Check for updates

OPEN ACCESS

EDITED AND REVIEWED BY Ana Cuenda, Spanish National Research Council (CSIC), Spain

*CORRESPONDENCE Ahmed M. Abdal Dayem, ahmed@konkuk.ac.kr

SPECIALTY SECTION This article was submitted to Signaling, a section of the journal Frontiers in Cell and Developmental Biology

RECEIVED 18 November 2022 ACCEPTED 21 November 2022 PUBLISHED 28 November 2022

CITATION

Abdal Dayem AM, Saadeldin IM and Zhang S (2022), Editorial: Crosslinking ROS signaling and stem cells. *Front. Cell Dev. Biol.* 10:1101802. doi: 10.3389/fcell.2022.1101802

COPYRIGHT

© 2022 Abdal Dayem, Saadeldin and Zhang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or

reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Editorial: Crosslinking ROS signaling and stem cells

Ahmed M. Abdal Dayem¹*, Islam M. Saadeldin^{2,3} and Shengliang Zhang⁴

¹Department of Stem Cell and Regenerative Biotechnology, KU Fusion Science and Technology Institute, Konkuk University, Seoul, South Korea, ²Lab. of Theriogenology, College of Veterinary Medicine, Chungnam National University, Daejeon, South Korea, ³Research Institute of Veterinary Medicine, Chungnam National University, Daejeon, South Korea, ⁴Legorreta Cancer Center, Department of Pathology and Laboratory Medicine, Warren Alpert Medical School, Brown University, Providence, RI, United States

KEYWORDS

reactive oxygen species, pluripotent stem cells, mesenchymal stem cells, signaling molecule, differentiation, flies

Editorial on the Research Topic Crosslinking ROS signaling and stem cells

Stem cells are broadly classified into adult stem cells (ASCs) and pluripotent stem cells (PSCs) which include embryonic stem cells (ESCs) and induced PSCs (iPSCs). ESCs can be obtained from the inner cell mass of the blastocysts and give rise to all the embryonic three germ layers, however, the critical ethical predicament limits ESCs clinical applications. iPSCs possess the same differentiation capacity as ESCs but avoid ESCs usage-associated ethical dilemmas. Therefore, the generation of iPSCs *via* the reprogramming of somatic cells with the transduction of key transcription factors of pluripotency (OCT4, SOX2, KLF4, c-MYC, and Lin-28) represents a breakthrough in the way of clinical application of stem cells in regenerative medicine.

Stem cells inhabit specialized microenvironments called "stem cell niches" that maintain a hypoxic and low level of reactive oxygen species (ROS) environment. The stem cell niche is essential for preserving stem cell stemness and self-renewal potency. ROS has been considered for a long time to be implicated in stress and various disorders and has been redefined as a vital signaling molecule with multifaceted functions in modulating stem cell biology and differentiation (Ogasawara & Zhang, 2009; Nugud, Sandeep, & El-Serafi, 2018).

ROS is an inclusive term that refers to a broad variety of oxidants generated from molecular oxygen. ROS is a family member that belongs to a group of reactive species called reactive nitrogen, sulfur, carbon, selenium, electrophile, and halogen species. They can engage in redox (reduction-oxidation) reactions and form oxidative alteration on biological macromolecules, and ultimately activate key redox signaling and its related biological functions. Two key ROS signaling molecules hydrogen peroxide (H_2O_2) and the superoxide anion radical (O_2^-), are produced primarily by NADPH oxidases and controlled by growth hormones, cytokines, cytokines, mitochondrial electron transport chain. There are more than 40 enzymes involved in this process. H_2O_2 is implicated in a vast majority of redox-associated modulation of biological activities (Reczek & Chandel, 2015).

ROS level not only affects stem cell differentiation capacity but also has a marked impact on stem cell self-renewal and proliferation capacity. Exposure to a low concentration of ROS results in beneficial signaling related to physiological events such as oxygen sensing, whereas a high ROS concentration is detrimental (Finkel, 2011; Tormos et al., 2011). Low ROS level represents an in vivo quiescent milieu for stem cells, which is essential for long-term selfrenewal, proliferation, and differentiation (Jang & Sharkis, 2007). There is a negative correlation between the ROS concentration and the expression level of the pluripotency markers, such as Oct4, Tra 1-60, Nanog, and Sox2, which ultimately augment the endoderm and mesoderm differentiation of ESCs. ESCs showed apoptotic changes when exposed to a high concentration of ROS (>150 µM) (Guo, Chakraborty, Rajan, Wang, & Huang, 2010). On the other hand, ESCs are resistant to a short exposure and a low concentration of ROS compared to the differentiated cells, such as fibroblasts.

The engagement of ROS in modulating the cell cycle has been reported in various multipotent stem cells (Le Belle et al., 2011; Lyublinskaya et al., 2015). In PSCs, the fluctuation in ROS level is associated with cell cycle progression. Ivanova detected the maximal ROS level at the S-phase of the cell cycle (Ivanova et al., 2021). Thus, the application of antioxidants leads to the abrogation of S-phase progression and the consequent disturbance in DNA synthesis and the related apoptotic changes.

ROS is also involved in the stem cell differentiation process. For instance, a high level of H_2O_2 induced by NOX4 is essential for the differentiation of ESCs into cardiac lineage (Xiao et al., 2009). Tormos et al. demonstrated the vital role of mitochondrial complex III in the early surge in the mitochondrial metabolism that leads to ROS generation, which induces the adipogenic differentiation of human mesenchymal stem cells (hMSCs) (Tormos et al., 2011).

The goal of this Research Topic was to untangle the underlying cross-link between ROS signaling and stem cell potency and biological functions. We selected and edited some interesting reviews and original research articles on the Research Topic. These studies demonstrate the impact of ROS levels and the modulation of stem cell characteristics and differentiation in humans and flies.

The cyclic mechanical stress was found to boost the ROS level and induce the expression of nuclear factor erythroid-2-related factor-2 (Nrf2) and its downstream molecules including heme oxygenase 1 (HO1) and NADPH dehydrogenase quinone 1 (NQO1), which ultimately promoted the osteogenic differentiation in human periodontal ligament stem cells (PDLSCs) (Xi et al., 2021). In this regard, Xi et al. performed a protein-protein interaction analysis and proved the interaction between Nrf2 and the phosphoinositide three kinase (PI3K)/ protein kinase B (Akt) pathway. This finding is the main attribute in cyclic mechanical stress-mediated PDLSCs' osteogenic differentiation.

A well-written review paper from Sinenko et al. covers the recent advances in the physiological functions of the intracellular

ROS as a signaling molecule and a secondary messenger of a wide range of cellular signaling in modulating the potency and differentiation of stem cells as well as the development of invertebrate *Drosophila*.

A study by Sheng et al. showed that a novel natural antioxidant compound, Caffeic acid (CA) efficiently delayed the aging events in the *Drosophila* gut, which is associated with the aberrant function of intestinal stem cells (ISCs) and the subsequent dysplasia. This beneficial antioxidant effect is attributed to the potency of CA to suppress oxidative stress-induced JNK signaling.

These studies underscored the importance of Drosophila as a proper genetic model for studying the mitochondrial electron transport chain and mitochondrial ROS-associated biological necessity translating functions and the of various pathophysiological aspects in Drosophila that are enigmatic in humans. While editing this Research Topicr, we hoped that the contributed research work would serve as an inspiration for scientists in the field to investigate further mechanisms and applications of ROS signaling in flies that could be translational for promoting ROS-mediated enhanced stem cell function and clinical applications.

Moreover, Wang et al. demonstrated the beneficial effects of the bone marrow mesenchymal stem cell-derived exosomes on the pathogenesis of osteoarthritis through targeting the competitive endogenous RNA (ceRNA) networks represented in LYRM4-AS1/GRPR/miR-6515-5p signaling pathway.

Author contributions

All authors listed have made a substantial, direct, and intellectual contribution to the work, and approved it for publication.

Funding

Saadeldin IM is supported by projects from the Ministry of Science and ICT through the National Research Foundation of Korea (NRF) (2022R1I1A1A01065412) and the Brain Pool program (grant No: 2021H1D3A2A02040098).

Acknowledgments

We acknowledge our corresponding universities and institutes for the support.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated

References

Finkel, T. (2011). Signal transduction by reactive oxygen species. J. Cell Biol. 194 (1), 7–15. doi:10.1083/jcb.201102095

Guo, Y. L., Chakraborty, S., Rajan, S. S., Wang, R., and Huang, F. (2010). Effects of oxidative stress on mouse embryonic stem cell proliferation, apoptosis, senescence, and self-renewal. *Stem Cells Dev.* 19 (9), 1321–1331. doi:10.1089/scd.2009.0313

Ivanova, J. S., Pugovkina, N. A., Neganova, I. E., Kozhukharova, I. V., Nikolsky, N. N., and Lyublinskaya, O. G. (2021). Cell cycle-coupled changes in the level of reactive oxygen species support the proliferation of human pluripotent stem cells. *STEM CELLS* 39 (12), 1671–1687. doi:10.1002/stem.3450

Jang, Y.-Y., and Sharkis, S. J. (2007). A low level of reactive oxygen species selects for primitive hematopoietic stem cells that may reside in the low-oxygenic niche. *Blood* 110 (8), 3056–3063. doi:10.1182/blood-2007-05-087759

Le Belle, J. E., Orozco, N. M., Paucar, A. A., Saxe, J. P., Mottahedeh, J., Pyle, A. D., et al. (2011). Proliferative neural stem cells have high endogenous ROS levels that regulate self-renewal and neurogenesis in a PI3K/Akt-dependant manner. *Cell stem Cell* 8 (1), 59–71. doi:10.1016/j.stem.2010.11.028

Lyublinskaya, O., Borisov, Y. G., Pugovkina, N., Smirnova, I., Obidina, J. V., Ivanova, J. S., et al. (2015). *Oxidative medicine and cellular longevity*.Reactive oxygen species are required for human mesenchymal stem cells to initiate proliferation after the quiescence exit organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Nugud, A., Sandeep, D., and El-Serafi, A. T. (2018). Two faces of the coin: Minireview for dissecting the role of reactive oxygen species in stem cell potency and lineage commitment. J. Adv. Res. 14, 73–79. doi:10.1016/j.jare.2018.05.012

Ogasawara, M. A., and Zhang, H. (2009). Redox regulation and its emerging roles in stem cells and stem-like cancer cells. *Antioxid. Redox Signal.* 11 (5), 1107–1122. doi:10.1089/ars.2008.2308

Reczek, C. R., and Chandel, N. S. (2015). ROS-dependent signal transduction. Curr. Opin. Cell Biol. 33, 8–13. doi:10.1016/j.ceb.2014.09.010

Tormos, Kathryn V., Anso, E., Hamanaka, Robert B., Eisenbart, J., Joseph, J., Kalyanaraman, B., et al. (2011). Mitochondrial complex III ROS regulate adipocyte differentiation. *Cell Metab.* 14 (4), 537–544. doi:10.1016/j.cmet. 2011.08.007

Xi, X., Zhao, Y., Liu, H., Li, Z., Chen, S., and Liu, D. (2021). Nrf2 activation is involved in osteogenic differentiation of periodontal ligament stem cells under cyclic mechanical stretch. *Exp. Cell Res.* 403 (2), 112598. doi:10.1016/j.yexcr.2021. 112598

Xiao, Q., Luo, Z., Pepe, A. E., Margariti, A., Zeng, L., and Xu, Q. (2009). Embryonic stem cell differentiation into smooth muscle cells is mediated by Nox4-produced H2O2. *Am. J. Physiol. Cell Physiol.* 296 (4), C711–C723. doi:10. 1152/ajpcell.00442.2008