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Mitochondrial dysfunction and alveolar type II epithelial cell senescence: The destroyer and rescuer of idiopathic pulmonary fibrosis

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Idiopathic pulmonary fibrosis (IPF) is a chronic respiratory disease with an unknown origin and complex pathogenic mechanisms. A deeper understanding of these mechanisms is essential for effective treatment. Pulmonary fibrosis is associated with the senescence of alveolar type II epithelial (ATII) cells. Additionally, ATII senescence can lead to a senescence-associated secretory phenotype, which affects cellular communication and disrupts lung tissue repair, contributing to the development of IPF. The role of mitochondrial dysfunction in senescence-related diseases is increasingly recognized. It can induce ATII senescence through apoptosis, impaired autophagy, and disrupted energy metabolism, potentially playing a key role in IPF progression. This article explores the therapeutic potential of targeting cellular senescence and mitochondrial dysfunction, emphasizing their significant roles in IPF pathogenesis.

KEYWORDS

idiopathic pulmonary fibrosis, cellular senescence, mitochondrial dysfunction, alveolar type 2 (AT2) cells, senotherapeutics

1 Introduction

Idiopathic pulmonary fibrosis (IPF) is an interstitial lung disease of unknown etiology. The disease is chronic, progressive, destructive, and irreversible. The hallmark feature of IPF is the progressive formation and remodeling of lung scarring (Sui et al., 2023). Although the exact course and primary cause of IPF remain unclear, it is now thought that the pathogenesis of the disease may be related to genetic, environmental, viral, or immune factors (Ellson et al., 2014).

Fibrosis is a class of diseases caused by chronic organ damage, characterized by tissue hardening and scarring, and is generally described as excessive pathological deposition of extracellular matrix (ECM) during wound healing (Hinz and Lagares, 2020). Collagen, fibronectin, laminin, and other substances make up the ECM, an intercellular matrix that is essential for healthy tissue healing. Although ECM deposition is an inevitable byproduct of wound healing, if tissue damage persists or recurs, ECM synthesis and remodeling will become uncontrollable, resulting in the formation of persistent fibrotic scars that impair organ function and normal

structure (Fischer et al., 2024). Damaged epithelial cells, endothelial cells, and innate fibroblasts are stimulated by injury and undergo transdifferentiation into myofibroblasts, which are capable of strong contraction and matrix formation (Hailiwu et al., 2023). Excessive synthesis and deposition of ECM components lead to tissue structural remodeling and dysfunction, which are caused by myofibroblasts (Bhatt et al., 2024), which are important cellular mediators in the development of fibrosis.

Notably, fibrosis may play a role in the occurrence and spread of cancer. For example, it can improve the mechanical support of tumors, protect tumor cells from immune system attacks, and promote tumor growth by changing local hemodynamics (Tomos et al., 2025). One study showed that the incidence of lung cancer in patients with pulmonary fibrosis was 4.8%–48%, while the incidence in the control group was only 2.0%–6.4%. This difference may be related to the chronic inflammation and lung tissue destruction experienced by patients with pulmonary fibrosis (Kato et al., 2018).

The prognosis of IPF patients is poor, with an average survival of only about 3-5 years after diagnosis (Raghu et al., 2014). Epidemiological studies have shown that people aged 65 years and above have the highest incidence of IPF, and the incidence increases with age (Maher et al., 2021). Compared with people aged 40 years, people aged 70 years and above have a 6.9-fold increased risk of developing the disease (Choi et al., 2018). Therefore, IPF is considered an aging-related disease, and aging is considered an important risk factor for IPF (Wan et al., 2024). Cell growth arrest and reduced replication capacity are hallmarks of aging, and aging makes the lungs susceptible to fibrosis by preventing alveolar progenitor cells from regenerating and cultivating a cellular environment that is conducive to fibrosis (Smith et al., 2022). The primary markers of the link between senescence and IPF include cellular senescence, the senescence-associated secretory phenotype (SASP), and immunosenescence (Salminen, 2025). Alveolar epithelial type II (ATII) cells in IPF patients exhibit pronounced signs of senescence.

Cellular senescence is a key hallmark of aging (López-Otín et al., 2023). In several interstitial fibrosis disorders, senescence—an irreversible cell cycle arrest—is a defining characteristic (Hernandez-Gonzalez et al., 2021). ATII cells are essential for maintaining lung homeostasis (K. Liu et al., 2024). Growing evidence suggests that ATII cell senescence plays a crucial role in the remodeling process of aging-related pulmonary fibrosis. Senescent ATII cells communicate with surrounding cells by secreting SASP factors, which propagate peripheral cellular senescence, promote ECM deposition, disrupt lung structure, impair lung function, and ultimately contribute to the onset of IPF (Yao et al., 2021). In other words, senescent ATII cells activate fibroblasts and myofibroblasts, further exacerbating fibrosis.

A well-established hallmark of cellular senescence is mitochondrial dysfunction (Suryadevara et al., 2024). The term "mitochondrial dysfunction" primarily refers to impaired energy metabolism caused by mitochondrial DNA (mtDNA) damage, disruption of the mitochondrial membrane, inhibition of the respiratory chain, and decreased enzyme activity (Rangarajan et al.,

2017). These factors trigger several interconnected damage processes. In addition to being crucial for cellular energy production, apoptosis, and redox balance, mitochondria play a significant role in cellular senescence. Senescence is characterized by mitochondrial alterations, including reduced oxidative phosphorylation (OXPHOS), decreased levels of adenosine 5'triphosphate (ATP) and nicotinamide adenine dinucleotide (NAD⁺), and an accumulation of reactive oxygen species (ROS), damage-associated molecular patterns (DAMPs), and metabolites from the tricarboxylic acid (TCA) cycle (Martini and Passos, 2023). Many diseases are associated with its dysfunction. Hallmark features of idiopathic pulmonary fibrosis include altered metabolic processes, increased oxidative stress, and reduced cell survival, all of which are caused by mitochondrial dysfunction (Larson-Casey et al., 2020). More and more studies have recognized the importance of mitochondrial dysfunction in aging. It can mediate AT II cell senescence, thereby impairing lung tissue regeneration and AT II cell differentiation capacity. This ultimately increases the difficulty of maintaining the AT barrier over time and leads to fibroblast barrier activation, which is crucial for the development of fibrotic scars and idiopathic pulmonary fibrosis (Perry et al., 2024).

Transforming growth factor β (TGF- β)/Smad signaling pathway, Wnt/ β -catenin signaling pathway, vascular endothelial growth factor (VEGF), fibroblast growth factor (FGF) and platelet-derived growth factor (PDGF) signaling pathways are currently the main focus of research on the pathophysiology and related treatment options of idiopathic pulmonary fibrosis at home and abroad (Claudia et al., 2020). IPF is improved by preventing fibroblast activation and excessive collagen production, reducing cell proliferation and migration, and enhancing blood vessels.

Pirfenidone and nintedanib, which also use this pathway, are both approved by the U.S. Food and Drug Administration (FDA) for the treatment of IPF (Claudia et al., 2020). These drugs do not completely prevent the progressive loss of lung function; instead, they are intended only as palliative therapies for pulmonary fibrosis. They can also cause gastrointestinal problems, photosensitivity, and abnormal laboratory results such as elevated aminotransferases. Because of these negative consequences, IPF patients are less likely to adhere to their medication regimen, which can lead to further disease progression and loss of lung function (Gulati and Luckhardt, 2020). Lung transplantation is the only treatment that can potentially cure IPF, but it is not suitable for most patients (Justice et al., 2019). In addition, the pathophysiology of IPF involves complex immune pathways, irreversible lung tissue fibrosis, and the effects of antifibrotic drugs have their own limitations (Allanore et al., 2024). All of these reasons lead to major challenges in the current treatment of IPF.

Related research is ongoing, and it is crucial to understand how mitochondrial dysfunction and AT II senescence lead to IPF. In order to provide new perspectives for future IPF research and treatment, we systematically summarized the mechanisms of IPF induced by mitochondrial dysfunction in recent years, summarized the pathways of ATII senescence involved in IPF, and elaborated on IPF treatment targeting mitochondrial function and ATII senescence (Figure 1).

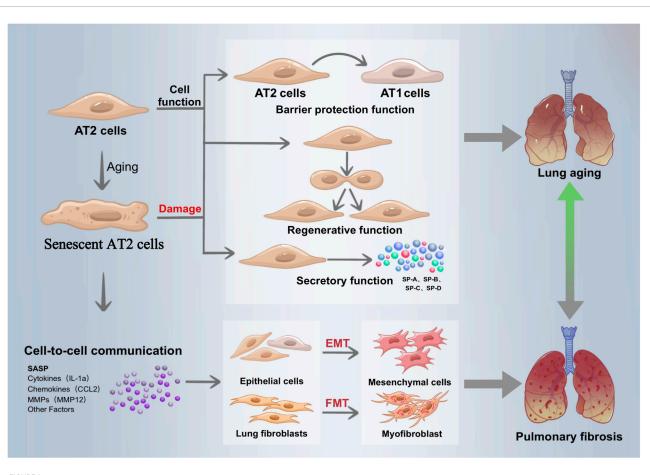


FIGURE 1

Senescent alveolar type 2 epithelial cells drive the development of IPE. Alveolar epithelial cells are divided into alveolar type 1 epithelial cells (ATI) and alveolar type 2 epithelial cells (ATI2). ATI participates in gas exchange and is found on the alveolar surface. With the ability of develop into ATI in order to restore the damaged ACE barrier, AT2 is a crucial component of the alveolar wall and the primary stem cell in the alveolar repair process. Its functions include barrier protection and regeneration. In addition, AT2 lowers alveolar surface tension control lung immunological activity, secretes a range of alveolar surfactants, and preserves lung function and alveolar structural stability. As people age, their capacity to generate the lung tissue is diminished, their AT2 barrier function is compromised and their ability to replenish alveolar epithelial stem cells is drastically diminished. By secreting SASP, senescent epithelial cells with AT2 cells as their primary constituent encourage the differentiation of epithelial cells into mesenchymel cells (EMT) stimulate neighboring fibroblasts into myofibroblasts (FMT). These action result in extracellular matrix secretion, collagen deposition, and lung tissue remodeling, all of which contribute to the pathological process of pulmonary fibrosis.

2 Senescent AT II in IPF: Involvement in cellular communication leads to abnormal lung tissue repair

2.1 Senescent ATII in IPF leads to abnormal lung tissue repair

One of the fundamental characteristics of cellular senescence is prolonged or permanent cell cycle arrest. Several factors, including telomere shortening, DNA damage, oxidative stress, senescence gene regulation, and epigenetic alterations, can trigger this process in IPF, ultimately limiting the regeneration of alveolar epithelial progenitor cells (Suryadevara et al., 2024). Single-cell RNA sequencing studies have shown that increased ATII cell senescence in the lung tissues of IPF patients accelerates pulmonary fibrosis by activating profibrotic myofibroblasts through multiple conventional mechanisms (Rubio et al., 2020). The prevailing consensus is that IPF is driven by ATII senescence, which leads to myofibroblast activation and ECM synthesis, resulting in fibrotic scarring and impaired lung tissue repair (Figure 2).

ATII cells play a crucial role in maintaining alveolar structure, stabilizing intrapulmonary function, and serving as the primary stem cells for alveolar repair. They can differentiate into ATI cells to restore the damaged alveolar barrier. According to Witschi's theory, lung fibrosis originates and progresses due to epithelial damage microfoci. If these microfoci are not repaired in a timely manner, the normal balance between fibroblasts and epithelial cells is disrupted, promoting fibrosis development (Haschek and Witschi, 1979).

When ATII cells undergo senescence, several age-related changes occur in lung tissue, including reduced alveolar epithelial stem cell renewal, impaired ATII function and differentiation capacity, defective lung tissue regeneration, increased expression of senescence markers such as p16 and p21, and altered β -galactosidase activity (SA- β -gal). The failure of senescent ATII cells to maintain the alveolar barrier, coupled with cell

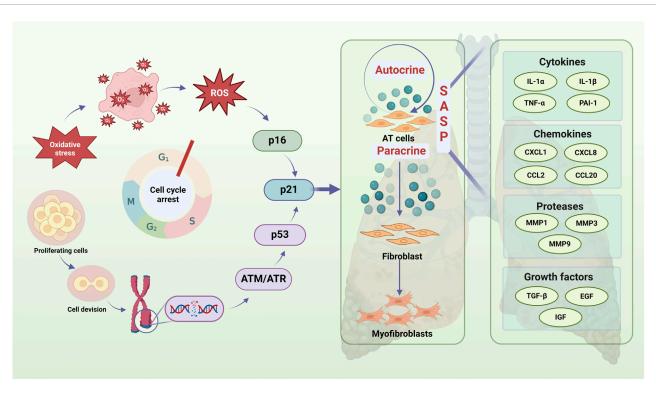


FIGURE 2

Senescent AT2 contributes to cellular communication, which exacerbates IPF by casing aberrant lung tissue repair. Oxidative stress and DNA damage are major contributions to AT2 cell cycle arrest (particularly G1 cycle) and regenerative failure during IPF. An imbalance between the production of free radical and the cell's capacity to scavenge them is known as oxidative stress, and its causes ROS buildup and P16 pathway activation. The continuous interference of endogenous and external stressors during cell division and proliferation can cause DNA damage, which sets off DNA damage responses, activates ATM/ATR and other DNA repair pathways, and ultimately influences P53 activation. The aforementioned two routes have the ability to trigger the P21 pathway, impact AT2 cells' autocrine and paracrine SASP, worsen AT2 cell senescence, and encourage peripheral fibroblasts to become myofibroblasts, all of which have an impact on the onset and progression of IPF. A crucial component of IPF cell communication, SASP contains growth factors, proteases, cytokines, chemokines, and more.

cycle arrest, leads to fibroblast activation, proliferation, and collagen deposition, ultimately contributing to fibrotic scarring (Liang et al., 2023).

In this study, we focused on the role of ATII cells in the development of IPF, their contribution to a pro-fibrotic cellular environment, their predisposition to fibrosis, and their regulation of cellular communication through SASP paracrine and autocrine signaling pathways.

2.2 Senescent AT II plays a pro-fibrotic role in accelerating IPF development through the involvement of SASP in cellular communication

Atypical lung tissue healing is triggered by the SASP, which primarily consists of growth factors, chemokines, and proinflammatory cytokines (Zeng et al., 2024a). Overexpression of SASP has been shown to strongly induce cellular senescence and promote epithelial-mesenchymal transition (EMT) through autocrine secretion (Chilosi et al., 2013). Additionally, SASP regulates the microenvironment in a paracrine manner, stimulating neighboring fibroblasts and myofibroblasts to excessively ECM, leading to lung tissue remodeling and, ultimately, the pathological progression of pulmonary fibrosis. Furthermore, SASP accelerates immune cell senescence and promotes chronic inflammation, which weakens immune function and impairs the clearance of inflammatory factors and senescent cells (Li et al., 2024). This creates a vicious cycle of senescence and inflammation. The immune system dysfunction caused by this cycle is referred to as immunosenescence (Dasgupta et al., 2024).

On one hand, SASP contributes to the progression of IPF through autocrine signaling. Yasunori Enomoto et al. demonstrated that DNA damage induced by bleomycin (BLM) activated p53 signaling in ATII cells, leading to TGF- β -mediated pro-fibrotic gene expression. This initiated a positive feedback loop of TGF- β signaling, which further exacerbated ATII senescence and contributed to IPF development (Enomoto et al., 2023).

On the other hand, SASP secreted by senescent ATII cells can induce senescence in nearby cells through paracrine signaling, playing a crucial role in lung aging and the progression of pulmonary fibrosis. Lehmann hypothesized that the reprogramming of alveolar epithelial cells by SASP components—such as interleukin-6 (IL-6), interleukin-1 β (IL-1 β), matrix metalloproteinase-12 (MMP-12), chemokine ligand 2 (CCL2), and keratinocyte growth factor—plays a major role in the pathophysiology of IPF. Their study also demonstrated that senescent ATII cells in mice with pulmonary fibrosis secrete higher levels of SASP, which in turn promotes fibroblast-to-myofibroblast transformation, thereby exacerbating fibrosis (Lehmann et al., 2017).

Furthermore, aging ATII cells have been found to promote massive proliferation and activation of fibroblasts and myofibroblasts through the expression of SASP factors such as PDGF, tumor necrosis factor (TNF), endothelin-1, connective tissue growth factor (CTGF), chemokine (C-X-C motif) ligand 12 (CXCL12), and plasminogen activator inhibitor-1 (PAI-1) (Rana et al., 2020). *In vitro* studies by *Rana* T demonstrated that PAI-1 serves as both a marker and mediator of cellular senescence. Notably, PAI-1 knockdown almost completely reversed bleomycin-induced ATII senescence and pulmonary fibrosis in mice (Li et al., 2023).

In conclusion, the autocrine and paracrine involvement of SASP in cellular communication disrupts lung tissue repair, leading senescent ATII cells to drive the development of IPF.

2.3 Other pathways

In addition, telomere dysfunction plays a crucial role in the involvement of senescent ATII cells in the development of IPF. A recent study demonstrated that telomerase inactivation due to *TERC* gene deletion not only accelerated ATII cell senescence but also promoted apoptosis and differentiation of ATII cells through both p53-dependent and p53-independent mechanisms (Zhang et al., 2021a). Other studies have shown that telomere dysfunction mediated by *TRF1* deletion can lead to mitochondrial damage and pulmonary remodeling *via* the ECM, with increased expression of senescence markers observed in ATII cells (Naikawadi et al., 2016).

Notably, the accumulation of DNA damage is one of the primary drivers of aging and age-related disorders. Studies have shown that YTHDC1 expression is reduced in both IPF patients and mouse models of pulmonary fibrosis, whereas YTHDC1 overexpression inhibits senescence and mitigates IPF *in vitro* and *in vivo* (Zhang et al., 2024).

These findings suggest that ATII cell senescence disrupts lung tissue repair, thereby accelerating the onset of IPF. They also highlight a promising avenue for further research into the role of ATII senescence in IPF progression. Targeting SASP secretion and intervening in ATII cell senescence could be valuable therapeutic strategies for pulmonary fibrosis, as these approaches may help slow or prevent IPF progression at an early stage.

3 The role of mitochondrial dysfunction in alveolar type II cell senescence and IPF progression

Mitochondrial dysfunction is a well-established hallmark of cellular aging (Suryadevara et al., 2024). Energy metabolism is one of the key functions of mitochondria. Most of the intracellular ATP is produced by mitochondria, which are also called the "powerhouses of the cell" because they help cells maintain a high ATP/ADP ratio, which is required to thermodynamically drive many metabolic events.

Many conditions, such as oxidative phosphorylation disorders, mtDNA mutations, and abnormal mitochondrial shape and

number, can be signs of mitochondrial dysfunction. All of these impairments exacerbate the process of cellular aging and may affect mitochondrial function, including energy production, redox balance, and calcium regulation (Van et al., 2020). In addition, mitochondrial dysfunction disrupts redox homeostasis (Seo et al., 2008), leading to accumulation of cytoplasmic NADH, and a decrease in the NAD/NADH ratio can lead to ATP depletion and cell cycle arrest (Wu et al., 2022).

The mitochondrial respiratory chain is essential for maintaining redox balance and intracellular signaling and is the main generator of intracellular ROS (Guan et al., 2024). Mitochondrial dysfunction is the root cause of excessive intracellular oxidative stress and can also affect the functioning of lysosomes, endoplasmic reticulum, and other organelles, promote autophagy, accelerate cell apoptosis, and ultimately lead to cell aging and death (Niforou et al., 2014; Zuk and Bonventre, 2016; Zhou et al., 2022). Additionally, elevated ROS act as endogenous DNA-damaging agents, inducing genetic instability and senescence-related gene alterations (Baranski et al., 2015), which in turn damage alveolar epithelial cells and compromise the epithelial barrier (Figure 3).

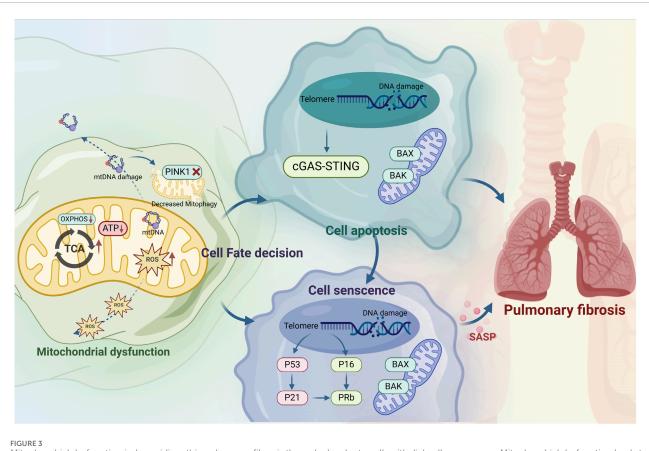
In IPF patients, abnormally enlarged and swollen mitochondria have been observed in ATII cells. This is attributed to the disruption of mitochondrial mass maintenance mechanisms, such as mitochondrial biogenesis and mitophagy, which can drive ATII cell senescence. Given their high metabolic demand due to surfactant production, ATII cells are particularly susceptible to mitochondrial dysfunction.

3.1 Decreased PINK1-mediated classical autophagy and energy metabolism

ATP production during aerobic respiration is closely linked to the structural and functional integrity of mitochondria. Cells utilize autophagy to remove dysfunctional mitochondria, thereby maintaining mitochondrial homeostasis and normal function (Boyman et al., 2020). However, autophagic activity declines with age (Harrington et al., 2023). Notably, 50% of lung mitochondria are found in ATII cells, making them particularly susceptible to age-related changes such as mitochondrial enlargement, cristae loss, endosome degradation, and reduced respiratory capacity (Sreedhar et al., 2020). Using TEM, Xia observed a significant increase in mitochondrial vacuolization and membrane rupture in senescent ATII cells (Ning et al., 2019), indicating mitochondrial dysfunction and impaired energy metabolism.

Mitochondrial dysfunction has been linked to decreased expression of PTEN-induced putative kinase 1 (PINK1), the primary regulator of mitochondrial homeostasis *in vivo*. Mitochondrial damage disrupts PINK1 translocation, leading to its activation on the outer mitochondrial membrane *via* autophosphorylation. Activated PINK1 recruits and activates the downstream autophagy protein E3 ubiquitin ligase Parkin, which enhances mitochondrial autophagy and mitigates epithelial cell senescence—an essential mechanism for limiting fibrosis (Sosulski et al., 2015). However, maintaining the PINK1-mediated autophagy pathway in senescent ATII cells is challenging (Bueno et al., 2015).

Mitochondrial autophagy reduces SASP factor release and alleviates cellular senescence (Chu et al., 2024). PINK1-mediated



Mitochondrial dysfunction induces idiopathic pulmonary fibrosis through alveolar type II epithelial cell senescence. Mitochondrial dysfunction leads to IPF through apoptosis and senescence. Mitochondrial dysfunction leads to abnormal energy metabolism, decreased mitochondrial ATP production, ROS accumulation, damage to mtDNA, and disruption of PINK1-mediated mitophagy. This process damages chromosomal DNA and activates the p53 pathways, which determines cell fates between apoptosis and senescence, but the specific mechanism is still unclear and may be related to stress. In apoptotic cells, p53 induces mitochondrial outer membrane permeabilization by regulating apoptotic pore formation, which allows cytochrome c release and caspase activation, leading to cell death. Chromosomal DNA damage activates the p53/p21 and p61/pRb pathways, induces cell cycle arrest and exacerbates cells senescence. The upregulation of the pro-survival pathway in senescent cells inhibits the formation of apoptotic pores, resulting in the release of miMOMP, sublethal apoptosis and mtDNA into cytoplasm, and mtDNA fragments are sensed by the cGAS-STING pathway, up-regulating the expression of inflammatory mediators, promoting cell senescence and exacerbating pulmonary fibrosis.

mitochondrial autophagy has been identified as a key factor in the pathophysiology of age-related lung diseases such as COPD and IPF (Bueno et al., 2015). Increasing evidence links mitochondrial autophagy and cellular senescence to the progression of IPF in the elderly (Wei et al., 2023a). In mouse lung tissue, PINK1 knockdown in ATII cells resulted in mitochondrial enlargement and dysfunction, impairing mitochondrial autophagy and increasing susceptibility to lung fibrosis (Chu et al., 2024). Thus, the regulation of mitochondrial autophagy plays a critical role in mitigating ATII cell senescence and preventing senescence-associated IPF by preserving mitochondrial homeostasis.

3.2 The role of cell apoptosis in alveolar type II cell senescence and IPF progression

Groundbreaking research in mitochondrial genetics has demonstrated that mitochondria release cytochrome C, a key component of the electron transport chain (ETC.), to induce apoptosis—a programmed cell death pathway distinct from cellular senescence (Victorelli et al., 2023). Mitochondria play a critical role in apoptosis regulation. Moreover, p53 activation is a pivotal step in aging. In response to various stimuli, p53 upregulates p21 to arrest the cell cycle and subsequently regulates transcriptional programs leading to apoptosis or cellular senescence (Huang et al., 2011). Senescent cells activate the p53/p21 and p16/pRb pathways, characterized by a prolonged DNA damage response (DDR). Additionally, apoptosis is primarily regulated by the Bcl-2 (B-cell lymphoma-2) protein family (Tian et al., 2024). In normal cells, Bcl-2 proteins localize to membrane structures such as the outer mitochondrial membrane (Rasmussen and Gama, 2020), where they interact with Bak and Bax (Bcl-2 Associated X protein) to prevent oligomer formation, maintain mitochondrial membrane integrity, and inhibit cytochrome C release, exerting anti-apoptotic effects (Samuel et al., 2010; Cao et al., 2023).

Mitochondrial dysfunction is a key driver of apoptosis in ATII cells with IPF. The ATM/ATR or AMPK pathway, activated by mitochondrial failure (Anand et al., 2020), phosphorylates p53, stabilizing it and enhancing its transcriptional activity. p53

disrupts the balance of Bcl-2 family proteins by activating proapoptotic factors and inhibiting anti-apoptotic proteins, leading to cytochrome C release and the extrusion of mtDNA into the cytoplasm through BAX/BAK pores (Schubert et al., 2024). mtDNA, as a damage-associated molecular pattern, binds to DNA pattern-recognition receptors, triggering the innate immune response *via* the cGAS-STING pathway (Li and Chen, 2018). *In vivo* studies have shown that cGAS-STING activation exacerbates apoptosis in alveolar epithelial cells (Huang et al., 2022). Moreover, persistent mitochondrial damage in IPF may exceed endogenous compensatory mechanisms, leading to chronic accumulation of dysfunctional mitochondria (Kapetanovic et al., 2015).

When ATII cells undergo apoptosis, pro-apoptotic proteins Bak and Bax, initially inhibited by Bcl-2, become activated, undergo conformational changes, and oligomerize on the outer mitochondrial membrane, increasing mitochondrial outer membrane permeability (MOMP) (Subas satish et al., 2024). Excessive MOMP promotes cellular senescence, triggering inflammation and SASP molecule secretion, including IL-6 and IL-8 (Victorelli et al., 2023; Garciaz et al., 2022).

ATII cells from IPF patients exhibit mitochondrial dysfunction and impaired autophagy (Bueno et al., 2015). *In vivo* studies have shown that PINK1-deficient mice develop similar mitochondrial abnormalities in ATII cells, leading to apoptosis and lung fibrosis (Bueno et al., 2015). Additionally, persistent stress can amplify mitochondrial damage (Schuliga et al., 2021). Mitochondrial dysfunction has been reported in IPF, connective tissue diseaseassociated interstitial lung disease (ILD), and experimental ILD models (Pokharel et al., 2024).

4 Targeting cellular senescence and mitochondrial dysfunction in IPF: A promising therapeutic approach

4.1 Targeting senescent AT II cells for the treatment of IPF

Senotherapeutics aim to address aging-related health issues by eliminating or suppressing senescent cells. This approach includes two main strategies: senolytics and senomorphics (Table 1). Senolytics induce apoptosis in senescent cells by targeting antiapoptotic pathways. For example, ABT-263 (Zhu et al., 2016) and ABT-737 (Yosef et al., 2016) bind to BCL-2, BCL-XL, and BCL-W, triggering apoptosis in senescent cells. Another senolytic, ABT-263, specifically inhibits BCL-2 and BCL-XL, effectively eliminating senescent cells. In vitro and in vivo studies confirm its anti-aging and anti-fibrotic properties, suggesting potential for treating age-related fibrotic diseases. Dasatinib (D) and quercetin (Q), when combined (D + Q), selectively induce apoptosis in senescent human cells without affecting non-senescent cells. This combination was the first identified anti-aging drug and has shown promise in improving age-related conditions in mice and in limited human trials (Justice et al., 2019).

Recent studies suggest that roxithromycin inhibits senescence through a NOX4-dependent mechanism, making it a potential treatment for IPF (Zhang et al., 2021b). However, researchers caution against its immediate use as a senolytic due to concerns

Targets of Action	Drug name	Verified path	PubMed ID
Senescent cells	ABT-263	vitro	26,711,051
		vivo and vitro	34,318,888
	ABT-737	vivo and vitro	27,048,913
	Dasatinib	clinical trials 30,616,998	
	Quercetin		30,616,998
	Roxithromycin	vivo and vitro	33,654,217
	SSK1	vivo and vitro	32,341,413
	Procyanidin C1	vivo and vitro	34,873,338
SASP	Rapamycin	vivo and vitro	28,371,119
	Resveratrol	vitro	28,329,136
	Metformin	vivo and vitro	26,990,999
		clinical trials	
	Aspirin	vitro	16,039,999
	Rutin	vivo and vitro	37,475,161

TABLE 1 Strategies targeting senescent AT II cells for the treatment of IPF.

about antimicrobial resistance. Instead, its properties could inform the development of future senolytic drugs. Another promising compound, senescence-specific killing compound 1 (SSK1), is activated by β -gal and removes senescent cells *via* p38/MAPK signaling. *In vivo* and *ex vivo* studies demonstrate its ability to alleviate IPF, reduce inflammation, and decrease senescence-related gene expression (Cai et al., 2020). Additionally, research on grape seed extract led to the successful isolation of proanthocyanidin C1, which has been shown in cellular and animal studies to effectively eliminate senescent cells and potentially extend lifespan.

Senomorphics, a newer anti-aging approach, mitigate the harmful effects of senescence by inhibiting SASPs rather than directly eliminating senescent cells. Representative senomorphics include metformin, rapamycin, and resveratrol (Wang et al., 2017; Menicacci et al., 2017; Noren hooten et al., 2016). Other potential senomorphics include aspirin, NF- κ B inhibitors, p38 MAPK inhibitors, JAK/STAT inhibitors, ATM inhibitors, and statins (Bode-Böger et al., 2005). Unlike senolytics, senomorphics primarily target SASP to prevent the paracrine/autocrine spread of senescence to neighboring and distant cells.

Rapamycin, for example, has demonstrated therapeutic effects on IPF in both *in vivo* and *ex vivo* studies by inhibiting the mTOR pathway, thereby reducing SASP production and inflammation while slowing cellular senescence (Chrienova et al., 2022). Similarly, rutin, a natural compound, has been identified as a potential senomorphic that suppresses SASP expression and may be used to treat aging-related diseases (H. Liu et al., 2024). By targeting the initial senescent cells, senomorphics not only prevent senescence from spreading but also disrupt the cycle that promotes further accumulation of senescent cells. While generally less potent than senolytics, natural polyphenols are gaining popularity due to their low toxicity and availability.

4.2 Targeting dysfunctional mitochondria for the treatment of IPF

The complex nature of mitochondrial dysfunction and its elusive phenotypic thresholds make it challenging to fully understand its role in disease. However, mitochondria remain key targets for therapeutic intervention. Mitochondrial failure contributes to pulmonary fibrosis by driving ATII cell senescence. Given this, targeting mitochondrial dysfunction offers a promising strategy to treat or delay IPF progression. Advances in mitochondrial biology have led to new therapeutic approaches, though human clinical trials remain limited, highlighting the need for further research. Potential treatments include antioxidants and drugs that enhance mitochondrial ATP production, reduce oxidative stress, and improve mitochondrial function (Wal et al., 2024). These strategies range from dietary interventions addressing nutritional deficiencies to pharmacological therapies that modulate mitochondrial dynamics, boost biogenesis, and mitigate oxidative damage (Table 2).

Reducing mitochondrial oxidative stress is a critical therapeutic goal. Inhibitors of mitochondrial fission play a key role in achieving this (Qi et al., 2013; Yang et al., 2024; Ko et al., 2021). Mdivi-1, for example, counteracts excess ROS by blocking Dynamin-Related Protein 1 (DRP1) GTPase activity, improving endothelial function and reducing inflammation in animal models (Qi et al., 2013). Similarly, the peptide P110 enhances mitochondrial function by inhibiting DRP1 (Qi et al., 2013). Senegenin has been shown in vivo to prevent oxidative stress-induced epithelial cell senescence and reduce lung fibrosis by modulating the Sirt1/Pgc-1a pathway (Zeng et al., 2024b). In BLM-induced lung fibrosis models, TH5487 reduced oxidative stress, promoted PINK1/Parkinmediated mitophagy, and alleviated mitochondrial dysfunction. Additionally, clinical research indicates that MitoQ scavenges free radicals, protecting cells from oxidative stress and enhancing cellular function.

Mitochondrial autophagy (mitophagy) plays a vital role in mitigating pulmonary fibrosis caused by ATII cell senescence. Tetrandrine (TET) has been shown to reduce lung inflammation and fibrosis by regulating mitophagy through the PINK1-Parkin signaling pathway (Chu et al., 2024). In MLE-12 cells, TET rescued impaired BLM-induced mitophagy by preventing the reduction of autophagy-related protein expression, while PINK1 gene knockdown abolished its effects (Chu et al., 2024). Naringenin has also been found to regulate mitophagy and alleviate pulmonary fibrosis *via* the ATF3/PINK1 pathway (Wei et al., 2023b). Several natural compounds, including spermidine, resveratrol, and urushiol A, support mitochondrial integrity by stimulating mitophagy and promoting biogenesis (Palikaras et al., 2018).

The cGAS/STING pathway is closely linked to aging and lung fibrosis, though its precise role remains unclear. Recent studies indicate that urushiol A-induced mitophagy reduces free cytoplasmic mtDNA activation of cGAS/STING, improving

Targets of Action	Drug name	Verified path	PubMed ID
oxidative stress	Mdivi 1	vivo	23,239,023
	P110	vivo	23,239,023
	Senegenin	vivo	38,929,114
	TH5487	vivo	38,929,114
	MitoQ	vivo	16,829,229
Mitophagy	TET	vivo and vitro	38,438,063
	Tetrandrine	vitro	38,438,063
	Naringin	vivo	36,688,958
	Spermidine	vivo and vitro	30,154,567
	Resveratrol	vivo and vitro	38,865,904
	Urolithin A		
	Magnokiol		
cGAS/STING	Urolithin A	vivo and vitro	30,154,567
	Przewalskin	vivo	38,237,513
	Harmine	vivo	38,924,867
	BAI 1	vivo	38,168,624
	MitoTam	clinical trials	29,786,070

TABLE 2 Strategies targeting mitochondrial dysfunction for the treatment of IPF.

mitochondrial function. Additionally, homopalol enhances SIRT3 deacetylation activity, activating SOD2's antioxidant function and OGG1-mediated DNA repair. This modulation of the cGAS/STING pathway helps prevent fibrosis and cellular senescence (Wu et al., 2024). Other compounds, such as purple salvia terpene ether and salvia divinorum extract, have been shown to reduce BLM-induced lung fibrosis by inhibiting TGF- β 1 signaling, oxidative stress, and collagen deposition (M. Zeng et al., 2024). Harmine has been confirmed *in vitro* to prevent pulmonary fibrosis by regulating DDR-associated genes and activating the TP53-Gadd45 α pathway (Gong et al., 2024).

Targeting inflammation linked to mitochondrial outer membrane permeabilization (miMOMP) may also be beneficial. For example, BAX inhibitor BAI-1 reduces mitochondrial BAX and BAK nanopores, lowering systemic inflammation and extending healthy lifespan in aged mice (Victorelli et al., 2023). Additionally, MitoTam, a mitochondria-targeted tamoxifen currently in clinical trials, has been shown to induce apoptosis in senescent cells and reduce senescence markers (p21 and p16) in the kidneys and lungs of aged mice (Hubackova et al., 2019).

Gene replacement therapy and gene editing technologies offer potential solutions for inherited mitochondrial disorders,

though a deeper understanding of the mitochondrial genome is required. Combination therapies have also gained attention; for instance, combining MitoQ with senolytics may enhance mitochondrial function while reducing senescent cell burden.

5 Conclusions and prospects

Although aging and IPF-a disease primarily affecting the elderly-are closely linked, cellular senescence in IPF differs significantly from normal physiological aging (Hagood, 2014). Normal senescence is a gradual, systemic decline, whereas in IPF, cellular senescence is rapid, localized, and pathologically driven, leading to alveolar epithelial failure and fibrosis (Confalonieri et al., 2022). In IPF, ATII cells are the primary targets of senescence, with mutation-driven, SASPmediated, and microenvironmental consequences that disrupt the alveolar barrier and activate fibroblasts, ultimately causing irreversible fibrosis (Confalonieri et al., 2022; Lei et al., 2024). In contrast, normal aging results in a slow decline in organ function and increased susceptibility to age-related diseases but is not inherently fatal. Therapeutic strategies for aging include mTOR inhibitors, antioxidants, anti-aging therapies, and calorie restriction, whereas IPF-specific treatments focus on antifibrotic agents and telomerase activation (Cătană et al., 2018). While approved antifibrotic drugs like pirfenidone and nintedanib slow IPF progression, they have side effects and cannot reverse lung function decline. Therefore, research on IPF aging could enhance pulmonary fibrosis management while offering new insights into anti-aging therapies.

Senolytic drugs targeting tumorigenic pathways have emerged as a promising class of therapies, though their long-term optimization remains challenging. Additionally, the chronic lung damage and inflammation in IPF may share mechanistic links with cancer, offering potential directions for anti-aging drug development. Targeting the "two-way vicious cycle" between mitochondrial dysfunction and ATII senescence may provide precision therapies for IPF. This cycle involves mitochondrial damage triggering senescence *via* ROS and energy depletion, while senescent cells further impair mitochondria and activate fibroblasts through SASP. While most evidence suggests ATII senescence promotes fibrosis, some SASP factors may inhibit collagen production, highlighting a complex, microenvironmentdependent effect.

Future research should explore novel drugs targeting this pathway to treat pulmonary fibrosis. Investigating the clinical potential of compounds that have shown efficacy in animal and cellular models could accelerate the development of effective IPF therapies. Mitochondria-targeted approaches align with precision medicine, offering tailored treatments that mitigate ATII cell senescence while minimizing adverse effects. However, the roles of many naturally occurring compounds in modulating mitochondrial autophagy, ATP production, and oxidative stress remain unclear. By targeting multiple pathways associated with mitochondrial dysfunction, these treatments could help counteract fibrosis progression.

Implementing a personalized medical approach requires clinically relevant biomarkers. A robust biomarker panel, monitored longitudinally, could facilitate early detection of treatment responses in clinical trials. However, challenges remain, including variability in staining techniques, sensitivity issues due to small sample sizes, and the need for independent validation. Most research is still limited to preclinical models, necessitating further human trials to assess the efficacy of senescence-targeting and mitochondrial therapies in IPF. Additionally, substances that show promise in treating fibrosislike conditions in other organs may not necessarily be effective against pulmonary fibrosis. Variability in experimental methods, reagents, and models further complicates research outcomes. The interactions between cellular senescence, mitochondrial dysfunction, and other disease factors remain poorly understood, and a major hurdle in IPF treatment is the lack of therapies that simultaneously target both ATII cell senescence and mitochondrial dysfunction.

Advancements in technology are deepening our understanding of IPF pathogenesis, which could optimize therapeutic strategies. As research progresses, new treatments targeting key molecular pathways are being developed, and clinical trials are gradually advancing. Gene therapies related to aging and pulmonary fibrosis are also emerging, propelled by advances in genetics and gene editing. Ultimately, IPF treatment is evolving toward greater precision and personalization, aiming to minimize side effects and improve patients' quality of life. With continued drug development and ongoing clinical trials, there is reason to be optimistic about the future of IPF treatment.

Author contributions

Suqi Liu: Investigation, Methodology, Software, Visualization, Writing-original draft, Writing-review and editing. QX: Supervision, Writing-review and editing, Methodology, Writing-original draft. Xuannian Li: Methodology, Supervision, Writing-review and editing, Writing-original draft. HL: Supervision, Writing-review and editing, Funding acquisition, Resources, Validation, Writing-original draft.

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References

Allanore, Y., Khanna, D., Smith, V., Aringer, M., Hoffmann-Vold, A. M., Kuwana, M., et al. (2024). Effects of nintedanib in patients with limited cutaneous systemic sclerosis and interstitial lung disease. *Rheumatol. Oxf.* 63 (3), 639–647. doi:10.1093/rheumatology/kead280

Anand, S. K., Sharma, A., Singh, N., and Kakkar, P. (2020). Entrenching role of cell cycle checkpoints and autophagy for maintenance of genomic integrity. *DNA Repair* (*Amst*) 86: 102748. doi:10.1016/j.dnarep.2019.102748

Baranski, O. A., Kalinichenko, V. V., and Adami, G. R. (2015). Increased FOXM1 expression can stimulate DNA repair in normal hepatocytes *in vivo* but also increases nuclear foci associated with senescence. *Cell Prolif.* 48 (1), 105–115. doi:10.1111/cpr.12153

Bhatt, H. N., Diwan, R., Estevao, I. L., Dong, R., Smith, J., Xiao, C., et al. (2024). Cadherin-11 targeted cell-specific liposomes enabled skin fibrosis treatment by inducing apoptosis [J]. *J. Control Release* 370: 110–123. doi:10.1016/j.jconrel.2024.04.029

Bode-BöGER, S. M., Martens-Lobenhoffer, J., TäGER, M., Schröder, H., and Scalera, F. (2005). Aspirin reduces endothelial cell senescence. *Biochem. Biophys. Res. Commun.* 334 (4), 1226–1232. doi:10.1016/j.bbrc.2005.07.014

Boyman, L., Karbowski, M., and Lederer, W. J. (2020). Regulation of mitochondrial ATP production: Ca(2+) signaling and quality control. *Trends Mol. Med.* 26 (1), 21–39. doi:10.1016/j.molmed.2019.10.007

Bueno, M., Lai, Y. C., Romero, Y., Brands, J., St Croix, C. M., Kamga, C., et al. (2015). PINK1 deficiency impairs mitochondrial homeostasis and promotes lung fibrosis. J. Clin. Invest. 125 (2), 521–538. doi:10.1172/JCI74942

 $\label{eq:stars} \begin{array}{l} Cai, Y., Zhou, H., Zhu, Y., Sun, Q., Ji, Y., Xue, A., et al. (2020). Elimination of senescent cells by β-galactosidase-targeted prodrug attenuates inflammation and restores physical function in aged mice. Cell Res. 30 (7), 574–589. doi:10.1038/s41422-020-0314-9 \end{array}$

Cao, Z., Yang, X., Li, T., Liu, Z., Li, P., Zhou, Y., et al. (2023). Molecular characterization and expression analysis of B-cell lymphoma-2 in Trachinotus ovatus and its role in apoptotic process. *Front. Immunol.* 14: 1129800. doi:10.3389/fimmu.2023.1129800

Cătană, C. S., Atanasov, A. G., and Berindan-Neagoe, I. (2018). Natural products with anti-aging potential: affected targets and molecular mechanisms. *Biotechnol. Adv.* 36 (6), 1649–1656. doi:10.1016/j.biotechadv.2018.03.012

Chilosi, M., Carloni, A., Rossi, A., and Poletti, V. (2013). Premature lung aging and cellular senescence in the pathogenesis of idiopathic pulmonary fibrosis and COPD/emphysema. *Transl. Res.* 162 (3), 156–173. doi:10.1016/j.trsl.2013.06.004

Choi, W. I., Dauti, S., Kim, H. J., Park, S. H., Park, J. S., and Lee, C. W. (2018). Risk factors for interstitial lung disease: a 9-year Nationwide population-based study [J]. *BMC Pulm. Med.* 18 (1), 96. doi:10.1186/s12890-018-0660-2

Chrienova, Z., Rysanek, D., Oleksak, P., Stary, D., Bajda, M., Reinis, M., et al. (2022). Discovery of small molecule mechanistic target of rapamycin inhibitors as anti-aging and anti-cancer therapeutics. *Front. Aging Neurosci.* 14: 1048260. doi:10.3389/fnagi.2022.1048260

Chu, L., Zhuo, J., Huang, H., Chen, W., Zhong, W., Zhang, J., et al. (2024). Tetrandrine alleviates pulmonary fibrosis by inhibiting alveolar epithelial cell senescence through PINK1/Parkin-mediated mitophagy. *Eur. J. Pharmacol.* 969: 176459. doi:10.1016/j.ejphar.2024.176459

Claudia, L., Alfonso, C., Lorenza, V., Laura, B., d'Alessandro, M., Cameli, P., et al. (2020). Common molecular pathways targeted by nintedanib in cancer and IPF: a bioinformatic study. *Pulm. Pharmacol. Ther.* 64: 101941. doi:10.1016/j.pupt.2020.101941

Confalonieri, P., Volpe, M. C., Jacob, J., Maiocchi, S., Salton, F., Ruaro, B., et al. (2022). Regeneration or repair? The role of alveolar epithelial cells in the pathogenesis of idiopathic pulmonary fibrosis (IPF). *Cells* 11 (13), 2095. doi:10.3390/cells11132095

Dasgupta, N., Lei, X., Shi, C. H., Arnold, R., Teneche, M. G., Miller, K. N., et al. (2024). Histone chaperone HIRA, promyelocytic leukemia protein, and p62/SQSTM1 coordinate to regulate inflammation during cell senescence. *Mol. Cell* 84 (17), 3271–3287.e8. doi:10.1016/j.molcel.2024.08.006

Ellson, C. D., Dunmore, R., Hogaboam, C. M., Sleeman, M. A., and Murray, L. A. (2014). Danger-associated molecular patterns and danger signals in idiopathic pulmonary fibrosis. *Am. J. Respir. Cell Mol. Biol.* 51 (2), 163–168. doi:10.1165/rcmb.2013-0366TR

Enomoto, Y., Katsura, H., Fujimura, T., Ogata, A., Baba, S., Yamaoka, A., et al. (2023). Autocrine TGF- β -positive feedback in profibrotic AT2-lineage cells plays

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a crucial role in non-inflammatory lung fibrogenesis. Nat. Commun. 14(1): 4956. doi:10.1038/s41467-023-40617-y

Fischer, A. G., Elliott, E. M., Brittian, K. R., Garrett, L., Sadri, G., Aebersold, J., et al. (2024). Matricellular protein CCN1 promotes collagen alignment and scar integrity after myocardial infarction. *Matrix Biol.* 133 14–32. doi:10.1016/j.matbio.2024.08.001

Garciaz, S., Guirguis, A. A., MüLLER, S., Brown, F. C., Chan, Y. C., Motazedian, A., et al. (2022). Pharmacologic reduction of mitochondrial iron triggers a noncanonical BAX/BAK-Dependent cell death. *Cancer Discov.* 12 (3), 774–791. doi:10.1158/2159-8290.CD-21-0522

Gong, Y., Wang, J., Pan, M., Zhao, Y., Zhang, H., Zhang, F., et al. (2024). Harmine inhibits pulmonary fibrosis through regulating DNA damage repair-related genes and activation of TP53-Gadd45α pathway. *Int. Immunopharmacol.* 138: 112542. doi:10.1016/j.intimp.2024.112542

Guan, B., Liu, Y., Xie, B., Zhao, S., Yalikun, A., Chen, W., et al. (2024). Mitochondrial genome transfer drives metabolic reprogramming in adjacent colonic epithelial cells promoting TGF β 1-mediated tumor progression. *Nat. Commun.* 15 (1), 3653. doi:10.1038/s41467-024-48100-y

Gulati, S., and Luckhardt, T. R. (2020). Updated evaluation of the safety, efficacy and tolerability of pirfenidone in the treatment of idiopathic pulmonary fibrosis. *Drug Healthc. Patient Saf.* 12: 85–94. doi:10.2147/DHPS.S224007

Hagood, J. S. (2014). Beyond the genome: epigenetic mechanisms in lung remodeling. *Physiol. (Bethesda)* 29 (3), 177–185. doi:10.1152/physiol.00048.2013

Hailiwu, R., Zeng, H., Zhan, M., Pan, T., Yang, H., and Li, P. (2023). Salvianolic acid A diminishes LDHA-driven aerobic glycolysis to restrain myofibroblasts activation and cardiac fibrosis via blocking Akt/GSK- 3β /HIF- 1α axis. *Phytother. Res.* 37 (10), 4540–4556. doi:10.1002/ptr.7925

Harrington, J. S., Ryter, S. W., Plataki, M., Price, D. R., and Choi, A. M. K. (2023). Mitochondria in health, disease, and aging. *Physiol. Rev.* 103 (4), 2349–2422. doi:10.1152/physrev.00058.2021

Haschek, W. M., and Witschi, H. (1979). Pulmonary fibrosis-a possible mechanism. *Toxicol. Appl. Pharmacol.* 51 (3), 475–487. doi:10.1016/0041-008x(79)90372-7

Hernandez-Gonzalez, F., Faner, R., Rojas, M., Agustí, A., Serrano, M., and Sellarés, J. (2021). Cellular senescence in lung fibrosis. *Int. J. Mol. Sci.* 22 (13), 7012. doi:10.3390/ijms22137012

Hinz, B., and Lagares, D. (2020). Evasion of apoptosis by myofibroblasts: a hallmark of fibrotic diseases. *Nat. Rev. Rheumatol.* 16 (1), 11–31. doi:10.1038/s41584-019-0324-5

Huang, S. X., Jaurand, M. C., Kamp, D. W., Whysner, J., and Hei, T. K (2011). Role of mutagenicity in asbestos fiber-induced carcinogenicity and other diseases. *J. Toxicol. Environ. Health B Crit. Rev.* 14 (1-4), 179–245. doi:10.1080/10937404.2011.556051

Huang, R., Shi, Q., Zhang, S., Lin, H., Han, C., Qian, X., et al. (2022). Inhibition of the cGAS-STING pathway attenuates lung ischemia/reperfusion injury via regulating endoplasmic reticulum stress in alveolar epithelial type II cells of rats. J. Inflamm. Res. 15: 5103–5119. doi:10.2147/JIR.S365970

Hubackova, S., Davidova, E., Rohlenova, K., Stursa, J., Werner, L., Andera, L., et al. (2019). Selective elimination of senescent cells by mitochondrial targeting is regulated by ANT2. *Cell Death Differ.* 26 (2), 276–290. doi:10.1038/s41418-018-0118-3

Justice, J. N., Nambiar, A. M., Tchkonia, T., LeBrasseur, N. K., Pascual, R., Hashmi, S. K., et al. (2019). Senolytics in idiopathic pulmonary fibrosis: results from a first-in-human, open-label, pilot study. *EBioMedicine* 40: 554–563. doi:10.1016/j.ebiom.2018.12.052

Kapetanovic, R., Bokil, N. J., and Sweet, M. J. (2015). Innate immune perturbations, accumulating DAMPs and inflammasome dysregulation: a ticking time bomb in ageing. *Ageing Res. Rev.* 24 (Pt A), 40–53. doi:10.1016/j.arr.2015.02.005

Kato, E., Takayanagi, N., Takaku, Y., Kagiyama, N., Kanauchi, T., Ishiguro, T., et al. (2018). Incidence and predictive factors of lung cancer in patients with idiopathic pulmonary fibrosis. *ERJ Open Res.* 4 (1), 2016. doi:10.1183/23120541.00111-2016

Ko, H. J., Tsai, C. Y., Chiou, S. J., Lai, Y. L., Wang, C. H., Cheng, J. T., et al. (2021). The phosphorylation status of drp1-ser637 by PKA in mitochondrial fission modulates mitophagy via PINK1/parkin to exert multipolar spindles assembly during mitosis. *Biomolecules* 11 (3), 424. doi:10.3390/biom11030424

Larson-Casey, J. L., He, C., and Carter, A. B. (2020). Mitochondrial quality control in pulmonary fibrosis. *Redox Biol.* 33: 101426. doi:10.1016/j.redox.2020.101426

Lehmann, M., Korfei, M., Mutze, K., Klee, S., Skronska-Wasek, W., Alsafadi, H. N., et al. (2017). Senolytic drugs target alveolar epithelial cell function and

attenuate experimental lung fibrosis ex vivo. Eur. Respir. J. 50 (2), 1602367. doi:10.1183/13993003.02367-2016

Lei, Y., Zhong, C., Zhang, J., Zheng, Q., Xu, Y., Li, Z., et al. (2024). Senescent lung fibroblasts in idiopathic pulmonary fibrosis facilitate non-small cell lung cancer progression by secreting exosomal MMP1. *Oncogene*, 44, 769-781. doi:10.1038/s41388-024-03236-5

Liang, J., Huang, G., Liu, X., Liu, N., Taghavifar, F., Dai, K., et al. (2023). Reciprocal interactions between alveolar progenitor dysfunction and aging promote lung fibrosis. *Elife* 12, e85415. doi:10.7554/eLife.85415

Li, J., Yang, D., Li, Z., Zhao, M., Wang, D., Sun, Z., et al. (2023). PINK1/Parkinmediated mitophagy in neurodegenerative diseases [J]. *Ageing Res. Rev.* 84: 101817. doi:10.1016/j.arr.2022.101817

Li, T., and Chen, Z. J. (2018). The cGAS-cGAMP-STING pathway connects DNA damage to inflammation, senescence, and cancer. *J. Exp. Med.* 215 (5), 1287–1299. doi:10.1084/jem.20180139

Liu, K., Meng, X., Liu, Z., Tang, M., Lv, Z., Huang, X., et al. (2024). Tracing the origin of alveolar stem cells in lung repair and regeneration. *Cell* 187 (10), 2428–2445.e20. doi:10.1016/j.cell.2024.03.010

Liu, H., Xu, Q., Wufuer, H., Li, Z., Sun, R., Jiang, Z., et al. (2024). Rutin is a potent senomorphic agent to target senescent cells and can improve chemotherapeutic efficacy. *Aging Cell* 23 (1), e13921. doi:10.1111/acel.13921

Li, Y., Cui, J., Liu, L., Hambright, W. S., Gan, Y., Zhang, Y., et al. (2024). mtDNA release promotes cGAS-STING activation and accelerated aging of postmitotic muscle cells. *Cell Death Dis.* 15 (7), 523. doi:10.1038/s41419-024-06863-8

López-Otín, C., Pietrocola, F., Roiz-Valle, D., Galluzzi, L., and Kroemer, G. (2023). Meta-hallmarks of aging and cancer. *Cell Metab.* 35 (1), 12–35. doi:10.1016/j.cmet.2022.11.001

Maher, T. M., Bendstrup, E., Dron, L., Langley, J., Smith, G., Khalid, J. M., et al. (2021). Global incidence and prevalence of idiopathic pulmonary fibrosis. *Respir. Res.* 22 (1), 197. doi:10.1186/s12931-021-01791-z

Martini, H., and Passos, J. F. (2023). Cellular senescence: all roads lead to mitochondria. *Febs J.* 290 (5), 1186–1202. doi:10.1111/febs.16361

Menicacci, B., Laurenzana, A., Chillà, A., Margheri, F., Peppicelli, S., Tanganelli, E., et al. (2017). Chronic resveratrol treatment inhibits MRC5 fibroblast SASP-related protumoral effects on melanoma cells. *J. Gerontol. A Biol. Sci. Med. Sci.* 72 (9), 1187–1195. doi:10.1093/gerona/glw336

Naikawadi, R. P., Disayabutr, S., Mallavia, B., Donne, M. L., Green, G., La, J. L., et al. (2016). Telomere dysfunction in alveolar epithelial cells causes lung remodeling and fibrosis. *JCI Insight* 1 (14), e86704. doi:10.1172/jci.insight.86704

Niforou, K., Cheimonidou, C., and Trougakos, I. P. (2014). Molecular chaperones and proteostasis regulation during redox imbalance. *Redox Biol.* 2: 323–332. doi:10.1016/j.redox.2014.01.017

Ning, X., Ji, X., Li, G., and Sang, N. (2019). Ambient PM(2.5) causes lung injuries and coupled energy metabolic disorder. *Ecotoxicol. Environ. Saf.* 170: 620–626. doi:10.1016/j.ecoenv.2018.12.028

Noren Hooten, N., Martin-Montalvo, A., Dluzen, D. F., Zhang, Y., Bernier, M., Zonderman, A. B., et al. (2016). Metformin-mediated increase in DICER1 regulates microRNA expression and cellular senescence. *Aging Cell* 15 (3), 572–581. doi:10.1111/acel.12469

Palikaras, K., Lionaki, E., and Tavernarakis, N. (2018). Mechanisms of mitophagy in cellular homeostasis, physiology and pathology. *Nat. Cell Biol.* 20 (9), 1013–1022. doi:10.1038/s41556-018-0176-2

Perry, A. S., Zhang, K., Murthy, V. L., Choi, B., Zhao, S., Gajjar, P., et al. (2024). Proteomics, human environmental exposure, and cardiometabolic risk. *Circ. Res.* 135 (1), 138–154. doi:10.1161/CIRCRESAHA.124.324559

Pokharel, M. D., Garcia-Flores, A., Marciano, D., Franco, M. C., Fineman, J. R., Aggarwal, S., et al. (2024). Mitochondrial network dynamics in pulmonary disease: bridging the gap between inflammation, oxidative stress, and bioenergetics. *Redox Biol.* 70: 103049. doi:10.1016/j.redox.2024.103049

Qi, X., Qvit, N., Su, Y. C., and Mochly-Rosen, D. (2013). A novel Drp1 inhibitor diminishes aberrant mitochondrial fission and neurotoxicity. *J. Cell Sci.* 126 (Pt 3), 789–802. doi:10.1242/jcs.114439

Raghu, G., Chen, S. Y., Yeh, W. S., Maroni, B., Li, Q., Lee, Y. C., et al. (2014). Idiopathic pulmonary fibrosis in US Medicare beneficiaries aged 65 years and older: incidence, prevalence, and survival, 2001-11. *Lancet Respir. Med.* 2 (7), 566–572. doi:10.1016/S2213-2600(14)70101-8

Rana, T., Jiang, C., Liu, G., Miyata, T., Antony, V., Thannickal, V. J., et al. (2020). PAI-1 regulation of TGF-β1-induced alveolar type II cell senescence, SASP secretion, and SASP-mediated activation of alveolar macrophages. *Am. J. Respir. Cell Mol. Biol.* 62 (3), 319–330. doi:10.1165/rcmb.2019-0071OC

Rangarajan, S., Bernard, K., and Thannickal, V. J. (2017). Mitochondrial dysfunction in pulmonary fibrosis. *Ann. Am. Thorac. Soc.* 14 (Suppl. ment_5), S383–S388. doi:10.1513/AnnalsATS.201705-370AW Rasmussen, M. L., and Gama, V. (2020). A connection in life and death: the BCL-2 family coordinates mitochondrial network dynamics and stem cell fate. *Int. Rev. Cell Mol. Biol.* 353: 255–284. doi:10.1016/bs.ircmb.2019.12.005

Rubio, K., Castillo-Negrete, R., and Barreto, G. (2020). Non-coding RNAs and nuclear architecture during epithelial-mesenchymal transition in lung cancer and idiopathic pulmonary fibrosis. *Cell Signal* 70: 109593. doi:10.1016/j.cellsig.2020.109593

Salminen, A. (2025). Cooperation between inhibitory immune checkpoints of senescent cells with immunosuppressive network to promote immunosenescence and the aging process. *Ageing Res. Rev.* 106: 102694. doi:10.1016/j.arr.2025.102694

Samuel, S., Tumilasci, V. F., Oliere, S., Nguyên, T. L. A., Shamy, A., Bell, J., et al. (2010). VSV oncolysis in combination with the BCL-2 inhibitor obatoclax overcomes apoptosis resistance in chronic lymphocytic leukemia. *Mol. Ther.* 18 (12), 2094–2103. doi:10.1038/mt.2010.188

Schubert, E., Mun, K., Larsson, M., Panagiotou, S., Idevall-Hagren, O., Svensson, C., et al. (2024). Complex regulation of mitochondrial signaling by human adenovirus minor capsid protein VI. *J. Virol.* 98 (7), e0035624. doi:10.1128/jvi.00356-24

Schuliga, M., Read, J., and Knight, D. A. (2021). Ageing mechanisms that contribute to tissue remodeling in lung disease. *Ageing Res. Rev.* 70: 101405. doi:10.1016/j.arr.2021.101405

Seo, A. Y., Xu, J., Servais, S., Hofer, T., Marzetti, E., Wohlgemuth, S. E., et al. (2008). Mitochondrial iron accumulation with age and functional consequences. *Aging Cell* 7 (5), 706–716. doi:10.1111/j.1474-9726.2008.00418.x

Smith, A. L. M., Whitehall, J. C., and Greaves, L. C. (2022). Mitochondrial DNA mutations in ageing and cancer. *Mol. Oncol.* 16 (18), 3276–3294. doi:10.1002/1878-0261.13291

Sosulski, M. L., Gongora, R., Danchuk, S., Dong, C., Luo, F., and Sanchez, C. G. (2015). Deregulation of selective autophagy during aging and pulmonary fibrosis: the role of TGF β 1. *Aging Cell* 14 (5), 774–783. doi:10.1111/acel.12357

Sreedhar, A., Aguilera-Aguirre, L., and Singh, K. K. (2020). Mitochondria in skin health, aging, and disease. *Cell Death Dis.* 11 (6), 444. doi:10.1038/s41419-020-2649-z

Subas Satish, H. P., Iyer, S., Shi, M. X., Wong, A. W., Fischer, K. C., Wardak, A. Z., et al. (2024). A novel inhibitory BAK antibody enables assessment of non-activated BAK in cancer cells. *Cell Death Differ.* 31 (6), 711–721. doi:10.1038/s41418-024-01289-3

Sui, J., Boatz, J. C., Shi, J., Hu, Q., Li, X., Zhang, Y., et al. (2023). Loss of ANT1 increases fibrosis and epithelial cell senescence in idiopathic pulmonary fibrosis. *Am. J. Respir. Cell Mol. Biol.* 69 (5), 556–569. doi:10.1165/rcmb.2022-0315OC

Suryadevara, V., Hudgins, A. D., Rajesh, A., Pappalardo, A., Karpova, A., Dey, A. K., et al. (2024). SenNet recommendations for detecting senescent cells in different tissues. *Nat. Rev. Mol. Cell Biol.* 25 (12), 1001–1023. doi:10.1038/ s41580-024-00738-8

Tian, X., Srinivasan, P. R., Tajiknia, V., Sanchez Sevilla Uruchurtu, A. F., Seyhan, A. A., Carneiro, B. A., et al. (2024). Targeting apoptotic pathways for cancer therapy. *J. Clin. Invest.* 134 (14), e179570. doi:10.1172/JC1179570

Tomos, I., Kanellopoulou, P., Nastos, D., and Aidinis, V. (2025). Pharmacological targeting of ECM homeostasis, fibroblast activation, and invasion for the treatment of pulmonary fibrosis. *Expert Opin. Ther. Targets* 29, 43-57. doi:10.1080/14728222.2025.2471579

Van, D. E. R. RIJT S., Molenaars, M., Mcintyre, R. L., Janssens, G. E., and Houtkooper, R. H. (2020). Integrating the hallmarks of aging throughout the tree of life: a focus on mitochondrial dysfunction. *Front. Cell Dev. Biol.* 8: 594416. doi:10.3389/fcell.2020.594416

Victorelli, S., Salmonowicz, H., Chapman, J., Martini, H., Vizioli, M. G., Riley, J. S., et al. (2023). Apoptotic stress causes mtDNA release during senescence and drives the SASP. *Nature* 622 (7983), 627–636. doi:10.1038/s41586-023-06621-4

Wal, P., Wal, A., Vig, H., Mahmood, D., and Khan, M. M. U. (2024). Potential applications of mitochondrial therapy with a focus on Parkinson's disease and mitochondrial transplantation. *Adv. Pharm. Bull.* 14 (1), 147–160. doi:10.34172/apb.2024.019

Wang, R., Yu, Z., Sunchu, B., Shoaf, J., Dang, I., Zhao, S., et al. (2017). Rapamycin inhibits the secretory phenotype of senescent cells by a Nrf2-independent mechanism. *Aging Cell* 16 (3), 564–574. doi:10.1111/acel.12587

Wan, R., Long, S., Ma, S., Yan, P., Li, Z., Xu, K., et al. (2024). NR2F2 alleviates pulmonary fibrosis by inhibition of epithelial cell senescence. *Respir. Res.* 25 (1), 154. doi:10.1186/s12931-024-02777-3

Wei, Y., You, Y., Zhang, J., Ban, J., Min, H., Li, C., et al. (2023a). Crystalline silicainduced macrophage pyroptosis interacting with mitophagy contributes to pulmonary fibrosis via modulating mitochondria homeostasis. *J. Hazard Mater* 454: 131562. doi:10.1016/j.jhazmat.2023.131562

Wei, Y., Sun, L., Liu, C., and Li, L. (2023b). Naringin regulates endoplasmic reticulum stress and mitophagy through the ATF3/PINK1 signaling axis to alleviate pulmonary fibrosis. *Naunyn Schmiedeb. Arch. Pharmacol.* 396 (6), 1155–1169. doi:10.1007/s00210-023-02390-z

Wu, S., Ai, Y., Huang, H., Wu, G., Zhou, S., Hong, W., et al. (2022). A synthesized olean-28,13 β -lactam targets YTHDF1-GLS1 axis to induce ROS-dependent metabolic crisis and cell death in pancreatic adenocarcinoma. *Cancer Cell Int.* 22 (1), 143. doi:10.1186/s12935-022-02562-6

Wu, W., Jia, H., Chen, S., Ma, X., Zhou, S., Qiu, L., et al. (2024). Inhibition of OGG1 ameliorates pulmonary fibrosis via preventing M2 macrophage polarization and activating PINK1-mediated mitophagy. *Mol. Med.* 30 (1), 72. doi:10.1186/s10020-024-00843-6

Yang, J., Ye, L., Cui, R., Zheng, K., Qiao, X., Wang, M., et al. (2024). Deoxynivalenol inhibits progenitor leydig cell development by stimulating mitochondrial fission in rats. *J. Agric. Food Chem.* 72 (18), 10616–10626. doi:10.1021/acs.jafc.4c01151

Yao, C., Guan, X., Carraro, G., Parimon, T., Liu, X., Huang, G., et al. (2021). Senescence of alveolar type 2 cells drives progressive pulmonary fibrosis. *Am. J. Respir. Crit. Care Med.* 203 (6), 707–717. doi:10.1164/rccm.202004-1274OC

Yosef, R., Pilpel, N., Tokarsky-Amiel, R., Biran, A., Ovadya, Y., Cohen, S., et al. (2016). Directed elimination of senescent cells by inhibition of BCL-W and BCL-XL. *Nat. Commun.* 7: 11190. doi:10.1038/ncomms11190

Zeng, M., Feng, A., Ren, Y., Zhang, Q., Wang, R., Zhang, B., et al. (2024). Salvia miltiorrhiza Bunge extract and Przewalskin ameliorate Bleomycin-induced pulmonary fibrosis by inhibition of apoptosis, oxidative stress and collagen deposition via the TGF- β 1 pathway. *Phytomedicine* 125: 155339. doi:10.1016/j.phymed.2024.155339

Zeng, Q., Gong, Y., Zhu, N., Shi, Y., Zhang, C., and Qin, L. (2024a). Lipids and lipid metabolism in cellular senescence: emerging targets for age-related diseases. *Ageing Res. Rev.* 97: 102294. doi:10.1016/j.arr.2024.102294

Zeng, Q., Luo, Y., Sang, X., Liao, M., Wen, B., Hu, Z., et al. (2024b). Senegenin attenuates pulmonary fibrosis by inhibiting oxidative-stress-induced epithelial cell senescence through activation of the sirt1/Pgc-1a signaling pathway. *Antioxidants (Basel)* 13, 675. doi:10.3390/antiox130606756).

Zhang, C., Chen, L., Xie, C., Wang, F., Wang, J., Zhou, H., et al. (2024). YTHDC1 delays cellular senescence and pulmonary fibrosis by activating ATR in an m6A-independent manner. *Embo J.* 43 (1), 61–86. doi:10.1038/ s44318-023-00003-2

Zhang, K., Wang, L., Hong, X., Chen, H., Shi, Y., Liu, Y., et al. (2021a). Pulmonary alveolar stem cell senescence, apoptosis, and differentiation by p53dependent and -independent mechanisms in telomerase-deficient mice. *Cells* 10 (11), 2892. doi:10.3390/cells10112892

Zhang, X., Dong, Y., Li, W. C., Tang, B. X., Li, J., and Zang, Y. (2021b). Roxithromycin attenuates bleomycin-induced pulmonary fibrosis by targeting senescent cells. *Acta Pharmacol. Sin.* 42 (12), 2058–2068. doi:10.1038/ s41401-021-00618-3

Zhou, J., Li, X. Y., Liu, Y. J., Feng, J., Wu, Y., Shen, H. M., et al. (2022). Full-coverage regulations of autophagy by ROS: from induction to maturation. *Autophagy* 18 (6), 1240–1255. doi:10.1080/15548627.2021.1984656

Zhu, Y., Tchkonia, T., Fuhrmann-Stroissnigg, H., Dai, H. M., Ling, Y. Y., Stout, M. B., et al. (2026). Identification of a novel senolytic agent, navitoclax, targeting the Bcl-2 family of anti-apoptotic factors. *Aging Cell* 15 (3), 428–435. doi:10.1111/acel.12445

Zuk, A., and Bonventre, J. V. (2016). Acute kidney injury. Annu. Rev. Med. 67: 293–307. doi:10.1146/annurev-med-050214-013407