Check for updates

### **OPEN ACCESS**

EDITED BY Qing K. Wang, Huazhong University of Science and Technology, China

REVIEWED BY

Fan Wang, St. Jude Children's Research Hospital, United States Tommaso Bucci, University of Liverpool, United Kingdom

\*CORRESPONDENCE Bohang Chen Inutcmcbh@163.com Wenjie Li Iwzhwh@163.com

RECEIVED 12 February 2024 ACCEPTED 26 March 2024 PUBLISHED 09 April 2024

### CITATION

Chen B, Wang C and Li W (2024) Serum albumin levels and risk of atrial fibrillation: a Mendelian randomization study. Front. Cardiovasc. Med. 11:1385223. doi: 10.3389/fcvm.2024.1385223

#### COPYRIGHT

© 2024 Chen, Wang and Li. This is an openaccess article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# Serum albumin levels and risk of atrial fibrillation: a Mendelian randomization study

### Bohang Chen<sup>1\*</sup>, Chuqiao Wang<sup>1</sup> and Wenjie Li<sup>2\*</sup>

<sup>1</sup>The First Clinical Medical College, Liaoning University of Traditional Chinese Medicine, Shenyang, Liaoning, China, <sup>2</sup>Department of Cardiovascular Medicine, Affiliated Hospital of Liaoning University of Traditional Chinese Medicine, Shenyang, Liaoning, China

**Objective:** Although several observational studies have linked serum albumin to cardiovascular disease and considered it as an important biomarker, little is known about whether increasing or maintaining serum albumin levels can effectively improve the prognosis of patients with atrial fibrillation. Therefore, this study aims to further explore the causal relationship between serum albumin and atrial fibrillation and its potential mechanism.

**Method:** Using data from large-scale genome-wide association studies, we conducted a two-sample Mendelian randomization (MR) analysis and a mediation MR analysis, using serum albumin as the exposure variable and atrial fibrillation as the outcome variable. We included 486 serum metabolites as potential mediating factors. To increase the robustness of the analysis, we applied five statistical methods, including inverse variance weighted, weighted median, MR-Egger, simple mode, and weighted mode. Validate the MR results using Bayesian weighted Mendelian randomization method.

**Result:** The results of the MR analysis indicate a significant inverse association between genetically predicted serum albumin concentration (g/L) and the risk of atrial fibrillation (Beta = -0.172, OR = 0.842, 95% CI: 0.753-0.941, p = 0.002). Further mediation MR analysis revealed that serum albumin may mediate the causal relationship with atrial fibrillation by affecting two serum metabolites, docosatrienoate and oleate/vaccenate, and the mediating effect was significant. In addition, all our instrumental variables showed no heterogeneity and level-multiplicity in the MR analysis. To verify the stability of the results, we also conducted a sensitivity analysis using the leave-one-out method, and the results further confirmed that our findings were robust and reliable. Finally, we conducted a validation using the Bayesian weighted Mendelian randomization method, which demonstrated the reliability of our causal inference results.

**Conclusion:** This study strongly demonstrates the causal relationship between serum albumin and reduced risk of atrial fibrillation through genetic methods, and reveals the key mediating role of two serum metabolites in this relationship. These findings not only provide a new perspective for our understanding of the role of serum albumin in atrial fibrillation, but also provide new ideas for the prevention and treatment strategies of atrial fibrillation.

### KEYWORDS

serum albumin, atrial fibrillation, causal relationship, Mendelian randomization study, mediating effect, serum metabolites

### 1 Introduction

Cardiovascular disease has become the leading cause of death worldwide. Epidemiological factors indicate that low serum albumin levels are independently associated with a variety of cardiovascular diseases, including coronary artery disease, heart failure, and atrial fibrillation, and have been established as effective and independent prognostic indicators for patients with cardiovascular disease (1). Through multiple long-term and large-scale cohort studies, a significant trend has emerged: a decrease in serum albumin levels is significantly correlated with a relative increase in the incidence of atrial fibrillation (2, 3). A comprehensive meta-analysis involving 9 trials and 32,130 subjects further strengthened this view. The analysis showed that compared to patients with low serum albumin levels, patients with high serum albumin levels had a significantly reduced risk of developing atrial fibrillation (OR: 0.62, 95% CI: 0.44–0.89, P = 0.009). Specifically, every 10 g/L increase in serum albumin level was associated with a 36% reduction in the relative risk (95% CI: 0.51–0.81,  $I^2 = 87\%$ , P < 0.001) (4). However, although observational studies have shown consistency in revealing the negative correlation between serum albumin levels and atrial fibrillation, there is still uncertainty regarding whether preventing and correcting low serum albumin has actual benefits for the prognosis or patients at risk of atrial fibrillation. In other words, the potential causal relationship between the two has not been clearly confirmed, which remains a focus of current controversy. In view of this, we urgently need to conduct more in-depth research to clarify the causal relationship between serum albumin levels and atrial fibrillation and its underlying mechanisms. This will provide important theoretical basis and practical guidance for future cardiovascular disease prevention, treatment, and rehabilitation. At the same time, it will also help us to more fully understand the important role of serum albumin in cardiovascular disease, laying a solid foundation for the development of new treatment strategies and methods.

Mendelian randomization (MR) is an effective method for evaluating the causal relationship between observable variable exposures or risk factors and clinical outcomes (5). In particular, when randomized controlled trials are limited in verifying causal relationships, and observational studies exhibit biased associations due to potential confounding variables or reverse causality, this genetic analysis method highlights its importance (6). In this study, we systematically collected published data and employed a two-sample Mendelian randomization analysis method to identify the causal relationship between serum albumin level and atrial fibrillation through rigorous data processing and analysis procedures, aiming to reveal the true and unbiased association between the two.

### 2 Materials and methods

### 2.1 Source of data

We collected the genome-wide association study (GWAS) dataset related to serum albumin level and atrial fibrillation by

visiting the website (https://gwas.mrcieu.ac.uk). Among them, the GWAS dataset for serum albumin levels (N = 400,938) was derived from a study utilizing exon-wide imputation, association, and fine-mapping (7). This study accurately inferred exon-wide variations by leveraging extensive haplotype sharing between 49,960 UK Biobank participants with exome sequencing and the rest of the cohort. The aim was to determine the causal effects of rare coding variants on 54 quantitative traits. The GWAS dataset for atrial fibrillation was obtained from a study that aggregated a large-scale individual data from the Atrial Fibrillation Genetics (AFGen) Consortium (Ncase = 15,979, Ncontrol = 102,776) (8). Through a comprehensive and meticulous meta-analysis of 33 studies, including GWAS, Exome-Wide Association Studies (ExWAS), and Rare Variant Association Studies (RVAS), a highquality GWAS dataset tailored for atrial fibrillation was constructed. This dataset not only encompasses common variants but also delves into the impact of rare variants on atrial fibrillation, further elucidating the genetic basis of the condition.

In addition, we also obtained a serum metabolite dataset from a large-scale GWAS study (ID range: GCST90199621-GCST90201020). This study comprehensively examined two European population-based surveys involving a total of 7,824 participants. With the help of advanced technology and strict quality control processes, the study explored the genetic basis of more than 400 blood metabolites, and ultimately identified 486 metabolites at the genetic level for our subsequent analysis (9). Among these metabolites, 309 are known and 177 are unknown. In order to more systematically understand these known metabolites, we summarized them into eight major metabolic pathways, each involving different biochemical processes (10).

In this analysis, no sample overlap was found among the three GWAS datasets we used. The above datasets provide us with rich genetic information and biochemical background, which helps us to explore the association between serum albumin and atrial fibrillation and its potential genetic and metabolic mechanisms in depth.

### 2.2 Research method

In this study, we used R4.2.2 statistical software combined with professional R packages such as TwoSampleMR and MR-PRESSO to conduct systematic MR analysis to further explore the possible causal relationship between serum albumin and atrial fibrillation. To ensure the accuracy and robustness of the research results, we comprehensively applied five advanced statistical methods, including inverse variance weighted, weighted median, MR-Egger, simple mode, and weighted mode methods. Among these methods, the inverse variance weighted (IVW) method became the main method for our MR analysis due to its efficient and stable characteristics.

Bayesian weighted Mendelian randomization is a statistical method that combines Bayesian inference with MR (11). It utilizes MR to eliminate the influence of confounding factors and employs Bayesian inference to integrate prior information with observed data, obtaining a posterior distribution for more accurate estimation of causal relationships. Specifically, this approach can weight the effect sizes of exposure factors based on prior information and sample data, ensuring that exposure factors with larger effects have a greater influence in the analysis. This enhances the comprehensiveness and reliability of our analytical results.

### 2.3 Instrumental variable assumption

In the process of using genetic variation to assess causal effects, selecting SNPs as instrumental variables is crucial. We must strictly follow three basic assumptions to ensure the accuracy and reliability of the study. First, the selected SNPs must be significantly correlated with the exposure factor; second, the association between the SNP and the outcome must be independent of any confounding factors; finally, the SNP can only be linked to the clinical outcome through the exposure factor (12).

To avoid bias due to linkage disequilibrium, we set strict screening criteria. Specifically, SNPs significantly associated with exposure factors, in the presence of linkage disequilibrium, need to satisfy an  $r^2$  value of less than 0.001 and a genetic distance of 10,000 kb. To ensure the robustness of the selected instrumental variables, we also conducted further testing using the F statistic. Only when the F value is greater than 10 can we be confident that there is no weak instrumental variable bias (13).

When screening SNPs associated with serum metabolites, we adopted a relatively loose statistical threshold (P < 1e-05). This strategy aims to ensure that as many exposure variants as possible can be captured even with a small number of significant SNPs. However, we did not lower the quality requirements for instrumental variables. Instead, we still adhere to strict linkage disequilibrium conditions ( $r^2$  value less than 0.01, genetic distance not exceeding 500 kb) and the F > 10 criterion for eliminating weak instrumental variables.

After a series of rigorous screening processes, we successfully extracted instrumental variables that met all conditions and included them in the scope of the study. At the same time, we also recorded detailed information about each SNP's effect allele (EA), allele effect value ( $\beta$ ), standard error (SE), and *P* value (14, 15).

### 2.4 Statistical method

We conducted detailed data analysis using statistical software version R4.2.2 and its dedicated R packages (TwoSampleMR and MR-PRESSO). To ensure the accuracy and consistency of the data, we carefully summarized the statistical information of the exposure factor and clinical outcome datasets, and checked the allelic correspondence of each SNP on the effect of exposure factors and clinical outcomes one by one. The IVW method, as the core analysis method, provides a comprehensive weighted assessment of potential causal effects by integrating the MR effect estimates of each SNP (16). This method is most reliable when there is no level heterogeneity in instrumental variables (17). At the same time, the weighted mode method can robustly estimate causal effects even when up to 50% of the information comes from genetic variants with invalid instrumental variables (18).

To test whether there is horizontal multilevel effect in instrumental variables, we used the MR-Egger regression method, which effectively estimates the impact of horizontal multilevel effect through the intercept term (19). To further detect and correct horizontal multilevel effect, we also used the MR-PRESSO test to achieve this goal by removing outliers (20). In assessing heterogeneity of instrumental variables, we used Cochran's *Q*-test for quantitative analysis. When the *P* value is less than 0.05, we determined that there is significant heterogeneity, and at this time, we will choose the IVW random effect model to estimate causal effects to ensure the robustness of the results. Throughout the analysis process, we set *P* < 0.05 as the threshold to determine whether the results have statistical significance.

### 3 MR result

# 3.1 Causal relationship between serum albumin level and atrial fibrillation

We set serum albumin levels as the exposure factor, atrial fibrillation as the study outcome, and set strict statistical significance thresholds (P < 5e-08). After screening, we have preliminarily identified 208 SNPs associated with serum albumin levels. However, based on our hypothesis of linkage disequilibrium, we further excluded 7 SNPs. These 7 SNPs are rs10743939, rs11871801, rs17151639, rs2696671, rs3744274, rs7797854 and rs930734. Next, to avoid the issue of weak instrument variable bias, we set a screening condition with an F-test value greater than 10. After this round of screening, we further eliminated 46 SNPs. These 36 SNPs are rs12209602, rs12453576, rs12516449, rs1675382, rs1782455, rs1809423, rs1973878, rs2228213, rs2301029, rs2736231, rs27744, rs28925904, rs3213868, rs33994242, rs3756772, rs3087243. rs4730221, rs55910200, rs57826934, rs61735533, rs62622830, rs648514, rs6684353, rs67611724, rs6873349, rs7031621, rs704017, rs7153110, rs725660, rs74010640, rs74397112, rs7443182, rs75009793, rs755731, rs76701589, rs7726159, rs8045125, rs8187658, rs843925, rs848486, rs869337, rs870526, rs921071, rs921835, rs9309325 and rs9772460. To ensure the purity of instrumental variables, that is, the association between SNP and outcome is not affected by confounding factors, we carefully searched the PhenoScanner database. The results showed that 36 SNPs were associated with thyroid disease, diabetes, hypertension, coronary artery disease and other potential confounding factors. These confounding factors may interfere with our research results, so we have decided to exclude these SNPs from the analysis. These 36 SNPs are rs4409785, rs1133400, rs6897932, rs2586886, rs460879, rs12243326, rs6486122, rs6861681, rs7586970, rs4804416, rs1076540, rs1378942, rs1229984, rs10468017, rs10828249, rs10517086, rs2923089, rs58148580, rs2972143, rs6935921, rs459193, rs646776, rs9088, rs4540292, rs2000999, rs2229357, rs653178, rs731839,

rs1128249, rs13107325, rs17036326, rs1122326, rs6831256, rs9987289, rs1260326 and rs28929474. Before conducting the MR analysis, we performed quality control on the 119 SNPs obtained through the aforementioned screening process, filtering out those with ambiguous base calls. Ambiguous base calls refer to situations where the specific base at a particular position cannot be unambiguously determined, which may result from sequencing errors, sample contamination, or other factors. The presence of ambiguous base calls can introduce bias into our analysis, affecting the accurate inference of genotypes and subsequently impacting the estimation of causal effects as well as the statistical power of the analysis. If ambiguous base calls are present, we will remove them to ensure the accuracy and reliability of our analysis results. We didn't find any ambiguous base calls, so no SNPs were removed in this step. Finally, we identified 119 robust SNPs as instrumental variables for subsequent MR analysis. Through this series of screenings for SNPs, we have ensured that our analysis has adequate statistical power.Detailed information of SNP is shown in Supplementary Table S1.

After a series of rigorous MR analyses, we consistently found a significant causal relationship between the increase in serum albumin levels and the reduction in the risk of atrial fibrillation. This conclusion was supported by five different MR analysis methods, all of which had an odds ratio (OR) less than 1, strongly supporting our research hypothesis. It is particularly noteworthy that we used the IVW method as the primary analysis method. The IVW method has high statistical power and can more accurately reveal potential causal relationships. The analysis results showed that there was a significant association between the increase in serum albumin levels and the reduction in the risk of atrial fibrillation (IVW: Beta = -0.172, OR = 0.842, 95% CI: 0.753-0.941, P = 0.002).

To further verify the consistency and reliability of the instrumental variables, we used IVW and MR-Egger regression methods to detect heterogeneity among instrumental variables. The results of both methods indicate that there is no significant heterogeneity among instrumental variables. In addition, we also confirmed the absence of horizontal multi-effect interference in instrumental variables using the MR-PRESSO test. To further verify the stability of the MR results, we conducted a sensitivity analysis using the leave-one-out method. By removing SNPs one by one and comparing the causal effects of the remaining SNPs with the MR analysis results of all SNPs, we found that the MR results remained highly stable. This indicates that the causal relationship between serum albumin and reduced risk of atrial fibrillation is not dominated by a single or small number of instrumental variables, but is based on comprehensive data analysis and reliable conclusions (Figure 1). The results of MR analysis are shown in Supplementary Table S2.

### 3.2 Causal relationship between serum metabolite levels and atrial fibrillation

We systematically explored the potential causal relationship between 486 serum metabolites and atrial fibrillation. To ensure the accuracy and reliability of the results, we set a screening criterion of P value less than 1e-05 to screen out SNPs significantly associated with each metabolite. At the same time, the condition of F value greater than 10 eliminated weak instrumental variables, ensuring the validity of the analysis.

In the process of in-depth analysis, we used five different analytical methods and required that the OR direction obtained by these methods must be consistent to enhance the confidence of the results. After a series of MR analyses, we found that a total of 26 serum metabolites had significant causal effects on atrial fibrillation. Specifically, kynurenine levels, pro-hydroxypro levels, docosatrienoate (22:3n3) levels, octanoylcarnitine (c8) levels, 5,6-dihydrouridine levels, tetradecanedioate (C14-DC) levels, cis-4-decenoylcarnitine (C10:1) levels, guaiacol levels, octadecanedioylcarnitine (C18-DC) levels, sulfate N-acetylglucosamine/n-acetylgalactosamine levels, trans 3,4methyleneheptanoate levels, taurodeoxycholic acid 3-sulfate levels, glycodeoxycholate 3-sulfate levels, oleate/vaccenate (18:1) levels, 1-linoleoyl-GPG (18:2) levels, 1-(1-enyl-palmitoyl)-2palmitoyl-GPC (P-16:0/16:0) levels, 1-linoleoyl-2-linolenoyl-GPC (18:2/18:3) levels had significant negative correlation with atrial fibrillation, which means they may help reduce the risk of atrial fibrillation. Homocitrulline levels, threonate levels, gamma-glutamylvaline 1-methyl-4-imidazoleacetate levels, levels, dimethylarginine (sdma + adma) levels, N-acetyl-3methylhistidine levels, sphingomyelin (d18:1/17:0, d17:1/18:0, d19:1/16:0) levels and 2-naphthol sulfate levels showed significant positive correlation with atrial fibrillation and may be potential risk factors for atrial fibrillation. To verify the robustness of this result, we conducted sensitivity analysis on these 26 serum metabolites. The sensitivity analysis results support our conclusion that the results are stable. Figure 2 shows us a forest plot of serum metabolites with significant causal relationships with atrial fibrillation. The results of MR analysis are presented in Supplementary Table S3.

# 3.3 Causal relationship between serum albumin level and related serum metabolites

We further analyzed the exposure factor of serum albumin and selected 26 serum metabolites previously identified as having genetic causal relationships with atrial fibrillation as outcome variables for in-depth analysis. To ensure the rigor of the results, we set strict statistical significance thresholds (P < 5e-08) and instrumental variable screening criteria (F value > 10) to eliminate potential weak instrumental variables. At the same time, we used five different Mendelian randomization analysis methods, and required that the direction of the ORs obtained by these methods be consistent to enhance the confidence of the results.

When conducting mediation analysis with Mendelian randomization, we pay special attention to the independence of the genetic pathways between exposure factors and mediating variables, as well as between mediating variables and outcome



variables. After careful screening, we confirmed that there are no identical SNPs in these two pathways that can be used as instrumental variables, thus satisfying the necessary conditions for conducting mediation analysis.

Through analysis using the IVW method, we revealed significant causal relationships between serum albumin and two specific serum metabolites. Specifically, there was a positive causal relationship between serum albumin and Docosatrienoate (22:3n3) levels (OR = 1.117, 95% CI: 1.001–1.247, p = 0.046), and similarly, a significant positive causal relationship was also

observed between serum albumin and Oleate/vaccenate (18:1) levels (OR = 1.155, 95% CI: 1.051–1.271, p = 0.002). The results of MR analysis are shown in Supplementary Tables S4, S5.

Combining our previous findings on the causal relationship between serum albumin levels and atrial fibrillation and serum metabolites and atrial fibrillation, we have reason to speculate that these two serum metabolites may play an important mediating role between serum albumin and atrial fibrillation (Figures 3, 4). The results of MR analysis are shown in Supplementary Tables S6, S7.

05

id.exposure	nsnp	method	pval	OR(95% CI)
GCST90199636	18	Weighted median	0.097	0.922 (0.838 to 1.015)
	18	Inverse variance weighted	0.044	0.932 (0.870 to 0.998)
	18	Weighted mode	0.279	0.937 (0.837 to 1.050)
GCST90199682 GCST90199687	23	Weighted median	0.027	1.110 (1.012 to 1.218)
	23	Inverse variance weighted	0.016 🛏 🛏	1.083 (1.015 to 1.155)
	23	Weighted mode	0.026	1.120 (1.020 to 1.229)
	18	Weighted median	0.026	1.158 (1.018 to 1.317)
	18	Inverse variance weighted	0.045	1.102 (1.002 to 1.212)
	18	Weighted mode	0.092	→ 1 177 (0 984 to 1 408)
GCST00100703	21	Weighted median	0.431	1 056 (0.922 to 1.211)
	21	Inverse verience weighted	0.047	1.112 (1.002 to 1.227)
	21	Inverse variance weighted	0.047	1.113 (1.002 to 1.237)
000700/00707	21	veighted mode	0.740	1.040 (0.827 to 1.309)
GCS190199707	23	Weighted median	0.345	0.980 (0.939 to 1.022)
	23	Inverse variance weighted	0.048	0.965 (0.931 to 1.000)
	23	Weighted mode	0.416 🔫	0.982 (0.942 to 1.025)
GCST90199710	21	Weighted median	0.055 -	0.904 (0.815 to 1.002)
	21	Inverse variance weighted	0.041 🛏	0.931 (0.869 to 0.997)
	21	Weighted mode	0.100	0.907 (0.812 to 1.013)
GCST90199717	24	Weighted median	0.029	1.080 (1.008 to 1.157)
	24	Inverse variance weighted	0.008	1.077 (1.019 to 1.137)
	24	Weighted mode	0.042	1.081 (1.007 to 1.161)
GCST90199720	22	Weighted median	0.086	0.948 (0.893 to 1.007)
CCST00100767	22	Inverse variance weighted	0.049	0.948 (0.899 to 1.000)
	22	Molable d d-	0.100	0.046 (0.099 to 1.000)
	22	vveignted mode	0.109	0.946 (0.887 to 1.009)
GCST90199767	25	vveighted median	0.079	0.905 (0.809 to 1.012)
	25	Inverse variance weighted	0.048	0.926 (0.858 to 0.999)
	25	Weighted mode	0.263	0.924 (0.806 to 1.058)
GCST90199806 GCST90199832	15	Weighted median	0.025	0.929 (0.871 to 0.991)
	15	Inverse variance weighted	0.009	0.936 (0.890 to 0.984)
	15	Weighted mode	0.027	0.924 (0.868 to 0.984)
	17	Weighted median	0.169	1.088 (0.965 to 1.228)
	17	Inverse variance weighted	0.048	1.090 (1.001 to 1.187)
	17	Weighted mode	0.241	1 090 (0 949 to 1 253)
CCST00100969	20	Weighted median	0.014	1.002 (1.018 to 1.171)
003190199008	20	weighted median	0.014	1.092 (1.010 to 1.171)
	20	Inverse variance weighted	0.016	1.067 (1.012 (0 1.125)
	20	vveighted mode	0.021	1.092 (1.020 to 1.171)
GCST90199880	23	Weighted median	0.092	0.947 (0.889 to 1.009)
	23	Inverse variance weighted	0.040	0.945 (0.895 to 0.997)
	23	Weighted mode	0.072	0.943 (0.887 to 1.002)
GCST90199940	20	Weighted median	0.053 +	0.882 (0.776 to 1.002)
	20	Inverse variance weighted	0.045	0.907 (0.824 to 0.998)
	20	Weighted mode	0.157 🔸	0.850 (0.685 to 1.055)
GCST90199992	20	Weighted median	0.019 +•+	0.934 (0.882 to 0.989)
	20	Inverse variance weighted	0.032	0.949 (0.906 to 0.995)
	20	Weighted mode	0.033	0.932 (0.877 to 0.989)
GCST90200021	14	Weighted median	0.073	0.916 (0.833 to 1.008)
	14	Inverse variance weighted	0.029	0.919 (0.852 to 0.991)
	14	Mojekted mode	0.023	0.919 (0.032 (0.0391)
0007000000	14	Weighted mode	0.004	0.040 (0.040 to 1.040)
GCS190200023	12	vveignted median	0.150	0.916 (0.812 to 1.034)
	12	Inverse variance weighted	0.028	0.899 (0.818 to 0.989)
	12	Weighted mode	0.201	0.884 (0.741 to 1.056)
GCST90200024	20	Weighted median	0.160	0.953 (0.891 to 1.019)
	20	Inverse variance weighted	0.039 🛏	0.946 (0.897 to 0.997)
	20	Weighted mode	0.140	0.951 (0.893 to 1.014)
GCST90200026	27	Weighted median	0.030	0.934 (0.878 to 0.993)
	27	Inverse variance weighted	0.028	0.950 (0.908 to 0.994)
	27	Weighted mode	0.039	0.939 (0.887 to 0.994)
GCST90200042	18	Weighted median	0.255	0.930 (0.822 to 1.053)
	18	Inverse variance weighted	0.002	0.886 (0.821 to 0.957)
	18	Weighted mode	0.321	0.936 (0.824 to 1.062)
0000000000	05	Weighted mode	0.021	4.007 (0.024 to 1.003)
GGS190200047	25	vveignted median	0.105	1.097 (0.981 to 1.226)
	25	inverse variance weighted	0.003	1.117 (1.038 to 1.201)
00070000000	25	Weighted mode	0.446	1.057 (0.918 to 1.218)
GCST90200074	18	Weighted median	0.016	0.913 (0.849 to 0.983)
	18	Inverse variance weighted	0.014	0.924 (0.868 to 0.984)
	18	Weighted mode	0.049	0.921 (0.854 to 0.994)
GCST90200083	27	Weighted median	0.099	0.949 (0.891 to 1.010)
	27	Inverse variance weighted	0.016	0.942 (0.898 to 0.989)
	27	Weighted mode	0.169	0.954 (0.893 to 1.018)
GCST90200095	20	Weighted median	0.233	0.959 (0.895 to 1.027)
000100200000	20	Inverse variance weighted	0.019	0.942 (0.897 to 0.990)
	20	Wolferse variance weighted	0.013	0.042 (0.007 to 0.000)
CORTOCOCAOE	20	vveignted mode	0.217	0.960 (0.903 to 1.022)
GCST90200195	29	Weighted median	0.067	0.938 (0.875 to 1.005)
	29	Inverse variance weighted	0.015 +++	0.943 (0.899 to 0.988)
	29	Weighted mode	0.022	0.927 (0.872 to 0.986)
GCST90200213	17	Weighted median	0.174	1.096 (0.960 to 1.251)
	17	Inverse variance weighted	0.049	1 104 (1 000 to 1 219)
		inverse vanance weighted	01040	1.101 (1.000 to 1.210)

FIGURE 2

Forest plot of serum metabolites with atrial fibrillation.



### 3.4 The mediating role of serum metabolites in the causal relationship between serum albumin and atrial fibrillation

After a series of rigorous MR analyses, we successfully constructed a causal relationship chain based on the relationship between serum albumin and atrial fibrillation, mediated by the relationship between serum albumin and specific serum metabolites, and the relationship between these serum metabolites and atrial fibrillation. This series of causal links deeply reveals the key mediating role of these serum metabolites in the relationship between serum albumin and atrial fibrillation (Figures 5, 6). To more accurately quantify this mediating effect, we conducted detailed calculations on these two specific serum metabolites. The results showed that this mediating effect had significant statistical significance. Specifically, Docosatrienoate levels played a mediating role in the causal relationship between serum albumin and atrial fibrillation, with a magnitude of effect of  $\beta = -0.008$ , SE = 3.271e-05, and a 95% confidence interval ranging from -0.008 to -0.007. This mediating effect accounted for 4.63% of the overall causal relationship, with a confidence interval of 4.59%-4.67%. Similarly, the mediating effect of Oleate/vaccenate levels was  $\beta = -0.017$ , SE = 7.157e-05, with a 95% confidence interval ranging from -0.018 to -0.017. This



mediating effect accounted for up to 10.17% of the overall causal relationship, with a confidence interval of 10.09%–10.25%.

# 3.5 Validation using the Bayesian-weighted Mendelian randomization method

Finally, we validated our analysis using the Bayesian-weighted Mendelian randomization method. The results showed a significant negative correlation between serum albumin and atrial fibrillation (Beta = -0.172, OR = 0.842, 95% CI: 0.751-0.943, P = 0.003).

Subsequently, we also verified the mediating role of serum metabolites in the causal relationship between serum albumin and atrial fibrillation. The results indicated a positive causal relationship between serum albumin and Docosatrienoate (22:3n3) levels (Beta = 0.113, OR = 1.120, 95% CI: 1.000–1.253, p = 0.049), as well as a significant positive causal relationship between serum albumin and Oleate/vaccenate (18:1) levels (Beta = -0.147, OR = 1.159, 95% CI: 1.050–1.279, p = 0.003). However, unfortunately, when verifying the causal relationship between the two metabolite levels, Docosatrienoate (22:3n3) and Oleate/vaccenate (18:1), with atrial fibrillation, we had to



acknowledge that the Bayesian-weighted Mendelian randomization method did not successfully validate the causal relationship between Docosatrienoate (22:3n3) levels and atrial fibrillation (Beta = -0.072, OR = 0.931, 95% CI: 0.866–1.001, p = 0.053). Although the Beta value was less than 0 and the OR value was less than 1, indicating a negative causal relationship between Docosatrienoate (22:3n3) levels and atrial fibrillation, the *P*-value did not appear to be significant. On the other hand, we have successfully verified the causal relationship between Oleate/vaccenate (18:1) levels and atrial fibrillation (Beta = -0.154, OR = 0.857, 95% CI: 0.778-0.945, p = 0.001), which means that elevated Oleate/vaccenate (18:1) levels will reduce the risk of atrial fibrillation.

### 4 Discussion

Atrial fibrillation is a frequent arrhythmic phenomenon characterized by rapid and chaotic pulsations in the atria, which subsequently interfere with the normal rhythm of the ventricles. This condition significantly impairs the overall function of the heart, not only causing serious impact on patients' quality of life, but also potentially inducing other potential health problems (21). In the general population, the incidence of atrial fibrillation is approximately 2%, but in the elderly population aged 80 years and above, this proportion rapidly rises to 10%–12%. Various factors, such as myocardial fibrosis, inflammatory processes,



oxidative stress, and genetic and behavioral factors, are involved in the development of atrial fibrillation to varying degrees, collectively weaving an intricate pathophysiological network (22). Albumin, a protein with a molecular weight of 69 kDa consisting of 585 amino acids, is one of the most abundant circulating proteins in the blood and accounts for more than half of the total serum volume (23). As a "transporter" of blood, albumin can not only bind to and transport a variety of endogenous and exogenous substances, but also serves as a core element in maintaining blood osmotic balance, which is crucial for ensuring the stable operation of the circulatory system. In addition, albumin also exhibits its multifaceted biological activities, including antioxidant, anti-inflammatory, and anti-platelet aggregation effects (24). In healthy adult populations, serum albumin concentration usually remains within a relatively stable range between 3.5 g/dl and 5 g/dl. It is worth noting that women's serum albumin levels tend to be slightly lower than those of men and show a gradual decreasing trend with age.

Previously, no study has used MR analysis to clarify the causal relationship between serum albumin level and the risk of atrial fibrillation. Our study is the first to fully utilize publicly available GWAS datasets for in-depth MR analysis, aiming to reveal the direct causal link between serum albumin level and atrial fibrillation. In addition, we further explored the rich genetic data of serum metabolites to explore potential mediating factors between the two. Our findings showed that a decrease in serum albumin level significantly increases the risk of atrial fibrillation, which is consistent with previous observational studies. This important finding provides new perspectives and conclusive evidence for a more comprehensive understanding of the pathogenesis of atrial fibrillation.

Although the potential mechanism of serum albumin reducing the risk of atrial fibrillation has not been conclusive, we can explore it from multiple perspectives. Firstly, the strong antioxidant capacity of albumin is a factor that cannot be ignored. The occurrence of atrial fibrillation is closely related to the increase in oxidative stress levels in the atrium. The excessive production of reactive oxygen species (ROS) is directly related to structural changes and electrophysiological remodeling of the heart, which not only plays a role in triggering atrial fibrillation, but also plays an important role in maintaining this arrhythmic state (25). Albumin contains a free thiol group, accounting for 80% of free thiols in plasma, which can selectively scavenge ROS, making albumin an important antioxidant and regarded as the most important antioxidant in whole blood (26, 27). In addition, serum albumin can carry nitric oxide and bilirubin in plasma, providing additional protection to the heart muscle and preventing oxidative stress (28).

Serum albumin may play another important role in preventing the risk of atrial fibrillation, which is inseparable from its inhibitory effect on inflammation and blocking the process of atherosclerosis. Inflammation exacerbates the electrophysiological and structural changes of the atrium through various pathways, thereby promoting the occurrence and maintenance of atrial fibrillation (29). Atherosclerosis will change the structure and function of gap junction proteins, increase the return of electrical signals and enhance the excitability of atrial tissue, further promoting the occurrence and development of atrial fibrillation (30). Endothelial dysfunction, as one of the early manifestations of atherosclerosis, is closely related to the adhesion and migration of leukocytes on the vascular endothelium, and is one of the important ways to trigger inflammation and atherosclerosis. Serum albumin can selectively intervene in this process, reduce adhesion between monocytes and endothelial cells, and thus alleviate inflammatory response (31). In addition, albumin can regulate the production of nitric oxide to improve endothelial function and alleviate persistent myocardial inflammation. This is achieved by reducing the messenger RNA and protein expression of myocardial nitric oxide synthase II (32). The decrease in serum albumin levels may affect the integrity of the endothelial layer of the heart, and supplementing with albumin can help maintain this important barrier and further reduce inflammatory responses (33). During the formation of atherosclerosis, the overactivity of platelets will release a variety of proinflammatory and growth factors, which will aggravate the formation of vasculitis and atherosclerosis. It is interesting that albumin can inhibit the ability of histone induced platelet aggregation activity, which is directly related to its concentration in the blood (34). In addition, albumin also has significant anticoagulant properties, which helps prevent changes in cardiac structure and

electrophysiology caused by atherosclerosis, thus maintaining the stability of circulating blood flow and normal function of the heart (35).

Docosatrienoate is a polyunsaturated fatty acid (PUFA) belonging to the omega-3 family, which plays an important role in the composition of cell membrane phospholipids. The level of docosatrienoate is affected by multiple factors, including diet, age, gender, health status, and genetics. It decreases with age or the progression of chronic diseases (36). Omega-3 PUFA has been shown to affect cardiovascular health by regulating gene expression and transcription, promoting the production of antiinflammatory lipids, regulating fat oxidation processes and free radical release, and inhibiting ectopic electrical activity (37-39). Specifically, it can directly act on ion channels in cardiomyocytes, stabilizing myocardial electrical activity by inhibiting the flow of specific ions (Na+, Ca2+, K+, etc.) and prolonging the relative refractory period of the atrium (40, 41). In addition, it can further optimize the function of ion channels by adjusting the fluidity of cardiomyocyte membranes, thereby enhancing the electrophysiological stability of myocardial tissue (42-44). Omega-3 PUFA can also effectively alleviate inflammation in cardiac tissue, reduce oxidative stress levels, and alleviate calcium overload in cardiomyocytes, which is attributed to its ability to inhibit the production and activity of pro-inflammatory cytokines while regulating nuclear factor activity (45-47). In addition, it has a optimizing effect on autonomic nervous function, balancing the activity of sympathetic and vagal nerves and improving endothelial function (45, 46). At the level of mitochondria, it can regulate mitochondrial calcium homeostasis, regulate mitochondrial gene expression, enhance mitochondrial respiration, and control ROS production, thereby protecting the heart from potential risks of mitochondrial damage and cell apoptosis in a comprehensive manner (48, 49). Finally, it is worth mentioning that omega-3 PUFA can precisely regulate the gene expression pattern of the heart and reduce the occurrence of arrhythmia (50, 51).

As a monounsaturated fatty acid (MUFA), oleate and its precursor oleic acid (OA) play an important role in the human circulatory system. They not only participate in fat metabolism as energy sources, but also constitute important components of cell membranes (52). Research has confirmed that OA can significantly reduce various cardiovascular risks (53). Its unique antioxidant properties are particularly noteworthy, as it can directly regulate the synthesis and activity of antioxidant enzymes, thereby helping to improve endothelial function (54). OA prevents the conversion of vasodilator molecule oxide nitric acid to harmful peroxynitrite under oxidative stress environment (55). OA can also further exert its antioxidant stress effects by reducing mediators associated with oxidative stress (56, 57). On the other hand, OA can regulate cholesterol transport and absorption, inhibit inflammatory factors, and thereby alleviate inflammation in the inner wall of blood vessels (58). In the immune system, OA also plays an important regulatory role. It can alter the fluidity of immune cell membranes (59), thereby affecting membrane-mediated signaling (60). In addition, OA can regulate the activity of immune cells by regulating multiple

cytoplasmic signaling pathways such as MAPK and PI3 K/Akt (61). In neutrophils, OA can reduce their migration ability (62). In macrophages, it can inhibit the production of pro-inflammatory factors and promote the secretion of anti-inflammatory factors (63–65). OA can also affect the physiological processes of T cells and regulate the function of Treg cells (66–68). OA can also regulate the expression of various genes to further affect the function and activity of immune cells (69, 70).

Vaccinate, as a unique conjugated base of vaccenic acid(VA), is generated through the deprotonation process of carboxylic acid groups. It is worth mentioning that VA is a geometric isomer of OA, which also has various beneficial characteristics for cardiovascular health (71). Through research on various animal models, it has been found that VA can significantly improve the risk factors of cardiovascular disease. This positive effect may be attributed to its ability to reduce plasma cholesterol, regulate blood lipid levels, and optimize the ratio of high and low density lipoprotein (72-74). In addition, VA also demonstrated potential in reducing pro-inflammatory markers, successfully reducing IL-2 and TNF in rat disease models  $\alpha$  The level of (75, 76). In addition, VA can regulate the production of arachidonic acid, which is a key bioactive substance, and has an important impact on regulating lipid metabolism, immune response, vascular function, platelet aggregation and other downstream metabolic pathways involved in atherosclerosis (77).

When exploring the relationship between serum albumin and serum metabolites, it is not difficult to discern the crucial role of albumin. Docosatrienoate, Oleate, and Vaccenate are all types of fatty acids. Our MR analysis revealed a significant positive correlation between serum albumin levels and these three fatty acids. However, it is inaccurate to say that serum albumin elevates fatty acid levels because serum albumin itself does not increase the synthesis or production of fatty acids. Instead, serum albumin's role in regulating fatty acids is primarily reflected in its binding, thereby exerting transport, homeostatic maintenance, and protective effects (78, 79).

Serum albumin in the human body is a vital plasma protein that serves as the main carrier for fatty acids. It possesses multiple binding sites for fatty acids, which exhibit varying affinities, including two high-affinity, five medium-affinity, and over 20 low-affinity binding sites (80, 81). As a result, serum albumin can tightly bind to multiple fatty acid moieties with varying degrees of affinity. Structurally, albumin is composed of a complex polypeptide chain that folds intricately to form several globular units. Interestingly, the strong-affinity fatty acid binding sites may be located precisely in the clefts between these globular units, providing an ideal binding environment for fatty acids (82). When fatty acids are present in their anionic form, they can bind tightly to albumin. This tight binding is primarily attributed to the non-polar interactions established between the fatty acid hydrocarbon chains and the uncharged amino acid side chains on albumin (83). It is worth noting that these binding sites exhibit a certain degree of flexibility, allowing their conformation to adjust flexibly to accommodate the binding needs of different fatty acids. This characteristic ensures that albumin can effectively bind to various fatty acids, playing a critical role in transport and regulation within the human body. This has been further confirmed by studies showing that when albumin is knocked out in mice, the concentration of free fatty acids in their plasma significantly decreases (84). This finding underscores the importance of albumin in maintaining fatty acid homeostasis and transport.

The binding of plasma albumin to fatty acids plays a crucial role in the human body. Firstly, the binding of albumin to fatty acids stabilizes their presence in the bloodstream, preventing their rapid degradation and ensuring their effective transport from the liver to other tissues and organs for utilization. Secondly, albumin actively participates in regulating fatty acid metabolism by binding to them, helping maintain a steady-state concentration of fatty acids in the plasma. Through precise binding and release of fatty acids, albumin regulates their distribution, utilization, and excretion in the body, supporting normal physiological functions of organs. Additionally, the binding action of albumin provides protection to fatty acids, shielding them from damage caused by oxidation and other harmful factors, thereby maintaining their biological activity. It is worth mentioning that serum albumin itself possesses significant antioxidant properties, further providing a protective barrier for fatty acids and reducing the risk of oxidative damage. In summary, the binding of plasma albumin to fatty acids not only ensures their stable transport and metabolism but also provides comprehensive protection for their safety and activity in the body.

More and more studies have revealed that there is a significant negative correlation between serum albumin levels and cardiovascular diseases, especially coronary atherosclerotic heart disease, heart failure and atrial fibrillation. Even after comprehensive consideration and correction of multiple risk factors, this association remains stable and evident (85, 86). It is worth mentioning that serum albumin, as a prognostic assessment provides information independent tool, of conventional prognostic markers and therefore has unique value (87). Especially in patients with atrial fibrillation who undergo ablation treatment, serum albumin levels can predict the risk of postoperative recurrence, further highlighting its important role in the pathological process of atrial fibrillation (88). A recent study has provided strong support for our findings, analyzing major electronic health record reports from 80 healthcare organizations in the United States between 2000 and 2023. Through this large-scale retrospective analysis of a joint research network, researchers have discovered that patients with low albumin levels face a significantly increased risk of cardiovascular disease. This increased risk is independent of factors such as age, gender, comorbidities, and other causes of hypoalbuminemia. Of particular concern is the strong association between low albumin levels measured during the acute phase and an increased risk of atrial fibrillation in clinical conditions characterized by increased oxidative stress, such as ischemic stroke. Specifically, a decrease in albumin levels is associated with an 11% increase in the risk of atrial fibrillation within 30 days of its occurrence (89).

In view of this, this study aims to deeply analyze the causal relationship between serum albumin and atrial fibrillation. We adopted the MR analysis method, which is a unique genetic perspective that can examine genetic variations closely related to risk factors (90). Through this approach, we hope to more accurately identify which risk factors directly contribute to the occurrence of atrial fibrillation.

Our study has the following advantages. Firstly, compared to traditional observational studies, MR analysis is more rigorous in exploring causal relationships and can effectively avoid the interference of reverse causality and confounding factors. Secondly, to ensure the accuracy and reliability of the study, we deliberately selected a large sample size, further enhancing the statistical power and minimizing the potential impact of insufficient data or uncertainties on the research results. Finally, we employed Bayesian Mendelian randomization for validation, enhancing the robustness and comprehensiveness of the findings, which aids in our more holistic understanding of the relationship between exposure factors and outcome variables.

Although we have made efforts to overcome various limitations in our research, we still need to face up to their existing limitations. When analyzing the causal relationship between serum albumin and atrial fibrillation, although we have tried our best to control other potential interfering factors, we still cannot completely rule out the possibility that SNPs related to serum albumin may affect atrial fibrillation through other indirect pathways. This potential link between this polymorphism and other traits may cause confounding effects, which poses a certain challenge to our causal inference. On the other hand, since we used summary statistics from GWAS only and not individual genotype data, the estimated causal effect may be biased due to various factors such as the use of single variants, genotype imputation errors, and heterogeneity of genetic effects across different populations. Even when F statistics >10 is used, the weak instrument variable problem still exists. Third one is, Although we have conducted MR analysis using large-scale GWAS data, more massive and comprehensive prospective cohort studies are needed in the future to further enhance the reliability of our research conclusions and gradually expand their scope of application.

### 5 Conclusion

In summary, our study strongly demonstrates the causal relationship between serum albumin and reduced risk of atrial fibrillation through genetic methods, and reveals the key mediating role of two serum metabolites in this relationship. These findings not only provide a new perspective for understanding the role of serum albumin in atrial fibrillation, but also provide innovative ideas and directions for the prevention and treatment of cardiovascular diseases such as atrial fibrillation.

### Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material, further inquiries can be directed to the corresponding authors.

### **Ethics statement**

Ethical review and approval was not required for the study on human participants in accordance with the local legislation and institutional requirements. Written informed consent from the [patients/participants OR patients/participants legal guardian/next of kin] was not required to participate in this study in accordance with the national legislation and the institutional requirements.

### Author contributions

BC: Conceptualization, Data curation, Formal Analysis, Funding acquisition, Investigation, Methodology, Project administration, Resources, Software, Supervision, Validation, Visualization, Writing – original draft, Writing – review & editing. CW: Conceptualization, Data curation, Formal Analysis, Funding acquisition, Investigation, Methodology, Project administration, Resources, Software, Supervision, Validation, Visualization, Writing – original draft, Writing – review & editing. WL: Conceptualization, Data curation, Formal Analysis, Funding acquisition, Investigation, Methodology, Project administration, Resources, Software, Supervision, Validation, Visualization, Resources, Software, Supervision, Validation, Visualization, Writing – original draft, Writing – review & editing.

### Funding

The author(s) declare no financial support was received for the research, authorship, and/or publication of this article.

### Acknowledgments

The authors sincerely thank the relevant researchers for sharing the original GWAS summary statistical data analyzed in this study. Thanks to all authors for their participation and contributions in this article.

### Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

### Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their

affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

### References

1. Liu Y, Liu H, Dong L, Chen J, Guo J. Prevalence of atrial fibrillation in hospitalized patients over 40 years old: ten-year data from the people's hospital of Peking University. *Acta Cardiol.* (2010) 65(2):221-4. doi: 10.2143/AC.65.2.2047057

2. Mukamal KJ, Tolstrup JS, Friberg J, Grønbaek M, Jensen G. Fibrinogen and albumin levels and risk of atrial fibrillation in men and women (the Copenhagen City heart study). *Am J Cardiol.* (2006) 98(1):75–81. doi: 10.1016/j.amjcard.2006.01. 067

3. Zhao D, Jiao H, Zhong X, Wang W, Li L. The association between serum albumin levels and related metabolic factors and atrial fibrillation: a retrospective study. *Medicine (Baltimore)*. (2022) 101(44):e31581. doi: 10.1097/MD.000000000031581

4. Wang Y, Du P, Xiao Q, Li J, Liu X, Tan J, et al. Relationship between serum albumin and risk of atrial fibrillation: a dose-response meta-analysis. *Front Nutr.* (2021) 8:728353. doi: 10.3389/fnut.2021.728353

5. Sekula P, Del Greco MF, Pattaro C, Köttgen A. Mendelian randomization as an approach to assess causality using observational data. *J Am Soc Nephrol.* (2016) 27 (11):3253–65. doi: 10.1681/ASN.2016010098

6. Davey Smith G, Hemani G. Mendelian randomization: genetic anchors for causal inference in epidemiological studies. *Hum Mol Genet*. (2014) 23(R1):R89–98. doi: 10. 1093/hmg/ddu328

7. Barton AR, Sherman MA, Mukamel RE, Loh PR. Whole-exome imputation within UK biobank powers rare coding variant association and fine-mapping analyses. *Nat Genet.* (2021) 53(8):1260–9. doi: 10.1038/s41588-021-00892-1

8. Christophersen IE, Rienstra M, Roselli C, Yin X, Geelhoed B, Barnard J, et al. Large-scale analyses of common and rare variants identify 12 new loci associated with atrial fibrillation. *Nat Genet.* (2017) 49(6):946–52. Erratum in: Nat Genet. 2017 49(8):1286. doi: 10.1038/ng.3843

9. Shin SY, Fauman EB, Petersen AK, Krumsiek J, Santos R, Huang J, et al. An atlas of genetic influences on human blood metabolites. *Nat Genet.* (2014) 46(6):543–50. doi: 10.1038/ng.2982

10. Krumsiek J, Suhre K, Evans AM, Mitchell MW, Mohney RP, Milburn MV, et al. Mining the unknown: a systems approach to metabolite identification combining genetic and metabolic information. *PLoS Genet.* (2012) 8(10):e1003005. doi: 10. 1371/journal.pgen.1003005

11. Zhao J, Ming J, Hu X, Chen G, Liu J, Yang C. Bayesian weighted Mendelian randomization for causal inference based on summary statistics. *Bioinformatics*. (2020) 36(5):1501–8. doi: 10.1093/bioinformatics/btz749

12. Thomas DC, Conti DV. Commentary: the concept of "Mendelian randomization". Int J Epidemiol. (2004) 33(1):21-5. doi: 10.1093/ije/dyh048

13. Gill D, Efstathiadou A, Cawood K, Tzoulaki I, Dehghan A. Education protects against coronary heart disease and stroke independently of cognitive function: evidence from Mendelian randomization. *Int J Epidemiol.* (2019) 48(5):1468–77. doi: 10.1093/ije/dyz200

14. Palmer TM, Lawlor DA, Harbord RM, Sheehan NA, Tobias JH, Timpson NJ, et al. Using multiple genetic variants as instrumental variables for modifiable risk factors. *Stat Methods Med Res.* (2012) 21(3):223–42. doi: 10.1177/0962280210394459

15. Levin MG, Judy R, Gill D, Vujkovic M, Verma SS, Bradford Y, et al. Genetics of height and risk of atrial fibrillation: a Mendelian randomization study. *PLoS Med.* (2020) 17(10):e1003288. doi: 10.1371/journal.pmed.1003288

16. Burgess S, Scott RA, Timpson NJ, Davey Smith G, Thompson SG, EPIC-InterAct Consortium. Using published data in Mendelian randomization: a blueprint for efficient identification of causal risk factors. *Eur J Epidemiol.* 2015 30 (7):543–52. doi: 10.1007/s10654-015-0011-z

17. Burgess S, Butterworth A, Thompson SG. Mendelian randomization analysis with multiple genetic variants using summarized data. *Genet Epidemiol.* (2013) 37 (7):658–65. doi: 10.1002/gepi.21758

18. Bowden J, Davey Smith G, Haycock PC, Burgess S. Consistent estimation in Mendelian randomization with some invalid instruments using a weighted median estimator. *Genet Epidemiol.* (2016) 40(4):304–14. doi: 10.1002/gepi.21965

19. Bowden J, Davey Smith G, Burgess S. Mendelian randomization with invalid instruments: effect estimation and bias detection through egger regression. *Int J Epidemiol.* (2015) 44(2):512–25. doi: 10.1093/ije/dyv080

### Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fcvm.2024. 1385223/full#supplementary-material

20. Verbanck M, Chen CY, Neale B, Do R. Detection of widespread horizontal pleiotropy in causal relationships inferred from Mendelian randomization between complex traits and diseases. *Nat Genet.* (2018) 50(5):693–8. Erratum in: Nat Genet. 2018 50(8):1196. doi: 10.1038/s41588-018-0099-7

21. Staerk L, Sherer JA, Ko D, Benjamin EJ, Helm RH. Atrial fibrillation: epidemiology, pathophysiology, and clinical outcomes. *Circ Res.* (2017) 120:1501–17. doi: 10.1161/CIRCRESAHA.117.309732

22. Schnabel RB, Yin X, Gona P, Larson MG, Beiser AS, McManus DD, et al. 50 Year trends in atrial fibrillation prevalence, incidence, risk factors, and mortality in the Framingham Heart Study: a cohort study. *Lancet.* (2015) 386(9989):154–62. doi: 10.1016/S0140-6736(14)61774-8

23. Goldwasser P, Feldman J. Association of serum albumin and mortality risk. J Clin Epidemiol. (1997) 50(6):693-703. doi: 10.1016/s0895-4356(97)00015-2

24. Arques S. Human serum albumin in cardiovascular diseases. Eur J Intern Med. (2018) 52:8–12. doi: 10.1016/j.ejim.2018.04.014

25. Lenaerts I, Driesen RB, Hermida N, Holemans P, Heidbüchel H, Janssens S, et al. Role of nitric oxide and oxidative stress in a sheep model of persistent atrial fibrillation. *Europace*. (2013) 15(5):754–60. Erratum in: Europace. 2013 15(11):1684. Blanco, Nerea Hermida [corrected to Hermida, Nerea]. doi: 10.1093/europace/eut012

26. Roche M, Rondeau P, Singh NR, Tarnus E, Bourdon E. The antioxidant properties of serum albumin. *FEBS Lett.* (2008) 582(13):1783–7. doi: 10.1016/j. febslet.2008.04.057

27. Anraku M, Chuang VT, Maruyama T, Otagiri M. Redox properties of serum albumin. *Biochim Biophys Acta.* (2013) 1830(12):5465–72. doi: 10.1016/j.bbagen. 2013.04.036

28. Stamler JS, Jaraki O, Osborne J, Simon DI, Keaney J, Vita J, et al. Nitric oxide circulates in mammalian plasma primarily as an S-nitroso adduct of serum albumin. *Proc Natl Acad Sci U S A*. (1992) 89(16):7674–7. doi: 10.1073/pnas.89.16.7674

29. Guo Y, Lip GY, Apostolakis S. Inflammation in atrial fibrillation. J Am Coll Cardiol. (2012) 60(22):2263–70. doi: 10.1016/j.jacc.2012.04.063

30. Batta A, Hatwal J, Batta A, Verma S, Sharma YP. Atrial fibrillation and coronary artery disease: an integrative review focusing on therapeutic implications of this relationship. *World J Cardiol.* (2023) 15(5):229–43. doi: 10.4330/wjc.v15.i5.229

31. Zhang WJ, Frei B. Albumin selectively inhibits TNF alpha-induced expression of vascular cell adhesion molecule-1 in human aortic endothelial cells. *Cardiovasc Res.* (2002) 55(4):820–9. doi: 10.1016/s0008-6363(02)00492-3

32. Walley KR, McDonald TE, Wang Y, Dai S, Russell JA. Albumin resuscitation increases cardiomyocyte contractility and decreases nitric oxide synthase II expression in rat endotoxemia. *Crit Care Med.* (2003) 31(1):187–94. doi: 10.1097/00003246-200301000-00029

33. Mendes RS, Pelosi P, Schultz MJ, Rocco PRM, Silva PL. Fluids in ARDS: more pros than cons. *Intensive Care Med Exp.* (2020) 8(Suppl 1):32. doi: 10.1186/s40635-020-00319-x

34. Lam FW, Cruz MA, Leung HC, Parikh KS, Smith CW, Rumbaut RE. Histone induced platelet aggregation is inhibited by normal albumin. *Thromb Res.* (2013) 132(1):69–76. doi: 10.1016/j.thromres.2013.04.018

35. Paar M, Rossmann C, Nusshold C, Wagner T, Schlagenhauf A, Leschnik B, et al. Anticoagulant action of low, physiologic, and high albumin levels in whole blood. *PLoS One.* (2017) 12(8):e0182997. doi: 10.1371/journal.pone.0182997

36. Kar S. Role of omega-3 fatty acids in the prevention of atrial fibrillation. *Rev Cardiovasc Med.* (2013) 14(2–4):e82–91. doi: 10.3909/ricm0620

37. Mollace V, Gliozzi M, Carresi C, Musolino V, Oppedisano F. Re-assessing the mechanism of action of n-3 PUFAs. *Int J Cardiol.* (2013) 170(2 Suppl 1):S8–11. doi: 10.1016/j.ijcard.2013.06.038

38. Endo J, Arita M. Cardioprotective mechanism of omega-3 polyunsaturated fatty acids. J Cardiol. (2016) 67(1):22-7. doi: 10.1016/j.jjcc.2015.08.002

39. Christou GA, Christou KA, Korantzopoulos P, Rizos EC, Nikas DN, Goudevenos JA. The current role of Omega-3 fatty acids in the management of atrial fibrillation. *Int J Mol Sci.* (2015) 16(9):22870–87. doi: 10.3390/ijms160922870

40. Allaire J, Couture P, Leclerc M, Charest A, Marin J, Lépine MC, et al. A randomized, crossover, head-to-head comparison of eicosapentaenoic acid and

docosahexaenoic acid supplementation to reduce inflammation markers in men and women: the comparing EPA to DHA (ComparED) study. *Am J Clin Nutr.* (2016) 104(2):280–7. doi: 10.3945/ajcn.116.131896

41. Roy J, Le Guennec JY. Cardioprotective effects of omega 3 fatty acids: origin of the variability. J Muscle Res Cell Motil. (2017) 38(1):25–30. doi: 10.1007/s10974-016-9459-z

42. Calder PC. Mechanisms of action of (n-3) fatty acids. J Nutr. (2012) 142 (3):592S-9S. doi: 10.3945/jn.111.155259

43. Fenton JI, Gurzell EA, Davidson EA, Harris WS. Red blood cell PUFAs reflect the phospholipid PUFA composition of major organs. *Prostaglandins Leukot Essent Fatty Acids*. (2016) 112:12–23. doi: 10.1016/j.plefa.2016.06.004

44. Jahangiri A, Leifert WR, Patten GS, McMurchie EJ. Termination of asynchronous contractile activity in rat atrial myocytes by n-3 polyunsaturated fatty acids. *Mol Cell Biochem*. (2000) 206(1-2):33–41. doi: 10.1023/a:1007025007403

45. Endo J, Sano M, Isobe Y, Fukuda K, Kang JX, Arai H, et al. 18-HEPE, an n-3 fatty acid metabolite released by macrophages, prevents pressure overload-induced maladaptive cardiac remodeling. *J Exp Med.* (2014) 211(8):1673–87. doi: 10.1084/ jem.20132011

46. Ramadeen A, Laurent G, dos Santos CC, Hu X, Connelly KA, Holub BJ, et al. n-3 polyunsaturated fatty acids alter expression of fibrotic and hypertrophic genes in a dog model of atrial cardiomyopathy. *Heart Rhythm*. (2010) 7(4):520–8. doi: 10.1016/j. hrthm.2009.12.016

47. Baum JR, Dolmatova E, Tan A, Duffy HS. Omega 3 fatty acid inhibition of inflammatory cytokine-mediated Connexin43 regulation in the heart. *Front Physiol.* (2012) 3:272. doi: 10.3389/fphys.2012.00272

48. Akar FG, O'Rourke B. Mitochondria are sources of metabolic sink and arrhythmias. *Pharmacol Ther.* (2011) 131(3):287-94. doi: 10.1016/j.pharmthera. 2011.04.005

49. Rohrbach S. Effects of dietary polyunsaturated fatty acids on mitochondria. *Curr Pharm Des.* (2009) 15(36):4103–16. doi: 10.2174/138161209789909692

50. Bordoni A, Astolfi A, Morandi L, Pession A, Danesi F, Di Nunzio M, et al. N-3 PUFAs modulate global gene expression profile in cultured rat cardiomyocytes. Implications in cardiac hypertrophy and heart failure. *FEBS Lett.* (2007) 581 (5):923–9. doi: 10.1016/j.febslet.2007.01.070

51. Deckelbaum RJ, Worgall TS, Seo T. n-3 fatty acids and gene expression. *Am J Clin Nutr.* (2006) 83(6 Suppl):1520S-5S. Erratum in: Am J Clin Nutr. 2006 84 (4):949. doi: 10.1093/ajcn/83.6.1520S.

52. Schwingshackl L, Hoffmann G. Monounsaturated fatty acids and risk of cardiovascular disease: synopsis of the evidence available from systematic reviews and meta-analyses. *Nutrients*. (2012) 4(12):1989–2007. doi: 10.3390/nu4121989

53. Huth PJ, Fulgoni VL 3rd, Larson BT. A systematic review of high-oleic vegetable oil substitutions for other fats and oils on cardiovascular disease risk factors: implications for novel high-oleic soybean oils. *Adv Nutr.* 2015 6(6):674–93. doi: 10. 3945/an.115.008979

54. Bhattacharjee B, Pal PK, Chattopadhyay A, Bandyopadhyay D. Oleic acid protects against cadmium induced cardiac and hepatic tissue injury in male wistar rats: a mechanistic study. *Life Sci.* (2020) 244:117324. doi: 10.1016/j.lfs.2020.117324

55. Chen J, Li Q, Zhang Y, Yang P, Zong Y, Qu S, et al. Oleic acid decreases the expression of a cholesterol transport-related protein (NPC1L1) by the induction of endoplasmic reticulum stress in CaCo-2 cells. *J Physiol Biochem.* (2011) 67 (2):153–63. doi: 10.1007/s13105-010-0058-y

56. Oh YT, Lee JY, Lee J, Kim H, Yoon KS, Choe W, et al. Oleic acid reduces lipopolysaccharide-induced expression of iNOS and COX-2 in BV2 murine microglial cells: possible involvement of reactive oxygen species, p38 MAPK, and IKK/NF-kappaB signaling pathways. *Neurosci Lett.* (2009) 464(2):93–7. doi: 10.1016/j.neulet.2009.08.040

57. Harvey KA, Walker CL, Xu Z, Whitley P, Pavlina TM, Hise M, et al. Oleic acid inhibits stearic acid-induced inhibition of cell growth and pro-inflammatory responses in human aortic endothelial cells. *J Lipid Res.* (2010) 51(12):3470–80. doi: 10.1194/jlr. M010371

58. Yang ZH, Nill K, Takechi-Haraya Y, Playford MP, Nguyen D, Yu ZX, et al. Differential effect of dietary supplementation with a soybean oil enriched in oleic acid versus linoleic acid on plasma lipids and atherosclerosis in LDLR-deficient mice. *Int J Mol Sci.* (2022) 23(15):8385. doi: 10.3390/ijms23158385

59. Speizer LA, Watson MJ, Brunton LL. Differential effects of omega-3 fish oils on protein kinase activities in vitro. *Am J Physiol.* (1991) 261(1 Pt 1):E109–14. doi: 10. 1152/ajpendo.1991.261.1.E109

60. Leekumjorn S, Cho HJ, Wu Y, Wright NT, Sum AK, Chan C. The role of fatty acid unsaturation in minimizing biophysical changes on the structure and local effects of bilayer membranes. *Biochim Biophys Acta*. (2009) 1788(7):1508–16. doi: 10.1016/j. bbamem.2009.04.002

61. Manosalva C, Mena J, Velasquez Z, Colenso CK, Brauchi S, Burgos RA, et al. Cloning, identification and functional characterization of bovine free fatty acid receptor-1 (FFAR1/GPR40) in neutrophils. *PLoS One.* (2015) 10(3):e0119715. doi: 10.1371/journal.pone.0119715

62. Mena SJ, Manosalva C, Carretta MD, Teuber S, Olmo I, Burgos RA, et al. Differential free fatty acid receptor-1 (FFAR1/GPR40) signalling is associated with gene expression or gelatinase granule release in bovine neutrophils. *Innate Immun.* (2016) 22(6):479–89. doi: 10.1177/1753425916656765

63. Hou Y, Wei D, Bossila EA, Zhang Z, Li S, Bao J, et al. FABP5 deficiency impaired macrophage inflammation by regulating AMPK/NF-κB signaling pathway. *J Immunol.* (2022) 209(11):2181–91. doi: 10.4049/jimmunol.2200182

64. Hong JH, Lee YC. Anti-inflammatory effects of cicadidae periostracum extract and oleic acid through inhibiting inflammatory chemokines using PCR arrays in LPS-induced lung inflammation in vitro. *Life (Basel).* (2022) 12(6):857. doi: 10. 3390/life12060857

65. Müller AK, Albrecht F, Rohrer C, Koeberle A, Werz O, Schlörmann W, et al. Olive oil extracts and oleic acid attenuate the LPS-induced inflammatory response in murine RAW264.7 macrophages but induce the release of prostaglandin E2. *Nutrients.* (2021) 13(12):4437. doi: 10.3390/nu13124437

66. Carrillo C, Giraldo M, Cavia MM, Alonso-Torre SR. Effect of oleic acid on storeoperated calcium entry in immune-competent cells. *Eur J Nutr.* (2017) 56(3):1077–84. doi: 10.1007/s00394-016-1157-5

67. Pompura SL, Wagner A, Kitz A, LaPerche J, Yosef N, Dominguez-Villar M, et al. Oleic acid restores suppressive defects in tissue-resident FOXP3 tregs from patients with multiple sclerosis. *J Clin Invest.* (2021) 131(2):e138519. doi: 10.1172/JCI138519

68. Singh V, Ubaid S. Role of silent information regulator 1 (SIRT1) in regulating oxidative stress and inflammation. *Inflammation*. (2020) 43(5):1589–98. Erratum in: Inflammation. 2021 44(5):2142. doi: 10.1007/s10753-020-01242-9

69. Christofides A, Konstantinidou E, Jani C, Boussiotis VA. The role of peroxisome proliferator-activated receptors (PPAR) in immune responses. *Metab Clin Exp.* (2021) 114:154338. doi: 10.1016/j.metabol.2020.154338

70. Medeiros-de-Moraes IM, Gonçalves-de-Albuquerque CF, Kurz ARM, Oliveira FMJ, de Abreu VHP, Torres RC, et al. Omega-9 oleic acid, the main compound of olive oil, mitigates inflammation during experimental sepsis. *Oxid Med Cell Longev.* (2018) 2018:6053492. doi: 10.1155/2018/6053492

71. Field CJ, Blewett HH, Proctor S, Vine D. Human health benefits of vaccenic acid. *Appl Physiol Nutr Metab.* (2009) 34(5):979–91. doi: 10.1139/H09-079

72. Lock AL, Horne CA, Bauman DE, Salter AM. Butter naturally enriched in conjugated linoleic acid and vaccenic acid alters tissue fatty acids and improves the plasma lipoprotein profile in cholesterol-fed hamsters. *J Nutr.* (2005) 135(8):1934–9. doi: 10.1093/jn/135.8.1934

73. Bauchart D, Roy A, Lorenz S, Chardigny JM, Ferlay A, Gruffat D, et al. Butters varying in trans 18:1 and cis-9,trans-11 conjugated linoleic acid modify plasma lipoproteins in the hypercholesterolemic rabbit. *Lipids.* (2007) 42(2):123–33. doi: 10. 1007/s11745-006-3018-0

74. Roy A, Chardigny JM, Bauchart D, Ferlay A, Lorenz S, Durand D, et al. Butters rich either in trans-10-C18:1 or in trans-11-C18:1 plus cis-9, trans-11 CLA differentially affect plasma lipids and aortic fatty streak in experimental atherosclerosis in rabbits. *Animal.* (2007) 1(3):467–76. doi: 10.1017/S175173110770530X

75. Blewett HJ, Gerdung CA, Ruth MR, Proctor SD, Field CJ. Vaccenic acid favourably alters immune function in obese JCR:IA-cp rats. *Br J Nutr.* (2009) 102 (4):526–36. doi: 10.1017/S0007114509231722

76. Wang Y, Jacome-Sosa MM, Ruth MR, Goruk SD, Reaney MJ, Glimm DR, et al. Trans-11 vaccenic acid reduces hepatic lipogenesis and chylomicron secretion in JCR: lA-cp rats. J Nutr. (2009) 139(11):2049–54. doi: 10.3945/jn.109.109488

77. Gebauer SK, Chardigny JM, Jakobsen MU, Lamarche B, Lock AL, Proctor SD, et al. Effects of ruminant trans fatty acids on cardiovascular disease and cancer: a comprehensive review of epidemiological, clinical, and mechanistic studies. *Adv Nutr.* (2011) 2(4):332–54. doi: 10.3945/an.111.000521

78. Litus EA, Permyakov SE, Uversky VN, Permyakov EA. Intrinsically disordered regions in serum albumin: what are they for? *Cell Biochem Biophys.* (2018) 76(1–2):39–57. doi: 10.1007/s12013-017-0785-6

79. van der Vusse GJ. Albumin as fatty acid transporter. Drug Metab Pharmacokinet. (2009) 24(4):300-7. doi: 10.2133/dmpk.24.300

80. Curry S, Brick P, Franks NP. Fatty acid binding to human serum albumin: new insights from crystallographic studies. *Biochim Biophys Acta*. (1999) 1441(2-3):131-40. doi: 10.1016/s1388-1981(99)00148-1

81. Linciano S, Moro G, Zorzi A, Angelini A. Molecular analysis and therapeutic applications of human serum albumin-fatty acid interactions. *J Control Release*. (2022) 348:115–26. doi: 10.1016/j.jconrel.2022.05.038

82. Spector AA. Fatty acid binding to plasma albumin. J Lipid Res. (1975) 16 (3):165-79. doi: 10.1016/S0022-2275(20)36723-7

83. Reichenwallner J, Hinderberger D. Using bound fatty acids to disclose the functional structure of serum albumin. *Biochim Biophys Acta.* (2013) 1830 (12):5382–93. doi: 10.1016/j.bbagen.2013.04.031

84. Abdollahi A, Dowden BN, Buhman KK, Zembroski AS, Henderson GC. Albumin knockout mice exhibit reduced plasma free fatty acid concentration and enhanced insulin sensitivity. *Physiol Rep.* (2022) 10(5):e15161. doi: 10.14814/phy2. 15161

85. Arques S. Serum albumin and cardiovascular disease: state-of-the-art review. Ann Cardiol Angeiol (Paris). (2020) 69(4):192–200. doi: 10.1016/j.ancard. 2020.07.012

86. Yuan HJ, Zhong X, Li Y, Xue YT, Jiao HC. Correlation between serum albumin and D-dimer levels in 909 patients with non-valvular atrial fibrillation: a retrospective study from a single center in China. *Med Sci Monit.* (2022) 28:e938511. doi: 10.12659/ MSM.938511

87. Zhong X, Jiao H, Zhao D, Teng J, Yang M. A retrospective study to determine the association between serum albumin levels and atrial fibrillation by sex in 950 patients from a single center in China. *Med Sci Monit.* (2022) 28:e935347. doi: 10. 12659/MSM.935347

88. Schamroth Pravda N, Golovchiner G, Goldenberg G, Plakht Y, Wiessman M, Tal S, et al. Albumin as a prognostic marker for atrial fibrillation recurrence following cryoballoon ablation of pulmonary venous. *J Clin Med.* (2022) 12(1):264. doi: 10. 3390/jcm12010264

89. Bucci T, Pastori D, Pignatelli P, Ntaios G, Abdul-Rahim AH, Violi F, et al. Albumin levels and risk of early cardiovascular complications after ischemic stroke: a propensity-matched analysis of a global federated health network. *Stroke.* (2024) 55(3):604–12. doi: 10.1161/STROKEAHA.123.044248

90. Larsson SC. Mendelian randomization as a tool for causal inference in human nutrition and metabolism. *Curr Opin Lipidol.* (2021) 32(1):1–8. doi: 10.1097/MOL. 00000000000721