



# VEX1 Influences mVSG Expression During the Transition to Mammalian Infectivity in *Trypanosoma brucei*

Eliane Tihon<sup>1†</sup>, Karinna Rubio-Peña<sup>1‡</sup>, Annick Dujeancourt-Henry<sup>1</sup>, Aline Cruzols<sup>2</sup>, Brice Rotureau<sup>2,3</sup> and Lucy Glover<sup>1\*</sup>

<sup>1</sup>Trypanosome Molecular Biology, Institut Pasteur, Université Paris Cité, Paris, France, <sup>2</sup>Trypanosome Transmission Group, Trypanosome Cell Biology Unit, INSERM U1201 and, Institut Pasteur, Paris, France, <sup>3</sup>Parasitology Lab, Institut Pasteur of Guinea, Conakry, Guinea

## OPEN ACCESS

### Edited by:

Luisa M. Figueiredo,  
Universidade de Lisboa, Portugal

### Reviewed by:

Nikolay Kolev,  
Yale University, United States  
Martin Craig Taylor,  
University of London, United Kingdom

### \*Correspondence:

Lucy Glover  
lucy.glover@pasteur.fr

### †Present address:

Eliane Tihon,  
Department of Human Pharma,  
Medical Affairs, SCS Boehringer  
Ingelheim Comm. V, Brussels, Belgium

‡These authors have contributed  
equally to this work

### Specialty section:

This article was submitted to  
Epigenomics and Epigenetics,  
a section of the journal  
Frontiers in Cell and Developmental  
Biology

Received: 09 January 2022

Accepted: 28 February 2022

Published: 05 April 2022

### Citation:

Tihon E, Rubio-Peña K,  
Dujeancourt-Henry A, Cruzols A,  
Rotureau B and Glover L (2022) VEX1  
Influences mVSG Expression During  
the Transition to Mammalian Infectivity  
in *Trypanosoma brucei*.  
Front. Cell Dev. Biol. 10:851475.  
doi: 10.3389/fcell.2022.851475

The *Trypanosoma (T) brucei* life cycle alternates between the tsetse fly vector and the mammalian host. In the insect, *T. brucei* undergoes several developmental stages until it reaches the salivary gland and differentiates into the metacyclic form, which is capable of infecting the next mammalian host. Mammalian infectivity is dependent on expression of the metacyclic variant surface glycoprotein genes as the cells develop into mature metacyclics. The VEX complex is essential for monoallelic *variant surface glycoprotein* expression in *T. brucei* bloodstream form, however, initiation of expression of the surface proteins genes during metacyclic differentiation is poorly understood. To better understand the transition to mature metacyclics and the control of metacyclic *variant surface glycoprotein* expression we examined the role of VEX1 in this process. We show that modulating VEX1 expression leads to a dysregulation of variant surface glycoprotein expression during metacyclogenesis, and that following both *in vivo* and *in vitro* metacyclic differentiation VEX1 relocalises from multiple nuclear foci in procyclic cells to one to two distinct nuclear foci in metacyclic cells - a pattern like the one seen in mammalian infective bloodstream forms. Our data suggest a role for VEX1 in the metacyclic differentiation process and their capacity to become infectious to the mammalian host.

**Keywords:** *Trypanosoma brucei*, VSG, antigenic variation, monoallelic expression, metacyclogenesis

## INTRODUCTION

The vector borne protozoan parasite *Trypanosoma (T) brucei* is the causative agent of Human African and Animal African Trypanosomiasis and remains today a pervasive public health issue in sub-Saharan Africa. Trypanosomes have a digenetic life cycle that transitions between the mammalian host and the tsetse fly (Glossinidae family) insect vector. During the life cycle, the trypanosome undergoes several important developmental transitions and includes the formation of up to 10 different morphological forms (Rotureau and Van Den Abbeele 2013) broadly grouped into trypomastigote and epimastigote morphotypes, and are defined by the relative positions of the nuclear and mitochondrial (kinetoplast) DNA in the cell (Hoare 1971).

Within the mammalian host, trypanosomes are covered in a dense layer of a single species of variant surface glycoprotein (VSG) and exist as proliferative slender forms or G<sub>1</sub> arrested stumpy forms (Matthews 2005). Stumpy forms are pre-adapted to survival in the insect host and differentiate into procyclic forms, replacing the VSG coat with a procyclin coat following a tsetse fly blood meal. The trypanosomes then colonise the cardia, posterior midgut (Rose et al., 2020) and eventually

migrate to the salivary glands (Van Den Abbeele et al., 1999; Sharma et al., 2008). The second major developmental transition in the tsetse fly is known as metacyclogenesis (Tetley et al., 1987; Rotureau et al., 2012), and occurs in the salivary glands. During this process epimastigote parasites that are attached to the salivary gland epithelium asymmetrically divide to produce pre-metacyclic cells that mature into mammalian infective metacyclic cells (Rotureau et al., 2012). Metacyclic trypanosomes acquire mammalian infectivity in the tsetse fly salivary gland where they begin to express a stage specific *metacyclic variant surface glycoprotein (mVSG)* gene (Tetley et al., 1987; Graham and Barry 1995). This mVSG coat is critical to the parasites ability to infect the mammalian host as it pre-adapts them for survival in the bloodstream (Vickerman 1969; Vickerman 1985). The life cycle is thus completed with the bite of a tsetse fly which deposits metacyclic trypanosomes into the mammalian host dermis (Caljon et al., 2016) where they differentiate and proliferate in the bloodstream and extracellular fluid and tissue (Capewell et al., 2016; Trindade et al., 2016). It is here, in the mammalian host, that the mVSG surface coat is replaced with a bloodstream form VSG coat (Stijlemans et al., 2016).

As with bloodstream form VSG expression, only one mVSG is present on the surface of the cell (Tetley et al., 1987; Ramey-Butler et al., 2015), and expression of mVSG genes transitions from multi-mVSG expression in pre-metacyclics to singular mVSG gene expression in mature metacyclic cells as monoallelic expression is established (Hutchinson et al., 2021). VSG expression is tightly controlled, and VSG genes are transcribed from highly specialised loci known as expression sites. The metacyclic expression site (MES) share some similarity to the bloodstream form expression site (BES) in that they both: contain a single VSG gene, are found at telomeres and are transcribed by RNA Pol-I (Barry et al., 1998; Ramey-Butler et al., 2015), and although MES and BES promoters are not conserved (Ginger et al., 2002), they are both recognised by the same CITFA Pol-I transcription factor (Kolev et al., 2017). However, while the BES is a polycistronic transcription unit composed of the VSG gene and several expression site associated genes (ESAGS), the MES is a monocistronic unit, harbouring only the mVSG gene (Alarcon et al., 1994). Very little is understood about the regulation of mVSG genes during the developmental transition to metacyclic form cells in the salivary gland of the tsetse fly. This is mostly due to the inability to culture quiescent and non-proliferative metacyclic cells that has hampered molecular studies on this life cycle stage. This has been overcome by ectopic over expression of RNA binding protein 6 (RBP6), in cultured procyclic cells which leads to the development of mammalian infective metacyclic forms (Kolev et al., 2012).

Several chromatin remodelling factors (Alsford et al., 2012), telomere binding proteins (Yang et al., 2009), the expression site body (ESB) (Navarro and Gull 2001), the inositol phosphate pathway (Cestari and Stuart 2015) and the histone chaperone CAF-1 (Faria et al., 2019) have all been implicated in the regulation and maintenance of bloodstream form VSG expression. The single active BES is also depleted of

nucleosomes (Figueiredo and Cross 2010; Stanne and Rudenko 2010). This suggests control at the level of transcription, elongation and that chromatin reorganisation is critical for singular VSG expression. A novel chromatin protein, TbSAP is required for silencing mVSG genes in bloodstream form cells (Davies et al., 2021), and a targeted RNAi screen revealed 22 positive and negative regulators required for the developmental transitions towards mammalian infectivity (Toh et al., 2021). The VEX complex (Glover et al., 2016; Faria et al., 2019) is a monoallelic regulator that restricts VSG transcription to a single telomere, recruits the RNA splicing machinery to ensure high levels of processing (Faria et al., 2021) and is also required for silencing mVSG genes in bloodstream form parasites. The VEX complex is composed of VEX1, VEX2 (Glover et al., 2016; Faria et al., 2019), is enriched as one to two foci within the nucleus with one immediately adjacent to the ESB, or the site of active VSG expression, within a telomere cluster (Glover et al., 2016; Faria et al., 2019). Following *in vitro* differentiation, the VEX complex initially relocates to the nuclear periphery, as has been reported for the active expression site (Landeira and Navarro 2007), but then redistributes within the nucleus. In the cultured procyclic insect stage cells, the VEX focus appears to be concomitant with all telomeres, suggesting that at least VEX1 differentially binds to telomeres in different life cycle stages (Glover et al., 2016), the purpose of this altered distribution remains unknown. Here we show that VEX1 is required for initiation of mVSG expression during metacyclogenesis and that VEX1 focal accumulation is life cycle stage-dependent.

## MATERIALS AND METHODS

### *Trypanosoma brucei* Growth and Manipulation

Procyclic stage *Trypanosoma brucei* PT1 (Trenaman et al., 2019) cells were grown in SDM-79 medium at 27°C. Cell density was determined using a haemocytometer. For transformation,  $2 \times 10^7$  cells were spun for 10 min at 1,000 g at room temperature the supernatant discarded and washed in 2 ml of prewarmed cytomix and spun as before. The cell pellet was resuspended in prewarmed 100  $\mu$ L cytomix solution (van den Hoff et al., 1992) with 10  $\mu$ g linearised DNA and placed in a 0.2 cm gap cuvette, and nucleofected (Lonza) using the X-014 program. The transfected cells were placed into 10 ml SDM-79 medium only and placed in an incubator to allow the cells to recover overnight. Serial dilutions plated out into 96 well plates at a 1:25, 1:50, and neat.  $1 \times 10^6$ /ml wild type or untransformed cells were added to the dilutions to condition the medium. Antat 1.1E (EATRO1125) cells were grown in HMI-11 medium at 37.4°C with 5% CO<sub>2</sub> and the density of cell cultures measured using a haemocytometer keeping cells below  $1 \times 10^5$  cells/ml. Transformation of cell lines was carried out by centrifuging  $2.5 \times 10^7$  cells at 1,000 g for 10 min at room temperature. The cell pellet was resuspended with 10  $\mu$ g linearized DNA in 100  $\mu$ L of warm cytomix solution (van den Hoff et al., 1992), placed in a cuvette (0.2 cm gap) and transformed using a Nucleofector™

**TABLE 1** | Primers used in this study.

Cloning	
RBP6 Forward	GATCAAGCTTATGTTCTACCCCAACAGCCCG
RBP6 Reverse	GATCGGATCTCAACCAGCGGCACCG
qPCR	
Actin F	GTACCACTGGCATTGTTCTCG
Actin R	CTTCATGAGATATTCGTCAGGTC
VEX1-4 F	ACGACCGAAGTTGTTGGGT
VEX1-4 R	TAACCTTCTGCTGCTGACCG
RBP6-4 F	TTTTGCCATGCGGAAGATGC
RBP6-4 R	GGGAACCCGCATGAACGTAT
mVSG 397 Forward	TGAAGCTGTGAAAGGGACAG
mVSG 397 Reverse	GAGGGCGAATTGTTTGTATTAGG
mVSG 531 Forward	GACGAAAGCCTGGGTAACATAAA
mVSG 531 Reverse	CCGCAGCTCGTTGATAGTATTG
mVSG 639 Forward	CCGACGATGAACACAGTTGA
mVSG 639 Reverse	TCTATGCCGTTCCGCTTTAC
mVSG 653 Forward	GGGCTGTTTCGCGACTAATA
mVSG 653 Reverse	CGTGGTGAAGTCTCCTGTTT
mVSG 1954 Forward	GCAGAGGCCTTAGCACTAAAT
mVSG 1954 Reverse	GGAGTTGACTTTCCTCCATCAG
mVSG559 F1	CAGAGCAAACCAGGCGCTG
mVSG559 R1	GTGTGTCCGCTGCAGTCG
mVSG559 F2	ACTGCCTGAGCTAAAGGCAGA
mVSG559 R2	ACCGCGTAGCCGTTAGTGTG
mVSG636 F1	ACGTTGGCAGCCAATGCAG
mVSG636 R1	AAGCTGCGCTACACCGTC
mVSG636 F2	ATCAGCCATCGGCGAACAAG
mVSG636 R2	CCAGCAACGTGCTAGCTGC
mVSG3591 F1	TTAACGGCGGCGACTGGCA
mVSG3591 R1	CGCTGGGCTGCCTTGACAA
mVSG3591 F2	AGCGACGATGGAGCCGTAA
mVSG3591 R2	CTTGCTTTGGCTGCCTGTG

(Lonza) (X-001 function). Transfected cells were recovered in 36 ml of warm HMI-11 at 37°C for 4–6 h, after which cells were plated out in 48 well plates at 1:5, 1:10, 1:50 and neat serial dilutions with the required drug selection. For metacyclic differentiation induction, the PT1 RBP6 overexpression cell line was grown in SDM-80 medium with 10% heat inactivated FBS, without glucose and 50 mM N-acetyl glucose-amine to block uptake of residual glucose molecules from the FBS (Dolezelova et al., 2020). RBP6 overexpression was induced with 10 µg/ml of tetracycline and the cell line maintained between  $2\text{--}5 \times 10^6$  cells/mL—exponential mid-log growth phase. Hygromycin and Blasticidin were selected at  $2 \mu\text{g ml}^{-1}$ ,  $2.5 \mu\text{g ml}^{-1}$ , and  $10 \mu\text{g ml}^{-1}$  respectively. Puromycin, phleomycin, hygromycin, blasticidin, and tetracycline were maintained at  $1 \mu\text{g ml}^{-1}$ .

## Cell Line Set Up

To construct PT1 RBP6 overexpression cell line, RBP6 was amplified from wild type genomic DNA using primers RBP6F and RBP6R (See **Table 1** for sequence) and cloned into pRPa vector (Alsford et al., 2005) using HindIII—BamH1. The resulting construct was linearized with AscI. To construct the constitutive VEX1 overexpression construct pRPaVEX1, we digested pRPaVEX1<sup>i6m</sup> and pRPaVEX1<sup>isl</sup> with HindIII—BamH1 and

ligated. pRPaVEX1<sup>i6m</sup>, pRPaVEX1<sup>isl</sup> and pNATVEX1<sup>12myc</sup> are described in (Glover et al., 2016).

## Immunofluorescence Microscopy

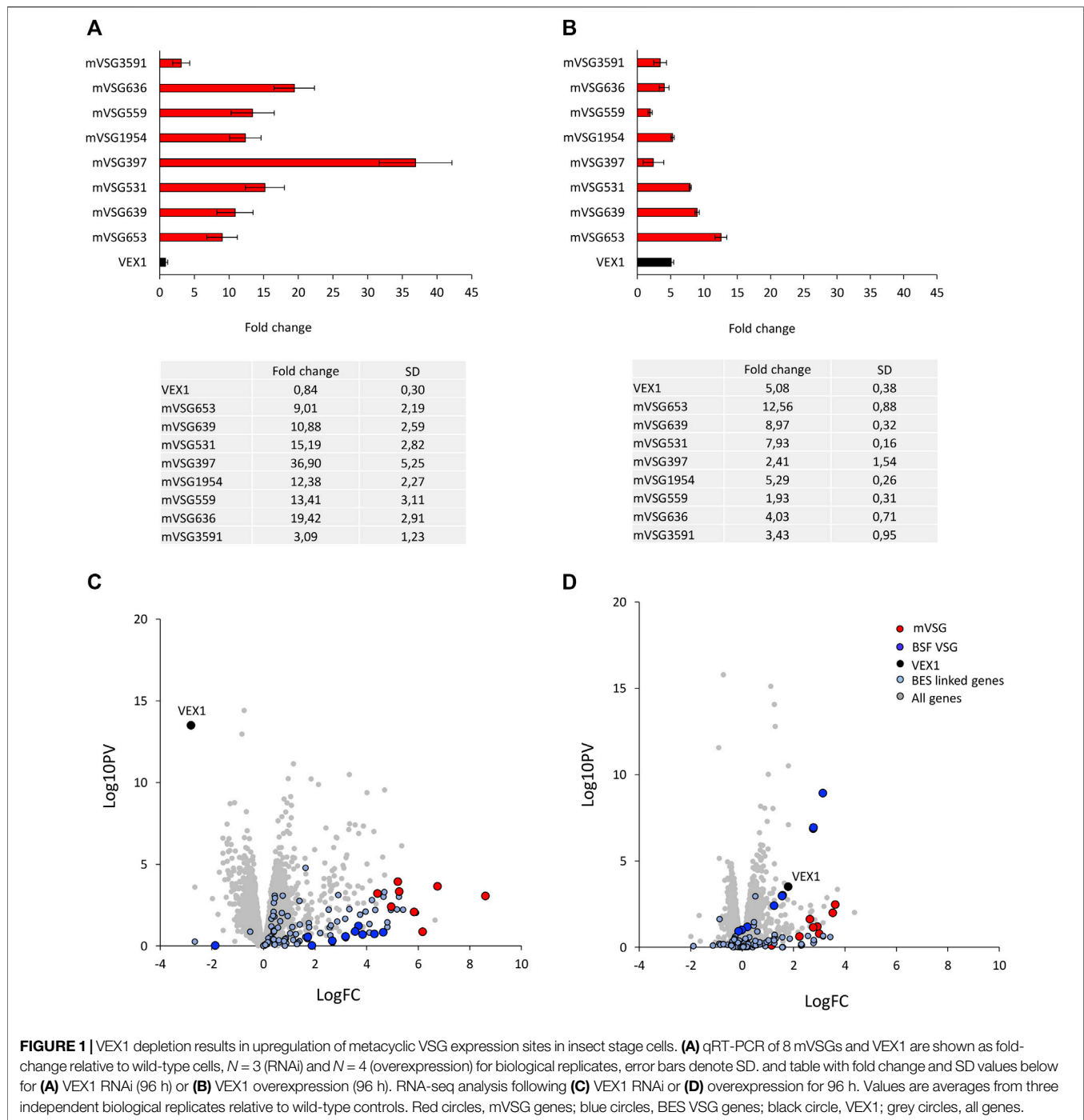
Immunofluorescence analysis was carried out using standard protocols as described previously (Glover et al., 2013). Rabbit α-myc (Cell Signalling, # 71D10) (1: 200) and rabbit α-CRD (1: 500) [Davids-Biotechnologies, (Zamze et al., 1988)]. Secondary anti-sera used were goat α-rabbit AlexaFluor<sup>®</sup> 555, goat α-rabbit AlexaFluor<sup>®</sup> 488 (1:1,000). Samples were mounted in Vectashield (Vector Laboratories) containing 4, 6-diamidino-2-phenylindole (DAPI). In *T. brucei*, DAPI-stained nuclear and mitochondrial DNA can be used as cytological markers (Woodward and Gull 1990); Images were captured using a ZEISS Imager 72 epifluorescence microscope with an Axiocam 506 mono camera and images were processed in ImageJ.

## RNA Analysis

RNA samples were taken at 0-, 4- and 8-days post RBP6 and VEX1 overexpression or knockdown induction. RNA was extracted from 50 ml of culture at  $2 \times 10^6$  cells/ml. RNA-seq was carried out on a BGISEq platform at The Beijing Genome Institute (BGI). Reads were mapped to a hybrid genome assembly consisting of the *T. brucei* 427 reference genome plus the bloodstream VSG-ESs (Hertz-Fowler et al., 2008; Cross et al., 2014; Muller et al., 2018). Bowtie 2-mapping was used with the parameters --very-sensitive --no-discordant --phred33. Alignment files were manipulated with SAMtools (Li et al., 2009). Per-gene read counts were derived using the Artemis genome browser (Carver et al., 2012); MapQ, 0. Read counts were normalised using edgeR and differential expression was determined with classic edgeR. RPKM values were derived from normalised read counts in edgeR (Robinson et al., 2010).

## qPCR and RT-qPCR Analysis

The expression levels of *RBP6*, *VEX1*, and *mVSG* genes was analysed by RT-qPCR using Luna Universal qPCR MasterMix (NEB) with 500 nM of primers. All primer pairs are listed in **Table 1**. RNA was extracted using a Qiagen RNeasy Kit and the samples were treated with DNase 1 for 1 h according to manufactures instructions and eluted in 30 µL of RNase free water. The samples were quantified using a Nanodrop (ThermoFisher). cDNA was prepared using SuperScript IV (ThermoFisher) following the supplier instructions from 1–2 µg RNA with a polyT primer. For each pair of primers (used at 500 nM), triplicates of each sample were run per plate (Hard-shell PCR Plates 96 well, thin wall; Bio-Rad), which were sealed with Microseal “B” Seals (BioRad). All experiments were run on a CFX96 Touch Real-time Detection system with a C1000 Touch Thermal cycler (Bio-Rad), using the following PCR cycling conditions: 95°C for 1 min, followed by 40 cycles of 95°C for 15 s and 60°C for 30 s (fluorescence intensity data collected at the end of the last step). Data was then analysed by relative quantification using the  $\Delta\Delta\text{Ct}$  method (CFX

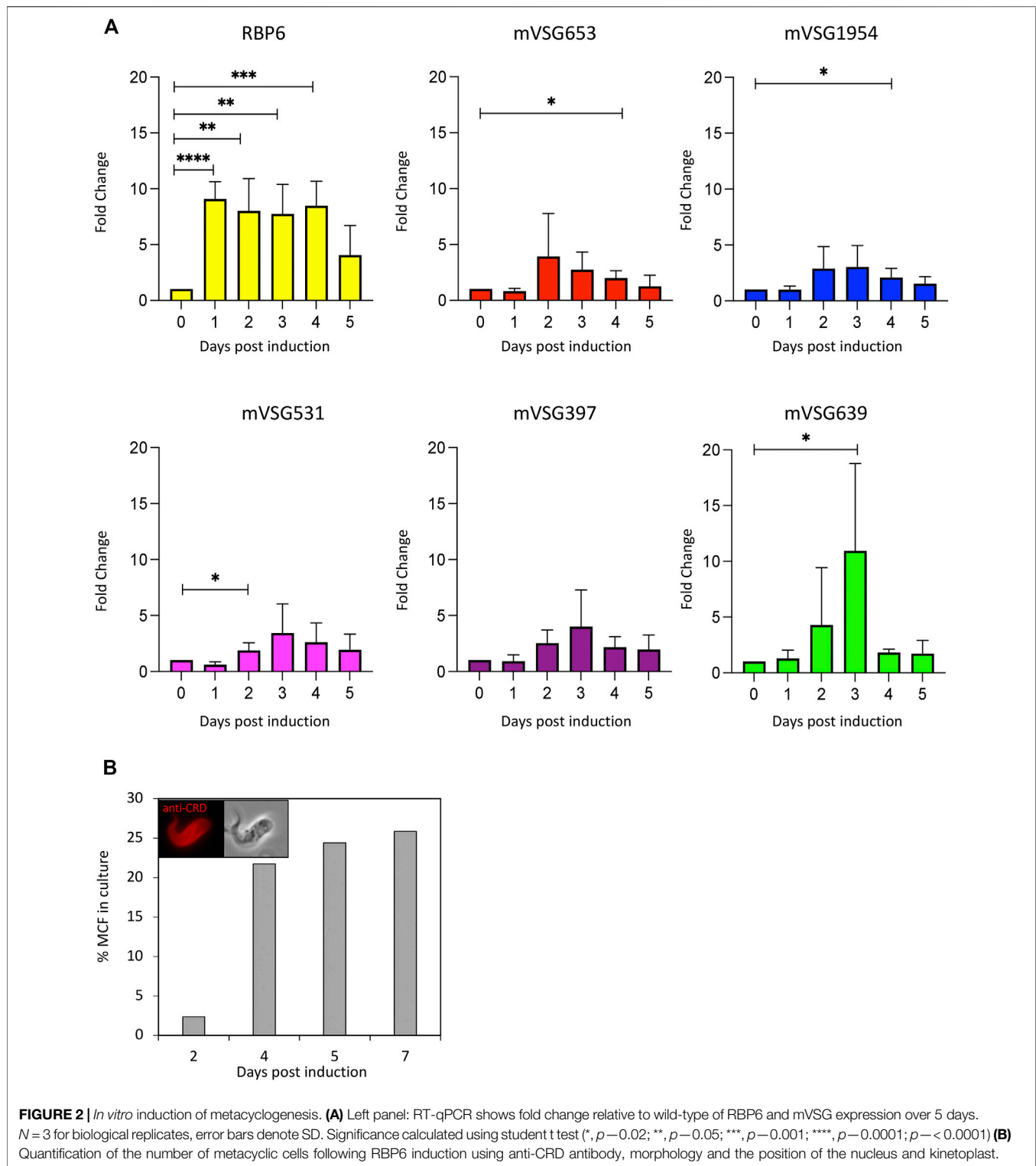


Maestro software—Bio-Rad) and Cq determination regression was used. In all cases, product abundance was determined relative to an actin control locus.

## Fly Infections

Tsetse flies (*Glossina morsitans*) were maintained at 27°C and 70% hygrometry in Roubaud cages, in groups of 50 male flies per cage. Pleomorphic trypanosome cell lines were maintained

at  $1 \times 10^5$  cells/ml density in HMI-9 medium plus 10% FBS at 37°C with 5% CO<sub>2</sub>. *In vitro* stumpy differentiation was induced in HMI-9, supplemented with 10% FBS without antibiotics, by adding 8-pCPT-2'-O-Me-5'-AMP (5 μM) (BioLog-Life Science Institut) to the culture 48 h before fly infection (Laxman et al., 2006). On the day of infection, trypanosomes were resuspended at  $10^6$  cells per ml in SDM79 with no antibiotics supplemented with 10 mM



glutathione prior infection (MacLeod et al., 2007). Flies were fed on infected media through a silicone membrane and maintained until dissection by feeding three times per week

on sheep's blood in heparin. Flies were starved for 2 days before dissection at day 28. Imaging was carried out using a ZEISS Imager 72 epifluorescence microscope with an Axiocam

**TABLE 2** | RT-qPCR values for RBP6, VEX1 and mVSG expression Fold change and SD values N = 3 (Parental), N = 4 (overexpression) and N = 2 (RNAi) for biological replicates.

	RBP6		mVSG653		mVSG1954		mVSG531		mVSG639		mVSG397	
	Fold change	SD	Fold change	SD	Fold change	SD	Fold change	SD	Fold change	SD	Fold change	SD
D0	1	0	1	0	1	0	1	0	1	0	1	0
D1	9.08	1.55	0.83	0.24	1.00	0.32	0.61	0.27	1.30	0.74	0.90	0.58
D2	8.01	2.89	3.93	3.85	2.90	1.96	1.90	0.67	4.30	5.14	2.55	1.16
D3	7.74	2.65	2.74	1.59	3.04	1.92	3.43	2.60	10.94	7.84	4.01	3.28
D4	8.47	2.20	1.99	0.68	2.09	0.83	2.63	1.70	9.26	14.87	2.17	0.95
D5	4.07	2.64	1.26	0.99	1.54	0.64	1.95	1.39	1.73	1.18	1.98	1.29

506 mono camera. Single images (for DIC) or multichannel stacks (for fluorescence) of images every 0.24  $\mu\text{m}$  were acquired. When maximum intensity Z-projections are presented, they were generated using Fiji (Schlindelin et al., 2012).

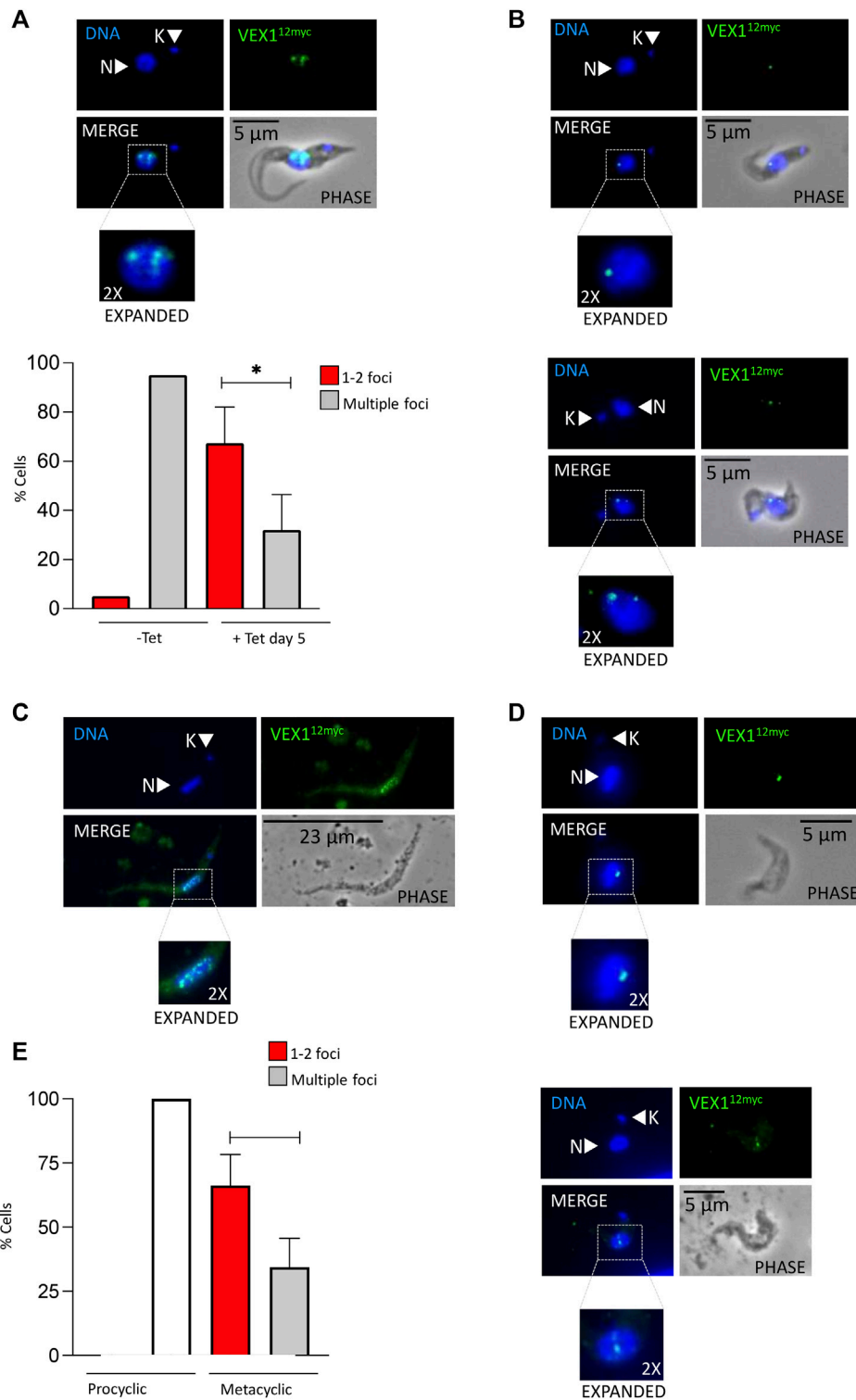
## RESULTS

### Loss of VEX1 Results in mVSG Expression in Insect Stage Cells

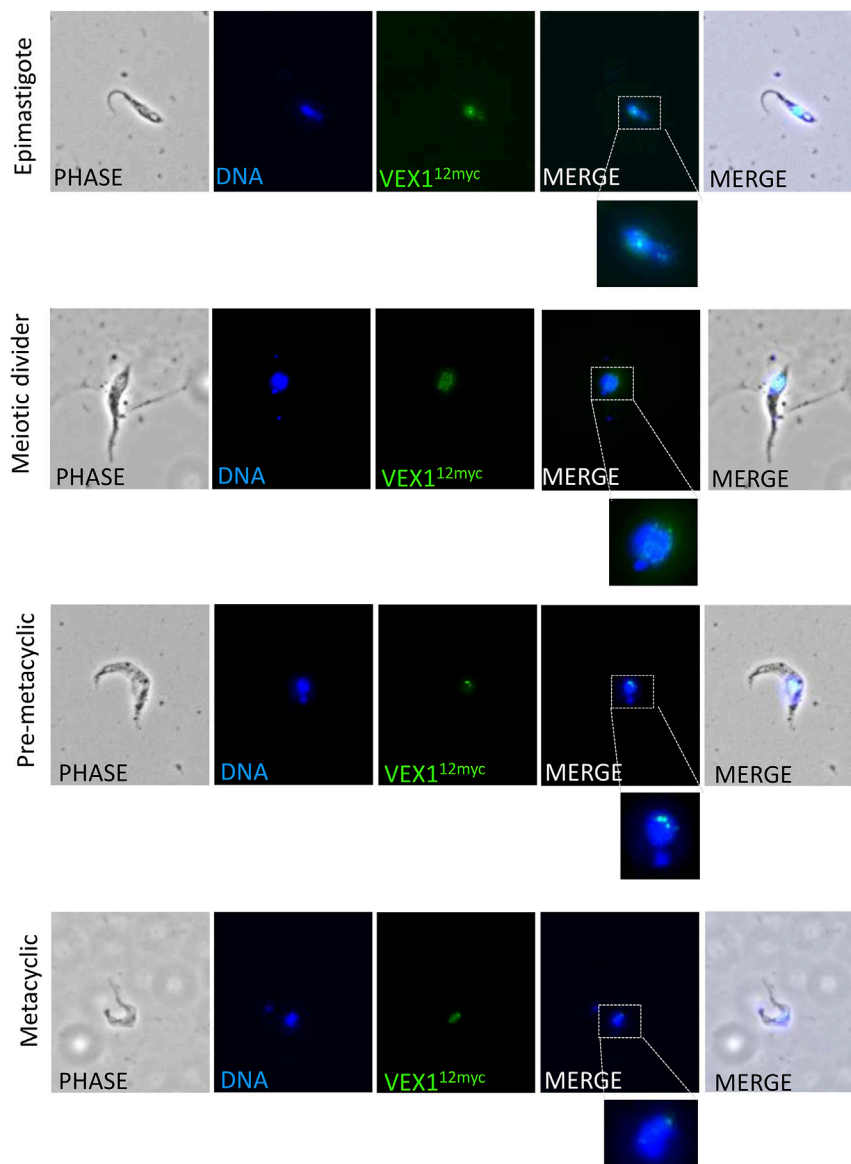
The transition between developmental stages is accompanied by changes in gene expression, including the silencing or activation of both the BESs and MESs. The bloodstream form to procyclic stage differentiation is marked by the replacement of the VSG surface coat with EP and GPEET procyclins (Acosta-Serrano et al., 2001). Given the VEX complex association with the VSG transcription compartment in the bloodstream form cells and that it redistributes upon differentiation (Glover et al., 2016; Faria et al., 2019; Faria et al., 2021), we wanted to ask whether VEX1 was required for silencing BES and MESs in the procyclic stage cells. We first assessed expression of the 8 mVSG genes (Muller et al., 2018) and VEX1 by RT-qPCR in VEX1 overexpression or knock down backgrounds (Figures 1A,B). VEX1 RNAi led to a greater derepression of the 8 mVSG genes (Figure 1A; Supplementary Figure S1A) compared to VEX1 overexpression (Figure 1B; Supplementary Figure S1B)—average fold change of 15,04 compared to 5,82 respectively. Given this striking difference between expression of the mVSG genes according to the VEX1 expression level, we wanted to determine whether this was similar for the BES VSGs. Transcriptomic analysis of VEX1 silenced by RNAi or overexpressed for 96 h showed similar patterns to the RT-qPCR, where RNAi led to a greater derepression of the 8 mVSG genes than overexpression, suggesting a primarily silencing function for VEX1 in procyclic stage cells. When we analysed BES linked genes, we found that overall, there was a subtler derepression as compared to the mVSG genes (Figures 1C,D), but this was more pronounced following VEX1 RNAi, again suggesting that VEX1 is primarily required for silencing in this life cycle stage. Our data point to VEX1 having a role as a negative regulator of VSG expression in procyclic stage cells, where knocking down expression of VEX1 from the cell has a stronger effect compared to overexpression. This suggests that VEX1 is required for efficient silencing of mVSG ES promoters in procyclic stage cells.

### VEX1 Focal Accumulation Is Life Cycle Stage Dependent

As we have shown VEX1 is required for efficient silencing of expression-site linked VSG genes in insect stage cells, we then asked whether the distribution of VEX1 in the nucleus changed depending on the life cycle stage in the tsetse fly, and specifically in metacyclic cells where a VSG is expressed. To assess VEX1 localisation in metacyclic cells, we used the inducible RBP6 expression system (Kolev et al., 2012). In this system, metacyclic cells are produced spontaneously rather than in a temporal order (Kolev et al., 2012; Ramey-Butler et al., 2015), allowing us to capture both early stage and mature metacyclic cells. We found, as previously shown, that induction of RBP6 stimulates metacyclogenesis *in vitro* and leads to the expression of mVSGs and the production of metacyclic cells in culture (Figure 2A). We assessed the expression of 5 mVSG genes and RBP6 by RT-qPCR and found a significant increase in mVSG653, mVSG1954, mVSG531 and mVSG639 expression over day 2 to day 4 (Figure 2A; Table 2). We then scored the number of metacyclic cells in culture using a pan-VSG antibody (anti-CRD). Between day 5 and 7 up to 25% of the culture were metacyclic cells (Figure 2B). Following *in vitro* differentiation to insect stage cells, the active BES relocates to the nuclear periphery (Landeira and Navarro 2007), while the VEX complex is redistributed from one—two nuclear foci to a multifocal distribution that appears to be concomitant with all telomeres (Glover et al., 2016). This differentiation step is also associated with a substantial increase in protein abundance VEX2 (Faria et al., 2019). A similar pattern was observed in the uninduced procyclic stage cells (Figure 3A), where VEX1 forms a multi-focal pattern in the nucleus. Following 5 days of RBP6 overexpression, metacyclic cells were identified in culture based on cell morphology and the position of the nucleus and kinetoplast. We found that in these metacyclic cells there were significantly more VEX1 forming one to two foci (Figures 3A,B; Procyclic cells 95% formed multiple foci versus 68% of metacyclic cells with 1-2 foci), suggesting that the pattern of VEX1 accumulation in the nucleus is life cycle stage specific. We then wanted to confirm these findings *in vivo*. Tsetse flies (*Glossina morsitans morsitans*) were infected with Antat 1.1E bloodstream form cells with natively tagged VEX1. As was



**FIGURE 3 |** VEX1 nuclear distribution is life-cycle specific. Immunofluorescence analysis of VEX1 localization **(A)** Upper panel: RBP6 uninduced culture derived procyclic cells. Lower Panel: Quantification of nuclear VEX1 foci Uninduced and day 5 induced RBP6 cultured procyclic and metacyclic cells ( $n \geq 100$ ). Non dividing cells were counted, morphology and the position of the nucleus and kinetoplast were used to count only procyclic cells for uninduced. The images correspond to maximal 3D projections of 0.24 μm stacks; scale bars 23 or 5 μm. Error bar, SD. Significance calculated using student t test (\*,  $p = 0.04$ ) **(B)** RBP6 induced culture derived metacyclic cells with either one or two foci **(C)** tsetse fly midgut derived late procyclic/meosocyclic cells **(D)** tsetse fly salivary gland derived metacyclic cells with either one or two VEX1 foci **(E)** Quantification of nuclear VEX1 foci in tsetse fly derived procyclic and metacyclic cells ( $n \geq 100$ ). The images correspond to maximal 3D projections of 0.24 μm stacks; scale bars 23 or 5 μm. Error bar, SD. Counts from 12 dissected tsetse flies. N, nucleus; K, kinetoplast. Significance calculated using student t test (\*,  $p = 0.02$ ).



**FIGURE 4** | VEX1 localization in Tsetse fly derived cells. Immunofluorescence analysis of tsetse fly derived trypanosome cells. The images correspond to maximal 3D projections of 0.24  $\mu\text{m}$  stacks; scale bars 5  $\mu\text{m}$ . N, nucleus; K, kinetoplast.

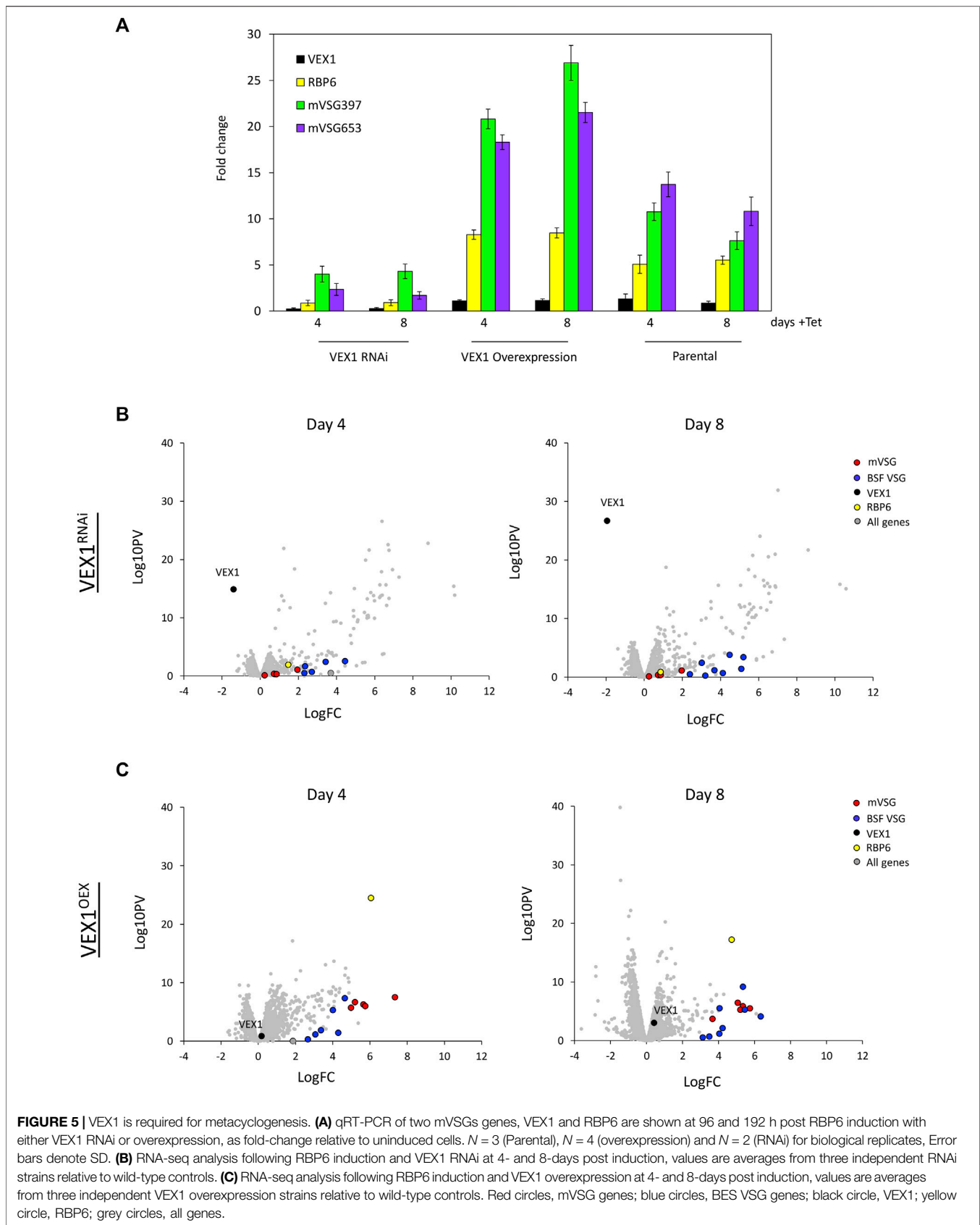
seen in the *in vitro* RBP6 system, VEX1 formed a multi focal pattern in midgut trypomastigote cells (procyclic and mesocyclic forms, **Figures 3C,E; Supplementary Figure S2**). Approximately 60% of metacyclic cells had one to two foci (**Figures 3D,E; Supplementary Figure S3**), and in approximately 40% we could detect 3 or more VEX1 foci, these may be early metacyclic where monoallelic *mVSG* expression is not yet established (Hutchinson et al., 2021). We also examined additional life cycle stages found throughout in the tsetse fly and found that VEX1 was distributed across the nucleus in a multi-focal pattern. Therefore, in metacyclic cells one to two VEX1 foci per nucleus was the dominated pattern (**Figure 4; Supplementary Figure S3**). We, therefore, see a similar

pattern of VEX1 distribution both *in vitro* and *in vivo*. The VEX1 localisation we see in metacyclic is reminiscent of that seen in bloodstream form cells (Glover et al., 2016; Faria et al., 2019; Faria et al., 2021) where the VEX complex is required for singular *VSG* expression.

### VEX1 Modulates *mVSG* Expression During Metacyclogenesis

The transition from epimastigote to metacyclic cells in the tsetse fly salivary gland results in the activation of MESS promoters and expression of *mVSG* genes (Graham and Barry 1995; Sharma et al., 2009). To determine the role of VEX1 in metacyclogenesis we established a doubly inducible TET ON system to





**TABLE 3** | RT-qPCR values for RBP6 and mVSG expression Fold change and SD values for N = 3 for biological replicates.

	VEX1		RBP6		mVSG397		mVSG653	
	AVE	SD	AVE	SD	AVE	SD	AVE	SD
Parental D4	0.86	1.21	5.52	4.44	7.64	6.30	10.81	10.00
Parental D8	1.32	0.38	5.07	2.07	10.76	5.25	13.73	10.18
VEX1 overexpression D4	1.15	0.37	8.47	4.15	26.89	13.78	21.52	9.68
VEX1 overexpression D8	1.09	0.48	8.29	4.47	20.82	27.83	18.30	14.28
VEX1 RNAi D4	0.28	0.09	0.91	0.54	4.30	3.45	1.70	1.95
VEX1 RNAi D8	0.22	0.30	0.87	0.60	4.00	3.25	2.35	1.04

simultaneously modulate *VEX1* expression levels and induce metacyclogenesis via *RBP6* overexpression (**Supplementary Figure S1C**). Initially we assessed the expression of two *mVSG* genes by RT-qPCR (**Figure 5A**; **Table 3**) at 4- and 8-days post induction along with *RBP6* and *VEX1*. In the parental cell line, with *RBP6* overexpression only, we see a 5-fold increase in *RBP6* expression and a 7–13 -fold increase in *mVSG397* and *mVSG653* expression (**Figure 5A**; **Table 3**). Strikingly, a reduction in *VEX1* resulted in poor *mVSG* expression as compared to the parental cell line (*RBP6* overexpression only), and surprisingly we also see a 6-fold reduction in *RBP6* expression (**Figure 5A**; **Table 3**). Conversely, when *VEX1* and *RBP6* are overexpressed, we see an 8-fold increase in *RBP6* expression and between 18–26-fold increase in *mVSG397* and *mVSG653* expression (**Figure 5A**; **Table 3**). We noted that the increase in *VEX1* overexpression cell line was subtle, but even in this context has a dramatic effect on both *mVSG* and *RBP6* expression levels (**Figure 5A**; **Table 3**).

### VEX1 Positively Regulates RBP6 and mVSG Expression During Metacyclogenesis

To further investigate the role of *VEX1* in metacyclogenesis we wanted to see what global changes were associated with *VEX1* RNAi or overexpression during metacyclogenesis. For this we performed transcriptomics analyses at day 4 and 8. Our analysis revealed that during metacyclogenesis, *VEX1* RNAi resulted in reduced *mVSG* expression and a concomitant reduction in the level of *RBP6* (**Figure 5B**) as we saw with the RT-qPCR (**Figure 5A**). This suggests that initiation of *mVSG* expression is crucial to this life cycle differentiating process. Using a pan-VSG antibody, we found that in cultures at day 4 and 8 cultures, no VSG expressing cells were seen (data not shown). We noted that there was a cohort of genes whose transcripts increased in abundance at day 4 and 8 (**Figure 5B**) following *RBP6* overexpression and *VEX1* RNAi. Analysis revealed a specific increase in abundance of silent BES linked genes, normally only expressed from the single active BES in bloodstream form cells (**Supplementary Figure S4A**). Differentiation from bloodstream form to procyclic form trypanosomes results in cessation of transcription of the *ESAG* and VSG genes from the active BES, however multiple BES promoters remain active a relatively equivalent level (Pays et al., 1989; Zomerdijk et al., 1990; Rudenko et al., 1994). Although BES promoters are active in procyclic cells, transcription terminates before the *ESAG7* gene (Zomerdijk et al., 1990; Rudenko et al., 1994). We then looked

with more detail at which BES-linked genes were significantly upregulated, we found that *ESAG 10*, 7, and 6 were 4 to 6-fold upregulated at all BESs, but this level of upregulation was not sustained (**Supplementary Figure S4C**, **S5**) suggesting transcription does not proceed across the whole BES. In fact, the BES-linked VSG genes were only on average 2-fold upregulated. We then looked at other Pol I transcribed genes and found that the procyclin associated genes *PAG1*, *PAG4*, and *PAG5* (Tb927.10.10240, Tb927.10.10210, Tb927.10.10230 respectively) showed an increase in transcript abundance (by 4, 6, 5-fold respectively on day 4) but not *EP1*, *EP2* *Procyclin* or *GPEET* (**Supplementary Figure S4A**). This suggests that during metacyclogenesis, *VEX1* is necessary for expression of *mVSG* genes and maintain silencing of BESs, but additional factors are required for full BES silencing in this life cycle stage. In contrast, following *VEX1* overexpression we see an increase in expression of *mVSG* genes and only moderate increase in BES linked VSG genes (**Figure 5C**), suggesting that *VEX1* promotes *mVSG* expression. Unlike with *VEX1* RNAi, we see a more restrained increase in transcript abundance of BES linked genes and the procyclin and procyclin associated genes (**Supplementary Figure S4B**). Across the BES, *ESAG 10*, 7, and 6 again showed the highest fold change, but by only 2-fold change on average, which was significantly lower than in the *VEX1* RNAi (**Supplementary Figure S4D**, **S5**). Strikingly though, *mVSG* genes show an average of 6-fold increase in transcript abundance (**Supplementary Figure S4D**), revealing a positive role for *VEX1* in *mVSG* transcription in metacyclic cells.

## DISCUSSION

The developmental transitions that trypanosomes undergo as they cycle between the tsetse fly and mammalian host are coupled to dramatic changes in gene expression and especially in surface antigen expression. Central to trypanosomes survival in the mammalian host is the expression of a unique VSG gene that forms a dense protective barrier on the surface of the cell—both in the initial stages of the infection and once established. Our understanding of what leads to mammalian infectivity in African trypanosomes has long been limited by our ability to study key developmental transitions, however, this has changed with the establishment of the *RBP6* overexpression system (Kolev et al., 2012), and more recently with single-cell RNA sequencing of tsetse fly derived trypanosomes (Vigneron et al., 2020; Hutchinson

et al., 2021). How these *VSG* genes are switched on in the tsetse fly salivary gland and the factors that control this process have remained unknown. Here we describe the role of VEX1 in the initiation of *mVSG* expression in metacyclic trypanosomes.

Localisation of VEX1 in the bloodstream form cells is intrinsically linked to function, with VEX1 accumulating at the single active BES and the spliced leader locus (Glover et al., 2016; Faria et al., 2019). The VEX complex undergoes a dramatic relocalisation from bloodstream form to insect stage procyclic form, potentially making the VEX complex available when *VSG* expression is reinitiated. Defining the localisation of VEX1 before and after metacyclogenesis is therefore key. Our *in vitro* and *in vivo* data show a relocalisation of VEX1 in insect stage cells associated with the expression of *mVSG* genes, suggesting that the VEX-complex may act similarly to that in bloodstream forms cells defining the single active MES. The variation in the number of VEX1 foci in metacyclic cells may represent the transition to single *mVSG* expression, but this remains to be shown.

Regulation of monoallelic *VSG* expression is lost when slender bloodstream forms differentiate into G<sub>1</sub> arrested stumpy forms in the mammalian host (Amiguet-Vercher et al., 2004) and the *VSG* coat is shed in the tsetse fly (Roditi et al., 1989). In the procyclic form cell, *VSG* promoters are silenced and repositioned to the nuclear periphery (Navarro et al., 1999; Landeira and Navarro 2007). Depletion of the VEX complex in bloodstream form cells leads to loss of monoallelic *VSG* regulation (Glover et al., 2016; Faria et al., 2019) and here, we show that loss of VEX1 results in *VSG* expression in insect stage procyclic cells (Figures 1A,C). We observed a stronger effect on the MES which may be due to the difference in size between the BES and MES, with the former being up to 60 kb in length and the latter only 5 kb, and the processivity of the polymerase across these loci.

How *VSG* transcription, in either metacyclic or bloodstream form cells, is initiated is unclear. From their single cell RNA-sequencing of tsetse fly derived cells Hutchinson et al., 2021 proposed a “race model” for the initiation and establishment of monoallelic *mVSG* expression. They show a two-step process governs the establishment of monoallelic expression; 1) transcription is initiated at multiple MESs but 2) a single MES eventually dominates (Hutchinson et al., 2021). This process invokes the recruitment of VEX1 and VEX2 by the MESs which in turn triggers a positive feedback loop to recruit the splicing machinery, defines the ESB and drives transcription. The MES with the highest transcription level, recruiting most of the VEX-complex, and thereby depriving the other MESs, would outcompete the others and be established as the single active MES (Glover et al., 2016; Faria et al., 2021; Hutchinson et al., 2021). Several studies in bloodstream form cells also suggest a transcriptional race for establishment of monoallelic *VSG* expression. Firstly, although transcription is initiated at all BESs, it is only elongated over one (Kassem et al., 2014); secondly, during a forced transcriptional switch, where the active BES is switched off and a silent BES is activated, transcription transiently increased across multiple silent BESs in a suggested “probing” of silent *VSG*-ES before the cell fully activates one BES and undergoes a switching event (Aresta-Branco et al., 2016). Our data revealed that VEX1 influences initiation of *mVSG* expression. Where VEX1 is depleted,

and *mVSG* expression is low, and metacyclogenesis fails (Figure 5). In fact, by increasing the abundance of VEX1, and presumably the VEX-complex during metacyclogenesis, not only does *mVSG* transcript abundance increase but so too does *RBP6* expression through a positive feedback loop (Figure 5).

In summary, we have shown that focal accumulation of VEX1 in the metacyclic trypanosome nucleus is dependent on the expression of a *VSG*, presumably defining the single active MES. Our findings have revealed that VEX1 modulates metacyclogenesis and that this life cycle differentiation step is dependent on the cell ability to initiate expression of *mVSG* genes.

## DATA AVAILABILITY STATEMENT

The datasets for VEX1 RNAi or overexpression in procyclic cells and VEX1 RNAi or overexpression in parallel with *RBP6* overexpression in procyclic cells for this study can be found on the ENA PRJEB49957 (ERP134502).

## AUTHOR CONTRIBUTIONS

ET, KR-P, BR, and LG conceived and designed the experiments. ET, AD-H, and KR-P performed the experiments. ET, KR-P, and LG analyzed the data. ET, KR-P, BR, and LG contributed reagents, materials and analysis tools. KR-P and LG wrote the paper. KR-P, ET, BR, and LG edited the paper.

## FUNDING

This project has received funding from the Institut Pasteur to LG. ET was supported by funding from the French Government Agence Nationale de la Recherche, ANR-17-CE12-0012 to LG. KR-P is funded by the French Government’s Investissement d’Avenir program Laboratoire d’Excellence Integrative Biology of Emerging Infectious Diseases (grant ANR-10-LABX-62-IBEID).

## ACKNOWLEDGMENTS

We would like to thank Sebastian Hutchinson for help with the RNA-seq analysis.

## SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fcell.2022.851475/full#supplementary-material>

**Supplementary Figure 1** | Cell line set up. Protein and immunofluorescence analysis of VEX1<sup>tryp</sup> (A) RNAi and (B) overexpression. (C) Protein analysis of *RBP6* overexpression and in conjunction with VEX1 overexpression or VEX1 RNAi.

**Supplementary Figure 2 |** VEX1 localization in tsetse fly salivary gland-derived metacyclic cells. Immunofluorescence analysis of tsetse fly midgut-derived late procyclic / mesocyclic cells. The images correspond to maximal 3D projections of 0.24  $\mu$ m stacks; scale bars 5  $\mu$ m. N, nucleus; K, kinetoplast.

**Supplementary Figure 3 |** VEX1 localization in Tsetse fly derived metacyclic cells. Immunofluorescence analysis of tsetse fly salivary gland derived late metacyclic cells. The images correspond to maximal 3D projections of 0.24  $\mu$ m stacks; scale bars 5  $\mu$ m. N, nucleus; K, kinetoplast.

**Supplementary Figure 4 |** VEX1 RNAi silencing and overexpression results in upregulation of BES linked genes during metacyclogenesis. **(A)** RNA-seq analysis following RBP6 induction and VEX1 RNAi at 4- and 8-days post induction, values are averages from three independent RNAi strains relative to wild-type controls. **(B)** RNA-seq analysis following RBP6 induction and VEX1 overexpression at 4 and 8 days, values are averages from three independent VEX1 overexpression strains relative to wild-type controls. Blue circles, BES linked genes, Black circle, VEX1; Yellow circle, RBP6; Pink circle, Procyclin and Procyclin associated genes; Grey

circles, all genes. **(C)** Upper panel: Schematic depicting a generic BES. Lower panel: Plot depicting the average fold change of BES linked genes following RBP6 overexpression and VEX1 RNAi. Error bar, SD. **(D)** Upper panel: Schematic depicting a generic BES. Lower panel: Plot depicting the average fold change of BES linked genes following RBP6 overexpression and VEX1 overexpression. Error bar, SD. Values are averages of three independent controls relative to wild-type control.

**Supplementary Figure 5 |** VEX1 RNAi and overexpression modulate the level of *ESAG7* and *ESAG6* during metacyclogenesis. **(A)** Plot depicting the average of *ESAG 10, 7, and 6* between VEX1 overexpression and RNAi on day 4 **(B)** Plot depicting the average of *ESAG 10, 7, and 6* between VEX1 overexpression and RNAi on day 8 **(C)** Plot depicting the fold change of each BES linked *ESAG6* following RBP6 overexpression and VEX1 overexpression or RNAi. **(D)** Plot depicting the fold change of each BES linked *ESAG7* following RBP6 overexpression and VEX1 overexpression or RNAi. Error bar, SD. Significance was calculated using an unpaired t test; (\*, P 0.03; \*\*\*, P, 0.0001; \*\*\*\*, P, < 0.0001).

## REFERENCES

- Acosta-Serrano, A., Vassella, E., Liniger, M., Renggli, C. K., Brun, R., Roditi, I., et al. (2001). The Surface Coat of Procyclic *Trypanosoma Brucei*: Programmed Expression and Proteolytic Cleavage of Procyclin in the Tsetse Fly. *Proc. Natl. Acad. Sci.* 98, 1513–1518. doi:10.1073/pnas.98.4.1513
- Alarcon, C. M., Son, H. J., Hall, T., and Donelson, J. E. (1994). A Monocistronic Transcript for a Trypanosome Variant Surface Glycoprotein. *Mol. Cell Biol.* 14, 5579–5591. doi:10.1128/mcb.14.8.5579
- Alsford, S., duBois, K., Horn, D., and Field, M. C. (2012). Epigenetic Mechanisms, Nuclear Architecture and the Control of Gene Expression in Trypanosomes. *Expert Rev. Mol. Med.* 14, e13. doi:10.1017/erm.2012.7
- Alsford, S., Kawahara, T., Glover, L., and Horn, D. (2005). Tagging a T. Brucei RRNA Locus Improves Stable Transfection Efficiency and Circumvents Inducible Expression Position Effects. *Mol. Biochem. Parasitol.* 144, 142–148. doi:10.1016/j.molbiopara.2005.08.009
- Amiguet-Vercher, A., Pérez-Morga, D., Pays, A., Poelvoorde, P., Van Xong, H., Tebabi, P., et al. (2004). Loss of the Mono-Allelic Control of the VSG Expression Sites during the Development of *Trypanosoma Brucei* in the Bloodstream. *Mol. Microbiol.* 51, 1577–1588. doi:10.1111/j.1365-2958.2003.03937.x
- Aresta-Branco, F., Pimenta, S., and Figueiredo, L. M. (2016). A Transcription-independent Epigenetic Mechanism Is Associated with Antigenic Switching in *Trypanosoma Brucei*. *Nucleic Acids Res.* 44, 3131–3146. doi:10.1093/nar/gkv1459
- Barry, J., Graham, S. V., Fotheringham, M., Graham, V. S., Kobryn, K., and Wymer, B. (1998). VSG Gene Control and Infectivity Strategy of Metacyclic Stage *Trypanosoma Brucei*. *Mol. Biochem. Parasitol.* 91, 93–105. doi:10.1016/s0166-6851(97)00193-x
- Caljon, G., Van Reet, N., De Trez, C., Vermeersch, M., Pérez-Morga, D., and Van Den Abbeele, J. (2016). The Dermis as a Delivery Site of *Trypanosoma Brucei* for Tsetse Flies. *Plos Pathog.* 12, e1005744. doi:10.1371/journal.ppat.1005744
- Capewell, P., Cren-Travaille, C., Marchesi, F., Johnston, P., Clucas, C., Benson, R. A., et al. (2016). The Skin Is a Significant but Overlooked Anatomical Reservoir for Vector-Borne African Trypanosomes. *Elife* 5, e17716. doi:10.7554/elifelife.17716
- Carver, T., Harris, S. R., Berriman, M., Parkhill, J., and McQuillan, J. A. (2012). Artemis: an Integrated Platform for Visualization and Analysis of High-Throughput Sequence-Based Experimental Data. *Bioinformatics* 28, 464–469. doi:10.1093/bioinformatics/btr703
- Cestari, I., and Stuart, K. (2015). Inositol Phosphate Pathway Controls Transcription of Telomeric Expression Sites in Trypanosomes. *Proc. Natl. Acad. Sci. USA* 112, E2803–E2812. doi:10.1073/pnas.1501206112
- Cross, G. A. M., Kim, H.-S., and Wickstead, B. (2014). Capturing the Variant Surface Glycoprotein Repertoire (The VSGnome) of *Trypanosoma Brucei* Lister 427. *Mol. Biochem. Parasitol.* 195, 59–73. doi:10.1016/j.molbiopara.2014.06.004
- Davies, C., Ooi, C.-P., Sioutas, G., Hall, B. S., Sidhu, H., Butter, F., et al. (2021). TbSAP Is a Novel Chromatin Protein Repressing Metacyclic Variant Surface Glycoprotein Expression Sites in Bloodstream Form *Trypanosoma Brucei*. *Nucleic Acids Res.* 49, 3242–3262. doi:10.1093/nar/gkab109
- Dolezelova, E., Kunzova, M., Dejung, M., Levin, M., Panicucci, B., Regnault, C., et al. (2020). Cell-based and Multi-Omics Profiling Reveals Dynamic Metabolic Repurposing of Mitochondria to Drive Developmental Progression of *Trypanosoma Brucei*. *Plos Biol.* 18, e3000741.
- Faria, J., Glover, L., Hutchinson, S., Boehm, C., Field, M. C., and Horn, D. (2019). Monoallelic Expression and Epigenetic Inheritance Sustained by a *Trypanosoma Brucei* Variant Surface Glycoprotein Exclusion Complex. *Nat. Commun.* 10, 3023. doi:10.1038/s41467-019-10823-8
- Faria, J., Luzak, V., Muller, L. S. M., Brink, B. G., Hutchinson, S., Glover, L., et al. (2021). Spatial Integration of Transcription and Splicing in a Dedicated Compartment Sustains Monogenic Antigen Expression in African Trypanosomes. *Nat. Microbiol.* 6 (3), 289–300. doi:10.1038/s41564-020-00833-4
- Figueiredo, L. M., and Cross, G. A. M. (2010). Nucleosomes Are Depleted at the VSG Expression Site Transcribed by RNA Polymerase I in African Trypanosomes. *Eukaryot. Cel* 9, 148–154. doi:10.1128/ec.00282-09
- Ginger, M. L., Blundell, P. A., Lewis, A. M., Browitt, A., Günzl, A., and Barry, J. D. (2002). *Ex Vivo* and *In Vitro* Identification of a Consensus Promoter for VSG Genes Expressed by Metacyclic-Stage Trypanosomes in the Tsetse Fly. *Eukaryot. Cel* 1, 1000–1009. doi:10.1128/ec.1.6.1000-1009.2002
- Glover, L., Alsford, S., and Horn, D. (2013). DNA Break Site at Fragile Subtelomeres Determines Probability and Mechanism of Antigenic Variation in African Trypanosomes. *Plos Pathog.* 9, e1003260. doi:10.1371/journal.ppat.1003260
- Glover, L., Hutchinson, S., Alsford, S., and Horn, D. (2016). VEX1 Controls the Allelic Exclusion Required for Antigenic Variation in Trypanosomes. *Proc. Natl. Acad. Sci. USA* 113, 7225–7230. doi:10.1073/pnas.1600344113
- Graham, S. V., and Barry, J. D. (1995). Transcriptional Regulation of Metacyclic Variant Surface Glycoprotein Gene Expression during the Life Cycle of *Trypanosoma Brucei*. *Mol. Cell Biol* 15, 5945–5956. doi:10.1128/mcb.15.11.5945
- Hertz-Fowler, C., Figueiredo, L. M., Quail, M. A., Becker, M., Jackson, A., Bason, N., et al. (2008). Telomeric Expression Sites Are Highly Conserved in *Trypanosoma Brucei*. *Plos one* 3, e3527. doi:10.1371/journal.pone.0003527
- Hoare, C. A. (1971). Rationalization of the Terminology for the Developmental Stages of Trypanosomatid Flagellates. *Med. Parasitol (Mosk)* 40, 307–309.
- Hutchinson, S., Foulon, S., Crouzols, A., Menafra, R., Rotureau, B., Griffiths, A. D., et al. (2021). The Establishment of Variant Surface Glycoprotein Monoallelic Expression Revealed by Single-Cell RNA-Seq of *Trypanosoma Brucei* in the Tsetse Fly Salivary Glands. *Plos Pathog.* 17, e1009904. doi:10.1371/journal.ppat.1009904
- Kassem, A., Pays, E., and Vanhamme, L. (2014). Transcription Is Initiated on Silent Variant Surface Glycoprotein Expression Sites Despite Monoallelic Expression in *Trypanosoma Brucei*. *Proc. Natl. Acad. Sci.* 111, 8943–8948. doi:10.1073/pnas.1404873111
- Kolev, N. G., Günzl, A., and Tschudi, C. (2017). Metacyclic VSG Expression Site Promoters Are Recognized by the Same General Transcription Factor That Is Required for RNA Polymerase I Transcription of Bloodstream Expression Sites. *Mol. Biochem. Parasitol.* 216, 52–55. doi:10.1016/j.molbiopara.2017.07.002

- Kolev, N. G., Ramey-Butler, K., Cross, G. A. M., Ullu, E., and Tschudi, C. (2012). Developmental Progression to Infectivity in *Trypanosoma Brucei* Triggered by an RNA-Binding Protein. *Science* 338, 1352–1353. doi:10.1126/science.1229641
- Ladeira, D., and Navarro, M. (2007). Nuclear Repositioning of the VSG Promoter during Developmental Silencing in *Trypanosoma Brucei*. *J. Cel. Biol.* 176, 133–139. doi:10.1083/jcb.200607174
- Laxman, S., Riechers, A., Sadilek, M., Schwede, F., and Beavo, J. A. (2006). Hydrolysis Products of cAMP Analogs Cause Transformation of *Trypanosoma brucei* from Slender to Stumpy-Like Forms. *PNAS* 103 (50), 19194–19199.
- Li, H., Handsaker, B., Wysoker, A., Fennell, T., Ruan, J., Homer, N., et al. Subgroup Genome Project Data Processing (2009). The Sequence Alignment/Map Format and SAMtools. *Bioinformatics* 25, 2078–2079. doi:10.1093/bioinformatics/btp352
- MacLeod, E. T., Maudlin, I., Darby, A. C., and Welburn, S. C. (2007). Antioxidants Promote Establishment of *Trypanosoma* Infections in Tsetse. *Parasitology* 134, 827–831. doi:10.1017/s0031182007002247
- Matthews, K. R. (2005). The Developmental Cell Biology of *Trypanosoma Brucei*. *J. Cel. Sci.* 118, 283–290. doi:10.1242/jcs.01649
- Müller, L. S. M., Cosentino, R. O., Förstner, K. U., Guizzetti, J., Wedel, C., Kaplan, N., et al. (2018). Genome Organization and DNA Accessibility Control Antigenic Variation in *Trypanosomes*. *Nature* 563, 121–125. doi:10.1038/s41586-018-0619-8
- Navarro, M., Cross, G. A., and Wirtz, E. (1999). *Trypanosoma Brucei* Variant Surface Glycoprotein Regulation Involves Coupled Activation/inactivation and Chromatin Remodeling of Expression Sites. *EMBO J.* 18, 2265–2272. doi:10.1093/emboj/18.8.2265
- Navarro, M., and Gull, K. (2001). A Pol I Transcriptional Body Associated with VSG Mono-Allelic Expression in *Trypanosoma Brucei*. *Nature* 414, 759–763. doi:10.1038/414759a
- Pays, E., Coquelet, H., Pays, A., Tebabi, P., and Steinert, M. (1989). *Trypanosoma Brucei*: Posttranscriptional Control of the Variable Surface Glycoprotein Gene Expression Site. *Mol. Cel. Biol.* 9, 4018–4021. doi:10.1128/mcb.9.9.4018
- Ramey-Butler, K., Ullu, E., Kolev, N. G., and Tschudi, C. (2015). Synchronous Expression of Individual Metacyclic Variant Surface Glycoprotein Genes in *Trypanosoma Brucei*. *Mol. Biochem. Parasitol.* 200, 1–4. doi:10.1016/j.molbiopara.2015.04.001
- Robinson, M. D., McCarthy, D. J., and Smyth, G. K. (2010). edgeR: A Bioconductor Package for Differential Expression Analysis of Digital Gene Expression Data. *Bioinformatics* 26, 139–140. doi:10.1093/bioinformatics/btp616
- Roditi, I., Schwarz, H., Pearson, T. W., Becroft, R. P., Liu, M. K., Richardson, J. P., et al. (1989). Procyclin Gene Expression and Loss of the Variant Surface Glycoprotein during Differentiation of *Trypanosoma Brucei*. *J. Cel. Biol.* 108, 737–746. doi:10.1083/jcb.108.2.737
- Rose, C., Casas-Sánchez, A., Dyer, N. A., Solórzano, C., Beckett, A. J., Middlehurst, B., et al. (2020). *Trypanosoma Brucei* Colonizes the Tsetse Gut via an Immature Peritrophic Matrix in the Proventriculus. *Nat. Microbiol.* 5, 909–916. doi:10.1038/s41564-020-0707-z
- Rotureau, B., Subota, I., Buisson, J., and Bastin, P. (2012). A New Asymmetric Division Contributes to the Continuous Production of Infective *Trypanosomes* in the Tsetse Fly. *Development* 139, 1842–1850. doi:10.1242/dev.072611
- Rotureau, B., and Van Den Abbeele, J. (2013). Through the Dark Continent: African *Trypanosome* Development in the Tsetse Fly. *Front. Cel. Infect. Microbiol.* 3, 53. doi:10.3389/fcimb.2013.00053
- Rudenko, G., Blundell, P. A., Taylor, M. C., Kieft, R., and Borst, P. (1994). VSG Gene Expression Site Control in Insect Form *Trypanosoma Brucei*. *EMBO J.* 13, 5470–5482. doi:10.1002/j.1460-2075.1994.tb06882.x
- Schindelin, J., Arganda-Carreras, I., Frise, E., Kaynig, V., Longair, M., Pietzsch, T., et al. (2012). Fiji: An Open-Source Platform for Biological-Image Analysis. *Nat. Methods* 9 (7), 672–682. doi:10.1038/nmeth.2019
- Sharma, R., Gluenz, E., Peacock, L., Gibson, W., Gull, K., and Carrington, M. (2009). The Heart of Darkness: Growth and Form of *Trypanosoma Brucei* in the Tsetse Fly. *Trends Parasitol.* 25, 517–524. doi:10.1016/j.pt.2009.08.001
- Sharma, R., Peacock, L., Gluenz, E., Gull, K., Gibson, W., and Carrington, M. (2008). Asymmetric Cell Division as a Route to Reduction in Cell Length and Change in Cell Morphology in *Trypanosomes*. *Protist* 159, 137–151. doi:10.1016/j.protis.2007.07.004
- Stanne, T. M., and Rudenko, G. (2010). Active VSG Expression Sites in *Trypanosoma Brucei* Are Depleted of Nucleosomes. *Eukaryot. Cel* 9, 136–147. doi:10.1128/ec.00281-09
- Stijlemans, B., Caljon, G., Van Den Abbeele, J., Van Ginderachter, J. A., Magez, S., and De Trez, C. (2016). Immune Evasion Strategies of *Trypanosoma Brucei* within the Mammalian Host: Progression to Pathogenicity. *Front. Immunol.* 7, 233. doi:10.3389/fimmu.2016.00233
- Tetley, L., Turner, C. M., Barry, J. D., Crowe, J. S., and Vickerman, K. (1987). Onset of Expression of the Variant Surface Glycoproteins of *Trypanosoma Brucei* in the Tsetse Fly Studied Using Immunoelectron Microscopy. *J. Cel Sci* 87 (Pt 2), 363–372. doi:10.1242/jcs.87.2.363
- Toh, J. Y., Nkouawa, A., Sánchez, S. R., Shi, H., Kolev, N. G., and Tschudi, C. (2021). Identification of Positive and Negative Regulators in the Stepwise Developmental Progression towards Infectivity in *Trypanosoma Brucei*. *Sci. Rep.* 11, 5755. doi:10.1038/s41598-021-85225-2
- Trenaman, A., Glover, L., Hutchinson, S., and Horn, D. (2019). A post-transcriptional Respiratome Regulator in *Trypanosomes*. *Nucleic Acids Res.* 47, 7063–7077. doi:10.1093/nar/gkz455
- Trindade, S., Rijo-Ferreira, F., Carvalho, T., Pinto-Neves, D., Guegan, F., Aresta-Branco, F., et al. (2016). *Trypanosoma Brucei* Parasites Occupy and Functionally Adapt to the Adipose Tissue in Mice. *Cell Host & Microbe* 19, 837–848. doi:10.1016/j.chom.2016.05.002
- Van Den Abbeele, J., Claes, Y., van Bockstaele, D., Le Ray, D., and Coosemans, M. (1999). *Trypanosoma Brucei* Spp. Development in the Tsetse Fly: Characterization of the post-mesocyclic Stages in the Foregut and Proboscis. *Parasitology* 118 (Pt 5), 469–478. doi:10.1017/s0031182099004217
- van den Hoff, M. J. B., Moorman, A. F. M., and Lamers, W. H. (1992). Electroporation in 'intracellular' Buffer Increases Cell Survival. *Nucl. Acids Res.* 20, 2902. doi:10.1093/nar/20.11.2902
- Vickerman, K. (1985). 'Developmental Cycles and Biology of Pathogenic *Trypanosomes*. *Br. Med. Bull.* 41, 105–114.
- Vickerman, K. (1969). On the Surface Coat and Flagellar Adhesion in *Trypanosomes*. *J. Cel Sci* 5, 163–193. doi:10.1242/jcs.5.1.163
- Vigneron, A., O'Neill, M. B., Weiss, B. L., Savage, A. F., Campbell, O. C., Kamhawi, S., et al. (2020). Single-cell RNA Sequencing of *Trypanosoma Brucei* from Tsetse Salivary Glands Unveils Metacyclogenesis and Identifies Potential Transmission Blocking Antigens. *Proc. Natl. Acad. Sci. USA* 117, 2613–2621. doi:10.1073/pnas.1914423117
- Woodward, R., and Gull, K. (1990). Timing of Nuclear and Kinetoplast DNA Replication and Early Morphological Events in the Cell Cycle of *Trypanosoma Brucei*. *J. Cel Sci* 95 (Pt 1), 49–57. doi:10.1242/jcs.95.1.49
- Yang, X., Figueiredo, L. M., Espinal, A., Okubo, E., and Li, B. (2009). RAP1 Is Essential for Silencing Telomeric Variant Surface Glycoprotein Genes in *Trypanosoma Brucei*. *Cell* 137, 99–109. doi:10.1016/j.cell.2009.01.037
- Zamze, S. E., Ferguson, M. A. J., Collins, R., Dwek, R. A., and Rademacher, T. W. (1988). Characterization of the Cross-Reacting Determinant (CRD) of the Glycosyl-Phosphatidylinositol Membrane Anchor of *Trypanosoma Brucei* Variant Surface Glycoprotein. *Eur. J. Biochem.* 176, 527–534. doi:10.1111/j.1432-1033.1988.tb14310.x
- Zomerdiijk, J. C., Ouellette, M., ten Asbroek, A. L., Kieft, R., Bommer, A. M., Clayton, C. E., et al. (1990). The Promoter for a Variant Surface Glycoprotein Gene Expression Site in *Trypanosoma Brucei*. *EMBO J.* 9, 2791–2801. doi:10.1002/j.1460-2075.1990.tb07467.x

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**Publisher's Note:** All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Tihon, Rubio-Peña, Dujeancourt-Henry, Crouzols, Rotureau and Glover. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.