



Role of Actin Cytoskeleton Reorganization in Polarized Secretory Traffic at the Immunological Synapse

Victor Calvo¹ and Manuel Izquierdo^{2*}

¹ Departamento de Bioquímica, Facultad de Medicina, Instituto de Investigaciones Biomédicas Alberto Sols, Consejo Superior de Investigaciones Científicas - Universidad Autónoma de Madrid (CSIC-UAM), Madrid, Spain, ² Instituto de Investigaciones Biomédicas Alberto Sols, Consejo Superior de Investigaciones Científicas - Universidad Autónoma de Madrid, Madrid, Spain

OPEN ACCESS

Edited by:

Pedro Roda-Navarro,
Universidad Complutense de
Madrid, Spain

Reviewed by:

Cosima T. Baldari,
University of Siena, Italy
Enrique Aguado,
University of Cádiz, Spain

*Correspondence:

Manuel Izquierdo
mizquierdo@iib.uam.es

Specialty section:

This article was submitted to
Cell Adhesion and Migration,
a section of the journal
Frontiers in Cell and Developmental
Biology

Received: 13 November 2020

Accepted: 11 January 2021

Published: 04 February 2021

Citation:

Calvo V and Izquierdo M (2021) Role
of Actin Cytoskeleton Reorganization
in Polarized Secretory Traffic at the
Immunological Synapse.
Front. Cell Dev. Biol. 9:629097.
doi: 10.3389/fcell.2021.629097

T cell receptor (TCR) and B cell receptor (BCR) stimulation by antigen presented on an antigen-presenting cell (APC) induces the formation of the immune synapse (IS), the convergence of secretory vesicles from T and B lymphocytes toward the centrosome, and the polarization of the centrosome to the immune synapse. Immune synapse formation is associated with an initial increase in cortical F-actin at the synapse, followed by a decrease in F-actin density at the central region of the immune synapse, which contains the secretory domain. These reversible, actin cytoskeleton reorganization processes occur during lytic granule degranulation in cytotoxic T lymphocytes (CTL) and cytokine-containing vesicle secretion in T-helper (Th) lymphocytes. Recent evidences obtained in T and B lymphocytes forming synapses show that F-actin reorganization also occurs at the centrosomal area. F-actin reduction at the centrosomal area appears to be involved in centrosome polarization. In this review we deal with the biological significance of both cortical and centrosomal area F-actin reorganization and some of the derived biological consequences.

Keywords: T lymphocytes, B lymphocytes, immune synapse, actin cytoskeleton, protein kinase C δ , centrosome, multivesicular bodies, FMNL1

INTRODUCTION

T and B lymphocyte activation by antigen-presenting cells (APC) takes place at a specialized cell to cell interface called the immunological synapse (IS). IS establishment by T and B lymphocytes is a very dynamic, plastic and critical event, acting as a tunable signaling platform that integrates spatial, mechanical and biochemical signals, involved in antigen-specific, cellular and humoral immune responses (Fooksman et al., 2010; de la Roche et al., 2016). The IS is described by the formation of a concentric, bullseye spatial pattern, termed the supramolecular activation complex (SMAC), upon cortical actin reorganization (Billadeau et al., 2007; Griffiths et al., 2010; Yuseff et al., 2013; Kuokkanen et al., 2015). This reorganization yields a central cluster of antigen receptors bound to antigen called central SMAC (cSMAC) and a surrounding adhesion molecule-rich ring, called peripheral SMAC (pSMAC), which appears to be crucial for adhesion with the APC (Monks et al., 1998; Fooksman et al., 2010). Surrounding

the pSMAC, at the edge of the contact area with the APC, is the distal SMAC (dSMAC), which consists of a circular array of dense filamentous actin (F-actin) (Griffiths et al., 2010; Le Flo'ch and Huse, 2015) (**Figure 1A**).

T cell receptor (TCR) and B cell receptor (BCR) stimulation by antigen presented by APC, together with accessory molecules interaction with their ligands on the APC, induces IS formation, convergence of T and B lymphocyte secretory vesicles toward the centrosome and, almost simultaneously, centrosome polarization to the IS (Huse, 2012; de la Roche et al., 2016). This leads to polarized secretion of extracellular vesicles, lytic granules, stimulatory cytokines, or lytic proteases (**Figure 1**). The centrosome is the major microtubule organization center (MTOC) in T and B lymphocytes, and consists of two centrioles surrounded by pericentriolar material (PCM), generating a radial organization of microtubules (Sanchez and Feldman, 2017). IS formation is associated with an initial increase in cortical F-actin at the IS (Billadeau et al., 2007), followed by a decrease in F-actin density at the central region of the IS that includes the cSMAC and contains the secretory domain (Griffiths et al., 2010; Ritter et al., 2015). Subsequently, F-actin recovery at the cSMAC leads to conclusion of lytic granule secretion in cytotoxic T lymphocytes (CTL) (Ritter et al., 2017). These reversible actin cytoskeleton reorganization processes occur during lytic granule secretion in CTL and cytokine-containing vesicle secretion in T-helper (Th) lymphocytes (Na et al., 2015; Ritter et al., 2017).

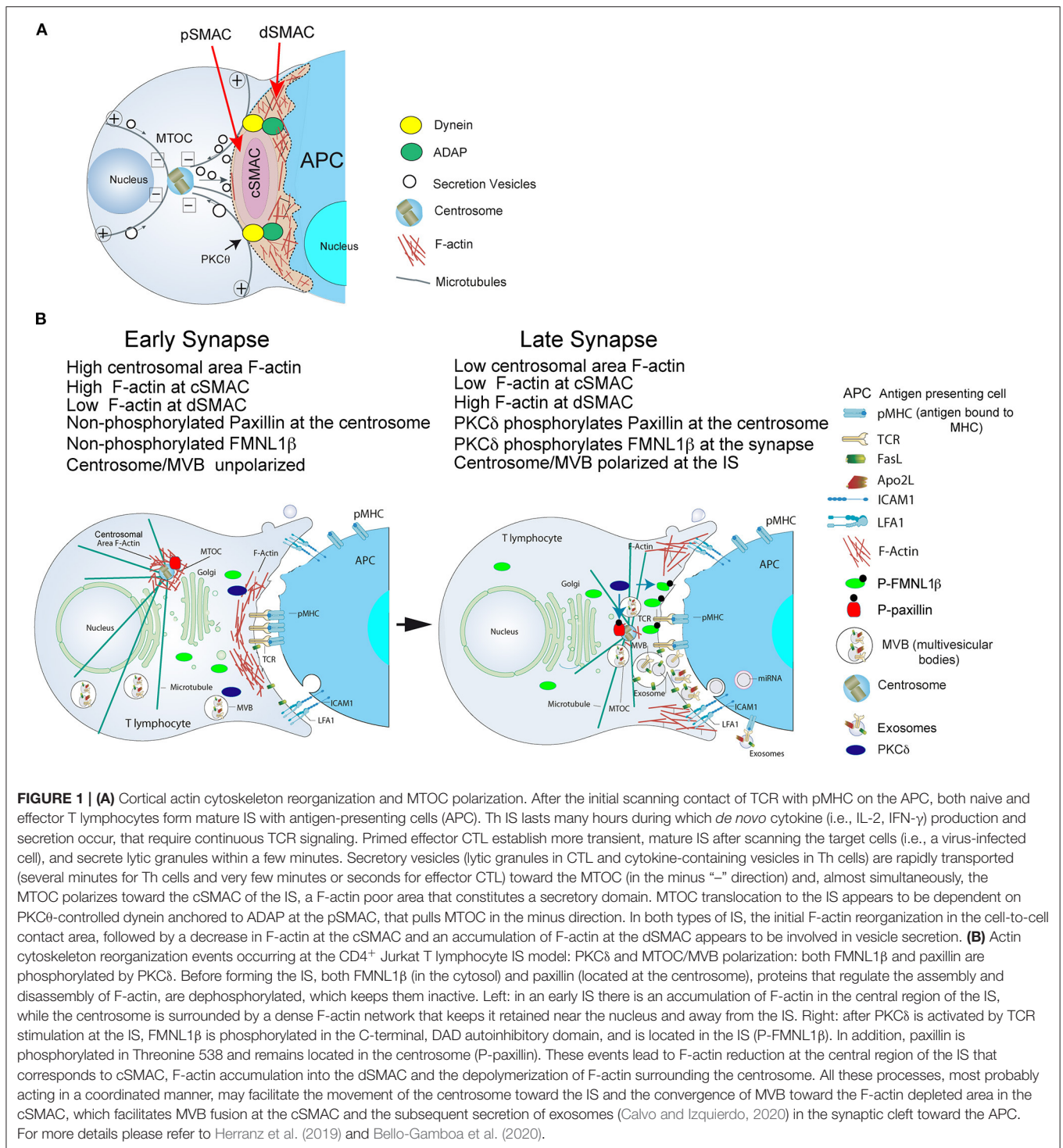
IMMUNE SYNAPSE MORPHOLOGY FORMED BY DIFFERENT IMMUNE CELLS

The IS has long been characterized by the general concentric architecture that adopts during its maturation (Le Flo'ch and Huse, 2015). The bullseye actin cytoskeleton architecture of the IS and the F-actin reorganization process are common to CD4⁺ Th lymphocytes, CD8⁺ CTL, B lymphocytes, and natural killer (NK) cells forming IS (Rak et al., 2011; Lagrue et al., 2013; Le Flo'ch and Huse, 2015). However, for space reasons in this review we deal only with IS made by T and B lymphocytes. The bullseye

pattern of a mature synapse includes redistribution of F-actin and surface receptors in concentric regions. In this context, radially symmetric spreading of the T lymphocyte over the surface of the APC is conducted by protrusive actin polymerization (Le Flo'ch and Huse, 2015; de la Roche et al., 2016), leading to TCR-pMHC interactions at the tip of these actin-rich interdigitations (de la Roche et al., 2016). This is accompanied by the formation of TCR microclusters in the synaptic membrane that, as TCR signaling is initiated, coalesce in the center of the synapse to form the cSMAC (Le Flo'ch and Huse, 2015; de la Roche et al., 2016). Thus, as the IS develops, retrograde (centripetal) F-actin flow drives TCR microclusters into the cSMAC, while F-actin intensively reorganizes into the peripheral ring that will become the dSMAC (Le Flo'ch and Huse, 2015). F-actin flow within the dSMAC promotes adhesion by clustering integrins such as LFA-1 in the pSMAC (Comrie et al., 2015), that eventually will interact with ICAM-1 located on the APC (**Figure 1B**), reinforcing adhesion between these cells and amplifying TCR signaling (Le Flo'ch and Huse, 2015; de la Roche et al., 2016). Remarkably, all the above mentioned immune cells share the capability to directionally secrete stimulatory cytokines, proteases, cytotoxic factors, or extracellular vesicles (including exosomes) at the IS (Calvo and Izquierdo, 2020). This F-actin structure for polarized secretion is thought to enhance the specificity and efficiency of the triggered biological responses (Le Flo'ch and Huse, 2015). Although the IS formed by all these immune cells share this general F-actin pattern, important differences in terms of stability, duration but also synaptic F-actin structure exist among CD4⁺ Th, CD8⁺ CTL, B lymphocytes, and NK cells (Murugesan et al., 2016; Carisey et al., 2018). For instance, whereas CD4⁺ cells form stable IS (from minutes up to several hours), that are necessary for both directional and continuous secretion of stimulatory cytokines (Ueda et al., 2011), IS made by primed CTL trigger the rapid polarization (from seconds to very few minutes) of their lytic granules or secretory lysosomes (SL) toward the IS (Griffiths et al., 2010; Huse, 2012). CTL form very transient IS, lasting only few minutes, until target cells are eliminated. Optimal CTL function is thought to require rapid and transient contacts in order to consecutively deliver as many successive lethal hits as possible to several target cells (Calvo and Izquierdo, 2020). Apart from these kinetic and stability differences, the canonical bullseye actin pattern was described in CD4⁺ T lymphocytes using lipid bilayers or upon interaction with B cells. However, a variation of this canonical pattern was observed when CD4⁺ T lymphocytes were challenged with dendritic cells as APC, resulting in a multifocal cell to cell IS (Brossard et al., 2005; Kumari et al., 2019). Thus, 3D spatial differences between the IS made by the same immune cell type exist. Moreover, under comparable stimulation conditions, differences in actin cytoskeleton spatial organization and dynamics among immortalized human and primary mouse and human T lymphocytes exist (Colin-York et al., 2019; Kumari et al., 2019), and there are differences in the organization and molecular mechanisms underlying these F-actin networks (Kumari et al., 2019) (**Table 1**).

Although IS architecture and dynamics are major determinants of antigen recognition and signaling by TCR and BCR, the molecular components that contribute to the

Abbreviations: 3D-SIM, 3D structured illumination microscopy; ADAP, adhesion and degranulation-adaptor protein; APC, antigen-presenting cell; BCR, B-cell receptor for antigen; C^c , center of mass; cSMAC, central supramolecular activation cluster; CTL, cytotoxic T lymphocytes; DAG, diacylglycerol; DGK α , diacylglycerol kinase α ; DGK ζ , diacylglycerol kinase ζ ; Dial1, Diaphanous-1; dSMAC, distal supramolecular activation cluster; F-actin, filamentous actin; FMNL1, formin-like 1; ILPs; invadosome-like protrusions; IS, immune synapse; ITAM, immunoreceptor tyrosine-based motifs; LAT, linker activation of T cells; LFA-1, lymphocyte function-associated antigen 1; LLSM, lattice light sheet microscopy; MHC, major histocompatibility complex; MVB, multivesicular bodies; MTOC, microtubule-organizing center; NK, natural killer; NPFs, actin nucleation promoting factors; PCM, pericentriolar material; PKC, protein kinase C; PKC θ , protein kinase C θ isoform; PKD, protein kinase D; PLC, phospholipase C; PKC δ , protein kinase C δ isoform; pSMAC, peripheral supramolecular activation cluster; ROI, region of interest; SEE, staphylococcal enterotoxin E; SIM, structured illumination microscopy; SL, secretory lysosomes; SLP76, SH2 domain-containing leukocyte protein of 76 kDa; SMAC, supramolecular activation cluster; STED, stimulated emission depletion; TCR, T-cell receptor for antigen; Th, T-helper; TIRFM, total internal reflection fluorescence microscopy; WASH; Wiskott-Aldrich syndrome protein and SCAR homolog; WASp, Wiskott-Aldrich syndrome protein; ZAP70, Syk-kinase zeta chain-associated protein of 70 kDa.



distinct F-actin patterns observed in IS formed by different immune cells remain largely unknown (Kumari et al., 2019). Indeed, this knowledge is required to understand how different immune cells acquire and develop their functional specialization. It has been speculated that the actin cytoskeleton can arbitrate a force balance across the IS

interface to regulate the dimensions and lifetimes of diverse subsynaptic zones that, in turn, may alter different T cell activation steps (Kumari et al., 2019). Remarkably, several F-actin regulatory proteins are different for each synapse subtype (summarized in Table 1), thus these differences may underlie the spatiotemporal differences in F-actin architecture

TABLE 1 | F-actin reorganization and polarized secretion events in T and B lymphocyte IS and involved proteins.

(*)	T lymphocyte IS		B lymphocyte IS	
	CTL/APC	CD4 ⁺ T cell /APC	B/APC	B /CD4 ⁺ T cell
F-actin reorganization at the IS	+ CDC42/WASP/ARP2/3(Billadeau et al., 2007; Sinai et al., 2010) TAGNL2 (Na et al., 2015)	+ CDC42/WASP/ARP2/3 (Chemin et al., 2012) dynamin 2 (Gomez et al., 2005) CDC42 (Stowers et al., 1995) FMNL1, Dia1 (Murugesan et al., 2016) HS1 (Gomez et al., 2006) TAGNL2 (Na et al., 2015)	+ CDC42/WASP/ARP2/3 Ezrin, Moesin(Kuokkanen et al., 2015)	+ TAGNL2(Na et al., 2016)
F-actin reduction at cSMAC and centrosome polarization	+ (Ritter et al., 2015, 2017) CDC42/IQGAP1 (Stowers et al., 1995; Stinchcombe et al., 2006)	+ PKC δ , FMNL1 β (Herranz et al., 2019; Bello-Gamboa et al., 2020) Dynein(Combs et al., 2006; Liu et al., 2013; Sanchez et al., 2019)	+ dynein, proteasome(Schnyder et al., 2011; Ibanez-Vega et al., 2019)	Unknown
F-actin reduction at centrosome and centrosome polarization	Unknown	+ PKC δ , paxillin (Bello-Gamboa et al., 2020) WASH, ARP2/3(Farina et al., 2016)	+ ARP2/3(Obino et al., 2016) Proteasome(Ibanez-Vega et al., 2019)	Unknown
Lytic granules and/or Exosome secretion	+ (Peters et al., 1989)	+ (Alonso et al., 2011)	+ (Yuseff et al., 2011; Kuokkanen et al., 2015)	+ (Muntasell et al., 2007)
Centrosome polarization	+ (Stinchcombe et al., 2006)	+ (Jeda et al., 2011)	+ (Yuseff et al., 2011)	+ (Duchez et al., 2011)
Mechanisms of centrosome polarization	Paxillin (Herreros et al., 2000; Robertson and Ostergaard, 2011) FMNL1, Dia1 (Gomez et al., 2007) CDC42/IQGAP1 (Stowers et al., 1995; Stinchcombe et al., 2006)	FMNL1, Dia1 (Gomez et al., 2007) PKC θ , dynein (Quann et al., 2009) CDC42 (Stowers et al., 1995) PKC δ , paxillin(Herranz et al., 2019; Bello-Gamboa et al., 2020)	ARP2/3 (Obino et al., 2016) Proteasome(Ibanez-Vega et al., 2019)	Unknown
DAG/DGK-control of centrosome polarization.	+ (Quann et al., 2009)	+ DAG, DGK α (Quann et al., 2009; Alonso et al., 2011) DAG, dynein(Liu et al., 2013) DAG/dynein/PKC (Sanchez et al., 2019)	+ DAG, DGK ζ (Merino-Cortés et al., 2020)	Unknown
PKC/PKD control of secretory granules/MVB traffic	+ PKC δ (Ma et al., 2007) PKC θ (Monks et al., 1998; Quann et al., 2011)	+ PKC δ (Herranz et al., 2019) PKD1/2 (Mazzeo et al., 2016)	+ PKC ζ (Siemasko et al., 1998; Yuseff et al., 2011) PKD1/3 (Mazzeo et al., 2016)	Unknown

(*) The quoted biological response in the first column corresponds to the response of the effector, first cell type for each indicated cell-cell synapse.

existing among different IS. It is out of the scope of this review to detail these differences, please refer to some excellent reviews on this subject, that include also data on F-actin reorganization in the IS made by NK cells (Billadeau et al., 2007; Lagrue et al., 2013; Hammer et al., 2018; Li et al., 2018; Blumenthal and Burkhardt, 2020).

SIGNALS REGULATING CORTICAL ACTIN REORGANIZATION IN THE IMMUNE SYNAPSE

cSMAC, pSMAC, and dSMAC formation characterizes a mature IS and is the basis of a signaling platform that integrates signals

and coordinates molecular interactions leading to both exocytic and endocytic processes, necessary for an appropriate antigen-specific immune response (Griffiths et al., 2010; Xie et al., 2013). Actin reorganization plays a central role in IS maintenance, as well as in antigen receptor-derived signaling (Billadeau et al., 2007). In brief, TCR and BCR stimulation, together with interaction of adhesion and co-stimulatory molecules at the IS, trigger early second messengers such as calcium and diacylglycerol (DAG) (Izquierdo and Cantrell, 1992). DAG regulates several protein kinase C (PKC) family members (including novel PKC members, PKC δ and PKC θ), protein kinase D (PKD1), and Ras/ERK2 pathway (Spitaler et al., 2006), leading to the activation of the two major actin regulatory pathways: the formin (FMNL1 and Dia) pathway involved in

F-actin nucleation, and the CDC42/WASP/ARP2/3 pathway involved in actin filament branching (Kühn and Geyer, 2014; Kumari et al., 2014). In the IS made by T lymphocytes, TCR engagement triggers multiple signaling pathways that regulate actin organization and rearrangements at the different F-actin networks inside and outside the IS (Billadeau et al., 2007; Kumari et al., 2014; Le Floch and Huse, 2015; Hammer et al., 2018). Thus, actin polymerization is a result of complex molecular interactions, and a diverse and complex set of TCR-downstream molecular pathways contributes to actin polymerization. Once the T cell is bound to the pMHC complex on the APC, the immunoreceptor tyrosine-based motifs (ITAM) in the cytosolic tail of the TCR-associated CD3 complex undergo phosphorylation, which then serve as docking sites for the Syk-kinase zeta chain-associated protein of 70 kDa (ZAP70) (Kumari et al., 2014). CD3-recruited ZAP70 then leads to phosphorylation and activation of two key adaptor molecules that associate with a variety of molecules in TCR-associated signaling complexes. First, ZAP70 phosphorylates linker activation of T cells (LAT), which in turn associates via Gads with the SH2 domain-containing leukocyte protein of 76 kDa (SLP76). SLP76 recruitment to phosphorylated LAT allows subsequent SLP76 phosphorylation by ZAP70. Phosphorylated LAT also directly interacts with PLC γ 1, a major regulator of Ca $^{2+}$ influx in response to TCR triggering. PLC γ is recruited via LAT interaction to the synaptic membrane and generates DAG at the IS. SLP76 acts as a scaffold for a multitude of actin effectors including Rho GTPases nucleotide exchange factor, Vav1, adaptor molecule Nck, and actin nucleation promoting factors (NPFs). Alternatively, Rho GTPases (CDC42, Rho, Rac) can also bind and activate actin NPFs (ARP2/3, WAVE), which in turn activate actin nucleation factors (ARP2/3, formins), ultimately leading to F-actin formation. In addition, PI3K activated by LAT/SLP76 activation produces PIP $_3$ at the plasma membrane that, by recruiting DOCK2 to the IS, activates Rac and, ultimately, WAVE/ARP2/3 (Le Floch and Huse, 2015). An intriguing feature of actin cytoskeleton regulation at the IS is the variety of both actin NPFs and actin nucleation factors involved in actin cytoskeleton assembly. This could be related to the complexity of the four F-actin networks contributing to the architecture of the IS described below (Hammer et al., 2018).

More recently, by using super-resolution imaging techniques, such as total internal fluorescence microscopy combined with 3D structured-illumination microscopy (TIRFM/3D-SIM) on functionalized stimulatory surfaces, it has been shown that following initial TCR-antigen interaction, at least four discrete F-actin networks form and maintain the shape and function of this canonical IS (Hammer et al., 2018; Blumenthal and Burkhardt, 2020): branched F-actin network at the dSMAC controlled by WAVE/ARP2/3 activity; actomyosin arc network at the pSMAC controlled by formin Dia1 and phosphorylated myosin; hypodense F-actin at the cSMAC controlled by ARP2/3 and probably formins such as FMNL1/Dia1; F-actin foci at the dSMAC and pSMAC controlled by WASP and HS1 (Hammer et al., 2018; Blumenthal and Burkhardt, 2020). Therefore, in the same cell type distinct regulators control discrete F-actin networks. The four F-actin networks at the T lymphocyte IS

are related to the three distinct functional and signaling areas at the IS (SMACs). Thus, the outer ring of the IS, the dSMAC, corresponds to the lamellipodium region of a migrating cell and forms quickly upon TCR stimulation and contains highly-branched actin filaments generated by the ARP2/3 activator WAVE2. Radially arranged within the dSMAC are bundles of linear actin filaments, generated by formin (Dia) activity near the edge of the spreading lymphocyte, that bend as they move inward the dSMAC, forming actomyosin arcs spanning the pSMAC. Therefore, these actomyosin arcs define the pSMAC, which is enriched in integrins. Actomyosin network disassembly at the pSMAC leads to a F-actin poor region in the center of the IS known as the cSMAC. cSMAC is associated with receptor internalization and provides a site for exocytic vesicle secretion (Figure 1). Finally, the fourth actin network consists of dense actin foci related to protrusive structures rich in F-actin called invadosome-like protrusions (ILPs) (Hammer et al., 2018; Blumenthal and Burkhardt, 2020). Recent evidences support the view that the described F-actin network complexity is, at least in part, based on the action of different F-actin regulatory pathways. For instance, branched F-actin network at the dSMAC is controlled by PI3K-PIP $_3$ -DOCK2-WAVE/ARP2/3, whereas F-actin foci at the cSMAC is regulated by ZAP70-LAT/SLP76-VAV-CDC42-WASP-ARP2/3 (Hammer et al., 2018).

SYNAPTIC ACTIN CYTOSKELETON CONTROL OF CENTROSOME AND SECRETION VESICLES POLARIZATION: RELEASE OF EXTRACELLULAR VESICLES AND LYTIC GRANULES

Regarding the mechanisms controlling centrosome polarization and the role of cortical actin reorganization in MTOC polarization, DAG production at the IS has been shown to be important for centrosome polarization in both CTL and CD4 $^{+}$ T lymphocytes forming IS (Quann et al., 2009). With respect to potential DAG effectors, DAG-activated PKC θ at the IS triggers the adhesion and degranulation-adaptor protein (ADAP)/dynein complex localization at the F-actin/integrin rich, pSMAC ring (Figure 1A). Since dynein is a minus end-directed microtubule motor, after being recruited to the IS it can bind microtubules and reorient the centrosome by minus end-directed motion (Combs et al., 2006; Quann et al., 2009, 2011; Liu et al., 2013) (Figure 1A). At the early stages of IS formation F-actin accumulates at the lymphocyte-APC contact area to generate filopodia and lamellipodia, that produce dynamic changes between extension and contraction in the lymphocyte over the APC surface (Le Floch and Huse, 2015). Subsequently, once IS growth has stabilized, cortical F-actin accumulates into the dSMAC, and F-actin reduction at the cSMAC appears to facilitate secretion toward the APC by focusing secretion vesicles on the IS (Stinchcombe et al., 2006), creating regions within the canonical bullseye IS structure (Figure 1A). Thus, F-actin reduction at the cSMAC does not simply allow secretion, since it apparently plays an active role in centrosome movement to the IS (Stinchcombe et al., 2006; Ritter et al., 2015; Sanchez et al., 2019). It is widely

thought that centrosome reorientation promotes cytotoxic and Th lymphocyte specificity by guiding secretory vesicles to the IS for directional secretion (Stinchcombe et al., 2006; Huse et al., 2008). However, in the context of secretory traffic at the IS, not always centrosome polarization is necessary for either secretory vesicle transport or secretion at the IS in CD4⁺ T lymphocytes (Chemin et al., 2012) or lytic granule polarization and secretion in CTLs (Ma et al., 2007; Bertrand et al., 2013; Nath et al., 2016). For instance, it has been shown that an early and rapid secretion phase of lytic granules constitutively positioned nearby the IS precedes centrosome polarization at the CTL-target cell IS (Bertrand et al., 2013). In addition, in PKC δ -KO mouse CTL, lytic granules did not polarize to the IS and, subsequently, CTL activity and target cell death were inhibited (Ma et al., 2007). However, centrosome polarization toward the IS was not affected by the absence of PKC δ (Ma et al., 2007). In contrast, in the synapses made by CD4⁺ Jurkat T lymphocytes, multivesicular bodies (MVB, a type of secretory vesicles involved in exosome secretion by CD4⁺ Jurkat and primary CD8⁺ and CD4⁺ T lymphocytes (Alonso et al., 2005, 2011)) and centrosome always co-migrated toward the IS, and MVB and centrosome did not polarize in PKC δ -interfered CD4⁺ Jurkat T lymphocytes and the centrosome/MTOC center of mass (MTOC^C) was coincident or very proximal to the MVB center of mass (MVB^C) regardless of polarization (Herranz et al., 2019; Bello-Gamboa et al., 2020). In summary, all these results broaden current views of CTL biology by revealing an extremely rapid lytic granule secretion step and by showing that centrosome polarization is dispensable for efficient lytic granule secretion. All these examples of segregation between centrosome movement and lytic granule polarization point out that centrosome repositioning, secretory vesicles traffic, and F-actin architecture and dynamics at several locations should be analyzed at the single cell level, to obtain a more complete view of the secretion process.

In addition, it has been recently described that centriole-deficient CTL exhibited reduced cytotoxicity due to an alteration in secretory granule biogenesis, although this deficient response was not due to impaired polarized secretion, since lytic granule traffic and secretion toward the IS remained unaffected (Tamzalit et al., 2020). Instead, it has been proposed that the defect was in part due to impaired F-actin reorganization at the IS produced by centriole deletion (Tamzalit et al., 2020), which points out an unexpected role of the intact centrosome in supporting synaptic F-actin architecture and dynamics. Interestingly, CTL lacking centriole formed synapses that lacked an obvious F-actin cleared region at the cSMAC, a similar phenotype to that found in PKC δ -interfered CD4⁺ Jurkat T lymphocytes (Herranz et al., 2019; Bello-Gamboa et al., 2020). Polarized secretion is known to occur in IS domains that have been cleared of F-actin and are, thus, accessible to secretion vesicles (Griffiths et al., 2010; Huse, 2012; Ritter et al., 2015; Herranz et al., 2019).

More recently, it has been shown that PKC δ -dependent F-actin clearing at the cSMAC and PKC δ -dependent phosphorylation of formin FMNL1 β at the IS, are involved in centrosome/MVB polarization leading to exosome secretion in CD4⁺ Jurkat T lymphocytes forming IS (Herranz et al., 2019; Bello-Gamboa et al., 2020). The formins Dia and FMNL1 are

constitutively inactive because they undergo intramolecular, autoinhibitory folding in the cytoplasm, which blocks their ability to nucleate and elongate actin filaments (Hammer et al., 2018). In this context, a different formin, FMNL2, is phosphorylated by PKC α and PKC δ at S1072, reversing its autoinhibition by the C-terminal, DAD auto-inhibitory domain and enhancing F-actin assembly, β 1-integrin endocytosis, and invasive motility (Wang et al., 2015). In the FMNL1 isoform FMNL1 β , S1086 is surrounded by a sequence displaying high homology to the one surrounding S1072 of FMNL2 (Wang et al., 2015; Bello-Gamboa et al., 2020). Our results support that IS-induced, PKC δ -dependent phosphorylation in FMNL1 β C-terminal region containing the auto-inhibitory domain (possibly at S1086) activates FMNL1 β and mediates centrosome polarization. Thus, PKC δ appears to regulate F-actin reorganization, most probably, by controlling FMNL1 β activation through phosphorylation at S1086 (Bello-Gamboa et al., 2020), as certain PKC isoforms activate FMNL2 activity (Wang et al., 2015).

CENTROSOMAL F-ACTIN AND CENTROSOME POLARIZATION

The centrosome nucleates and anchors microtubules and is thus considered to be the principal MTOC. In addition, few years ago it was discovered that the centrosome organizes a local F-actin network and should be considered as a F-actin organizing center (Farina et al., 2016). Isolated centrosomes from Jurkat T lymphocytes efficiently nucleate actin filaments, and the centrosome is associated *in vivo* with an actin network, and these results were extended to other cell types (Farina et al., 2016; Plessner et al., 2019). Moreover, actin filament nucleation at the centrosome is mediated by the nucleation-promoting factor Wiskott-Aldrich syndrome protein (WASP) and SCAR homolog (WASH), in combination with the ARP2/3 complex (Farina et al., 2016). Pericentriolar material seems to modulate F-actin network by regulating WASH/ARP2/3 recruitment to the centrosome (Farina et al., 2016).

Centrosomal F-actin and B Lymphocytes

Centrosome-associated ARP2/3 locally nucleates F-actin, which is needed for centrosome tethering to the nucleus (Obino et al., 2016). Upon B lymphocyte activation with BCR-ligand-coated beads as a synapse model, ARP2/3 is partially depleted from the centrosome, as a result of its recruitment to the IS, where it regulates cortical F-actin. This leads to a reduction in F-actin nucleation at the centrosome and thereby allows its detachment from the nucleus and polarization to the IS (Obino et al., 2016). Thus, centrosomal F-actin depletion appears to be crucial allowing centrosome polarization toward the IS during BCR stimulation in B lymphocytes (Obino et al., 2016). Thus, both *in vitro* and living-cell experiments support this new view of centrosome as a genuine and plastic F-actin-organizing center. However, the precise function of the F-actin network at the centrosome is not well understood. In the same B lymphocyte model, F-actin depletion around the centrosome, F-actin reorganization at

the IS, and centrosome polarization depend on proteasome activity (Ibanez-Vega et al., 2019). By inhibiting proteasome activity, an inhibition of F-actin dismantling around centrosome correlated with the inhibition of centrosome polarization toward the B lymphocyte synapse (Ibanez-Vega et al., 2019). Thus, it appears that at least two mechanisms controlling centrosomal area F-actin co-exist in B lymphocytes, and both regulate centrosome polarization.

Centrosomal F-actin and T Lymphocytes

We have shown that F-actin clearing at the cSMAC and centrosomal area F-actin depletion, respectively, mediated by PKC δ -dependent phosphorylation of FMNL1 β or paxillin, are associated with centrosome/MVB polarization and exosome secretion in CD4⁺ Jurkat T lymphocytes forming IS (Herranz et al., 2019; Bello-Gamboa et al., 2020) (**Figure 1B**). Although PKC δ appeared to regulate centrosomal area F-actin, FMNL1 β did not appear to participate in this regulation (Bello-Gamboa et al., 2020). A possible PKC δ downstream effector involved in centrosomal area F-actin reorganization could be the actin regulatory protein paxillin, whose phosphorylation at threonine 538 (T538) by PKC δ leads to actin cytoskeleton depolymerization and regulates integrin-mediated adhesion and migration of B lymphoid cells (Romanova et al., 2010). Moreover, the centrosome cannot polarize to the IS in paxillin-interfered CTL (Robertson and Ostergaard, 2011). In addition, paxillin phosphorylation is required for CTL lytic granule secretion (Robertson et al., 2005), and both paxillin (Herrerros et al., 2000) and PKC δ (Fanning et al., 2005) are localized at the centrosome. Thus, in CD4⁺ Jurkat T lymphocytes forming IS, we have found that PKC δ -dependent paxillin phosphorylation may govern a F-actin reorganization network different from F-actin at the IS, such as centrosomal F-actin, that may also contribute to the diminished centrosome polarization observed in PKC δ -interfered Jurkat T lymphocyte clones (**Figure 1B**). This PKC δ -dependent, paxillin-regulated mechanism for centrosomal area F-actin reorganization appears to co-exist in CD4⁺ Jurkat T lymphocytes with the PKC δ -dependent, FMNL1 β -regulated mechanism for cortical F-actin reorganization explained above. More research is necessary (i.e., experiments involving phospho-deficient and phospho-mimetic mutants of paxillin at T538 and/or FMNL1 β at S1086) to establish the relative contribution of these mechanisms to the polarization processes.

In addition, impaired F-actin reorganization at the IS was produced by centriole deletion in CTL (Tamzalit et al., 2020), which points out an unexpected role for the intact centrosome and/or centrosomal F-actin in supporting synaptic F-actin architecture and dynamics. Moreover, lower centrosomal actin filament densities enhanced microtubule growth at the centrosome (Inoue et al., 2019), that decisively affected cell adhesion and spreading. These results, together with the fact that ARP2/3 is partially depleted from the centrosome as a result of its recruitment to the IS (Farina et al., 2016), suggest an unsuspected direct or indirect interaction (i.e., competition for F-actin regulators such as ARP2/3), between cortical and centrosomal area F-actin networks, that in turn may regulate tubulin cytoskeleton at different subcellular locations. The

distinct F-actin networks may be functionally interconnected by coordinated activation of different actin assembly factors (Bello-Gamboa et al., 2020) (**Figure 1B**), competition for the same regulatory factor (Obino et al., 2016) and/or actin monomer availability (Suarez and Kovar, 2016). Pericentriolar material seemed to modulate the centrosomal F-actin network by regulating ARP2/3 and WASH recruitment to the centrosome (Farina et al., 2016). Since F-actin reorganization at the remaining proteinaceous pericentriolar material (PCM) area, containing pericentrin and γ -tubulin, was not analyzed in centriole-deficient CTL (Tamzalit et al., 2020), it will be interesting to study pericentriolar area F-actin in detail in these cells. In addition, considering that PKC δ is located in the centrosome (Fanning et al., 2005; Ma et al., 2008a), but also in lytic granules in CTL (Ma et al., 2008b), it is conceivable that PKC δ , directly or indirectly, may coordinately regulate both centrosomal area and synaptic F-actin networks.

CURRENT RESEARCH GAPS

Lipid Bilayer Synapse Model

Most of what we know about the formation, organization, and dynamics of the four described F-actin and actomyosin networks at the IS results from high spatio-temporal resolution image analysis of T cells engaged with activating surfaces such as coated glass and planar lipid bilayers, which position these networks in the ideal imaging plane, avoiding the Z spatial dimension (Hammer et al., 2018). This approach is certainly somewhat reductionist since it is not possible to guarantee that all the molecular interactions occurring in a real, cell to cell synapse will also occur upon interaction with the coated glass or the lipid bilayer (Fooksman et al., 2010). In fact, supported lipid bilayers do not completely imitate the complex and irregular surface of an APC or target cell, possibly causing non-physiological interactions in the IS (Bertolet and Liu, 2016). Thus, although studies using supported planar bilayers are powerful in terms of resolution and sensitivity, it is important to test the predictions of these model systems using *in vitro* or *in vivo* cell-cell systems in order to extend the results to a more physiologic scenario (Dustin, 2009).

Actin Cytoskeleton in Primary vs. Immortalized T Cells and Different Synapse Subtypes

Striking differences in F-actin architecture and dynamics at the IS have been found between human primary CD4⁺ and immortalized CD4⁺ T lymphocytes, such as Jurkat cells, under comparable activation conditions on lipid bilayers (Colin-York et al., 2019; Kumari et al., 2019). In contrast, no major differences were found between the synaptic architecture of Jurkat cells and mouse CD8⁺ primary CTLs (Murugesan et al., 2016) on stimulatory lipid bilayers. F-actin network dynamics and mechanics are likely to be different in these cells, and influence how T cells employ mechanical cues in different ways during antigen recognition (Kumari et al., 2019). Thus, caution should be taken when generalizing the cellular mechanisms underlying

the variety of IS patterns and motility behaviors to specific T cell subtypes and across different species (Kumari et al., 2019). It is conceivable that these differences also exist in different real cell-cell synapses, although they have not been observed yet (Colin-York et al., 2019; Kumari et al., 2019). In addition, while PKC δ regulates centrosomal area F-actin upon IS formation by CD4⁺ Jurkat cells (Bello-Gamboa et al., 2020), more research is necessary to extend these results to both CD8⁺ and CD4⁺ primary T lymphocytes forming IS. While TIRFM combined with super-resolution 3D-SIM provided probably the best live-cell images of synapses on lipid bilayer models (Murugesan et al., 2016; Colin-York et al., 2019), the best live-cell images of F-actin architecture and dynamics within T cells engaged with an APC were obtained using high temporal resolution, lattice light sheet microscopy (LLSM) in a diffraction-limited mode (Ritter et al., 2015; Fritzsche et al., 2017). An option to improve the image spatial resolution is taking advantage of the better XY resolution of microscopes with respect to the axial (Z) resolution (Calvo and Izquierdo, 2018), by using LLSM combined with 3D-SIM. Other options are placing the T lymphocyte on top of an APC by using “pairing and coupling” microfluidic devices (Jang et al., 2015), or on a very flat APC in which the IS is located in the XY optical plane (Wang et al., 2018). These last two options will indeed provide real cell-cell synapse models. While LLSM images in a diffraction-limited mode (Ritter et al., 2015; Fritzsche et al., 2017) showed with remarkable definition the branched F-actin network dynamics in the dSMAC, including its contribution to a flow of F-actin up the sides of the APC-bound T cell, none of the four F-actin networks observed by TIRFM combined with 3D-SIM in lipid bilayers were visible using LLSM (Hammer et al., 2018). These latter F-actin networks, all of which are much fainter F-actin structures than those observed in the dSMAC (Hammer et al., 2018), will probably become discernible in the future by using one or more of the cell setups and super-resolution imaging methods described above.

Centrosomal F-actin Network Characterization and Measurements

Although in the original publication the authors defined the existence of a “centrosomal” F-actin network, it should be underlined this is an operative definition that does not specify the extension and/or the limits of such a network (Farina et al., 2016). In fact, the authors arbitrarily defined a 2 μm -diameter region of interest (ROI) including the centrosome for F-actin fluorescence intensity measurements. Subsequent publications have incorporated for centrosomal F-actin measurements the use of a 1.6–2 μm diameter, centrosome-centered ROI (Obino et al., 2016; Ibanez-Vega et al., 2019; Bello-Gamboa et al., 2020). However, a more appropriate term could be “centrosomal area” F-actin, to remark that measures were referred to an area centered at the centrosome and surely larger than the centrosome itself. Indeed, this is a more accurate term that includes the complexity of organelle interconnection. This is an important issue for centrosomal area F-actin evaluation, since the imaging techniques used in these publications (confocal microscopy, TIRFM, epifluorescence microscopy plus deconvolution) do not

allow enough resolution to sustain that F-actin assembles at the pericentrosomal matrix instead of other membrane-bound organelles included in this area (Obino et al., 2016; Bello-Gamboa et al., 2019; Ibanez-Vega et al., 2019). Since interphase centrosomes are smaller than mitotic centrosomes (Decker et al., 2011) and cells may be at any cell cycle phase, it is difficult to establish a fixed centrosomal diameter. If a 2 μm -diameter centrosomal area ROI is analyzed, it cannot be ruled out that other F-actin-regulating organelles are included in this area, such as the Golgi, endosomes, or MVB (Colon-Franco et al., 2011; Bello-Gamboa et al., 2020). MVB are also involved in actin polymerization at the IS during intracellular reorganization (Calabia-Linares et al., 2011). Electron microscopy images show that vesicles and endosomes are located nearby the centrosomes (Ueda et al., 2011). The use of a pericentriolar marker, together with new imaging super-resolution techniques, would facilitate the study of the centrosomal area and the specific localization and dynamics of centrosomal F-actin. Indeed, in the future, emerging and promising techniques, such as LLSM (Ritter et al., 2015; Fritzsche et al., 2017), combined with non-diffraction limited, super-resolution microscopy (Fritzsche et al., 2017; Calvo and Izquierdo, 2018), may help to a better definition of centrosomal area F-actin structure and function, as it occurred for the four recently defined, synaptic F-actin networks that contribute to maintain the shape and function of the canonical IS (Section Signals Regulating Cortical Actin Reorganization In The Immune Synapse). In this context, TIRFM or TIRFM combined with 3D-SIM are exceptionally useful imaging techniques to study both F-actin cytoskeleton and secretion vesicle degranulation on the XY plane of coated glass or lipid bilayer (Rak et al., 2011; Murugesan et al., 2016; Sinha et al., 2016; Carisey et al., 2018), due to its high signal-to-noise ratio and improved spatial resolution below the diffraction limit (Calvo and Izquierdo, 2018). However, this procedure is somewhat limited since only an illuminated homogeneous surface can be used to stimulate cells and TIRFM would dismiss subcellular structures (i.e., centrosome) or molecules located beyond a minor Z distance from the stimulatory surface (>200–300 nm). Thus, centrosome movement in the Z dimension from distal subcellular locations cannot be imaged, although its last stages approaching the IS can be properly imaged. One possibility to circumvent this problem is to limit centrosome movement in the Z dimension by using the cell setups explained above. Indeed, other techniques as the already mentioned diffraction-limited LLSM, combined with 2D or 3D stimulated emission depletion (STED) super-resolution microscopy (Fritzsche et al., 2017), may provide in the future new tools to address the study of centrosomal area F-actin organization and dynamics at the nanoscale level.

POTENTIAL FUTURE DEVELOPMENTS IN THE FIELD. IMAGING THE IMMUNOLOGICAL SYNAPSE

For adequate IS imaging by fluorescence microscopy, harmonizing temporal and spatial resolutions, overcoming

spatial constraints due to imaging in Z optical axis, improving signal-to-noise ratio, and solving the photobleaching and cytotoxicity inherent to any live cell imaging, are required (Combs and Shroff, 2017; Calvo and Izquierdo, 2018). Please refer to some excellent reviews dealing with the more relevant fluorescence microscopy techniques used in cell biology in general (Combs and Shroff, 2017; Lambert and Waters, 2017; Sahl et al., 2017), and specifically for IS imaging (Rossy et al., 2013; Calvo and Izquierdo, 2018; Herranz et al., 2019), since it is out of the scope of this review to deal with these relevant technical issues. In this context, it is remarkable that to date difficulties in visualization of both primary and transformed T lymphocyte models at high spatio-temporal resolution have somewhat limited our understanding of the principles underlying T lymphocyte subtype-specific activation (Kumari et al., 2019). However, visualization and quantification of actin cytoskeleton and the underlying patterns and dynamics has evolved to a significant degree due to the advances in microscopy regarding spatiotemporal resolution (Kumari et al., 2019). Currently, we only have a limited knowledge of how actin cytoskeleton organization affects distinct synaptic patterning and signaling. This is due to several facts, including the low transfection efficiency of F-actin reporters in primary lymphocytes, the rapid kinetics of changes, the reduced area of cell-cell synapses, and the active, highly plastic and irregular cell-cell synaptic interface that precludes image capture at high spatiotemporal resolution (Kumari et al., 2019; Blumenthal and Burkhardt, 2020). In the future, the development of emerging and promising techniques such as LLSM (Ritter et al., 2015) and new 3D live-cell super-resolution microscopy (Fritzsche et al., 2017), combined with some useful probes for F-actin in living cells (Lukinavicius et al., 2013, 2014; Bello-Gamboa et al., 2020) will transform how we image cellular and protein dynamics during IS interactions. These advances will indeed shed more light into our knowledge of these processes.

Thus, some techniques of choice have been specifically used for IS imaging and to overcome the mentioned caveats. Planar lipid bilayers and coverslips or beads-coated with surface proteins or agonistic antibodies are good options. These approaches reduce a 3D complex structure such a cell-cell IS to only two dimensions (XY), enabling high-resolution imaging techniques such as TIRFM (Huppa and Davis, 2003) and, since stimulation occurs at a homogenous, well-defined Z position, image capture at high spatial resolution becomes feasible. If the imaged cell is flat enough, or the Z dimension-restricted cell setups described above are used, secretory vesicle movement at the XY focus plane is a centripetal convergence toward the cSMAC area and can be conveniently imaged and analyzed (Fooksman et al., 2010; Sinha et al., 2016). Using this technique for some structures contained and reorganizing within the IS (i.e., F-actin), the images obtained by using anti-TCR-coated coverslips and lipid bilayers and TIRFM and TIRFM-SIM combination probably exhibit, by far, the highest definition and spatial resolution obtained to date (Murugesan et al., 2016; Sinha et al., 2016). The opportunity to change the composition of the lipid bilayer or the stimulatory antibodies by loading antigens, accessory molecules, changing lipids, etc., allows for reconstitution approaches,

increasing the flexibility of this strategy (Huppa and Davis, 2003), although centrosomal area F-actin imaging in living cells will require to develop alternative strategies such as LLSM combined with super-resolution imaging techniques (i.e., STED, 3D-SIM), harboring higher temporal resolution and full competence in the Z dimension.

Apart of the described role of F-actin regions and SMACs in vesicle secretion obtained by high-resolution microscopy, emerging evidences obtained thanks to high-resolution live imaging microscopy support that F-actin-driven and maintained structures such as T cell microvilli, acting as finger-like membrane protrusions or invadosome-like protrusions, may participate in sensing pMHC on APCs, acting as bona fide “synptosomes” (Sage et al., 2012; Kim et al., 2018; Kim and Jun, 2019), or acting as interfacial protrusions at the IS contact area to facilitate lytic granule secretion and CTL activity (Tamzalit et al., 2019). The formation, maintenance and activity of the later protrusions, as SMACs architecture and functions, both depend on WASP and ARP2/3 activity. The fact that some of the IS actin networks consist of dense actin foci related to protrusive structures rich in F-actin, called invadosome-like protrusions (ILPs) (Hammer et al., 2018; Blumenthal and Burkhardt, 2020), supports that these protrusions should also be analyzed in the IS and actin cytoskeleton studies. Moreover, T lymphocyte microvilli should also be considered not only as structures involved in surveying antigen on APCs or target cells, but also in signaling to APCs or target cells. Thus, these structures are related with relevant immune regulation mechanisms previously discovered (Kim and Jun, 2019). More research involving state-of-the-art microscopy techniques is necessary to understand the mechanisms controlling their generation and function.

CONCLUDING REMARKS

Cells precisely control the formation and the dynamics of both tubulin and actin cytoskeleton networks to coordinate important processes, including motility, cell division, endocytosis and polarized secretion. In addition, cells coordinate the formation of distinct F-actin networks from a general cytosolic pool of actin monomers (Suarez and Kovar, 2016). The available literature concerning the centrosomal subcellular localization and actin cytoskeleton dynamics described here and elsewhere (Dogterom and Koenderink, 2019) demonstrate the existence of a relevant connection between tubulin and actin cytoskeletons and centrosome/MVB polarized traffic and function. Although these links between F-actin and microtubule dynamics are intriguing, very little is known about their molecular bases and functional relevance (Le Floc’h and Huse, 2015). In addition, recent evidences demonstrate that the different F-actin networks appear to be co-ordinately regulated and interconnected. Close coordination between centrosome and centrosomal area F-actin with synaptic F-actin could facilitate the efficient organization of synaptic responses in space and time (Tamzalit et al., 2020). In addition, inhibitor studies indicate that the four discrete cortical actin networks more recently described at the IS (Section Signals Regulating Cortical Actin Reorganization In The Immune

Synapse) largely function independently of one another, although there is some coordinate control due to competition for free actin monomer (Hammer et al., 2018). How these distinct cortical and non-cortical networks are regulated, how they interact with the TCR signaling network, as well as their interconnections constitute an intriguing and challenging issue to be addressed in the future. The application of super-resolution microscopy in this context will enable, together with conventional biochemistry techniques, to tackle some of these issues through directly analyzing the interactions between the cytoskeletons and other cell proteins at immune synapses.

AUTHOR CONTRIBUTIONS

VC and MI: conceived the manuscript, writing of the manuscript, approved its final content, conceptualization, and writing—review and editing. MI: writing original draft preparation. Both authors contributed to the article and approved the submitted version.

REFERENCES

- Alonso, R., Mazzeo, C., Rodriguez, M. C., Marsh, M., Fraile-Ramos, A., Calvo, V., et al. (2011). Diacylglycerol kinase alpha regulates the formation and polarisation of mature multivesicular bodies involved in the secretion of Fas ligand-containing exosomes in T lymphocytes. *Cell Death Differ.* 18, 1161–1173. doi: 10.1038/cdd.2010.184
- Alonso, R., Rodriguez, M. C., Pindado, J., Merino, E., Merida, I., and Izquierdo, M. (2005). Diacylglycerol kinase alpha regulates the secretion of lethal exosomes bearing Fas ligand during activation-induced cell death of T lymphocytes. *J. Biol. Chem.* 280, 28439–28450. doi: 10.1074/jbc.M501112200
- Bello-Gamboa, A., Izquierdo, J. M., Velasco, M., Moreno, S., Garrido, A., Meyers, L., et al. (2019). Imaging the human immunological synapse. *J. Vis. Exp.* 154:e60312. doi: 10.3791/60312
- Bello-Gamboa, A., Velasco, M., Moreno, S., Herranz, G., Ilie, R., Huetos, S., et al. (2020). Actin reorganization at the centrosomal area and the immune synapse regulates polarized secretory traffic of multivesicular bodies in T lymphocytes. *J. Extracell. Vesicles* 9:1759926. doi: 10.1080/20013078.2020.1759926
- Bertolet, G., and Liu, D. (2016). The planar lipid bilayer system serves as a reductionist approach for studying NK cell immunological synapses and their functions. *Methods Mol. Biol.* 1441, 151–165. doi: 10.1007/978-1-4939-3684-7_13
- Bertrand, F., Muller, S., Roh, K. H., Laurent, C., Dupre, L., and Valitutti, S. (2013). An initial and rapid step of lytic granule secretion precedes microtubule organizing center polarization at the cytotoxic T lymphocyte/target cell synapse. *Proc. Natl. Acad. Sci. U. S. A.* 110, 6073–6078. doi: 10.1073/pnas.1218640110
- Billadeau, D. D., Nolz, J. C., and Gomez, T. S. (2007). Regulation of T-cell activation by the cytoskeleton. *Nat. Rev. Immunol.* 7, 131–143. doi: 10.1038/nri2021
- Blumenthal, D., and Burkhardt, J. K. (2020). Multiple actin networks coordinate mechanotransduction at the immunological synapse. *J. Cell Biol.* 219:e201911058. doi: 10.1083/jcb.201911058
- Brossard, C., Feuillet, V., Schmitt, A., Randriamampita, C., Romao, M., Raposo, G., et al. (2005). Multifocal structure of the T cell - dendritic cell synapse. *Eur. J. Immunol.* 35, 1741–1753. doi: 10.1002/eji.200425857
- Calabia-Linares, C., Robles-Valero, J., de la Fuente, H., Perez-Martinez, M., Martin-Cofreces, N., Alfonso-Perez, M., et al. (2011). Endosomal clathrin drives actin accumulation at the immunological synapse. *J. Cell Sci.* 124 (Pt 5), 820–830. doi: 10.1242/jcs.078832
- Calvo, V., and Izquierdo, M. (2018). Imaging polarized secretory traffic at the immune synapse in living T lymphocytes. *Front. Immunol.* 9:684. doi: 10.3389/fimmu.2018.00684

FUNDING

This research was funded by grants from the Spanish Ministerio de Economía y Competitividad (MINECO), Plan Nacional de Investigación Científica (SAF2016-77561-R and PID2020-114148RB-I00) to MI, which was in part granted with FEDER funding (EC), corresponding to the Programa Estatal de Investigación, Desarrollo e Innovación Orientada a los Retos de la Sociedad.

ACKNOWLEDGMENTS

The authors apologize for not including some relevant references due to space limitations. We acknowledge all the past and present members of the lab for their generous contribution. We acknowledge the support of the publication fee by the CSIC Open Access Publication Support Initiative through its Unit of Information Resources for Research (URICI).

- Calvo, V., and Izquierdo, M. (2020). Inducible polarized secretion of exosomes in T and B lymphocytes. *Int. J. Mol. Sci.* 21:2631. doi: 10.3390/ijms21072631
- Carisey, A. F., Mace, E. M., Saeed, M. B., Davis, D. M., and Orange, J. S. (2018). Nanoscale dynamism of actin enables secretory function in cytolytic cells. *Curr. Biol.* 28, 489–502.e9. doi: 10.1016/j.cub.2017.12.044
- Chemin, K., Bohineust, A., Dogniaux, S., Turret, M., Guegan, S., Miro, F., et al. (2012). Cytokine secretion by CD4+ T cells at the immunological synapse requires Cdc42-dependent local actin remodeling but not microtubule organizing center polarity. *J. Immunol.* 189, 2159–2168. doi: 10.4049/jimmunol.1200156
- Colin-York, H., Kumari, S., Barbieri, L., Cords, L., and Fritzsche, M. (2019). Distinct actin cytoskeleton behaviour in primary and immortalised T-cells. *J. Cell Sci.* 133:jcs232322. doi: 10.1242/jcs.232322
- Colon-Franco, J. M., Gomez, T. S., and Billadeau, D. D. (2011). Dynamic remodeling of the actin cytoskeleton by FMNL1gamma is required for structural maintenance of the Golgi complex. *J. Cell Sci.* 124 (Pt 18), 3118–3126. doi: 10.1242/jcs.083725
- Combs, C. A., and Shroff, H. (2017). Fluorescence microscopy: a concise guide to current imaging methods. *Curr. Protoc. Neurosci.* 79, 2.1.1–2.1.25. doi: 10.1002/cpns.29
- Combs, J., Kim, S. J., Tan, S., Ligon, L. A., Holzbaur, E. L., Kuhn, J., et al. (2006). Recruitment of dynein to the Jurkat immunological synapse. *Proc. Natl. Acad. Sci. U. S. A.* 103, 14883–14888. doi: 10.1073/pnas.0600914103
- Comrie, W. A., Babich, A., and Burkhardt, J. K. (2015). F-actin flow drives affinity maturation and spatial organization of LFA-1 at the immunological synapse. *J. Cell Biol.* 208, 475–491. doi: 10.1083/jcb.201406121
- de la Roche, M., Asano, Y., and Griffiths, G. M. (2016). Origins of the cytolytic synapse. *Nat. Rev. Immunol.* 16, 421–432. doi: 10.1038/nri.2016.54
- Decker, M., Jaensch, S., Pozniakovskiy, A., Zinke, A., O'Connell, K. F., Zachariae, W., et al. (2011). Limiting amounts of centrosome material set centrosome size in *C. elegans* embryos. *Curr. Biol.* 21, 1259–1267. doi: 10.1016/j.cub.2011.06.002
- Dogterom, M., and Koenderink, G. H. (2019). Actin-microtubule crosstalk in cell biology. *Nat. Rev. Mol. Cell Biol.* 20, 38–54. doi: 10.1038/s41580-018-0067-1
- Duchez, S., Rodrigues, M., Bertrand, F., and Valitutti, S. (2011). Reciprocal polarization of T and B cells at the immunological synapse. *J. Immunol.* 187, 4571–4580. doi: 10.4049/jimmunol.1100600
- Dustin, M. L. (2009). Supported bilayers at the vanguard of immune cell activation studies. *J. Struct. Biol.* 168, 152–160. doi: 10.1016/j.jsb.2009.05.007
- Fanning, A., Volkov, Y., Freeley, M., Kelleher, D., and Long, A. (2005). CD44 cross-linking induces protein kinase C-regulated migration of human T lymphocytes. *Int. Immunol.* 17, 449–458. doi: 10.1093/intimm/dxh225

- Farina, F., Gaillard, J., Guerin, C., Couste, Y., Sillibourne, J., Blanchoin, L., et al. (2016). The centrosome is an actin-organizing centre. *Nat. Cell Biol.* 18, 65–75. doi: 10.1038/ncb3285
- Fooksman, D. R., Vardhana, S., Vasiliver-Shamis, G., Liese, J., Blair, D. A., Waite, J., et al. (2010). Functional anatomy of T cell activation and synapse formation. *Annu. Rev. Immunol.* 28, 79–105. doi: 10.1146/annurev-immunol-030409-101308
- Fritzsche, M., Fernandes, R. A., Chang, V. T., Colin-York, H., Clausen, M. P., Felce, J. H., et al. (2017). Cytoskeletal actin dynamics shape a ramifying actin network underpinning immunological synapse formation. *Sci. Adv.* 3:e1603032. doi: 10.1126/sciadv.1603032
- Gomez, T. S., Hamann, M. J., McCarney, S., Savoy, D. N., Lubking, C. M., Heldebrandt, M. P., et al. (2005). Dynamin 2 regulates T cell activation by controlling actin polymerization at the immunological synapse. *Nat. Immunol.* 6, 261–270. doi: 10.1038/nri1168
- Gomez, T. S., Kumar, K., Medeiros, R. B., Shimizu, Y., Leibson, P. J., and Billadeau Daniel, D. (2007). Formins regulate the actin-related protein 2/3 complex-independent polarization of the centrosome to the immunological synapse. *Immunity* 26, 177–190. doi: 10.1016/j.immuni.2007.01.008
- Gomez, T. S., McCarney, S. D., Carrizosa, E., Labno, C. M., Comiskey, E. O., Nolz, J. C., et al. (2006). HSI functions as an essential actin-regulatory adaptor protein at the immune synapse. *Immunity* 24, 741–752. doi: 10.1016/j.immuni.2006.03.022
- Griffiths, G. M., Tsun, A., and Stinchcombe, J. C. (2010). The immunological synapse: a focal point for endocytosis and exocytosis. *J. Cell Biol.* 189, 399–406. doi: 10.1083/jcb.201002027
- Hammer, J. A., Wang, J. C., Saeed, M., and Pedrosa, A. T. (2018). Origin, organization, dynamics, and function of actin and actomyosin networks at the T cell immunological synapse. *Annu. Rev. Immunol.* 37, 201–224. doi: 10.1146/annurev-immunol-042718-041341
- Herranz, G., Aguilera, P., Davila, S., Sanchez, A., Stancu, B., Gomez, J., et al. (2019). Protein kinase C delta regulates the depletion of actin at the immunological synapse required for polarized exosome secretion by T cells. *Front. Immunol.* 10:851. doi: 10.3389/fimmu.2019.00851
- Herreros, L., Rodriguez-Fernandez, J. L., Brown, M. C., Alonso-Lebrero, J. L., Cabanas, C., Sanchez-Madrid, F., et al. (2000). Paxillin localizes to the lymphocyte microtubule organizing center and associates with the microtubule cytoskeleton. *J. Biol. Chem.* 275, 26436–26440. doi: 10.1074/jbc.M003970200
- Huppa, J. B., and Davis, M. M. (2003). T-cell-antigen recognition and the immunological synapse. *Nat. Rev. Immunol.* 3, 973–983. doi: 10.1038/nri1245
- Huse, M. (2012). Microtubule-organizing center polarity and the immunological synapse: protein kinase C and beyond. *Front. Immunol.* 3:235. doi: 10.3389/fimmu.2012.00235
- Huse, M., Quann, E. J., and Davis, M. M. (2008). Shouts, whispers and the kiss of death: directional secretion in T cells. *Nat. Immunol.* 9, 1105–1111. doi: 10.1038/nri.f.215
- Ibanez-Vega, J., Del Valle Batalla, F., Saez, J. J., Soza, A., and Yuseff, M. I. (2019). Proteasome dependent actin remodeling facilitates antigen extraction at the immune synapse of B cells. *Front. Immunol.* 10:225. doi: 10.3389/fimmu.2019.00225
- Inoue, D., Obino, D., Pineau, J., Farina, F., Gaillard, J., Guerin, C., et al. (2019). Actin filaments regulate microtubule growth at the centrosome. *EMBO J.* 38:e99630. doi: 10.15252/embj.201899630
- Izquierdo, M., and Cantrell, D. A. (1992). T-cell activation. *Trends Cell Biol.* 2, 268–271. doi: 10.1016/0962-8924(92)90199-W
- Jang, J. H., Huang, Y., Zheng, P., Jo, M. C., Bertolet, G., Zhu, M. X., et al. (2015). Imaging of cell-cell communication in a vertical orientation reveals high-resolution structure of immunological synapse and novel PD-1 dynamics. *J. Immunol.* 195, 1320–1330. doi: 10.4049/jimmunol.1403143
- Kim, H. R., and Jun, C. D. (2019). T cell microvilli: sensors or senders? *Front. Immunol.* 10:1753. doi: 10.3389/fimmu.2019.01753
- Kim, H. R., Mun, Y., Lee, K. S., Park, Y. J., Park, J. S., Park, J. H., et al. (2018). T cell microvilli constitute immunological synaptosomes that carry messages to antigen-presenting cells. *Nat. Commun.* 9:3630. doi: 10.1038/s41467-018-06090-8
- Kühn, S., and Geyer, M. (2014). Formins as effector proteins of Rho GTPases. *Small GTPases* 5:e29513. doi: 10.4161/sgtp.29513
- Kumari, S., Colin-York, H., Irvine, D. J., and Fritzsche, M. (2019). Not all T cell synapses are built the same way. *Trends Immunol.* 40, 977–980. doi: 10.1016/j.it.2019.09.009
- Kumari, S., Curado, S., Mayya, V., and Dustin, M. L. (2014). T cell antigen receptor activation and actin cytoskeleton remodeling. *Biochim. Biophys. Acta* 1838, 546–556. doi: 10.1016/j.bbammem.2013.05.004
- Kuokkanen, E., Sustar, V., and Mattila, P. K. (2015). Molecular control of B cell activation and immunological synapse formation. *Traffic* 16, 311–326. doi: 10.1111/tra.12257
- Laguerre, K., Carisey, A., Oszmiana, A., Kennedy, P. R., Williamson, D. J., Cartwright, A., et al. (2013). The central role of the cytoskeleton in mechanisms and functions of the NK cell immune synapse. *Immunol. Rev.* 256, 203–221. doi: 10.1111/imr.12107
- Lambert, T. J., and Waters, J. C. (2017). Navigating challenges in the application of superresolution microscopy. *J. Cell Biol.* 216, 53–63. doi: 10.1083/jcb.201610011
- Le Floch, A., and Huse, M. (2015). Molecular mechanisms and functional implications of polarized actin remodeling at the T cell immunological synapse. *Cell. Mol. Life Sci.* 72, 537–556. doi: 10.1007/s00018-014-1760-7
- Li, J., Yin, W., Jing, Y., Kang, D., Yang, L., Cheng, J., et al. (2018). The coordination between B cell receptor signaling and the actin cytoskeleton during B cell activation. *Front. Immunol.* 9:3096. doi: 10.3389/fimmu.2018.03096
- Liu, X., Kapoor, T. M., Chen, J. K., and Huse, M. (2013). Diacylglycerol promotes centrosome polarization in T cells via reciprocal localization of dynein and myosin II. *Proc. Natl. Acad. Sci. U. S. A.* 110, 11976–11981. doi: 10.1073/pnas.1306180110
- Lukinavicius, G., Reymond, L., D'Este, E., Masharina, A., Gottfert, F., Ta, H., et al. (2014). Fluorogenic probes for live-cell imaging of the cytoskeleton. *Nat. Methods* 11, 731–733. doi: 10.1038/nmeth.2972
- Lukinavicius, G., Umezawa, K., Olivier, N., Honigsmann, A., Yang, G., Plass, T., et al. (2013). A near-infrared fluorophore for live-cell super-resolution microscopy of cellular proteins. *Nat. Chem.* 5, 132–139. doi: 10.1038/nchem.1546
- Ma, J. S., Haydar, T. F., and Radoja, S. (2008b). Protein kinase C delta localizes to secretory lysosomes in CD8+ CTL and directly mediates TCR signals leading to granule exocytosis-mediated cytotoxicity. *J. Immunol.* 181, 4716–4722. doi: 10.4049/jimmunol.181.7.4716
- Ma, J. S., Monu, N., Shen, D. T., Mecklenbrauker, I., Radoja, N., Haydar, T. F., et al. (2007). Protein kinase C delta regulates antigen receptor-induced lytic granule polarization in mouse CD8+ CTL. *J. Immunol.* 178, 7814–7821. doi: 10.4049/jimmunol.178.12.7814
- Ma, W., Koch, J. A., and Viveiros, M. M. (2008a). Protein kinase C delta (PKCdelta) interacts with microtubule organizing center (MTOC)-associated proteins and participates in meiotic spindle organization. *Dev. Biol.* 320, 414–425. doi: 10.1016/j.ydbio.2008.05.550
- Mazzeo, C., Calvo, V., Alonso, R., Merida, I., and Izquierdo, M. (2016). Protein kinase D1/2 is involved in the maturation of multivesicular bodies and secretion of exosomes in T and B lymphocytes. *Cell Death Differ.* 23, 99–109. doi: 10.1038/cdd.2015.72
- Merino-Cortés, S. V., Gardeta, S. R., Roman-García, S., Martínez-Riaño, A., Pineau, J., Liebana, R., et al. (2020). Diacylglycerol kinase ζ promotes actin cytoskeleton remodeling and mechanical forces at the B cell immune synapse. *Sci. Signal.* 13:eaaw8214. doi: 10.1126/scisignal.aaw8214
- Monks, C. R. F., Freiberg, B. A., Kupfer, H., Sciaky, N., and Kupfer, A. (1998). Three-dimensional segregation of supramolecular activation clusters in T cells. *Nature* 395, 82–86. doi: 10.1038/25764
- Muntasell, A., Berger, A. C., and Roche, P. A. (2007). T cell-induced secretion of MHC class II-peptide complexes on B cell exosomes. *EMBO J.* 26, 4263–4272. doi: 10.1038/sj.emboj.7601842
- Murugesan, S., Hong, J., Yi, J., Li, D., Beach, J. R., Shao, L., et al. (2016). Formin-generated actomyosin arcs propel T cell receptor microcluster movement at the immune synapse. *J. Cell Biol.* 215, 383–399. doi: 10.1083/jcb.201603080
- Na, B.-R., Kim, H.-R., Piragyte, I., Oh, H.-M., Kwon, M.-S., Akber, U., et al. (2015). TAGLN2 regulates T cell activation by stabilizing the actin cytoskeleton at the immunological synapse. *J. Cell Biol.* 209, 143–162. doi: 10.1083/jcb.201407130
- Na, B. R., Kwon, M. S., Chae, M. W., Kim, H. R., Kim, C. H., Jun, C. D., et al. (2016). Transgelin-2 in B-cells controls T-cell activation by stabilizing T cell - B cell conjugates. *PLoS One* 11:e0156429. doi: 10.1371/journal.pone.0156429

- Nath, S., Christian, L., Tan, S. Y., Ki, S., Ehrlich, L. I. R., and Poenie, M. (2016). Dynein separately partners with NDE1 and dynactin to orchestrate T cell focused secretion. *J. Immunol.* 197, 2090–2101. doi: 10.4049/jimmunol.1600180
- Obino, D., Farina, F., Malbec, O., Saez, P. J., Maurin, M., Gaillard, J., et al. (2016). Actin nucleation at the centrosome controls lymphocyte polarity. *Nat. Commun.* 7:10969. doi: 10.1038/ncomms10969
- Peters, P. J., Geuze, H. J., Van der Donk, H. A., Slot, J. W., Griffith, J. M., Stam, N. J., et al. (1989). Molecules relevant for T cell-target cell interaction are present in cytolytic granules of human T lymphocytes. *Eur. J. Immunol.* 19, 1469–1475. doi: 10.1002/eji.1830190819
- Plessner, M., Knerr, J., and Grosse, R. (2019). Centrosomal actin assembly is required for proper mitotic spindle formation and chromosome congression. *iScience* 15, 274–281. doi: 10.1016/j.isci.2019.04.022
- Quann, E. J., Liu, X., Altan-Bonnet, G., and Huse, M. (2011). A cascade of protein kinase C isozymes promotes cytoskeletal polarization in T cells. *Nat. Immunol.* 12, 647–654. doi: 10.1038/ni.2033
- Quann, E. J., Merino, E., Furuta, T., and Huse, M. (2009). Localized diacylglycerol drives the polarization of the microtubule-organizing center in T cells. *Nat. Immunol.* 10, 627–635. doi: 10.1038/ni.1734
- Rak, G. D., Mace, E. M., Banerjee, P. P., Svitkina, T., and Orange, J. S. (2011). Natural killer cell lytic granule secretion occurs through a pervasive actin network at the immune synapse. *PLoS Biol.* 9:e1001151. doi: 10.1371/journal.pbio.1001151
- Ritter, A. T., Asano, Y., Stinchcombe, J. C., Dieckmann, N. M., Chen, B. C., Gawden-Bone, C., et al. (2015). Actin depletion initiates events leading to granule secretion at the immunological synapse. *Immunity* 42, 864–876. doi: 10.1016/j.immuni.2015.04.013
- Ritter, A. T., Kapnick, S. M., Murugesan, S., Schwartzberg, P. L., Griffiths, G. M., and Lippincott-Schwartz, J. (2017). Cortical actin recovery at the immunological synapse leads to termination of lytic granule secretion in cytotoxic T lymphocytes. *Proc. Natl. Acad. Sci. U. S. A.* 114:201710751. doi: 10.1073/pnas.1710751114
- Robertson, L. K., Mireau, L. R., and Ostergaard, H. L. (2005). A role for phosphatidylinositol 3-kinase in TCR-stimulated ERK activation leading to paxillin phosphorylation and CTL degranulation. *J. Immunol.* 175, 8138–8145. doi: 10.4049/jimmunol.175.12.8138
- Robertson, L. K., and Ostergaard, H. L. (2011). Paxillin associates with the microtubule cytoskeleton and the immunological synapse of CTL through its leucine-aspartic acid domains and contributes to microtubule organizing center reorientation. *J. Immunol.* 187, 5824–5833. doi: 10.4049/jimmunol.1003690
- Romanova, L. Y., Holmes, G., Bahte, S. K., Kovalchuk, A. L., Nelson, P. J., Ward, Y., et al. (2010). Phosphorylation of paxillin at threonine 538 by PKCdelta regulates LFA1-mediated adhesion of lymphoid cells. *J. Cell Sci.* 123 (Pt 9), 1567–1577. doi: 10.1242/jcs.060996
- Rossy, J., Pigeon, S. V., Davis, D. M., and Gaus, K. (2013). Super-resolution microscopy of the immunological synapse. *Curr. Opin. Immunol.* 25, 307–312. doi: 10.1016/j.coi.2013.04.002
- Sage, P. T., Varghese, L. M., Martinelli, R., Sciuto, T. E., Kamei, M., Dvorak, A. M., et al. (2012). Antigen recognition is facilitated by invadosome-like protrusions formed by memory/effector T cells. *J. Immunol.* 188, 3686–3699. doi: 10.4049/jimmunol.1102594
- Sahl, S. J., Hell, S. W., and Jakobs, S. (2017). Fluorescence nanoscopy in cell biology. *Nat. Rev. Mol. Cell Biol.* 18, 685–701. doi: 10.1038/nrm.2017.71
- Sanchez, A. D., and Feldman, J. L. (2017). Microtubule-organizing centers: from the centrosome to non-centrosomal sites. *Curr. Opin. Cell Biol.* 44, 93–101. doi: 10.1016/j.cob.2016.09.003
- Sanchez, E., Liu, X., and Huse, M. (2019). Actin clearance promotes polarized dynein accumulation at the immunological synapse. *PLoS ONE* 14:e0210377. doi: 10.1371/journal.pone.0210377
- Schnyder, T., Castello, A., Feest, C., Harwood, N. E., Oellerich, T., Urlaub, H., et al. (2011). B cell receptor-mediated antigen gathering requires ubiquitin ligase Cbl and adaptors Grb2 and Dok-3 to recruit dynein to the signaling microcluster. *Immunity* 34, 905–918. doi: 10.1016/j.immuni.2011.06.001
- Siemasko, K., Eisfelder, B. J., Williamson, E., Kabak, S., and Clark, M. R. (1998). Cutting edge: signals from the B lymphocyte antigen receptor regulate MHC class II containing late endosomes. *J. Immunol.* 160, 5203–5208.
- Sinai, P., Nguyen, C., Schatzle, J. D., and Wülfing, C. (2010). Transience in polarization of cytolytic effectors is required for efficient killing and controlled by Cdc42. *Proc. Natl. Acad. Sci.* 107:11912. doi: 10.1073/pnas.0913422107
- Sinha, S., Hoshino, D., Hong, N. H., Kirkbride, K. C., Grega-Larson, N. E., Seiki, M., et al. (2016). Cortactin promotes exosome secretion by controlling branched actin dynamics. *J. Cell Biol.* 214, 197–213. doi: 10.1083/jcb.201601025
- Spitaler, M., Emslie, E., Wood, C. D., and Cantrell, D. (2006). Diacylglycerol and protein kinase D localization during T lymphocyte activation. *Immunity* 24, 535–546. doi: 10.1016/j.immuni.2006.02.013
- Stinchcombe, J. C., Majorovits, E., Bossi, G., Fuller, S., and Griffiths, G. M. (2006). Centrosome polarization delivers secretory granules to the immunological synapse. *Nature* 443, 462–465. doi: 10.1038/nature05071
- Stowers, L., Yelon, D., Berg, L. J., and Chant, J. (1995). Regulation of the polarization of T cells toward antigen-presenting cells by Ras-related GTPase CDC42. *Proc. Natl. Acad. Sci.* 92:5027. doi: 10.1073/pnas.92.11.5027
- Suarez, C., and Kovar, D. R. (2016). Internetwork competition for monomers governs actin cytoskeleton organization. *Nat. Rev. Mol. Cell Biol.* 17, 799–810. doi: 10.1038/nrm.2016.106
- Tamzalit, F., Tran, D., Jin, W., Boyko, V., Bazzi, H., Kepecs, A., et al. (2020). Centrioles control the capacity, but not the specificity, of cytotoxic T cell killing. *Proc. Natl. Acad. Sci. U. S. A.* 117, 4310–4319. doi: 10.1073/pnas.1913220117
- Tamzalit, F., Wang, M. S., Jin, W., Tello-Lafoz, M., Boyko, V., Heddlston, J. M., et al. (2019). Interfacial actin protrusions mechanically enhance killing by cytotoxic T cells. *Sci. Immunol.* 4:eaav5445. doi: 10.1126/sciimmunol.aav5445
- Ueda, H., Mophew, M. K., McIntosh, J. R., and Davis, M. M. (2011). CD4+ T-cell synapses involve multiple distinct stages. *Proc. Natl. Acad. Sci. U. S. A.* 108, 17099–17104. doi: 10.1073/pnas.1113703108
- Wang, J. C., Bolger-Munro, M., and Gold, M. R. (2018). “Imaging the interactions between B cells and antigen-presenting cells,” in *B Cell Receptor Signaling: Methods and Protocols*, ed C. Liu (New York, NY: Springer New York), 131–161.
- Wang, Y., Arjonen, A., Pouwels, J., Ta, H., Pausch, P., Bange, G., et al. (2015). Formin-like 2 promotes β 1-integrin trafficking and invasive motility downstream of PKC α . *Dev. Cell* 34, 475–483. doi: 10.1016/j.devcel.2015.06.015
- Xie, J., Tato, C. M., and Davis, M. M. (2013). How the immune system talks to itself: the varied role of synapses. *Immunol. Rev.* 251, 65–79. doi: 10.1111/imr.12017
- Yuseff, M. I., Pierobon, P., Reversat, A., and Lennon-Dumenil, A. M. (2013). How B cells capture, process and present antigens: a crucial role for cell polarity. *Nat. Rev. Immunol.* 13, 475–486. doi: 10.1038/nri3469
- Yuseff, M. I., Reversat, A., Lankar, D., Diaz, J., Fanget, I., Pierobon, P., et al. (2011). Polarized secretion of lysosomes at the B cell synapse couples antigen extraction to processing and presentation. *Immunity* 35, 361–374. doi: 10.1016/j.immuni.2011.07.008

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Calvo and Izquierdo. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.