



# Diacylglycerol Kinases (DGKs): Novel Targets for Improving T Cell Activity in Cancer

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Diacylglycerol kinases (DGKs) are a family of enzymes that catalyze the metabolism of diacylglycerol (DAG). Two isoforms of DGK, DGK $\alpha$ , and DGK $\zeta$ , specifically regulate the pool of DAG that is generated as a second messenger after stimulation of the T cell receptor (TCR). Deletion of either isoform in mouse models results in T cells bearing a hyperresponsive phenotype and enhanced T cell activity against malignancy. Whereas, DGK $\zeta$  appears to be the dominant isoform in T cells, rationale exists for targeting both isoforms individually or coordinately. Additional work is needed to rigorously identify the molecular changes that result from deletion of DGKs in order to understand how DAG contributes to T cell activation, the effect of DGK inhibition in human T cells, and to rationally develop combined immunotherapeutic strategies that target DGKs.

**Keywords:** diacylglycerol, diacylglycerol kinase, immunotherapy, CD8+ T cell, T cell receptor

## DIACYLGLYCEROL KINASES IN T CELL RECEPTOR SIGNAL TRANSDUCTION

Diacylglycerol kinases (DGKs) represent a family of enzymes that catalyze phosphorylation of the membrane lipid sn-1,2 diacylglycerol (DAG) to form phosphatidic acid (PA) (Eichmann and Lass, 2015). In T cells, DAG is formed downstream of the T cell receptor (TCR) after activation of the gamma 1 isoform of phospholipase C (PLC $\gamma$ 1) and cleavage of phosphatidylinositol 4,5-bisphosphate (PIP<sub>2</sub>) into DAG and an additional second messenger, inositol 1,4,5-trisphosphate (IP<sub>3</sub>) (Krishna and Zhong, 2013). Whereas, IP<sub>3</sub> is important in facilitating release of calcium from the endoplasmic reticulum, DAG interacts with other proteins important in TCR signal transduction, such as Protein kinase C (predominantly  $\theta$  isoform in T cells, but also isoforms  $\epsilon$  and  $\eta$ ; Quann et al., 2011) and the Ras activating protein RasGRP1 (Krishna and Zhong, 2013). Biochemically, targeting the activity of DGKs in T cells, either by germline deletion, or with chemical inhibitors, results in enhanced and sustained signaling downstream of T cells, as assessed by prolonged phosphorylation of downstream molecules, such as extracellular signal-related kinases 1/2 (ERK1/2; Zhong et al., 2003; Olenchock et al., 2006; Riese et al., 2011). Although, three isoforms of DGK are known to be present within T cells (DGK $\alpha$ , DGK $\delta$ , and DGK $\zeta$ ), only two, DGK $\alpha$  and DGK $\zeta$ , are thought to play an important role in facilitating DAG metabolism downstream of the TCR (Joshi and Koretzky, 2013). The function of DGK $\delta$  is unknown in T cells; its role in facilitating metabolic flexibility between lipid and carbohydrate utilization suggest that it may regulate pools of DAG unrelated to TCR signaling (Chibalin et al., 2008). The signaling

changes resulting from the absence of DGK $\alpha$  or DGK $\zeta$  alter the transcriptional program of activated T cells. For instance, generation of the transcription factor AP1, which is dependent on Ras/ERK signaling, is decreased in stimulated Jurkat T cells overexpressing DGK $\zeta$  (Zhong et al., 2002). Similarly, NF- $\kappa$ B (nuclear factor kappa-light-chain-enhancer of activated B cells), a critical transcription factor activated downstream of PKC $\theta$  in T cells, is present at increased levels after stimulation of DGK $\zeta$ -deficient lymphocytes as compared to DGK $\zeta$ -replete cells (Schmidt et al., 2013), though other data suggests that the regulation of NF- $\kappa$ B may, in some instances, be positively regulated by DGKs (Yang et al., 2016).

The change in activation of transcription factors in stimulated T cells after manipulation of DGK activity correlates with changes in T cell activation markers and function. For instance, overexpression of DGK $\alpha$  or DGK $\zeta$  in Jurkat T cells results in decreased expression of the activation marker CD69 after stimulation through the TCR complex (Sanjuán et al., 2001; Zhong et al., 2002), and overexpression of DGK $\alpha$  induces a state of decreased functional activity resembling an anergy-like state (Zha et al., 2006). In contrast, deletion of DGK $\alpha$  or DGK $\zeta$  results in T cells with enhanced production of effector cytokines, such as IL2 and IFN $\gamma$ , and enhanced proliferation (Zhong et al., 2003; Olenchock et al., 2006). Inhibition of DGK $\alpha$  also allows T cells to overcome TCR signaling defects present in human X-linked lymphoproliferative disease (XLP-1) resulting from the loss of SAP [signaling lymphocytic activation molecule (SLAM)-associated protein (Baldanzi et al., 2011; Ruffo et al., 2016)], along with uncontrolled effector T cell expansion after exposure to Epstein Barr virus (EBV) characteristic of this disease (Ruffo et al., 2016).

## TARGETING DIACYLGLYCEROL KINASES TO ENHANCE T CELL ANTI-TUMOR ACTIVITY

Given the enhanced functional activity conferred by loss of DGKs in T cells, our group and others have tested the hypothesis that these proteins may serve as useful targets for enhancing T cell anti-tumor activity. Recently, strategies to target negative regulators of T cells to enhance their anti-tumor activity have been successfully translated from basic science studies into clinical care (Byrne et al., 2015; Sharma and Allison, 2015; Shin and Ribas, 2015; Callahan et al., 2016). Although, antibodies directed against CTLA-4 and PD-1 are the most prominent examples of therapies that have generated clinical responses in human malignancy, there is significant interest in identifying additional inhibitory regulators of T cells to combine with existing approaches and to use in instances where blockade of PD-1 and other immune checkpoints is ineffective (Restifo et al., 2016). We focused our studies on DGK $\zeta$ , since that enzyme appears to represent the dominant isoform in T cells, based on a direct comparison of TCR signal strength between T cells deficient in either DGK $\alpha$  or DGK $\zeta$  (Joshi et al., 2013). Using an EL4-ova subcutaneous model system to permit tracking of immune responses, we observed that DGK $\zeta$ <sup>-/-</sup> mice had an

increased frequency of tumor rejection, along with a trend toward increased number of tumor-specific CD8<sup>+</sup> T cells (Riese et al., 2011). Additionally, we demonstrated that adoptively transferred naïve (Riese et al., 2011) or activated (Riese et al., 2013) tumor-specific effector T cells displayed increased activation by tumor and resultant inhibition of tumor growth. While these studies relied on strong-antigen driven tumor models, it is likely that loss of DGKs also enhances T cell anti-tumor activity in tumors with low-grade antigens, since DAG-mediated activation of RasGRP1 regulates the threshold for T cell activation (Das et al., 2009), and earlier studies implicated a role for Ras in Jurkat T cell activation mediated by low grade TCR stimulus (Perez de Castro et al., 2004).

DGK $\alpha$  has also been evaluated as a potential target to improve T cell activity against tumor, based on the observation that DGK $\alpha$  is upregulated in certain inhibited T cell conditions, such as anergy (Zha et al., 2006), and that DGK $\alpha$  is upregulated in tumor-infiltrating lymphocytes in human renal cell carcinoma (Prinz et al., 2012). For instance, our own study using adoptive transfer of CAR (chimeric antigen receptor)-T cells demonstrated similar increases in efficacy (compared to wild type T cells) between T cells deficient in DGK $\alpha$  or DGK $\zeta$  in the treatment of murine mesothelioma (Riese et al., 2013). Additionally, a study testing the importance of DGK $\alpha$  in glioblastoma multiforme (GBM) cells found that concurrent administration of the relatively non-specific DGK $\alpha$  inhibitor R59022 resulted in decreased growth of intracranially injected GBM tumors. Although, a preponderance of evidence suggested that the decreased tumor growth in this model resulted from inhibition of DGK $\alpha$  within the tumor cells, modulation of immune activity was not assessed, and could have been contributory (Dominguez et al., 2013). DGKs also play a role in limiting the activity of NK cells isolated from tumors in patients with renal cell carcinoma, since the addition of either IL-2 or DGK inhibitor (R59022) to culture media improves the impaired function of tumor-associated NK cells (Prinz et al., 2014). Recently, a more specific inhibitor for DGK $\alpha$  has been developed that may be useful to extend these studies into additional tumor models (Liu et al., 2016). Although a direct comparison has not to-date been performed comparing tumor growth in DGK $\alpha$ <sup>-/-</sup> and DGK $\zeta$ <sup>-/-</sup> mice, future studies will undoubtedly provide additional comparisons between the two genotypes with respect to T cell anti-tumor immunity.

## MECHANISM OF ENHANCED T CELL ANTI-TUMOR ACTIVITY IN DGK $\zeta$ -DEFICIENT T CELLS

Enhanced anti-tumor activity observed in DGK $\zeta$ -deficient T cells was initially thought to result from increased cytokine production generated after TCR stimulation; however, it is clear that intrinsic insensitivity to inhibitory signals in the tumor microenvironment is also an important determinant. In broad terms, T cell inhibitory factors can be broadly separated into two groups: those that inhibit T cells by directly inhibiting proximal TCR signal transduction, and those that inhibit T cells independent of attenuation of TCR signaling. Examples

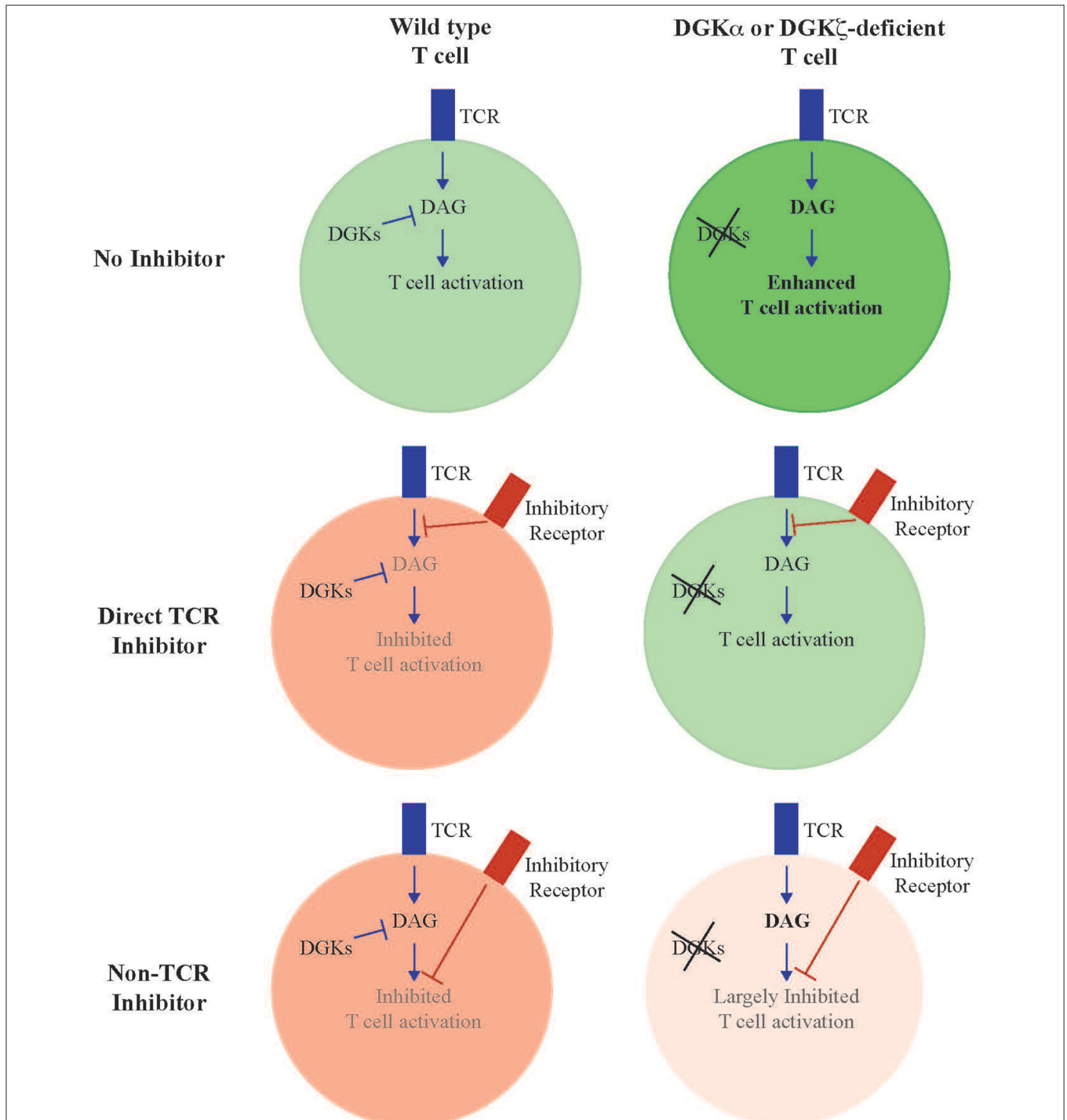
of immunosuppressant pathways that facilitate direct inhibition of TCR signaling include PD-1 (Chemnitz et al., 2004; Parry et al., 2005; Yokosuka et al., 2012), Lag3 (Okazaki et al., 2011), Prostaglandin E<sub>2</sub> (PGE<sub>2</sub>) (Wehbi and Taskén, 2016), adenosine 2A receptor (Linnemann et al., 2009; Linden and Cekic, 2012), and as we have recently identified, TGFβ (Arumugam et al., 2015; Newman et al., 2016). Whereas, PD-1 and TGFβ inhibit TCR signaling by directly or indirectly recruiting inhibitory tyrosine phosphatases, such as SHP-1 or SHP-2 (Src homology region 2 domain-containing phosphatase-1/2) to the cell surface, adenosine 2A, and PGE<sub>2</sub> receptors activate protein kinase A (PKA) leading to Csk-mediated inhibition of the proximal activating tyrosine kinase Lck (Newick et al., 2016). In contrast, Lag3 appears to oppose TCR signaling via a KIEELE motif that acts through an unclear mechanism (Freeman and Sharpe, 2012). In either case, these inhibitory factors act to oppose activation events facilitated by proximal tyrosine protein kinases, such as Lck or Zap-70, that are responsible for initiating TCR signal cascades and are upstream of DAG generation. In contrast are inhibitory factors that act predominantly by inhibiting T cells independent of TCR activation and DAG generation. These include a subset of immune checkpoint receptors, such as Tim3 (Jones et al., 2008; Lee et al., 2011), and CTLA-4. Although some reports have suggested direct proximal inhibition of TCR mediated by CTLA-4 (Lee et al., 1998), CTLA-4 more likely functions predominantly as a sink to sequester CD80 and CD86, the ligands of the co-stimulatory molecule CD28 (Green et al., 1994; Walunas et al., 1996; van der Merwe et al., 1997; Collins et al., 2002). T cells deficient in DGKs may demonstrate differential sensitivity to inhibition mediated by dependently or independently of TCR signaling (Figure 1, Table 1). For instance, a test of numerous T cell inhibitory pathways revealed that, in contrast to wild type T cells, T cells lacking DGKζ demonstrate reduced inhibition of IFNγ production in the presence of high concentrations of TGFβ, Prostaglandin E<sub>2</sub> (PGE<sub>2</sub>) or adenosine (Riese et al., 2013). This suggests that one can predict, in broad terms, how DGK-deficient T cells will respond to various immunosuppressive pathways within the tumor microenvironment, in that one would anticipate DGK-deficient T cells to be insensitive to inhibitors that directly attenuate TCR signaling, such as PD-1 or Lag3, but sensitive to inhibitory pathways that do not directly interfere with TCR signaling, such as Tim3 or CTLA-4. This model also predicts which immune targets might be optimally paired with agents that block DGKs, such that one would anticipate little synergy with direct regulators of TCR signaling, but potential excellent synergy with TCR-independent inhibitors. Experiments are currently ongoing to test these hypotheses.

## POTENTIAL ISSUES ARISING FROM TARGETING DGKS

The diverse set of cellular functions governed by DAG and PA, including signal transduction (Mérida et al., 2008), lipid biogenesis (Shulga et al., 2013), and membrane trafficking (Cho and Stahelin, 2005), presents challenges for broadly targeting

DGK activity therapeutically, especially since the enzymatic specificity of the 10 mammalian DGKs is highly conserved, with the exception of DGKε (Jennings et al., 2015). Thus, isolation of a compound with specificity toward an individual isoform may prove difficult, a problem exacerbated by the paucity of structural information available about eukaryotic DGKs. Although, a prokaryotic form of DGK has been solved structurally (Li et al., 2013), it is sufficiently divergent from eukaryotic isoforms that it provides little value in predicting active site topography of eukaryotic DGKs. For instance, prokaryotic DGKA is not limited to enzymatic activity against lipids, but can also catalyze reactions using glycerol and water as substrates for phosphorylation (Ullrich et al., 2011). Therefore, approaches to target non-catalytic domains of DGKζ may provide the best means to achieve isoform specificity. The structural domains of DGKζ have been well defined, and include a C1 domain, a MARCKS domain, an ankyrin repeat domain, and a C-terminal PDZ-binding domain (Joshi and Koretzky, 2013). Of these domains, only the MARCKS domain and ankyrin repeat domain are unique to DGKζ among the 10 DGK family members (Joshi and Koretzky, 2013). Furthermore, only the MARCKS domain, a substrate for serine/threonine phosphorylation by PKCα (Topham et al., 1998), is required for DGKζ function (Santos et al., 2002). Thus, targeting the MARCKS domain may be an effective strategy for therapeutic targeting of DGKζ, in a manner that confers specificity.

Apart from concerns with isotype specificity, therapeutic targeting of DGKα and DGKζ could result in deleterious “on-target” effects, such as enhanced cellular proliferation and autoimmunity. As expected, mice with deletions of both DGKα and DGKζ generate T cells with enhanced TCR signaling downstream of DAG, as well as more potent effector functions after *in vitro* stimulation relative to single knockouts or wild type T cells (Guo et al., 2008; Riese et al., 2013). However, the activated level of Ras/Erk signal transduction in double knockout mice (DKO) results in thymic lymphomagenesis (Guo et al., 2008). This pro-malignant potential of enhanced DAG signaling is consistent with data from human patients with T cell acute lymphoblastic leukemia (T-ALL), in which RasGRP1, the Ras activating protein activated by DAG, has been found to be frequently overexpressed (Hartzell et al., 2013). Thus, caution must be used if DGK germline deletion (e.g., using CRISPR-based approaches) is used as a means to target DGKs in adoptive T cell therapies. Generating auto-immune disease may also be an issue when targeting DGKs. Although mice deficient in DGKα, DGKζ, or both do not develop overt autoimmunity, it is likely that the mice would develop enhanced T cell responses in autoimmune models such as experimental autoimmune encephalitis (EAE), especially given the role DGKs play in limiting activation of Mnk1/2-mediated activation and development of encephalitis in the EAE model (Gorentla et al., 2013). A possible explanation for the lack of overt autoimmunity in DGKζ or DKO mice may be the concurrent enhanced generation of natural T regs in these mice (Joshi et al., 2013), although this remains speculative, and raises the additional consideration that thymic development of T cells may be impacted by manipulation of DGK activity. For instance, constitutive expression of a membrane-bound form of DGKα



**FIGURE 1 | Model of enhanced T cell activity in DGK-deficient T cells.** DGK $\alpha$  and DGK $\zeta$  function to metabolize DAG generated downstream of TCR activation. Deletion of either or both isoforms results in T cells with enhanced TCR signal transduction downstream of DAG and resultant enhanced effector functions ("No inhibitor"). Loss of DGK confers resistance to inhibitory pathways that function through direct interference of TCR signal transduction, since these inhibitory events occur proximal to generation of DAG ("Direct TCR Inhibitor"). We predict that T cells deficient in DGKs remain sensitive to inhibitory pathways that mediate their effect independent of interference with TCR signal transduction ("Non-TCR Inhibitor"), or for pathways that inhibit TCR signaling at rate limiting steps downstream of DAG formation.

**TABLE 1 | Relative insensitivity to inhibitory stimuli of CD8+ T cells.**

Inhibitory stimuli/receptor	Inhibitor-mediated fold ↓ in IFN $\gamma$ by CD8+ T cell		Does inhibitory pathway directly impact TCR signaling?	References
	WT	DGK $\zeta$ <sup>-/-</sup>		
TGF $\beta$	35x	1x	Yes	Arumugam et al., 2015
PGE <sub>2</sub>	2x	1.5x	Yes	Riese et al., 2013
Adenosine	2x	1.2x	Yes	Riese et al., 2013
PD-1	4x	Unknown	Yes	Parry et al., 2005
Lag3	2x	Unknown	Yes	Hannier et al., 1998; Huang et al., 2015
Tim-3	None	Unknown	No	Lee et al., 2011
CTLA-4	2x	Unknown	Uncertain	Krummel and Allison, 1995; Walunas and Bluestone, 1998

leads to accumulation of immature CD8+ “single positive” T cells within the thymus in addition to peripheral lymphopenia (Almena et al., 2013), and DKO mice demonstrate a severe impairment in thymic development of invariant NKT cells, a subset of innate T cells (Shen et al., 2011). Ongoing experiments are needed to test whether DGK-deficient mice will develop more severe disease than wild type counterparts after autoimmune challenges, and to determine whether subtle alterations are present in the thymic development of other conventional T cell subsets.

## CONCLUSIONS

As immunotherapeutic approaches come to the forefront of cancer treatment, there is an increased need to evaluate proteins and molecules that inhibit the immune system, especially in T cells. Diacylglycerol kinases should warrant a high degree of consideration in these targeting strategies, ideally with the development of small molecule inhibitors. In the meanwhile, it may be advantageous to move forward with gene manipulation in adoptive cellular therapies. The ability to very specifically target DGKs within only the transferred T cells will minimize systemic side effects. It will also be possible to include suicide genes to

enable destruction of the transferred T cells, should it become necessary. A better understanding of the changes that result from acute and long-term targeting of DGKs should help discern the effectiveness of this strategy both alone, and in combination with other therapies designed to induce immune cell activation.

## AUTHOR CONTRIBUTIONS

All authors contributed to the conception and design of the manuscript and contributed writing to the paper.

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## REFERENCES

- Almena, M., Andrada, E., Liebana, R., and Merida, I. (2013). Diacylglycerol metabolism attenuates T-cell receptor signaling and alters thymocyte differentiation. *Cell Death Dis.* 4, e912. doi: 10.1038/cddis.2013.396
- Arumugam, V., Bluemn, T., Wesley, E., Schmidt, A. M., Kambayashi, T., Malarkannan, S., et al. (2015). TCR signaling intensity controls CD8+ T cell responsiveness to TGF- $\beta$ . *J. Leukoc. Biol.* 98, 703–712. doi: 10.1189/jlb.2HIMA1214-578R
- Baldanzi, G., Pighini, A., Bettio, V., Rainero, E., Traini, S., Chianale, F., et al. (2011). SAP-mediated inhibition of diacylglycerol kinase  $\alpha$  regulates TCR-induced diacylglycerol signaling. *J. Immunol.* 187, 5941–5951. doi: 10.4049/jimmunol.1002476
- Byrne, K. T., Vonderheide, R. H., Jaffee, E. M., and Armstrong, T. D. (2015). Special conference on tumor immunology and immunotherapy: a new chapter. *Cancer Immunol. Res.* 3, 590–597. doi: 10.1158/2326-6066.CIR-15-0106
- Callahan, M. K., Postow, M. A., and Wolchok, J. D. (2016). Targeting T cell co-receptors for cancer therapy. *Immunity* 44, 1069–1078. doi: 10.1016/j.immuni.2016.04.023
- Chemnitz, J. M., Parry, R. V., Nichols, K. E., June, C. H., and Riley, J. L. (2004). SHP-1 and SHP-2 associate with immunoreceptor tyrosine-based switch motif of programmed death 1 upon primary human T cell stimulation, but only receptor ligation prevents T cell activation. *J. Immunol.* 173, 945–954.
- Chibalin, A. V., Leng, Y., Vieira, E., Krook, A., Björnholm, M., Long, Y. C., et al. (2008). Downregulation of diacylglycerol kinase delta contributes to hyperglycemia-induced insulin resistance. *Cell* 132, 375–386. doi: 10.1016/j.cell.2007.12.035
- Cho, W., and Stahelin, R. V. (2005). Membrane-protein interactions in cell signaling and membrane trafficking. *Annu. Rev. Biophys. Biomol. Struct.* 34, 119–151. doi: 10.1146/annurev.biophys.33.110502.133337
- Collins, A. V., Brodie, D. W., Gilbert, R. J. C., Iaboni, A., Manso-Sancho, R., Walse, B., et al. (2002). The interaction properties of costimulatory molecules revisited. *Immunity* 17, 201–210.

- Das, J., Ho, M., Zikherman, J., Govern, C., Yang, M., Weiss, A., et al. (2009). Digital signaling and hysteresis characterize ras activation in lymphoid cells. *Cell* 136, 337–351. doi: 10.1016/j.cell.2008.11.051
- Dominguez, C. L., Floyd, D. H., Xiao, A., Mullins, G. R., Kefas, B. A., Xin, W., et al. (2013). Diacylglycerol kinase  $\alpha$  is a critical signaling node and novel therapeutic target in glioblastoma and other cancers. *Cancer Discov.* 3, 782–797. doi: 10.1158/2159-8290.CD-12-0215
- Eichmann, T. O., and Lass, A. (2015). DAG tales: the multiple faces of diacylglycerol—stereochemistry, metabolism, and signaling. *Cell. Mol. Life Sci.* 72, 3931–3952. doi: 10.1007/s00018-015-1982-3
- Freeman, G. J., and Sharpe, A. H. (2012). A new therapeutic strategy for malaria: targeting T cell exhaustion. *Nat. Immunol.* 13, 113–115. doi: 10.1038/ni.2211
- Gorentla, B. K., Krishna, S., Shin, J., Inoue, M., Shinohara, M. L., Grayson, J. M., et al. (2013). Mnk1 and 2 are dispensable for T cell development and activation but important for the pathogenesis of experimental autoimmune encephalomyelitis. *J. Immunol.* 190, 1026–1037. doi: 10.4049/jimmunol.1200026
- Green, J. M., Noel, P. J., Sperling, A. I., Walunas, T. L., Gray, G. S., Bluestone, J. A., et al. (1994). Absence of B7-dependent responses in CD28-deficient mice. *Immunity* 1, 501–508.
- Guo, R., Wan, C.-K., Carpenter, J. H., Mousallem, T., Boustany, R.-M. N., Kuan, C.-T., et al. (2008). Synergistic control of T cell development and tumor suppression by diacylglycerol kinase alpha and zeta. *Proc. Natl. Acad. Sci. U.S.A.* 105, 11909–11914. doi: 10.1073/pnas.0711856105
- Hannier, S., Tournier, M., Bismuth, G., and Triebel, F. (1998). CD3/TCR complex-associated lymphocyte activation gene-3 molecules inhibit CD3/TCR signaling. *J. Immunol.* 161, 4058–4065.
- Hartzell, C., Ksionda, O., Lemmens, E., Coakley, K., Yang, M., Dail, M., et al. (2013). Dysregulated RasGRP1 responds to cytokine receptor input in T cell leukemogenesis. *Sci. Signal.* 6, ra21. doi: 10.1126/scisignal.2003848
- Huang, R. Y., Eppolito, C., Lele, S., Shrikant, P., Matsuzaki, J., and Odunsi, K. (2015). LAG3 and PD1 co-inhibitory molecules collaborate to limit CD8+ T cell signaling and dampen antitumor immunity in a murine ovarian cancer model. *Oncotarget* 6, 27359–27377. doi: 10.18632/oncotarget.4751
- Jennings, W., Doshi, S., D'Souza, K., and Epan, R. M. (2015). Molecular properties of diacylglycerol kinase-epsilon in relation to function. *Chem. Phys. Lipids* 192, 100–108. doi: 10.1016/j.chemphyslip.2015.06.003
- Jones, R. B., Ndhlovu, L. C., Barbour, J. D., Sheth, P. M., Jha, A. R., Long, B. R., et al. (2008). Tim-3 expression defines a novel population of dysfunctional T cells with highly elevated frequencies in progressive HIV-1 infection. *J. Exp. Med.* 205, 2763–2779. doi: 10.1084/jem.20081398
- Joshi, R. P., and Koretzky, G. A. (2013). Diacylglycerol kinases: regulated controllers of T cell activation, function, and development. *Int. J. Mol. Sci.* 14, 6649–6673. doi: 10.3390/ijms14046649
- Joshi, R. P., Schmidt, A. M., Das, J., Pytel, D., Riese, M. J., Lester, M., et al. (2013). The  $\zeta$  isoform of diacylglycerol kinase plays a predominant role in regulatory T cell development and TCR-mediated ras signaling. *Sci. Signal.* 6, ra102. doi: 10.1126/scisignal.2004373
- Krishna, S., and Zhong, X. (2013). Role of diacylglycerol kinases in T cell development and function. *Crit. Rev. Immunol.* 33, 97–118.
- Krummel, M. F., and Allison, J. P. (1995). CD28 and CTLA-4 have opposing effects on the response of T cells to stimulation. *J. Exp. Med.* 182, 459–465.
- Lee, J., Su, E. W., Zhu, C., Hainline, S., Phuah, J., Moroco, J. A., et al. (2011). Phosphotyrosine-dependent coupling of Tim-3 to T-cell receptor signaling pathways. *Mol. Cell. Biol.* 31, 3963–3974. doi: 10.1128/MCB.05297-11
- Lee, K. M., Chuang, E., Griffin, M., Khattri, R., Hong, D. K., Zhang, W., et al. (1998). Molecular basis of T cell inactivation by CTLA-4. *Science* 282, 2263–2266.
- Li, D., Lyons, J. A., Pye, V. E., Vogeley, L., Aragão, D., Kenyon, C. P., et al. (2013). Crystal structure of the integral membrane diacylglycerol kinase. *Nature* 497, 521–524. doi: 10.1038/nature12179
- Linden, J., and Cekic, C. (2012). Regulation of lymphocyte function by adenosine. *Arterioscler. Thromb. Vasc. Biol.* 32, 2097–2103. doi: 10.1161/ATVBAHA.111.226837
- Linnemann, C., Schildberg, F. A., Schurich, A., Diehl, L., Hegenbarth, S. I., Endl, E., et al. (2009). Adenosine regulates CD8 T-cell priming by inhibition of membrane-proximal T-cell receptor signalling. *Immunology* 128, e728–e737. doi: 10.1111/j.1365-2567.2009.03075.x
- Liu, K., Kunii, N., Sakuma, M., Yamaki, A., Mizuno, S., Sato, M., et al. (2016). A novel diacylglycerol kinase  $\alpha$ -selective inhibitor, CU-3, induces cancer cell apoptosis and enhances immune response. *J. Lipid Res.* 57, 368–379. doi: 10.1194/jlr.M062794
- Mérida, I., Avila-Flores, A., and Merino, E. (2008). Diacylglycerol kinases: at the hub of cell signalling. *Biochem. J.* 409, 1–18. doi: 10.1042/BJ20071040
- Newick, K., O'Brien, S., Sun, J., Kapoor, V., Maceyko, S., Lo, A., et al. (2016). Augmentation of CAR T-cell trafficking and anti-tumor efficacy by blocking protein kinase A. *Localization* 4, 541–551. doi: 10.1158/2326-6066.CIR-15-0263
- Newman, D. K., Fu, G., Adams, T., Arumugam, V., Bluemn, T., and Riese, M. J. (2016). An unexpected role for PECAM-1 in TGF beta-mediated inhibition of T cell function. *Sci. Signal.* 9, 1–12.
- Okazaki, T., Okazaki, I.-M., Wang, J., Sugiura, D., Nakaki, F., Yoshida, T., et al. (2011). PD-1 and LAG-3 inhibitory co-receptors act synergistically to prevent autoimmunity in mice. *J. Exp. Med.* 208, 395–407. doi: 10.1084/jem.20100466
- Olenchock, B. A., Guo, R., Carpenter, J. H., Jordan, M., Topham, M. K., Koretzky, G. A., et al. (2006). Disruption of diacylglycerol metabolism impairs the induction of T cell anergy. *Nat. Immunol.* 7, 1174–1181. doi: 10.1038/ni1400
- Parry, R. V., Chemnitz, J. M., Frauwirth, K. A., Lanfranco, A. R., Braunstein, I., Kobayashi, S. V., et al. (2005). CTLA-4 and PD-1 receptors inhibit T-cell activation by distinct mechanisms. *Mol. Cell. Biol.* 25, 9543–9553. doi: 10.1128/MCB.25.21.9543-9553.2005
- Perez de Castro, I., Bivona, T. G., Philips, M. R., and Pellicer, A. (2004). Ras activation in Jurkat T cells following low-grade stimulation of the T-cell receptor is specific to N-Ras and occurs only on the Golgi apparatus. *Mol. Cell. Biol.* 24, 3485–3496.
- Prinz, P. U., Mandler, A. N., Brech, D., Masouris, I., Oberneder, R., and Noessner, E. (2014). NK-cell dysfunction in human renal carcinoma reveals diacylglycerol kinase as key regulator and target for therapeutic intervention. *Int. J. Cancer* 135, 1832–1841. doi: 10.1002/ijc.28837
- Prinz, P. U., Mandler, A. N., Masouris, I., Durner, L., Oberneder, R., and Noessner, E. (2012). High DGK- $\alpha$  and disabled MAPK pathways cause dysfunction of human tumor-infiltrating CD8+ T cells that is reversible by pharmacologic intervention. *J. Immunol.* 188, 5990–6000. doi: 10.4049/jimmunol.1103028
- Quann, E. J., Liu, X., Altan-Bonnet, G., and Huse, M. (2011). A cascade of protein kinase C isozymes promotes cytoskeletal polarization in T cells. *Nat. Immunol.* 12, 647–654. doi: 10.1038/ni.2033
- Restifo, N. P., Smyth, M. J., and Snyder, A. (2016). Acquired resistance to immunotherapy and future challenges. *Nat. Rev. Cancer* 16, 121–126. doi: 10.1038/nrc.2016.2
- Riese, M. J., Grewal, J., Das, J., Zou, T., Patil, V., Chakraborty, A. K., et al. (2011). Decreased diacylglycerol metabolism enhances ERK activation and augments CD8+ T cell functional responses. *J. Biol. Chem.* 286, 5254–5265. doi: 10.1074/jbc.M110.171884
- Riese, M. J., Wang, L.-C. S., Moon, E. K., Joshi, R. P., Ranganathan, A., June, C. H., et al. (2013). Enhanced effector responses in activated CD8+ T cells deficient in diacylglycerol kinases. *Cancer Res.* 73, 3566–3577. doi: 10.1158/0008-5472.CAN-12-3874
- Ruffo, E., Malacarne, V., Larsen, S. E., Das, R., Patrussi, L., Wülfing, C., et al. (2016). Inhibition of diacylglycerol kinase  $\alpha$  restores restimulation-induced cell death and reduces immunopathology in XLP-1. *Sci. Transl. Med.* 8, 321ra7. doi: 10.1126/scitranslmed.aad1565
- Sanjuán, M. A., Jones, D. R., Izquierdo, M., and Mérida, I. (2001). Role of diacylglycerol kinase alpha in the attenuation of receptor signaling. *J. Cell Biol.* 153, 207–220.
- Santos, T., Carrasco, S., Jones, D. R., Mérida, I., and Eguinoa, A. (2002). Dynamics of diacylglycerol kinase zeta translocation in living T-cells. Study of the structural domain requirements for translocation and activity. *J. Biol. Chem.* 277, 30300–30309. doi: 10.1074/jbc.M200999200
- Schmidt, A. M., Zou, T., Joshi, R. P., Lechner, T. M., Pimentel, M. A., Sommers, C. L., et al. (2013). Diacylglycerol kinase  $\zeta$  limits the generation of natural regulatory T cells. *Sci. Signal.* 6, ra101. doi: 10.1126/scisignal.2004411
- Sharma, P., and Allison, J. P. (2015). The future of immune checkpoint therapy. *Science* 348, 56–61. doi: 10.1126/science.aaa8172
- Shen, S., Wu, J., Srivatsan, S., Gorentla, B. K., Shin, J., Xu, L., et al. (2011). Tight regulation of diacylglycerol-mediated signaling is critical for proper invariant NKT cell development. *J. Immunol.* 187, 2122–2129. doi: 10.4049/jimmunol.1100495

- Shin, D. S., and Ribas, A. (2015). The evolution of checkpoint blockade as a cancer therapy: what's here, what's next? *Curr. Opin. Immunol.* 33, 23–35. doi: 10.1016/j.coi.2015.01.006
- Shulga, Y. V., Loukov, D., Ivanova, P. T., Milne, S. B., Myers, D. S., Hatch, G. M., et al. (2013). Diacylglycerol kinase delta promotes lipogenesis. *Biochemistry* 52, 7766–7776. doi: 10.1021/bi401178y
- Topham, M. K., Bunting, M., Zimmerman, G. A., McIntyre, T. M., Blackshear, P. J., and Prescott, S. M. (1998). Protein kinase C regulates the nuclear localization of diacylglycerol kinase-zeta. *Nature* 394, 697–700. doi: 10.1038/29337
- Ullrich, S. J., Hellmich, U. A., Ullrich, S., and Glaubitz, C. (2011). Interfacial enzyme kinetics of a membrane bound kinase analyzed by real-time MAS-NMR. *Nat. Chem. Biol.* 7, 263–270. doi: 10.1038/nchembio.543
- van der Merwe, P. A., Bodian, D. L., Daenke, S., Linsley, P., and Davis, S. J. (1997). CD80 (B7-1) binds both CD28 and CTLA-4 with a low affinity and very fast kinetics. *J. Exp. Med.* 185, 393–403.
- Walunas, T. L., Bakker, C. Y., and Bluestone, J. A. (1996). CTLA-4 ligation blocks CD28-dependent T cell activation. *J. Exp. Med.* 183, 2541–2550.
- Walunas, T. L., and Bluestone, J. A. (1998). CTLA-4 regulates tolerance induction and T cell differentiation *in vivo*. *J. Immunol.* 187, 3466–3474.
- Wehbi, V. L., and Taskén, K. (2016). Molecular mechanisms for cAMP-mediated immunoregulation in T cells - role of anchored protein Kinase A signaling units. *Front. Immunol.* 7:222. doi: 10.3389/fimmu.2016.00222
- Yang, J., Zhang, P., Krishna, S., Wang, J., Lin, X., Huang, H., et al. (2016). Unexpected positive control of NF $\kappa$ B and miR-155 by DGK $\alpha$  and  $\zeta$  ensures effector and memory CD8+ T cell differentiation. *Oncotarget.* 7, 33744–33764. doi: 10.18632/oncotarget.8164
- Yokosuka, T., Takamatsu, M., Kobayashi-Imanishi, W., Hashimoto-Tane, A., Azuma, M., and Saito, T. (2012). Programmed cell death 1 forms negative costimulatory microclusters that directly inhibit T cell receptor signaling by recruiting phosphatase SHP2. *J. Exp. Med.* 209, 1201–1217. doi: 10.1084/jem.20112741
- Zha, Y., Marks, R., Ho, A. W., Peterson, A. C., Janardhan, S., Brown, I., et al. (2006). T cell anergy is reversed by active Ras and is regulated by diacylglycerol kinase-alpha. *Nat. Immunol.* 7, 1166–1173. doi: 10.1038/ni1394
- Zhong, X.-P., Hainey, E. A., Olenchock, B. A., Jordan, M. S., Maltzman, J. S., Nichols, K. E., et al. (2003). Enhanced T cell responses due to diacylglycerol kinase zeta deficiency. *Nat. Immunol.* 4, 882–890. doi: 10.1038/ni958
- Zhong, X.-P., Hainey, E. A., Olenchock, B. A., Zhao, H., Topham, M. K., and Koretzky, G. A. (2002). Regulation of T cell receptor-induced activation of the Ras-ERK pathway by diacylglycerol kinase zeta. *J. Biol. Chem.* 277, 31089–31098. doi: 10.1074/jbc.M203818200

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

MR and SA are co-holders of a pending patent application on targeting DGKs for adoptive cellular therapies.

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