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Discovery of paradoxical genes: reevaluating the prognostic impact of overexpressed genes in cancer

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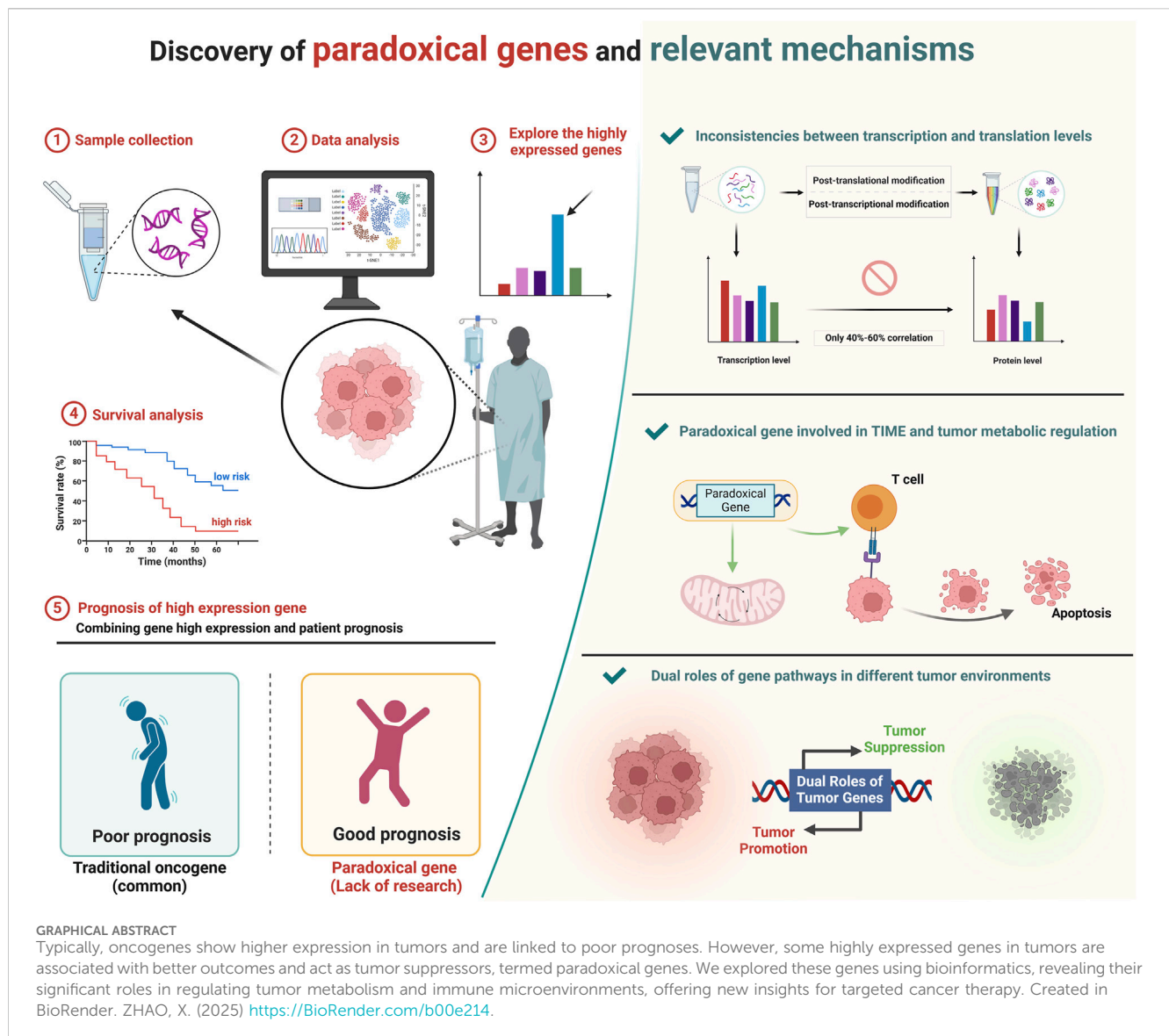
Oncogenes are typically overexpressed in tumor tissues and often linked to poor prognosis. However, recent advancements in bioinformatics have revealed that many highly expressed genes in tumors are associated with better patient outcomes. These genes, which act as tumor suppressors, are referred to as “paradoxical genes.” Analyzing The Cancer Genome Atlas (TCGA) confirmed the widespread presence of paradoxical genes, and KEGG analysis revealed their role in regulating tumor metabolism. Mechanistically, discrepancies between gene and protein expression—affected by pre- and post-transcriptional modifications—may drive this phenomenon. Mechanisms like upstream open reading frames and alternative splicing contribute to these inconsistencies. Many paradoxical genes modulate the tumor immune microenvironment, exerting tumor-suppressive effects. Further analysis shows that the stage- and tumor-specific expression of these genes, along with their environmental sensitivity, influence their dual roles in various signaling pathways. These findings highlight the importance of paradoxical genes in resisting tumor progression and maintaining cellular homeostasis, offering new avenues for targeted cancer therapy.

KEYWORDS

paradoxical genes, bioinformatics, tumor metabolism, discordant gene-protein abundance, tumor immune microenvironment, signaling pathway

1 Introduction

Traditionally, bioinformatics analyses often utilize the differential gene abundance between tumor and normal tissues to screen for target genes (Golub et al., 1999). Typically, genes with significantly higher abundance in tumors than normal tissues are classified as oncogenes and become focal points of research (Bishop, 1991; Croce, 2008). However, a new understanding has emerged with the proliferation of comprehensive databases such as TCGA. Researchers increasingly recognize that not all genes are highly expressed in tumors act as promoters of cancer (Cao et al., 2021; Yan et al., 2019; Boor et al., 2020; Martinez-Turtos et al., 2022). In fact, some genes show high abundance in tumor tissues but are associated with good patients prognosis (Cao et al., 2021; Yan et al., 2019; Boor et al., 2020; Martinez-Turtos et al., 2022). This finding challenges the traditional understanding of tumor biology. Although public databases provide evidence that genes which are highly expressed in tumor tissues and serve tumor suppressor roles are prevalent, they have not been broadly recognized or systematically analyzed by the scientific community. Our work pioneers the classification and definition of these genes as “paradoxical genes,” and it delves deeply into the reasons and context for the existence of these paradoxical genes.



The discrepancy between mRNA and protein levels may be a reason for the emergence of paradoxical genes (Vogel and Marcotte, 2012). This difference is mainly due to post-transcriptional and -translational modifications, which are crucial in the dynamic regulation of gene expression (Vogel and Marcotte, 2012). Post-transcriptional modifications include processes such as alternative splicing, enabling a single gene to generate multiple mRNA variants, thereby expanding the diversity of the proteome (Vogel and Marcotte, 2012; Wahl et al., 2009). The upstream open reading frames (uORFs) can significantly regulate the translation of the main open reading frame (ORF) (Barbosa et al., 2013; Johnstone et al., 2016). Concurrently, post-translational modifications such as phosphorylation and ubiquitination further diversify protein functions and regulation (Hershko and Ciechanover, 1998; Hunter, 2007). However, within the context of TCGA, the focus is solely on the measurement of mRNA abundance (Cancer Genome Atlas Research Network, 2008; Author anonymous, 2012; Weinstein et al., 2013). This is typically quantified using RNA sequencing data, which provides detailed information on the levels of mRNA present

in a given sample (Cancer Genome Atlas Research Network, 2008; Author anonymous, 2012; Weinstein et al., 2013). Inconsistencies between gene and protein levels can potentially distort patient prognosis results, contributing to the emergence of paradoxical genes (Akbari et al., 2014).

The tumor immune microenvironment (TIME) is critical for suppressing tumor progression, through its complex network of immune cells, stromal cells, signaling molecules, and extracellular matrix components (Joyce and Fearon, 2015; Fridman et al., 2012; Quail and Joyce, 2013). This dynamic environment can promote or inhibit tumor growth, depending on the balance of pro- and anti-tumor factors (Schreiber et al., 2011). Key players include cytotoxic T lymphocytes (CTLs), natural killer (NK) cells, dendritic cells, B cells, proinflammatory cytokines, and chemokines (Schreiber et al., 2011; Smyth et al., 2002; Palucka and Banchereau, 2012; Nelson, 2010; Müller et al., 2009; Topalian et al., 2015). There is currently evidence that some Paradoxical genes are involved in regulating TIME and inhibiting tumor progression (Cao et al., 2021; Yan et al., 2019; Boor et al., 2020; Martinez-Turtos et al., 2022). The

regulatory influence of paradoxical genes on TIME elucidates the mechanism underlying their tumor suppressor effects.

The expression of genes and pathways exhibits context-dependent effects, also known as context specificity, which refer to the phenomenon where the function or behavior of a gene varies depending on the specific context in which it operates (Feil and Fraga, 2012; Huang, 2009; Beyer et al., 2007). In the context of cancer, the role of a gene can vary significantly depending on factors such as the type of cancer, the tissue in which the tumor originates, and the stage of the cancer (Vogelstein and Kinzler, 2004; Hanahan and Weinberg, 2011; Stratton et al., 2009). For instance, genes associated with signaling pathways like TGF β , NOTCH, and NF- κ B demonstrate differential expression across various tumor tissues. The TGF β pathway presents a dual role, acting as a tumor suppressor in early stages and a promoter in advanced stages, with the expression varying significantly in breast, pancreatic, and colorectal cancers (Massagué, 2008). Similarly, the NOTCH signaling pathway, critical for cell fate determination and differentiation, shows oncogenic as well as tumor-suppressive functions, depending on the cancer type, as observed in breast cancer (BRCA), T-cell acute lymphoblastic leukemia, and lung cancer (Stylianou et al., 2006; Weng et al., 2004; Kopan and Ilgan, 2009). Often, the NF- κ B pathway, a key regulator of inflammation and immune responses, is dysregulated in BRCA, multiple myeloma, and colorectal cancer, where it facilitates cell proliferation, survival, and chronic inflammation (Karin, 2006; Annunziata et al., 2007; Greten et al., 2004). The specific roles and expression patterns of these similar pathways in different tumor environments are also one of the reasons for the widespread existence of paradoxical genes.

In this article, by exploring the mechanism of paradoxical genes formation, we seek to broaden the current understanding of tumor biology and provide new ideas for tumor treatment and research.

2 Paradoxical genes emerge as a key focus in bioinformatics research

When genes highly expressed in cancer compared to normal tissues, are identified through bioinformatics analyses, it often implies that these genes may act as drivers of oncogenesis, serve as diagnostic or prognostic biomarkers, or act as predictive markers for treatment response (Niu et al., 2022; Liu Y. et al., 2021). Indeed, the relationship between gene expression and cancer prognosis can be complex and sometimes counterintuitive (Hanahan and Weinberg, 2011). Contrary to what might be expected, high expression of certain genes in tumors can be associated with a more favorable prognosis (Hanahan and Weinberg, 2011). This paradoxical finding highlights the complexity of cancer biology, revealing that genes may play multifaceted roles in tumorigenesis and cancer progression (Hanahan and Weinberg, 2011). For instance, some genes highly expressed in tumors could participate in immune response activation, DNA repair mechanisms, or cellular differentiation processes, potentially inhibiting tumor growth or spread, thereby improving patient outcomes (Blagih et al., 2020; Williams and Schumacher, 2016; Kalluri and Weinberg, 2009). Researchers face a challenge of dissecting the dual roles that some genes, acting as oncogenes in

certain contexts while serving protective or suppressive functions in others (Hanahan and Weinberg, 2011).

2.1 Paradoxical genes exhibit differential and uneven expression across various tumors

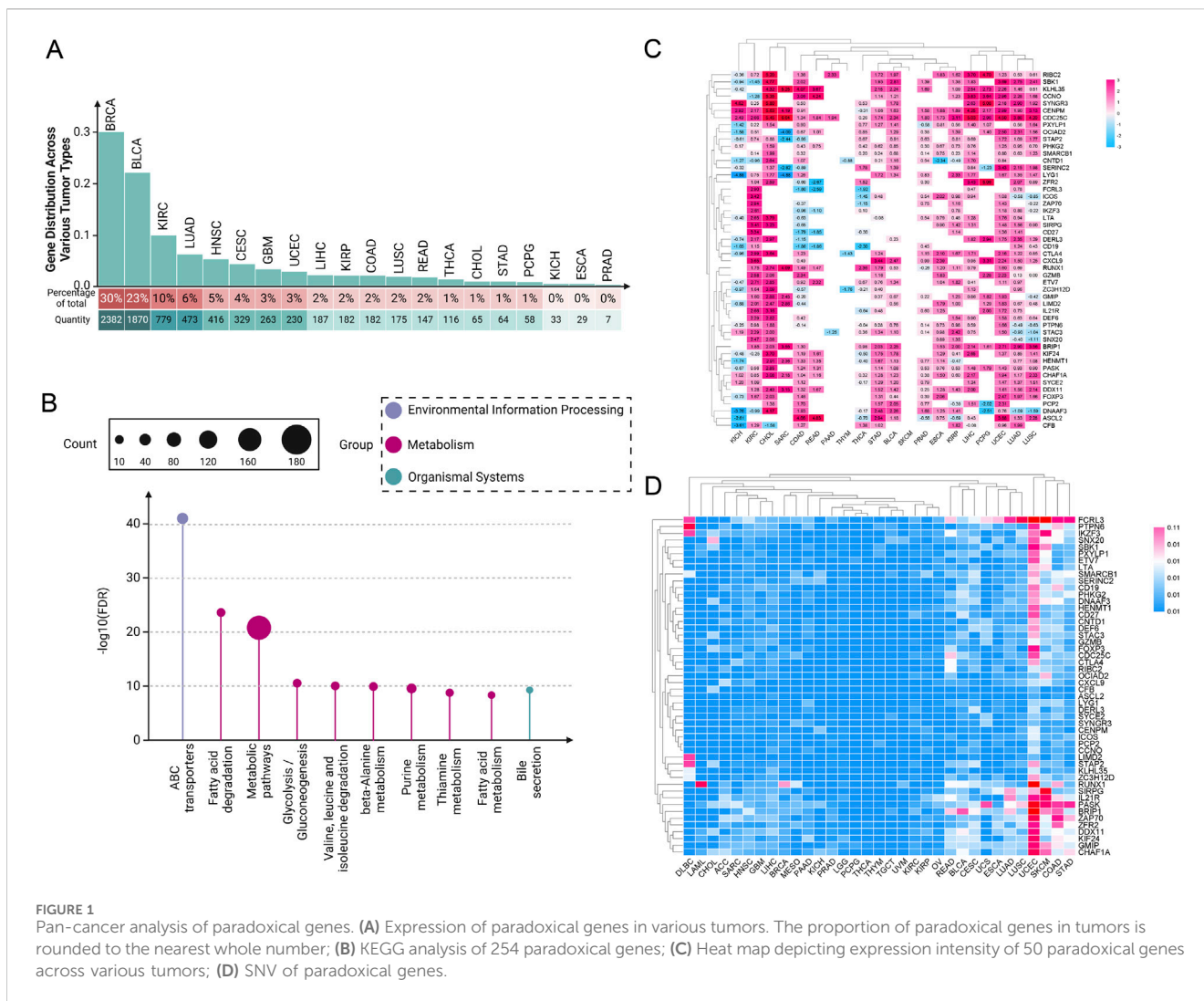
In recent years, our understanding of the molecular underpinnings of cancer has been revolutionized by the integration of genomic databases such as TCGA into cancer research (Tomczak et al., 2015). TCGA provides an extensive compilation of genetic mutations, gene expression data, and epigenetic alterations across thousands of tumors, spanning over 30 human tumor types (Weinstein et al., 2013; Tomczak et al., 2015). Through our analysis of the TCGA database, our team has determined that the role of paradoxical genes cannot be overlooked. We conducted a detailed analysis of the distribution of paradoxical genes across various tumor types, ranking them by the number of highly expressed genes within each tumor category (Figure 1A). We observed that this type of gene is ubiquitously present across various cancers. Notably, BRCA, bladder urothelial carcinoma, and kidney renal clear cell carcinoma (KIRC) exhibit the highest expression levels of these tumor suppressor genes. In contrast, kidney chromophobe (KICH), esophageal carcinoma (ESCA), and prostate adenocarcinoma (PRAD) have significantly lower expression of these genes.

This disparity suggests a complex regulatory mechanism involved in the expression of paradoxical genes, which the tumor microenvironment (TME) and specific oncogenic pathways could influence. The high expression levels of paradoxical genes in BRCA, BLCA, and KIRC suggest that these cancers possibly utilize these genes to balance between tumor suppression and oncogenic activity, potentially as a response to oncogenic stress or other cellular pressures. In contrast, the reduced expression of paradoxical genes in KICH, ESCA, and PRAD might indicate a loss of this balancing mechanism, possibly contributing to more aggressive tumor behavior. These findings provide a crucial direction for future research into the mechanisms regulating paradoxical genes and their role in cancer progression.

2.2 Kyoto encyclopedia of genes and genomes analysis of paradoxical genes: insights into their relationship with tumor metabolism

To investigate the functional mechanisms of paradoxical genes further, we initially screened 254 paradoxical genes for a KEGG analysis (Figure 1B). A common characteristic among these genes is their high expression in at least three tumor cell groups, correlating with improved patient prognosis. Our KEGG analysis revealed that these genes are extensively involved in various metabolism-related pathways, suggesting it as a primary mechanism through which paradoxical genes influence tumor prognosis.

Notably, liver X receptor (LXR) genes, including LXR α and LXR β , are a few examples of this phenomenon (Han et al., 2023; Wang et al., 2023). LXRs are nuclear receptors involved in lipid metabolism, inflammation, and cholesterol homeostasis (Zelcer and



Tontonoz, 2006). In the context of cancer, LXRs have demonstrated a dual role in tumor prognosis, influenced by their regulatory impact on metabolic pathways and immune responses in the tumor microenvironment (Han et al., 2023; Wang et al., 2023). LXR genes can play a role in inhibiting tumor progression (Zhang et al., 2014; Nguyen-Vu et al., 2013). This is primarily mediated through their anti-inflammatory effects within the tumor microenvironment. High expression of LXR genes causes upregulation of cholesterol efflux transporters such as ATP-binding cassette transporter A1 (ABCA1) and ABCG1, which facilitate cholesterol efflux and reduce lipid accumulation within macrophages, thus attenuating the inflammatory response associated with tumor progression (Joseph et al., 2003; Wang et al., 2006). Furthermore, LXR activation has been linked to the suppression of inflammatory cytokine production by immune cells, leading to a less conducive environment for tumor growth (Joseph et al., 2003; Fessler, 2016; Chawla et al., 2001). A study demonstrated that LXRs activation disrupts BRCA cell proliferation by downregulating the expression of genes involved in cell growth and proliferation, particularly those regulated by the E2F family of transcription factors (Nguyen-Vu et al., 2013). The activation of LXRs leads to the downregulation of key genes involved in the cell

cycle, DNA replication, and other critical processes for cancer cell division (Nguyen-Vu et al., 2013). This effect is partly mediated through the regulation of E2F2, highlighting a potential mechanism by which LXRs inhibit proliferation in cancer cells (Nguyen-Vu et al., 2013).

Conversely, LXRs can promote tumor growth through several mechanisms. Their activation leads to upregulation of genes involved in lipid biosynthesis, such as SREBP-1c (Sterol Regulatory Element-Binding Protein 1c) (Okazaki et al., 2010; Jeong et al., 2021). SREBP-1c is a crucial transcription factor that enhances the expression of genes required for fatty acid and triglyceride synthesis (Okazaki et al., 2010; Jeong et al., 2021). In many cancers, particularly those with high lipid requirements like breast cancer, this can contribute to tumor cell proliferation and survival by ensuring a steady supply of essential lipids that are critical for membrane synthesis, energy storage, and signaling (Santos and Schulze, 2012; Menendez and Lupu, 2007; Swinnen et al., 2006; Zadra et al., 2013; Fukuchi et al., 2004; Vedin et al., 2009). Our research (Figure 1A) demonstrated that paradoxical genes are prevalently expressed in breast cancer, which also supports the notion that the modulation of these genes, particularly their regulation of tumor lipid metabolism, is fundamental to their anticancer effects.

In our previous investigation of clear cell renal cell carcinoma (ccRCC), we further examined the dual role of LXR (Wu et al., 2019). Our findings suggest that LXR functions as a balance gene, where both heightened and diminished expression levels can exert inhibitory effects on ccRCC progression (Wu et al., 2019). Specifically, both LXR agonists and inverse agonists inhibit cell proliferation and colony formation. The LXR agonist LXR623 downregulates low-density lipoprotein receptor (LDLR) and upregulates ABCA1, causing a decline in intracellular cholesterol and induced apoptosis (Wu et al., 2019). Conversely, the LXR inverse agonist SR9243 downregulates key FA synthesis proteins, including sterol regulatory SREBP-1c, FA synthase, and stearoyl-CoA desaturase 1 (SCD1), leading to decreased FA content and apoptosis in ccRCC (Wu et al., 2019). This phenomenon illustrates that alterations in cancer metabolism are a pivotal factor in mediating the regulatory effects of paradoxical genes on tumor prognosis.

2.3 Expression intensity and single nucleotide variations (SNV) analysis of paradoxical genes across different tumors

Subsequently, we screened out 50 paradoxical genes for pan-cancer analysis (Figure 1C). Their common feature is that they are highly expressed in ≥ 4 groups of tumor cells, and are associated with better patient prognosis. The expression intensity of these genes was analyzed in 20 different tumor tissues, and we found widespread overexpression, including, but not limited to, KIRC, cholangiocarcinoma (CHOL), stomach adenocarcinoma (STAD), BLCA, PRAD, ESCA, and liver hepatocellular carcinoma (LIHC), highlighting their potential as pan-cancer prognostic markers. Interestingly, subsequent analysis of SNVs in these genes showed that mutations are infrequent, a characteristic not generally observed in traditional oncogenes (Figure 1D). This observation further supports the hypothesis that the prognostic effect of paradoxical genes is mediated through mechanisms distinct from those employed by oncogenes, potentially driving tumor evolution towards less aggressive and more treatable forms. Nonetheless, there are currently very few reports on the occurrence of highly expressed tumor suppressor genes in tumors. Uncovering previously underappreciated complexities in the relationship between gene expression and cancer prognosis is critical.

3 The relationship between gene abundance and protein abundance: is there always a direct correlation?

The central dogma of molecular biology, formulated by Francis Crick, describes the flow of genetic information from DNA to RNA to protein through the processes of transcription and translation (Crick, 1970). While it is generally hypothesized that higher mRNA levels correlate with higher protein levels, this relationship is influenced by several factors (Vogel and Marcotte, 2012; Schwanhäusser et al., 2011; Tian et al., 2004). Post-transcriptional regulation can modify mRNA stability and translation efficiency, as well as the sequence features of the mRNA itself, such as upstream

ORFs, can affect how efficiently it is translated (Barbosa et al., 2013; Sonenberg and Hinnebusch, 2009). Protein stability and degradation processes further modulate the levels of functional protein in the cell (Ciechanover and Kwon, 2015; Sherman and Goldberg, 2001). Although studies have typically shown a positive correlation between mRNA and protein levels, the variability suggests that multiple mechanisms, including translation efficiency and protein stability, are significant in determining final protein levels in biological systems (Schwanhäusser et al., 2011; Liu et al., 2016). Several studies have reported average correlation coefficients around 40%–60%, indicating that mRNA levels indicate protein abundance but are far from perfectly predictive due to various biological and methodological confounders (Vogel and Marcotte, 2012; Perl et al., 2017; Kosti et al., 2016) (Figure 2). This makes studies involving the gene level resulting from bioinformatics analysis somewhat one-sided.

3.1 The insights from the effect of upstream open reading frames

The concept of mRNA translation primarily involves coding mRNA into proteins by ribosomes, a process central to gene expression (Kozak, 1999; Jackson et al., 2010). Typically, this decoding focuses on the ORF that starts with a start codon (usually AUG) and ends with a stop codon (Jackson et al., 2010; Kozak, 2001). The discovery and study of uORFs have expanded our understanding of translational regulation and its complexities (Barbosa et al., 2013; Calvo et al., 2009; Wethmar, 2014). uORFs are alternative ORFs located upstream of the in the 5' untranslated region (5'UTR) of main coding sequence of an mRNA (Calvo et al., 2009). These uORFs can play a significant role in the regulation of translation of the main ORF (Barbosa et al., 2013; McGillivray et al., 2018).

uORFs are initiated when a ribosome recognizes and binds to a start codon (usually AUG, but sometimes a near-cognate codon) at the 5'UTR of an mRNA (Silva et al., 2019). The presence of a uORF upstream of the main coding sequence can alter ribosomal scanning and initiation dynamics, consequently affecting the translation of the downstream ORF (Hinnebusch et al., 2016; Morris and Geballe, 2000). After a uORF is translated, ribosomes can either dissociate from the mRNA or resume scanning for another start codon (Kozak, 2005). Several factors affect the ability of ribosomes to re-initiate translation at the downstream main ORF depends on, including the length of the uORF, the distance between the cistrons (the gap between the uORF and the main ORF), and the sequence context around the stop codon of the uORF and the start codon of the main ORF (Jackson et al., 2010; Ivanov et al., 2010; Kozak, 1987) (Figure 2). Often, efficient re-initiation is contingent upon the ribosomal retention of initiation factors during the uORF translation. The impact of a uORF on the main ORF translation can vary dramatically depending on its sequence and context (Pavitt, 2005). Some uORFs exhibit features that stall ribosomal function or slow translation, potentially enhancing or inhibiting the translation of the main ORF (Caliskan et al., 2015). For instance, Phan et al. elucidate how conserved uORFs in the 5'UTR of Polo-like kinase 4 (PLK4) mRNA play a crucial role in controlling the translation of PLK4, thereby regulating the duplication of centriole in primordial

Differences between mRNA abundance and protein abundance

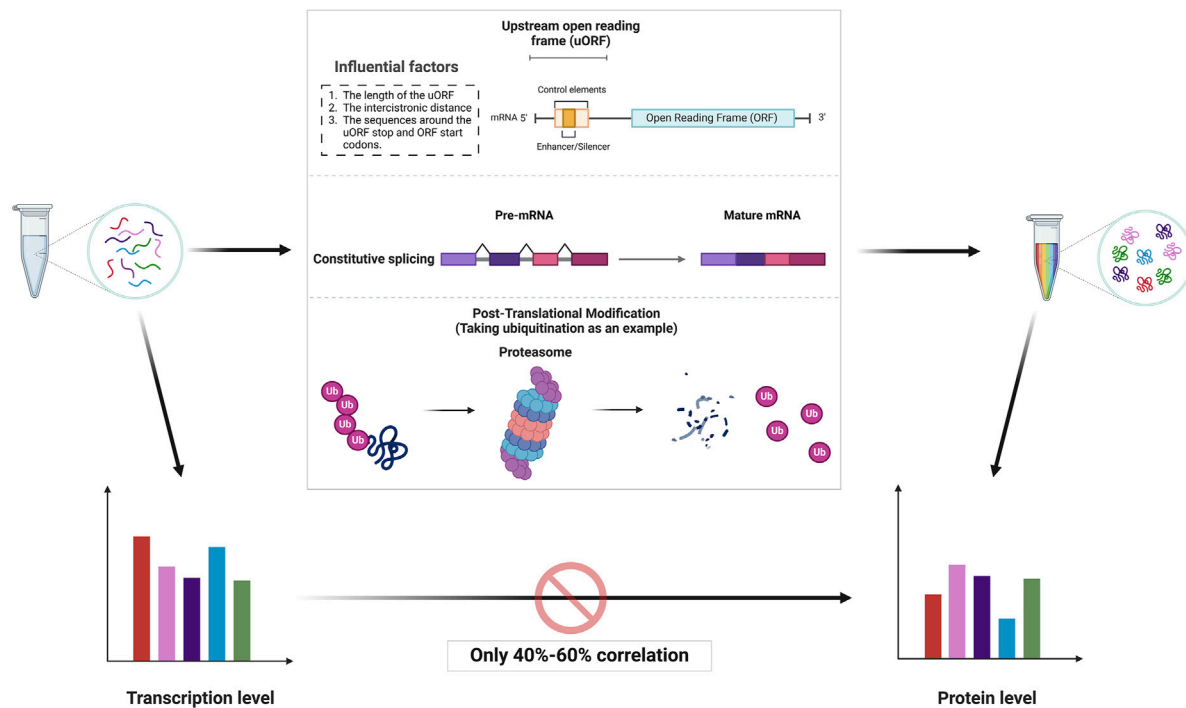


FIGURE 2 Post-transcriptional and post-translational modifications contribute to discrepancies between mRNA abundance and protein abundance. Created in BioRender. ZHAO, X. (2025) <https://BioRender.com/q14t025>.

germ cells (PGCs) and preserving genomic integrity (Phan et al., 2022). This translational control mechanism prevents excessive PLK4 synthesis, vital for preventing centriole amplification and associated mitotic errors, highlighting a specific requirement for uORF in regulating the balance of PLK4 levels during germ cell development (Phan et al., 2022). A recent study by Cieřla et al. reveals that the regulation of SF3B1 protein levels through ALKBH5-driven N6-methyladenosine demethylation in the 5'UTR influences its translation, driving splicing mechanisms that impact DNA repair and epigenetic regulation (Cieřla et al., 2023). These studies demonstrate the critical role of post-transcriptional modifications in the expression of final protein.

3.2 The insights from alternative splicing

Alternative splicing is a post-transcriptional regulatory mechanism that contributes significantly to proteomic diversity and gene expression regulation in eukaryotic organisms (Nilsen and Graveley, 2010; Wang et al., 2008; Black, 2003; Barash et al., 2010). It involves the selective inclusion or exclusion of pre-mRNA segments (exons) during the RNA splicing process, resulting in multiple distinct mRNA transcripts from a single gene (Chen and Manley, 2009; Kornbliht et al., 2013) (Figure 2). This process can affect the quantity as well as the functionality of the encoded proteins (Kornbliht et al., 2013). When determining RNA abundance using technologies like RNA-seq, reads mapping to a

gene are typically aggregated to estimate the overall abundance of the gene (Wang et al., 2009; Mortazavi et al., 2008). This standard approach does not differentiate between the various transcripts produced by alternative splicing (Ozsolak and Milos, 2011; Trapnell et al., 2009). Consequently, even if the total mRNA of a gene remains constant, changes in the splicing patterns can lead to proteins with significantly altered types and functions (Black, 2003; Kalsotra and Cooper, 2011). This is a critical factor to consider in gene expression analysis because while the quantitative measure (total RNA transcripts) might not show variation, the qualitative changes (different splice variants) can have profound biological ramifications (Nilsen and Graveley, 2010; Barash et al., 2010). Meanwhile, specific conditions or stimuli might induce changes in splicing patterns without altering the overall mRNA levels (Wang et al., 2008; David and Manley, 2010). Such differential splicing events can produce protein variants with differing, sometimes opposing, functions (Wang et al., 2008; David and Manley, 2010). Some splice variants may include or exclude sequences with regulatory elements affecting translation efficiency, such as internal ribosome entry sites or uORFs (Barbosa et al., 2013; Sonenberg and Hinnebusch, 2009).

In a recent study, researchers demonstrated the significant role of alternative splicing in the regulation of chromatin dynamics, particularly through the manipulation of histone deacetylase (HDAC)7 splicing downstream of T cell signaling pathways (Agosto et al., 2023). Notably, the longer HDAC7 isoform, induced by the RNA-binding protein CUGBP Elav-like family

member 2, enhances the expression of key T cell surface proteins such as CD3, CD28, and CD69, highlighting the broad implications of alternative splicing on histone modification and gene regulatory mechanisms in T cells (Agosto et al., 2023). Particularly in studies related to diseases such as cancer, where splicing patterns can be drastically altered, researchers must consider the total expression level of a gene as well as the expression levels of individual splice variants.

3.3 The insights from post-translational regulation

Differences in protein abundance and gene abundance are largely caused by post-translational modifications (PTMs). PTMs like ubiquitination and phosphorylation can target proteins for degradation, leading to lower protein levels despite high mRNA expression (Hershko and Ciechanover, 1998; Hunter, 2007; Cohen, 2000; Ciechanover, 2005; Deshaies and Ferrell, 2001). Conversely, protein levels rise when modifications protect proteins from degradation. PTMs also modulate protein activity, producing active or inactive forms that do not directly correlate with mRNA levels (Figure 2). For instance, phosphorylated proteins often exhibit functions or stabilities different from their non-phosphorylated counterparts (Hunter, 2007; Olsen et al., 2006; Manning et al., 2002). Modifications such as phosphorylation, methylation, and acetylation also impact protein-protein interactions, altering binding affinities and affecting signaling pathways and cellular processes independent of gene expression (Ross et al., 2023; Nishi et al., 2014; Duan and Walther, 2015). The regulatory mechanism of post-translational modifications is comprehensively summarized in Table 1.

Integrative multi-omics analysis highlights the significant impact of PTMs on the differences in protein and gene levels, particularly during the human cell cycle (Parkes and Niranjana, 2019). While mRNA and translation data explain some variations in protein abundance, the remaining inconsistencies are primarily due to PTMs, which adjust protein levels post-synthesis (Parkes and Niranjana, 2019). In a study focusing on triple-negative breast cancer (TNBC), researchers identified tumor endothelial marker 8 (TEM8) as a key indicator of breast tumor-initiating cells (Xu et al., 2021). The study also highlighted the binding of estrogen receptor α to the promoter region of the ubiquitin E3 enzyme ankyrin repeat and SOCS box containing 10 (ASB10). It also activates ASB10 transcription, and ASB10 interacts with TEM8, thereby affecting the ubiquitination of TEM8 and ultimately affecting the TEM8 protein level (Xu et al., 2021). Thus, Indirect evidence indicates that variations in TEM8 mRNA and protein expression across BRCA subtypes may be attributed to post-translational modifications (Xu et al., 2021).

4 Regulation of tumor immune microenvironment by paradoxical genes

The process of TIME begins with the recognition of tumor-specific antigens by antigen-presenting cells, like dendritic cells, which capture and present these neoantigens to naïve T cells in lymph nodes, thereby initiating T-cell activation (Mellman et al.,

2011). This activation triggers a series of immune responses, including the release of chemokines that attract more effector immune cells (such as CTLs, NK cells, and macrophages) to the tumor site, effectively infiltrating the tumor (Joyce and Fearon, 2015). Within the TIME, a pivotal change occurs as effector T cells reprogram immunosuppressive cells and alter the metabolic environment, diminishing the suppressive function of regulatory immune cells such as regulatory T cells (Tregs), myeloid-derived suppressor cells (Ho et al., 2015). This culminates in the direct cytotoxic attack on tumor cells by CTLs and NK cells, utilizing mechanisms like perforin and granzyme release to induce tumor cell apoptosis (Trapani and Smyth, 2002). Furthermore, some activated T cells differentiate into memory T cells, providing long-term surveillance and a rapid response mechanism against tumor recurrence (Sallusto et al., 2004) (Figure 3).

Recently, several scholars have dedicated their research efforts to understanding how paradoxical genes influence prognosis through the regulation of the TIME. Cao et al. analyzed data from TCGA and the gene expression omnibus, revealing that the expression of C-X-C motif chemokine ligand (CXCL)11 was elevated in colon cancer tissues compared to healthy tissues, and higher levels of CXCL11 correlated with improved survival outcomes (Cao et al., 2021). Furthermore, assessment of three independent datasets, including TCGA and two single-cell RNA sequencing datasets from Gene Expression Omnibus, in addition to immunohistochemistry data from a COAD patient cohort demonstrated that this tumor suppressor effect possibly due to its association with an increased presence of antitumor immune cells (CD8⁺ T cells, and CD56 NK cells), underscored CXCL11's role in modulating the TIME (4). Furthermore, HERV-H LTR-associating 2 (HHLA2), a newly identified member of the B7 immune checkpoint family, is also a typical paradoxical gene (Yan et al., 2019). HHLA2 is minimally expressed in normal pancreatic tissues but shows significant upregulation from precancerous stages to invasive pancreatic ductal adenocarcinoma (PDAC), according to immunohistochemistry analyses on tissue microarrays (Yan et al., 2019). In 77.17% of PDAC cases, the expression of HHLA2 is strongly associated with an improved post-surgical prognosis, indicating functions of HHLA2 as a costimulatory ligand in pancreatic cancer, activating CD8⁺ T cell proliferation and improving patient prognosis (Yan et al., 2019). A subsequent study also presented a similar point; immunohistochemical analysis on tissue micro-arrays from surgically resected tumors of 122 pancreatic and 72 ampullary cancer patients revealed HHLA2 expression in 67% of pancreatic and 93% of ampullary tumors, associating enhanced expression with improved post-surgical outcomes, including delayed cancer recurrence and improved cancer-specific survival (Boor et al., 2020). Similarly, the study by Martinez-Turtos et al. highlights that overexpression of inositol-requiring enzyme 1 α (IRE1 α) in murine colorectal and Lewis lung carcinoma cells in syngeneic immunocompetent mice, leads to a tumor-suppressive phenotype (Martinez-Turtos et al., 2022). This anti-tumoral effect is attributed to the RNase activity of IRE1 α , which induces apoptosis in tumor cells, enhances adaptive anti-cancer immunosurveillance through XBP1 mRNA splicing, and regulates IRE1-dependent degradation of RNA (RIDD) (Martinez-Turtos et al., 2022). However, in addition to the tumor suppressive role of IRE1 α , its tumor-promoting role is also evident in preclinical

TABLE 1 Overview of post-translational modifications in protein regulation.

| PTM | Mechanism | Regulation direction | Biological effects | Clinical significance | References |
|-----------------|--|------------------------------------|--|--|---|
| Phosphorylation | Addition of phosphate groups to amino acids (Ser, Thr, Tyr) | Activate or inhibit | Regulates enzyme activity, signal transduction, cell cycle, apoptosis | Targeting phosphorylation pathways is a strategy in cancer therapy | Hunter (2007), Singh et al. (2017) |
| Ubiquitination | Attachment of ubiquitin to lysine residues | Usually leads to degradation | Controls protein turnover, modulates signaling pathways, cellular stress responses | Central in neurodegenerative diseases like Alzheimer's and Parkinson's, and cancer therapies | Popovic et al. (2014) |
| Acetylation | Addition of acetyl groups to lysine residues | Can activate or stabilize proteins | Influences gene expression, enzyme activity, protein stability, and metabolic regulation | Targeted by HDAC inhibitors in cancer treatment | Shvedunova and Akhtar (2022), Drazic et al. (2016), He et al. (2023) |
| Methylation | Addition of methyl groups to lysine or arginine | Can activate or repress | Affects protein interaction, stability, DNA binding, and transcriptional regulation | Targeted by drugs that modify methylation dynamics (inhibitors of methyltransferases and demethylases) | Greer and Shi (2012), Dawson and Kouzarides (2012), Klose and Zhang (2007) |
| Sumoylation | Addition of SUMO proteins to lysine residues | Typically inhibits | Regulates nuclear-cytosolic transport, transcriptional activity, DNA repair | Implicated in cancer and heart disease | Flotho and Melchior (2013), Geiss-Friedlander and Melchior (2007), Chang and Yeh (2020) |
| Prenylation | Attachment of lipid groups (farnesyl or geranylgeranyl) to cysteine residues at the C-terminus of proteins | Generally activates | Facilitates membrane attachment, affects protein localization and function in signaling | Targeted in anti-cancer therapies, especially in Ras-related cancers | Jung and Bachmann (2023), Baranyi et al. (2020) |

PTM, post-translational modifications; HDAC, histone deacetylase; SUMO, small ubiquitin-like modifier; Ser, serine; Thr, threonine; Tyr, tyrosine.

models of various cancers, including TNBC, PDAC, and colon cancer (Harnoss et al., 2020; Garcia-Carbonero et al., 2018; Xie et al., 2019). This duality suggests that the impact of Paradoxical genes on cancer prognosis may be multifaceted and not solely affected by the TIME (Figure 3).

5 Dual roles of paradoxical genes and associated signaling pathways in tumors

Certain genes and their associated signaling pathways exhibit bidirectional effects on tumor prognosis, transitioning between inhibiting and promoting tumor progression. This dualistic behavior is also one of the key factors contributing to the emergence of paradoxical genes (Hanahan and Weinberg, 2011; Sadikovic et al., 2008). This biphasic regulatory effect can depend on various factors, including tumor stage, tumor-specific expression, and TME (Hanahan and Weinberg, 2011; Plaks et al., 2015; Gerlinger et al., 2012; Marusyk et al., 2012; Bray, 2016). The impact of gene expression can vary by cancer type and the tissue of origin (Stratton et al., 2009). Genes beneficial in one type of cancer might be deleterious in another (Vogelstein and Kinzler, 2004; Hanahan and Weinberg, 2011; Sadikovic et al., 2008). The role of E-cadherin in tumor suppression is well-established in BRCA due to its function in maintaining cell-cell adhesion and inhibiting metastasis (Padmanaban et al., 2019). However, in other cases, such as gastric cancer, the expression of can be associated with different outcomes based on additional factors like the presence of specific mutations or the overall state of cellular adhesion molecules (Becker et al., 1994). We focuses on signaling pathways with biphasic

regulatory effects, including the TGF β , NOTCH, and NF- κ B pathways (Figure 4). Genes associated with these pathways are key contributors to the development of paradoxical genes, which exhibit peculiar behaviors during gene expression and regulation.

6 Stage-specific expression of paradoxical genes: insights from the transforming growth factor- β signaling pathways

The expression of tumor suppressor genes exhibits significant variability across different cancer stages, contributing to the phenomenon of paradoxical genes (Sherr and McCormick, 2002). For instance, in the early stages of cancer, key genes like TP53 and RB1 play a crucial role in maintaining cellular integrity by regulating DNA damage repair and controlling cell cycle progression (Sherr and McCormick, 2002). However, as the tumor evolves, these genes often become inactivated due to mutations, deletions, or epigenetic modifications, leading to unchecked cell proliferation and advancement to more aggressive stages (Li et al., 1997; Cantley and Neel, 1999). PTEN, which regulates the PI3K/AKT signaling pathway, is commonly altered in cancers such as prostate and breast cancer, thereby facilitating tumor growth and survival (Li et al., 1997; Cantley and Neel, 1999; Salmena et al., 2008). In later stages, the suppression of tumor suppressor genes can precipitate metastasis and resistance to treatment, severely worsening the prognosis (Valastyan and Weinberg, 2011; Gottesman, 2002; Hanahan and Weinberg, 2000). Notably, the TGF- β pathway is recognized for its dual role in oncogenesis, acting as a tumor suppressor in initial stages while potentially fostering cancer

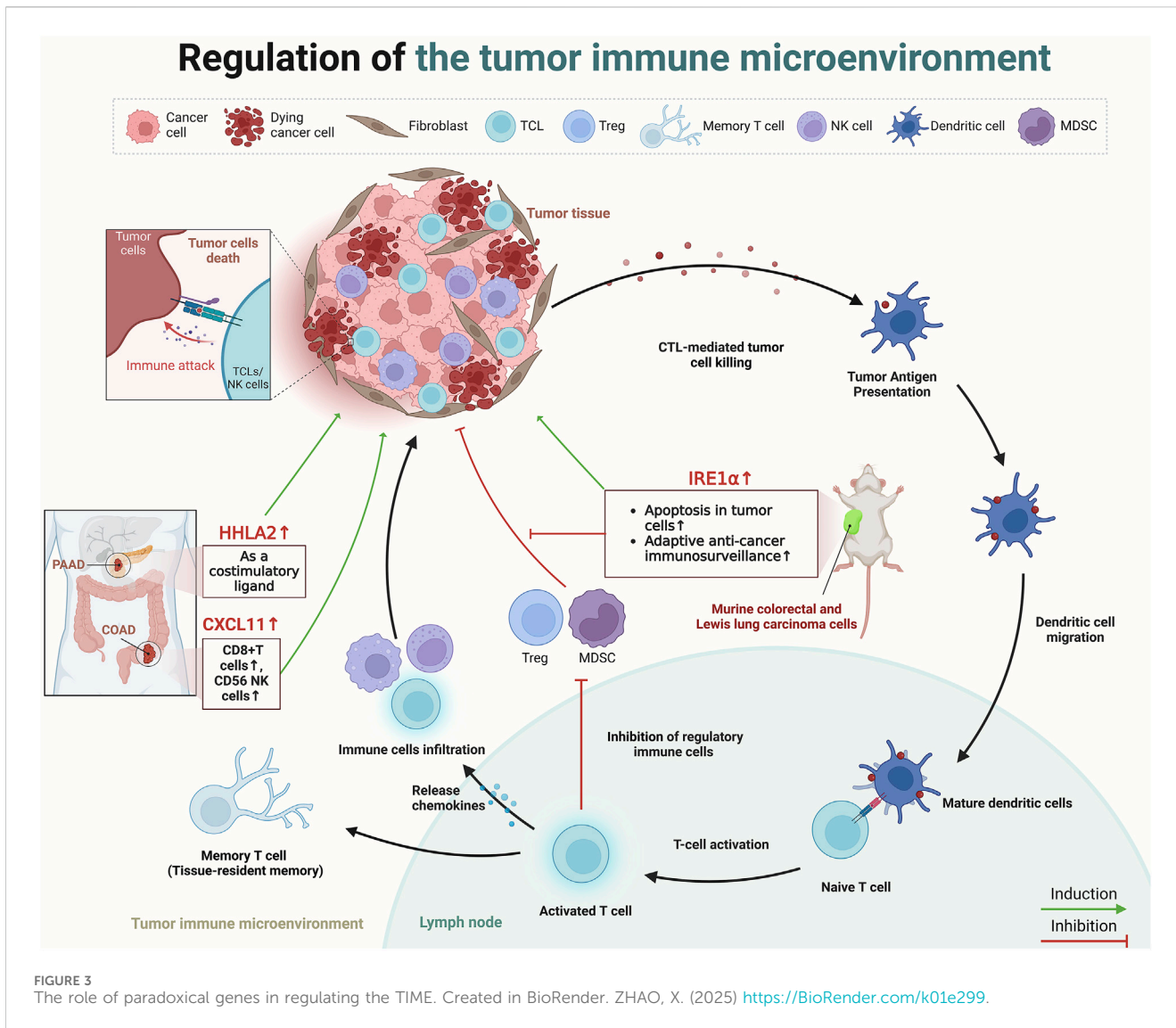


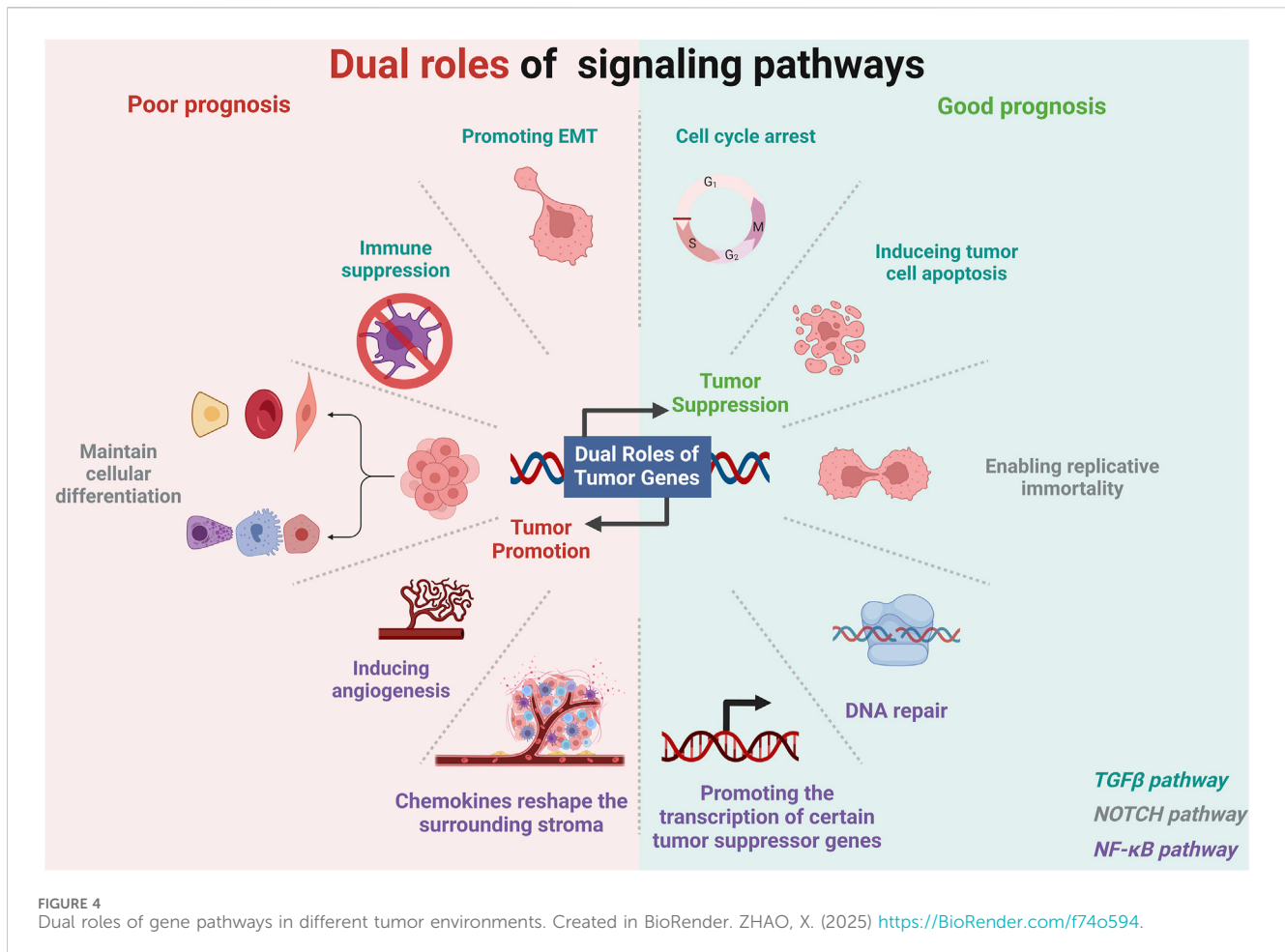
FIGURE 3 The role of paradoxical genes in regulating the TIME. Created in BioRender. ZHAO, X. (2025) <https://BioRender.com/k01e299>.

progression in advanced stages (Massagué, 2008). Within this pathway, SMAD family members, including SMAD2 and SMAD4, are pivotal in relaying TGF-β signals that suppress cell division (Derynck and Zhang, 2003).

The TGFβ pathway suppresses tumors in the early stages of tumor development mainly by maintaining cellular homeostasis (including cell cycle arrest and apoptosis) and preventing uncontrolled cell proliferation (Colak and Ten Dijke, 2017). TGFβ regulates the cell cycle by inhibiting the transition from the G1 phase to the S phase, thereby preventing DNA replication and cell division, which is facilitated through the upregulation of cyclin-dependent kinase (CDK) inhibitors, which deactivate CDKs essential for cell cycle progression (Derynck, 1994; Decker et al., 2021). TGFβ also induces apoptosis, or programmed cell death through the activation of death-associated proteins and modulation of apoptosis-related genes, eliminating cells with potentially harmful mutations (Zhao et al., 2018; Schulte-Hermann et al., 1992). It also maintains cellular differentiation and proper morphology, thereby inhibiting the epithelial-mesenchymal transition (EMT), a critical process in cancer metastasis (Hao et al., 2019). Furthermore, TGFβ

exerts anti-inflammatory effects within the cellular environment, regulating immune cell activity and cytokine production to suppress chronic inflammation, thus preventing tumor growth (Viel et al., 2016; Coussens and Werb, 2002).

In advanced cancer, TGF-β primarily acts as a tumor promoter. TGFβ promotes the EMT, a critical process for metastasis, by regulating transcription factors like Snail, Slug, and Twist that modify adhesion and migration properties of the cell (Peng et al., 2022; Wang et al., 2013; Ang et al., 2023). Simultaneously, TGF-β exerts systemic immune suppression and inhibits host immunosurveillance and also regulates the infiltration of inflammatory/immune cells and cancer-associated fibroblasts in the TME, causing direct changes in tumor cells (Yang et al., 2010). Neutralizing TGF-β enhances CD8+ T-cell- and NK-cell-mediated anti-tumor immune responses and increases the neutrophil-attracting chemokine production, leading to the recruitment and activation of neutrophils with an antitumor phenotype (Yang et al., 2010). It also interacts with cancer-associated fibroblasts and mesenchymal stem cells within the TME to remodel the extracellular matrix, increasing tumor stiffness and spreading cancer (Arima et al., 2023) (Figure 4).



The development of inhibitors that target TGF β signaling is a promising treatment approach for cancers where TGF β promotes tumor growth and metastasis (Derynck and Akhurst, 2007; Herberitz et al., 2015). These inhibitors generally block TGF β receptors, preventing the downstream signaling cascades that lead to oncogenic effects (Herberitz et al., 2015). SB-431542 was initially identified as a potent and specific inhibitor of the activin receptor-like kinase (ALK)4, ALK5, and ALK7 type I receptors of the TGF- β superfamily, effectively and selectively inhibiting activin and TGF- β signaling without impacting BMP signaling or other divergent pathways like extracellular signal-regulated kinase (ERK), c-Jun N-terminal kinase (JNK), or p38 mitogen activated protein kinase (Inman et al., 2002). In recent years, galunisertib (LY2157299 monohydrate), a selective, small molecule that can be taken orally to inhibit TGF- β receptor I kinase, exhibits antitumor activity across various cancer models, including breast, colon, lung, and HCC, by specifically downregulating SMAD2 phosphorylation and inhibiting the canonical TGF- β pathway (Herberitz et al., 2015). This drug is currently being evaluated in clinical trials in an intermittent dosing regimen (14 days on/14 days off, on a 28-day cycle) as part of monotherapy or in combination with other antitumor treatments to balance efficacy and safety, targeting cancers with high unmet medical needs like glioblastoma, pancreatic cancer, and HCC (Herberitz et al., 2015; Akhurst and Hata, 2012; Nadal et al.,

2023). Clinical trials investigating combinations of TGF β inhibitors with programmed cell death protein 1 (PD-1)/programmed death-ligand 1 (PD-L1) inhibitors are exploring this approach, showing promising results in improving anti-tumor immunity and patient outcomes (Wrzesinski et al., 2007). Hence, understanding the tumor stage and the specific role of TGF β is crucial to determining when and how to target this pathway effectively. Personalizing treatment based on the genetic and molecular profiles of individual tumors could optimize the efficacy of TGF β inhibitors and minimize adverse effects.

7 Tumor-specific expression of paradoxical genes: insights from the NOTCH signaling pathway

The NOTCH signaling pathway is a crucial cell communication mechanism that influences various biological processes, such as differentiation, proliferation, apoptosis, and stem cell maintenance (Bray, 2016). This pathway involves NOTCH receptors (NOTCH1-4) interacting with delta-like (DLL1, DLL3, DLL4) or Jagged (JAG1, JAG2) ligands on adjacent cells, initiating proteolytic cleavages that release the NOTCH intracellular domain (NICD) (Bray, 2016). This domain moves to the nucleus, where it converts recombination signal binding protein for immunoglobulin kappa J region from a repressor to

an activator, with mastermind-like proteins, initiating transcription of target genes families like HES and HEY (Bray, 2016). The finely tuned regulation of this pathway, which includes endocytic trafficking and post-translational modifications, is essential for maintaining cellular and tissue homeostasis (Fortini, 2009). The dual nature of NOTCH signalling in cancer biology—acting as a tumor suppressor in some contexts while promoting tumor progression in others—underscores the complexity of its signaling pathways and their diverse effects on cancer etiology and progression (Valdez and Xin, 2013) (Figure 4).

In specific cancers like those of skin and liver, NOTCH signaling is essential for maintaining cellular differentiation and tissue architecture (Panelos and Massi, 2009; Kawaguchi and Kaneko, 2021). Specifically, in squamous cell carcinoma, increased NOTCH signaling is associated with reduced tumor formation and progression (Panelos et al., 2008). Impaired NOTCH signaling, as demonstrated by the expression of the pan-NOTCH inhibitor dominant negative mastermind-like (DNMAML)1 in conditional transgenic mice, leads to hyperplastic epidermis and spontaneous development of cutaneous squamous cell carcinoma (SCC) and actinic keratoses, suggesting a protective role of canonical NOTCH signaling against cutaneous SCC (Proweller et al., 2006). Meanwhile, NOTCH1 signaling significantly inhibits the growth of HCC by inducing cell cycle arrest at the G (0)/G (1) phase and promoting apoptosis, by downregulating key cell cycle proteins and upregulating p21 and p53, while also suppressing antiapoptotic B-cell lymphoma 2(Bcl-2) expression (Giovannini et al., 2016; Giovannini et al., 2014; Kim et al., 2017; Qi et al., 2003; Viatour et al., 2011). However, the role of NOTCH signaling is paradoxically reversed in other types of cancers. In T-cell acute lymphoblastic leukemia (T-ALL) and certain breast cancers, NOTCH activation enhances cell proliferation, survival, and stemness, thereby promoting tumor growth and survival (Weng et al., 2004; Reedijk et al., 2005). Besides, the NOTCH signaling pathway also presents a biological dual nature widely in various cancers (Table 2).

8 The impact of environmental sensitivity on the formation of paradoxical genes

The environmental sensitivity of suppressor genes refers to the fact that the expression and function of these genes are affected by the TME, including factors such as hypoxia and acidity, which in turn affects cancer progression and cellular behavior (Gatenby and Gillies, 2004; Webb et al., 2011). Additionally, immune cells within this microenvironment release a variety of cytokines and growth factors that significantly impact cancer dynamics (Coussens and Werb, 2002; Grivennikov et al., 2010; de Visser et al., 2006). Specifically, certain immune-derived factors may suppress tumor cells, while others might activate signaling pathways that induce tumor cells (Coussens and Werb, 2002; Grivennikov et al., 2010; de Visser et al., 2006). These regulatory effects of environmental sensitivity on tumor suppressor genes undoubtedly contributed to the emergence of paradoxical genes.

8.1 Insights from hypoxia

Hypoxia within the TME critically influences the progression of cancer by modulating tumor suppressor genes, primarily via the

stabilization of hypoxia-inducible factors (HIFs) such as HIF-1 α and HIF-2 α (Semenza, 2003; Keith et al., 2011). Under low oxygen conditions, these transcription factors translocate to the nucleus, activating genes that drive angiogenesis, metabolism, cell survival, and invasion (Semenza, 2003). The suppression of the VHL gene under hypoxic conditions leads to unregulated HIF activity, promoting the secretion of angiogenic factors like vascular endothelial growth factor and platelet-derived growth factor, which are instrumental in tumor growth and proliferation (Semenza, 2003). Hypoxia plays a significant role in the regulation of tumor suppressor genes. For instance, Chen et al. discovered that Hypoxia-inducible HIF-1 α directly interacts with Mdm2, enhancing the *in vivo* association between p53 and HIF-1 alpha and acting as a mediator in their indirect interaction, which is crucial for the stabilization and activation of p53 in response to hypoxic stress (Chen et al., 2003). Furthermore, they found that HIF-1 alpha inhibits the Mdm2-mediated ubiquitination and nuclear export of p53, thereby protecting p53 from degradation and facilitating its role in transcriptional activation under hypoxic conditions (Chen et al., 2003). Additionally, PTEN, a crucial regulator of the PI3K/AKT signaling pathway, is downregulated by microRNAs like miR-21, which are themselves upregulated under hypoxic conditions. This indicates that hypoxia indirectly plays a significant role in the regulation of PTEN through the modulation of microRNA levels (Cascio et al., 2010; Kulshreshtha et al., 2007; Meng et al., 2007).

Hypoxic stress within solid tumors profoundly influences epigenetic regulation, particularly affecting DNA methylation and histone modifications (Shahrzad et al., 2007; Krieg et al., 2010). For instance, Watson et al. investigated the effects of chronic hypoxia in prostate cells and identified significant epigenetic changes, including increased global DNA methylation and H3K9 histone acetylation, associated with an altered cellular phenotype characterized by enhanced apoptotic resistance, cellular senescence, and increased invasiveness (Watson et al., 2009). These findings suggest that chronic hypoxia induces genome-wide adjustments in DNA methylation and histone modifications, potentially promoting and maintaining a hypoxic-adapted cellular phenotype that may contribute to tumor development (Watson et al., 2009). Meanwhile, Krieg et al. demonstrated that HIF-1 α regulates the histone demethylase JMJD1A, which enhances the expression of hypoxia-responsive genes and promotes tumor growth (Krieg et al., 2010). This study highlights the critical role of HIF-1 α in modifying histone enzymes under hypoxic conditions, thereby contributing to the dynamic regulation of gene expression in cancer cells (Krieg et al., 2010). These epigenetic modifications could further facilitate the emergence of paradoxical genes by modulating the expression diversity of tumor suppressor genes.

These molecular alterations under hypoxic stress lead to severe consequences, including enhanced angiogenesis, facilitating metastatic spread, altered cellular metabolism favoring cancer cell survival in low oxygen conditions, and increased resistance to conventional therapies (Semenza, 2003; Harris, 2002; Semenza, 2010). Targeting these adaptations has led to novel therapeutic strategies aimed at the hypoxic niche—such as the development of inhibitors that block HIFs, strategies to restore the functions of inactivated tumor suppressor pathways, and the use of hypoxia-activated prodrugs (Semenza, 2003; Wilson and Hay, 2011; Brown

TABLE 2 Summary of the tumor-promoting or tumor-inhibiting effects of the NOTCH signaling pathway in various cancers.

| Effect on tumors | Cancer type | Predominant NOTCH receptor | Role of notch signaling pathway | Signaling pathways involved | Mechanism of action | References |
|------------------|---------------------------|----------------------------|---|---|--|--|
| Promotion | Breast cancer | NOTCH1, NOTCH2, NOTCH3 | Promotes cell proliferation, survival, stemness, and contributes to treatment resistance and metastasis in breast cancer | JAG1-NOTCH1, NOTCH1-PTEN-ERK1/2, HER2, PKC α -JAG1-NOTCH, Notch3, FYN-STAT5-NOTCH2 | Enhances proliferation, stem cell survival, and metastasis; modulates resistance to treatments; promotes aggressive tumor traits | Reedijk et al. (2005), Baker et al. (2018), Pandya et al. (2016), Saran et al. (2023), Leontovich et al. (2018), Lee et al. (2018) |
| | Colon cancer | NOTCH1, NOTCH3 | Activates pathways that facilitate tumor cell survival and maintain stem cells | Wnt/ β -catenin, Hedgehog | Enhances tumor cell survival, supports stem cell maintenance, and represses secretory cell differentiation | Ishiguro et al. (2017), Bertrand et al. (2012), Brzozowa-Zasada (2022) |
| | T lymphoblastic neoplasms | NOTCH1 | Activates oncogenic pathways in T-cell neoplasms, contributes to chromosomal translocations, and influences transcriptional regulation in T-cell leukemia | Chromosomal translocations involving TAN-1 (NOTCH1), convergence with transcription factors like Ikaros | Induces oncogenic transformations in T-cell neoplasms, facilitates leukemia progression, and influences transcriptional regulation | Ellisen et al. (1991), Bellavia et al. (2007) |
| | T-ALL | NOTCH1, NOTCH3 | Serves as a biomarker and strengthens NOTCH signaling in T-ALL, indicating aggressive disease and supporting leukemia maintenance | Related to NOTCH signaling pathways, specifically NOTCH3/Jagged1 signaling | Serves as a biomarker indicating aggressive disease, reinforces Notch signaling, and promotes disease progression and maintenance | Bardelli et al. (2021), Pelullo et al. (2014) |
| | NSCLC | NOTCH1, NOTCH3 | Activates EGFR and other pathways, contributes to metastasis and stemness, and is associated with poor prognosis in NSCLC | EGFR, RFC4/Notch1 signaling, Notch3 overexpression | Enhances NSCLC metastasis, stemness, and tumor progression; correlates with poor prognosis | Pancewicz-Wojtkiewicz (2016), Liu et al. (2021b), Ye et al. (2013) |
| | CCRCC | NOTCH1, NOTCH3 | Influences renal cancer cell proliferation and increases metastasis risk and tumor growth | Cell cycle regulation and HIF-2 α , PI3K/Akt signaling | NOTCH3 regulates cell cycle progression and HIF-2 α ; NOTCH1 linked to increased metastasis risk and promotes tumor growth via PI3K/Akt | Han et al. (2020), Ai et al. (2012), Xu et al. (2012) |
| | Glioma | NOTCH1, NOTCH3 | Regulates several pathways to promote tumor growth, stemness, and invasion in gliomas | EGFR/c-myc, TGF β /Hippo/Notch | Enhances glioma cell invasion, self-renewal, and growth; interacts with multiple pathways to promote tumor development | Zhao et al. (2017), Xing et al. (2015), Pierfelice et al. (2011), Yi et al. (2019) |
| | Prostate cancer | NOTCH1, NOTCH3 | Influences tumor growth and progression in prostate cancer by altering cellular behaviors and gene expression | Changes in NOTCH expression during development and tumorigenesis | Supports tumor growth and development through modulation of signaling pathways | Carvalho et al. (2014), Shou et al. (2001) |
| Both | HNSCC | NOTCH1 | Influences tumor cell plasticity and contributes to HNSCC progression and suppression | EGFR, γ -secretase, Mutational landscape | Modulates tumor cell behavior and survival through interactions with EGFR and γ -secretase; linked to survival outcomes | Kalafut et al. (2021), Li et al. (2007), Stransky et al. (2011), Wirth et al. (2018) |

(Continued on following page)

TABLE 2 (Continued) Summary of the tumor-promoting or tumor-inhibiting effects of the NOTCH signaling pathway in various cancers.

| Effect on tumors | Cancer type | Predominant NOTCH receptor | Role of notch signaling pathway | Signaling pathways involved | Mechanism of action | References |
|------------------|-----------------|----------------------------|--|---|--|---|
| Suppression | HCC | NOTCH1, NOTCH3 | Inhibits HCC growth through cell cycle arrest, apoptosis induction, and modulation of key molecular pathways | Hippo, Wnt/ β -catenin, JNK, Cyclin G1, MDM2, miR-221 | Induces cell cycle arrest and apoptosis; suppresses tumor growth by modulating signaling pathways | Giovannini et al. (2016) , Giovannini et al. (2014) , Kim et al. (2017) , Qi et al. (2003) , Viator et al. (2011) , Sui et al. (2017) |
| | Cervical cancer | NOTCH1 | Downregulation of Notch1 signaling is required for sustained HPV-E6/E7 expression in cervical cancer | HPV-E6/E7 expression, Notch1 signaling | Sustains HPV-E6/E7 expression and facilitates malignant transformation by specifically modulating Notch1 signaling | Talora et al. (2002) |
| | Neuroblastoma | NOTCH1, NOTCH2 | Induces growth arrest in neuroblastoma cells by activating NOTCH signaling | Delta-Notch, Phox2B mutations | Induces cell cycle arrest and inhibits neuroblastoma cell growth through activation of Notch signaling | Zage et al. (2012) , van Limpt et al. (2005) |

T-ALL, T-cell acute lymphoblastic leukemia; NSCLC, non-small cell lung cancer; CCRCC, clear cell renal cell carcinoma; HCC, hepatocellular carcinoma; HNSCC, head and neck squamous cell carcinoma; TGF, transforming growth factor; HPV, human papillomavirus; EGFR, epidermal growth factor receptor; HIF, hypoxia-inducible factor; PKCa, protein kinase C α Phox2B, paired-like homeobox 2B; RFC4, replication factor C subunit 4; PI3K, phosphoinositide 3-kinase.

and Wilson, 2004; Wigerup et al., 2016). Additionally, addressing the downstream effects of tumor suppressor gene suppression, such as the use of PI3K inhibitors in cases of PTEN loss, offers a refined approach to disrupt the survival mechanisms employed by tumor cells in the hypoxic TME (Sansal and Sellers, 2004; Janku et al., 2018). These approaches leverage the modulation of tumor suppressor genes within the hypoxic tumor microenvironment to enhance the efficacy of cancer treatment.

8.2 Insights from acidosis

Acidosis within the TME significantly also impacts the cancer cell behavior (Webb et al., 2011; Damaghi et al., 2013). This condition stems from metabolic alterations in tumor cells, notably the high rates of glycolysis leading to excessive lactic acid production, even in the presence of oxygen (Webb et al., 2011; Gillies et al., 2008). This metabolic shift results in an accumulation of lactic acid, leading to a marked decrease in pH within the surrounding tissue (Gillies and Gatenby, 2007). Meanwhile, Riemann et al. demonstrated that an acidic tumor microenvironment induces reactive oxygen species (ROS) formation which then activate mitogen-activated protein kinases (MAPK) signaling in cancer cells (Riemann et al., 2011). This activation leads to phosphorylation of the transcription factor CREB via p38, altering transcriptional activity and potentially sustaining tumorigenic changes even after cells return to a normal environment (Riemann et al., 2011). Additionally, the acidic environment can further lead to epigenetic changes, affecting DNA methylation and histone modifications, potentially leading to the silencing of tumor suppressor genes or the activation of oncogenes (Thorne et al., 2009; Kulis and Esteller, 2010). These insights highlight the regulatory influence of pH factors on tumor prognosis and related signaling pathways, thereby creating conditions that are favorable for the emergence of paradoxical genes.

8.3 Insights from immune signaling pathways modulation

Immune modulation can have profound effects on the expression and function of tumor suppressor genes (Oeckinghaus et al., 2011). A key player in this regulatory network is the NF- κ B signaling pathway, which orchestrates responses that can either inhibit or promote tumor progression based on the surrounding cellular context (Karin et al., 2002; Baud and Karin, 2009). NF- κ B promotes oncogenesis by upregulating the expression of genes that promote cell proliferation and inhibit programmed cell death. Key targets include genes encoding anti-apoptotic proteins, such as Bcl-2, B-cell lymphoma-extra large (Bcl-xL), and inhibitor of apoptosis proteins, cell cycle regulators (such as cyclin D1 and c-Myc), and growth factors that together foster an environment conducive to the survival and proliferation of cancer cells (Annunziata et al., 2007; Karin et al., 2002; Baud and Karin, 2009). This role of NF- κ B has been extensively documented in cancers like multiple myeloma, where it contributes directly to the survival and proliferation of malignant cells under chemotherapeutic stress (Annunziata et al., 2007). Additionally, NF- κ B is a critical regulator of the TME by stimulating the production of pro-inflammatory cytokines and chemokines such as TNF- α , interleukin (IL)-6, and IL-8. These molecules aid in reshaping the surrounding stroma, promoting angiogenesis, and facilitating tumor cell invasion and metastasis (Karin, 2006). Furthermore, NF- κ B helps recruit and activate various immune cells within the TME that support tumor growth rather than combat it, thus contributing to tumor progression and the suppression of effective anti-tumor immune responses (Karin, 2006). NF- κ B also contributes to the ability of tumor cells to evade immune surveillance. It modulates the expression of molecules affecting the immune response, such as major histocompatibility complex molecules and PD-L1, a ligand for the PD-1 receptor on T cells, which inhibits T cell function. NF- κ B promotes an

immunosuppressive microenvironment by enhancing the expression of PD-L1 on tumor cells, allowing tumor cells to escape detection and destruction by the immune system (Greten and Karin, 2004).

The NF- κ B also plays key role as a suppressor of tumor development under certain contexts (Perkins, 2012). This suppression is primarily evident during the early stages of cancer and involves mechanisms that maintain cellular homeostasis and inhibit malignant transformations (Perkins, 2012). NF- κ B contributes to genomic stability maintenance by regulating the expression of genes involved in DNA repair and cell cycle checkpoints. This function prevents the accumulation of genetic mutations that could otherwise lead to oncogenesis (Volcic et al., 2012). NF- κ B can also induce cellular senescence, a permanent cell cycle arrest that functions as a barrier against the proliferation of potentially cancerous cells. This dual role of promoting DNA repair and senescence helps to suppress early tumor development and progression (Janssens and Tschopp, 2006). In specific cellular contexts, NF- κ B can activate the transcription of certain tumor suppressor genes. For example, NF- κ B induces the expression of GADD45 β , a stress-response gene that plays a crucial role in DNA repair and cell cycle regulation (Jarome et al., 2015; Al Tarrass et al., 2024; De Smaele et al., 2001). By activating such genes, NF- κ B contributes to the activation of mechanisms that can curb uncontrolled cell growth and promote apoptotic pathways in cells that have undergone malignant transformation (De Smaele et al., 2001) (Figure 4).

The ubiquitous role of the NF- κ B signaling pathway in cancer makes it a significant target for therapeutic intervention (Karin, 2006). Current therapeutic modalities focus on NF- κ B aim to mitigate its tumor-promoting actions while preserving or enhancing its tumor-suppressive capabilities (Karin and Greten, 2005). Therapeutic agents that inhibit NF- κ B can potentially reduce tumor-associated inflammation and diminish the supportive TME that fosters cancer cell survival and metastasis (Moreau et al., 2011). For instance, the use of proteasome inhibitors such as bortezomib, which prevents the degradation of I κ B (inhibitor of NF- κ B), thus inhibiting NF- κ B activation, has been found effective in treating multiple myeloma by reducing NF- κ B mediated survival signals (Moreau et al., 2011). Additionally, strategies to modulate the role of NF- κ B in immune suppression are being explored; modulation of NF- κ B is being studied in the context of enhancing the effectiveness of immunotherapies, such as checkpoint inhibitors (Lim et al., 2016). By suppressing NF- κ B-induced PD-L1 expression on tumor cells, these therapies can enhance T-cell activity against tumors, through a dual approach by directly inhibiting tumor cell survival mechanisms while boosting anti-tumor immunity (Lim et al., 2016). These examples illustrate the importance of the therapeutic targeting of NF- κ B is significant due to its ability to alter the TME, reduce tumor resistance to conventional therapies, and improve the outcomes of immunotherapeutic approaches (Karin, 2006; Greten and Karin, 2004; Gilmore and Herscovitch, 2006).

9 Conclusion

Traditionally, tumor suppressor genes such as TP53, retinoblastoma 1, and PTEN are well-known for their roles in regulating vital cellular processes, including DNA repair, cell cycle

progression, and apoptosis (Salmena et al., 2008; Vogelstein et al., 2000; Kaelin, 1997). The TP53 gene, often described as the “guardian of the genome,” is implicated in nearly half of the occurrence of all human cancers due to its critical functions in DNA repair and cell cycle regulation (Vogelstein et al., 2000). Similarly, RB1 controls the G1/S transition in the cell cycle, and PTEN counteracts the PI3K/AKT signaling pathway to influence cell survival (Salmena et al., 2008; Kaelin, 1997). Typically, the loss of function in these genes, whether through mutations, deletions, or epigenetic alterations, lead to the uncontrolled cell growth that characterizes cancer (Salmena et al., 2008; Vogelstein et al., 2000; Kaelin, 1997).

Contrary to long-standing scientific beliefs that these genes are rarely overexpressed in tumor tissues, recent advancements and the continuous enrichment of gene network databases, including TCGA have confirmed that the potential overexpression of tumor suppressor genes in such tissues. This phenomenon may be influenced by variations in tumor mutational burden and the interplay between compensatory regulatory mechanisms and the tumor microenvironment. Tumors with high tumor mutational burden often exhibit increased genomic instability, which can lead to the upregulation of tumor suppressor genes as part of a compensatory response. This has been consistently demonstrated in database studies as well as in basic experimental research. This phenomenon has been consistently demonstrated through both database analyses and fundamental experimental studies. While many studies focus primarily on gene expression at the transcriptional level, neglecting the corresponding expression at the protein level can result in unbalanced and potentially biased interpretations of biological effects. Nonetheless, given that non-coding genes vastly outnumber coding genes, this imbalance is likely only a secondary factor contributing to the existence of contradictory genes. Moreover, as previously discussed, substantial evidence supports the notion that variations in pathway expression across distinct tissue environments underscore the presence of contradictory genes. Of particular relevance is the stage-specific expression of these genes. Notably, the overexpression of tumor suppressor genes frequently occurs during the early stages of tumorigenesis, functioning as part of the cellular response to oncogenic stress (Sakaguchi et al., 2003; Ohuchida et al., 2006; Braig et al., 2005; Brambilla et al., 1999). For instance, S100A11 has been identified as a paradoxical gene (Sakaguchi et al., 2003). Sakaguchi et al. demonstrated that S100C/A11 is a critical mediator of calcium-induced growth inhibition in human epidermal keratinocytes by facilitating the phosphorylation-induced nuclear translocation of S100C/A11, thereby halting cell growth (Sakaguchi et al., 2003). However, Ohuchida et al. investigated the expression of the tumor suppressor gene S100A11 across various stages of pancreatic carcinogenesis (Ohuchida et al., 2006). Their research revealed overexpression of S100A11 in the early stages of pancreatic cancer development, such as in intraductal papillary mucinous neoplasms and pancreatic intraepithelial neoplasia (Ohuchida et al., 2006). However, its expression diminishes as the disease progresses advances (Ohuchida et al., 2006). Similarly, the overexpression of the tumor suppressor gene p16INK4a has been documented in early-stage tumors, where it has a crucial role in inducing cellular senescence and halting the proliferation of potentially cancerous cells (Braig et al., 2005; Brambilla et al., 1999). Similar studies not only confirm the existence of paradoxical genes but also emphasize their potential antagonistic

effects on tumor progression during tumorigenesis. Paradoxical genes strive to maintain cellular homeostasis. As discussed in our previous work, this antagonistic effect may be linked to alterations in tumor metabolism, TIME, and related signaling pathways, suggesting a potential self-protective mechanism of the body against tumors. We propose that this might represent a form of intrinsic tumor suppression. This overexpression may arise from mechanisms such as subclonal heterogeneity or attempts to balance rapid proliferation and genomic instability. However, once this antagonistic equilibrium is disrupted, paradoxical genes may no longer be able to counteract the progression, leading to their diminished expression and the subsequent unchecked advancement of the tumor towards increased malignancy. Therefore, enhancing the expression of these tumor suppressor genes at early stages could offer promising potential for effective tumor treatment.

In summary, this review introduces the innovative concept of paradoxical genes, highlights the crucial role of tumor suppressor genes in targeted cancer therapy, and provides a theoretical framework for treating cancer by exploiting the balanced interplay between oncogenes and tumor suppressor genes.

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DL: Writing—original draft, Writing—review and editing. LL: Visualization, Writing—original draft, Writing—review and editing. XC: Supervision, Writing—original draft, Writing—review and editing. GW: Conceptualization, Investigation, Writing—original draft.

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Glossary

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|---------------------------------|--|--------------------------------|--|
| ABCA1 | ATP-Binding Cassette Transporter A1 | KIRC | Kidney Renal Clear Cell Carcinoma |
| ALK | Activin Receptor-Like Kinase | LDLR | Low-Density Lipoprotein Receptor |
| ASB10 | Ankyrin Repeat and SOCS Box Containing 10 | LIHC | Liver Hepatocellular Carcinoma |
| Bcl-2 | B-Cell Lymphoma 2 | LXR | Liver X Receptor |
| Bcl-xL | B-Cell Lymphoma-extra Large | MDSCs | Myeloid-Derived Suppressor Cells |
| BLCA | Bladder Urothelial Carcinoma | MHC | Major Histocompatibility Complex |
| BRCA | Breast Cancer | NF-κB | Nuclear Factor Kappa Light Chain Enhancer of Activated B Cells |
| ccRCC | Clear Cell Renal Cell Carcinoma | NICD | Notch Intracellular Domain |
| CDK | Cyclin-Dependent Kinase | NK | Natural Killer |
| CELF2 | CUGBP Elav-Like Family Member 2 | NGS | Next-Generation Sequencing |
| CHOL | Cholangiocarcinoma | ORF | Open Reading Frame |
| COAD | Colon Adenocarcinoma | PD-1 | Programmed Cell Death Protein 1 |
| CTLs | Cytotoxic T Lymphocytes | PD-L1 | Programmed Death-Ligand 1 |
| CXCL | C-X-C Motif Chemokine Ligand | PDAC | Pancreatic Ductal Adenocarcinoma |
| CXCR | C-X-C Motif Chemokine Receptor | Phox2B | Paired-Like Homeobox 2B |
| DEA | Differential Expression Analysis | PI3K | Phosphoinositide 3-Kinase |
| DLL | Delta-Like | PKCα | Protein Kinase C α |
| DNMAML | Dominant Negative Mastermind-Like | PLK4 | Polo-Like Kinase 4 |
| EGFR | Epidermal Growth Factor Receptor | PRAD | Prostate Adenocarcinoma |
| EMT | Epithelial-Mesenchymal Transition | PTMs | Post-Translational Modifications |
| ERK | Extracellular Signal-Regulated Kinase | QC | Quality Control |
| ESCA | Esophageal Carcinoma | RB1 | Retinoblastoma 1 |
| FA | Fatty Acid | RFC4 | Replication Factor C Subunit 4 |
| FDA | Food and Drug Administration | RIDD | Regulated IRE1-Dependent Decay of RNA |
| GO | Gene Ontology | SCC | Squamous Cell Carcinoma |
| GTEx | Genotype-Tissue Expression | SCD1 | Stearoyl-CoA Desaturase 1 |
| GADD45β | Growth Arrest and DNA Damage-Inducible β | SNPs | Single Nucleotide Polymorphisms |
| HDAC | Histone Deacetylase | SREBP-1c | Sterol Regulatory Element-Binding Protein 1c |
| HHLA2 | HERV-H LTR-Associating 2 | SUMO | Small Ubiquitin-like Modifier |
| HGP | Human Genome Project | T-ALL | T-cell Acute Lymphoblastic Leukemia |
| HIF | Hypoxia-Inducible Factor | TCGA | The Cancer Genome Atlas |
| HNSCC | Head and Neck Squamous Cell Carcinoma | TGFβ | Transforming Growth Factor β |
| HPV | Human Papillomavirus | TEM8 | Tumor Endothelial Marker 8 |
| IAPs | Inhibitor of Apoptosis Proteins | Thr | Threonine |
| ICGC | International Cancer Genome Consortium | TIME | Tumor Immune Microenvironment |
| IRES | Internal Ribosome Entry Sites | TNBC | Triple-Negative Breast Cancer |
| IκB | Inhibitor of NF- κ B | Tyr | Tyrosine |
| IPMN | Intraductal Papillary Mucinous Neoplasms | uORFs | Upstream Open Reading Frames |
| IRE | Inositol-Requiring Enzyme | | |
| JNK | c-Jun N-terminal Kinase | | |
| KEGG | Kyoto Encyclopedia of Genes and Genomes | | |
| KICH | Kidney Chromophobe | | |