



OPEN ACCESS

EDITED AND REVIEWED BY
Valerie Kouskoff,
The University of Manchester,
United Kingdom

*CORRESPONDENCE
Tong Ming Liu,
✉ dbsluimt@yahoo.com

RECEIVED 14 September 2023
ACCEPTED 09 October 2023
PUBLISHED 13 October 2023

CITATION
Liu TM and Wu Y (2023), Editorial:
Advancing gene and cell therapy using
human mesenchymal stem cells.
Front. Cell Dev. Biol. 11:1294460.
doi: 10.3389/fcell.2023.1294460

COPYRIGHT
© 2023 Liu and Wu. This is an open-
access article distributed under the terms
of the [Creative Commons Attribution
License \(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use, distribution or
reproduction in other forums is
permitted, provided the original author(s)
and the copyright owner(s) are credited
and that the original publication in this
journal is cited, in accordance with
accepted academic practice. No use,
distribution or reproduction is permitted
which does not comply with these terms.

Editorial: Advancing gene and cell therapy using human mesenchymal stem cells

Tong Ming Liu^{1*} and Yingnan Wu^{2,3}

¹Institute of Molecular and Cell Biology (IMCB), Agency for Science, Technology and Research (A*STAR), Singapore, Singapore, ²Department of Orthopaedic Surgery, Yong Loo Lin School of Medicine, National University of Singapore, Singapore, Singapore, ³NUS Tissue Engineering Program, Life Sciences Institute, National University of Singapore, Singapore, Singapore

KEYWORDS

mesenchymal stem cells, senescence, cell therapy, gene therapy, MSC-EVs

Editorial on the Research Topic

Advancing gene and cell therapy using human mesenchymal stem cells

Human mesenchymal stem cells (hMSCs) have emerged as an attractive cell source for regenerative medicine. MSCs are adult stem cells, which reside in almost all adult tissues and birth-associated tissues. As one of the most clinically used stem cells, MSCs have been used in over 1,500 clinical trials to treat over 30 diseases. However, primary MSCs have limited life span, resulting in limited cells (Liu et al., 2013; Liu et al., 2020; Liu, 2021). During *in vitro* expansion, MSCs gradually lose differentiation potential and become senescent, which greatly compromises their therapeutic applications. Gene and cell therapy of MSCs offers a promising treatment option for regenerative medicine. This Research Topic contains a collection of articles exploring MSC senescence, iMSCs from iPSCs, chondrogenesis of iMSCs, and the application of MSCs or MSC-Exos for repairing heart injuries and treating stress urinary incontinence.

MSCs become senescent during *in vitro* expansion. To understand the molecular basis of MSC senescence and identify biomarkers used to evaluate MSC senescence, Yang et al. re-analyzed sequencing data in the public domain and established a regulatory network of “transcription factor (TF)-miRNA-Target” associated with the senescence and quality status of MSCs during passage. This study provides insights into key molecules for evaluating the passage-dependent replicative senescence of MSCs and deciphers the mechanism of miRNAs regulating the senescence of MSCs during *in vitro* expansion. This study provides passage-dependent indicators for monitoring the quality of MSCs during the clinical application.

Induced pluripotent stem cells (iPSCs) can self-renew unlimitedly and differentiated into any type of cells, including MSCs. Therefore iPSC-MSCs (iMSCs) can provide unlimited iPSC-MSCs to overcome limited MSCs with primary MSCs and represent an alternative source to MSCs (Liu et al., 2020; Liu, 2021). However, the use of iPSCs for regenerative medicine has concern over safety due to possible iPSCs contamination in iMSCs causing tumors. In this Research Topic, Al-Akashi et al. used the Brequinar (BRQ) to eradicate the leftover iPSCs and purify iPSC-MSCs. BRQ is a key enzyme in *de novo* pyrimidine biosynthesis, which inhibits dihydroorotate dehydrogenase (DHODH) to prevent the growth of 3D iPSC aggregates. The authors found that BRQ can be used to eradicate the remaining undifferentiated hiPSCs, there were no changes in survival, differentiation

potential and gene expression between BRQ-treated iMSCs and non-treated iMSCs, suggesting that BRQ can effectively purify iMSCs from hiPSCs, this will reduce concern over safety of iMSC based therapy. In addition, iMSC quality is critical to clinical application. Using a stepwise protocol to differentiate iPSCs toward MSCs (iMSCs) via the induction of neural crest cells, [Zujur et al.](#) compared two types of xeno-free culture media, XSF and T1 media to generate iMSCs from iPSCs. They also reported that TD-198946 increased the size of chondrogenic spheroids and improved chondrogenesis of iMSCs, shown by enhanced chondrogenic genes and stains. TD-198946 is a thienopyridone derivative and has been shown to be a potent chondrogenic factor. Subcutaneous transplantation of chondrogenic spheroids from iMSCs showed that cartilage extracellular matrix increased without a noticeable effect on hypertrophy. This study provides insights into generating good quality of iMSCs for cartilage tissue engineering.

Extracellular vesicles secreted by MSCs (MSC-EVs) have recently been shown as a promising alternative to MSCs for regenerative medicine ([Yang et al., 2023](#)). MSC-EVs are extracellular nanovesicles containing bioactive proteins, nucleic acids, lipids, enzymes and metabolites. Compared with the direct use of MSCs, MSC-EVs have advantages in higher safety, lower immunogenicity, lower immune rejection, effortless storage and the ability to cross biological barriers and avoid complications. [Zhu et al.](#) reviewed the applications of exosomes derived from MSCs for the repair of heart injuries. MSC exosomes (MSC-Exos) decrease the apoptosis of myocardial cells and fibrosis, prevent the infiltration of inflammatory factors and increase the formation of blood vessels. The authors discussed MSCs exosomes with a specific focus on the role of MSC-Exos miRNA in myocardial injury repair. Non-coding RNAs (NC-RNAs) in MSC-Exos are affected by various pretreatments, such as drugs, chemicals and hypoxia. The authors further discussed the cardiac protective effects of long non-coding RNAs (lncRNAs) in MSC-Exos and underlying molecular mechanisms of MSC-Exos. This review provides valuable insights for advancements in heart tissue engineering research using MSC-EVs.

Stress urinary incontinence (SUI) is the unintentional loss of urine caused by increased abdominal pressure during physical activity, which adversely affects the quality of life of patients. Current available therapies are not effective in all patients, MSCs represent a promising therapy for treating SUI. [Liu et al.](#) reviewed

MSC-based treatments of SUI and summarized four treatment strategies, including single MSC type therapy, MSC-based combination therapy, MSC secretome treatment, and other factors affecting MSC therapy. This review discussed the molecular mechanisms underlying MSC mediated SUI treatment. Although evidence supports the safety and effectiveness of MSCs or MSC-EVs for treating SUI, there are limitations with current studies, including small sample sizes, most data from animal studies and few from clinical trials, short-term results and unclear mechanism of action, etc. Therefore, more research and clinical trials are needed for a better understanding and optimization of these treatments.

Author contributions

TL: Conceptualization, Project administration, Supervision, Writing—original draft, Writing—review and editing. YW: Writing—original draft.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This work was funded by the A*STAR Career Development Award (C210112047 to TL) and pre-gap funding (I22D1AG050 to TL).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

- Liu, T. M., Ng, W. M., Tan, H. S., Vinitha, D., Yang, Z., Fan, J. B., et al. (2013). Molecular basis of immortalization of human mesenchymal stem cells by combination of p53 knockdown and human telomerase reverse transcriptase overexpression. *Stem Cells Dev.* 22 (2), 268–278. doi:10.1089/scd.2012.0222
- Liu, T. M., Yildirim, E. D., Li, P., Fang, H. T., Denslin, V., Kumar, V., et al. (2020). Ascorbate and iron are required for the specification and long-term self-renewal of human skeletal mesenchymal stromal cells. *Stem Cell Rep.* 14, 210–225. doi:10.1016/j.stemcr.2020.01.002
- Liu, T. M. (2021). Application of mesenchymal stem cells derived from human pluripotent stem cells in regenerative medicine. *World J. Stem cells* 13 (12), 1826–1844. doi:10.4252/wjsc.v13.i12.1826
- Yang, Y., Wu, Y., Yang, D., Neo, S. H., Kadir, N. D., Goh, D., et al. (2023). Secretive derived from hypoxia preconditioned mesenchymal stem cells promote cartilage regeneration and mitigate joint inflammation via extracellular vesicles. *Bioact. Mater* 27, 98–112. doi:10.1016/j.bioactmat.2023.03.017