



# The Airway Pathobiome in Complex Respiratory Diseases: A Perspective in Domestic Animals

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Respiratory infections in domestic animals are a major issue for veterinary and livestock industry. Pathogens in the respiratory tract share their habitat with a myriad of commensal microorganisms. Increasing evidence points towards a respiratory pathobiome concept, integrating the dysbiotic bacterial communities, the host and the environment in a new understanding of respiratory disease etiology. During the infection, the airway microbiota likely regulates and is regulated by pathogens through diverse mechanisms, thereby acting either as a gatekeeper that provides resistance to pathogen colonization or enhancing their prevalence and bacterial co-infectivity, which often results in disease exacerbation. Insight into the complex interplay taking place in the respiratory tract between the pathogens, microbiota, the host and its environment during infection in domestic animals is a research field in its infancy in which most studies are focused on infections from enteric pathogens and gut microbiota. However, its understanding may improve pathogen control and reduce the severity of microbial-related diseases, including those with zoonotic potential.

**Keywords:** domestic animals, pathobiome, respiratory infectious diseases, airway microbiota, public health

## INTRODUCTION

Complex respiratory diseases are highly prevalent and can be life threatening in domestic animals in which a prompt diagnosis and targeted treatments are essential (Ericsson et al., 2016; Bond et al., 2017; Oladunni et al., 2019; Ericsson et al., 2020). Besides impacting animal health and welfare, they cause a significant health burden worldwide including high treatment costs, high morbidity, premature mortality, decreased performance and severe consequences to public health and the environment (Kuiken et al., 2005; Holt et al., 2011; Ericsson et al., 2016; Bond et al., 2017; Oladunni et al., 2019; Blakebrough-Hall et al., 2020; Ericsson et al., 2020). For example, the bovine respiratory disease complex (BRDC) is a leading cause of morbidity and economic losses in wealthy countries which ranges from 30% in Belgium (van Leenen et al., 2020) to 49% in Switzerland and up to 80% in the U.S.A. (Hilton, 2014). Similarly, the porcine respiratory disease complex (PRDC) in finishing pigs continues to grow (Qin et al., 2018) with a morbidity rate ranging from 10% in Denmark (Hansen et al., 2010) to 40% in the U.S.A (Harms et al., 2002). As for common livestock animals, the equine respiratory disease complex (ERDC) is an important respiratory infection in horses

(Wasko et al., 2011) that affects up to 66% of the equine population (Wasko et al., 2011) and can quickly disqualify a horse from racing, showing, training or other activities for periods of time ranging from a few days to months. In companion animals, the canine infectious respiratory disease complex (CIRDC) affects 66% of the dogs studied in Europe, including both peat and kenneled dogs (Mitchell et al., 2017). The feline respiratory disease complex has been described as one of the most importance cause of morbidity for cats in U.S.A, with reported incidence as high as 30% (Wagner et al., 2018). Finally, the respiratory disease complex in poultry remains widespread and has become endemic in different countries causing subclinical infections, mild respiratory symptoms and high production losses in birds either raised for meat or eggs (Awad et al., 2014; Guabiraba and Schouler, 2015; Patel et al., 2018; Samy and Naguib, 2018).

Our view on the dynamics of airway diseases has now been broadened to include an additional aspect of the complex system: the microbiota of the host and its environment (Bernardo-Cravo et al., 2020). The properties of the host microbiome have led to the concept of the holobiome in which the host and its microbial communities are merged into a symbiotic superorganism and later, to the concept of pathobiome to further consider microbiome communities in disease state (Vayssier-Taussat et al., 2014). The importance of the pathobiome concept arose from human studies in which disruption of a health-promoting and stable gut microbiome results in dysbiosis— a microbiome community acting as pathogenic entity (Bass et al., 2019). In this concept, the pathogen and host microbiome are assembled, leading to similar issues raised for the holobiome (Bernardo-Cravo et al., 2020).

Much work is required to identify the mechanisms underlying the microorganism relations and perturbations of a balanced and healthy microbiome that lead to a pathobiome (Bass et al., 2019). Commensal microbiome might act as a gatekeeper that provides resistance to infection on the mucosal surface and spreading to the lungs (Man et al., 2017; Li et al., 2019) or enhance disease exacerbation.

The number of studies addressing the role of the respiratory bacterial communities in animal health and disease is limited and almost entirely restricted to animals farmed for foods such as cattle and pigs. Moreover, although microbiota is composed of bacteria, protists, fungi, archaea and viruses, most studies in domestic animals have focused solely on bacterial microbiota.

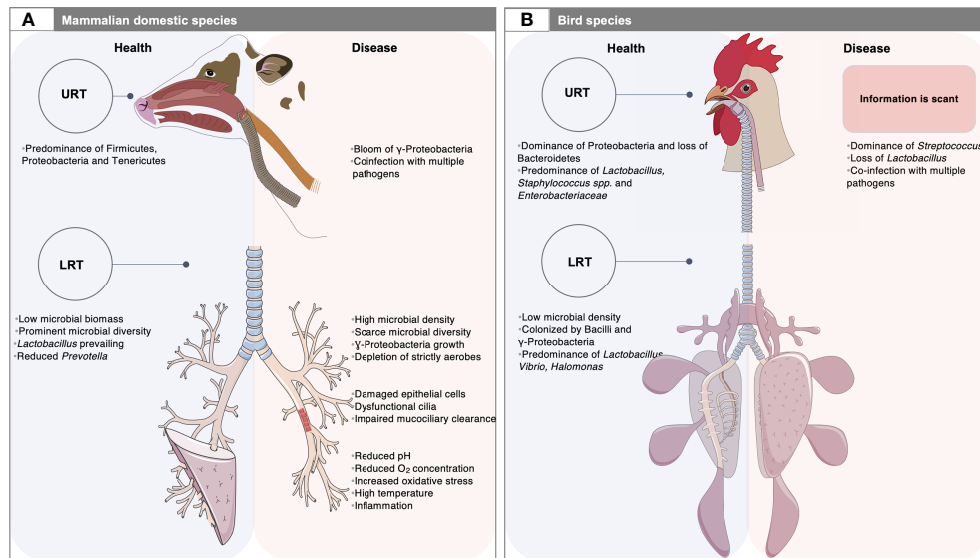
In cows and feedlot cattle, several studies have reported how a healthy respiratory microbiota is established in the airways and surveyed which host and environmental factors drive it (Timsit et al., 2016; Hall et al., 2017; Holman et al., 2017; Nicola et al., 2017; Zeineldin et al., 2017a; Holman et al., 2018; Stroebel et al., 2018; Amat et al., 2019; McMullen et al., 2020; Zeineldin et al., 2020a). Analogously, predicting when and how respiratory microbiome breaks down is at the heart of several studies in swine (Cortes et al., 2018; Zeineldin et al., 2018; Jakobsen et al., 2019; Megahed et al., 2019; Mou et al., 2019; Wang et al., 2019; Pirolo et al., 2021). For instance, it appears as though that during weaning, several microorganisms act

synergistically to mediate the BRDC (Gaeta et al., 2017; Klima et al., 2019; McMullen et al., 2019; Zeineldin et al., 2019) and PRDC (Wang et al., 2018; Li et al., 2021). Aside from ruminants and swine, efforts have focused on characterizing the composition patterns of the upper and lower airways in sheep (Glendinning et al., 2016; Glendinning et al., 2017a), horses (Bond et al., 2017), animals domesticated for companionship (Ericsson et al., 2016; Vientoos-Plotts et al., 2017; Vientós-Plotts et al., 2017; Fastrès et al., 2019; Fastrès et al., 2020b) and commercial birds (Shabbir et al., 2014; Glendinning et al., 2017b; Johnson et al., 2018; Ngunjiri et al., 2019; Abundo et al., 2020; Taylor et al., 2020; Abundo et al., 2021; Kursa et al., 2021).

Determining the underlying causes of respiratory illness is complicated. Species of veterinary interest are subjected to different host variables, environments and pathogens, which could all play a role in disease, either alone or in concert. However, we show evidence supporting the existence of an interplay between respiratory pathogens, commensal microbiota, host and environment in the respiratory apparatus of domestic animals. We untangle whether the airway microbiota act as a gatekeeper that provides resistance to pathogen colonization, thus ensuring resiliency and health in domestic animals. Moreover, we provide an overview of the covariation between airway microbiota, host and environment factors within and between species and we suggest potential applications of this knowledge in veterinary medicine. Lastly, we outline avenues to understand if gut microbiota-derived metabolites could be key molecular mediators of the microbiota-gut-lung axis and affect the onset of the respiratory diseases. Thus, the exploration of the relationships between respiratory pathogens, hosts and respiratory microbiota in domestic animals is timely and novel.

## DO DIFFERENCES IN THE RESPIRATORY SYSTEM CONTRIBUTE TO MICROBIOME COMPOSITION? MAMMALS VERSUS BIRDS

It is remarkable that mammals and birds, the two great classes of vertebrates capable of sustained high oxygen consumption, present many distinct differences (morphologic, physiologic and mechanic) in the respiratory tract (West et al., 2007) (**Figure 1**). For example, in mammals, the lower respiratory tract (LRT) comprises the trachea, the primary bronchi and the lungs, whereas in birds it involves the syrinx, the air sacs distributed throughout the body, the bronchi, the bronchioles and lungs (Nochi et al., 2018; Abundo et al., 2020; Abundo et al., 2021). Unlike mammals, the avian lungs are essentially rigid and tubular without alveoli (Bernhard et al., 2004) and have a unidirectional airflow in which the lungs are ventilated *via* air sacs (Nochi et al., 2018). By contrast, the mammalian lungs have reciprocating ventilation with large terminal air spaces (alveoli) and reduced airflow in peripheral structures (Bernhard et al., 2004). Additionally, the blood-gas barrier of the avian lung is



**FIGURE 1** | Airway microbiota: a matter of animal species, physiology and diseased settings. **(A)** The respiratory tract of domestic mammals is mainly colonized by Firmicutes, Proteobacteria, Tenericutes and Bacteroidetes. At steady state (left, blue rectangle), the microbial biomass in the low respiratory tract (LRT) is low and likely depends on the balance between migration of bacteria from the upper respiratory tract (URT) and the mucociliary and immune clearance. During respiratory disease (right; red rectangle), the LRT bacterial propagation outpaces the capacity of the airways to clear the microorganisms, which often results in increased microbial density and dysbiosis. Dysfunctional cilia and enhanced mucus production may contribute to reduced clearance and trapping of microorganisms within airways. An outgrowth of  $\gamma$ -Proteobacteria and loss of Firmicutes is observed in mammals, together with a limited microbial diversity, and the exacerbation of respiratory inflammation and loss of mucosal barrier function. In this context, co-infections by other pathogens or pathobionts are frequent. This polymicrobial disease involves microorganisms that act synergistically, or in succession to mediate complex disease processes. Studying such questions is challenging due to the complexity of sample collection. The exploration of LRT microbiota is based on invasive sampling techniques such as bronchoalveolar lavage, bronchus mucosal scraping and tracheal aspirates. Swabs of the deep nasal cavity, nasopharynx, oropharynx, paranasal sinus and tonsil are usually taken to explore the URT site; **(B)** The URT of birds is qualitatively similar to that of mammals. However, the nasal and oral cavities communicate with each other through the choanal cleft region. Their URT is mainly colonized by Firmicutes and Proteobacteria. At the genus level, a healthy ecosystem mainly includes highly dominant taxa, namely *Lactobacillus*, *Staphylococcus* spp. and members of the *Enterobacteriaceae* family. Unlike mammals, the LRT comprises the syrinx and the air sacs distributed throughout the body. In a stable state, the microbial density in the lung is very low. The LRT is mainly colonized by Bacilli and  $\gamma$ -Proteobacteria. At the genus level, *Lactobacillus* is the dominant bacterial taxon, followed by *Vibrio* and *Halomonas*. Yet, information related to the microbiota composition under respiratory diseases is scant. The assessment of microbiota in the URT is based on choanal swabs, nasal cavity wash and upper and lower tracheal washes, whereas the description of the LRT microbiota depends on lower respiratory lavage. The picture of mammalian LRT was downloaded from smart Servier Medical Art <https://smart.servier.com> without changes. Servier Medical Art by Servier is licensed under a Creative Commons Attribution 3.0 License. Written permission for publication of the chicken figure and chicken respiratory system drawing has been taken.

approximately 56-67% thinner than that of a mammal of the same body mass while the respiratory surface area is approximately 15% greater (Maina et al., 1989), which altogether have the potential for more efficient gas exchange (West et al., 2007). As a result, the environment of the LRT is very different between mammal and birds (West et al., 2007). The upper respiratory tract (URT) of birds is qualitatively similar to that of mammals. It has a nasal cavity with communicating sinuses, a larynx supported by cartilaginous plates and the trachea, although the nasal and oral cavities communicate with each other through the choanal cleft region, which appears in birds to be related to the incomplete secondary palate (Figure 1).

These host factors that themselves change across birds and mammals likely play an important role in structuring the airway microbiomes across species. It is possible that some other aspects of adaptation to flight have a net effect on microbiome composition, diversity and function in birds. For instance, the large mass-specific gas uptake by the avian respiratory system (Brown et al., 1997), as well as the large trachea length, the

decreased body mass, the higher metabolic rate, the higher average body temperature (40-41°C) and the higher resting heart rate (~245 beats/min) (Brown et al., 1997) probably influence the airway microbiome. The reduction in genome size compared to all amniotes, coupled with a shrinkage of large numbers of genes involved in the immune function (Lovell et al., 2014) might also consistently impact the microbiome composition and function.

## HOST DETERMINANTS IN A MICROBE-DOMINATED WORLD: A MATTER OF SPECIES

The importance of host-related factors to the diversity of respiratory microbiomes is still poorly understood. However, we have observed that more closely related mammals tend to present a similar degree of taxa composition in their

microbiomes, with likely downstream effects on host immunity and metabolic potential (**Table 1**). This may be due to the breathing patterns in mammals and the fact that the anatomy and physiology of both the upper and lower respiratory tract do not differ greatly among mammal species (**Figure 1**). In healthy bovine individuals, the URT microbiota (based on nasal and nasopharyngeal swab samples) is dominated by Proteobacteria, a group of bacteria suggested to have high functional variability (Bradley and Pollard, 2017), followed by Tenericutes, Firmicutes and Bacteroidetes (Gaeta et al., 2017; McMullen et al., 2019). Proteobacteria, Firmicutes, and Bacteroidetes are the three most dominant phyla in the URT bacterial communities in piglets (Slifierz et al., 2015; Correa-Fiz et al., 2016; Wang et al., 2019). Precisely, the composition of the tonsillar and oropharyngeal microbiome of adult pigs resembled that of the nasal microbiota, with Proteobacteria and Firmicutes as the predominant phyla (Lowe et al., 2012; Wang et al., 2018). Similarly, the URT microbiome of healthy horses is mainly dominated by Proteobacteria, Actinobacteria, Firmicutes, and Bacteroidetes (Bond et al., 2017). The most common phyla colonizing the nasal cavity of healthy dogs are Proteobacteria, followed by Firmicutes and Bacteroidetes (Tress et al., 2017). Proteobacteria is also the most abundant phylum in the cat respiratory tract, reaching up to 60% and 62% in the oropharyngeal swabs and BAL, respectively (Vientoos-Plotts et al., 2017).

A noticeable and consistent overlap between URT and LRT profile of microbial communities was observed in mammals (**Table 1**). In ruminants, it is likely the result of regurgitating of feedstuffs during rumination and the formation of aerosols during eructation (Klima et al., 2019). Thus, the oral microbiota in cattle probably acts as secondary reservoirs for the lung (McMullen et al., 2020). No information is available on the relationship between the URT and LRT in pigs, and the existing results of LRT microbiota are divergent at the phyla level (Huang et al., 2019; Jiang et al., 2019; Li et al., 2021). In line with the observation in ruminants, studies in cats and dogs support micro-aspiration of pharyngeal secretions as a primary route of microbial colonization of the LRT (Vientos-Plotts et al., 2017; Vientos-Plotts et al., 2019; Fastrès et al., 2020b). In healthy horses, the similarity between nose and lung microbiomes is expected for biological reasons (Bond et al., 2017), such as the large minute ventilation (60 L/min at rest) of these obligate nasal breathers and the fact that they have complete separation of the nasopharynx and oropharynx owing to their long velum, except when swallowing (Fillion-Bertrand et al., 2019). The end result of this interplay is the microbiome of lungs closely resembles that of the oropharynx in the vast majority of healthy mammals. However, it is not clear to what extent the deep airways are stably colonized by specific taxa or whether the microorganisms are in a dynamic state of flux being permanently cleared and repopulated from the URT.

In poultry, Firmicutes, Proteobacteria, Bacteroides, Actinobacteria and Tenericutes are the main well-documented phyla in the upper respiratory (Sohail et al., 2015; Glendinning et al., 2017b; Ngunjiri et al., 2019; Abundo et al., 2021; Kursa et al., 2021) (**Table 1**). In a stable state, the URT is colonized by Bacilli and Clostridia, whereas the LRT, including air sacs, is populated by Bacilli and  $\gamma$ -Proteobacteria (Abundo et al., 2021). However, birds

seem to have lost an association with Bacteroidetes but retained an association with facultative anaerobes species from the Proteobacteria phylum (Taylor et al., 2020). The loss of Bacteroidetes likely increases the proportion of transient environmental microorganisms and decreases the observed degree of host specificity and resilience (Taylor et al., 2020). Indeed, this could also help explain the dominance of Proteobacteria, which make up a large proportion of the airborne microbiome in birds (Cáliz et al., 2018), a source to which flighted animals are constantly exposed. At finer grains, birds present fewer obligate anaerobes and more facultative anaerobes compared to mammals. *Lactobacillus* spp. is the main dominant and stable colonizer of the upper respiratory tract (nasal cavity and choanal cleft), albeit varying relative abundances (Sohail et al., 2015; Glendinning et al., 2017b; Johnson et al., 2018; Abundo et al., 2020; Mulholland et al., 2021). *Lactobacilli* are followed by members of the family *Enterobacteriaceae* and of *Staphylococcus* spp. and *Escherichia-Shigella* (Glendinning et al., 2017b; Ngunjiri et al., 2019; Kursa et al., 2021). In the LRT, *Lactobacillus* are also the dominant bacterial taxa (Johnson et al., 2018; Abundo et al., 2021), outranking *Vibrio* and *Halomonas*, as determined by lower respiratory lavage in turkeys, chicken broilers and chicken layers raised in commercial settings (Abundo et al., 2021).

Collectively, these observations support the emerging idea that the nasal cavity in birds, which has the highest microbiota richness, serves as the mainland from which other respiratory sites are colonized (Abundo et al., 2020). The constant movement of fluid and air in the choana can play a major role in equilibrating the microbiome of URT with that of the gut or environment in birds and secondly, it can be responsible for the vast majority of the constant microbial seeding of the lower airways from both oral and nasal cavity. Bacterial overlap between the respiratory tract and gut has been observed in healthy layers raised in commercial farms (Ngunjiri et al., 2019) and turkeys (Taylor et al., 2020; Kursa et al., 2021), possibly through aerosolization of fecal bacteria and the contact with the litter environment.

Other host-related factors that might affect respiratory microbiota variations among mammals and birds involve mucus secretion, gas concentrations, osmolality, temperature, pH, nutrients such as iron and vitamins, surfactant secretion, blood flow, as well as extracellular DNA (Zhu et al., 2019). Additionally, the IgA secretion and the innate and adaptive immune recognition, including the antimicrobial peptides and the sensing of microbes, are endogenous forces that contribute to regulating the host-microbiota interactions in the respiratory system (Wypych et al., 2019).

## ENVIRONMENTAL STRESSORS AS DRIVERS OF THE RESPIRATORY MICROBIOTA COMPOSITION AND DYNAMICS ACROSS SPECIES

In mammalian hosts, the newborn animals are considered sterile *in utero* during a normal pregnancy, but during the first hours of

**TABLE 1 |** Recent metagenomic studies on the respiratory microbiota of domestic animals.

Specie	Cohort description	Sample type	Platform and sequenced region	Main phyla	Genera	$\alpha$ -diversity	Reference
Bovine	Healthy Holstein calves (n=32) vs calves with pneumonia (n=16), otitis (n=28) or both diseases (n=5). Time series: d3, d24, d28 and d35 of life. Age: 3 to 35 days of age	NPS	Illumina Miseq (V4)	Proteobacteria (30%-80%), Tenericutes (1%-10%), Firmicutes (1-20%), Bacteroidetes (1-10%)	<i>Mannheimia</i> (3-25%), <i>Mycoplasma</i> (3-18%), <i>Moraxella</i> (2-18%), <i>Psychrobacter</i> (3-14%), <i>Pseudomonas</i> (1-13%). Abundances of <i>Mannheimia</i> , <i>Moraxella</i> , and <i>Mycoplasma</i> were significantly higher in diseased vs healthy animals	$\alpha$ -diversity indices were not different between healthy and diseased calves	(Lima et al., 2016)
Bovine	Angus-beef cattle from weaning to 40 days after arrival at a feedlot (n=30). Time series: weaning, d0, d40	DNS	Illumina Miseq (V3)	Tenericutes (53.2%), Proteobacteria (34.7%), Firmicutes (4.2%), Bacteroidetes (3.7%), Actinobacteria (3.4%)	<i>Mycoplasma</i> ( <i>M. dispar</i> and <i>M. bovirhinis</i> , 52%), <i>Pasteurella</i> (11%), <i>Ralstonia</i> (10%), <i>Moraxella</i> (9%)	No differences in $\alpha$ -diversity over time	(Timsit et al., 2016)
Bovine	Healthy Holstein heifer calves at 14d (n=10) and 28d (n=19) vs BRDC at 14d (n=6) and 28d (n=6). Age: 3 to 35 days of age	NPS	Illumina Miseq (Not specified)	Proteobacteria, Firmicutes, Actinobacteria, Bacteroidetes, Tenericutes	Healthy: <i>Mannheimia haemolytica</i> (7%), <i>Psychrobacter</i> (2-5%), <i>Actinobacillus</i> (3-6%), <i>Mycoplasma</i> (2-12%), BRDC: <i>Pseudomonas fluorescens</i> (0.02 - 13%), <i>Mycoplasma</i> (8%), <i>Mannheimia haemolytica</i> (6%), <i>Eubacterium</i> (6%), <i>Psychrobacter</i> (5%). No significant differences were detected between groups	Not specified	(Gaeta et al., 2017)
Bovine	Healthy post-weaned Piedmontese calves (n=11) vs calves with respiratory diseases (n=8)	DNS, TA	Illumina Miseq (V4)	DNS: Proteobacteria (36.1%), Tenericutes (27.7%), Firmicutes (18.4%), Bacteroidetes (10.1%), Actinobacteria (6.3%) TA: Tenericutes (78%), Proteobacteria (11%), Fusobacteria (4%), Bacteroidetes (4%), Actinobacteria (3%)	DNS: <i>Mycoplasma</i> (27%), <i>Moraxella</i> (17%), <i>Aggregatibacter</i> (4%), <i>Sphingomonas</i> (3%), <i>Corynebacterium</i> (1.3%), <i>Psychrobacter</i> (1.2%). TA: <i>Mycoplasma</i> (77%), <i>Pasteurella multocida</i> (7%), <i>Mannheimia</i> (1.6%), <i>Bacteroides</i> (1.5%), <i>Ureaplasma</i> (1.3%). No differences between healthy and the BRDC-affected calves	$\alpha$ -diversity indices were different between TA and DNS samples	(Nicola et al., 2017)
Bovine	Angus weaned beef calves fed with Selenium-biofortified alfalfa hay (Se) for 9 weeks (n=30) vs control (n=15). Age: 6.5 to 9 months old	DNS	Illumina Miseq (V4)	Control group: Proteobacteria (44%), Bacteroidetes (24%), Firmicutes (19%); Se group: Bacteroidetes (29%), Proteobacteria (27%), Firmicutes (23%)	Control group: <i>Moraxellaceae</i> (19%), <i>Chitinophagaceae</i> (13%), <i>Aggregatibacter</i> (11%), <i>Weeksellaceae</i> (7%) Se group: <i>Chitinophagaceae</i> (23%), <i>Moraxellaceae</i> (9%) <i>Ruminococcaceae</i> (6%)	Se group tended to have an enriched nasal microbiota	(Hall et al., 2017)
Bovine	Angus x Hereford heifers transport to a feedlot (n=14). Time series: d0, d2, d7 and d14. Age: 8 months old	NPS	Illumina Miseq (V4)	Firmicutes (22-42%), Proteobacteria (10-30%), Tenericutes (5-20%), Actinobacteria (5-12%)	<i>Mycoplasma</i> (0.003-86%), <i>Psychrobacter</i> (0.55-90%), <i>Clostridium</i> , <i>Flavobacterium</i> , <i>Bacteroides</i> , <i>Rikenellaceae</i> , <i>Mannheimia</i> , <i>Acinetobacter</i> , <i>Corynebacterium</i> , <i>Moraxella</i> , <i>Pasteurella</i> , <i>Streptococcus</i>	Richness increased following feedlot placement (d2)	(Holman et al., 2017)
Bovine	Healthy Charolais calves (n = 8). Age: 6 to 8-month-old	DNS, BALF	Illumina Miseq (V3-V4)	DNS: Actinobacteria (43.9%), Proteobacteria (15.9%), Tenericutes (12.1%), Bacteroidetes (8.8%), Fusobacteria (0.1%) BALF: Proteobacteria (33.3%), Bacteroidetes (18.2%), Tenericutes (8.8%), Actinobacteria (4.9%), Fusobacteria (3.4%)	DNS: <i>Rathayibacter</i> (24.41%), <i>Mycoplasma</i> (12.20%), <i>Corynebacterium</i> (10.37%), <i>Clostridium</i> (2.53%), <i>Prevotella</i> (0.28%) BALF: <i>Mycoplasma</i> (9.48%), <i>Bibersteinia</i> (14.41%), <i>Prevotella</i> (11.69%), <i>Clostridium</i> (6.85%), <i>Rathayibacter</i> (0.07%)	No differences in $\alpha$ -diversity indices between DNS and BALF samples	(Zeineldin et al., 2017b)
Bovine	Healthy weaned feedlot cattle (n=60) vs cattle with bronchopneumonia (BP;	NPS, TA	Illumina Miseq (V4)	Tenericutes (47%), Proteobacteria (26%), Firmicutes (21%)	TA healthy group: <i>Mycoplasma</i> (46%), <i>Lactococcus</i> (18%), <i>Histophilus</i> (10%), <i>Moraxella</i> (6%), <i>Mannheimia</i> (6%)	Lower $\alpha$ -diversity in the NPS and TA of BP vs control	(Timsit et al., 2018)

(Continued)

TABLE 1 | Continued

Specie	Cohort description	Sample type	Platform and sequenced region	Main phyla	Genera	$\alpha$ -diversity	Reference
	n=60). Mixed-breed beef steers. Body weight: ~260 kg				Only in NPS healthy group: <i>Corynebacterium</i> (0.54%), <i>Jeotgalicoccus</i> (0.69%), <i>Psychrobacter</i> (0.52%), <i>Planomicrobium</i> (0.43%) NPS, TA of BP group: <i>Mycoplasma bovis</i> (46%), <i>Mannheimia haemolytica</i> (5.6%), <i>Pasteurella multocida</i> (2.18%)		
Bovine	Weaned Angus-cross beef heifers transported directly to a feedlot (RANC; n=30) vs heifers co-mingled at an auction market for 24 h before being placed in a feedlot (AUCTION; n=30). Time series: d0, d2, d7 and d30 after arrival	NPS, TA	Illumina Miseq (V4)	NPS: Tenericutes (41%), Proteobacteria (32%), Firmicutes (5%) TA: Tenericutes (50%), Proteobacteria (14%), Firmicutes (9%), Actinobacteria (4%)	NPS: <i>Mycoplasma</i> (40.8%), <i>Moraxella</i> (18.7%), <i>Pasteurella</i> (6.8%), <i>Mannheimia</i> (3.8%) TA: <i>Mycoplasma</i> (50.4%), <i>Pasteurella</i> (5.7%), <i>Moraxella</i> (3.1%)	No differences in $\alpha$ -diversity between RANC and AUCTION, but it decreased over time	(Stroebel et al., 2018)
Bovine	Commercial feedlot cattle injected with antimicrobials (n=20) vs in-feed antibiotics (n=20). Age: 0 to 12 months	NPS	Illumina Miseq (V3-V4)	Not specified	<i>Psychrobacter</i> (20-55%), <i>Pseudomonas</i> (1-50%), <i>Mycoplasma</i> (1-15%), <i>Moraxella</i> (0-10%)	Non specified	(Holman et al., 2018)
Bovine	Healthy feedlot cattle (n=3) vs cattle with BRDC (n=15)	BALF	HiSeq 2500 (WMS)	Not specified	Healthy: <i>Bacillus</i> (6%), <i>Enterococcus faecium</i> (5%), <i>Brachyspira hamptonii</i> (4%), <i>Bacillus obstructivus</i> (3%), <i>Streptococcus pneumoniae</i> (2%) BRDC: <i>Mannheimia haemolytica</i> (18.8%), <i>Mycoplasma bovis</i> (5%), <i>Mycoplasma dispar</i> (3%), <i>Enterococcus faecalis</i> (2%), <i>Bacillus</i> VT-16-64 (2%)	Not specified	(Klima et al., 2019)
Bovine	Weaned Angus × Herford cross calves transported to feedlot (n=13). Time series: auction market (day 0), and feedlot placement (day 2, 7, 14)	NPS	Illumina Miseq (V4)	Proteobacteria (36.1%), Firmicutes (20.1%), Tenericutes (19.3%), Actinobacteria (12.7%), Bacteroidetes (8.6%)	<i>Acinetobacter</i> (1-4%), <i>Bacteroides</i> (1-2%), <i>Bifidobacterium</i> (0-2%), <i>Corynebacterium</i> (0-9%), <i>Jeotgalicoccus</i> (1-2%), <i>Mannheimia</i> (0-1%), <i>Methanobrevibacter</i> (1-5%), <i>Moraxella</i> (0-10%)	Increase of richness following transport to an auction market and feedlot	(Amat et al., 2019)
Bovine	Angus × Herford steers transported to a feedlot and injected with antimicrobials (n=12) vs in-feed antibiotics (n=12) and untreated (n=12). Time series: d0, d2, d5, d12, d19 and d34. Age: weaned calves	NPS	Illumina Miseq (V4)	Not specified	Untreated: <i>Acinetobacter</i> (2-5%), <i>Corynebacterium</i> (2-6%), <i>Jeotgalicoccus</i> (1-3%), <i>Moraxella</i> (1-20%), unclassified <i>Ruminococcaceae</i> (2-10%), <i>Mycoplasma</i> (1-20%), <i>Planomicrobium</i> (2.5-5%), <i>Psychrobacter</i> (0-10%)	Richness and $\alpha$ -diversity increased following transport to the feedlot	(Holman et al., 2019)
Bovine	Healthy calves (n=82) vs BRDC calves (n=82). Time series: 3-12d, 13-20d, 21-44d. Animals were raised without antimicrobials. Age: newly received feedlot cattle of about ~218 kg	DNS	Illumina Miseq (V4)	Proteobacteria (69%), Tenericutes (23%), Firmicutes (3%), Actinobacteria (2%), Bacteroidetes (2%)	Healthy: <i>Moraxella</i> (22%), <i>Mycoplasma</i> (17%), <i>Histophilus</i> (16.62%) BRDC group: <i>Mycoplasma</i> (27%), <i>Histophilus</i> (21%), <i>Moraxella</i> (18%)	Species richness was lower in BRDC compared to healthy	(McMullen et al., 2019)
Bovine	Healthy pre-weaned calves fed 2 different milk-feeding programs (n=40). Age: 3.5 ± 1.15 days of age. BW: 39.3 ± 4.25 kg	DNS	Illumina Miseq (V1-V3)	Tenericutes (29.5%), Firmicutes (19.3%), Actinobacteria (19%), Proteobacteria (16%), Bacteroidetes (11.5%), Fusobacteria (2.5%)	<i>Pseudoclavibacter</i> (13.8%), <i>Mycoplasma</i> (29.5%). Differences between feeding regimes were observed for <i>Streptococcaceae</i> family and <i>Histophilus</i>	No differences in $\alpha$ -diversity between treatments	(Maynou et al., 2019)
Bovine	Healthy calves (n=9) vs BRDC affected calves treated with tilimicosin (n=9). Age: 6 to 8 months of age.	NPS	Illumina Miseq (V1-V3)	Firmicutes (27.07%), Actinobacteria (24.51%), Tenericutes (16.05%), and Proteobacteria (14.43%).	<i>Mycoplasma</i> (18.73%), <i>Microbacteriaceae</i> (9.36%), <i>Acinetobacter</i> (7.35%), <i>Corynebacterium</i> (6.36%)	No differences in $\alpha$ -diversity between groups	(Zeineldin et al., 2020b)

(Continued)

TABLE 1 | Continued

Specie	Cohort description	Sample type	Platform and sequenced region	Main phyla	Genera	$\alpha$ -diversity	Reference
Sheep	Suffolk cross sheep sampled at 3 spatially disparate segmental bronchi (n=6). Time series: day 0, 1 month, and 3 months. Age: 20 months old	SBT	Illumina Miseq (V2-V3)	BRDC-affected and treated calves showed significant decrease in Actinobacteria Not specified	<i>Staphylococcus</i> (16%), <i>Corynebacterium</i> (9%), <i>Jeotgalicoccus</i> (5%), <i>Streptococcus</i> (5%)	No difference in richness or $\alpha$ -diversity between different lung sites	(Glendinning et al., 2016)
Sheep	Scottish mule x Suffolk sheep (n=40). Age: 7 weeks old	OPS, BALF	Illumina Miseq (V2-V3)	Not specified	OPS: <i>Pasteurellaceae</i> (22%), <i>Mannheimia</i> (14%), <i>Fusobacterium</i> (11%), <i>Bibersteinia trehalosi</i> (8%), <i>Neisseriaceae</i> (7%), <i>Moraxella</i> (6%), <i>Bibersteinia</i> (5%) BALF: <i>Staphylococcus equorum</i> (13%), <i>Staphylococcus sciuri</i> (6%), <i>Mannheimia</i> (5%), <i>Prevotella</i> (5%)	No difference in richness or $\alpha$ -diversity	(Glendinning et al., 2017a)
Pig	Healthy commercial pigs (n=30) vs pigs with Glässer's disease (n=40). Age: 3-4 weeks old (before weaning)	DNS	Illumina Miseq (V3-V4)	Healthy: Proteobacteria (32.5%), Firmicutes (21.1%), Tenericutes (2.2%), Actinobacteria (1.3%) Glässer: Proteobacteria (44%), Firmicutes (18.5%), Tenericutes (2.2%), Actinobacteria (1.3%)	unclassified <i>Pasteurellaceae</i> (27.0%), <i>Moraxella</i> (17.2%), <i>Weeksella</i> (12.9%), <i>Haemophilus</i> (6.1%), Healthy: <i>Moraxella</i> (13.6%), <i>Enhydrobacter</i> (3.1%), <i>Haemophilus</i> (2.8%) Glässer: <i>Moraxella</i> (22.2%), <i>Haemophilus</i> (9.4%), <i>Enhydrobacter</i> (4.3%)	Healthy status was associated to higher species richness and $\alpha$ -diversity	(Correa-Fiz et al., 2016)
Pig	Piglets treated by Ceftiofur Crystalline free acid (CCFA, n=4), Ceftiofur hydrochloride (CHC, n=4), Tulathromycin (TUL, n=4), Oxytetracycline (OTC, n=4), or Procaine Penicillin G (PPG, n=4). Time series: day 0, 1, 3, 7, and 14 after dosing. Age: 8-week-old	DNS	Illumina Miseq (V1-V3)	Firmicutes (46.46%), Proteobacteria (31.87%), Bacteroidetes (9.64%)	<i>Moraxella</i> (21.52%), <i>Clostridium</i> (19.74%), <i>Streptococcus</i> (10.93%), <i>Calothrix</i> (5.43%), <i>Prevotella</i> (4.49%)	The $\alpha$ -diversity was not affected by treatment	(Zeineldin et al., 2018)
Pig	Cross-bred Yorkshire x Hampshire healthy newborn piglets (n=28). Time series: 8h post-birth, 1, 2, 3 and 4 weeks of age	TS	Illumina Miseq (V4)	Not specified	Post-born piglets were colonized by <i>Streptococcus</i> , <i>Staphylococcus</i> , <i>Moraxella</i> , <i>Rothia</i> , and <i>Pasteurellaceae</i> . By 1 week of age, members of the <i>Pasteurellaceae</i> , <i>Moraxellaceae</i> , and <i>Streptococcaceae</i> families were the most dominant	Not specified	(Cortes et al., 2018)
Pig	Healthy cross-bred pigs (n=10) vs pigs with Porcine respiratory disease (PRDC) disease (n=23). Age: 2-3 weeks of age (before weaning)	NPS	Illumina Miseq (V3-V4)	Firmicutes (53.11%), Proteobacteria (27.89%), Bacteroidetes (12.17%), Fusobacteria (3.15%), Actinobacteria (2.29%)	Healthy: <i>Lactobacillus</i> (30.44-37.28%), <i>Streptococcus</i> (23.60-32.26%), <i>Actinobacillus</i> (11.48-15.09%), <i>Bergeyella</i> (3.10%), <i>Escherichia-Shigella</i> (2.24%) PRDC: <i>Streptococcus</i> (22.22-26.25%), <i>Actinobacillus</i> (9.66-17.63%), <i>Moraxella</i> (12.02-15.82%), <i>Veillonella</i> (7.16%), <i>Bergeyella</i> (4.60-7.46%), <i>Fusobacterium</i> (4.64%), <i>Porphyromonas</i> (4.31%), <i>Escherichia-Shigella</i> (4.19%)	No differences in $\alpha$ -diversity and richness between the healthy and PRDC groups	(Wang et al., 2018)
Pig	Healthy Duroc Landrace Yorkshire cross-breed piglets (n=10) vs piglets challenged with Porcine	BALF	Illumina Miseq (V3-V4)	Healthy: Firmicutes (79.8-89.8%), Tenericutes (0.18-2.4%), Proteobacteria (5.3-13.4%)	Healthy: <i>Anoxybacillus</i> , <i>Caloramator</i> PRRSV: <i>Haemophilus parasuis</i> (35-48%), <i>Mycoplasma hyorhinis</i> (27-41%), <i>Bacteroides</i> (4-11%), <i>Chloroplast</i>	Reduced $\alpha$ -diversity in the PRRSV group	(Jiang et al., 2019)

(Continued)

TABLE 1 | Continued

Specie	Cohort description	Sample type	Platform and sequenced region	Main phyla	Genera	$\alpha$ -diversity	Reference
	reproductive and respiratory syndrome (PRRSV) (n=10). Age: 8 to 10 weeks of age			PRRSV: Proteobacteria (40.8-49%), Firmicutes (2.4-8.8%), Tenericutes (27-40.0%)	(1-3%), unclassified <i>Chitinophagaceae</i> (1-7%)		
Pig	Cross-bred pigs with lung-lesion and raised under natural conditions (n=20). Age: 240 days of age	BALF	Illumina Miseq (V3-V4)	Proteobacteria (34.2%), Tenericutes (22.3%), Bacteroidetes (18.8%), Firmicutes (18.1%)	<i>Mycoplasma</i> (13.0%), <i>Methylotenera</i> (10.9%), <i>Ureaplasma</i> (9.2%), <i>Phyllobacterium</i> (5.3%), <i>Prevotella</i> (4.0%), <i>Sphingobium</i> (3.2%), <i>Lactobacillus</i> (3.0%), <i>Thermus</i> (2.7%), <i>Streptococcus</i> (2.4%), <i>Haemophilus</i> (1.8%)	Reduced $\alpha$ -diversity in lungs with higher level of lesions	(Huang et al., 2019)
Pig	Commercial healthy pigs at slaughter (n=10)	TS	Illumina Miseq (V3-V4)	Proteobacteria (30-40%), Firmicutes (30%), Fusobacteria (20%), Bacteroidetes (10-20%). No differences between tonsil surface and in deep tonsil tissue	Surface of tonsils: <i>Actinobacillus</i> Deep tonsils: <i>Yersinia</i> , <i>Pasteurella</i> .	No difference in $\alpha$ -diversity between the surface and the deep tonsil tissue	(Jakobsen et al., 2019)
Pig	Duroc×Landrace×Yorkshire growing pigs exposed to different levels of gaseous ammonia (n=72). Body weight ~30 kg	DNS, TS	Illumina Miseq (V3-V4)	Proteobacteria (36.4%), Firmicutes (34.8%), Bacteroidetes (19.9%), Actinobacteria (4.1%)	<i>Moraxella</i> , <i>Pseudomonas</i> , <i>Lactobacillus</i> , <i>Prevotella</i> , <i>Bacteroides</i> , <i>Megasphaera</i> , <i>Streptococcus</i> , <i>Rothia</i> , <i>Allobaculum</i> , <i>Blautia</i> , <i>Oscillospira</i>	Ammonia concentration decreased the $\alpha$ -diversity	(Wang et al., 2019)
Pig	Commercial weaned pigs housed to simple slatted-floor (S, n=75) vs complex straw-based rearing ecosystem (C, n=75). Sampling time: from 164 days post-weaning at the time of slaughter	BMS	Illumina Miseq (V3-V4)	Firmicutes (58.2%), Proteobacteria (30%) Actinobacteria phylum was more abundant in C-raised pigs compared to S-raised pigs	<i>Bacteroidetes</i> , <i>Clostridium</i> , <i>Streptococcus</i> <i>Anaerotruncus</i> (8%) was higher in S-raised pigs whereas <i>Bacteroidetes</i> (24.3%) and <i>Chitinophagaceae</i> (15.4%) were higher in C-raised pigs	The S ecosystem increased the $\alpha$ -diversity	(Megahed et al., 2019)
Pig	Post-weaned pigs with oxytetracycline parenterally administered (n=21) or in feed (n=22), and the non-medicated feed group (n=22). Time series: days 0 (before start of treatment), 4, 7, 11, and 14	DNS, TS	Illumina Miseq (V4)		DNS untreated: <i>Streptococcus</i> (23.61%), <i>Moraxellaceae</i> (29.74%), <i>Actinobacillus</i> (9.30%), <i>Moraxella</i> (3.36%), <i>Lactobacillus</i> (2.43%), DNS antibiotic: <i>Moraxellaceae</i> (32.84%), <i>Streptococcus</i> (12.68%), <i>Moraxella</i> (5.01%), <i>Lactobacillus</i> (2.15%) Tonsils untreated: <i>Veillonella</i> (18.27%), <i>Streptococcus</i> (16.31%), <i>Actinobacillus</i> (15.46%), <i>Bacteroides</i> (13.91%), <i>Fusobacterium</i> (6.95%) Tonsils antibiotic: <i>Actinobacillus</i> (22.90%), <i>Bacteroides</i> (17%), <i>Veillonella</i> (14.03%), <i>Streptococcus</i> (13.60%), <i>Fusobacterium</i> (7.09%)	Nasal $\alpha$ -diversity with antibiotic treatment was lower compared to control group. In tonsils, $\alpha$ -diversity was not affected by treatment	(Mou et al., 2019)
Pig	Healthy cross-bred pigs (n=30) vs pigs with Glässer's disease, n=51). Age: weaning pigs (3-4 weeks of age)	DNS	Illumina Miseq (V3-V4)	Not specified	Glässer: <i>Corynebacterium</i> , <i>Clostridium</i> XI, <i>Escherichia/Shigella</i> , Healthy: <i>Odoribacter</i> , <i>Planobacterium</i> , <i>Phascolarctobacterium</i> .	Not specified	(Mahmmod et al., 2020)
Pig	Healthy duroc×Landrace×Yorkshire growing pigs (n=8) vs diseased pigs with PRDC (n=20). Age: 270 ± 3 days of age	BALF	Illumina Miseq (V3-V4)	Healthy: Proteobacteria (59%), Firmicutes (28.55%), Tenericutes (9.94%), Bacteroidetes (2%) Diseased: Proteobacteria (54%), Firmicutes (34%), Tenericutes (3%), Bacteroidetes (7%)	Healthy: Higher abundance of <i>Lactococcus</i> , <i>Enterococcus</i> , <i>Staphylococcus</i> , <i>Lactobacillus</i> . Diseased: Enhanced richness of <i>Streptococcus</i> , <i>Haemophilus</i> , <i>Pasteurella</i> , <i>Bordetella</i>	$\alpha$ -diversity was lower in healthy individuals than in diseased group	(Li et al., 2021)

(Continued)



TABLE 1 | Continued

Specie	Cohort description	Sample type	Platform and sequenced region	Main phyla	Genera	$\alpha$ -diversity	Reference
Bird	Birds raised at 3 different farms in Pakistan (n=14)	TA, BALF	454 Roche	Proteobacteria, Firmicutes, Tenericutes, Actinobacteria, Bacteroidetes	Farm A: unclassified $\gamma$ Proteobacteria, Farm B: <i>Avibacterium</i> , Farm C: unclassified <i>Enterobacteriaceae</i> , <i>Pseudomonas</i>	The $\alpha$ -diversity was higher in farms B and C than farm A	(Shabbir et al., 2014)
Bird	Healthy commercial chickens aged 2 days (n = 5), 3 weeks (n = 5) and 30 months (n = 6)	DNS, BALF	Illumina Miseq (V2-V3)		DNS: <i>Staphylococcus</i> (8.0%), <i>Lactobacillus</i> (6.2%), unclassified <i>Enterobacteriaceae</i> (6.0%), <i>Faecalibacterium prausnitzii</i> (5.0%), <i>Staphylococcus equorum</i> (5.0%), <i>Lactobacillus reuteri</i> (4.4%) BALF: <i>Pseudomonas</i> (20.7%), <i>Achromobacter</i> (4.8%), <i>Lactobacillus</i> (4.8%), <i>Turicibacter</i> (4.7%), SMB53 (3.6%). The 30-month-old birds showed lower <i>lactobacilli</i> but higher <i>Jeotgalicoccus</i> , <i>Staphylococcus</i> and <i>smb53</i>	The richness and $\alpha$ -diversity were different between DNS and BALF. Richness raised with age	(Glendinning et al., 2017b)
Bird	Commercial Cobb 500 broilers from different flocks (n=120). Three grow-out cycles from 0 to 42 d of age. A cross-sectional sampling with different ages (n=90)	TA	Illumina Miseq (V4)	Not specified	<i>Lactobacillus</i> , <i>Escherichia/Shigella</i> , <i>Staphylococcus</i> , <i>Corynebacterium</i> , unclassified <i>Moraxellaceae</i> , unclassified <i>Ruminococcaceae</i> , unclassified <i>Clostridiales</i> , <i>Ruminococcus</i> , <i>Psychrobacter</i> , <i>Blautia</i>	Not specified	(Johnson et al., 2018)
Bird	Hy-Line W-36 commercial layers (n=181). Nine grow-out cycles from 5 weeks to > 17 weeks of age	DNS, TA	Illumina Miseq (V3-V4)	Not specified	DNS: <i>Staphylococcus</i> , <i>Enterobacteriaceae</i> , unclassified <i>Lactobacillaceae</i> , <i>Lactobacillus reuteri</i> , <i>Faecalibacterium prausnitzii</i> , <i>Deinococcus</i> , unclassified <i>Burkholderiaceae</i> TA: <i>Lactobacillus</i> , <i>Clostridium</i> , <i>Enterococcus</i> , <i>Escherichia-Shigella</i> , <i>Callibacterium</i> , <i>Imrithobacterium</i> , <i>Staphylococcus</i> , <i>Streptococcus</i> , <i>Deinococcus</i> , <i>Corynebacterium</i> , and <i>Staphylococcus</i>	Not specified	(Ngunjiri et al., 2019)
Bird	Healthy commercial turkeys sampled at 3 time points during brooding (1, 3, and 5 weeks) and grow-out (8, 12, and 16 weeks; n=104).	DNS, TA	Illumina Miseq (V4)	DNS: Firmicutes and Actinobacteria TA: Proteobacteria	<i>Deinococcus</i> , <i>Corynebacterium</i> , and <i>Staphylococcus</i>	The $\alpha$ -diversity in the nasal cavity or trachea did not change with age	(Taylor et al., 2020)
Bird	Healthy ale Arbor Acres broilers exposed to 4 different levels of ammonia for 21 days (n=228). Age: 22 days of age	TA	Illumina Miseq (V3-V4)	Firmicutes (70%), Proteobacteria (15%)	<i>Lactobacillus</i> decreased under different levels of ammonia exposure, whereas <i>Faecalibacterium</i> , <i>Ruminococcus</i> , unclassified <i>Lachnospiraceae</i> , <i>Ruminococcaceae</i> UCG-014, <i>Streptococcus</i> and <i>Blautia</i> increased with ammonia levels	The $\alpha$ -diversity decreased with the ammonia levels	(Zhou et al., 2021)
Horse	Healthy horses (n=3) vs healthy horses treated with dexamethasone (n = 3) and vs horses with Inflammatory Airway Disease (IAD; n = 7).	DNS, BALF	Illumina Miseq (V3-V4)	Proteobacteria (43.85%), Actinobacteria (21.63%), Firmicutes (16.82%), Bacteroidetes (13.24%)	<i>Sphingomonas</i> (15.65%), <i>Pseudomonas</i> (14.57%), <i>Pantoea</i> (11.68%), <i>Knoellia</i> (2.80%), <i>Agrococcus</i> (2.61%) <i>Arthrobacter</i> (2.61%). <i>Streptococcus</i> was higher in IAD horses. Dexamethasone had no effect	No differences in $\alpha$ -diversity. Decrease in richness in BALF	(Bond et al., 2017)
Horse	Healthy adult horses (n=6) vs asthmatic horses (n=6). Animals were at pasture (low antigen exposure), housed indoors and fed good- quality hay ("moderate exposure") or	DNS, BALF	Illumina Miseq (Not specified)	DNS: Proteobacteria, Firmicutes BALF: Proteobacteria, Firmicutes, Bacteroidetes, Actinobacteria, Verrucomicrobia	DNS: <i>Pasteurella multocida</i> (39.5%), <i>Actinobacillus</i> (23.2%) BALF: <i>Enterobacteriaceae</i> (4%), <i>Pasteurella multocida</i> (3.0%), <i>Comamonadaceae</i> (2%), <i>Actinobacillus</i> (1.9%), <i>Staphylococcus</i> (1%)	DNS had higher richness and $\alpha$ -diversity than BALF samples	(Fillion-Bertrand et al., 2019)

(Continued)

TABLE 1 | Continued

Specie	Cohort description	Sample type	Platform and sequenced region	Main phyla	Genera	$\alpha$ -diversity	Reference
Horse	poor-quality hay ("high exposure") Healthy horses (n=23) vs horses affected with primary (n=14) and secondary sinusitis (n=62). Age: 2 - 30 years old	PS	Illumina Miseq (V4)	Proteobacteria (63%, 23.7–99.0%), Firmicutes (14%, 0.38–64%), Actinobacteria (5.19, 0–62%)	Healthy: <i>Pseudomonas</i> (14.6%, 0.2–50.2%), <i>Delftia</i> (7.8%, 0–26.2%) <i>Stenotrophomonas</i> (6.7%, 0.1–18.2%) <i>Dokdonella</i> (5.1%, 0–80.8%) <i>Aggregatibacter</i> (4.7%, 0–71.0%) <i>Acinetobacter</i> (3.6%, 0–20.3%) <i>Achromobacter</i> (2.7%, 0–20.6%). Sinusitis: <i>Streptococcus equi</i> subsp. <i>Zooepidemicus</i> , <i>Fusobacterium</i>	No differences in Simpson diversity index between healthy and sinusitis groups	(Beste et al., 2019)
Horse	Healthy horses (n=4) vs horses with naturally occurring mild asthma (n = 16). Body weight ~435-612 kg Horses were treated with nebulized dexamethasone for 14 d.	NPS, BALF	Illumina Miseq (V4)	NPS: Proteobacteria (37.81%), Bacteroidetes (25.71%), Actinobacteria (17.77%), Firmicutes (17.9%)	<i>Hymenobacter</i> (16.51%), <i>Staphylococcus</i> (13.49%), <i>Pedobacter</i> (6.33%), <i>Moraxella</i> (4.50%), <i>Sphingomonas</i> (3.75%), <i>Pseudomonas</i> (3.62%)	Nebulized dexamethasone treatment decreased $\alpha$ -diversity in the nasopharynx	(Bond et al., 2020)
Dog	Healthy female dogs (n=16). Age: 2 - 8 years of age	DNS, NPS, BALF	Illumina Miseq (V4)	DNS: Proteobacteria (55.40%), Actinobacteria (0.73%) NPS: Proteobacteria (60.79%), Tenericutes (7.04%), Actinobacteria (0.41%) BALF: Proteobacteria (88.53%), Actinobacteria (2.61%)	DNS: unclassified <i>Pasteurellaceae</i> (7.20%), <i>Brevundimonas diminuta</i> (2.69%), <i>Cardiobacteriales</i> (2.11%) NPS: <i>Porphyromonas</i> (8.32%), <i>Fusobacterium</i> (2.96%), unclassified <i>Weeksellaceae</i> (2.70%), unclassified <i>Neisseriaceae</i> (3.36%) BALF: <i>Brevundimonas diminuta</i> (22.48%), <i>Sphingopyxis alaskensis</i> (2.37%), unclassified <i>Pasteurellaceae</i> (2.36%), <i>Propionibacteriaceae</i> unclassified (1.9%), unclassified <i>Bradyrhizobiaceae</i> (1.23%), unclassified <i>Methylobacteriaceae</i> (1.68%)	BALF in healthy dogs had equivalent richness to DNS	(Ericsson et al., 2016)
Dog	Healthy dogs (n = 23), dogs with malignant nasal neoplasia (n = 16), and dogs with chronic rhinitis (n = 8). Age ~6 years old	DNS	Illumina Miseq (V4)	Healthy: Proteobacteria (83.4%, 37.4%– 98.5%), Firmicutes (4.8%, 0.4–20.8%), Bacteroidetes (2.6%, 0.1–12.5%), Cyanobacteria (2.1%, 0.0–11.6%), Actinobacteria (2.1%, 0.1–8.6%).	Healthy group: <i>Moraxella</i> (59.2%), <i>Phyllobacterium</i> (3.4%), family <i>Cardiobacteriaceae</i> (2.1%), <i>Staphylococcus</i> (1.7%). Diseased group: <i>Moraxella</i> (34.5%, 0.7–77.3%), order Streptophyta (6.4%, 0.0–16.6%), <i>Riemerella</i> (4.4%, 0.0–25.3%), family <i>Pasteurellaceae</i> (2.9%, 0.2–17.1%)	Shannon diversity index was lower for the healthy dogs than for the diseased dogs	(Tress et al., 2017)
Dog	Healthy client-owned dogs (n=5) vs client-owned dogs diagnosed with bacterial pneumonia (n=15)	OPS, BALF	Illumina Miseq (V4)	Not specified	OPS healthy: <i>Escherichia-Shigella</i> (15.33%), <i>Prevotella</i> (15.21%), <i>Streptococcus</i> (10.88%), <i>Bacteroides</i> (10.96%), <i>Mycoplasma canis</i> (7.67%), <i>Acinetobacter</i> (3.90%), BALF healthy: <i>Streptococcus</i> (96.4%) BALF healthy: <i>Pseudomonas stutzeri</i> (29.5%), <i>Acinetobacter</i> (20.9%), and <i>Brevundimonas</i> (20.1%) BALF diseased: <i>Acinetobacter</i> (22.3%), <i>Bradyrhizobium</i> (9.3%), <i>Brevundimonas</i> (8.3%), <i>Agrobacterium radiobacter</i> (7.1%)	Richness was decreased with pneumonia	(Vientós-Plotts et al., 2019)
Dog	Healthy research dogs (n=16) vs client-owned dogs diagnosed with Chronic Bronchitis (n=53). Age: between 8 and 9 years old	BALF	Illumina Miseq (V4)	Not specified	BALF healthy: <i>Streptococcus</i> (96.4%) BALF healthy: <i>Pseudomonas stutzeri</i> (29.5%), <i>Acinetobacter</i> (20.9%), and <i>Brevundimonas</i> (20.1%) BALF diseased: <i>Acinetobacter</i> (22.3%), <i>Bradyrhizobium</i> (9.3%), <i>Brevundimonas</i> (8.3%), <i>Agrobacterium radiobacter</i> (7.1%)	Diseased dogs had lower richness than healthy research dogs	(Ericsson et al., 2020)
Dog	Healthy west highland white terriers (n=6) vs west highland white terriers with idiopathic pulmonary fibrosis (n=11)	BALF	Illumina Miseq (V1–V3)	Proteobacteria, Actinobacteria, Firmicutes, Bacteroidetes	Healthy group: <i>Cutibacterium</i> , <i>Staphylococcus</i> , <i>Streptococcus</i> , <i>Pseudomonas</i> , <i>Corynebacterium</i> , unclassified <i>Pasteurellaceae</i> , <i>Acinetobacter</i> , <i>Conchiformibius</i> , <i>Flavobacterium</i> , <i>Porphyromonas</i> .	Lower $\alpha$ -diversity in diseased dogs compared to healthy	(Fastrès et al., 2020a)

(Continued)

TABLE 1 | Continued

Specie	Cohort description	Sample type	Platform and sequenced region	Main phyla	Genera	$\alpha$ -diversity	Reference
Dog	Healthy dogs of different breeds (n=45) vs dogs diagnosed with idiopathic pulmonary fibrosis (n=11) exposed to different living conditions	BALF	Illumina Miseq (V1–V3)	Healthy: Proteobacteria, Actinobacteria, Firmicutes, Bacteroidetes	Diseased group: <i>Brochothrix</i> , <i>Curvibacter</i> , <i>Pseudarcicella</i> , unclassified <i>Flavobacteriaceae</i> BALF healthy: <i>Cutibacterium</i> , <i>Staphylococcus</i> , <i>Streptococcus</i> , <i>Pseudomonas</i> , <i>Corynebacterium_1</i> , <i>Pasteurellaceae</i> genus, <i>Acinetobacter</i> , <i>Conchiformibius</i> , <i>Flavobacterium</i> and <i>Porphyromonas</i> (all together ~23% of bacterial population) BALF diseased: predominance of <i>Rhodoluna</i> , <i>Brochothrix</i> , <i>Curvibacter</i> , <i>Pseudarcicella</i> , <i>Flavobacteriaceae</i> family	No differences between living conditions for the $\alpha$ -diversity and the evenness. No differences between healthy and diseased dogs	(Fastrès et al., 2020b)
Cat	Healthy cats (n=6). Time series: day 0, week 2, and week 10. Age < 1 year old	OPS, BALF	Illumina Miseq (V4)	OPS Proteobacteria (60%, 54.78–74.28) BALF: Proteobacteria (62.36%, 27.26–81.65%)	OPS <i>Pasteurellaceae</i> (15.99%), <i>Moraxellaceae</i> (14.79%), <i>Porphyromonadaceae</i> (12.45%), <i>Pseudomonadaceae</i> (10.21%), <i>Paraprevotellaceae</i> (5.46%). BALF: <i>Pseudomonadaceae</i> (34.24%), <i>Sphingobacteriaceae</i> (22.56%), <i>Bradyrhizobiaceae</i> (15.86%), <i>Bifidobacterium</i> (35.93%), <i>Bifidobacterium animalis</i> (23.29%), <i>Streptococcus</i> (25.53%), <i>Lactobacillus zeae</i> (11.64%), <i>Lactobacillus</i> (3.21%)	OPS was richer than BALF. No differences in richness between time points	(Vientoos-Plotts et al., 2017)
Cat	Probiotic administration in healthy cats (n=6). Time series: baseline and after probiotic administration. Age < 1 year old	OPS, BALF	Illumina Miseq (V4)	Actinobacteria (59.27%), Firmicutes (40.67%)	<i>Bifidobacterium</i> (35.93%), <i>Bifidobacterium animalis</i> (23.29%), <i>Streptococcus</i> (25.53%), <i>Lactobacillus zeae</i> (11.64%), <i>Lactobacillus</i> (3.21%)	Probiotic increased the richness in the OPS and BALF	(Vientós-Plotts et al., 2017)
Cat	Healthy cats (n = 28) vs cats with nasal neoplasia (n = 16), and cats with feline upper respiratory tract disease (FURTD; n = 15). Age ranged from 6 months to 14.0 years	DNS	Illumina Miseq (V4)	Proteobacteria, Firmicutes, and Bacteroidetes	Healthy: <i>Moraxella</i> (33%), unclassified <i>Bradyrhizobiaceae</i> (11.3%), <i>Sediminibacterium</i> (5.8%), <i>Staphylococcus</i> (4.3%), <i>Pseudomonas</i> (2.3%) FURTD: <i>Moraxella</i> (38.6%), unclassified <i>Bradyrhizobiaceae</i> (8.8%), <i>Staphylococcus</i> (6.3%), <i>Chlamydia</i> (5.7%), <i>Pasteurella</i> (5.7%), <i>Sediminibacterium</i> (4.6%), <i>Bibersteinia</i> (4%) Neoplasia: <i>Moraxella</i> (15.8%), unclassified <i>Bradyrhizobiaceae</i> (20.6%), unclassified <i>Chitinophagaceae</i> (7.3%), <i>Phyllobacterium</i> (6.6%), <i>Pasteurella</i> (5.1%)	No differences between health and diseased cats	(Dorn et al., 2017)

BALF, Bronchoalveolar lavage fluid; BMS, Bronchus mucosal scraping; DNS, deep nasal swap; NPS, nasopharyngeal swabs; OPS, Oropharyngeal swabs; PS, Paranasal sinus; SBT, Segmental bronchi throat; TA, tracheal aspiration; TS, tonsil swabs; WMS, whole metagenome sequencing.

life, a wide range of microorganisms are acquired after the passage through the birth canal, during suckling and maternal care, and from the immediate environment at delivery, such as pen material, feed and feces. Consequently, the microbiota of the calf URT is highly similar to the maternal vaginal microbiota (Lima et al., 2019). Immediately after birth, the airway microbiota in ruminants evolves and reaches a maximum of diversity within one month of age (Timsit et al., 2017; Lima et al., 2019). The taxa organisms detected in the LRT shift from Proteobacteria and Firmicutes to Bacteroidetes and Tenericutes across age (Lima et al., 2019). Such microbiota changes are thought to be intertwined with immune system maturation that promotes tolerance to environmental allergens

(Ramírez-Labrada et al., 2020). If the colonization process is disrupted, the animal may develop a dysbiotic microbiota, causing a predisposition to contracting complex respiratory disease. An elegant review by Zeineldin et al. (2019) about ruminants declared that changes in diet (Hall et al., 2017; Maynou et al., 2019), antimicrobial use (Collie et al., 2015; Timsit et al., 2017; Holman et al., 2018; Holman et al., 2019; McMullen et al., 2019), pathogen exposure (Holman et al., 2015), as well as early life management procedures, including weaning, vaccination, commingling, long-distance transportation and housing (Timsit et al., 2016; Holman et al., 2017; Timsit et al., 2017; Zeineldin et al., 2017a; McMullen et al., 2018; Amat et al., 2019) are major contributing factors to the initial seeding

disruption of the airway microbiota and have been associated with various health outcomes later in life (**Figure 2**).

Along the same lines, the nasal microbiota of piglets following delivery resembles that of the sow and depends on the route by which the pig is delivered and the feeding type (Wang et al., 2013a). The tonsillar microbiota of piglets following birth resembles the sow vaginal and teat skin microbiota, indicating that these two body sites contribute to the colonization of the URT (Pena Cortes et al., 2018). However, no studies have examined the initial seeding and development of the LRT microbiome in swine. Thereafter, the microbiota begins to stabilize and progress toward an adult-like composition 2–3 weeks after the weaning (Slifierz et al., 2015; Cortes et al., 2018). As previously described in the gut (Mach et al., 2015), weaning is the critical period for the establishment of a robust and stable adult-like microbiota composition in piglets (Pirolo et al., 2021). Alongside weaning, other key environmental factors can modify the composition of the URT microbiota early in life, including the addition or removal of feed antibiotics (Zeineldin et al., 2018; Correa-Fiz et al., 2019; Mou et al., 2019; Zeineldin et al., 2019), gaseous ammonia concentration (Michiels et al., 2015; Wang et al., 2019), type of floor (Megahed et al., 2019) or feeding strategies (Weese et al., 2014).

The way in which different bacteria populate the respiratory microbiota is unknown in horses. Yet, it is not demonstrated that mode of birth, type of diet and antibiotic use have profound effects on the respiratory microbiome of newborn up to some months or years of age in horses. The age at which the respiratory microbiota acquires an adult-like configuration is still unclear. Maternal separation at weaning has a pivotal influence on the gut microbiome composition in foals (Mach et al., 2017). This likely holds true for the respiratory microbiota. Our previous findings in the gut (Mach et al., 2020; Mach et al., 2021a; Mach et al., 2021b) also highlight the possibility that the airway microbiota may be particularly sensitive to where a horse lives and what a horse does. In fact, short-term changes in housing and forage type alters the pulmonary microbiota in horses (Fillion-Bertrand et al., 2019), as well as the contact with animals and people who work at horse facilities, veterinary health care and medication (Kauter et al., 2019).

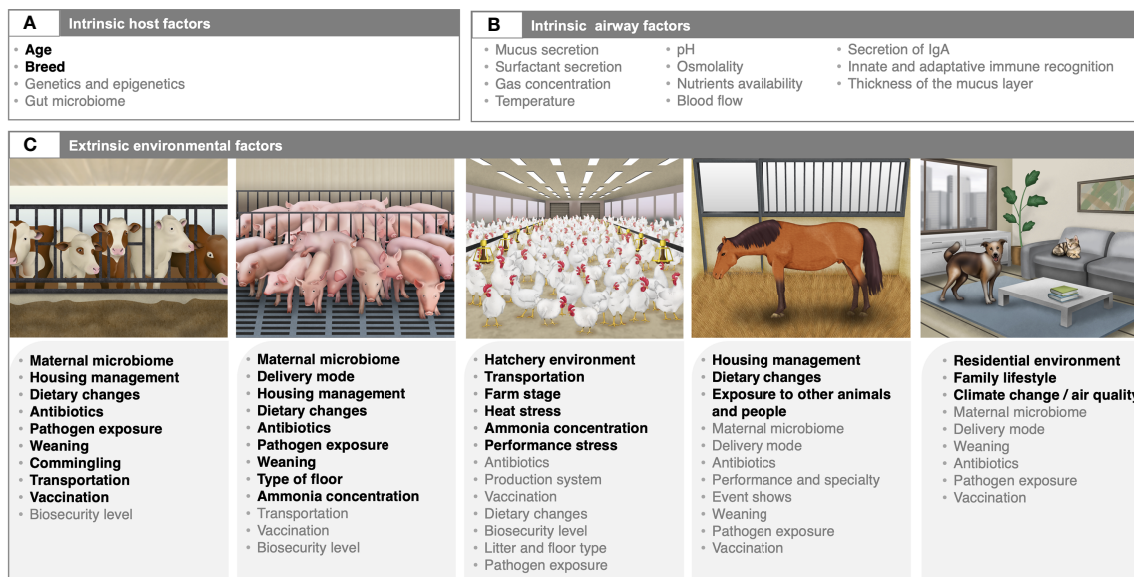
In a similar manner, published literature on the effects of birth mode on the respiratory microbial composition during the first years of life in companion animals is scant. Whether C-section is associated with lower bacterial diversity and linked with allergy is also unclear. Nonetheless, the combined effect of passive (living environment) and active (lifestyle) factors on the airway microbiota in companion animals from developed countries have started to receive attention for its role in complex respiratory diseases, especially asthma (**Figure 2**). At least, dogs living in urban environments (worse air quality/air pollution) and exposed to an urban-type lifestyle (e.g., living closed in apartments for hours and in a single-person family without other pets) may present an altered microbiota and be more susceptible to respiratory diseases compared to those living in a rural environment and farming lifestyle (Lehtimäki et al., 2018). In agreement with the posited hypothesis, climate change that affects air quality has been shown to lead to alterations in

microbial communities in the dog airways (Ericsson et al., 2020). Therefore, the living conditions are suspected to play a role in the airway microbiota in dogs, while no differences are found between types of breeds (Fastrès et al., 2020b). This assumed interplay between urban-type lifestyle, microbial exposure, host microbiota and respiratory disease needs to be confirmed in cats. The information of whether the microbiota of urban cats is more alike than rural ones is still lacking.

Concerning the airway microbiota in birds, the initial colonization of commercially hatched chickens is mainly dependent on the hatchery environment and transport to the farm (Brugman et al., 2018; Kubasova et al., 2019). Notably, some microorganisms can be acquired in the pre-hatching phase, directly from the mother in the oviduct of the hen, or from the environment through the pores in the eggshell (Kers et al., 2018). After birth, lower airway microbiota of commercial chickens kept indoors gradually assembles, without clear separation between brooding, growing and laying stages, while nasal microbiota shifts drastically after the birds transitioned to the laying stage (17 weeks onward) (Ngunjiri et al., 2019). Yet, despite the results by Ngunjiri et al. (2019), little is known about what drives the acquisition and development of the respiratory microbiota during early in life in poultry. Information is scant when focusing on chicks that have any form of contact with adult hen microbiota, chicks that live outdoors throughout their whole life or free-range organic chicken. Exposure to chronic heat-stress (Sohail et al., 2015), atmosphere ammonia concentrations (Liu et al., 2020; Zhou et al., 2021), housing and environmental conditions (Kursa et al., 2021), as well as performance stress (Kursa et al., 2021) are known to shape the composition of the respiratory tract microbiota in domestic birds. As for gut, environmental factors such as biosecurity level, disease onset, litter and feed access may each influence the poultry airway microbiota establishment after hatching (Kers et al., 2018) (**Figure 2**). Antibiotic treatments and vaccinations, which are often administered *via* spray or eye/nose drops, may also affect the development of the microbiota.

## **FRENEMIES: LACTOBACILLUS AND PROTEOBACTERIA IN COMPLEX RESPIRATORY DISEASES**

Host mechanisms for pathobiome regulation in the respiratory apparatus might be different between mammalian animal hosts and birds, but they have in common the aim of both preventing the invasion by pathogens and managing symbiotic microorganisms to maximize the benefits for the host. Precisely, the commensal microbiota may directly inhibit the persistence, transmission and evolution of pathogens through mechanisms such as competing for nutrients, bactericidal activity, metabolic inhibition, biofilm formation, quorum sensing, disruption of signaling molecules, and spatial occlusion (Man et al., 2017; Zhu et al., 2019). Additionally, host systemic and respiratory immune response (Man et al., 2017; Zaneveld et al., 2017; Amat et al., 2019; Budden et al., 2019; Li et al., 2019) may indirectly affect the interaction between



**FIGURE 2** | Intrinsic and extrinsic factors affecting the composition and development of the airway microbiota in domestic animals. The respiratory tract microbiota composition and development are dynamic and are shaped by various intrinsic host factors (A), as well as intrinsic airway milieu conditions (B), and environmental variables (C). Factors highlighted in bold are based on published literature. For ruminants and swine, factors are mainly extracted from the review by Zeineldin et al. (2019) and Pirolo et al. (2021), respectively. Variables colored in grey are those that illustrate directional hypothesis made by the authors. Written permission for publication of the domestic animals' drawings in the figure has been taken, except for the horse drawing. Horse drawing has been obtained from Mach et al. (2020).

commensals and pathogens. None of these mechanisms are wholly effective, but they all reflect common actions by commensal microbiota to neutralize threats to their hosts and, by extension, to themselves.

At this time, several examples of possibly actionable mechanisms of competition and cooperation have been reported in ruminants. For instance, microbiota of the alveolar epithelium in adult sheep synthesizes antimicrobial anionic peptides that quickly bind to and inactivate *Mannheimia haemolytica* (Heidari et al., 2002). In cattle, Amat et al. (2019) suggested that the presence of bacteria from the *Lactobacillaceae* family have a competitive exclusion effect on the BRDC-associated *Pasteurellaceae* family. These results were supported by inhibition assays, in which strains within the families of *Lactobacillaceae*, *Streptococcaceae*, and *Enterococcaceae* displayed *in vitro* antimicrobial activity (Amat et al., 2019). Additionally, *Lactobacillus* were shown to inhibit the growth of *Mannheimia haemolytica* *in vitro* (Amat et al., 2017). Taken together, commensal species from the *Lactobacillaceae* family may offer resistance to pathogens directly by producing antimicrobials and beneficial metabolites (e.g. lactic acid and SCFA) or indirectly through keeping cells of the innate and adaptive immune system on the tip of their toes, ready to respond rapidly to pathogenic threats when required.

In pigs, the supplementation with the Lactobacillales *Enterococcus faecium* as probiotic carried direct adsorptive trapping of swine influenza viruses (H1N1 and H3N2 subtypes) by direct physical interaction and by reinforcing innate antiviral defense through mediators such as IL-6,

TNF- $\alpha$ , IL-10, IFN- $\alpha$  and TLR-3 (Wang et al., 2013b). Evidence that probiotics may be effective in reducing severity, duration, and incidence of respiratory illnesses in herd-based species was stated as being low to very low (Suez et al., 2019).

While an increased prevalence of *Lactobacillus* is a potential diagnostic signature of eubiosis in ruminants and swine, respiratory disorders in those farm animals are often accompanied by an abnormal expansion of Proteobacteria (Table 1). More specifically,  $\gamma$ -Proteobacteria, a phylum containing a wide variety of pathobionts, is frequently enriched in the respiratory tract of infected individuals of cattle and pigs, thereby increasing the likelihood of pathogenic responses. In fact, numerous studies in cattle have linked a remarkable loss in  $\alpha$ -diversity to a gain in Proteobacteria during disease. For example, the URT of BRDC-affected calves presented higher abundance of *Moraxella*, *Streptococcus*, *Haemophilus*, *Neisseria* (Holman et al., 2015), *Pseudomonas* (Gaeta et al., 2017), *Solibacillus* and *Pasteurella* spp. (Zeineldin et al., 2017a). Within the  $\gamma$ -Proteobacteria, *Pasteurella* spp. was also increased by 1.5 in calves with pneumonia, as diagnosed by ultrasonography, compared with calves without pneumonia (Raabis et al., 2021). The lower  $\alpha$ -diversity of the nasopharyngeal and tracheal bacterial communities of cattle with bronchopneumonia likely explained why they were colonized by pathogens such as *Mycoplasma bovis*, *Mannheimia haemolytica* and *Pasteurella multocida* (Timsit et al., 2018). Parallel work by Lima et al. (2016) demonstrated a marked increase in abundance of the *Mannheimia*, *Moraxella*, and *Mycoplasma* genera in the URT of

three days-old calves coupled to the development of pneumonia and otitis media in calves during the first 60 days of life.

Comparably, a significant gain in the Proteobacteria and loss in Firmicutes, species richness and  $\alpha$ -diversity was seen in the nasal microbiota of piglets with Glässer's disease (Correa-Fiz et al., 2016). Interestingly, the *Pasteurellaceae* was one of the main families responsible for the higher abundance in Proteobacteria, with *Haemophilus parasuis*, being clearly increased, as well as the *Mycoplasmataceae* family (Correa-Fiz et al., 2016). The pro-inflammatory bacteria *Moraxella* spp., was significantly increased in the oropharynx of suckling pigs with porcine respiratory disease (Wang et al., 2018), whilst Proteobacteria such as *Sphingobium*, *Haemophilus* and *Phyllobacterium* co-occurred with *Mycoplasma* and *Ureaplasma* in lungs with severe lesions.

Consistent with results observed in pigs and cows, horses infected with equine herpesvirus 1 (EHV-1) had lower nasal bacterial richness, evenness and diversity than healthy horses, conjointly with higher abundance of Proteobacteria (Gomez et al., 2021).

Broadly, a sustained increase in abundance of the phylum Proteobacteria seems a biomarker of dysbiosis in the respiratory tract of cattle, pigs and horses. Whether Proteobacteria are flourishing better when the immune system is stimulated, or whether they actively contribute to the host immune pathology under pathogen infection has still to be explored. As Proteobacteria can tolerate oxidative stress (Million and Raoult, 2018), their expansion is likely linked to the onset of mucosa inflammation following primary infection. Additionally, they encode the metabolic capacity to utilize inflammatory byproducts to survive and prosper under acid and low oxygen conditions (Huffnagle et al., 2017), outcompeting bacteria that lack the metabolic capacity to benefit from inflammation. However, the phylum Proteobacteria, named after the Greek god Proteus, has the largest phylogenetic structure, including 116 validated families of bacteria (Shin et al., 2015). As the name suggests, this phylum presents a large range of morphologies and versatile functions (Shin et al., 2015). Indeed, in cats and dogs, *Pseudomonas* and *Acinetobacter*, microorganisms belonging to the phylum Proteobacteria, probably exert a protective function in the lung of companion animals **Table 1**). Bacteria in those two genera likely have multiple high-affinity adhesins that mediate binding and biofilm formation preventing the growth of potential pathogens in the lung of cats and dogs (Vientoos-Plotts et al., 2017; Fastrès et al., 2020b). In addition, *Acinetobacter* could play an anti-inflammatory role in the lungs (Fastrès et al., 2020b). Despite arguments regarding beneficial functions of Proteobacteria, a blooming pattern of Proteobacteria following disruption of homeostasis by environmental or host factors can further facilitate inflammation or invasion by exogenous pathogens in mammals.

Yet, the nature of Proteobacteria in the respiratory pathobiome in poultry remains largely unknown. Only Liu et al. (2020), have recently evoked a relationship between the increase of Proteobacteria under seven days of ammonia exposure and respiratory tract diseases in broilers.

## COMPLEX RESPIRATORY DISEASE AND COMORBIDITIES: THE MULTIPLE PATHOGEN DISEASE PARADIGM IN A PATHOBIOME CONTEXT

As central finding from respiratory tract microbiota is that most complex airway diseases are caused by pathogens that can be common members of a symbiome in the absence of disease. The well-characterized BRDC is an example of how commensal inhabitants of the respiratory tract (namely *Mannheimia haemolytica*, *Histophilus somni*, *Pasteurella multocida*, *Trueperella pyogenes*, *Mycoplasma bovis*, *Arcanobacterium pyogenes*, *Mycoplasma dispar*, *Ureaplasma diversum*, and *Mycoplasma bovirhinis*) proliferate in the URT and invade the lung via inhalation under specific conditions (Griffin et al., 2010). Likewise, the URT of clinically asymptomatic pigs often harbors pathobionts such *Mycoplasma hyopneumoniae*, *Haemophilus parasuis*, *Actinobacillus pleuropneumoniae*, *Actinobacillus suis*, *Pasteurella multocida*, *Bordetella bronchiseptica* and *Streptococcus suis* (Luehrs et al., 2017; Vötsch et al., 2018) that can all prone to disease. In dogs, many potential pathogens involved with CIRDC, e.g., *Mycoplasma cynos*, *Bordetella bronchiseptica* and *Streptococcus* spp. are common members of the respiratory microbiota (Day et al., 2020). Similarly, the URT is also a reservoir of opportunistic pathogens in domestic birds. Potential respiratory pathogens, namely *Avibacterium*, *Gallibacterium*, *Mycoplasma*, and *Ornithobacterium*, are found in the URT of turkeys (Kursa et al., 2021).

The next complexity that we can bring in is that the modifications of the airway microbiota resulting from primary virus infections often act as a disturbance for secondary infections (Zeineldin et al., 2019). The one pathogen-one disease paradigm is often insufficient to elucidate many of the respiratory diseases (Vayssier-Taussat et al., 2014; Vayssier-Taussat et al., 2015).

A plethora of examples in swine illustrates the framework in which co-infection occurs. Swine influenza virus enhances the morbidity of *Streptococcus suis* infection by decreasing mucociliary clearance, damaging epithelial cells, and by facilitating its adherence, colonization and invasion in the lungs (Meng et al., 2016). Swine influenza virus also compensates for the lack of suliyisin (cytotoxic protein secreted by *Streptococcus suis*) in the adherence and invasion process of suliyisin-negative *Streptococcus suis* (Meng et al., 2019). Similarly, a synergism between nasal *Staphylococcus aureus* and pathobionts such as *Pasteurella multocida* and *Klebsiella* spp. have also been reported in pigs (Espinosa-Gongora et al., 2016). Co-occurrence between the porcine reproductive and respiratory syndrome virus (PRRSV), *Haemophilus parasuis* and *Mycoplasma hyorhinis* in pig lungs is repeatedly observed (Jiang et al., 2019). Contrastingly, the virulence of one pathogen may be reduced by co-infection with another, as it has been shown for the bacterium *Escherichia/Shigella* in the respiratory tract of weaning piglets, which inhibited the virulence of *Haemophilus parasuis* or the presence of *Phascolarctobacterium* in the respiratory tract (Mahmmod et al., 2020). On the host side, respiratory pathogens can also modulate

innate and adaptive immune responses (e.g. weakening monocyte activity, suppressing phagocytic capacity of alveolar macrophages, among others) that therefore promote secondary bacterial infection (Zeineldin et al., 2019). For example, PRRSV infection of bone marrow-derived dendritic cells regulated the innate immune system by partially acting on the phagocytosis of *S. suis*, but mainly by modulating the development of an exacerbated inflammatory response (Auray et al., 2016). Similarly, *in vitro* co-infection of swine epithelial cells with *S. suis* and swine influenza virus showed an important synergy between the two pathogens regarding the up-regulation of genes coding for inflammatory mediators (Dang et al., 2014). Cells co-infected with PRRSV and porcine circovirus produced more TNF than cells infected with just one of the pathogens (Shi et al., 2010), whereas an exacerbated inflammatory response was described in pigs co-infected with PRRSV and *Mycoplasma hyopneumoniae* (Thanawongnuwech et al., 2004).

Another case in point are the low pathogenic avian influenza viruses (LPAIV) that, during outbreaks, even in the antibiotic era, are coupled to co-infections by pathogens such as *Mycoplasma gallisepticum*, *Mycoplasma synoviae*, *Ornithobacterium rhinotracheale*, avian pathogenic *Escherichia coli* (APEC) and *Staphylococcus aureus*, which are responsible for a higher mortality rate (Much et al., 2002; Belkasmi et al., 2020) and a marked reduction in egg production of laying hens (Umar et al., 2016). Moreover, it appears as though that concurrent pathogens such as infectious bronchitis virus (IBV), the APEC, the avian pneumovirus and the LPAIV act synergistically or cumulatively to mediate complex infectious processes (Awad et al., 2014; Guabiraba and Schouler, 2015; Patel et al., 2018).

## THE GUT-LUNG AXIS DURING RESPIRATORY DISEASES: KEY TO UNDERSTANDING HOLOBIONT FUNCTIONING

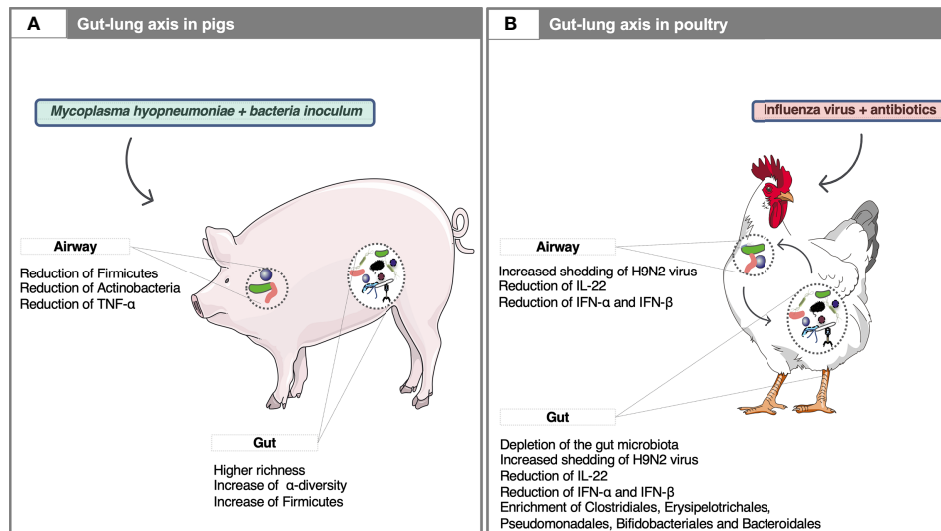
The pathophysiology of the complex respiratory diseases in domestic animals seems more complex than previously assumed. It has been recently posited that respiratory comorbidities might be partly modulated by the bidirectional inter-organ communication with the gastrointestinal tract, referred to as the gut-lung axis (Dang and Marsland, 2019). This new field of research is now investigating how gut microbiota modulates the onset of respiratory infections (Niederwerder, 2017) and whether the airway microbiota in turn, influence host epithelial and immune cells to adjust inflammatory responses at distal sites such gut (Budden et al., 2017; Wypych et al., 2019).

The gut of ruminants (Ramayo-Caldas et al., 2019), pigs (Mach et al., 2015; Ramayo-Caldas et al., 2016), horses (Mach et al., 2020; Mach et al., 2021a), companion animals (Alessandri et al., 2020) and chickens (Rychlik, 2020) accommodates a complex community of microorganisms that live in a commensal relationship with their hosts and provide significant contributions to the metabolism and immune

responses. It might be hypothesized that in symbiosis, gut microbiota ensures the uptake of microbial metabolites that favorably affects the immune and endocrine pathways involved in respiratory disease and progression, whereas the lungs maintain inflammatory homeostasis in the gut by controlling immune response. Even if the mechanisms bridging gut microbiota with the alterations of respiratory disease outcome are still poorly understood in domestic animals, a growing body of research in swine supports that the crosstalk between the gut and airways microbiome. The gut microbiome composition in growing pigs following an exposure to a non-pathogenic oral microbial inoculum modulated microbial communities and was beneficial upon challenge with *Mycoplasma hyopneumoniae* (Schachtschneider et al., 2013). Similarly, the gut microbiome diversity and composition in piglets determined the respiratory disease progression in pigs after *M. hyopneumoniae* inoculation [Surendran Nair et al. (2019); **Figure 3A**]. Likewise, increased fecal microbiome diversity was associated with improved health outcomes after experimental co-infection with PRRSV and porcine circovirus type 2 (PCV2) (Niederwerder et al., 2016; Ober et al., 2017; Niederwerder et al., 2018).

Several recent studies in domestic birds have also tackled this broader question of the gut-lung axis. Antibiotic treatment in day-old layer chickens resulted in a significant depletion of the gut microbiota, a down-regulation of the type I interferon response and higher shedding of LPAIV of H9N2 subtype in the oropharynx and cloacal section (Yitbarek et al., 2018a). A similar study (Yitbarek et al., 2018b) reported that depletion of the gut microbiota in young chickens using antibiotic treatment increased both oropharyngeal and cloacal levels of the LPAIV of H9N2 subtype and decreased the expression of IFN- $\alpha$  and IFN- $\beta$  in the gut and respiratory tract (**Figure 3B**). Likewise, depletion of gut microbiota increased the severity of Marek's disease in infected chickens, coupled to an increase in the transcription of IFN- $\alpha$ , IFN- $\beta$  and IFN- $\gamma$  in the bursa of *Fabricius* at four days post-infection (Bavananthasivam et al., 2021). In another study developed in chickens, *Lactobacillus salivarius* intake alleviated lung inflammation injury caused by *Mycoplasma gallisepticum* infection and increased host defense against *Escherichia coli* by improved gut microbiota composition (Wang et al., 2021). Antibiotic-treated ducks had increased levels of intestinal highly pathogenic avian influenza virus (HPAIV) of H5N9 subtype associated with a reduced antiviral immune response, but no higher viral titers in the respiratory tract (Figueroa et al., 2020). Finally, the oral administration of *E. coli* O86:B7 expressing high levels of galactose- $\alpha$ -1,3-galactose ( $\alpha$ -Gal) in turkeys was found to abrogate anti- $\alpha$ -Gal IgA response in lungs and to protect against experimental *Aspergillus fumigatus* infection (Mateos Hernández et al., 2020). The authors suggested that gut microbiota generates  $\alpha$ -Gal-specific antigen-specific regulatory T cells (Tregs) in the gut, which can then migrate to the lungs, promoting disease tolerance which prevents the systemic upregulation of pro-inflammatory cytokines (Mateos Hernández et al., 2020).

As outlined above, knowledge regarding the crosstalk mechanisms by which the airway microbiota affects immune



**FIGURE 3** | Examples of the gut-lung axis cross-talk in domestic animals. **(A)** The gut microbiota composition affects systemic immune responses and lowers the severity of *M. hyopneumoniae* infection in the respiratory tract of pigs. For example, the administration of oral microbial inoculum increased Firmicutes such as *Roseburia*, *Barnesiella*, *Blautia*, *Dorea* in the gut and reduced the relative abundance of Actinobacteria and Firmicutes phyla in the airways microbiota, as well as the levels of TNF- $\alpha$  in the lungs and their lesions (Schachtschneider et al., 2013). Young pigs with low microbial diversity in the gut showed severe lung lesions on exposure to *M. hyopneumoniae*, whereas the opposite trend was observed in piglets with higher SCFA producing taxa in the gut (e.g., *Ruminococcus*, *Prevotella*, *Ruminiclostridium*, and *Oscillospira*) (Surendran Nair et al., 2019). In addition to having local effects in the gut, SCFAs enter the circulation, modulate bone-marrow hematopoiesis and thereby can promote regulatory or pro-inflammatory responses in the lung (Surendran Nair et al., 2019); **(B)** In H9N2 subtype LPAIV infected chickens, microbiota gut depletion using antibiotics have shown to reduce IFN response, which plays an important role in innate responses to viral infections in the gut and airways and increase the influenza virus shedding from the upper respiratory and gastrointestinal tracts (Yitbarek et al., 2018a). Along with IFNs, treatment of chickens with antibiotics for 12 days resulted in reduced interleukin 22 expression in the respiratory tract after gut microbiota depletion and in enrichment with class Erysipelotrichia, Bacteroidia and with order Clostridiales, Erysipelotrichales, Pseudomonadales, Bifidobacteriales and Bacteroidales (Yitbarek et al., 2018b). Written permission for publication of the chicken drawing in the figure has been taken. The pictures of the pig, bacteria and viruses were downloaded from smart Servier Medical Art <https://smart.servier.com> without changes. Servier Medical Art by Servier is licensed under a Creative Commons Attribution 3.0 License.

response in the gut is scant (Brown et al., 2017; Mazel-Sanchez et al., 2019). It is postulated that gut microbiota can reach occasionally the lungs through aspiration of vomit or esophageal reflux, therefore exposing pattern-recognition receptors (PRRs) expressed by host cells to peptidoglycans or lipopolysaccharides and stimulating the immune response. Alternatively, these microbe-associated molecular patterns can be transported *via* the circulation together with other gut-derived microbial metabolites known to have immunomodulatory properties such as short-chain fatty acids (SCFA) or indole derivatives (Maslowski et al., 2009; Wypych et al., 2019). Avian influenza virus infections have shown to alter the intestinal microbiota in domestic poultry and aquatic birds (Zhao et al., 2018). However, these alterations are likely the outcome of the viral replication in the avian intestinal tract, rather than a cross-talk between respiratory and intestinal mucosal surfaces.

## THE ZONOTIC POTENTIAL OF INFLUENZA A VIRUSES: A CRAWLING THREAT

Because species of veterinary interest are in regular and increasingly close contact with humans, especially in

developing countries, understanding the environmental and host determinants that allow pathogens to develop in the respiratory system of livestock and occasionally to be transmitted to and cause diseases in humans is also a relevant research questions today (Peiris et al., 2012). Influenza A viruses, which are commonly found during the onset of complex respiratory diseases in livestock, highlight the problem of devastating zoonotic infections that can arise from veterinary species such as avian and mammalian animal hosts (Kuiken et al., 2005; Mostafa et al., 2018). For instance, in one study involving 125 farrow-to-finish pig hirs in France, Influenza A viruses of H1N1 subtype was reportedly found during respiratory disorders (Fablet et al., 2012). Of particular concern is the emergence of the LPAIV of H7Nx subtypes infecting humans, which are frequently reported in commercial poultry, backyards and live bird markets in numerous countries in Europe, Asia and Africa (Abdelwhab et al., 2014; Richard et al., 2014). While H7N2, H7N3 and H7N7 subtypes LPAIV occasionally infect humans causing mild, if any, clinical manifestations (Gao et al., 2013), the emerged H7N9 subtype LPAIV in China causes a high mortality rate in humans (40%) and have pandemic potential (Horman et al., 2018). Additionally, a growing threat to both animal and human health is emerging from the H9N2 subtype LPAIV, endemic in



poultry in many areas of Eurasia and Africa (Peacock et al., 2019).

## CONCLUSIONS

The last decade has brought substantial insights into the characterization of the respiratory microbiota in species of veterinary interest. Sterility of the airways has been rejected. The mechanisms behind the colonization of the respiratory system in the different species have started to be explored, as well as the microbial communities occupying the different niches in the airways. Yet nothing is known about the role eukaryotes, archaea and phages play in the respiratory tract in domestic animals.

Collectively, the results obtained in the last years suggest some degree of convergence in the airway respiratory microbiota of mammals. At the phylum level, mammals harbor Firmicutes, Bacteroidetes and Proteobacteria in the upper and lower respiratory system. This is juxtaposed with domestic birds, in which we observed a reduction of Bacteroidetes but increased abundance of Proteobacteria, sustaining the host-specific microbe associations. Although these differences are not yet fully understood, the hypothesis that birds are merely “feathered” mammals with a few specific differences related to flight and production of large yolky eggs is an erroneous assumption. Therefore, not all of the results obtained on one species can be directly translated to the other ones.

The increase in the prevalence of respiratory disorders in farm and companion animals have reached epidemic statistical levels in wealthy countries that generate a greater need to monitor and control their impact on morbidity and mortality. The increasing demands for livestock products coupled to the intensification of marked-oriented systems in many countries, climate change, ecology incursion, as well as movements of people, animals and products increase the risk of complex respiratory disease development worldwide (Kuiken et al., 2005), even if data for regions in South America, Asia, Middle-East and Africa is scant. The prevention and treatment of respiratory diseases are of paramount importance, not only because they can induce immunosuppression and render animals susceptible to opportunistic infections, but also because it may increase the risk of cross-species transmissions, foodborne and zoonotic diseases.

Disorders in the respiratory tract are compatible with a “ménage à quatre” situation, as they depend on the intricate interactions between the pathogen(s), multiple symbionts of the respiratory tract, the host response and the environmental conditions. There is evidence showing that various environmental factors such as abrupt weaning, antibiotic administration and housing management often disrupt the respiratory microbial ecosystem and increase susceptibility to infections in animal species used for food. Under unfavorable environmental conditions, the symbionts themselves can act as opportunistic pathogens or not provide the same degree of protection. An example hereof is that the respiratory disease

complex in bovine, porcine and commercial birds often imply pathobionts. Pigs, ruminants and birds, which frequently switch environments through their intense production lives, are more prone to exhibit respiratory disorders. Modulation of husbandry environment may facilitate the manipulation of the pathobiotic systems. Beyond farm animals, urbanized lifestyle, featuring restricted animal contact and house confinement promotes changes in airway microbiota in both companion animals and horses.

The commensal microbiota composition may affect the pathogen infectivity or the expansion of pathobionts beyond a level of tolerance, either directly *via* the secretion of molecules with antimicrobial activity or indirectly *via* immune-mediated modifications. Of the remaining questions are: which taxa or mixtures of these, if any, make animals more susceptible or resistant to respiratory disorders in each of the domestic species. Thus far, data in mammals have identified members of the *Lactobacillaceae* family as key protective hub taxa in the respiratory tract. Conversely, the expansion in Proteobacteria coupled with a drop in Firmicutes and in diversity has been associated with a pathobiome context in cows, pigs and horses. Proteobacteria seem benign when they are in minor proportion, whereas, under the onset of disease, they trigger inflammatory responses.

The blooming of Proteobacteria could be a potential biomarker for respiratory disease states in mammals and birds, however, trends and nuances are noted. Species within the Proteobacteria phylum have a protective role in immune response against infection or inflammation, as shown in companionship animals. Therefore, one can speculate that bacteria from the phylum Proteobacteria have as yet unidentified functions, and so a better understanding of such functions is key to identifying their symbiotic or their pathological relations with hosts.

To date, re-establishment of respiratory eubiosis after infections and preventive treatment with probiotics has not been demonstrated in domestic animals. Nonetheless, manipulation of gut microbiota in swine and domestic birds has shown to shape their immune response and interfere with the course of respiratory disease. The existence of a gut-lung axis paves the way for new approaches in the management of respiratory disease in domestic animals.

## FUTURE PERSPECTIVES

Despite significant efforts towards understanding the link between the pathogen(s), the commensal host-microbiota and the environment, this research field has just started to explore the mechanisms by which pathobiome disrupts respiratory homeostasis in animals. An improved understanding of the spatial distribution and temporal changes from a commensal microbiota to a pathobiome state would provide valuable information (biomarkers) to implement preventive measures or treatments, before observation of clinical signs or symptoms. Both symbiome and pathobiome vary over time and between individuals. Thus, longitudinal studies focusing

on the aforementioned quadrangular interactions are needed to understand the role that the symbiome and pathobiome may have in agent-associated disease outbreaks, as well as the forces at play. These should concentrate on gaining a better understanding of the shifts of the microbiota composition and function in different sections of the respiratory and gastrointestinal systems, and whether the microbiota shifts occur before, and cause a change in respiratory health status, or if they are a consequence of a change in health due to a different host and environmental factors. Expanding shotgun metagenomic sequencing and functional metagenomics (e.g., metatranscriptomics, metaproteomics, metabolomics) of the respiratory tract is required to address this important issue. Advances in sequencing and database annotation are critical for further advances in the field that have the potential to optimize the methods to mitigate diseases, including those related to emerging zoonotic pathogens.

## REFERENCES

- Abdelwhab E. M., Veits J., and Mettenleiter T. C. (2014). Prevalence and Control of H7 Avian Influenza Viruses in Birds and Humans. *Epidemiol. Infect.* 142, 896–920. doi: 10.1017/S0950268813003324
- Abundo M. E., Ngunjiri J. M., Taylor K. J. M., Ji H., Ghorbani A., Kc M., et al. (2020). Evaluation of Sampling Methods for the Study of Avian Respiratory Microbiota. *Avian Dis.* 64, 277–285. doi: 10.1637/aviandiseases-D-19-00200
- Abundo M. E. C., Ngunjiri J. M., Taylor K. J. M., Ji H., Ghorbani A., Mahesh K. C., et al. (2021). Assessment of Two DNA Extraction Kits for Profiling Poultry Respiratory Microbiota From Multiple Sample Types. *PLoS One* 16, e0241732. doi: 10.1101/2020.10.21.348508
- Alessandri G., Argentini C., Milani C., Turrone F., Cristina Ossiprandi M., van Sinderen D., et al. (2020). Catching a Glimpse of the Bacterial Gut Community of Companion Animals: A Canine and Feline Perspective. *Microb. Biotechnol.* 13, 1708–1732. doi: 10.1111/1751-7915.13656
- Amat S., Holman D. B., Timsit E., Schwinghamer T., and Alexander T. W. (2019). Evaluation of the Nasopharyngeal Microbiota in Beef Cattle Transported to a Feedlot, With a Focus on Lactic Acid-Producing Bacteria. *Front. Microbiol.* 10, 1988–2003. doi: 10.3389/fmicb.2019.01988
- Amat S., Subramanian S., Timsit E., and Alexander T. W. (2017). Probiotic Bacteria Inhibit the Bovine Respiratory Pathogen Mannheimia Haemolytica Serotype 1 In Vitro. *Lett. Appl. Microbiol.* 64, 343–349. doi: 10.1111/lam.12723
- Auray G., Lachance C., Wang Y., Gagnon C. A., Segura M., and Gottschalk M. (2016). Transcriptional Analysis of PRRSV-infected Porcine Dendritic Cell Response to Streptococcus Suis Infection Reveals Up-Regulation of Inflammatory-Related Genes Expression. *PLoS One* 11, e0156019. doi: 10.1371/journal.pone.0156019
- Awad F., Chhabra R., Baylis M., and Ganapathy K. (2014). An Overview of Infectious Bronchitis Virus in Chickens. *Worlds Poult. Sci. J.* 70, 375–384. doi: 10.1017/S0043933914000385
- Bass D., Stentiford G. D., Wang H. C., Koskella B., and Tyler C. R. (2019). The Pathobiome in Animal and Plant Diseases. *Trends Ecol. Evol.* 34, 996–1008. doi: 10.1016/j.tree.2019.07.012
- Bavananthasivam J., Astill J., Matsuyama-Kato A., Taha-Abdelaziz K., Shojadoost B., and Sharif S. (2021). Gut Microbiota is Associated With Protection Against Marek's Disease Virus Infection in Chickens. *Virology* 553, 122–130. doi: 10.1016/j.virol.2020.10.011
- Belkasmı S. F. Z., Fellahi S., Touzani C. D., Faraji F. Z., Maaroufi I., Delverdi M., et al. (2020). Co-Infections of Chickens With Avian Influenza Virus H9N2 and Moroccan Italy 02 Infectious Bronchitis Virus: Effect on Pathogenesis and Protection Conferred by Different Vaccination Programmes. *Avian Pathol.* 49, 21–28. doi: 10.1080/03079457.2019.1656328
- Bernardo-Cravo A. P., Schmeller D. S., Chatzinotas A., Vredenburg V. T., and Loyau A. (2020). Environmental Factors and Host Microbiomes Shape Host-Pathogen Dynamics. *Trends Parasitol.* 36, 616–633. doi: 10.1016/j.pt.2020.04.010

## AUTHOR CONTRIBUTIONS

NM designed the structure of this review and wrote the manuscript. EB, LN, and CC have contributed to the reviewing and editing of the manuscript. All authors contributed to the article and approved the submitted version.

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- Bernhard W., Haslam P. L., and Floros J. (2004). From Birds to Humans: New Concepts on Airways Relative to Alveolar Surfactant. *Am. J. Respir. Cell Mol. Biol.* 30, 6–11. doi: 10.1165/rcmb.2003-0158TR
- Beste K. J., Lawhon S. D., Chamoun-Emanuelli A. M., Duff A. H., Coleman M. C., Griffin C. E., et al. (2019). Culture-Independent and Dependent Evaluation of the Equine Paranasal Sinus Microbiota in Health and Disease. *Equine Vet. J.* 0, 1–9. doi: 10.1111/evj.13168
- Blakebrough-Hall C., McMeniman J. P., and González L. A. (2020). An Evaluation of the Economic Effects of Bovine Respiratory Disease on Animal Performance, Carcass Traits, and Economic Outcomes in Feedlot Cattle Defined Using Four BRD Diagnosis Methods. *J. Anim. Sci.* 98, skaa005. doi: 10.1093/jas/skaa005
- Bond S. L., Timsit E., Workentine M., Alexander T., and Léguillette R. (2017). Upper and Lower Respiratory Tract Microbiota in Horses: Bacterial Communities Associated With Health and Mild Asthma (Inflammatory Airway Disease) and Effects of Dexamethasone. *BMC Microbiol.* 17, 1–11. doi: 10.1186/s12866-017-1092-5
- Bond S. L., Workentine M., Hundt J., Gilkerson J. R., and Léguillette R. (2020). Effects of Nebulized Dexamethasone on the Respiratory Microbiota and Mycobacteria and Relative Equine herpesvirus-1, 2, 4, 5 in an Equine Model of Asthma. *J. Vet. Intern. Med.* 34, 307–321. doi: 10.1111/jvim.15671
- Bradley P. H., and Pollard K. S. (2017). Proteobacteria Explain Significant Functional Variability in the Human Gut Microbiome. *Microbiome* 5 (1) 36–59. doi: 10.1186/s40168-017-0244-z
- Brown R. E., Brain J. D., and Wang N. (1997). The Avian Respiratory System: A Unique Model for Studies of Respiratory Toxicosis and for Monitoring Air Quality. *Environ. Heal.* 105, 188–200. doi: 10.1289/ehp.97105188
- Brown R. L., Sequeira R. P., and Clarke T. B. (2017). The Microbiota Protects Against Respiratory Infection Via GM-CSF Signaling. *Nat. Commun.* 8, 1512–1523. doi: 10.1038/s41467-017-01803-x
- Brugman S., Ikeda-Ohtsubo W., Braber S., Folkerts G., Pieterse C. M. J., and Bakker P. A. H. M. (2018). A Comparative Review on Microbiota Manipulation: Lessons From Fish, Plants, Livestock, and Human Research. *Front. Nutr.* 5, 80. doi: 10.3389/fnut.2018.00080
- Budden K. F., Gellatly S. L., Wood D. L. A., Cooper M. A., Morrison M., Hugenholtz P., et al. (2017). Emerging Pathogenic Links Between Microbiota and the Gut-Lung Axis. *Nat. Rev. Microbiol.* 15, 55. doi: 10.1038/nrmicro.2016.142
- Budden K. F., Shukla S. D., Rehman S. F., Bowerman K. L., Keely S., Hugenholtz P., et al. (2019). Functional Effects of the Microbiota in Chronic Respiratory Disease. *Lancet Respir. Med.* 7, 907–920. doi: 10.1016/S2213-2600(18)30510-1
- Cáliz J., Triadó-Margarit X., Camarero L., and Casamayor E. O. (2018). A Long-Term Survey Unveils Strong Seasonal Patterns in the Airborne Microbiome Coupled to General and Regional Atmospheric Circulations. *Proc. Natl. Acad. Sci. U. S. A.* 115, 12229–12234. doi: 10.1073/pnas.1812826115
- Collie D., Glendinning L., Govan J., Wright S., Thornton E., Tennant P., et al. (2015). Lung Microbiota Changes Associated With Chronic Pseudomonas Aeruginosa Lung Infection and the Impact of Intravenous Colistimethate Sodium. *PLoS One* 82, 3225–3232. doi: 10.1371/journal.pone.0142097

- Correa-Fiz F., Fraile L., and Aragon V. (2016). Piglet Nasal Microbiota at Weaning may Influence the Development of Glässer's Disease During the Rearing Period. *BMC Genomics* 17, 404–418. doi: 10.1186/s12864-016-2700-8
- Correa-Fiz F., Gonçalves dos Santos J. M., Illas F., and Aragon V. (2019). Antimicrobial Removal on Piglets Promotes Health and Higher Bacterial Diversity in the Nasal Microbiota. *Sci. Rep.* 9, 6545–6554. doi: 10.1038/s41598-019-43022-y
- Cortes L. C. P., LeVeque R. M., Funk J. A., Marsh T. L., and Mulks M. H. (2018). Development of the Tonsil Microbiome in Pigs and Effects of Stress on the Microbiome. *Front. Vet. Sci.* 5, 220. doi: 10.3389/fvets.2018.00220
- Dang Y., Lachance C., Wang Y., Gagnon C. A., Savard C., Segura M., et al. (2014). Transcriptional Approach to Study Porcine Tracheal Epithelial Cells Individually or Dually Infected With Swine Influenza Virus and Streptococcus Suis. *BMC Vet. Res.* 10, 86. doi: 10.1186/1746-6148-10-86
- Dang A. T., and Marsland B. J. (2019). Microbes, Metabolites, and the Gut–Lung Axis. *Mucosal Immunol.* 12, 843–850. doi: 10.1038/s41385-019-0160-6
- Day M. J., Carey S., Clercx C., Kohn B., Marsilio F., Thiry E., et al. (2020). Aetiology of Canine Infectious Respiratory Disease Complex and Prevalence of its Pathogens in Europe. *J. Comp. Pathol.* 176, 86–108. doi: 10.1016/j.jcpa.2020.02.005
- Dorn E. S., Tress B., Suchodolski J. S., Nisar T., Ravindran P., Weber K., et al. (2017). Bacterial Microbiome in the Nose of Healthy Cats and in Cats With Nasal Disease. *PLoS One* 12, 1–23. doi: 10.1371/journal.pone.0180299
- Ericsson A. C., Personett A. R., Grobman M. E., Rindt H., and Reinero C. R. (2016). Composition and Predicted Metabolic Capacity of Upper and Lower Airway Microbiota of Healthy Dogs in Relation to the Fecal Microbiota. *PLoS One* 11, e0154646. doi: 10.1371/journal.pone.0154646
- Ericsson A. C., Personett A. R., Rindt H., Grobman M. E., and Reinero C. R. (2020). Respiratory Dysbiosis and Population-Wide Temporal Dynamics in Canine Chronic Bronchitis and non-Inflammatory Respiratory Disease. *PLoS One* 15, 1–18. doi: 10.1371/journal.pone.0228085
- Espinosa-Gongora C., Larsen N., Schønning K., Fredholm M., and Guardabassi L. (2016). Differential Analysis of the Nasal Microbiome of Pig Carriers or non-Carriers of Staphylococcus Aureus. *PLoS One* 11, e0160331. doi: 10.1371/journal.pone.0160331
- Fablet C., Marois-Créhan C., Simon G., Grasland B., Jestin A., Kobisch M., et al. (2012). Infectious Agents Associated With Respiratory Diseases in 125 Farrow-to-Finish Pig Herds: A Cross-Sectional Study. *Vet. Microbiol.* 157, 152–163. doi: 10.1016/j.vetmic.2011.12.015
- Fastrès A., Canonne M. A., Taminiau B., Billen F., Garigliany M. M., Daube G., et al. (2020a). Analysis of the Lung Microbiota in Dogs With Bordetella Bronchiseptica Infection and Correlation With Culture and Quantitative Polymerase Chain Reaction. *Vet. Res.* 51, 46. doi: 10.1186/s13567-020-00769-x
- Fastrès A., Taminiau B., Vangrinsven E., Tutunaru A. C., Moysse E., Farnir F., et al. (2019). Effect of an Antimicrobial Drug on Lung Microbiota in Healthy Dogs. *Heliyon* 5, e02802. doi: 10.1016/j.heliyon.2019.e02802
- Fastrès A., Vangrinsven E., Taminiau B., Tutunaru A.-C., Jabri H., Daube G., et al. (2020b). Assessment of Lung Microbiota in Healthy Dogs: Influence of the Type of Breed, Living Conditions and Canine Idiopathic Pulmonary Fibrosis. *BMC Microbiol.* 20, 84. doi: 10.1186/s12866-020-01784-w
- Figueroa T., Bessière P., Coggon A., Bouwman K. M., van der Woude R., Delverdier M., et al. (2020). The microbiota Contributes to the Control of Highly Pathogenic H5N9 Influenza Virus Replication in Ducks. *J. Virol.* 94, e00289–e00220. doi: 10.1128/jvi.00289-20
- Fillion-Bertrand G., Dickson R. P., Boivin R., Lavoie J. P., Huffnagle G. B., and Leclere M. (2019). Lung Microbiome is Influenced by the Environment and Asthmatic Status in an Equine Model of Asthma. *Am. J. Respir. Cell Mol. Biol.* 60, 189–197. doi: 10.1165/rcmb.2017-0228OC
- Gaeta N. C., Lima S. F., Teixeira A. G., Ganda E. K., Oikonomou G., Gregory L., et al. (2017). Deciphering Upper Respiratory Tract Microbiota Complexity in Healthy Calves and Calves That Develop Respiratory Disease Using Shotgun Metagenomics. *J. Dairy Sci.* 100, 1445–1458. doi: 10.3168/jds.2016-11522
- Gao R., Cao B., Hu Y., Feng Z., Wang D., Hu W., et al. (2013). Human Infection With a Novel Avian-Origin Influenza A (H7N9) Virus. *N. Engl. J. Med.* 368, 1888–1897. doi: 10.1056/nejmoa1304459
- Glendinning L., Collie D., Wright S., Rutherford K. M. D., and McLachlan G. (2017a). Comparing Microbiotas in the Upper Aerodigestive and Lower Respiratory Tracts of Lambs. *Microbiome* 5, 145. doi: 10.1186/s40168-017-0364-5
- Glendinning L., McLachlan G., and Vervelde L. (2017b). Age-Related Differences in the Respiratory Microbiota of Chickens. *PLoS One* 12, e0188455. doi: 10.1371/journal.pone.0188455
- Glendinning L., Wright S., Pollock J., Tennant P., Collie D., and McLachlan G. (2016). Variability of the Sheep Lung Microbiota. *Appl. Environ. Microbiol.* 82, 3225–3238. doi: 10.1128/AEM.00540-16
- Gomez D. E., Arroyo L. G., Lillie B., and Scott Weese J. (2021). Nasal Bacterial Microbiota During an Outbreak of Equine Herpesvirus 1 at a Farm in Southern Ontario. *Can. J. Vet. Res.* 85, 3–11.
- Griffin D., Chengappa M. M., Kuszak J., and McVey D. S. (2010). Bacterial Pathogens of the Bovine Respiratory Disease Complex. *Vet. Clin. North Am. - Food Anim. Pract.* 26, 381–394. doi: 10.1016/j.cvfa.2010.04.004
- Guabiraba R., and Schouler C. (2015). Avian Colibacillosis: Still Many Black Holes. *FEMS Microbiol. Lett.* 362, 1–8. doi: 10.1093/femsle/fnv118
- Hall J. A., Isaiah A., Estill C. T., Pirelli G. J., and Suchodolski J. S. (2017). Weaned Beef Calves Fed Selenium-Biofortified Alfalfa Hay Have an Enriched Nasal Microbiota Compared With Healthy Controls. *PLoS One* 12, e0179217. doi: 10.1371/journal.pone.0179215
- Hansen M. S., Pors S. E., Jensen H. E., Bille-Hansen V., Bisgaard M., Flachs E. M., et al. (2010). An Investigation of the Pathology and Pathogens Associated With Porcine Respiratory Disease Complex in Denmark. *J. Comp. Pathol.* 143, 120–131. doi: 10.1016/j.jcpa.2010.01.012
- Harms P. A., Halbur P. G., and Sorden S. D. (2002). Three Cases of Porcine Respiratory Disease Complex Associated With Porcine Circovirus Type 2 Infection. *J. Swine Heal. Prod.* 10, 27–30.
- Heidari M., Hamir A., Cutlip R. C., and Brogden K. A. (2002). Antimicrobial Anionic Peptide Binds In Vivo to Mannheimia (Pasteurella) Haemolytica Attached to Ovine Alveolar Epithelium. *Int. J. Antimicrob. Agents* 20, 69–72. doi: 10.1016/S0924-8579(02)00048-1
- Hilton W. M. (2014). BRD in 2014: Where Have We Been, Where are We Now, and Where do We Want to Go? *Anim. Heal. Res. Rev.* 151, 120–122. doi: 10.1017/S1466252314000115
- Holman D. B., McAllister T. A., Topp E., Wright A. D. G., and Alexander T. W. (2015). The Nasopharyngeal Microbiota of Feedlot Cattle That Develop Bovine Respiratory Disease. *Vet. Microbiol.* 180, 90–95. doi: 10.1016/j.vetmic.2015.07.031
- Holman D. B., Timsit E., Amat S., Abbott D. W., Buret A. G., and Alexander T. W. (2017). The Nasopharyngeal Microbiota of Beef Cattle Before and After Transport to a Feedlot. *BMC Microbiol.* 17, 1–12. doi: 10.1186/s12866-017-0978-6
- Holman D. B., Timsit E., Booker C. W., and Alexander T. W. (2018). Injectable Antimicrobials in Commercial Feedlot Cattle and Their Effect on the Nasopharyngeal Microbiota and Antimicrobial Resistance. *Vet. Microbiol.* 214, 140–147. doi: 10.1016/j.vetmic.2017.12.015
- Holman D. B., Yang W., and Alexander T. W. (2019). Antibiotic Treatment in Feedlot Cattle: A Longitudinal Study of the Effect of Oxytetracycline and Tulathromycin on the Fecal and Nasopharyngeal Microbiota. *Microbiome* 7, 1–14. doi: 10.1186/s40168-019-0696-4
- Holt H. R., Alarcon P., Velasova M., Pfeiffer D. U., and Wieland B. (2011). Bpex Pig Health Scheme: A Useful Monitoring System for Respiratory Disease Control in Pig Farms? *BMC Vet. Res.* 7, 82. doi: 10.1186/1746-6148-7-82
- Horman W. S. J., Nguyen T. H. O., Kedzierska K., Bean A. G. D., and Layton D. S. (2018). The Drivers of Pathology in Zoonotic Avian Influenza: The Interplay Between Host and Pathogen. *Front. Immunol.* 9, 1812. doi: 10.3389/fimmu.2018.01812
- Huang T., Zhang M., Tong X., Chen J., Yan G., Fang S., et al. (2019). Microbial Communities in Swine Lungs and Their Association With Lung Lesions. *Microb. Biotechnol.* 12, 289–304. doi: 10.1111/1751-7915.13353
- Huffnagle G. B., Dickson R. P., and Lukacs N. W. (2017). The Respiratory Tract Microbiome and Lung Inflammation: A Two-Way Street. *Mucosal Immunol.* 10, 299. doi: 10.1038/mi.2016.108
- Jakobsen A. M., Bahl M. I., Buschhardt T., Hansen T. B., Al-Soud W. A., Breyndrod A. D., et al. (2019). Bacterial Community Analysis for Investigating Bacterial Transfer From Tonsils to the Pig Carcass. *Int. J. Food Microbiol.* 295, 8–18. doi: 10.1016/j.ijfoodmicro.2019.02.003
- Jiang N., Liu H., Wang P., Huang J., Han H., and Wang Q. (2019). Illumina MiSeq Sequencing Investigation of Microbiota in Bronchoalveolar Lavage Fluid and

- Cecum of the Swine Infected With PRRSV. *Curr. Microbiol.* 76, 222–230. doi: 10.1007/s00284-018-1613-y
- Johnson T. J., Youmans B. P., Noll S., Cardona C., Evans N. P., Peter Karnezos T., et al. (2018). A Consistent and Predictable Commercial Broiler Chicken Bacterial Microbiota in Antibiotic-Free Production Displays Strong Correlations With Performance. *Appl. Environ. Microbiol.* 84, 1–18. doi: 10.1128/AEM.00362-18
- Kauter A., Epping L., Semmler T., Antao E.-M., Kannapin D., Stoeckle S. D., et al. (2019). The Gut Microbiome of Horses: Current Research on Equine Enteric Microbiota and Future Perspectives. *Anim. Microbiome* 1, 1–15. doi: 10.1186/s42523-019-0013-3
- Kers J. G., Velkers F. C., Fischer E. A. J., Hermes G. D. A., Stegeman J. A., and Smidt H. (2018). Host and Environmental Factors Affecting the Intestinal Microbiota in Chickens. *Front. Microbiol.* 9, 235. doi: 10.3389/fmicb.2018.00235
- Klima C. L., Holman D. B., Ralston B. J., Stanford K., Zaheer R., Alexander T. W., et al. (2019). Lower Respiratory Tract Microbiome and Resistome of Bovine Respiratory Disease Mortalities. *Microb. Ecol.* 78, 446–456. doi: 10.1007/s00248-019-01361-3
- Kubasova T., Kollarcikova M., Crhanova M., Karasova D., Cejkova D., Sebkova A., et al. (2019). Contact With Adult Hen Affects Development of Caecal Microbiota in Newly Hatched Chicks. *PLoS One* 14, e0212446. doi: 10.1371/journal.pone.0212446
- Kuiken T., Leighton F. A., Fouchier R. A. M., LeDuc J. W., Peiris J. S. M., Schudel A., et al. (2005). Pathogen Surveillance in Animals. *Science* 309, 1680–1681. doi: 10.1126/science.1113310
- Kursa O., Tomczyk G., Sawicka-Durkalec A., Giza A., and Slomiany-Szwarc M. (2021). Bacterial Communities of the Upper Respiratory Tract of Turkeys. *Sci. Rep.* 11, 1–11. doi: 10.1038/s41598-021-81984-0
- Lehtimäki J., Sinkko H., Hielm-Björkman A., Salmela E., Tiira K., Laatikainen T., et al. (2018). Skin Microbiota and Allergic Symptoms Associate With Exposure to Environmental Microbes. *Proc. Natl. Acad. Sci. U. S. A.* 115, 4897–4902. doi: 10.1073/pnas.1719785115
- Lima S. F., de Souza Bicalho M. L., and Bicalho R. C. (2019). The Bos Taurus Maternal Microbiome: Role in Determining the Progeny Early-Life Upper Respiratory Tract Microbiome and Health. *PLoS One* 14, e0208014. doi: 10.1371/journal.pone.0208014
- Li N., Ma W. T., Pang M., Fan Q. L., and Hua J. L. (2019). The Commensal Microbiota and Viral Infection: A Comprehensive Review. *Front. Immunol.* 10, 1551–1567. doi: 10.3389/fimmu.2019.01551
- Lima S. F., Teixeira A. G. V., Higgins C. H., Lima F. S., and Bicalho R. C. (2016). The Upper Respiratory Tract Microbiome and its Potential Role in Bovine Respiratory Disease and Otitis Media. *Sci. Rep.* 6, 1–12. doi: 10.1038/srep29050
- Liu Q. X., Zhou Y., Li X. M., Ma D. D., Xing S., Feng J. H., et al. (2020). Ammonia Induce Lung Tissue Injury in Broilers by Activating NLRP3 Inflammasome Via Escherichia/Shigella. *Poult. Sci.* 99, 3402–3410. doi: 10.1016/j.psj.2020.03.019
- Li Z., Wang X., Di D., Pan R., Gao Y., Xiao C., et al. (2021). Comparative Analysis of the Pulmonary Microbiome in Healthy and Diseased Pigs. *Mol. Genet. Genomics* 296, 21–31. doi: 10.1007/s00438-020-01722-5
- Lovell P. V., Wirthlin M., Wilhelm L., Minx P., Lazar N. H., Carbone L., et al. (2014). Conserved Syntenic Clusters of Protein Coding Genes are Missing in Birds. *Genome Biol.* 15, 565. doi: 10.1186/s13059-014-0565-1
- Lowe B., Marsh T., Isaacs-Cosgrove N., Kirkwood R., Kiupel M., and Mulks M. (2012). Defining the “Core Microbiome” of the Microbial Communities in the Tonsils of Healthy Pigs. *BMC Microbiol.* 12, 20. doi: 10.1186/1471-2180-12-20
- Luehrs A., Siegenthaler S., Grützner N., Kuhnert P., and Nathues H. (2017). Occurrence of Mycoplasma Hyorhinis Infections in Fattening Pigs and Association With Clinical Signs and Pathological Lesions of Enzootic Pneumonia. *Vet. Microbiol.* 203, 1–5. doi: 10.1016/j.vetmic.2017.02.001
- Mach N., Berri M., Estellé J., Levenez F., Lemonnier G., Denis C., et al. (2015). Early-Life Establishment of the Swine Gut Microbiome and Impact on Host Phenotypes. *Environ. Microbiol. Rep.* 7, 554–569. doi: 10.1111/1758-2229.12285
- Mach N., Foury A., Kittelmann S., Reigner F., Moroldo M., Ballester M., et al. (2017). The Effects of Weaning Methods on Gut Microbiota Composition and Horse Physiology. *Front. Physiol.* 8, 535. doi: 10.3389/fphys.2017.00535
- Mach N., Lansade L., Bars-Cortina D., Dhome-Pollet S., Foury A., Moisan M.-P., et al. (2021a). Gut Microbiota Resilience in Horse Athletes Following Holidays Out to Pasture. *Sci. Rep.* 11, 5007–5023. doi: 10.1038/s41598-021-84497-y
- Mach N., Moroldo M., Rau A., Lecardonnell J., Le Moyec L., Robert C., et al. (2021b). Understanding the Holobiont: Crosstalk Between Gut Microbiota and Mitochondria During Long Exercise in Horse. *Front. Mol. Biosci.* 8, 656204–656221. doi: 10.3389/fmolb.2021.656204
- Mach N., Ruet A., Clark A., Bars-Cortina D., Ramayo-Caldas Y., Crisci E., et al. (2020). Priming for Welfare: Gut Microbiota is Associated With Equitation Conditions and Behavior in Horse Athletes. *Sci. Rep.* 10, 8311–8330. doi: 10.1038/s41598-020-65444-9
- Mahmoud Y. S., Correa-Fiz F., and Aragon V. (2020). Variations in Association of Nasal Microbiota With Virulent and non-Virulent Strains of Glaesserella (Haemophilus) Parasuis in Weaning Piglets. *Vet. Res.* 51, 7. doi: 10.1186/s13567-020-0738-8
- Maina J. N., King A. S., and Settle G. (1989). An Allomet- Ric Study of Pulmonary Morphometric Parameters in Birds, With Mammalian Comparisons. *Philos. Trans. R. Soc B Biol. Sci.* 326, 1–57. doi: 10.1098/rstb.1989.0104
- Man W. H., De Steenhuijsen P. W. A. A., and Bogaert D. (2017). The Microbiota of the Respiratory Tract: Gatekeeper to Respiratory Health. *Nat. Rev. Microbiol.* 15, 259–270. doi: 10.1038/nrmicro.2017.14
- Maslowski K. M., Vieira A. T., Ng A., Kranich J., Sierro F., Yu D., et al. (2009). Regulation of Inflammatory Responses by Gut Microbiota and Chemoattractant Receptor GPR43. *Nature* 461, 1282–1286. doi: 10.1038/nature08530
- Mateos Hernández L., Risco-Castillo V., Torres-Maravilla E., Bermúdez-Humarán L., Rakotobe S., Galon C., et al. (2020). Gut Microbiota Abrogates Anti- $\alpha$ -Gal IgA Response in Lungs and Protects Against Experimental Aspergillus Infection in Poultry. *Vaccines* 8, 285. doi: 10.3390/vaccines8020285
- Maynou G., Chester-Jones H., Bach A., and Terré M. (2019). Feeding Pasteurized Waste Milk to Preweaned Dairy Calves Changes Fecal and Upper Respiratory Tract Microbiota. *Front. Vet. Sci.* 6, 159. doi: 10.3389/fvets.2019.00159
- Mazel-Sanchez B., Yildiz S., and Schmolke M. (2019). Ménage À Trois: Virus, Host, and Microbiota in Experimental Infection Models. *Trends Microbiol.* 27, 440–452. doi: 10.1016/j.tim.2018.12.004
- McMullen C., Alexander T. W., Léguillette R., Workentine M., and Timsit E. (2020). Topography of the Respiratory Tract Bacterial Microbiota in Cattle. *Microbiome* 8, 91. doi: 10.1186/s40168-020-00869-y
- McMullen C., Orsel K., Alexander T. W., van der Meer F., Plastow G., and Timsit E. (2018). Evolution of the Nasopharyngeal Bacterial Microbiota of Beef Calves From Spring Processing to 40 Days After Feedlot Arrival. *Vet. Microbiol.* 225, 139–148. doi: 10.1016/j.vetmic.2018.09.019
- McMullen C., Orsel K., Alexander T. W., van der Meer F., Plastow G., and Timsit E. (2019). Comparison of the Nasopharyngeal Bacterial Microbiota of Beef Calves Raised Without the Use of Antimicrobials Between Healthy Calves and Those Diagnosed With Bovine Respiratory Disease. *Vet. Microbiol.* 231, 56–62. doi: 10.1016/j.vetmic.2019.02.030
- Megahed A., Zeineldin M., Evans K., Maradiaga N., Blair B., Aldridge B., et al. (2019). Impacts of Environmental Complexity on Respiratory and Gut Microbiome Community Structure and Diversity in Growing Pigs. *Sci. Rep.* 9, 13773. doi: 10.1038/s41598-019-50187-z
- Meng F., Tong J., Vötsch D., Peng J. Y., Cai X., Willenborg M., et al. (2019). Viral Coinfection Replaces Effects of Sulysin on Streptococcus Suis Adherence to and Invasion of Respiratory Epithelial Cells Grown Under Air-Liquid Interface Conditions. *Infect. Immun.* 87, 1–12. doi: 10.1128/IAI.00350-19
- Meng F., Wu N. H., Seitz M., Herrler G., and Valentin-Weigand P. (2016). Efficient Sulysin-Mediated Invasion and Apoptosis in Porcine Respiratory Epithelial Cells After Streptococcal Infection Under Air-Liquid Interface Conditions. *Sci. Rep.* 6, 26748. doi: 10.1038/srep26748
- Michiels A., Piepers S., Ulens T., Van Ransbeeck N., Del Pozo Sacristán R., Sierens A., et al. (2015). Impact of Particulate Matter and Ammonia on Average Daily Weight Gain, Mortality and Lung Lesions in Pigs. *Prev. Vet. Med.* 121, 99–107. doi: 10.1016/j.prevetmed.2015.06.011
- Million M., and Raoult D. (2018). Linking Gut Redox to Human Microbiome. *Hum. Microbiome J.* 10, 27–32. doi: 10.1016/j.humic.2018.07.002
- Mitchell J. A., Cardwell J. M., Leach H., Walker C. A., Le Poder S., Decaro N., et al. (2017). European Surveillance of Emerging Pathogens Associated With Canine Infectious Respiratory Disease. *Vet. Microbiol.* 212, 31–38. doi: 10.1016/j.vetmic.2017.10.019
- Mostafa A., Abdelwhab E. M., Mettenleiter T. C., and Pleschka S. (2018). Zoonotic Potential of Influenza A Viruses: A Comprehensive Overview. *Viruses* 10, 1–38. doi: 10.3390/v10090497

- Mou K. T., Allen H. K., Alt D. P., Trachsel J., Hau S. J., Coetzee J. F., et al. (2019). Shifts in the Nasal Microbiota of Swine in Response to Different Dosing Regimens of Oxytetracycline Administration. *Vet. Microbiol.* 237, 108386. doi: 10.1016/j.vetmic.2019.108386
- Much P., Winner F., Stipkovits L., Rosengarten R., and Citti C. (2002). *Mycoplasma Gallisepticum*: Influence of Cell Invasiveness on the Outcome of Experimental Infection in Chickens. *FEMS Immunol. Med. Microbiol.* 34, 181–186. doi: 10.1016/S0928-8244(02)00378-4
- Mulholland K. A., Robinson M. G., Keeler S. J., Johnson T. J., Weber B. W., and Keeler C. L. (2021). Metagenomic Analysis of the Respiratory Microbiome of a Broiler Flock From Hatching to Processing. *Microorganisms* 9, 1–18. doi: 10.3390/microorganisms9040721
- Ngunjiri J. M., Taylor K. J. M., Abundo M. C., Jang H., Elaish M., Mahesh K. C., et al. (2019). Farm Stage, Bird Age, and Body Site Dominantly Affect the Quantity, Taxonomic Composition, and Dynamics of Respiratory and Gut Microbiota of Commercial Layer Chickens. *Appl. Environ. Microbiol.* 85, 1–17. doi: 10.1128/AEM.03137-18
- Nicola I., Cerutti F., Grego E., Bertone I., Gianella P., D'Angelo A., et al. (2017). Characterization of the Upper and Lower Respiratory Tract Microbiota in Piedmontese Calves. *Microbiome* 5, 152. doi: 10.1186/s40168-017-0372-5
- Niederwerder M. C. (2017). Role of the Microbiome in Swine Respiratory Disease. *Vet. Microbiol.* 209, 97–106. doi: 10.1016/j.vetmic.2017.02.017
- Niederwerder M. C., Constance L. A., Rowland R. R., Abbas W., Fernando S. C., Potter M. L., et al. (2018). Fecal Microbiota Transplantation is Associated With Reduced Morbidity and Mortality in Porcine Circovirus Associated Disease. *Front. Microbiol.* 9, 1631–1649. doi: 10.3389/fmicb.2018.01631
- Niederwerder M. C., Jaing C. J., Thissen J. B., Cino-Ozuna A. G., McLoughlin K. S., and Rowland R. R. (2016). Microbiome Associations in Pigs With the Best and Worst Clinical Outcomes Following Co-Infection With Porcine Reproductive and Respiratory Syndrome Virus (PRRSV) and Porcine Circovirus Type 2 (PCV2). *Vet. Microbiol.* 188, 1–11. doi: 10.1016/j.vetmic.2016.03.008
- Nochi T., Jansen C. A., Toyomizu M., and van Eden W. (2018). The Well-Developed Mucosal Immune Systems of Birds and Mammals Allow for Similar Approaches of Mucosal Vaccination in Both Types of Animals. *Front. Nutr.* 5, 60. doi: 10.3389/fnut.2018.00060
- Ober R. A., Thissen J. B., Jaing C. J., Cino-Ozuna A. G., Rowland R. R., and Niederwerder M. C. (2017). Increased Microbiome Diversity at the Time of Infection is Associated With Improved Growth Rates of Pigs After Co-Infection With Porcine Reproductive and Respiratory Syndrome Virus (PRRSV) and Porcine Circovirus Type 2 (PCV2). *Vet. Microbiol.* 208, 203–211. doi: 10.1016/j.vetmic.2017.06.023
- Oladunni F. S., Horohov D. W., and Chambers T. M. (2019). EHV-1: A Constant Threat to the Horse Industry. *Front. Microbiol.* 10, 2668. doi: 10.3389/fmicb.2019.02668
- Patel J. G., Patel B. J., Patel S. S., Raval S. H., Parmar R. S., Joshi D. V., et al. (2018). Metagenomic of Clinically Diseased and Healthy Broiler Affected With Respiratory Disease Complex. *Data Br.* 19, 82–85. doi: 10.1016/j.dib.2018.05.010
- Peacock T. P., James J., Sealy J. E., and Iqbal M. (2019). A Global Perspective on h9n2 Avian Influenza Virus. *Viruses* 11, 1–28. doi: 10.3390/v11070620
- Peiris J. S. M., Poon L. L. M., and Guan Y. (2012). Surveillance of Animal Influenza for Pandemic Preparedness. *Science* 335, 1173–1174. doi: 10.1126/science.1219936
- Pena Cortes L. C., Leveque R. M., Funk J., Marsh T. L., and Mulks M. H. (2018). Development of the Tonsillar Microbiome in Pigs From Newborn Through Weaning. *BMC Microbiol.* 18, 35. doi: 10.1186/s12866-018-1176-x
- Pirollo M., Espinosa-Gongora C., Bogaert D., and Guardabassi L. (2021). The Porcine Respiratory Microbiome: Recent Insights and Future Challenges. *Anim. Microbiome* 3, 9. doi: 10.1186/s42523-020-00070-4
- Qin S., Ruan W., Yue H., Tang C., Zhou K., and Zhang B. (2018). Viral Communities Associated With Porcine Respiratory Disease Complex in Intensive Commercial Farms in Sichuan Province, China. *Sci. Rep.* 8, 13341. doi: 10.1038/s41598-018-31554-8
- Raabis S. M., Quick A. E., Skarlupka J. H., Suen G., and Ollivett T. L. (2021). The Nasopharyngeal Microbiota of Prewaned Dairy Calves With and Without Ultrasonographic Lung Lesions. *J. Dairy Sci.* 21, 27–28. doi: 10.3168/jds.2020-19096
- Ramayo-Caldas Y., Mach N., Lepage P., Levenez F., Denis C., Lemonnier G., et al. (2016). Phylogenetic Network Analysis Applied to Pig Gut Microbiota Identifies an Ecosystem Structure Linked With Growth Traits. *ISME J.* 10, 2973–2977. doi: 10.1038/ismej.2016.77
- Ramayo-Caldas Y., Zingaretti L., Popova M., Estellé J., Bernard A., Pons N., et al. (2019). Identification of Rumen Microbial Biomarkers Linked to Methane Emission in Holstein Dairy Cows. *J. Anim. Breed. Genet.* 00, 1–11. doi: 10.1111/jbg.12427
- Ramírez-Labrada A. G., Isla D., Artal A., Arias M., Rezusta A., Pardo J., et al. (2020). The Influence of Lung Microbiota on Lung Carcinogenesis, Immunity, and Immunotherapy. *Trends Cancer* 6, 86–97. doi: 10.1016/j.trecan.2019.12.007
- Richard M., De Graaf M., and Herfst S. (2014). Avian Influenza A Viruses: From Zoonosis to Pandemic. *Future Virol.* 9, 513–524. doi: 10.2217/fvl.14.30
- Rychlik I. (2020). Composition and Function of Chicken Gut Microbiota. *Animals (basel)* 10 (1), 103–123. doi: 10.3390/ani10010103
- Samy A., and Naguib M. M. (2018). Avian Respiratory Coinfection and Impact on Avian Influenza Pathogenicity in Domestic Poultry: Field and Experimental Findings. *Vet. Sci.* 5, 23. doi: 10.3390/vetsci5010023
- Schachtschneider K. M., Yeoman C. J., Isaacson R. E., White B. A., Schook L. B., and Pieters M. (2013). Modulation of Systemic Immune Responses Through Commensal Gastrointestinal Microbiota. *PLoS One* 8, e53969. doi: 10.1371/journal.pone.0053969
- Shabbir M. Z., Malys T., Ivanov Y. V., Park J., Bakr Shabbir M. A., Rabbani M., et al. (2014). Microbial Communities Present in the Lower Respiratory Tract of Clinically Healthy Birds in Pakistan. *Poult. Sci.* 94, 612–620. doi: 10.3382/ps/pev010
- Shi K. C., Guo X., Ge X. N., Liu Q., and Yang H. C. (2010). Cytokine mRNA Expression Profiles in Peripheral Blood Mononuclear Cells From Piglets Experimentally Co-Infected With Porcine Reproductive and Respiratory Syndrome Virus and Porcine Circovirus Type 2. *Vet. Microbiol.* 140, 155. doi: 10.1016/j.vetmic.2009.07.021
- Shin N. R., Whon T. W., and Bae J. W. (2015). Proteobacteria: Microbial Signature of Dysbiosis in Gut Microbiota. *Trends Biotechnol.* 33 (9), 496–503. doi: 10.1016/j.tibtech.2015.06.011
- Slifizer M. J., Friendship R. M., and Weese J. S. (2015). Longitudinal Study of the Early-Life Fecal and Nasal Microbiotas of the Domestic Pig. *BMC Microbiol.* 15, 184. doi: 10.1186/s12866-015-0512-7
- Sohail M. U., Hume M. E., Byrd J. A., Nisbet D. J., Shabbir M. Z., Ijaz A., et al. (2015). Molecular Analysis of the Caecal and Tracheal Microbiome of Heat-Stressed Broilers Supplemented With Prebiotic and Probiotic. *Avian Pathol.* 44, 67–74. doi: 10.1080/03079457.2015.1004622
- Stroebel C., Alexander T., Workentine M. L., and Timsit E. (2018). Effects of Transportation to and Co-Mingling at an Auction Market on Nasopharyngeal and Tracheal Bacterial Communities of Recently Weaned Beef Cattle. *Vet. Microbiol.* 223, 126–133. doi: 10.1016/j.vetmic.2018.08.007
- Suez J., Zmora N., Segal E., and Elinav E. (2019). The Pros, Cons, and Many Unknowns of Probiotics. *Nat. Med.* 25, 716–729. doi: 10.1038/s41591-019-0439-x
- Surendran Nair M., Eucker T., Martinson B., Neubauer A., Victoria J., Nicholson B., et al. (2019). Influence of Pig Gut Microbiota on *Mycoplasma Hypopneumoniae* Susceptibility. *Vet. Res.* 50, 86. doi: 10.1186/s13567-019-0701-8
- Taylor K. J. M., Ngunjiri J. M., Abundo M. C., Jang H., Elaish M., Ghorbani A., et al. (2020). Respiratory and Gut Microbiota in Commercial Turkey Flocks With Disparate Weight Gain Trajectories Display Differential Compositional Dynamics. *Appl. Environ. Microbiol.* 86, e00431–e00420. doi: 10.1128/AEM.00431-20
- Thanawongnuweh R., Thacker B., Halbur P., and Thacker E. L. (2004). Increased Production of Proinflammatory Cytokines Following Infection With Porcine Reproductive and Respiratory Syndrome Virus and *Mycoplasma Hypopneumoniae*. *Clin. Diagn. Lab. Immunol.* 11, 901–908. doi: 10.1128/CDLI.11.5.901-908.2004
- Timsit E., Workentine M., Crepieux T., Miller C., Regev-Shoshani G., Schaefer A., et al. (2017). Effects of Nasal Instillation of a Nitric Oxide-Releasing Solution or Parenteral Administration of Tilmicosin on the Nasopharyngeal Microbiota of Beef Feedlot Cattle at High-Risk of Developing Respiratory Tract Disease. *Res. Vet. Sci.* 115, 117–124. doi: 10.1016/j.rvsc.2017.02.001
- Timsit E., Workentine M., Schryvers A. B., Holman D. B., van der Meer F., and Alexander T. W. (2016). Evolution of the Nasopharyngeal Microbiota of Beef

- Cattle From Weaning to 40 Days After Arrival at a Feedlot. *Vet. Microbiol.* 187, 75–81. doi: 10.1016/j.vetmic.2016.03.020
- Timsit E., Workentine M., van der Meer F., and Alexander T. (2018). Distinct Bacterial Metacommunities Inhabit the Upper and Lower Respiratory Tracts of Healthy Feedlot Cattle and Those Diagnosed With Bronchopneumonia. *Vet. Microbiol.* 221, 105–113. doi: 10.1016/j.vetmic.2018.06.007
- Tress B., Dorn E. S., Suchodolski J. S., Nisar T., Ravindran P., Weber K., et al. (2017). Bacterial Microbiome of the Nose of Healthy Dogs and Dogs With Nasal Disease. *PLoS One* 12, e0176736. doi: 10.1371/journal.pone.0180299
- Umar S., Guerin J. L., and Ducatez M. F. (2016). Low Pathogenic Avian Influenza and Coinfecting Pathogens: A Review of Experimental Infections in Avian Models. *Avian Dis.* 61, 3–15. doi: 10.1637/11514-101316-review
- van Leenen K., Van Driessche L., De Cremer L., Masmeijer C., Boyen F., Deprez P., et al. (2020). Comparison of Bronchoalveolar Lavage Fluid Bacteriology and Cytology in Calves Classified Based on Combined Clinical Scoring and Lung Ultrasonography. *Prev. Vet. Med.* 176, 104901. doi: 10.1016/j.prevetmed.2020.104901
- Vayssier-Taussat M., Albina E., Citti C., Cosson J. F., Jacques M. A., Lebrun M. H., et al. (2014). Shifting the Paradigm From Pathogens to Pathobiome New Concepts in the Light of Meta-Omics. *Front. Cell. Infect. Microbiol.* 4, 29. doi: 10.3389/fcimb.2014.00029
- Vayssier-Taussat M., Kazimirova M., Hubalek Z., Hornok S., Farkas R., Cosson J. F., et al. (2015). Emerging Horizons for Tick-Borne Pathogens: From the “One Pathogen-One Disease” Vision to the Pathobiome Paradigm. *Future Microbiol.* 10, 2033–2043. doi: 10.2217/fmb.15.114
- Vientoos-Plotts A. I., Ericsson A. C., Rindt H., Grobman M. E., Graham A., Bishop K., et al. (2017). Dynamic Changes of the Respiratory Microbiota and its Relationship to Fecal and Blood Microbiota in Healthy Young Cats. *PLoS One* 12, 1–17. doi: 10.1371/journal.pone.0173818
- Vientos-Plotts A. I., Ericsson A. C., Rindt H., and Reinero C. R. (2017). Oral Probiotics Alter Healthy Feline Respiratory Microbiota. *Front. Microbiol.* 8, 1287. doi: 10.3389/fmicb.2017.01287
- Vientos-Plotts A. I., Ericsson A. C., Rindt H., and Reinero C. R. (2019). Respiratory Dysbiosis in Canine Bacterial Pneumonia: Standard Culture vs. Microbiome Sequencing. *Front. Vet. Sci.* 6, 354. doi: 10.3389/fvets.2019.00354
- Vötsch D., Willenborg M., Weldearegay Y. B., and Valentin-Weigand P. (2018). *Streptococcus Suis* - The “Two Faces” of a Pathobiont in the Porcine Respiratory Tract. *Front. Microbiol.* 9, 480. doi: 10.3389/fmicb.2018.00480
- Wagner D. C., Kass P. H., and Hurley K. F. (2018). Cage Size, Movement in and Out of Housing During Daily Care, and Other Environmental and Population Health Risk Factors for Feline Upper Respiratory Disease in Nine North American Animal Shelters. *PLoS One* 13 (1), e0190140. doi: 10.1371/journal.pone.0190140
- Wang Q., Cai R., Huang A., Wang X., Qu W., Shi L., et al. (2018). Comparison of Oropharyngeal Microbiota in Healthy Piglets and Piglets With Respiratory Disease. *Front. Microbiol.* 9, 3218–3229. doi: 10.3389/fmicb.2018.03218
- Wang Z., Chai W., Burwinkel M., Twardziok S., Wrede P., Palissa C., et al. (2013b). Inhibitory Influence of *Enterococcus Faecium* on the Propagation of Swine Influenza A Virus In Vitro. *PLoS One* 8, e53043. doi: 10.1371/journal.pone.0053043
- Wang T., He Q., Yao W., Shao Y., Li J., and Huang F. (2019). The Variation of Nasal Microbiota Caused by Low Levels of Gaseous Ammonia Exposure in Growing Pigs. *Front. Microbiol.* 10, 1083–1097. doi: 10.3389/fmicb.2019.01083
- Wang J., Ishfaq M., and Li J. (2021). *Lactobacillus Salivarius* Ameliorated *Mycoplasma Gallisepticum*-Induced Inflammatory Injury and Secondary *Escherichia Coli* Infection in Chickens: Involvement of Intestinal Microbiota. *Vet. Immunol. Immunopathol.* 233, 110192. doi: 10.1016/j.vetimm.2021.110192
- Wang M., Radlowski E. C., Monaco M. H., Fahey G. C., Gaskins H. R., and Donovan S. M. (2013a). Mode of Delivery and Early Nutrition Modulate Microbial Colonization and Fermentation Products in Neonatal Piglets. *J. Nutr.* 143, 795–803. doi: 10.3945/jn.112.173096
- Wasko A. J., Barkema H. W., Nicol J., Fernandez N., Logie N., and Léguillette R. (2011). Evaluation of a Risk-Screening Questionnaire to Detect Equine Lung Inflammation: Results of a Large Field Study. *Equine Vet. J.* 43 (2), 145–152. doi: 10.1111/j.2042-3306.2010.00150.x
- Weese J. S., Sliifer M., Jalali M., and Friendship R. (2014). Evaluation of the Nasal Microbiota in Slaughter-Age Pigs and the Impact on Nasal Methicillin-Resistant *Staphylococcus Aureus* (MRSA) Carriage. *BMC Vet. Res.* 10, 1–10. doi: 10.1186/1746-6148-10-69
- West J. B., Watson R. R., and Fu Z. (2007). Major Differences in the Pulmonary Circulation Between Birds and Mammals. *Respir. Physiol. Neurobiol.* 157, 382–390. doi: 10.1016/j.resp.2006.12.005
- Wypych T. P., Wickramasinghe L. C., and Marsland B. J. (2019). The Influence of the Microbiome on Respiratory Health. *Nat. Immunol.* 20, 1279–1290. doi: 10.1038/s41590-019-0451-9
- Yitbarek A., Alkie T., Taha-Abdelaziz K., Astill J., Rodriguez-Lecompte J. C., Parkinson J., et al. (2018a). Gut Microbiota Modulates Type I Interferon and Antibody-Mediated Immune Responses in Chickens Infected With Influenza Virus Subtype H9N2. *Benef. Microbes* 9, 417–427. doi: 10.3920/BM2017.0088
- Yitbarek A., Taha-Abdelaziz K., Hodgins D. C., Read L., Nagy É., Weese J. S., et al. (2018b). Gut Microbiota-Mediated Protection Against Influenza Virus Subtype H9N2 in Chickens is Associated With Modulation of the Innate Responses. *Sci. Rep.* 8, 13189. doi: 10.1038/s41598-018-31613-0
- Zaneveld J. R., McMinds R., and Thurber R. V. (2017). Stress and Stability: Applying the Anna Karenina Principle to Animal Microbiomes. *Nat. Microbiol.* 2, 17121. doi: 10.1038/nmicrobiol.2017.121
- Zeineldin M., Aldridge B., Blair B., Kancer K., and Lowe J. (2018). Microbial Shifts in the Swine Nasal Microbiota in Response to Parenteral Antimicrobial Administration. *Microb. Pathog.* 121, 210–217. doi: 10.1016/j.micpath.2018.05.028
- Zeineldin M., Elolimy A. A., and Barakat R. (2020a). Meta-Analysis of Bovine Respiratory Microbiota: Link Between Respiratory Microbiota and Bovine Respiratory Health. *FEMS Microbiol. Ecol.* 96, 127. doi: 10.1093/femsec/fiaa127
- Zeineldin M., Lowe J., and Aldridge B. (2019). Contribution of the Mucosal Microbiota to Bovine Respiratory Health. *Trends Microbiol.* 27, 753. doi: 10.1016/j.tim.2019.04.005
- Zeineldin M., Lowe J., and Aldridge B. (2020b). Effects of Tilmicosin Treatment on the Nasopharyngeal Microbiota of Feedlot Cattle With Respiratory Disease During the First Week of Clinical Recovery. *Front. Vet. Sci.* 7, 115. doi: 10.3389/fvets.2020.00115
- Zeineldin M., Lowe J., de Godoy M., Maradiaga N., Ramirez C., Ghanem M., et al. (2017a). Disparity in the Nasopharyngeal Microbiota Between Healthy Cattle on Feed, at Entry Processing and With Respiratory Disease. *Vet. Microbiol.* 208, 30–37. doi: 10.1016/j.vetmic.2017.07.006
- Zeineldin M. M., Lowe J. F., Grimmer E. D., De Godoy M. R. C., Ghanem M. M., Abd El-Raof Y. M., et al. (2017b). Relationship Between Nasopharyngeal and Bronchoalveolar Microbial Communities in Clinically Healthy Feedlot Cattle. *BMC Microbiol.* 17, 1–11. doi: 10.1186/s12866-017-1042-2
- Zhao N., Wang S., Li H., Liu S., Li M., Luo J., et al. (2018). Influence of Novel Highly Pathogenic Avian Influenza A (H5N1) Virus Infection on Migrating Whooper Swans Fecal Microbiota. *Front. Cell. Infect. Microbiol.* 8, 46. doi: 10.3389/fcimb.2018.00046
- Zhou Y., Zhang M., Liu Q., and Feng J. (2021). The Alterations of Tracheal Microbiota and Inflammation Caused by Different Levels of Ammonia Exposure in Broiler Chickens. *Poult. Sci.* 100, 685–696. doi: 10.1016/j.psj.2020.11.026
- Zhu X., Dordet-Frisoni E., Gillard L., Ba A., Hygonenq M. C., Sagné E., et al. (2019). Extracellular DNA: A Nutritional Trigger of *Mycoplasma Bovis* Cytotoxicity. *Front. Microbiol.* 10, 2753–2766. doi: 10.3389/fmicb.2019.02753

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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