



OPEN ACCESS

EDITED BY

Senka Blažetić,
Josip Juraj Strossmayer University of Osijek,
Croatia

REVIEWED BY

Diana Escalante-Alcalde,
National Autonomous University of Mexico,
Mexico

Barbara Viljetić,
Josip Juraj Strossmayer University of Osijek,
Croatia

*CORRESPONDENCE

Dragana Fabris,
✉ dragana.fabris@mef.hr
Ivana Karmelić,
✉ ivana.karmelic@mef.hr

[†]These authors have contributed equally to this work and share first authorship

RECEIVED 19 July 2024

ACCEPTED 11 November 2024

PUBLISHED 11 December 2024

CITATION

Karmelić I, Jurilj Sajko M, Sajko T, Rotim K and Fabris D (2024) The role of sphingolipid rheostat in the adult-type diffuse glioma pathogenesis. *Front. Cell Dev. Biol.* 12:1466141. doi: 10.3389/fcell.2024.1466141

COPYRIGHT

© 2024 Karmelić, Jurilj Sajko, Sajko, Rotim and Fabris. This is an open-access article distributed under the terms of the [Creative Commons Attribution License \(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

The role of sphingolipid rheostat in the adult-type diffuse glioma pathogenesis

Ivana Karmelić^{1*†}, Mia Jurilj Sajko^{2†}, Tomislav Sajko², Krešimir Rotim² and Dragana Fabris^{1*}

¹Department of Medical Chemistry, Biochemistry and Clinical Chemistry, School of Medicine, University of Zagreb, Zagreb, Croatia, ²Department of Neurosurgery, University Hospital Center "Sestre milosrdnice", Zagreb, Croatia

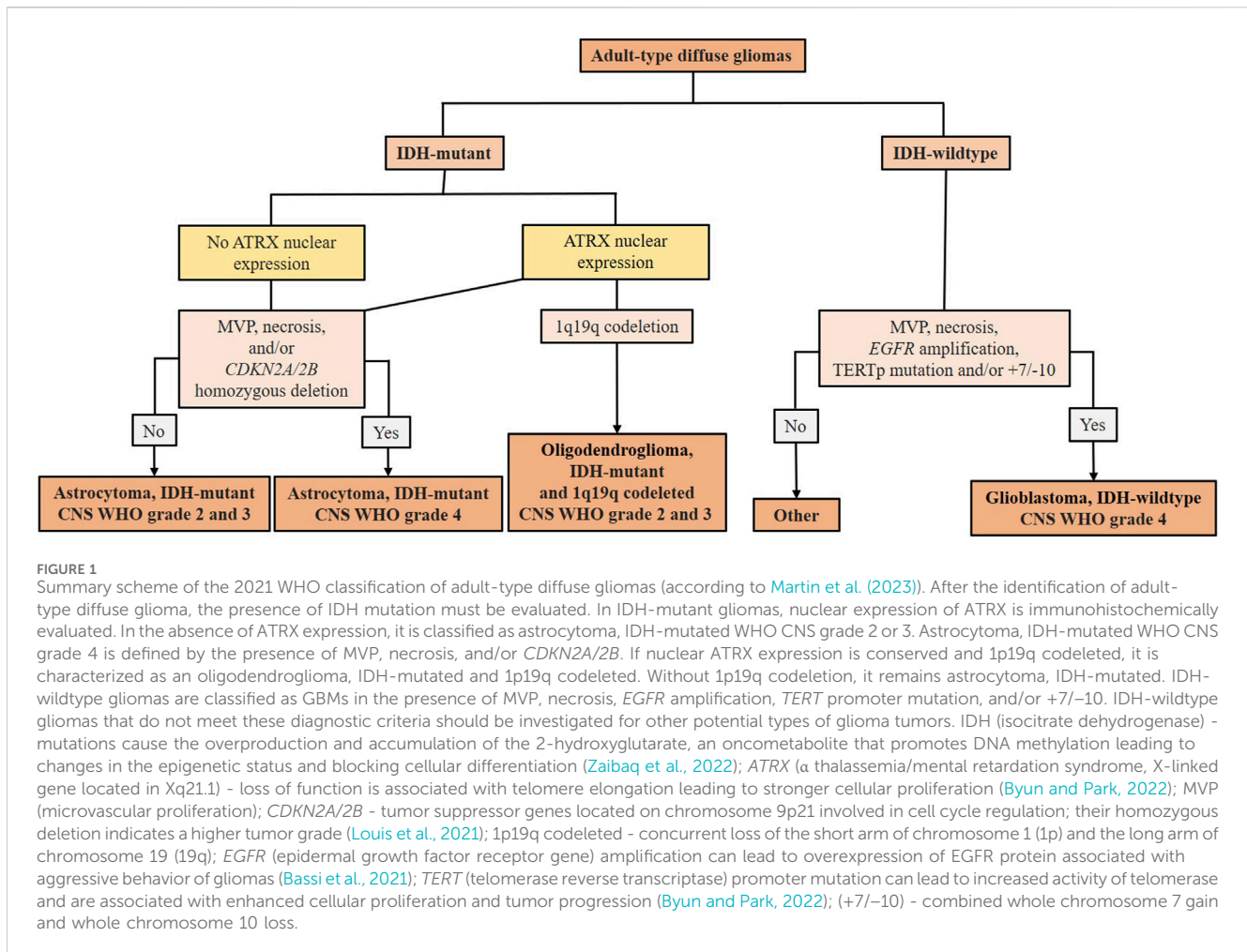
Gliomas are highly aggressive primary brain tumors, with glioblastoma multiforme being the most severe and the most common one. Aberrations in sphingolipid metabolism are a hallmark of glioma cells. The sphingolipid rheostat represents the balance between the pro-apoptotic ceramide and pro-survival sphingosine-1-phosphate (S1P), and in gliomas it is shifted toward cell survival and proliferation, promoting gliomas' aggressiveness, cellular migration, metastasis, and invasiveness. The sphingolipid rheostat can be altered by targeting enzymes that directly or indirectly affect the ratio of ceramide to S1P, leading to increased ceramide or decreased S1P levels. Targeting the sphingolipid rheostat offers a potential therapeutic pathway for glioma treatment which can be considered through reducing S1P levels or modulating S1P receptors to reduce cell proliferation, as well as through increasing ceramide levels to induce apoptosis in glioma cells. Although the practical translation into clinical therapy is still missing, sphingolipid rheostat targeting in gliomas has been of great research interest in recent years with several interesting achievements in the glioma therapy approach, offering hope for patients suffering from these vicious malignancies.

KEYWORDS

diffuse glioma, glioblastoma, sphingosine-1-phosphate, ceramides, sphingolipid rheostat

1 Introduction

Gliomas, the most prevalent and aggressive form of intracranial cancer, account for over 80% of malignant brain tumors, with glioblastoma multiforme (GBM) as the most common and severe subtype (Ostrom et al., 2014). Despite the significant research interest and certain progress in glioma treatment in recent years, the prognosis for patients with gliomas, especially GBM, remains very poor (Ma et al., 2024). The latest fifth edition of the World Health Organization (WHO) tumor classification of the central nervous system (CNS) introduced significant updates, incorporating molecular and genetic parameters for more precise clinical categorization (Louis et al., 2021). Sphingolipids, especially glycosphingolipids, are abundantly in the brain and integral to cell membrane structure and cellular signaling (Hannun and Obeid, 2018). Abnormalities in sphingolipid metabolism have been implicated in promoting gliomas' aggressiveness, strongly affecting numerous characteristics of tumor phenotype, such as cellular migration, metastasis, and invasiveness (Zaibaq et al., 2022). Their main metabolic enzymes have been actively investigated as new potential targets in glioma therapy development (Wang



et al., 2022). The “sphingolipid rheostat” concept highlights the opposing roles of ceramide, which induces apoptosis, and sphingosine-1-phosphate (S1P), which promotes proliferation (Cuveillier et al., 1996). This balance between the pro-apoptotic ceramide and pro-survival S1P is crucial in determining the cell’s fate, and in glioma it is altered towards the pro-survival S1P signaling, promoting uncontrolled cell proliferation and invasiveness (Zaibaq et al., 2022). This mini-review explores the complex and interconnected metabolic effects of sphingolipids in glioma progression, focusing on new achievements in sphingolipid rheostat targeting as an innovative glioma therapy approach, alone or combined with the established current glioma therapies to improve glioma, particularly GBM patient outcomes.

2 Adult-type diffuse gliomas classification

Diffuse gliomas are the most common type of primary CNS tumors in adults (Lucke-Wold et al., 2024; Martin et al., 2023). The fifth edition of the WHO classification of CNS tumors (CNS5) introduced significant changes by adding molecular and genetic markers to the previously histology based classification, making it more precise and clinically relevant (Louis et al., 2021). According to

WHO CNS5, there are three main types of adult-type diffuse gliomas: (I) astrocytoma, isocitrate dehydrogenase (IDH)-mutant; (II) oligodendroglioma, IDH-mutant and 1p/19q codeleted; and (III) glioblastoma, IDH-wildtype (Louis et al., 2021; Gisina et al., 2022) (Figure 1). IDH-mutant diffuse astrocytic tumors are considered as astrocytoma, IDH-mutant CNS WHO grades 2, 3, or 4 (Louis et al., 2021). Oligodendrogliomas, IDH-mutant and 1p/19q codeleted are classified as CNS WHO grade 2 or 3 (Louis et al., 2021). IDH-wildtype diffuse astrocytic tumors are now classified as GBM CNS WHO grade 4 if there is microvascular proliferation (MVP) or necrosis and if they meet 1 or more of 3 specific key molecular criteria (Figure 1), even if they show low-grade histological features (Louis et al., 2021). Further evaluation is necessary to classify other IDH-wildtype gliomas that do not meet these molecular criteria (Louis et al., 2021; Meyer et al., 2021). Astrocytoma, IDH-mutant CNS WHO grade 4 are no longer classified as GBMs due to distinct molecular and epigenetic profiles (Figure 1) and different clinical behaviors (Zaibaq et al., 2022; Rajaratnam et al., 2020). The most malignant and aggressive of the adult-type diffuse gliomas are the astrocytoma, IDH-mutant CNS WHO grade 3 and 4, and GBM, IDH-wildtype CNS WHO grade 4, considered higher-grade gliomas (HGG) (Martin et al., 2023). Glioblastoma accounts for nearly 15% of all brain tumors, with approximately

80% being primary tumors, mainly occurring in older patients, while secondary GBMs develop from lower-grade gliomas primarily in younger patients (Louis et al., 2021). Despite aggressive standard therapy, including radical surgery, radiotherapy and temozolomide (TMZ) chemotherapy, and even with adjuvant chemotherapy, the median overall survival of remains 14–20 months (Ma et al., 2024; Stupp et al., 2005). Other therapies, such as carmustine wafers placed in surgical cavity, anti-VEGF antibodies, PDGF and EGFR inhibitors, and immunotherapy, extend survival for only a few months (Norden and Wen, 2006). Recurrence remains nearly inevitable due to GBM's aggressiveness and therapy resistance, driven by brain tumor-initiating cells (BTICs) (Landis et al., 2018). Lower-grade gliomas, such as astrocytoma, IDH-mutant CNS WHO grade 2 and oligodendroglioma, IDH-mutant and 1q19q codeleted CNS WHO grade 2, have a better prognosis, but their treatment is also challenging as most of them tend to progress to HGG over time (Yu et al., 2020).

3 Sphingolipid metabolism

Sphingolipids are ubiquitous components of eukaryotic cell membranes, especially abundant in human neural tissue, where they contribute (Gault et al., 2010) to membrane structure and serve as signaling molecules in processes like apoptosis, proliferation, angiogenesis, and vesicular trafficking (Ogretmen, 2017). They are commonly divided into simple, sphingoid bases and ceramides, and complex sphingolipids, such as glycosphingolipids, sphingomyelins and sulphatides (Hannun and Obeid, 2018).

Sphingoid bases, long-chain aliphatic amino alcohols, form the structural backbone of sphingolipids and serve as signaling molecules. Their chain length can vary from 12 to 26 carbons, with sphingosine (d18:1) and sphinganine (d18:0) predominating in mammalian tissue; sphingosine arise from sphingolipid degradation, while sphinganine is a precursor in *de novo* sphingolipid biosynthesis (Pruett et al., 2008).

Ceramides, bioactive sphingolipids are synthesized by six ceramide synthase isoforms (CERS1-6) which bind fatty acids of varying chain lengths to a sphingoid base, resulting in ceramides with diverse functions and tissue distributions (Riebeling et al., 2003; Raichur, 2020). In human brain tissue, 18-carbon fatty acid ceramides are the most abundant (Pruett et al., 2008). Ceramides are synthesized either *de novo* or via hydrolysis from complex sphingolipids and are further catabolized by ceramidases to form sphingoid bases, serving as a key intermediates and precursors for all complex sphingolipids (Zheng et al., 2006; Mullen et al., 2012). Bioactive sphingoid bases and ceramides disrupt pro-survival cellular signaling pathways inducing apoptosis by various activating mechanisms (Nganga et al., 2018). Extracellularly oriented ceramides produced by acid sphingomyelinase (sphingomyelin phosphodiesterase 1, SMPD1) in lipid rafts activate membrane receptor clustering with different tumor necrosis factors triggering strong apoptotic signal responses (Dumitru et al., 2009). Intracellularly, ceramides trigger apoptosis by affecting outer mitochondrial membrane permeability and regulating apoptosis-related molecules like phosphatases, kinases, and phospholipase (Gault et al., 2010; Dadsena et al., 2019) During

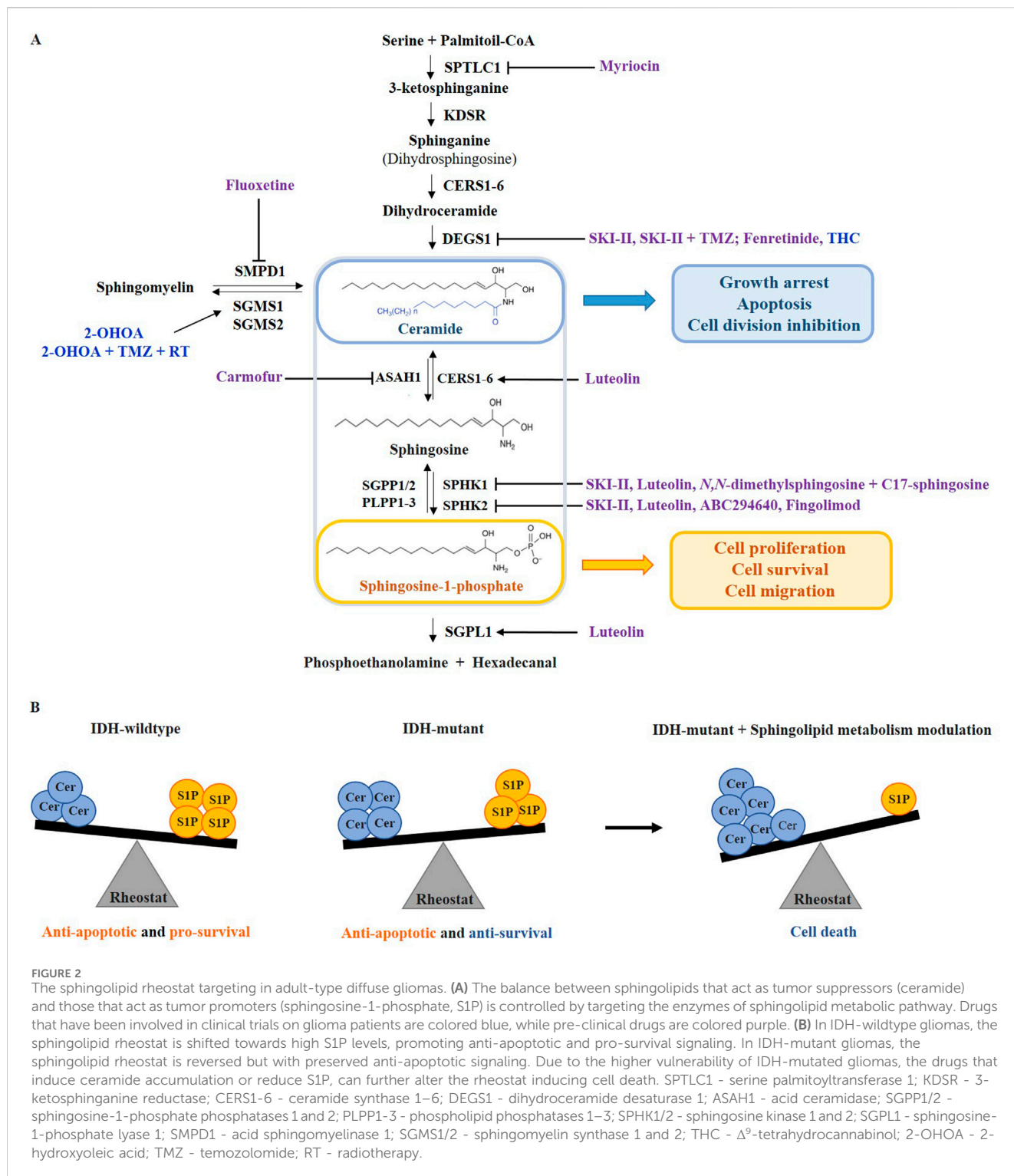
oncogenic transformation, cells often evade apoptosis as ceramidases become more active, hydrolyzing ceramides into sphingosine and fatty acid (Leverly, 2005).

Sphingosine kinases 1 and 2 (SPHK1/2) phosphorylate sphingosine to S1P, with SPHK1 being localized in the cytoplasm and SPHK2 in the nucleus, endoplasmic reticulum (ER), mitochondria, and, in cancer cells, in the plasma membrane (Diaz Escarcega et al., 2021). While SPHK1 has mainly pro-survival roles, SPHK2 can promote both pro-apoptotic and pro-survival signals depending on its localization in the cell. SPHK2 can be translocated to the nucleus where the formation of S1P inhibits histone deacetylase 1 and 2 (HDAC1/2) activity, leading to epigenetic changes in gene expression (Diaz Escarcega et al., 2021). Sphingosine-1-phosphate act as a second messenger regulating cell growth, differentiation, migration, and apoptosis by binding to G protein-coupled receptors, S1PR1-5, with variable tissue expression and important roles in development, aging, and pathologies (Merrill, 2011; Sousa et al., 2023; Blaho and Hla, 2014). S1P controls various signaling pathways, including angiogenesis and proliferation, by activating cytosolic effectors, phospholipases, and kinases (Pyne and Pyne, 2000). It is inactivated by ER-enzymes S1P-phosphatases 1 and 2 (SGPP1/2) and phospholipid phosphatases 1–3 (PLPP1-3), or by S1P lyase 1 (SGPL1) localized in the ER and Golgi, but also in plasma membrane, where it directly influences extracellular S1P concentrations (Sousa et al., 2023; Tang and Brindley, 2020) (Figure 2A).

4 Sphingolipid rheostat in gliomas

The sphingolipid rheostat concept refers to the balance between pro-apoptotic ceramides and sphingosine, and pro-survival S1P, where ceramide and sphingosine are directly correlated to the downregulation of S1P and vice versa (Cuvillier et al., 1996) (Figure 2A). Bioactive sphingolipid metabolites may serve as biomarkers for cancer malignancy, progression, and metastasis (Newton et al., 2015) (Figure 2B).

Overexpression of SPHK1 and the resulting production of S1P correlated with malignancy, poor prognosis, and shorter survival time in different types of gliomas, especially GBMs (Young and Van Brocklyn, 2007; Li et al., 2022). Increased SPHK1 and decreased SGPP1/2 expression were a hallmark of cultured GBM cells with upregulation of SPHK1 promoting growth and metastasis of GBM cells (Zaibaq et al., 2022; Abuhusain et al., 2013). Upregulated acid ceramidase (*N*-acyl sphingosine amidohydrolase 1, ASAH1) and SPHK1 were detected in GBM tissues compared to normal brain as well as higher ASAH1 activity in BTICs (Abuhusain et al., 2013). S1P was reported to be, on average, 9-fold higher in GBM tissues compared to normal human brain tissue and correlated with glioma grade, while C18 ceramide was 5-fold lower in GBM tissues (Abuhusain et al., 2013). Fast-proliferating glioblastoma stem cells (GSCs) degrade ceramide and convert sphingosine to S1P more rapidly than slow-proliferating GSCs, and consequently release a significantly higher amount of S1P into the extracellular environment (Marfia et al., 2014). GBM cells and BTICs also release S1P which interacts with S1PRs influencing diverse cellular processes including cell migration,



differentiation, proliferation, survival against apoptosis and angiogenesis (Zaibaq et al., 2022; Hawkins et al., 2020; Bassi et al., 2021). Moreover, overexpression of S1PR1-3 is observed in glioma cells (Hawkins et al., 2020). S1P also modulates EGFR expression in both cancer and non-cancer cells (Bassi et al., 2021).

Ceramide is considered a tumor suppressor lipid that can induce anti-proliferative and apoptotic responses in various tumor types,

including gliomas (Zaibaq et al., 2022; Bassi et al., 2023). Ceramide levels are often reduced in gliomas due to increased degradation or altered synthesis and are significantly lower in HGG, correlating with malignant progression and poor patient survival (Riboni et al., 2002). The expression of various ceramide types and the balance among them are considered critical for different cancer cell progressions, including glioma. Generally, long-chained ceramides (C16, C18, C20) are considered as pro-apoptotic, while

very-long chained ceramides (particularly C24 and C24:1) are associated with proliferative effects (Melero-Fernandez de Mera et al., 2022). In GBM tissue, C18-ceramide levels have been reported to decrease by up to 70%, while reconstitution of C18-ceramide induced cell death in human GBM cells resistant to TMZ (Das et al., 2018).

Recent findings indicate that IDH-wildtype gliomas exhibit a stronger imbalance in sphingolipid rheostat than IDH-mutant gliomas, resulting in elevated ceramide and sphingosine production, which may be the reason why IDH-mutant gliomas are less aggressive than IDH-wildtype (Dowdy et al., 2020). Moreover, several researches have shown that in IDH-mutant gliomas, the sphingolipid balance can be additionally shifted towards increasing pro-apoptotic ceramide production or towards decreasing pro-survival S1P production by targeting enzymes involved in the sphingolipid metabolism like SPHK1/2 and ceramidase, which reduces cancer hallmarks in IDH-mutant gliomas (Zaibaq et al., 2022; Dowdy et al., 2020; Tea et al., 2020) (Figure 2B).

4.1 Current improvements in sphingolipid rheostat targeting in glioma therapy

The sphingolipid rheostat can be altered by targeting enzymes that directly affect the ratio of ceramide to S1P, or by targeting other pathway enzymes that indirectly lead to increased ceramide or decreased S1P levels (Zaibaq et al., 2022; Tea et al., 2020). In gliomas, sphingolipid rheostat is out of balance promoting cell survival, thus providing a rich source of potential therapy targets. Ionizing radiation and TMZ, as current standard therapy for glioma patients, affect the sphingolipid pathway by activating SMPD1 which hydrolyzes sphingomyelin to pro-apoptotic ceramide (Hawkins et al., 2020; Tea et al., 2020).

Preclinical studies have shown that SPHK1 inhibitors can suppress glioma growth by reducing S1P levels. Still, their use in clinical trials is limited due to the conflicting data on the cytotoxic effects on different tumor cell models (Zaibaq et al., 2022; Sousa et al., 2023). Inhibition of SPHK1 selectively blocked angiogenesis, but it did not induce cell death while targeting SPHK1/2 both with dual SPHK inhibitor SKI-II proved to be more efficient in decreasing S1P production and promoting anticancer activity (Sousa et al., 2023; French et al., 2006). Additionally, SKI-II acts as a noncompetitive inhibitor of dihydroceramide desaturase 1 (DEGS1), an enzyme that catalyzes the introduction of a double bond into dihydroceramide, converting it into ceramide. The treatment of GBM cells with SKI-II resulted in the accumulation of dihydroceramide and depletion of S1P, which reduced cell proliferation and induced autophagy (Cingolani et al., 2014). SKI-II was found to have a synergistic effect with TMZ and induced cell death in the TMZ-resistant GBM cell lines (Sousa et al., 2023; Cingolani et al., 2014). Both SKI-II alone and combined with TMZ therapy are still in the preclinical testing phase (Sousa et al., 2023).

ABC294640 (Opaganib) as the first-in-class selective competitive inhibitor of SPHK2 has no off-targets on protein kinases and decreases levels of S1P in tissue cultures (Lewis et al., 2018). Dowdy et al. (Dowdy et al., 2020) demonstrated that in IDH-

mutant glioma subtypes, the combination of SPHK1 inhibitor *N,N*-dimethylsphingosine with C17 sphingosine reduced S1P production and enhanced ceramide and sphingosine accumulation. This induced growth arrest and apoptosis, specifically in IDH-mutant gliomas highlighting a metabolic vulnerability characterized by elevated ceramides and decreased SPHK2 expression in IDH-mutant compared to IDH-wildtype gliomas.

Fenretinide, a synthetic retinoid derivative, induces apoptosis in HGG cell lines by indirectly inhibiting DEGS1 (Puduvalli et al., 1999). Although additional testing led to a phase II clinical trial in patients with recurrent glioma and GBM, fenretinide was ineffective at the administered concentrations (Puduvalli et al., 1999; Puduvalli et al., 2004).

Preclinical studies and one pilot study have shown that THC (Δ^9 -tetrahydrocannabinol) has biostatic effects on recurrent GBM by inhibiting DEGS1 while sparing non-transformed astroglial cells (Velasco et al., 2007; Del Pulgar et al., 2002). The combination of THC and CBD (cannabidiol) in 1:1 ratio (Sativex oral spray) with TMZ is currently in a phase II clinical trial (ARISTOCRAT trial) for GBM therapy (Bhaskaran et al., 2024).

Acid ceramidase (ASAH1) is the key enzyme that reduces ceramide levels by catalyzing its hydrolysis to sphingosine, thus also controlling the pool of sphingosine that can be converted to S1P (Doan et al., 2017a). It has been reported that ASAH1 is upregulated in GBM tissue and correlates with worse patient outcomes and radioresistance (Doan et al., 2017b). *In vitro* inhibition of ASAH1 enzyme with **carbamofur**, a fluorouracil derivative, decreased the growth of TMZ-resistant GBM cells and effectively killed GSCs increasing ceramide levels and inducing apoptosis (Hawkins et al., 2023; Dementiev et al., 2019).

The most recent research by Navone et al (Navone et al., 2023) showed that **luteolin**, a natural flavonoid, inhibited the expression of SPHK1/2 while increasing the expression of both SGPL1 and CERS6, which decreased cell viability and survival of TMZ-resistant GSCs.

Myriocin, a competitive inhibitor of serine palmitoyltransferase 1 (SPTLC1), the first enzyme in *de novo* sphingolipid biosynthesis, can inhibit the proliferation of different cancer types by altering the sphingolipid rheostat (Lee et al., 2012; Yaguchi et al., 2017). Treating GBM cells with myriocin led to a 35% decrease in S1P levels (Bernhart et al., 2015). Synthetic sphingolipid analog, **ingolimid** (FTY720) was designed to improve the biostatic properties of myriocin, while also acting as a competitive inhibitor of SPHK2 (Zaibaq et al., 2022; Paugh et al., 2003). Fingolimod-1-phosphate inhibits HDAC1/2 and enhances histone acetylation leading to epigenetic regulation of specific genes that increase sensitivity to chemotherapeutics while also acting as an S1PR1 antagonist in the CNS lymph nodes (Hawkins et al., 2020; Pyne and Pyne, 2020). Carbamofur, luteolin, myriocin, and fingolimod are still in the preclinical phase of research (Zaibaq et al., 2022).

Sphingomyelin is a crucial component of membrane lipid rafts essential for signaling. It is synthesized by sphingomyelin synthase 1 and 2 (SGMS1/2) isoenzymes, with SGMS1 being the primary isoform present at significantly lower levels in cancer cell membranes, including GBM, and reduced expression of SGMS1 in gliomas is associated with poorer prognostic outcomes (Fernández-García et al., 2019; Lladó et al., 2014).

Major progress in the therapy of glioma was achieved with synthetic **2-hydroxyoleic acid** (2-OHOA), which proved to be a highly specific activator of the SGMS1 isoform with a significantly higher bioavailability and lower toxicity compared to TMZ when administered in therapeutic doses (Lopez et al., 2023). It has been tested on glioma cell lines, preclinical animal studies, and clinical trials on patients are in phase I-II (Zaibaq et al., 2022; Fernández-García et al., 2019; Lopez et al., 2023). Currently, phase IIB-III trials are testing the combination of 2-OHOA, TMZ, and radiotherapy for GBM (NIHR, 2021), representing the closest a novel sphingolipid modulator has come to potential approval as a glioma therapy.

Recent preclinical research indicates that antidepressant **fluoxetine** (Prozac), a selective serotonin reuptake inhibitor (SSRI), can inhibit SMPD1 and induce cell death in glioma by accumulating sphingomyelin, which results in inhibition of oncogenic EGFR signaling and activating lysosomal stress (Bi et al., 2021). Combining fluoxetine, but not other SSRI inhibitors, with TMZ caused massive GBM cell death and complete tumor regression in mice, and it is now being tested in early phase I clinical trial (UCSD, 2024).

4.2 Future perspectives

Even though many exciting findings have been revealed in recent years regarding sphingolipid rheostat targeting in gliomas, they have still not been translated into viable therapy (Zaibaq et al., 2022). Due to the very complex and interconnected sphingolipid metabolism, it is very challenging to influence the activity of one enzyme or metabolic product and not disrupt the expression or metabolic fate of another. Further obstacles to the clinical application of sphingolipid-targeted therapies include challenges with drug delivery across the blood-brain barrier, lack of target specificity that leads to systemic toxicity, tumor heterogeneity (especially in GBMs), and the absence of reliable biomarkers to identify patients who would benefit most from sphingolipid-based treatments (Rahman and Ali, 2024). To overcome these challenges, research is focused on developing targeted delivery systems, combination therapies, and reliable biomarkers for patient selection (Janneh, 2024). While sphingolipid-targeted therapies represent a promising direction for glioma treatment, their potential can be further enhanced by integrating with existing therapeutic strategies. Combining SPHK inhibitors or ceramide analogs with standard treatments such as TMZ or immunotherapy may produce synergistic effects. Furthermore, the integration of genetic profiling in treatment by identifying specific sphingolipid enzyme profiles in individual tumors could enable a more personalized therapeutic approach (Scorsetti et al., 2022; Zhou et al., 2024). This personalized approach could be particularly valuable in IDH-mutant gliomas, where genetic variations in sphingolipid metabolism may influence treatment outcomes (Kayabolen et al., 2021). A still insufficiently studied field in sphingolipid-based glioma therapy is the role of ceramide variability, as different ceramide chain lengths may have opposing effects, despite the general association of ceramides with pro-apoptotic functions (Melero-Fernandez de Mera et al., 2022; Hartmann et al., 2012). Another interesting potential lies in

the less abundant sphingoid base sphinganine (d18:0), although structurally similar to sphingosine, has distinct metabolic origins and lacks sphingosine's apoptotic signaling abilities (Farley et al., 2024). On the other hand, sphinganine-1-phosphate exhibits strong signaling properties and binds to the S1PR1 even stronger than S1P, yet lacks S1P's cytoprotective and proliferative effects (Van Brocklyn et al., 1998). The signaling roles of sphinganine and sphinganine-1-phosphate are yet to be clarified offering an interesting and unexplored field.

5 Conclusion

The sphingolipid rheostat is a critical regulator of cell fate in health and disease. In glioma cells, aberrant expression of enzymes of the sphingolipid metabolism leads the cells to escape apoptosis and promote survival and invasiveness. In recent years, several targeted therapies have been proposed that shift the rheostat towards apoptosis and tumor suppression. Targeting the sphingolipid rheostat for glioma treatment provides not only the first-line therapy options for gliomas but is also responsible for sensitizing particular types of gliomas, such as IDH-mutant glioma, for stronger chemotherapeutic response (Dowdy et al., 2020). Given the high complexity of gliomas, particularly GBM, and the limitations of treatment options available so far, a combination of targeting sphingolipid rheostat with some of the current therapies and with genetic profiling for personalized patient selection offers a promising approach to improve outcomes of patients suffering from these vicious tumors.

Author contributions

IK: Conceptualization, Investigation, Supervision, Visualization, Writing—original draft, Writing—review and editing. MJ: Investigation, Writing—original draft, Writing—review and editing. TS: Writing—review and editing. KR: Writing—review and editing. DF: Conceptualization, Investigation, Supervision, Visualization, Writing—original draft, Writing—review and editing.

Funding

The author(s) declare that financial support was received for the research, authorship, and/or publication of this article. This research was funded by Adris Foundation grant “SphingoMark and Thrombectomy” to DF, grant no. 13127 and by the University of Zagreb Support grant “Sphingolipidomics of stroke thrombi and serum in ischemic stroke” to IK, grant no. 10106-23-2513.

Conflict of interest

The authors declare that the research was conducted without any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated

organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

- Abuhusain, H. J., Matin, A., Qiao, Q., Shen, H., Kain, N., Day, B. W., et al. (2013). A metabolic shift favoring sphingosine 1-phosphate at the expense of ceramide controls glioblastoma angiogenesis. *J. Biol. Chem.* 288, 37355–37364. doi:10.1074/jbc.M113.494740
- Bassi, R., Brambilla, S., Tringali, C., and Giussani, P. (2021). Extracellular sphingosine-1-phosphate downstream of EGFR increases human glioblastoma cell survival. *Int. J. Mol. Sci.* 22 (13), 6824. doi:10.3390/ijms22136824
- Bassi, R., Dei Cas, M., Tringali, C., Compostella, F., Paroni, R., and Giussani, P. (2023). Ceramide is involved in temozolomide resistance in human glioblastoma U87MG overexpressing EGFR. *Int. J. Mol. Sci.* 24 (20), 15394. doi:10.3390/ijms242015394
- Bernhart, E., Damm, S., Wintersperger, A., Nuschold, C., Brunner, A. M., Plastira, I., et al. (2015). Interference with distinct steps of sphingolipid synthesis and signaling attenuates proliferation of U87MG glioma cells. *Biochem. Pharmacol.* 96 (2), 119–130. doi:10.1016/j.bcp.2015.05.007
- Bhaskaran, D., Savage, J., Patel, A., Collinson, F., Mant, R., Boele, F., et al. (2024). A randomised phase II trial of temozolomide with or without cannabinoids in patients with recurrent glioblastoma (ARISTOCRAT): protocol for a multi-centre, double-blind, placebo-controlled trial. *BMC Cancer* 24 (1), 83. doi:10.1186/s12885-023-11792-4
- Bi, J., Khan, A., Tang, J., Armando, A. M., Wu, S., Zhang, W., et al. (2021). Targeting glioblastoma signaling and metabolism with a re-purposed brain-penetrant drug. *Cell Rep.* 37 (5), 109957. doi:10.1016/j.celrep.2021.109957
- Blaho, V. A., and Hla, T. (2014). Thematic Review Series: lysophospholipids and their Receptors: an update on the biology of sphingosine 1-phosphate receptors. *J. Lipid Res.* 55 (8), 1596–1608. doi:10.1194/jlr.R046300
- Byun, Y. H., and Park, C.-K. (2022). Classification and diagnosis of adult glioma: a scoping review. *Brain and NeuroRehabilitation* 15 (3), e23. doi:10.12786/bn.2022.15.e23
- Cingolani, F., Casasampere, M., Sanlehi, P., Casas, J., Bujons, J., and Fabrias, G. (2014). Inhibition of dihydroceramide desaturase activity by the sphingosine kinase inhibitor SKI II. *J. Lipid Res.* 55 (8), 1711–1720. doi:10.1194/jlr.M049759
- Cuvillier, O., Pirianov, G., Kleuser, B., Vanek, P. G., Cosot, O. A., Gutkind, J. S., et al. (1996). Suppression of ceramide-mediated programmed cell death by sphingosine-1-phosphate. *Nat* 381 (6585), 800–803. doi:10.1038/381800a0
- Dadsena, S., Bockelmann, S., Mina, J. G. M., Hassan, D. G., Korneev, S., Razzera, G., et al. (2019). Ceramides bind VDAC2 to trigger mitochondrial apoptosis. *Nat. Commun.* 10 (1), 1832–1912. doi:10.1038/s41467-019-09654-4
- Das, A., Zdzislaw, S., Cachia, D., Patel, S. J., and Ogretmen, B. (2018). Exth-51. C18-Ceramide analogue drug overcomes resistance to temozolomide in glioblastoma. *Neuro Oncol.* 20 (Suppl. 1_6), vi96. doi:10.1093/neuonc/ny148.399
- Del Pulgar, T. G., Velasco, G., Sánchez, C., Haro, A., and Guzmán, M. (2002). *De novo*-synthesized ceramide is involved in cannabinoid-induced apoptosis. *Biochem. J.* 363 (Pt 1), 183–188. doi:10.1042/0264-6021:3630183
- Dementiev, A., Joachimiak, A., Nguyen, H., Gorelik, A., Illes, K., Shabani, S., et al. (2019). Molecular mechanism of inhibition of acid ceramidase by carmofur. *J. Med. Chem.* 62 (2), 987–992. doi:10.1021/acs.jmedchem.8b01723
- Diaz Escarcega, R., McCullough, L. D., and Tsvetkov, A. S. (2021). The functional role of sphingosine kinase 2. *Front. Mol. Biosci.* 8, 683767. doi:10.3389/fmolb.2021.683767
- Doan, N. B., Alhajala, H., Al-Gizawi, M. M., Mueller, W. M., Rand, S. D., Connelly, J. M., et al. (2017a). Acid ceramidase and its inhibitors: a *de novo* drug target and a new class of drugs for killing glioblastoma cancer stem cells with high efficiency. *Oncotarget* 8 (68), 112662–112674. doi:10.18632/oncotarget.22637
- Doan, N. B., Nguyen, H. S., Al-Gizawi, M. M., Mueller, W. M., Sabbadini, R. A., Rand, S. D., et al. (2017b). Acid ceramidase confers radioresistance to glioblastoma cells. *Oncol. Rep.* 38 (4), 1932–1940. doi:10.3892/or.2017.5855
- Dowdy, T., Zhang, L., Celiku, O., Movva, S., Lita, A., Ruiz-Rodado, V., et al. (2020). Sphingolipid pathway as a source of vulnerability in IDH1mut glioma. *Cancers (Basel)* 12, 2910. doi:10.3390/cancers12102910
- Dumitru, C. A., Weller, M., and Gulbins, E. (2009). Ceramide metabolism determines glioma cell resistance to chemotherapy. *J. Cell Physiol.* 221 (3), 688–695. doi:10.1002/jcp.21907
- Farley, S., Stein, F., Haberkant, P., Tafesse, F. G., and Schultz, C. (2024). Trifunctional sphinganine: a new tool to dissect sphingolipid function. *ACS Chem. Biol.* 19 (2), 336–347. doi:10.1021/acscchembio.3c00554
- Fernández-García, P., Rosselló, C. A., Rodríguez-Lorca, R., Beteta-Göbel, R., Fernández-Díaz, J., Lladó, V., et al. (2019). The opposing contribution of SMS1 and SMS2 to glioma progression and their value in the therapeutic response to 2HOHA. *Cancers* 11 (1), 88. doi:10.3390/cancers11010088
- French, K. J., Upson, J. J., Keller, S. N., Zhuang, Y., Yun, J. K., and Smith, C. D. (2006). Antitumor activity of sphingosine kinase inhibitors. *J. Pharmacol. Exp. Ther.* 318 (2), 596 LP–603. doi:10.1124/jpet.106.101345
- Gault, C. R., Obeid, L. M., and Hannun, Y. A. (2010). An overview of sphingolipid metabolism: from synthesis to breakdown. *Adv. Exp. Med. Biol.* 688, 1–23. doi:10.1007/978-1-4419-6741-1_1
- Gisina, A., Kholodenko, I., Kim, Y., Abakumov, M., Lupatov, A., and Yarygin, K. (2022). Glioma stem cells: novel data obtained by single-cell sequencing. *Int. J. Mol. Sci.* 23 (22), 14224. doi:10.3390/ijms232214224
- Hannun, Y. A., and Obeid, L. M. (2018). Sphingolipids and their metabolism in physiology and disease. *Nat. Rev. Mol. Cell Biol.* 19 (3), 175–191. doi:10.1038/nrm.2017.107
- Hartmann, D., Lucks, J., Fuchs, S., Schiffmann, S., Schreiber, Y., Ferreirós, N., et al. (2012). Long chain ceramides and very long chain ceramides have opposite effects on human breast and colon cancer cell growth. *Int. J. Biochem. Cell Biol.* 44 (4), 620–628. doi:10.1016/j.biocel.2011.12.019
- Hawkins, C. C., Ali, T., Ramanadham, S., and Hjelmeland, A. B. (2020). Sphingolipid metabolism in glioblastoma and metastatic brain tumors: a review of sphingomyelinases and sphingosine-1-phosphate. *Biomolecules* 10 (10), 1357–1423. doi:10.3390/biom10101357
- Hawkins, C. C., Jones, A. B., Gordon, E. R., Harsh, Y., Ziebro, J. K., Willey, C. D., et al. (2023). Carmofur prevents cell cycle progression by reducing E2F8 transcription in temozolomide-resistant glioblastoma cells. *Cell Death Discov.* 9 (1), 451. doi:10.1038/s41420-023-01738-x
- Janneh, A. H. (2024). Sphingolipid signaling and complement activation in glioblastoma: a promising avenue for therapeutic intervention. *Biochem.* 4 (2), 126–143. doi:10.3390/biochem4020007
- Kayabolen, A., Yilmaz, E., and Bagci-Onder, T. (2021). IDH mutations in glioma: double-edged sword in clinical applications? *Biomed* 9 (7), 799. doi:10.3390/biomed9070799
- Landis, C. J., Tran, A. N., Scott, S. E., Griguer, C., and Hjelmeland, A. B. (2018). The pro-tumorigenic effects of metabolic alterations in glioblastoma including brain tumor initiating cells. *Biochim. Biophys. Acta - Rev. Cancer* 1869 (2), 175–188. doi:10.1016/j.bbcan.2018.01.004
- Lee, Y. S., Choi, K. M., Lee, S., Sin, D. M., Yoo, K. S., Lim, Y., et al. (2012). Myriocin, a serine palmitoyltransferase inhibitor, suppresses tumor growth in a murine melanoma model by inhibiting *de novo* sphingolipid synthesis. *Cancer Biol. Ther.* 13 (2), 92–100. doi:10.4161/cbt.13.2.18870
- Leverly, S. B. (2005). Glycosphingolipid structural analysis and glycosphingolipidomics. *Methods Enzymol.* 405 (1998), 300–369. doi:10.1016/S0076-6879(05)05012-3
- Lewis, C. S., Voelkel-johnson, C., and Smith, C. D. (2018). Targeting sphingosine kinases for the treatment of cancer. *Adv. Cancer Res.* 140, 295–325. doi:10.1016/bs.acr.2018.04.015
- Li, W., Cai, H., Ren, L., Yang, Y., Yang, H., Liu, J., et al. (2022). Sphingosine kinase 1 promotes growth of glioblastoma by increasing inflammation mediated by the NF-κB/IL-6/STAT3 and JNK/PTX3 pathways. *Acta Pharm. Sin. B* 12 (12), 4390–4406. doi:10.1016/j.apsb.2022.09.012
- Lladó, V., López, D. J., Iburguen, M., Alonso, M., Soriano, J. B., Escríbá, P. V., et al. (2014). Regulation of the cancer cell membrane lipid composition by NaChOleate: effects on cell signaling and therapeutic relevance in glioma. *Biochim. Biophys. Acta - Biomembr.* 1838 (6), 1619–1627. doi:10.1016/j.bbmem.2014.01.027
- Lopez, J., Lai-Kwon, J., Molife, R., Welsh, L., Tunariu, N., Roda, D., et al. (2023). A Phase 1/2A trial of idroxiolic acid: first-in-class sphingolipid regulator and glioma cell autophagy inducer with antitumor activity in refractory glioma. *Br. J. Cancer* 129 (5), 811–818. doi:10.1038/s41416-023-02356-1
- Louis, D. N., Perry, A., Wesseling, P., Brat, D. J., Cree, I. A., Figarella-Branger, D., et al. (2021). The 2021 WHO classification of tumors of the central nervous system: a summary. *Neuro Oncol.* 23 (8), 1231–1251. doi:10.1093/neuonc/noab106
- Lucke-Wold, B., Rangwala, B. S., Shafique, M. A., Siddiq, M. A., Mustafa, M. S., Danish, F., et al. (2024). Focus on current and emerging treatment options for glioma: a

- comprehensive review. *World J. Clin. Oncol.* 15 (4), 482–495. doi:10.5306/wjco.v15.i4.482
- Ma, S., Pan, X., Gan, J., Guo, X., He, J., Hu, H., et al. (2024). DNA methylation heterogeneity attributable to a complex tumor immune microenvironment prompts prognostic risk in glioma. *Epigenetics* 19 (1), 2318506. doi:10.1080/15592294.2024.2318506
- Marfia, G., Campanella, R., Navone, S. E., Di Vito, C., Riccitelli, E., Hadi, L. A., et al. (2014). Autocrine/paracrine sphingosine-1-phosphate fuels proliferative and stemness qualities of glioblastoma stem cells. *Glia* 62 (12), 1968–1981. doi:10.1002/glia.22718
- Martin, K. C., Ma, C., and Yip, S. (2023). From theory to practice: implementing the WHO 2021 classification of adult diffuse gliomas in neuropathology diagnosis. *Brain Sci.* 13 (5), 817. doi:10.3390/brainsci13050817
- Melero-Fernandez de Mera, R. M., Villaseñor, A., Rojo, D., Carrión-Navarro, J., Gradillas, A., Ayuso-Sacido, A., et al. (2022). Ceramide composition in exosomes for characterization of glioblastoma stem-like cell phenotypes. *Front. Oncol.* 11, 788100. doi:10.3389/fonc.2021.788100
- Merrill, A. H. (2011). Sphingolipid and glycosphingolipid metabolic pathways in the era of sphingolipidomics. *Chem. Rev. Am. Chem. Soc.* 111, 6387–6422. doi:10.1021/cr2002917
- Meyer, N., Henkel, L., Linder, B., Zielke, S., Tascher, G., Trautmann, S., et al. (2021). Autophagy activation, lipotoxicity and lysosomal membrane permeabilization synergize to promote pimizide- and loperamide-induced glioma cell death. *Autophagy* 17 (11), 3424–3443. doi:10.1080/15548627.2021.1874208
- Mullen, T. D., Hannun, Y. A., and Obeid, L. M. (2012). Ceramide synthases at the centre of sphingolipid metabolism and biology. *Biochem. J.* 441 (3), 789–802. doi:10.1042/BJ20111626
- Navone, S. E., Guarnaccia, L., Rizzaro, M. D., Begani, L., Barilla, E., Alotta, G., et al. (2023). Role of luteolin as potential new therapeutic option for patients with glioblastoma through regulation of sphingolipid rheostat. *Int. J. Mol. Sci.* 2024 25 (1), 130. doi:10.3390/ijms25010130
- Newton, J., Lima, S., Maceyka, M., and Spiegel, S. (2015). Revisiting the sphingolipid rheostat: evolving concepts in cancer therapy. *Exp. Cell Res.* 333 (2), 195–200. doi:10.1016/j.yexcr.2015.02.025
- Nganga, R., Oleinik, N., and Ogretmen, B. (2018). Mechanisms of ceramide-dependent cancer cell death. *Adv. Cancer Res.* 140, 1–25. doi:10.1016/bs.acr.2018.04.007
- NIHR (2021). LAM-561 in addition to radiation therapy and temozolomide for glioblastoma-adjuvant Licensing and market availability plans Currently in phase II/III clinical trials. Available at: <https://io.nihr.ac.uk/tech-briefings/lam-561-in-addition-to-radiation-therapy-and-temozolomide-for-glioblastoma-adjuvant/> (Accessed January 11, 2022).
- Norden, A. D., and Wen, P. Y. (2006). Glioma therapy in adults. *Neurologist* 12 (6), 279–292. doi:10.1097/01.nrl.0000250928.26044.47
- Ogretmen, B. (2017). Sphingolipid metabolism in cancer signalling and therapy. *Nat. Rev. Cancer* 18 (1), 33–50. doi:10.1038/nrc.2017.96
- Ostrom, Q. T., Bauchet, L., Davis, F. G., Deltour, I., Fisher, J. L., Langer, C. E., et al. (2014). The epidemiology of glioma in adults: a “state of the science” review. *Neuro Oncol.* 16 (7), 896–913. doi:10.1093/neuonc/nou087
- Paugh, S. W., Payne, S. G., Barbour, S. E., Milstien, S., and Spiegel, S. (2003). The immunosuppressant FTY720 is phosphorylated by sphingosine kinase type 2. *FEBS Lett.* 554 (1–2), 189–193. doi:10.1016/s0014-5793(03)01168-2
- Pruett, S. T., Bushnev, A., Hagedorn, K., Adiga, M., Haynes, C. A., Sullards, M. C., et al. (2008). Biodiversity of sphingoid bases (“sphingosines”) and related amino alcohols. *J. Lipid Res.* 49, 1621–1639. doi:10.1194/jlr.R800012-JLR200
- Puduvalli, V. K., Saito, Y., Xu, R., Kouraklis, G. P., Levin, V. A., and Kyritsis, A. P. (1999). Fenretinide activates caspases and induces apoptosis in gliomas. *Clin. Cancer Res.* 5 (8), 2230–2235.
- Puduvalli, V. K., Yung, W. K. A., Hess, K. R., Kuhn, J. G., Groves, M. D., Levin, V. A., et al. (2004). Phase II study of fenretinide (NSC 374551) in adults with recurrent malignant gliomas: a North American brain tumor consortium study. *J. Clin. Oncol.* 22 (21), 4282–4289. doi:10.1200/JCO.2004.09.096
- Pyne, N. J., and Pyne, S. (2020). Recent advances in the role of sphingosine 1-phosphate in cancer. *FEBS Lett.* 594 (22), 3583–3601. doi:10.1002/1873-3468.13933
- Pyne, S., and Pyne, N. J. (2000). Sphingosine 1-phosphate signalling in mammalian cells. *Biochem. J.* 349 (2), 385–402. doi:10.1042/0264-6021:3490385
- Rahman, M. A., and Ali, M. M. (2024). Recent treatment strategies and molecular pathways in resistance mechanisms of antiangiogenic therapies in glioblastoma. *Cancers* 16 (17), 2975. doi:10.3390/cancers16172975
- Raichur, S. (2020). Ceramide synthases are attractive drug targets for treating metabolic diseases. *Front. Endocrinol. (Lausanne)* 11, 483. doi:10.3389/fendo.2020.00483
- Rajaratnam, V., Islam, M. M., Yang, M., Slaby, R., Ramirez, H. M., and Mirza, S. P. (2020). Glioblastoma: pathogenesis and current status of chemotherapy and other novel treatments. *Cancers* 12 (4), 937. doi:10.3390/cancers12040937
- Riboni, L., Campanella, R., Bassi, R., Villani, R., Gaini, S. M., Martinelli-Boneschi, F., et al. (2002). Ceramide levels are inversely associated with malignant progression of human glial tumors. *Glia* 39 (2), 105–113. doi:10.1002/glia.10087
- Riebeling, C., Allegood, J. C., Wang, E., Merrill, A. H., and Futerman, A. H. (2003). Two mammalian longevity assurance gene (LAG1) family members, trh1 and trh4, regulate dihydroceramide synthesis using different fatty acyl-CoA donors. *J. Biol. Chem.* 278 (44), 43452–43459. doi:10.1074/jbc.M307104200
- Scorsetti, M., Ducray, F., Razis, E. D., Gaca-Tabaszewska, M., Bogusiewicz, J., and Bojko, B. (2022). Metabolomic and lipidomic profiling of gliomas—a new direction in personalized therapies. *Cancers (Basel)* 14 (20), 5041. doi:10.3390/cancers14205041
- Sousa, N., Geiß, C., Bindila, L., Lieberwirth, I., Kim, E., and Régner-Vigouroux, A. (2023). Targeting sphingolipid metabolism with the sphingosine kinase inhibitor SKI-II overcomes hypoxia-induced chemotherapy resistance in glioblastoma cells: effects on cell death, self-renewal, and invasion. *BMC Cancer* 23 (1), 762–823. doi:10.1186/s12885-023-11271-w
- Stupp, R., Mason, W. P., van den Bent, M. J., Weller, M., Fisher, B., Taphoorn, M. J. B., et al. (2005). Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. *N. Engl. J. Med.* 352 (10), 987–996. doi:10.1056/NEJMoa043330
- Tang, X., and Brindley, D. N. (2020). Lipid phosphate phosphatases and cancer. *Biomolecules* 10 (9), 1263. doi:10.3390/biom10091263
- Tea, M. N., Poonnoose, S. I., and Pitson, S. M. (2020). Targeting the sphingolipid system as a therapeutic direction for glioblastoma. *Cancers (Basel)* 12 (1), 111. doi:10.3390/cancers12010111
- UCSD (2024). Brain tumor trial → fluoxetine and cytotoxic lysosomal stress in glioma (FLIRT). Available at: <https://clinicaltrials.ucsd.edu/trial/NCT05634707> (Accessed October 30, 2024).
- Van Brocklyn, J. R., Lee, M. J., Menzelev, R., Olivera, A., Edsall, L., Cuvillier, O., et al. (1998). Dual actions of sphingosine-1-phosphate: extracellular through the gi-coupled receptor edg-1 and intracellular to regulate proliferation and survival. *J. Cell Biol.* 142 (1), 229–240. doi:10.1083/jcb.142.1.229
- Velasco, G., Carracedo, A., Blázquez, C., Lorente, M., Aguado, T., Haro, A., et al. (2007). Cannabinoids and gliomas. *Mol. Neurobiol.* 36 (1), 60–67. doi:10.1007/s12035-007-0002-5
- Wang, Z., Ge, X., Shi, J., Lu, B., Zhang, X., and Huang, J. (2022). SPTSSA is a prognostic marker for glioblastoma associated with tumor-infiltrating immune cells and oxidative stress. *Oxid. Med. Cell Longev.* 2022 (1), 6711085. doi:10.1155/2022/6711085
- Yaguchi, M., Shibata, S., Satomi, Y., Hirayama, M., Adachi, R., Asano, Y., et al. (2017). Antitumor activity of a novel and orally available inhibitor of serine palmitoyltransferase. *Biochem. Biophys. Res. Commun.* 484 (3), 493–500. doi:10.1016/j.bbrc.2017.01.075
- Young, N., and Van Brocklyn, J. R. (2007). Roles of sphingosine-1-phosphate (S1P) receptors in malignant behavior of glioma cells. Differential effects of S1P2 on cell migration and invasiveness. *Exp. Cell Res.* 313 (8), 1615–1627. doi:10.1016/j.yexcr.2007.02.009
- Yu, D., Xuan, Q., Zhang, C., Hu, C., Li, Y., Zhao, X., et al. (2020). Metabolic alterations related to glioma grading based on metabolomics and lipidomics analyses. *Metabolites* 10 (12), 478–511. doi:10.3390/metabo10120478
- Zaibaq, F., Dowdy, T., and Larion, M. (2022). Targeting the sphingolipid rheostat in gliomas. *Int. J. Mol. Sci.* 23 (16), 9255. doi:10.3390/ijms23169255
- Zheng, W., Kollmeyer, J., Symolon, H., Momin, A., Munter, E., Wang, E., et al. (2006). Ceramides and other bioactive sphingolipid backbones in health and disease: lipidomic analysis, metabolism and roles in membrane structure, dynamics, signaling and autophagy. *Biochim. Biophys. Acta* 1758 (12), 1864–1884. doi:10.1016/j.bbamem.2006.08.009
- Zhou, Y., Tao, L., Qiu, J., Xu, J., Yang, X., Zhang, Y., et al. (2024). Tumor biomarkers for diagnosis, prognosis and targeted therapy. *Signal Transduct. Target Ther.* 9 (1), 132–186. doi:10.1038/s41392-024-01823-2