021). Additionally, prebiotic D-serine has shown potential as both a therapeutic target and a biomarker for acute kidney injury (AKI), highlighting the complex relationship between gut microbiota and kidney health (Nakade et al., 2018). In essence, probiotics, prebiotics, and synbiotics offer substantial therapeutic benefits for kidney diseases by ameliorating disruptions in gut microbiota and rectifying associated metabolic disorders.

3.6 Influence of COVID-19 on gut microbiota composition

Dysbiosis of the gut microbiome can precipitate alterations in immune responses and elevated concentrations of proinflammatory cytokines, potentially culminating in a cytokine storm during severe acute respiratory viral infections (Figure 6). Emerging studies have revealed disruptions in the gut microbiome among COVID-19 patients, both during and after the disease course. Notable findings include diminished bacterial diversity in older individuals, a decrease in beneficial microbes, an increase in facultative anaerobic bacteria, and a decrease in the production of key metabolites (Bosco and Noti, 2021). This phenomenon may be associated with the higher mortality rate observed in older individuals compared to younger populations in the context of COVID-19 (Casas-Deza et al., 2021). Immunomodulatory bacteria, such as Faecalibacterium prausnitzii, Eubacterium rectale, and bifidobacterial species, are reduced in patients with COVID-19 (Yeoh et al., 2021). Elevated populations of Burkholderia contaminans, Bacteroides nordii, Bifidobacterium longum, and Blautia spp. were observed; CAG 257 causes severity in patients with COVID-19 (Sun et al., 2022b). Another study revealed that the Burkholderia cepacia complex, Staphylococcus epidermidis, or Mycoplasma sp. were the most abundant in severely ill patients (Zhong et al., 2021b). Zuo et al. (Zuo et al., 2020) reported that the abundance of Coprobacillus, Clostridium ramosum, and Clostridium hathewayi was positively correlated with COVID-19 severity. In contrast, there was an inverse correlation between Faecalibacterium prausnitzii and disease severity in 15 hospitalized patients.

International efforts have implemented comprehensive COVID-19 vaccination campaigns. However, the emergence of variant strains poses a potential risk to the effectiveness of these vaccines. Additionally, emerging evidence suggests that the gut microbiome may play a critical role in naturally enhancing vaccine responses and modulating immune functions within the intestinal environment (Zimmermann, 2023). A balanced gut microbiota is crucial for combating COVID-19, as it contributes to maintaining a stable immune system that can effectively counteract viral infection. An optimal gut microbiota facilitates the regulation of pro-inflammatory and anti-inflammatory metabolites, thereby preventing excessive inflammation and attenuating the severity of COVID-19 (Chen et al., 2021).

4 Concluding observations

The involvement of gut bacteria as significant etiological agents in several human pathological conditions is paramount. As research continues to uncover the intricate relationships between gut microbiota and human health, it becomes increasingly evident that addressing microbial imbalances could offer novel avenues for disease prevention and treatment. Future research endeavors should concentrate on overcoming prevailing methodological constraints and expanding our comprehension of gut microbiome dynamics to optimally utilize its potential for enhancing human



health. Despite its importance, progress in elucidating the microbiome's etiological mechanisms and in creating therapeutic interventions derived from it has been impeded by the limitations inherent in current analytical approaches. Contemporary research on the gut microbiome predominantly employs cultureindependent metagenomic next-generation sequencing (mNGS) techniques (Chung et al., 2018). Although mNGS offers valuable insights into the etiological role of the gut microbiome in disease, it is not without constraints (Liu et al., 2022b). This necessitated supplementary culture-dependent experiments (Malla et al., 2018). The essence of such culture-based studies traces back to Koch's postulates, which necessitate culturing potential etiological agents to establish disease causality (Walker et al., 2006). To fully comprehend the etiological significance of the gut microbiome, pure cultures of the implicated species are indispensable. Such cultures facilitate whole-genome sequencing and enrich metagenome sequencing data resolution (Liu et al., 2022a).

Nevertheless, many gut microbes classified as "unculturable" remain inaccessible to current cultivation techniques, primarily due to the lack of appropriate culture media. Concurrently, the scientific community faces a major challenge in creating a comprehensive gut microbe repository, which is intrinsically linked to the unavailability of a universally applicable culture medium suitable for the diverse microbial entities within the human gut microbiome. Advancing our understanding necessitates prioritizing the development of a complete gut microbe library and the formulation of universal culture media. These limitations are the principal barriers to fully comprehending the etiological roles of the gut microbiome in human diseases. Addressing these challenges is essential for unlocking the full potential of microbiome research and developing effective therapeutic interventions.

Author contributions

MI: Resources, Writing – original draft, Methodology. NM: Resources, Writing – original draft, Methodology. SH: Conceptualization, Supervision, Writing – review & editing. HC: Resources, Writing – review & editing, Project administration, Funding acquisition.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This research was funded by the Korea Basic Science Institute (KBSI) grants A439111, C442400 and C430000. This research was also supported by the Bio & Medical Technology Development Program of the National Research Foundation (NRF) funded by the Korean government (MSIT) (No. RS-2023-00224099).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated

References

Abt, M. C., and Pamer, E. G. (2014). Commensal bacteria mediated defenses against pathogens. Curr. Opin. Immunol. 29, 16–22. doi: 10.1016/j.coi.2014.03.003

Afroz, K. F., Reyes, N., Young, K., Parikh, K., Misra, V., and Alvina, K. (2021). Altered gut microbiome and autism like behavior are associated with parental high salt diet in male mice. *Sci. Rep.* 11, 8364. doi: 10.1038/s41598-021-87678-x

Afzaal, M., Saeed, F., Shah, Y. A., Hussain, M., Rabail, R., Socol, C. T., et al. (2022). Human gut microbiota in health and disease: Unveiling the relationship. *Front. Microbiol.* 13, 999001. doi: 10.3389/fmicb.2022.999001

Ahmadian, F., Razmpoosh, E., Ejtahed, H. S., Javadi, M., Mirmiran, P., and Azizi, F. (2022). Effects of probiotic supplementation on major cardiovascular-related parameters in patients with type-2 diabetes mellitus: a secondary-data analysis of a randomized double-blind controlled trial. *Diabetol. Metab. Syndr.* 14, 52. doi: 10.1186/ s13098-022-00822-z

Alipour Nosrani, E., Tamtaji, O. R., Alibolandi, Z., Sarkar, P., Ghazanfari, M., Azami Tameh, A., et al. (2021). Neuroprotective effects of probiotics bacteria on animal model of Parkinson's disease induced by 6-hydroxydopamine: A behavioral, biochemical, and histological study. *J. Immunoassay Immunochem* 42, 106–120. doi: 10.1080/ 15321819.2020.1833917

Amiri, P., Hosseini, S. A., Ghaffari, S., Tutunchi, H., Ghaffari, S., Mosharkesh, E., et al. (2021). Role of Butyrate, a Gut Microbiota Derived Metabolite, in Cardiovascular Diseases: A comprehensive narrative review. *Front. Pharmacol.* 12, 837509. doi: 10.3389/fphar.2021.837509

Aniwan, S., Tremaine, W. J., Raffals, L. E., Kane, S. V., and Loftus, E. V. Jr. (2018). Antibiotic use and new-onset inflammatory bowel disease in olmsted county, minnesota: A population-based case-control study. *J. Crohns Colitis* 12, 137-144. doi: 10.1093/ecco-jcc/jjx135

Aoki, R., Kamikado, K., Suda, W., Takii, H., Mikami, Y., Suganuma, N., et al. (2017). A proliferative probiotic Bifidobacterium strain in the gut ameliorates progression of metabolic disorders via microbiota modulation and acetate elevation. *Sci. Rep.* 7, 43522. doi: 10.1038/srep43522

Balolong, M. P., Bautista, R. L. S., Ecarma, N. C. A., Balolong, E. C., Hallare, A. V., and Elegado, F. B. (2017). Evaluating the anti-obesity potential of 4B1, a probiotic strain isolated from balao-balao, a traditional Philippine fermented food. *Int. Food Res. J.* 24, 819–824.

Bezirtzoglou, E., Tsiotsias, A., and Welling, G. W. (2011). Microbiota profile in feces of breast- and formula-fed newborns by using fluorescence in *situ* hybridization (FISH). *Anaerobe* 17, 478–482. doi: 10.1016/j.anaerobe.2011.03.009

Blaustein, R. A., Seed, P. C., and Hartmann, E. M. (2021). Biotransformation of Doxorubicin Promotes Resilience in Simplified Intestinal Microbial Communities. *mSphere* 6 (3), e0006821. doi: 10.1128/mSphere.00068-21

Bosco, N., and Noti, M. (2021). The aging gut microbiome and its impact on host immunity. *Genes Immun.* 22, 289-303. doi: 10.1038/s41435-021-00126-8

Bozkurt, H. S., and Kara, B. (2020). A new treatment for ulcerative colitis: Intracolonic and xyloglucan application. *Eur. J. Inflammation* 18. doi: 10.1177/ 2058739220942626

Bravo, J. A., Forsythe, P., Chew, M. V., Escaravage, E., Savignac, H. M., Dinan, T. G., et al. (2011). Ingestion of Lactobacillus strain regulates emotional behavior and central GABA receptor expression in a mouse via the vagus nerve. *Proc. Natl. Acad. Sci. U.S.A.* 108, 16050–16055. doi: 10.1073/pnas.1102999108

Buffington, S. A., Di Prisco, G. V., Auchtung, T. A., Ajami, N. J., Petrosino, J. F., and Costa-Mattioli, M. (2016). Microbial reconstitution reverses maternal diet-induced social and synaptic deficits in offspring. *Cell* 165, 1762–1775. doi: 10.1016/ j.cell.2016.06.001

Bullich, C., Keshavarzian, A., Garssen, J., Kraneveld, A., and Perez-Pardo, P. (2019). Gut vibes in parkinson's disease: the microbiota-gut-brain axis. *Mov. Disord. Clin. Pract.* 6, 639–651. doi: 10.1002/mdc3.12840

Burokas, A., Arboleya, S., Moloney, R. D., Peterson, V. L., Murphy, K., Clarke, G., et al. (2017). Targeting the microbiota-gut-brain axis: prebiotics have anxiolytic and antidepressant-like effects and reverse the impact of chronic stress in mice. *Biol. Psychiatry* 82, 472–487. doi: 10.1016/j.biopsych.2016.12.031

Caminero, A., Herran, A. R., Nistal, E., Perez-Andres, J., Vaquero, L., Vivas, S., et al. (2014). Diversity of the cultivable human gut microbiome involved in gluten metabolism: isolation of microorganisms with potential interest for coeliac disease. *FEMS Microbiol. Ecol.* 88, 309–319. doi: 10.1111/fem.2014.88.issue-2

organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Cani, P. D., Neyrinck, A. M., Fava, F., Knauf, C., Burcelin, R. G., Tuohy, K. M., et al. (2007). Selective increases of bifidobacteria in gut microflora improve high-fat-dietinduced diabetes in mice through a mechanism associated with endotoxaemia. *Diabetologia* 50, 2374–2383. doi: 10.1007/s00125-007-0791-0

Casas-Deza, D., Bernal-Monterde, V., Aranda-Alonso, A. N., Montil-Miguel, E., Julian-Gomara, A. B., Letona-Gimenez, L., et al. (2021). Age-related mortality in 61,993 confirmed COVID-19 cases over three epidemic waves in Aragon, Spain. Implications for vaccination programmes. *PloS One* 16, e0261061. doi: 10.1371/journal.pone.0261061

Chang, C. S., and Kao, C. Y. (2019). Current understanding of the gut microbiota shaping mechanisms. J. BioMed. Sci. 26, 59. doi: 10.1186/s12929-019-0554-5

Chen, J., Hall, S., and Vitetta, L. (2021). Altered gut microbial metabolites could mediate the effects of risk factors in Covid-19. *Rev. Med. Virol.* 31, 1–13. doi: 10.1002/rmv.2211

Chen, L., Xu, X., Wu, X., Cao, H., Li, X., Hou, Z., et al. (2022). A comparison of the composition and functions of the oral and gut microbiotas in Alzheimer's patients. *Front. Cell Infect. Microbiol.* 12. doi: 10.3389/fcimb.2022.942460

Chen, W., Zhang, S., Wu, J., Ye, T., Wang, S., Wang, P., et al. (2020). Butyrateproducing bacteria and the gut-heart axis in atherosclerosis. *Clin. Chim. Acta* 507, 236– 241. doi: 10.1016/j.cca.2020.04.037

Chen, X., D'souza, R., and Hong, S. T. (2013). The role of gut microbiota in the gutbrain axis: current challenges and perspectives. *Protein Cell* 4, 403–414. doi: 10.1007/ s13238-013-3017-x

Chen, Y. T., Yang, N. S., Lin, Y. C., Ho, S. T., Li, K. Y., Lin, J. S., et al. (2018). A combination of Lactobacillus Mali APS1 and dieting improved the efficacy of obesity treatment via manipulating gut microbiome in mice. *Sci. Rep.* 8, 6153. doi: 10.1038/s41598-018-23844-y

Cheng, W. T., Kantilal, H. K., and Davamani, F. (2020). The mechanism of bacteroides fragilis toxin contributes to colon cancer formation. *Malays J. Med. Sci.* 27, 9–21. doi: 10.21315/mjms

Chlebicz-Wojcik, A., and Slizewska, K. (2021). Probiotics, prebiotics, and synbiotics in the irritable bowel syndrome treatment: A review. *Biomolecules* 11. doi: 10.3390/biom11081154

Choi, W. J., Dong, H. J., Jeong, H. U., Ryu, D. W., Song, S. M., Kim, Y. R., et al. (2020). Lactobacillus plantarum LMT1-48 exerts anti-obesity effect in high-fat dietinduced obese mice by regulating expression of lipogenic genes. *Sci. Rep.* 10, 869. doi: 10.1038/s41598-020-57615-5

Chu, C., Yu, L., Li, Y., Guo, H., Zhai, Q., Chen, W., et al. (2023). Meta-analysis of randomized controlled trials of the effects of probiotics in Parkinson's disease. *Food Funct.* 14, 3406–3422. doi: 10.1039/D2FO03825K

Chung, H. J., Nguyen, T. T. B., Kim, H. J., and Hong, S. T. (2018). Gut microbiota as a missing link between nutrients and traits of human. *Front. Microbiol.* 9, 1510. doi: 10.3389/fmicb.2018.01510

Chung, H. J., Yu, J. G., Lee, I. A., Liu, M. J., Shen, Y. F., Sharma, S. P., et al. (2016). Intestinal removal of free fatty acids from hosts by Lactobacilli for the treatment of obesity. *FEBS Open Bio* 6, 64–76. doi: 10.1002/2211-5463.12024

Collado, M. C., Calabuig, M., and Sanz, Y. (2007). Differences between the fecal microbiota of coeliac infants and healthy controls. *Curr. Issues Intest Microbiol*, 8, 9–14.

Corrêa-Oliveira, R., Fachi, J. L., Vieira, A., Sato, F. T., and Vinolo, M. A. (2016). Regulation of immune cell function by short-chain fatty acids. *Clin. Transl. Immunol.* 5, e73. doi: 10.1038/cti.2016.17

Cox, L. M., and Weiner, H. L. (2018). Microbiota signaling pathways that influence neurologic disease. *Neurotherapeutics* 15, 135–145. doi: 10.1007/s13311-017-0598-8

Davies, G. A., Bryant, A. R., Reynolds, J. D., Jirik, F. R., and Sharkey, K. A. (2006). Prion diseases and the gastrointestinal tract. *Can. J. Gastroenterol.* 20, 18–24. doi: 10.1155/2006/184528

Degruttola, A. K., Low, D., Mizoguchi, A., and Mizoguchi, E. (2016). Current understanding of dysbiosis in disease in human and animal models. *Inflammation Bowel Dis.* 22, 1137–1150. doi: 10.1097/MIB.00000000000750

Dehghan, P., Pourghassem Gargari, B., and Asghari Jafar-Abadi, M. (2014). Oligofructose-enriched inulin improves some inflammatory markers and metabolic endotoxemia in women with type 2 diabetes mellitus: a randomized controlled clinical trial. *Nutrition* 30, 418-423. doi: 10.1016/j.nut.2013.09.005

Depommier, C., Everard, A., Druart, C., Plovier, H., Van Hul, M., Vieira-Silva, S., et al. (2019). Supplementation with Akkermansia muciniphila in overweight and obese

human volunteers: a proof-of-concept exploratory study. Nat. Med. 25, 1096-1103. doi: 10.1038/s41591-019-0495-2

De Sousa Moraes, L. F., Grzeskowiak, L. M., De Sales Teixeira, T. F., and Gouveia Peluzio Mdo, C. (2014). Intestinal microbiota and probiotics in celiac disease. *Clin. Microbiol. Rev.* 27, 482–489. doi: 10.1128/CMR.00106-13

Dickerson, F. B., Stallings, C., Origoni, A., Katsafanas, E., Savage, C. L., Schweinfurth, L. A., et al. (2014). Effect of probiotic supplementation on schizophrenia symptoms and association with gastrointestinal functioning: a randomized, placebo-controlled trial. *Prim Care Companion CNS Disord.* 16. doi: 10.4088/PCC.13m01579

Ding, Y., Bu, F., Chen, T., Shi, G., Yuan, X., Feng, Z., et al. (2021). A next-generation probiotic: Akkermansia muciniphila ameliorates chronic stress-induced depressive-like behavior in mice by regulating gut microbiota and metabolites. *Appl. Microbiol. Biotechnol.* 105, 8411–8426. doi: 10.1007/s00253-021-11622-2

Di Vincenzo, F., Del Gaudio, A., Petito, V., Lopetuso, L. R., and Scaldaferri, F. (2024). Gut microbiota, intestinal permeability, and systemic inflammation: a narrative review. *Intern. Emerg. Med.* 19, 275–293. doi: 10.1007/s11739-023-03374-w

Doll, J. P. K., Vazquez-Castellanos, J. F., Schaub, A. C., Schweinfurth, N., Kettelhack, C., Schneider, E., et al. (2022). Fecal microbiota transplantation (FMT) as an adjunctive therapy for depression-case report. *Front. Psychiatry* 13, 815422. doi: 10.3389/fpsyt.2022.815422

Dooling, S. W., Sgritta, M., Wang, I. C., Duque, A., and Costa-Mattioli, M. (2022). The effect of limosilactobacillus reuteri on social behavior is independent of the adaptive immune system. *mSystems* 7, e0035822. doi: 10.1128/msystems.00358-22

Drabińska, N., Krupa-Kozak, U., and Jarocka-Cyrta, E. (2020). Intestinal Permeability in Children with Celiac Disease after the Administration of Oligofructose-Enriched Inulin into a Gluten-Free Diet-Results of a Randomized, Placebo-Controlled, Pilot Trial. *Nutrients* 12. doi: 10.3390/nu12061736

Duan, M., Wang, Y., Zhang, Q., Zou, R., Guo, M., and Zheng, H. (2021). Characteristics of gut microbiota in people with obesity. *PloS One* 16, e0255446. doi: 10.1371/journal.pone.0255446

Dunn, A. B., Jordan, S., Baker, B. J., and Carlson, N. S. (2017). The maternal infant microbiome: considerations for labor and birth. *MCN Am. J. Matern Child Nurs.* 42, 318–325. doi: 10.1097/NMC.0000000000373

Dupont, H. L., Suescun, J., Jiang, Z. D., Brown, E. L., Essigmann, H. T., Alexander, A. S., et al. (2023). Fecal microbiota transplantation in Parkinson's disease-A randomized repeat-dose, placebo-controlled clinical pilot study. *Front. Neurol.* 14, 1104759. doi: 10.3389/fneur.2023.1104759

Engevik, M. A., Luk, B., Chang-Graham, A. L., Hall, A., Herrmann, B., Ruan, W., et al. (2019). Bifidobacterium dentium Fortifies the Intestinal Mucus Layer via Autophagy and Calcium Signaling Pathways. *mBio* 10 (3), e01087-e01019. doi: 10.1128/mBio.01087-19

Fang, J., Wang, H., Zhou, Y., Zhang, H., Zhou, H., and Zhang, X. (2021). Slimy partners: the mucus barrier and gut microbiome in ulcerative colitis. *Exp. Mol. Med.* 53, 772–787. doi: 10.1038/s12276-021-00617-8

Farida, E., Nuraida, L., Giriwono, P. E., and Jenie, B. S. L. (2020). Lactobacillus rhamnosus Reduces Blood Glucose Level through Downregulation of Gluconeogenesis Gene Expression in Streptozotocin-Induced Diabetic Rats. *Int. J. Food Sci.* 2020, 6108575. doi: 10.1155/2020/6108575

Farup, P. G., Rudi, K., and Hestad, K. (2016). Faecal short-chain fatty acids - a diagnostic biomarker for irritable bowel syndrome? *BMC Gastroenterol.* 16, 51. doi: 10.1186/s12876-016-0446-z

Fernández-Tomé, S., Marin, A. C., Ortega Moreno, L., Baldan-Martin, M., Mora-Gutérrez, I., Lanas-Gimeno, A., et al. (2019). Immunomodulatory Effect of Gut Microbiota-Derived Bioactive Peptides on Human Immune System from Healthy Controls and Patients with Inflammatory Bowel Disease. *Nutrients* 11, 2605. doi: 10.3390/nu11112605

Fernandes, D., and Andreyev, J. (2022). The role of the human gut microbiome in inflammatory bowel disease and radiation enteropathy. *Microorganisms* 10. doi: 10.3390/microorganisms10081613

Filosa, S., Di Meo, F., and Crispi, S. (2018). Polyphenols-gut microbiota interplay and brain neuromodulation. *Neural Regener. Res.* 13, 2055–2059. doi: 10.4103/1673-5374.241429

Forsberg, G., Fahlgren, A., Horstedt, P., Hammarstrom, S., Hernell, O., and Hammarstrom, M. L. (2004). Presence of bacteria and innate immunity of intestinal epithelium in childhood celiac disease. *Am. J. Gastroenterol.* 99, 894–904. doi: 10.1111/j.1572-0241.2004.04157.x

Fukuda, S., Toh, H., Hase, K., Oshima, K., Nakanishi, Y., Yoshimura, K., et al. (2011). Bifidobacteria can protect from enteropathogenic infection through production of acetate. *Nature* 469 (7331), 543–547. doi: 10.1038/nature09646

Gacesa, R., Kurilshikov, A., Vich Vila, A., Sinha, T., Klaassen, M. A. Y., Bolte, L. A., et al. (2022). Environmental factors shaping the gut microbiome in a Dutch population. *Nature* 604, 732–739. doi: 10.1038/s41586-022-04567-7

Gao, R., Zhu, C., Li, H., Yin, M., Pan, C., Huang, L., et al. (2018). Dysbiosis signatures of gut microbiota along the sequence from healthy, young patients to those with overweight and obesity. *Obes. (Silver Spring)* 26, 351–361. doi: 10.1002/oby.22088

Garg, P. (2021). Inflammation in irritable bowel syndrome (IBS): role of psyllium fiber supplementation in decreasing inflammation and physiological management of IBS. *Turk J. Gastroenterol.* 32, 108–110. doi: 10.5152/tjg

Geng, S., Yang, L., Cheng, F., Zhang, Z., Li, J., Liu, W., et al. (2019). Gut microbiota are associated with psychological stress-induced defections in intestinal and blood-brain barriers. *Front. Microbiol.* 10:3067. doi: 10.3389/fmicb.2019.03067

Gerhardt, S., and Mohajeri, M. H. (2018). Changes of colonic bacterial composition in parkinson's disease and other neurodegenerative diseases. *Nutrients* 10. doi: 10.3390/ nu10060708

Ghaderi, A., Banafshe, H. R., Mirhosseini, N., Moradi, M., Karimi, M. A., Mehrzad, F., et al. (2019). Clinical and metabolic response to vitamin D plus probiotic in schizophrenia patients. *BMC Psychiatry* 19, 77. doi: 10.1186/s12888-019-2059-x

Gorecki, A. M., Preskey, L., Bakeberg, M. C., Kenna, J. E., Gildenhuys, C., Macdougall, G., et al. (2019). Altered gut microbiome in parkinson's disease and the influence of lipopolysaccharide in a human alpha-synuclein over-expressing mouse model. *Front. Neurosci.* 13, 839. doi: 10.3389/fnins.2019.00839

Gozd-Barszczewska, A., Koziol-Montewka, M., Barszczewski, P., Mlodzinska, A., and Huminska, K. (2017). Gut microbiome as a biomarker of cardiometabolic disorders. *Ann. Agric. Environ. Med.* 24, 416–422. doi: 10.26444/aaem/75456

Grossi, E., Melli, S., Dunca, D., and Terruzzi, V. (2016). Unexpected improvement in core autism spectrum disorder symptoms after long-term treatment with probiotics. *SAGE Open Med. Case Rep.* 4, 2050313X16666231. doi: 10.1177/ 2050313X16666231

Guarino, M. P. L., Altomare, A., Emerenziani, S., Di Rosa, C., Ribolsi, M., Balestrieri, P., et al. (2020). Mechanisms of action of prebiotics and their effects on gastro-intestinal disorders in adults. *Nutrients* 12. doi: 10.3390/nu12041037

Guo, C., Zhou, Q., Li, M., Zhou, L., Xu, L., Zhang, Y., et al. (2020). Breastfeeding restored the gut microbiota in caesarean section infants and lowered the infection risk in early life. *BMC Pediatr.* 20, 532. doi: 10.1186/s12887-020-02433-x

Ha, S., Oh, D., Lee, S., Park, J., Ahn, J., Choi, S., et al. (2021). Altered gut microbiota in korean children with autism spectrum disorders. *Nutrients* 13. doi: 10.3390/ nu13103300

Haiser, H. J., Gootenberg, D. B., Chatman, K., Sirasani, G., Balskus, E. P., and Turnbaugh, P. J. (2013). Predicting and manipulating cardiac drug inactivation by the human gut bacterium Eggerthella lenta. *Science*. 341 (6143), 295–298. doi: 10.1126/ science.1235872

Hasan, N., and Yang, H. (2019). Factors affecting the composition of the gut microbiota, and its modulation. *PeerJ* 7, e7502. doi: 10.7717/peerj.7502

Holscher, H. D. (2017). Dietary fiber and prebiotics and the gastrointestinal microbiota. *Gut Microbes* 8 (2), 172–184. doi: 10.1080/19490976.2017.1290756

Hou, K., Wu, Z. X., Chen, X. Y., Wang, J. Q., Zhang, D., Xiao, C., et al. (2022). Microbiota in health and diseases. *Signal Transduct Target Ther.* 7, 135. doi: 10.1038/ s41392-022-00974-4

Hsieh, P. S., Ho, H. H., Hsieh, S. H., Kuo, Y. W., Tseng, H. Y., Kao, H. F., et al. (2020). Lactobacillus salivarius AP-32 and Lactobacillus reuteri GL-104 decrease glycemic levels and attenuate diabetes-mediated liver and kidney injury in db/db mice. *BMJ Open Diabetes Res. Care* 8. doi: 10.1136/bmjdrc-2019-001028

Hsieh, F. C., Lee, C. L., Chai, C. Y., Chen, W. T., Lu, Y. C., and Wu, C. S. (2013). Oral administration of Lactobacillus reuteri GMNL-263 improves insulin resistance and ameliorates hepatic steatosis in high fructose-fed rats. *Nutr. Metab. (Lond)* 10, 35. doi: 10.1186/1743-7075-10-35

Hu, B., Das, P., Lv, X., Shi, M., Aa, J., Wang, K., et al. (2022). Effects of 'Healthy' Fecal microbiota transplantation against the deterioration of depression in fawn-hooded rats. *mSystems* 7, e0021822. doi: 10.1128/msystems.00218-22

Hu, X. F., Zhang, W. Y., Wen, Q., Chen, W. J., Wang, Z. M., Chen, J., et al. (2019). Fecal microbiota transplantation alleviates myocardial damage in myocarditis by restoring the microbiota composition. *Pharmacol. Res.* 139, 412–421. doi: 10.1016/ j.phrs.2018.11.042

Husebye, E., Hellström, P. M, Sundler, F., Chen, J., and Midtvedt, T. (2001). Influence of microbial species on small intestinal myoelectric activity and transit in germ-free rats. *Am. J. Physiol. Gastrointest. Liver Physiol.* 280 (3), G368–80. doi: 10.1152/ajpgi.2001.280.3.G368

Inlender, T., Nissim-Eliraz, E., Stavely, R., Hotta, R., Goldstein, A. M., Yagel, S., et al. (2021). Homeostasis of mucosal glial cells in human gut is independent of microbiota. *Sci. Rep.* 11 (1), 12796. doi: 10.1038/s41598-021-92384-9

Jang, H. M., Lee, K. E., and Kim, D. H. (2019). The Preventive and Curative Effects of Lactobacillus reuteri NK33 and Bifidobacterium adolescentis NK98 on Immobilization Stress-Induced Anxiety/Depression and Colitis in Mice. *Nutrients* 11. doi: 10.3390/ nu11040819

Jasemi, S., Emaneini, M., Fazeli, M. S., Ahmadinejad, Z., Nomanpour, B., Sadeghpour Heravi, F., et al. (2020). Toxigenic and non-toxigenic patterns I and biofilm-forming ability in Bacteroides fragilis strains isolated from patients diagnosed with colorectal cancer. *Gut Pathog.* 12, 28. doi: 10.1186/s13099-020-00366-5

Jia, L., Li, D., Feng, N., Shamoon, M., Sun, Z., Ding, L., et al. (20177046). Antidiabetic Effects of Clostridium butyricum CGMCC0313.1 through Promoting the Growth of Gut Butyrate-producing Bacteria in Type 2 Diabetic Mice. *Sci. Rep.* 7. doi: 10.1038/s41598-017-07335-0

Jie, Z., Xia, H., Zhong, S. L., Feng, Q., Li, S., Liang, S., et al. (2017). The gut microbiome in atherosclerotic cardiovascular disease. *Nat. Commun.* 8, 845. doi: 10.1038/s41467-017-00900-1

Joseph, N., Clayton, J. B., Hoops, S. L., Linhardt, C. A., Mohd Hashim, A., Mohd Yusof, B. N., et al. (2020). Alteration of the gut microbiome in normal and overweight school children from selangor with lactobacillus fermented milk administration. *Evol. Bioinform. Online* 16, 1176934320965943. doi: 10.1177/1176934320965943

Kabeerdoss, J., Sankaran, V., Pugazhendhi, S., and Ramakrishna, B. S. (2013). Clostridium leptum group bacteria abundance and diversity in the fecal microbiota of patients with inflammatory bowel disease: a case-control study in India. *BMC Gastroenterol.* 13, 20. doi: 10.1186/1471-230X-13-20

Kabouridis, P. S., Lasrado, R., McCallum, S., Chng, S. H., Snippert, H. J., Clevers, H., et al. (2015). Microbiota controls the homeostasis of glial cells in the gut lamina propria. *Neuron* 85 (2), 289–295. doi: 10.1016/j.neuron.2014.12.037

Kang, S., Denman, S. E., Morrison, M., Yu, Z., Dore, J., Leclerc, M., et al. (2010). Dysbiosis of fecal microbiota in Crohn's disease patients as revealed by a custom phylogenetic microarray. *Inflammation Bowel Dis.* 16, 2034–2042. doi: 10.1002/ibd.21319

Kang, N. R., and Kim, M. D. (2011). Tardive dyskinesia: treatment with aripiprazole. *Clin. Psychopharmacol. Neurosci.* 9, 1–8. doi: 10.9758/cpn.2011.9.1.1

Kang, D., Li, Z. P., and Ji, G. E. (2018). Anti-obesity effects of a mixture of fermented ginseng, BORI, and CH88 in high-fat diet-fed mice. *J. Microbiol. Biotechnol.* 28, 688–696. doi: 10.4014/jmb.1801.01016

Kang, V., Wagner, G. C., and Ming, X. (2014). Gastrointestinal dysfunction in children with autism spectrum disorders. *Autism Res.* 7, 501–506. doi: 10.1002/aur.1386

Karlsson, F. H., Fak, F., Nookaew, I., Tremaroli, V., Fagerberg, B., Petranovic, D., et al. (20121245). Symptomatic atherosclerosis is associated with an altered gut metagenome. *Nat. Commun.* 3. doi: 10.1038/ncomms2266

Kasai, C., Sugimoto, K., Moritani, I., Tanaka, J., Oya, Y., Inoue, H., et al. (2015). Comparison of the gut microbiota composition between obese and non-obese individuals in a Japanese population, as analyzed by terminal restriction fragment length polymorphism and next-generation sequencing. *BMC Gastroenterol.* 15, 100. doi: 10.1186/s12876-015-0330-2

Kedia, S., Virmani, S., Vuyyuru, S. K., Kumar, P., Kante, B., Sahu, P., et al. (2022). Faecal microbiota transplantation with anti-inflammatory diet (FMT-AID) followed by anti-inflammatory diet alone is effective in inducing and maintaining remission over 1 year in mild to moderate ulcerative colitis: a randomised controlled trial. *Gut* 71, 2401– 2413. doi: 10.1136/gutjn1-2022-327811

Kim, S., Shin, Y. C., Kim, T. Y., Kim, Y., Lee, Y. S., Lee, S. H., et al. (2021). Mucin degrader Akkermansia muciniphila accelerates intestinal stem cell-mediated epithelial development. *Gut Microbes* 13 (1), 1–20. doi: 10.1080/19490976.2021.1892441

Kim, E. S., Yoon, B. H., Lee, S. M., Choi, M., Kim, E. H., Lee, B. W., et al. (2022). Fecal microbiota transplantation ameliorates atherosclerosis in mice with C1q/TNF-related protein 9 genetic deficiency. *Exp. Mol. Med.* 54, 103–114. doi: 10.1038/s12276-022-00728-w

Kok, C. R., Rose, D., and Hutkins, R. (2023). Predicting personalized responses to dietary fiber interventions: opportunities for modulation of the gut microbiome to improve health. *Annu. Rev. Food Sci. Technol.* 14, 157–182. doi: 10.1146/annurev-food-060721-015516

Koppel, N., Maini Rekdal, V., and Balskus, E. P. (2017). Chemical transformation of xenobiotics by the human gut microbiota. *Science(New York N.Y.)* 356 (6344), eaag2770. doi: 10.1126/science.aag2770

Koopen, A., Witjes, J., Wortelboer, K., Majait, S., Prodan, A., Levin, E., et al. (2022). Duodenal Anaerobutyricum soehngenii infusion stimulates GLP-1 production, ameliorates glycaemic control and beneficially shapes the duodenal transcriptome in metabolic syndrome subjects: a randomised double-blind placebo-controlled crossover study. *Gut* 71, 1577–1587. doi: 10.1136/gutjnl-2020-323297

Kostic, A. D., Gevers, D., Pedamallu, C. S., Michaud, M., Duke, F., Earl, A. M., et al. (2012). Genomic analysis identifies association of Fusobacterium with colorectal carcinoma. *Genome Res.* 22, 292–298. doi: 10.1101/gr.126573.111

Kuai, X. Y., Yao, X. H., Xu, L. J., Zhou, Y. Q., Zhang, L. P., Liu, Y., et al. (2021). Evaluation of fecal microbiota transplantation in Parkinson's disease patients with constipation. *Microb. Cell Fact* 20, 98. doi: 10.1186/s12934-021-01589-0

Lakshmanan, A. P., Al Za'abi, M., Ali, B. H., and Terranegra, A. (2021). The influence of the prebiotic gum acacia on the intestinal microbiome composition in rats with experimental chronic kidney disease. *BioMed. Pharmacother.* 133, 110992. doi: 10.1016/j.biopha.2020.110992

Lapaquette, P., Glasser, A. L., Huett, A., Xavier, R. J., and Darfeuille-Michaud, A. (2010). Crohn's disease-associated adherent-invasive E. coli are selectively favoured by impaired autophagy to replicate intracellularly. *Cell Microbiol.* 12, 99–113. doi: 10.1111/cmi.2010.12.issue-1

Larsen, N., Vogensen, F. K., Van Den Berg, F. W., Nielsen, D. S., Andreasen, A. S., Pedersen, B. K., et al. (2010). Gut microbiota in human adults with type 2 diabetes differs from non-diabetic adults. *PloS One* 5, e9085. doi: 10.1371/journal.pone.0009085

Lee, Y. S., Lee, D., Park, G. S., Ko, S. H., Park, J., Lee, Y. K., et al. (2021). Lactobacillus plantarum HAC01 ameliorates type 2 diabetes in high-fat diet and streptozotocininduced diabetic mice in association with modulating the gut microbiota. *Food Funct.* 12, 6363–6373. doi: 10.1039/D1FO00698C

Lew, L. C., Hor, Y. Y., Yusoff, N. A. A., Choi, S. B., Yusoff, M. S. B., Roslan, N. S., et al. (2019). Probiotic Lactobacillus plantarum P8 alleviated stress and anxiety while enhancing memory and cognition in stressed adults: A randomised, double-blind, placebo-controlled study. *Clin. Nutr.* 38, 2053–2064. doi: 10.1016/j.clnu.2018.09.010

Li, Q., Chang, Y., Zhang, K., Chen, H., Tao, S., and Zhang, Z. (2020b5450). Implication of the gut microbiome composition of type 2 diabetic patients from northern China. *Sci. Rep.* 10. doi: 10.1038/s41598-020-62224-3

Li, N., Chen, H., Cheng, Y., Xu, F., Ruan, G., Ying, S., et al. (2021a). Fecal microbiota transplantation relieves gastrointestinal and autism symptoms by improving the gut microbiota in an open-label study. *Front. Cell. Infect. Microbiol.* 11, 759435. doi: 10.3389/fcimb.2021.801376

Li, H., He, J., and Jia, W. (2016a). The influence of gut microbiota on drug metabolism and toxicity. *Expert Opin. Drug Metab. Toxicol.* 12, 31–40. doi: 10.1517/17425255.2016.1121234

Li, B., He, Y., Ma, J., Huang, P., Du, J., Cao, L., et al. (2019a). Mild cognitive impairment has similar alterations as Alzheimer's disease in gut microbiota. *Alzheimers Dement* 15, 1357–1366. doi: 10.1016/j.jalz.2019.07.002

Li, J., Lin, S., Vanhoutte, P. M., Woo, C. W., and Xu, A. (2016b). Akkermansia muciniphila protects against atherosclerosis by preventing metabolic endotoxemiainduced inflammation in apoe-/- mice. *Circulation* 133, 2434–2446. doi: 10.1161/ CIRCULATIONAHA.115.019645

Li, B., Luo, X. F., Liu, S. W., Zhao, N., Li, H. N., Zhang, W., et al. (2020a). Abdominal massage reduces visceral hypersensitivity via regulating GDNF and PI3K/AKT signal pathway in a rat model of irritable bowel syndrome. *Evid Based Complement Alternat Med.* 2020, 3912931. doi: 10.1155/2020/3912931

Li, S., Song, J., Ke, P., Kong, L., Lei, B., Zhou, J., et al. (2021b9743). The gut microbiome is associated with brain structure and function in schizophrenia. *Sci. Rep.* 11. doi: 10.1038/s41598-021-89166-8

Li, F., Wang, M., Wang, J., Li, R., and Zhang, Y. (2019b). Alterations to the gut microbiota and their correlation with inflammatory factors in chronic kidney disease. *Front. Cell Infect. Microbiol.* 9, 206. doi: 10.3389/fcimb.2019.00206

Lin, Y. C., Chen, Y. T., Hsieh, H. H., and Chen, M. J. (2016). Effect of APS1 and L. M1 on obesity and glucose homeostasis in diet-induced obese mice. *J. Funct. Foods* 23, 580–589. doi: 10.1016/j.jff.2016.03.015

Lin, H., Guo, Q., Wen, Z., Tan, S., Chen, J., Lin, L., et al. (2021a). The multiple effects of fecal microbiota transplantation on diarrhea-predominant irritable bowel syndrome (IBS-D) patients with anxiety and depression behaviors. *Microb. Cell Fact* 20, 233. doi: 10.1186/s12934-021-01720-1

Lin, T. Y., Wu, P. H., Lin, Y. T., and Hung, S. C. (2021b). Gut dysbiosis and mortality in hemodialysis patients. *NPJ Biofilms Microbiomes* 7, 20. doi: 10.1038/s41522-021-00191-x

Liu, S., Li, E., Sun, Z., Fu, D., Duan, G., Jiang, M., et al. (2019). Altered gut microbiota and short chain fatty acids in Chinese children with autism spectrum disorder. *Sci. Rep.* 9, 287. doi: 10.1038/s41598-018-36430-z

Liu, S., Moon, C. D., Zheng, N., Huws, S., Zhao, S., and Wang, J. (2022a). Opportunities and challenges of using metagenomic data to bring uncultured microbes into cultivation. *Microbiome* 10, 76. doi: 10.1186/s40168-022-01272-5

Liu, H. N., Wu, H., Chen, Y. Z., Chen, Y. J., Shen, X. Z., and Liu, T. T. (2017). Altered molecular signature of intestinal microbiota in irritable bowel syndrome patients compared with healthy controls: A systematic review and meta-analysis. *Dig Liver Dis.* 49, 331–337. doi: 10.1016/j.dld.2017.01.142

Liu, Y., Zhang, R., Yao, B., Yang, J., Ge, H., Zheng, S., et al. (2022b). Metagenomics next-generation sequencing provides insights into the causative pathogens from critically ill patients with pneumonia and improves treatment strategies. *Front. Cell Infect. Microbiol.* 12, 1094518. doi: 10.3389/fcimb.2022.1094518

Lkhagva, E., Chung, H. J., Ahn, J. S., and Hong, S. T. (2021a). Host factors affect the gut microbiome more significantly than diet shift. *Microorganisms* 9. doi: 10.3390/microorganisms9122520

Lkhagva, E., Chung, H. J., Hong, J., Tang, W. H. W., Lee, S. I., Hong, S. T., et al. (2021b). The regional diversity of gut microbiome along the GI tract of male C57BL/6 mice. *BMC Microbiol.* 21, 44. doi: 10.1186/s12866-021-02099-0

Lopez-Siles, M., Martinez-Medina, M., Suris-Valls, R., Aldeguer, X., Sabat-Mir, M., Duncan, S. H., et al. (2016). Changes in the abundance of faecalibacterium prausnitzii phylogroups I and II in the intestinal mucosa of inflammatory bowel disease and patients with colorectal cancer. *Inflammation Bowel Dis.* 22, 28–41. doi: 10.1097/ MIB.0000000000000590

Luo, S., He, L., Zhang, H., Li, Z., Liu, C., and Chen, T. (2022). Arabinoxylan from rice bran protects mice against high-fat diet-induced obesity and metabolic inflammation by modulating gut microbiota and short-chain fatty acids. *Food Funct.* 13, 7707–7719. doi: 10.1039/D2FO00569G

Mahbub, N. U., Islam, M. M., Hong, S.-T., and Chung, H.-J. (2024). Dysbiosis of the gut microbiota and its effect on α -synuclein and prion protein misfolding: consequences for neurodegeneration. *Frontiers in Cellular and Infection Microbiology* 14. doi: 10.3389/fcimb.2024.1348279

Majeed, M., Nagabhushanam, K., Natarajan, S., Sivakumar, A., Ali, F., Pande, A., et al. (2016). Bacillus coagulans MTCC 5856 supplementation in the management of diarrhea predominant Irritable Bowel Syndrome: a double blind randomized placebo controlled pilot clinical study. *Nutr. J.* 15, 21. doi: 10.1186/s12937-016-0140-6

Malla, M. A., Dubey, A., Kumar, A., Yadav, S., Hashem, A., and Abd Allah, E. F. (2018). Exploring the human microbiome: the potential future role of next-generation sequencing in disease diagnosis and treatment. *Front. Immunol.* 9:2868. doi: 10.3389/fimmu.2018.02868

Mańkowska-Wierzbicka, D., Stelmach-Mardas, M., Gabryel, M., Tomczak, H., Skrzypczak-Zielińska, M., Zakerska-Banaszak, O., et al. (2020). The effectiveness of multi-session FMT treatment in active ulcerative colitis patients: A pilot study. *Biomedicines.* 8, 268. doi: 10.3390/biomedicines8080268

Marasco, G., Di Biase, A. R., Schiumerini, R., Eusebi, L. H., Iughetti, L., Ravaioli, F., et al. (2016). Gut microbiota and celiac disease. *Dig Dis. Sci.* 61, 1461–1472. doi: 10.1007/s10620-015-4020-2

Martinez-Cuesta, M. C., Del Campo, R., Garriga-Garcia, M., Pelaez, C., and Requena, T. (2021). Taxonomic characterization and short-chain fatty acids production of the obese microbiota. *Front. Cell Infect. Microbiol.* 11, 598093. doi: 10.3389/fcimb.2021.598093

Martoni, C. J., Srivastava, S., and Leyer, G. J. (2020). Lactobacillus acidophilus DDS-1 and Bifidobacterium lactis UABla-12 Improve Abdominal Pain Severity and Symptomology in Irritable Bowel Syndrome: Randomized Controlled Trial. *Nutrients* 12. doi: 10.3390/nu12020363

Mazur, M., Furgala, A., Jablonski, K., Mach, T., and Thor, P. (2012). Autonomic nervous system activity in constipation-predominant irritable bowel syndrome patients. *Med. Sci. Monit* 18, CR493–CR499. doi: 10.12659/MSM.883269

Meng, F., Zhang, F., Meng, M., Chen, Q., Yang, Y., Wang, W., et al. (2023). Effects of the synbiotic composed of mangiferin and Lactobacillus reuteri 1-12 on type 2 diabetes mellitus rats. *Front. Microbiol.* 14, 1158652. doi: 10.3389/fmicb.2023.1158652

Milani, C., Duranti, S., Bottacini, F., Casey, E., Turroni, F., Mahony, J., et al. (2017). The first microbial colonizers of the human gut: composition, activities, and health implications of the infant gut microbiota. *Microbiol. Mol. Biol. Rev.* 81. doi: 10.1128/MMBR.00036-17

Mirsepasi-Lauridsen, H. C., Vallance, B. A., Krogfelt, K. A., and Petersen, A. M. (2019). Escherichia coli pathobionts associated with inflammatory bowel disease. *Clin. Microbiol. Rev.* 32. doi: 10.1128/CMR.00060-18

Miyoshi, M., Ogawa, A., Higurashi, S., and Kadooka, Y. (2014). Anti-obesity effect of Lactobacillus gasseri SBT2055 accompanied by inhibition of pro-inflammatory gene expression in the visceral adipose tissue in diet-induced obese mice. *Eur. J. Nutr.* 53, 599–606. doi: 10.1007/s00394-013-0568-9

Mohajeri, M. H., La Fata, G., Steinert, R. E., and Weber, P. (2018). Relationship between the gut microbiome and brain function. *Nutr. Rev.* 76, 481–496. doi: 10.1093/ nutrit/nuy009

Nagao-Kitamoto, H., and Kamada, N. (2017). Host-microbial cross-talk in inflammatory bowel disease. *Immune Netw.* 17, 1–12. doi: 10.4110/in.2017.17.1.1

Nagpal, R., Neth, B. J., Wang, S., Craft, S., and Yadav, H. (2019). Modified Mediterranean-ketogenic diet modulates gut microbiome and short-chain fatty acids in association with Alzheimer's disease markers in subjects with mild cognitive impairment. *EBioMedicine* 47, 529–542. doi: 10.1016/j.ebiom.2019.08.032

Nakabayashi, I., Nakamura, M., Kawakami, K., Ohta, T., Kato, I., Uchida, K., et al. (2011). Effects of synbiotic treatment on serum level of p-cresol in haemodialysis patients: a preliminary study. *Nephrol. Dial Transplant.* 26, 1094–1098. doi: 10.1093/ndt/gfq624

Nakade, Y., Iwata, Y., Furuichi, K., Mita, M., Hamase, K., Konno, R., et al. (2018). Gut microbiota-derived D-serine protects against acute kidney injury. *JCI Insight* 3. doi: 10.1172/jci.insight.97957

Nandwana, V., Nandwana, N. K., Das, Y., Saito, M., Panda, T., Das, S., et al. (2022). The role of microbiome in brain development and neurodegenerative diseases. *Molecules* 27. doi: 10.3390/molecules27113402

Natarajan, N., and Pluznick, J. L. (2014). From microbe to man: the role of microbial short chain fatty acid metabolites in host cell biology. *Am. J. Physiol. Cell Physiol.* 307, C979–C985. doi: 10.1152/ajpcell.00228.2014

Nemet, I., Saha, P. P., Gupta, N., Zhu, W., Romano, K. A., Skye, S. M., et al. (2020). A cardiovascular disease-linked gut microbial metabolite acts via adrenergic receptors. *Cell* 180 862-877, e822. doi: 10.1016/j.cell.2020.02.016

Neufeld, K. A., Kang, N., Bienenstock, J., and Foster, J. A. (2011a). Effects of intestinal microbiota on anxiety-like behavior. *Commun. Integr. Biol.* 4, 492–494. doi: 10.4161/ cib.15702

Neufeld, K. M., Kang, N., Bienenstock, J., and Foster, J. A. (2011b). Reduced anxietylike behavior and central neurochemical change in germ-free mice. *Neurogastroenterol Motil.* 23 255-264, e119. doi: 10.1111/nmo.2011.23.issue-3

Neyrinck, A. M., Possemiers, S., Druart, C., Van De Wiele, T., De Backer, F., Cani, P. D., et al. (2011). Prebiotic effects of wheat arabinoxylan related to the increase in bifidobacteria, Roseburia and Bacteroides/Prevotella in diet-induced obese mice. *PloS One* 6, e20944. doi: 10.1371/journal.pone.0020944

Nguyen, T. T. B., Chung, H. J., Kim, H. J., and Hong, S. T. (2019). Establishment of an ideal gut microbiota to boost healthy growth of neonates. *Crit. Rev. Microbiol.* 45, 118–129. doi: 10.1080/1040841X.2018.1561643

Niibo, M., Shirouchi, B., Umegatani, M., Morita, Y., Ogawa, A., Sakai, F., et al. (2019). Probiotic Lactobacillus gasseri SBT2055 improves insulin secretion in a diabetic rat model. *J. Dairy Sci.* 102, 997–1006. doi: 10.3168/jds.2018-15203

Oh, D., and Cheon, K. A. (2020). Alteration of gut microbiota in autism spectrum disorder: an overview. *Soa Chongsonyon Chongsin Uihak* 31, 131–145. doi: 10.5765/jkacap.190039

Okubo, R., Koga, M., Katsumata, N., Odamaki, T., Matsuyama, S., Oka, M., et al. (2019). Effect of bifidobacterium breve A-1 on anxiety and depressive symptoms in

schizophrenia: A proof-of-concept study. J. Affect. Disord. 245, 377–385. doi: 10.1016/j.jad.2018.11.011

Olivares, M., Castillejo, G., Varea, V., and Sanz, Y. (2014). Double-blind, randomised, placebo-controlled intervention trial to evaluate the effects of Bifidobacterium longum CECT 7347 in children with newly diagnosed coeliac disease. *Br. J. Nutr.* 112, 30–40. doi: 10.1017/S0007114514000609

Pantoja-Feliciano, I. G., Clemente, J. C., Costello, E. K., Perez, M. E., Blaser, M. J., Knight, R., et al. (2013). Biphasic assembly of the murine intestinal microbiota during early development. *ISME J.* 7, 1112–1115. doi: 10.1038/ismej.2013.15

Park, J., Kim, M., Kang, S. G., Jannasch, A. H., Cooper, B., Patterson, J., et al. (2015). Short-chain fatty acids induce both effector and regulatory T cells by suppression of histone deacetylases and regulation of the mTOR-S6K pathway. *Mucosal Immunol.* 8 (1), 80–93. doi: 10.1038/mi.2014.44

Patrono, E., Svoboda, J., and Stuchlik, A. (2021). Schizophrenia, the gut microbiota, and new opportunities from optogenetic manipulations of the gut-brain axis. *Behav. Brain Funct.* 17, 7. doi: 10.1186/s12993-021-00180-2

Peddinti, V., Avaghade, M. M., Suthar, S. U., Rout, B., Gomte, S. S., Agnihotri, T. G., et al. (2024). Gut instincts: Unveiling the connection between gut microbiota and Alzheimer's disease. *Clin. Nutr. ESPEN* 60, 266–280. doi: 10.1016/j.clnesp.2024.02.019

Perez-Munoz, M. E., Arrieta, M. C., Ramer-Tait, A. E., and Walter, J. (2017). A critical assessment of the "sterile womb" and "*in utero* colonization" hypotheses: implications for research on the pioneer infant microbiome. *Microbiome* 5, 48. doi: 10.1186/s40168-017-0268-4

Perry, R. J., Peng, L., Barry, N. A., Cline, G. W., Zhang, D., Cardone, R. L., et al. (2016). Acetate mediates a microbiome-brain-beta-cell axis to promote metabolic syndrome. *Nature* 534, 213–217. doi: 10.1038/nature18309

Pesce, M., Seguella, L., Del Re, A., Lu, J., Palenca, I., Corpetti, C., et al. (2022). Nextgeneration probiotics for inflammatory bowel disease. *Int. J. Mol. Sci.* 23. doi: 10.3390/ ijms23105466

Pfeiffer, R. F. (1998). Gastrointestinal dysfunction in Parkinson's disease. *Clin. Neurosci.* 5, 136-146.

Phan, J., Nair, D., Jain, S., Montagne, T., Flores, D. V., Nguyen, A., et al. (2021). Alterations in gut microbiome composition and function in irritable bowel syndrome and increased probiotic abundance with daily supplementation. *mSystems* 6, e0121521. doi: 10.1128/mSystems.01215-21

Qiao, Y., Sun, J., Xia, S., Tang, X., Shi, Y., and Le, G. (2014). Effects of resveratrol on gut microbiota and fat storage in a mouse model with high-fat-induced obesity. *Food Funct.* 5, 1241–1249. doi: 10.1039/c3fo60630a

Quigley, E. M. M. (2017). Microbiota-brain-gut axis and neurodegenerative diseases. Curr. Neurol. Neurosci. Rep. 17, 94. doi: 10.1007/s11910-017-0802-6

Rajilic-Stojanovic, M., Biagi, E., Heilig, H. G., Kajander, K., Kekkonen, R. A., Tims, S., et al. (2011). Global and deep molecular analysis of microbiota signatures in fecal samples from patients with irritable bowel syndrome. *Gastroenterology* 141, 1792–1801. doi: 10.1053/j.gastro.2011.07.043

Ramezani, A., Massy, Z. A., Meijers, B., Evenepoel, P., Vanholder, R., and Raj, D. S. (2016). Role of the gut microbiome in uremia: A potential therapeutic target. *Am. J. Kidney Dis.* 67, 483–498. doi: 10.1053/j.ajkd.2015.09.027

Ranganathan, N., Patel, B. G., Ranganathan, P., Marczely, J., Dheer, R., Pechenyak, B., et al. (2006). *In vitro* and in *vivo* assessment of intraintestinal bacteriotherapy in chronic kidney disease. *ASAIO J.* 52, 70–79. doi: 10.1097/01.mat.0000191345.45735.00

Ranganathan, N., Ranganathan, P., Friedman, E. A., Joseph, A., Delano, B., Goldfarb, D. S., et al. (2010). Pilot study of probiotic dietary supplementation for promoting healthy kidney function in patients with chronic kidney disease. *Adv. Ther.* 27, 634–647. doi: 10.1007/s12325-010-0059-9

Rao, J., Xie, R., Lin, L., Jiang, J., Du, L., Zeng, X., et al. (2021). Fecal microbiota transplantation ameliorates gut microbiota imbalance and intestinal barrier damage in rats with stress-induced depressive-like behavior. *Eur. J. Neurosci.* 53, 3598–3611. doi: 10.1111/ejn.15192

Rasmussen, H. S., Holtug, K., and Mortensen, P. B. (1988). Degradation of amino acids to short-chain fatty acids in humans. An in *vitro* study. *Scand. J. Gastroenterol.* 23, 178–182. doi: 10.3109/00365528809103964

Rastall, R. A., and Gibson, G. R. (2015). Recent developments in prebiotics to selectively impact beneficial microbes and promote intestinal health. *Curr. Opin. Biotechnol.* 32, 42–46. doi: 10.1016/j.copbio.2014.11.002

Riedel, S., Pheiffer, C., Johnson, R., Louw, J., and Muller, C. J. F. (2021). Intestinal barrier function and immune homeostasis are missing links in obesity and type 2 diabetes development. *Front. Endocrinol. (Lausanne)* 12, 833544. doi: 10.3389/fendo.2021.833544

Saha, L. (2014). Irritable bowel syndrome: pathogenesis, diagnosis, treatment, and evidence-based medicine. *World J. Gastroenterol.* 20, 6759–6773. doi: 10.3748/wjg.v20.i22.6759

Saiyasit, N., Chunchai, T., Prus, D., Suparan, K., Pittayapong, P., Apaijai, N., et al. (2020). Gut dysbiosis develops before metabolic disturbance and cognitive decline in high-fat diet-induced obese condition. *Nutrition* 69, 110576. doi: 10.1016/j.nut.2019.110576

Saji, N., Murotani, K., Hisada, T., Tsuduki, T., Sugimoto, T., Kimura, A., et al. (2019). The relationship between the gut microbiome and mild cognitive impairment in

patients without dementia: a cross-sectional study conducted in Japan. Sci. Rep. 9, 19227. doi: 10.1038/s41598-019-55851-y

Salvi, P. S., and Cowles, R. A. (2021). Butyrate and the Intestinal Epithelium: Modulation of Proliferation and Inflammation in Homeostasis and Disease. *Cells* 10 (7), 1775. doi: 10.3390/cells10071775

Sampson, T. R., Debelius, J. W., Thron, T., Janssen, S., Shastri, G. G., Ilhan, Z. E., et al. (2016). Gut microbiota regulate motor deficits and neuroinflammation in a model of parkinson's disease. *Cell* 167 1469-1480, e1412. doi: 10.1016/j.cell.2016.11.018

Sanchez-Alcoholado, L., Castellano-Castillo, D., Jordan-Martinez, L., Moreno-Indias, I., Cardila-Cruz, P., Elena, D., et al. (20171936). Role of gut microbiota on cardio-metabolic parameters and immunity in coronary artery disease patients with and without type-2 diabetes mellitus. *Front. Microbiol.* 8. doi: 10.3389/ fmicb.2017.01936

Sanchez-Alcoholado, L., Ramos-Molina, B., Otero, A., Laborda-Illanes, A., Ordonez, R., Medina, J. A., et al. (2020). The role of the gut microbiome in colorectal cancer development and therapy response. *Cancers (Basel)* 12. doi: 10.3390/cancers12061406

Sanekommu, H., Taj, S., Mah Noor, R., Umair Akmal, M., Akhtar, R., Hossain, M., et al. (2023). Probiotics and fecal transplant: an intervention in delaying chronic kidney disease progression? *Clin. Pract.* 13, 881–888. doi: 10.3390/clinpract13040080

Sasso, J. M., Ammar, R. M., Tenchov, R., Lemmel, S., Kelber, O., Grieswelle, M., et al. (2023). Gut microbiome-brain alliance: A landscape view into mental and gastrointestinal health and disorders. ACS Chem. Neurosci. 14, 1717–1763. doi: 10.1021/acschemneuro.3c00127

Schellekens, H., Torres-Fuentes, C., Van De Wouw, M., Long-Smith, C. M., Mitchell, A., Strain, C., et al. (2021). Bifdobacterium longum counters the effects of obesity: Partial successful translation from rodent to human. *EBioMedicine* 63, 103176. doi: 10.1016/j.ebiom.2020.103176

Schwartz, D. L. (2016). Quercetin as an augmentation agent in schizophrenia. J. Clin. Psychopharmacol. 36, 282–283. doi: 10.1097/JCP.000000000000498

Sears, C. L. (2009). Enterotoxigenic Bacteroides fragilis: a rogue among symbiotes. *Clin. Microbiol. Rev.* 22, 349–369. doi: 10.1128/CMR.00053-08

Segal, A., Zlotnik, Y., Moyal-Atias, K., Abuhasira, R., and Ifergane, G. (2021). Fecal microbiota transplant as a potential treatment for Parkinson's disease - A case series. *Clin. Neurol. Neurosurg.* 207, 106791. doi: 10.1016/j.clineuro.2021.106791

Seksik, P., Rigottier-Gois, L., Gramet, G., Sutren, M., Pochart, P., Marteau, P., et al. (2003). Alterations of the dominant faecal bacterial groups in patients with Crohn's disease of the colon. *Gut* 52, 237–242. doi: 10.1136/gut.52.2.237

Sgritta, M., Dooling, S. W., Buffington, S. A., Momin, E. N., Francis, M. B., Britton, R. A., et al. (2019). Mechanisms underlying microbial-mediated changes in social behavior in mouse models of autism spectrum disorder. *Neuron* 101 246-259, e246. doi: 10.1016/j.neuron.2018.11.018

Shaaban, S. Y., El Gendy, Y. G., Mehanna, N. S., El-Senousy, W. M., El-Feki, H. S. A., Saad, K., et al. (2018). The role of probiotics in children with autism spectrum disorder: A prospective, open-label study. *Nutr. Neurosci.* 21, 676–681. doi: 10.1080/1028415X.2017.1347746

Shaidullov, I. F., Sorokina, D. M., Sitdikov, F. G., Hermann, A., Abdulkhakov, S. R., and Sitdikova, G. F. (2021). Short chain fatty acids and colon motility in a mouse model of irritable bowel syndrome. *BMC Gastroenterol.* 21, 37. doi: 10.1186/s12876-021-01613-y

Shao, Y., Forster, S. C., Tsaliki, E., Vervier, K., Strang, A., Simpson, N., et al. (2019). Stunted microbiota and opportunistic pathogen colonization in caesarean-section birth. *Nature* 574, 117–121. doi: 10.1038/s41586-019-1560-1

Sharma, S., and Tripathi, P. (2019). Gut microbiome and type 2 diabetes: where we are and where to go? *J. Nutr. Biochem.* 63, 101–108. doi: 10.1016/j.jnutbio.2018.10.003

Shemin, D., Bostom, A. G., Laliberty, P., and Dworkin, L. D. (2001). Residual renal function and mortality risk in hemodialysis patients. *Am. J. Kidney Dis.* 38, 85–90. doi: 10.1053/ajkd.2001.25198

Shin, S., and Cho, K. Y. (2020). Altered gut microbiota and shift in bacteroidetes between young obese and normal-weight korean children: A cross-sectional observational study. *BioMed. Res. Int.* 2020, 6587136. doi: 10.1155/2020/6587136

Shin, D. S., Rhee, K. J., and Eom, Y. B. (2020). Effect of Probiotic Clostridium butyricum NCTC 7423 Supernatant on Biofilm Formation and Gene Expression of Bacteroides fragilis. *J. Microbiol. Biotechnol.* 30, 368–377. doi: 10.4014/jmb.2001.01027

Shirouchi, B., Nagao, K., Umegatani, M., Shiraishi, A., Morita, Y., Kai, S., et al. (2016). Probiotic Lactobacillus gasseri SBT2055 improves glucose tolerance and reduces body weight gain in rats by stimulating energy expenditure. *Br. J. Nutr.* 116, 451–458. doi: 10.1017/S0007114516002245

Shulman, R. J., Hollister, E. B., Cain, K., Czyzewski, D. I., Self, M. M., Weidler, E. M., et al. (2017). Psyllium fiber reduces abdominal pain in children with irritable bowel syndrome in a randomized, double-blind trial. *Clin. Gastroenterol. Hepatol.* 15 712-719, e714. doi: 10.1016/j.cgh.2016.03.045

Silk, D. B., Davis, A., Vulevic, J., Tzortzis, G., and Gibson, G. R. (2009). Clinical trial: the effects of a trans-galactooligosaccharide prebiotic on faecal microbiota and symptoms in irritable bowel syndrome. *Aliment Pharmacol. Ther.* 29, 508–518. doi: 10.1111/j.1365-2036.2008.03911.x

Sobhani, I., Bergsten, E., Couffin, S., Amiot, A., Nebbad, B., Barau, C., et al. (2019). Colorectal cancer-associated microbiota contributes to oncogenic epigenetic signatures. *Proc. Natl. Acad. Sci. U.S.A.* 116, 24285–24295. doi: 10.1073/pnas.1912129116 Sohrabi, M., Sahu, B., Kaur, H., Hasler, W. A., Prakash, A., and Combs, C. K. (2022). Gastrointestinal changes and alzheimer's disease. *Curr. Alzheimer Res.* 19, 335–350. doi: 10.2174/1567205019666220617121255

Soto, C., and Satani, N. (2011). The intricate mechanisms of neurodegeneration in prion diseases. *Trends Mol. Med.* 17, 14–24. doi: 10.1016/j.molmed.2010.09.001

Steenbergen, L., Sellaro, R., Van Hemert, S., Bosch, J. A., and Colzato, L. S. (2015). A randomized controlled trial to test the effect of multispecies probiotics on cognitive reactivity to sad mood. *Brain Behav. Immun.* 48, 258–264. doi: 10.1016/ j.bbi.2015.04.003

Strati, F., Cavalieri, D., Albanese, D., De Felice, C., Donati, C., Hayek, J., et al. (2017). New evidences on the altered gut microbiota in autism spectrum disorders. *Microbiome* 5, 24. doi: 10.1186/s40168-017-0242-1

Stroup, T. S., and Gray, N. (2018). Management of common adverse effects of antipsychotic medications. *World Psychiatry* 17, 341–356. doi: 10.1002/wps.20567

Suganya, K., and Koo, B. S. (2020). Gut-brain axis: role of gut microbiota on neurological disorders and how probiotics/prebiotics beneficially modulate microbial and immune pathways to improve brain functions. *Int. J. Mol. Sci.* 21. doi: 10.3390/ ijms21207551

Sun, Z., Song, Z. G., Liu, C., Tan, S., Lin, S., Zhu, J., et al. (2022b). Gut microbiome alterations and gut barrier dysfunction are associated with host immune homeostasis in COVID-19 patients. *BMC Med.* 20, 24. doi: 10.1186/s12916-021-02212-0

Sun, J., Wang, F., Hu, X., Yang, C., Xu, H., Yao, Y., et al. (2018). Clostridium butyricum Attenuates Chronic Unpredictable Mild Stress-Induced Depressive-Like Behavior in Mice via the Gut-Brain Axis. *J. Agric. Food Chem.* 66, 8415–8421. doi: 10.1021/acs.jafc.8b02462

Sun, H., Zhao, F., Liu, Y., Ma, T., Jin, H., Quan, K., et al. (2022a). Probiotics synergized with conventional regimen in managing Parkinson's disease. *NPJ Parkinsons Dis.* 8, 62. doi: 10.1038/s41531-022-00327-6

Surdea-Blaga, T., Baban, A., and Dumitrascu, D. L. (2012). Psychosocial determinants of irritable bowel syndrome. *World J. Gastroenterol.* 18, 616–626. doi: 10.3748/wjg.v18.i7.616

Takayama, F., Taki, K., and Niwa, T. (2003). Bifidobacterium in gastro-resistant seamless capsule reduces serum levels of indoxyl sulfate in patients on hemodialysis. *Am. J. Kidney Dis.* 41, S142–S145. doi: 10.1053/ajkd.2003.50104

Taki, K., Takayama, F., and Niwa, T. (2005). Beneficial effects of Bifidobacteria in a gastroresistant seamless capsule on hyperhomocysteinemia in hemodialysis patients. *J. Ren Nutr.* 15, 77–80. doi: 10.1053/j.jrn.2004.09.028

Tamburini, S., Shen, N., Wu, H. C., and Clemente, J. C. (2016). The microbiome in early life: implications for health outcomes. *Nat. Med.* 22 (7), 713–722. doi: 10.1038/nm.4142

Taniya, M. A., Chung, H. J., Al Mamun, A., Alam, S., Aziz, M. A., Emon, N. U., et al. (2022). Role of gut microbiome in autism spectrum disorder and its therapeutic regulation. *Front. Cell Infect. Microbiol.* 12, 915701. doi: 10.3389/fcimb.2022.915701

Thirion, F., Speyer, H., Hansen, T. H., Nielsen, T., Fan, Y., Le Chatelier, E., et al. (2023). Alteration of gut microbiome in patients with schizophrenia indicates links between bacterial tyrosine biosynthesis and cognitive dysfunction. *Biol. Psychiatry Glob Open Sci.* 3, 283–291. doi: 10.1016/j.bpsgos.2022.01.009

Thomas, S., Izard, J., Walsh, E., Batich, K., Chongsathidkiet, P., Clarke, G., et al. (2017). The host microbiome regulates and maintains human health: A primer and perspective for non-microbiologists. *Cancer Res.* 77, 1783–1812. doi: 10.1158/0008-5472.CAN-16-2929

Thompson-Chagoyan, O. C., Maldonado, J., and Gil, A. (2005). Aetiology of inflammatory bowel disease (IBD): role of intestinal microbiota and gut-associated lymphoid tissue immune response. *Clin. Nutr.* 24, 339–352. doi: 10.1016/j.clnu.2005.02.009

Tian, P., Chen, Y., Zhu, H., Wang, L., Qian, X., Zou, R., et al. (2022). Bifidobacterium breve CCFM1025 attenuates major depression disorder via regulating gut microbiome and tryptophan metabolism: A randomized clinical trial. *Brain Behav. Immun.* 100, 233–241. doi: 10.1016/j.bbi.2021.11.023

Troseid, M., Andersen, G. O., Broch, K., and Hov, J. R. (2020). The gut microbiome in coronary artery disease and heart failure: Current knowledge and future directions. *EBioMedicine* 52, 102649. doi: 10.1016/j.ebiom.2020.102649

Ullah, H., Arbab, S., Tian, Y., Liu, C.-Q., Chen, Y., Qijie, L., et al. (2023). The gut microbiota-brain axis in neurological disorder. *Frontiers in Neuroscience* 17. doi: 10.3389/fnins.2023.1225875

Van De Merwe, J. P., Schroder, A. M., Wensinck, F., and Hazenberg, M. P. (1988). The obligate anaerobic faecal flora of patients with Crohn's disease and their first-degree relatives. *Scand. J. Gastroenterol.* 23, 1125–1131. doi: 10.3109/00365528809090179

Van Tilburg, M. A., Palsson, O. S., and Whitehead, W. E. (2013). Which psychological factors exacerbate irritable bowel syndrome? Development of a comprehensive model. *J. Psychosom Res.* 74, 486–492. doi: 10.1016/j.jpsychores.2013.03.004

Venkataraman, R., Madempudi, R. S., Neelamraju, J., Ahire, J. J., Vinay, H. R., Lal, A., et al. (2021). Effect of multi-strain probiotic formulation on students facing examination stress: a double-blind, placebo-controlled study. *Probiotics Antimicrob. Proteins* 13, 12–18. doi: 10.1007/s12602-020-09681-4

Verdam, F. J., Fuentes, S., De Jonge, C., Zoetendal, E. G., Erbil, R., Greve, J. W., et al. (2013). Human intestinal microbiota composition is associated with local and systemic inflammation in obesity. *Obes. (Silver Spring)* 21, E607–E615. doi: 10.1002/oby.20466

Vinelli, V., Biscotti, P., Martini, D., Del Bo, C., Marino, M., Merono, T., et al. (2022). Effects of dietary fibers on short-chain fatty acids and gut microbiota composition in healthy adults: A systematic review. *Nutrients* 14. doi: 10.3390/nu14132559

Vinolo, M. A., Rodrigues, H. G., Nachbar, R. T., and Curi, R. (2011). Regulation of inflammation by short chain fatty acids. *Nutrients* 3, 858–876. doi: 10.3390/nu3100858

Vitetta, L., Llewellyn, H., and Oldfield, D. (2019). Gut dysbiosis and the intestinal microbiome: streptococcus thermophilus a key probiotic for reducing uremia. *Microorganisms* 7. doi: 10.3390/microorganisms7080228

Vogt, N. M., Kerby, R. L., Dill-Mcfarland, K. A., Harding, S. J., Merluzzi, A. P., Johnson, S. C., et al. (2017). Gut microbiome alterations in Alzheimer's disease. *Sci. Rep.* 7, 13537. doi: 10.1038/s41598-017-13601-y

Voigt, A. Y., Costea, P. I., Kultima, J. R., Li, S. S., Zeller, G., Sunagawa, S., et al. (2015). Temporal and technical variability of human gut metagenomes. *Genome Biol.* 16, 73. doi: 10.1186/s13059-015-0639-8

Walker, L., Levine, H., and Jucker, M. (2006). Koch's postulates and infectious proteins. Acta Neuropathol. 112, 1–4. doi: 10.1007/s00401-006-0072-x

Wang, R. X., Lee, J. S., Campbell, E. L., and Colgan, S. P. Microbiota-derived butyrate dynamically regulates intestinal homeostasis through regulation of actin-associated protein synaptopodin. *Proc. Natl. Acad. Sci. U. S. A.* (2020) 117 (21), 11648–11657. doi: 10.1073/pnas.1917597117

Wang, J., Chen, J., Li, L., Zhang, H., Pang, D., Ouyang, H., et al. (2022). Clostridium butyricum and Bifidobacterium pseudolongum Attenuate the Development of Cardiac Fibrosis in Mice. *Microbiol. Spectr.* 10, e0252422. doi: 10.1128/spectrum.02524-22

Xiao, L. W., and Zhao, F. Q. (2023). Microbial transmission, colonisation and succession: from pregnancy to infancy. *Gut* 72, 772–786. doi: 10.1136/gutjnl-2022-328970

Xie, X., He, Y., Li, H., Yu, D., Na, L., Sun, T., et al. (2019). Effects of prebiotics on immunologic indicators and intestinal microbiota structure in perioperative colorectal cancer patients. *Nutrition* 61, 132–142. doi: 10.1016/j.nut.2018.10.038

Xu, Z., Liu, Z., Dong, X., Hu, T., Wang, L., Li, J., et al. (2018). Fecal microbiota transplantation from healthy donors reduced alcohol-induced anxiety and depression in an animal model of chronic alcohol exposure. *Chin. J. Physiol.* 61, 360–371. doi: 10.4077/CJP.2018.BAH633

Xu, H., Wang, J., Cai, J., Feng, W., Wang, Y., Liu, Q., et al. (2019). Protective Effect of Lactobacillus rhamnosus GG and its Supernatant against Myocardial Dysfunction in Obese Mice Exposed to Intermittent Hypoxia is Associated with the Activation of Nrf2 Pathway. Int. J. Biol. Sci. 15, 2471–2483. doi: 10.7150/ijbs.36465

Xu, M., Wang, C., Krolick, K. N., Shi, H., and Zhu, J. (2020). Difference in post-stress recovery of the gut microbiome and its altered metabolism after chronic adolescent stress in rats. *Sci. Rep.* 10, 3950. doi: 10.1038/s41598-020-60862-1

Xue, L. J., Yang, X. Z., Tong, Q., Shen, P., Ma, S. J., Wu, S. N., et al. (2020). Fecal microbiota transplantation therapy for Parkinson's disease: A preliminary study. *Med.* (*Baltimore*) 99, e22035. doi: 10.1097/MD.00000000022035

Yan, A., Culp, E., Perry, J., Lau, J. T., MacNeil, L. T., Surette, M. G., et al. (2018). Transformation of the Anticancer Drug Doxorubicin in the Human Gut Microbiome. *ACS Infect. Dis.* 4 (1), 68–76. doi: 10.1021/acsinfecdis.7b00166

Yeoh, Y. K., Zuo, T., Lui, G. C., Zhang, F., Liu, Q., Li, A. Y., et al. (2021). Gut microbiota composition reflects disease severity and dysfunctional immune responses in patients with COVID-19. *Gut* 70, 698–706. doi: 10.1136/gutjnl-2020-323020

You, S., Ma, Y., Yan, B., Pei, W., Wu, Q., Ding, C., et al. (2022). The promotion mechanism of prebiotics for probiotics: A review. *Front. Nutr.* 9, 1000517. doi: 10.3389/ fnut.2022.1000517

Yu, F., Feng, X., Li, X., Luo, Y., Wei, M., Zhao, T., et al. (2021). Gut-derived metabolite phenylacetylglutamine and white matter hyperintensities in patients with acute ischemic stroke. *Front. Aging Neurosci.* 13, 675158. doi: 10.3389/fnagi.2021.675158

Yuan, S., Wang, K. S., Meng, H., Hou, X. T., Xue, J. C., Liu, B. H., et al. (2023). The gut microbes in inflammatory bowel disease: Future novel target option for pharmacotherapy. *BioMed. Pharmacother.* 165, 114893. doi: 10.1016/j.biopha.2023.114893

Zaharuddin, L., Mokhtar, N. M., Muhammad Nawawi, K. N., and Raja Ali, R. A. (2019). A randomized double-blind placebo-controlled trial of probiotics in postsurgical colorectal cancer. *BMC Gastroenterol.* 19, 131. doi: 10.1186/s12876-019-1047-4

Zhai, C., Ahn, J. S., Islam, M. M., Lkhagva, E., Chung, H. J., and Hong, S. T. (2022a). Comparative analysis of original and replaced gut microbiomes within same individuals identified the intestinal microbes associated with weight gaining. *Microorganisms* 10. doi: 10.3390/microorganisms10051062

Zhai, T., Wang, P., Hu, X., and Zheng, L. (2022b). Probiotics bring new hope for atherosclerosis prevention and treatment. *Oxid. Med. Cell Longev* 2022, 3900835. doi: 10.1155/2022/3900835

Zhang, J., Ni, Y., Qian, L., Fang, Q., Zheng, T., Zhang, M., et al. (2021). Decreased abundance of akkermansia muciniphila leads to the impairment of insulin secretion and glucose homeostasis in lean type 2 diabetes. *Adv. Sci. (Weinh)* 8, e2100536. doi: 10.1002/advs.202100536

Zhang, Y., Si, X., Yang, L., Wang, H., Sun, Y., and Liu, N. (2022). Association between intestinal microbiota and inflammatory bowel disease. *Anim. Model. Exp. Med.* 5, 311–322. doi: 10.1002/ame2.12255

Zhang, J. L., Wang, S. B., Zeng, Z., Qin, Y. X., Shen, Q., and Li, P. L. (2020). Antidiabetic effects of 01 through improving hepatic insulin sensitivity in type 2 diabetic rat model. *J. Funct. Foods* 67. doi: 10.1016/j.jff.2020.103843

Zhang, W., Xu, J. H., Yu, T., and Chen, Q. K. (2019). Effects of berberine and metformin on intestinal inflammation and gut microbiome composition in db/db mice. *BioMed. Pharmacother.* 118, 109131. doi: 10.1016/j.biopha.2019.109131

Zhen, J., Zhou, Z., He, M., Han, H. X., Lv, E. H., Wen, P. B., et al. (2023). The gut microbial metabolite trimethylamine N-oxide and cardiovascular diseases. *Front. Endocrinol. (Lausanne)* 14, 1085041. doi: 10.3389/fendo.2023.1085041

Zheng, P., Zeng, B., Liu, M., Chen, J., Pan, J., Han, Y., et al. (2019). The gut microbiome from patients with schizophrenia modulates the glutamate-glutamine-GABA cycle and schizophrenia-relevant behaviors in mice. *Sci. Adv.* 5 (2), eaau831710.1126/sciadv.aau8317.

Zhong, H., Wang, J., Abdullah, Hafeez, M. A., Guan, R., and Feng, F. (2021a). Lactobacillus plantarum ZJUFB2 prevents high fat diet-induced insulin resistance in association with modulation of the gut microbiota. *Front. Nutr.* 8, 754222. doi: 10.3389/ fnut.2021.754222

Zhong, H., Wang, Y., Shi, Z., Zhang, L., Ren, H., He, W., et al. (2021b). Characterization of respiratory microbial dysbiosis in hospitalized COVID-19 patients. *Cell Discovery* 7, 23. doi: 10.1038/s41421-021-00257-2

Zhou, X., Shang, G. S., Tan, Q., He, Q., Tan, X., Park, K. Y., et al. (2021). Effect of Lactobacillus fermentum TKSN041 on improving streptozotocin-induced type 2 diabetes in rats. *Food Funct.* 12, 7938–7953. doi: 10.1039/D1FO01571K

Zhu, Y., Li, Q., and Jiang, H. (2020). Gut microbiota in atherosclerosis: focus on trimethylamine N-oxide. *APMIS* 128, 353–366. doi: 10.1111/apm.13038

Zimmermann, P. (2023). The immunological interplay between vaccination and the intestinal microbiota. *NPJ Vaccines* 8, 24. doi: 10.1038/s41541-023-00627-9

Zuo, T., and Ng, S. C. (20182247). The gut microbiota in the pathogenesis and therapeutics of inflammatory bowel disease. *Front. Microbiol.* 9. doi: 10.3389/fmicb.2018.02247

Zuo, T., Zhang, F., Lui, G. C. Y., Yeoh, Y. K., Li, A. Y. L., Zhan, H., et al. (2020). Alterations in gut microbiota of patients with COVID-19 during time of hospitalization. *Gastroenterology* 159 944-955, e948.x. doi: 10.1053/ j.gastro.2020.05.048