



OPEN ACCESS

EDITED BY Michael Brunner, St. Josefskrankenhaus, Germany

REVIEWED BY

Norbert Szentandrássy, University of Debrecen, Hungary Martin Manninger, Medical University of Graz, Austria

*CORRESPONDENCE Julian Wolfes

[†]These authors have contributed equally to this work

RECEIVED 11 January 2024 ACCEPTED 12 February 2024 PUBLISHED 22 February 2024

Wolfes J, Uphoff J, Kemena S, Wegner F, Rath B, Eckardt L, Frommeyer G and Ellermann C (2024) Divergent electrophysiologic action of dapagliflozin and empagliflozin on ventricular and atrial tachyarrhythmias in isolated rabbit hearts. Front Cardiovasc Med 11:1369250 doi: 10.3389/fcvm.2024.1369250

© 2024 Wolfes, Uphoff, Kemena, Wegner, Rath, Eckardt, Frommeyer and Ellermann. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted. provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Divergent electrophysiologic action of dapagliflozin and empagliflozin on ventricular and atrial tachyarrhythmias in isolated rabbit hearts

Julian Wolfes*, Jan Uphoff, Sven Kemena, Felix Wegner, Benjamin Rath, Lars Eckardt, Gerrit Frommeyer[†] and Christian Ellermann[†]

Department of Cardiology II, Electrophysiology, University Hospital Münster, Münster, Germany

Background: The use of SGLT-2 inhibitors has revolutionized heart failure therapy. Evidence suggests a reduced incidence of ventricular and atrial arrhythmias in patients with dapagliflozin or empagliflozin treatment. It is unclear to what extent the reduced arrhythmia burden is due to direct effects of the SGLT2 inhibitors or is solely a marker of improved cardiac function.

Methods: One hundred five rabbit hearts were allocated to eight groups and retrogradely perfused, employing a Langendorff setup. Action potential duration at 90% of repolarization (APD₉₀), QT intervals, effective refractory periods, conduction velocity, and dispersion of repolarization were obtained with monophasic action potential catheters. A model for tachyarrhythmias was established with the I_{Kr} blocker erythromycin for QT prolongation associated proarrhythmia as well as the potassium channel opener pinacidil for a short-QT model. An atrial fibrillation (AF) model was created with isoproterenol and acetylcholine. With increasing concentrations of both SGLT2 inhibitors, reductions in QT intervals and APD₉₀ were observed, accompanied by a slight in ventricular arrhythmia episodes. During drug-induced proarrhythmia, empagliflozin succeeded in decreasing QT intervals, APD₉₀, and VT burden whereas dapagliflozin demonstrated no significant effects. In the presence of pinacidil induced arrhythmogenicity, neither SGLT2 inhibitor had a significant impact on cardiac electrophysiology. In the AF setting, perfusion with dapagliflozin showed significant suppression of AF in the course of restitution of electrophysiological parameters whereas empagliflozin showed no significant effect on atrial fibrillation incidence.

Conclusion: In this model, empagliflozin and dapagliflozin demonstrated opposite antiarrhythmic properties. Empagliflozin reduced ventricular tachyarrhythmias whereas dapagliflozin showed effective suppression of atrial arrhythmias.

KEYWORDS

SGLT2, dapagliflozin, empagliflozin, langendorff, atrial fibrillation, short QT syndrome, long QT syndrome, arrhythmia

Introduction

The introduction of SGLT2-inhibitors has revolutionized heart failure therapy not only in heart failure with reduced ejection fraction (1, 2) but also in heart failure with preserved ejection fraction (3, 4). The impressive data of recent randomized trials has resulted in a recent update of the ESC guidelines in heart failure (5). Previous studies indicated a reduced incidence of ventricular tachyarrhythmias (VA) and atrial fibrillation (AF) with SGLT2-inhibitor therapy (6). To date, it is unclear whether the antiarrhythmic potency of SGLT2-inhibitors is due to direct interactions with cardiomyocytes or is merely a marker of improved cardiac function. Experimental data on the effect of SGLT2-inhibitors on ventricular and atrial electrophysiology is scarce. Antiarrhythmic effects have been described for dapagliflozin at the ventricular (7-9) level in different animal models. Similarly, ventricular antiarrhythmic effects have been described for empagliflozin in rat (10-12) and mouse models (13, 14) but also rabbit Langendorff models (15), whereas atrial effects are less recorded. Therefore, this study aimed at evaluating acute effects of the two SGLT2-inhibitors dapagliflozin and empagliflozin on atrial and ventricular electrophysiology in an established model.

Methods

All experimental protocols were approved by the local animal care committee and the local federal institution (Landesamt für Natur, Umwelt und Verbraucherschutz Nordrhein-Westfalen, file number: 81-02.05.50.21.004). The experimental Langendorff whole-heart setup has been extensively described by our group in previous publications (16–19).

In summary, a total of 105 rabbit hearts were obtained after the animals were euthanized by exsanguination. The hearts were aortically clamped into a Langendorff apparatus and retrogradely perfused employing warmed and oxygenated (95% O₂, 5% CO₂) KrebsisHenseleit buffer (NaCl 118 mM, NaHCO₃ 24.88 mM, D-glucose 5.55 mM, KCl 4.70 mM, Na-pyruvate 2 mM, CaCl₂ 1.80 mM, KH₂PO₄ 1.18 mM, MgSO₄ 0.83 mM). From here on, hearts were divided into two different experimental groups depending on whether the ventricular or atrial electrophysiological effects of SGLT-2 inhibitors were to be studied.

Experiments on ventricular electrophysiology and tachyarrhythmias

Seven electrophysiological catheters were placed epicardially for the detection of monophasic action potentials and one endocardial catheter was placed in the left ventricle. Furthermore, a pseudo 12 lead ECG was recorded from the warming-bath surrounding the heart. Mechanical AV nodal ablation was performed. Thereafter, the pacing protocol was started:

First, the unstimulated ventricular escape rate of the hearts was determined. Subsequently, pacing was performed with seven different cycle lengths between 900 and 300 ms while the cycle length-dependent monophasic action potentials and QT intervals were determined.

Subsequently, pacing with a short-coupled extrastimulus was performed to determine the effective refractory period (ERP). In addition, repetitive burst stimulations (3 cycles of 1,000 ms at $1~{\rm Hz}$) were used to record ventricular vulnerability. This was followed by perfusion with hypokalemic KHB (K+1.5 mM) to determine arrhythmia susceptibility in a hypokalemic environment.

Action potential duration at 90% of repolarization (APD $_{90}$) was measured between the fastest upstroke and 90% of repolarization. Spatial dispersion of repolarization was determined by the difference between the maximum and the minimum APD of the eight simultaneously recorded monophasic action potentials. Post-repolarization refractoriness (PRR) was calculated as the difference between ERP and APD $_{90}$.

Experiments on atrial electrophysiology and tachyarrhythmias

Five epicardial MAP catheters were placed on both atria with two recording MAPs on each atrium and one MAP for stimulation at the interseptal region. The hearts were paced with a cycle-length (CL) of 350, 250, and 150 ms for 1 min to stabilize the action potential before recording 16 consecutive beats at each CL for atrial action potential duration (aAPD₉₀) analysis. The atrial effective refractory period (aERP) and atrial post-repolarization refractoriness (aPRR) were determined analogously to the ventricular experiments. Burst pacing (10 cycles of 1,000 ms at 1 Hz) was utilized to evaluate arrhythmia susceptibility.

The 105 hearts were allocated to eight groups: In group 1 (Dapa), 12 hearts (n = 12) were treated with 3 μ M dapagliflozin. Thereafter, $5\,\mu M$ dapagliflozin was infused followed by $10\,\mu M$ dapagliflozin. In group 2 (Ery + Dapa) 12 hearts (n = 12) were perfused with $300\,\mu M$ erythromycin followed by $5\,\mu M$ dapagliflozin. In group 3 (Pina + Dapa), the 12 hearts (n = 12)were perfused with 1 μM pinacidil followed by 5 μM dapagliflozin. Group 4 (IsoACh + Dapa) employed the atrial setup in 13 hearts (n = 13). After the determination of the electrophysiological determinants at baseline, hearts were perfused with a combination of isoproterenol (1 µM) and acetylcholine $(1 \mu M)$ to increase the occurrence of AF. The combination of both drugs has been previously tested on this (20) and other experimental models (21). After 15 min of incubation, the aforementioned pacing protocol was employed again. Afterwards, the hearts were additionally perfused with 3 μM dapagliflozin, and electrophysiological parameters were determined. Each drug intervention underwent a 15-min run-in period before the electrophysiologic study was performed.

In group 5 (Empa), 13 hearts (n = 13) were treated with 1 μ M empagliflozin. Thereafter, 3 μ M empagliflozin was infused followed

by 5 μ M empagliflozin. In group 6 (Ery + Empa) 17 hearts (n=17) were perfused with 300 μ M erythromycin followed by 3 μ M empagliflozin. In group 7 (Pina + Empa), the 14 hearts (n=14) were perfused with 1 μ M pinacidil followed by 3 μ M empagliflozin. Group 8 (IsoACh + Empa) employed the atrial setup in 12 hearts (n=12). Analogous to group 4, after determination of baseline parameters, perfusion with 1 μ M IsoACh followed by perfusion with 3 μ M Empa was performed.

Electrograms and action potentials were recorded on a multichannel recorder and digitalized at a rate of 1 kHz with a 12-bit resolution. Variables are shown as mean \pm standard deviation. Statistical analyses and graphic visualizations were performed employing Graphpad Prism Version 9. Drug effects on mean APD₉₀, QT interval, spatial dispersion of repolarization of all cycle length, effective refractory periods and arrhythmia incidence were analyzed employing repeated measures ANOVA for multiple comparisons. P values < 0.05 were considered to be statistically significant.

Results

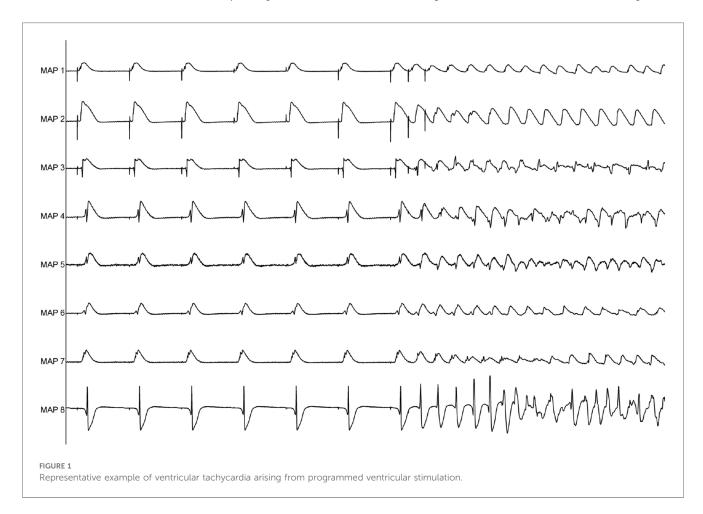
Dapagliflozin

Perfusion with dapagliflozin caused a significant shortening of QT intervals [baseline: 270 ± 51 ms; $3 \,\mu M$ Dapa: 243 ± 50 ms

(p < 0.05); 5 μM Dapa: 223 ± 43 ms (p < 0.05); 10 μM Dapa: 227 ± 46 ms (p < 0.05)] and APD₉₀ intervals [baseline: 178 ± 36 ms; 3 μM Dapa: 156 ± 33 ms (p < 0.05); 5 μM Dapa: 141 ± 27 ms (p < 0.05); 10 μM Dapa: 152 ± 35 ms (p < 0.05)] in rabbit ventricles, while spatial dispersion of repolarization was not significantly altered. Furthermore, perfusion with dapagliflozin caused a significant shortening of ERP while the PRR shortening reached statistical significance only under 10 μM Dapa. The incidence of ventricular arrhythmia episodes under programmed stimulation (representative example in Figure 1) was tendentially but not significantly increased [baseline: 2 episodes; 3 μM Dapa: 6 episodes (p = ns); 5 μM Dapa: 11 episodes (p = ns); 10 μM Dapa: 18 episodes (p = ns)] (Figure 2).

Dapagliflozin in a model of QT prolongation associated VA

Erythromycin (Ery) was employed to induce a medical LQT2 syndrome. The perfusion with Ery caused a significant prolongation of QT interval [baseline: 258 ± 40 ms; +300 μ M Ery: 286 ± 49 ms (p < 0.05)] and APD₉₀ (baseline: 195 ± 39 ms; +300 μ M Ery: 206 ± 44 ms (p < 0.05). The additional perfusion with Dapa did not cause a significant change in QT interval [+5 μ M Dapa: 284 ± 45 ms (p = 0.97)] and APD₉₀ [+5 μ M Dapa: 203 ± 40 ms (p = 0.15)]. These observations were paralleled



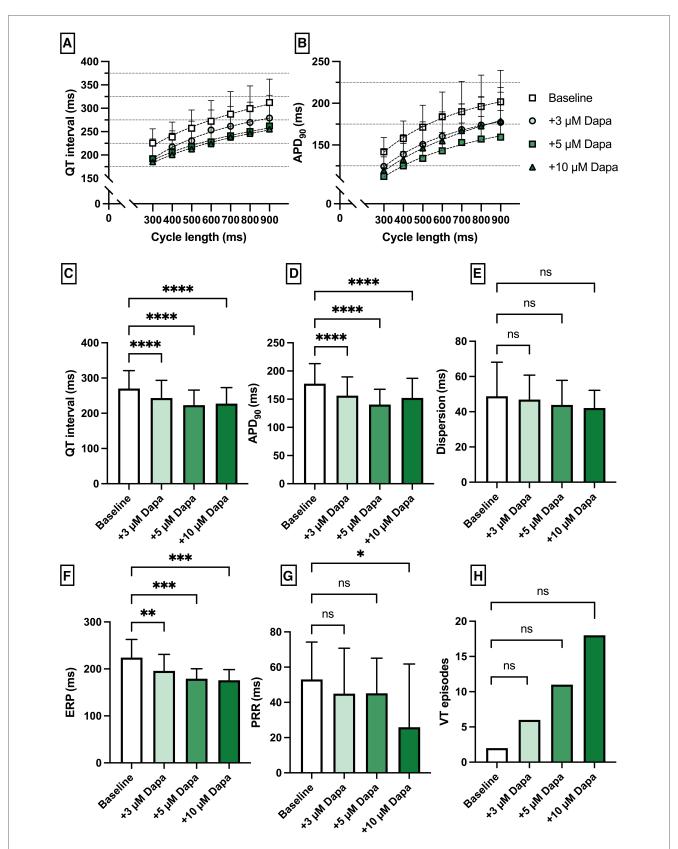


FIGURE 2
(A) Cycle-length dependent QT intervals and (B) action potential durations (APD₉₀) under baseline conditions (empty square) and after treatment with $3\,\mu\text{M}$ (light green circle), $5\,\mu\text{M}$ (green square) and $10\,\mu\text{M}$ (dark green triangle) dapagliflozin (dapa). (C) Overall QT interval and (D) APD₉₀. Concentration-dependent effect of dapagliflozin on (E) spatial dispersion of repolarization, (F) effective refractory period (ERP), (G) post-repolarization refractoriness (PRR) and (H) number of ventricular tachycardia (VT)/fibrillation (VF) induced by programmed ventricular stimulation $(*=p<0.005, **=p\leq0.001, **=p\leq0.001, **=p\leq0.0001)$.

by a significant increase in spatial dispersion of repolarization under Ery without significant changes under additional perfusion with Dapa. While ERP and PRR were not significantly altered under both perfusions the arrhythmia susceptibly under programmed stimulation was significantly increased under Ery [baseline: 1 episode; $+300 \mu M$ Ery: 36 episodes of torsade de pointes tachycardia (p < 0.05); $+5 \mu M$ Dapa: 33 episodes (p = ns)] (Figure 3).

Dapagliflozin in a model of QT abbreviation associated VA

Dapagliflozin in an atrial fibrillation model

Perfusion with 1 μ M isoproterenol and 1 μ M acetylcholine (IsoACh) was employed to raise susceptibility for atrial fibrillation (AF). Perfusion with IsoACh led to a significant shortening of aAPD₉₀ [baseline: 102 ± 19 ms; + 1 μ M IsoACh: 57 ± 9 ms (p < 0.05)] and perfusion with Dapa caused aAPD₉₀ prolongation [+3 μ M Dapa: 70 ± 15 ms (p < 0.05)]. The aERP was significantly shortened under IsoACh and returned to baseline duration under Dapa. The incidence of AF episodes was significantly increased under IsoACh and AF susceptibility was significantly suppressed by Dapa [baseline: 1 episode; 1 μ M IsoACh: 71 episodes (p < 0.05); + 3 μ M Dapa: 30 episodes (p < 0.05)] (Figure 5).

Empagliflozin

Perfusion with empagliflozin caused a significant shortening of QT intervals [baseline: 262 ± 39 ms; 1 μ M Empa: 232 ± 55 ms (p<0.05); 3 μ M Empa: 214 ± 39 ms (p<0.05); 5 μ M Empa: 193 ± 38 ms (p<0.05)] and APD₉₀ intervals [baseline: 172 ± 36 ms; 1 μ M Empa: 145 ± 31 ms (p<0.05); 3 μ M Empa: 127 ± 25 ms (p<0.05); 5 μ M Empa: 122 ± 28 ms (p<0.05)]. Spatial dispersion of repolarization was overall not significantly altered. Furthermore, perfusion with empagliflozin caused a significant

shortening of ERP while the PRR was not significantly altered. The incidence of ventricular arrhythmia episodes under programmed stimulation was tendentially increased while only the highest concentration of empagliflozin reached the threshold for statistical significance [baseline: 4 episodes; 1 μ M Empa: 12 episodes (p = ns); 3 μ M Empa: 8 episodes (p = ns); 5 μ M Empa: 29 episodes (p < 0.05)] (Figure 6).

Empagliflozin in a model of QT prolongation associated VA

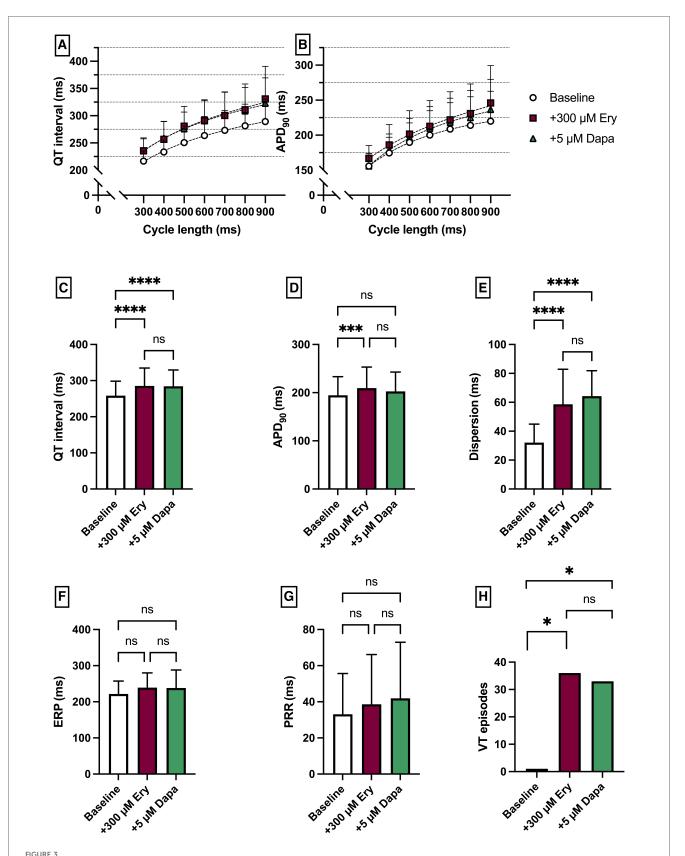
Perfusion with Ery caused a significant prolongation of the QT interval [baseline: 271 ± 42 ms; +300 μ M Ery: 287 ± 56 ms (p < 0.05)] and APD₉₀ [baseline: 186 ± 41 ms; +300 μ M Ery: 199 ± 48 ms (p < 0.05)]. The additional perfusion with Empa caused a significant shortening in QT interval [+3 μ M Empa: 277 ± 51 ms (p < 0.05)] and APD₉₀ [+3 μ M Empa: 172 ± 40 ms (p < 0.05)]. These observations were paralleled by a significant increase in spatial dispersion of repolarization under Ery without significant changes under additional perfusion with Empa. The ERP was significantly increased under Ery and not significantly altered under Empa. The PRR was not significantly altered under both perfusions. The arrhythmia susceptibly under programmed stimulation was significantly increased under Ery and significantly suppressed by Empa [baseline: 5 episodes; +300 μ M Ery: 41 episodes (p < 0.05); +3 μ M Empa: 17 episodes (p < 0.05)] (Figure 7).

Empagliflozin in a model of QT abbreviation associated VA

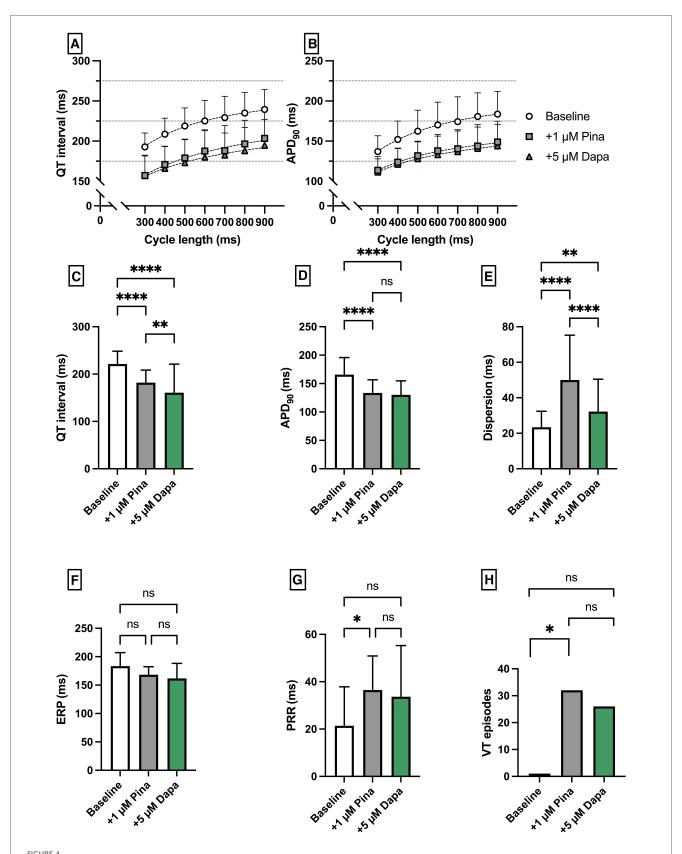
Perfusion with Pina caused a significant shortening of QT interval [baseline: $287 \pm 39 \text{ ms}$; $+ 1 \mu \text{M}$ Pina: $231 \pm 35 \text{ ms}$ (p < 0.05)] and APD₉₀ [baseline: $196 \pm 42 \text{ ms}$; $+ 1 \mu \text{M}$ Pina: $144 \pm 30 \text{ ms}$ (p < 0.05)]. The additional perfusion with Empa caused a significant shortening of the APD₉₀ interval [+3 μM Empa: $118 \pm 40 \text{ ms}$ (p < 0.05)] while the QT interval was not significantly altered [+3 μM Empa: $222 \pm 31 \text{ ms}$ (p = ns)]. The dispersion was significantly increased under Pina and not significantly altered under Empa. The incidence of ventricular arrhythmia episodes under perfusion with Pina [baseline: 3 episodes; $+ 1 \mu \text{M}$ Pina: -40 = pisodes (-10 = pisodes (-1

Empagliflozin in an atrial fibrillation model

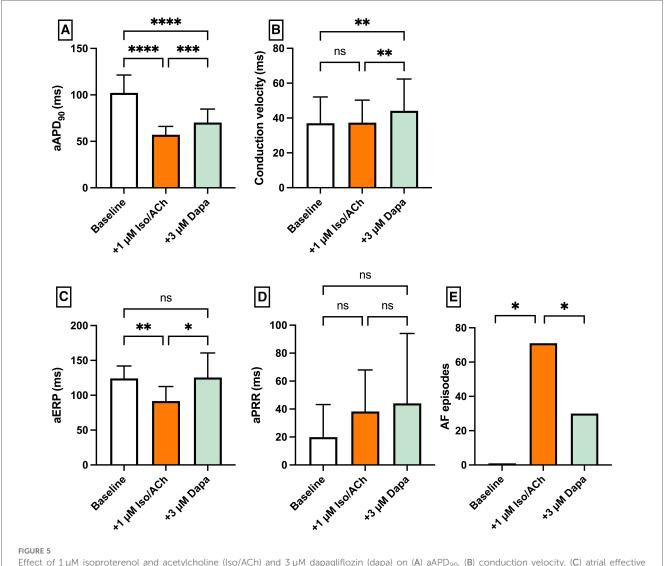
Perfusion with IsoACh led to a significant shortening of aAPD $_{90}$ [baseline: 109 ± 17 ms; +1 μ M IsoACh: 69 ± 20 ms (p<0.05)] while perfusion with Empa did not cause a significant change in aAPD $_{90}$ [+3 μ M Empa: 60 ± 12 ms (p=ns)]. Conduction velocity and eERP were not altered under both perfusions. The incidence of AF episodes was not significantly



(A) Cycle-length dependent QT intervals and (B) action potential durations (APD₉₀) under baseline conditions (empty circles) and after treatment with 300 μ M erythromycin (Ery) (purple square) and additional perfusion with 5 μ M (green triangle) dapagliflozin (dapa). (C) Overall QT interval and (D) APD₉₀. Effect of erythromycin and dapagliflozin on (E) spatial dispersion of repolarization, (F) effective refractory period (ERP), (G) post-repolarization refractoriness (PRR) and (H) number of ventricular tachycardia (VT)/fibrillation (VF) induced by programmed ventricular stimulation (* = p < 0.05, ** = $p \le 0.01$, ** = $p \le 0.001$, ** = $p \le 0.0001$).



(A) Cycle-length dependent QT intervals and (B) action potential durations (APD₉₀) under baseline conditions (empty circles) and after treatment with 1 μ M pinacidil (pina) (grey square) and additional perfusion with 5 μ M (green triangle) dapagliflozin (dapa). (C) Overall QT interval and (D) APD₉₀. Effect of pinacidil and dapagliflozin on (E) spatial dispersion of repolarization, (F) effective refractory period (ERP), (G) post-repolarization refractoriness (PRR) and (H) number of ventricular tachycardia (VT)/fibrillation (VF) induced by programmed ventricular stimulation (* = p < 0.05, ** = p ≤ 0.01, ** = p ≤ 0.001).



Effect of 1 μ M isoproterenol and acetylcholine (Iso/ACh) and 3 μ M dapagliflozin (dapa) on (A) aAPD₉₀, (B) conduction velocity, (C) atrial effective refractory period (aERP), (D) atrial post-repolarization refractoriness (aPRR) and (E) atrial fibrillation (AF) episodes (* = p < 0.05, ** = p ≤ 0.001, *** = p ≤ 0.0001).

increased under IsoACh and AF susceptibility was not significantly altered by Empa [baseline: 2 episodes; 1 μ M IsoACh: 27 episodes (p = ns); + 3 μ M Empa: 32 episodes (p = ns)] (Figure 9).

Discussion

The present experimental study on atrial and ventricular electrophysiology and arrhythmias of dapagliflozin and empagliflozin demonstrated the following main results:

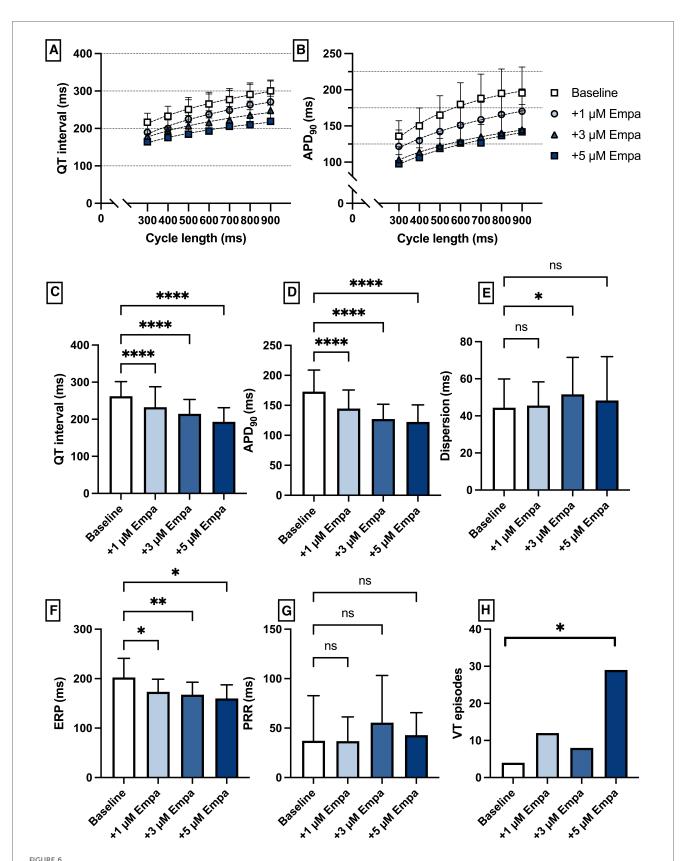
- (1) Empagliflozin showed antiarrhythmic effects in a model of QT prolongation associated proarrhythmia.
- (2) In a model of QT shortening as well as in an AF model, no significant effects of empagliflozin occurred.
- (3) In contrast, dapagliflozin showed no antiarrhythmic effects at the ventricular level in the drug-induced LQTS and SQTS.

However, significant antiarrhythmic effects were demonstrated in the atrial fibrillation model.

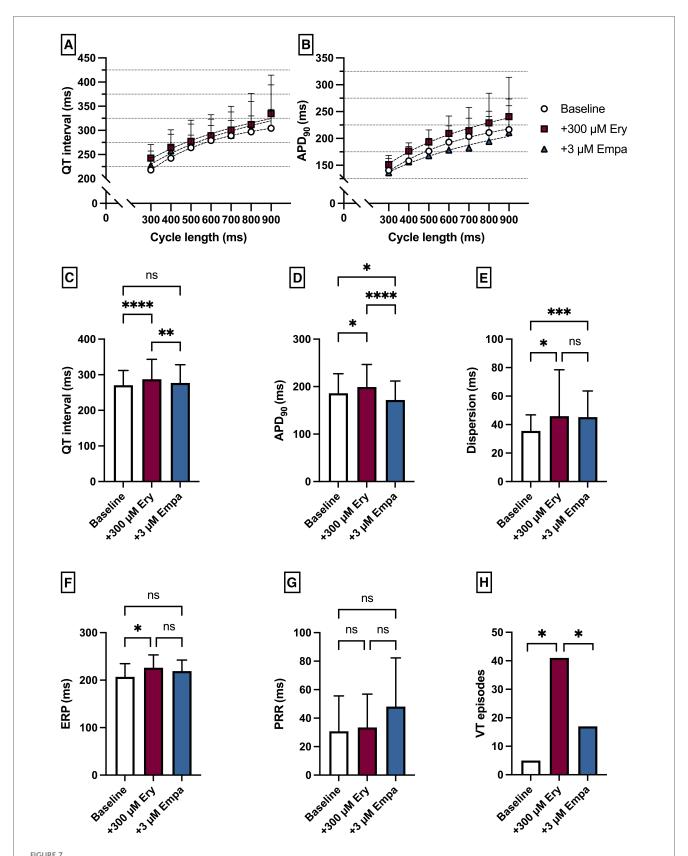
Noteworthy, the concentrations of dapagliflozin and empagliflozin used in this study are consistent with previously published plasma concentrations of dapagliflozin (22) and empagliflozin (23).

Electrophysiologic effects of dapagliflozin

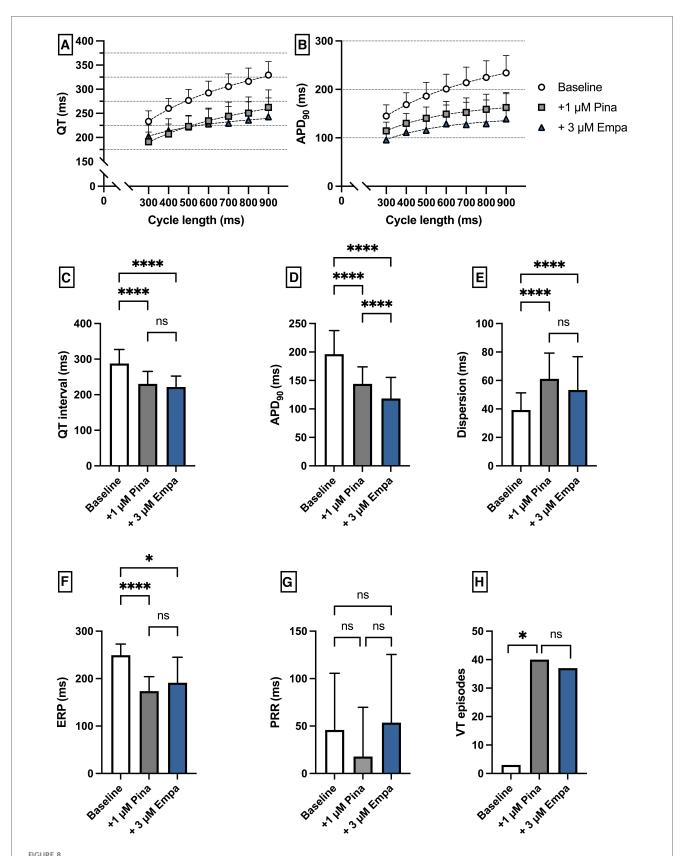
Perfusion with dapagliflozin significantly shortened cardiac repolarization with shortening of the QT interval and action potential duration. However, the number of VT episodes was not significantly reduced but rather stable with even a trend to more VT episodes. Similar effects of action potential shortening were demonstrated by Durak et al. (8) and Qin et al. (24), who



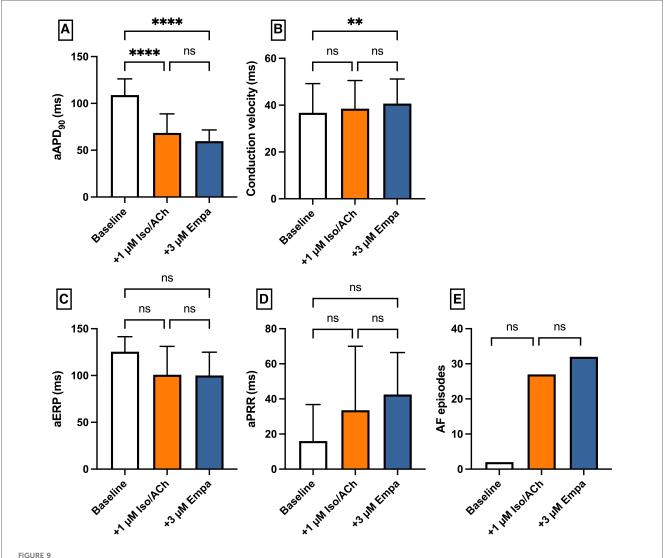
(A) Cycle-length dependent QT intervals and (B) action potential durations (APD₉₀) under baseline conditions (empty square) and after treatment with 1 μ M (light blue circle), 3 μ M (blue triangle) and 5 μ M (dark blue square) empagliflozin (empa). (C) Overall QT interval and (D) APD₉₀. Concentration-dependent effect of empagliflozin on (E) spatial dispersion of repolarization, (F) effective refractory period (ERP), (G) post-repolarization refractoriness (PRR) and (H) number of ventricular tachycardia (VT)/fibrillation (VF) induced by programmed ventricular stimulation (* = p < 0.05, ** = p ≤ 0.001, *** = p ≤ 0.0001).



(A) Cycle-length dependent QT intervals and (B) action potential durations (APD₉₀) under baseline conditions (empty circles) and after treatment with 300 μ M erythromycin (Ery) (purple square) and additional perfusion with 3 μ M (blue triangle) empagliflozin (empa). (C) Overall QT interval and (D) APD₉₀. Effect of erythromycin and empagliflozin on (E) spatial dispersion of repolarization, (F) effective refractory period (ERP), (G) post-repolarization refractoriness (PRR) and (H) number of ventricular tachycardia (VT)/fibrillation (VF) induced by programmed ventricular stimulation (* = p < 0.05, ** = $p \le 0.01$, ** = $p \le 0.001$, *** = $p \le 0.0001$).



(A) Cycle-length dependent QT intervals and (B) action potential durations (APD₉₀) under baseline conditions (empty circles) and after treatment with 1 μ M pinacidil (pina) (grey square) and additional perfusion with 3 μ M (blue triangle) empagliflozin (empa). (C) Overall QT interval and (D) APD₉₀. Effect of pinacidil and empagliflozin on (E) spatial dispersion of repolarization, (F) effective refractory period (ERP), (G) post-repolarization refractoriness (PRR) and (H) number of ventricular tachycardia (VT)/fibrillation (VF) induced by programmed ventricular stimulation (* = p < 0.05, ** = p ≤ 0.001, *** = p ≤ 0.001, *** = p ≤ 0.0001).



Effect of $1 \,\mu\text{M}$ isoproterenol and acetylcholine (Iso/ACh) and $3 \,\mu\text{M}$ empagliflozin (empa) on (A) aAPD₉₀, (B) conduction velocity, (C) atrial effective refractory period (aERP), (D) atrial post-repolarization refractoriness (aPRR) and (E) atrial fibrillation (AF) episodes (* = p < 0.05, ** = p \leq 0.01, *** = p \leq 0.001).

attributed them most likely to an induction of potassium channels. As expected, following the application of the IKr blocker erythromycin, a significant increase in QT interval and APD as well as dispersion occurred, which was accompanied by a significant increase in VA episodes. Perfusion with dapagliflozin did not result in any relevant alteration of APD and QT intervals in this setting, and there was no significant alteration in the number of VA episodes. It is possible that these differences between the previous studies and our observations result from the significantly shorter exposure time of dapagliflozin compared with the previous studies. Perfusion with the potassium channel opener pinacidil resulted in the expected decrease in APD and QT time with a parallel increase in arrhythmia incidence (25, 26). In the same model dapagliflozin showed no overall relevant effects on cardiac electrophysiology.

Atrial effects of dapagliflozin

In the AF model, dapagliflozin showed an effective reduction in AF episodes. Mechanisms here appear to be the prolongation of the effective refractory period and the restitution of action potential duration. In this study, dapagliflozin's electrophysiologic characteristics resemble those of different class I antiarrhythmic drugs (20, 25, 27). In line with this study, late sodium channel inhibition has been identified as a potential molecular target of different SGLT2 inhibitors in ventricular myocytes (28).

Ventricular effects of empagliflozin

Perfusion with empagliflozin significantly shortened QT time and action potential duration. However, the number of VA

episodes tended to be significantly increased only at high concentrations. Similar effects in terms of action potential shortening were also observed by Xue et al. (13) in a mouse model of myocardial infarction. Furthermore, perfusion with the IKr blocker erythromycin resulted in an increase in QT and APD with a concomitant increase in VA incidence. Here, additive perfusion with empagliflozin resulted in a reduction in QT and APD with a concomitant reduction in VA incidence. The antiarrhythmic effects observed in this model are consistent with observations by Baris et al, who showed that in rats in a sotalol-induced proarrhythmic environment, additive perfusion with empagliflozin led to a decrease in QT (12). Possible mechanisms of this observation in this case could be the previously described suppression of calcium sparks (29) and lowering of intracellular sodium (30). Pathologically prolonged QT is also proarrhythmic in the course of ischemic cardiomyopathies (31) and in animal models of heart failure with preserved ejection fraction (32), providing a possible explanation for direct antiarrhythmic effects of empagliflozin in these conditions. Perfusion with pinacidil resulted in the expected decrease in APD and QT interval with a parallel increase in arrhythmia incidence. The proarrhythmia in a short QT syndrome is usually due to a heterogeneous increase in the repolarizing currents with the resulting increase in dispersion as well as a shortened refractory period with consecutively increased ventricular vulnerability (25, 33). In this setting empagliflozin showed no overall relevant effects on cardiac electrophysiology.

Atrial effects of empagliflozin

In the atrial fibrillation model, empagliflozin did not show an effective reduction in AF episodes, in contrast to dapagliflozin. Conversely, the group of Shao et al. (11) demonstrated a significant reduction in AF episodes with longer-term empagliflozin use in a rat atrial fibrillation model. This seems most likely to be caused by the different durations of application with transcriptional effects and reduction of atrial fibrosis. Of note, a recent meta-analysis suggests a lower incidence of AF in patients treated with dapagliflozin compared to empagliflozin (34).

Limitations

Even though the Langendorff model of the isolated rabbit heart is an established model for the investigation of electrophysiological mechanisms (35), its applicability to the human heart remains limited. It should also be noted that variations in baseline values between the groups are most likely due to inter-individual differences and that there were significant differences in AF frequency between the dapagliflozin and empagliflozin groups. Nevertheless, the tendential effects remain largely unaffected by these differences. Furthermore, our model does not allow direct cellular electrophysiology or the observation of transcriptional effects due to the temporal relationship between perfusion and recording. This is of particular importance, as the group of Bers et al. (36) were able to demonstrate transcriptional inhibitory effects of empagliflozin on the late sodium channel influx $I_{\rm NaL}$.

Conclusion

Empagliflozin showed antiarrhythmic effects in the course of an erythromycin-induced LQT model. In the SQT model as well as the atrial fibrillation model, there was no pronounced effect of perfusion with empagliflozin.

Dapagliflozin showed no antiarrhythmic effects at the ventricular level in the drug-induced LQTS and SQTS. However, significant antiarrhythmic effects were demonstrated in the atrial fibrillation model.

Data availability statement

The datasets presented in this article are not readily available because data available on request from the authors. Requests to access the datasets should be directed to julian.wolfes@ukmuenster.de.

Ethics statement

The animal study was approved by Landesamt für Natur-, Umwelt- und Verbraucherschutz Nordrhein-Westfalen. The study was conducted in accordance with the local legislation and institutional requirements.

Author contributions

JW: Conceptualization, Data curation, Formal Analysis, Investigation, Writing – original draft, Writing – review & editing. JU: Data curation, Investigation, Writing – review & editing. SK: Data curation, Investigation, Writing – review & editing. FW: Formal Analysis, Validation, Writing – review & editing. BR: Formal Analysis, Methodology, Writing – review & editing. LE: Conceptualization, Data curation, Funding acquisition, Methodology, Project administration, Writing – review & editing. GF: Conceptualization, Data curation, Funding acquisition, Methodology, Project administration, Writing – review & editing. CE: Conceptualization, Data curation, Formal Analysis, Funding acquisition, Methodology, Project administration, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This research was funded by Hans und Gertie Fischer -Stiftung.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated

organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

- 1. McMurray JJV, Solomon SD, Inzucchi SE, Kober L, Kosiborod MN, Martinez FA, et al. Dapagliflozin in patients with heart failure and reduced ejection fraction. $N \, Engl \, J \, Med. \, (2019) \, 381(21):1995-2008. \, doi: \, 10.1056/NEJMoa1911303$
- 2. Packer M, Anker SD, Butler J, Filippatos G, Pocock SJ, Carson P, et al. Cardiovascular and renal outcomes with empagliflozin in heart failure. *N Engl J Med.* (2020) 383(15):1413–24. doi: 10.1056/NEJMoa2022190
- 3. Solomon SD, McMurray JJV, Claggett B, de Boer RA, DeMets D, Hernandez AF, et al. Dapagliflozin in heart failure with mildly reduced or preserved ejection fraction. $N\ Engl\ J\ Med.\ (2022)\ 387(12):1089-98.\ doi: 10.1056/NEJMoa2206286$
- 4. Anker SD, Butler J, Filippatos G, Ferreira JP, Bocchi E, Bohm M, et al. Empagliflozin in heart failure with a preserved ejection fraction. *N Engl J Med.* (2021) 385(16):1451–61. doi: 10.1056/NEJMoa2107038
- 5. McDonagh TA, Metra M, Adamo M, Gardner RS, Baumbach A, Bohm M, et al. 2023 focused update of the 2021 ESC guidelines for the diagnosis and treatment of acute and chronic heart failure. *Eur Heart J.* (2023) 44(37):3627–39. doi: 10.1093/eurheartj/ehad195
- 6. Fernandes GC, Fernandes A, Cardoso R, Penalver J, Knijnik L, Mitrani RD, et al. Association of SGLT2 inhibitors with arrhythmias and sudden cardiac death in patients with type 2 diabetes or heart failure: a meta-analysis of 34 randomized controlled trials. Heart Rhythm. (2021) 18(7):1098–105. doi: 10.1016/j.hrthm.2021. 03.028
- 7. Wu J, Liu T, Shi S, Fan Z, Hiram R, Xiong F, et al. Dapagliflozin reduces the vulnerability of rats with pulmonary arterial hypertension-induced right heart failure to ventricular arrhythmia by restoring calcium handling. *Cardiovasc Diabetol.* (2022) 21(1):197. doi: 10.1186/s12933-022-01614-5
- 8. Durak A, Olgar Y, Degirmenci S, Akkus E, Tuncay E, Turan B. A SGLT2 inhibitor dapagliflozin suppresses prolonged ventricular-repolarization through augmentation of mitochondrial function in insulin-resistant metabolic syndrome rats. *Cardiovasc Diabetol.* (2018) 17(1):144. doi: 10.1186/s12933-018-0790-0
- 9. Wang FZ, Wei WB, Li X, Huo JY, Jiang WY, Wang HY, et al. The cardioprotective effect of the sodium-glucose cotransporter 2 inhibitor dapagliflozin in rats with isoproterenol-induced cardiomyopathy. *Am J Transl Res.* (2021) 13 (9):10950–61.
- 10. Hu Z, Ju F, Du L, Abbott GW. Empagliflozin protects the heart against ischemia/reperfusion-induced sudden cardiac death. *Cardiovasc Diabetol.* (2021) 20(1):199. doi: 10.1186/s12933-021-01392-6
- 11. Shao Q, Meng L, Lee S, Tse G, Gong M, Zhang Z, et al. Empagliflozin, a sodium glucose co-transporter-2 inhibitor, alleviates atrial remodeling and improves mitochondrial function in high-fat diet/streptozotocin-induced diabetic rats. *Cardiovasc Diabetol.* (2019) 18(1):165. doi: 10.1186/s12933-019-0964-4
- 12. Baris VO, Dincsoy AB, Gedikli E, Zirh S, Muftuoglu S, Erdem A. Empagliflozin significantly prevents the doxorubicin-induced acute cardiotoxicity via non-antioxidant pathways. *Cardiovasc Toxicol.* (2021) 21(9):747–58. doi: 10.1007/s12012-021-09665-v
- 13. Xue G, Yang X, Zhan G, Wang X, Gao J, Zhao Y, et al. Sodium-glucose cotransporter 2 inhibitor empagliflozin decreases ventricular arrhythmia susceptibility by alleviating electrophysiological remodeling post-myocardial-infarction in mice. Front Pharmacol. (2022) 13:988408. doi: 10.3389/fphar.2022. 988408
- Mustroph J, Wagemann O, Lucht CM, Trum M, Hammer KP, Sag CM, et al. Empagliflozin reduces Ca/calmodulin-dependent kinase II activity in isolated ventricular cardiomyocytes. ESC Heart Fail. (2018) 5(4):642–8. doi: 10.1002/ehf2.
- 15. Azam MA, Chakraborty P, Si D, Du B, Masse S, Lai PFH, et al. Anti-arrhythmic and inotropic effects of empagliflozin following myocardial ischemia. *Life Sci.* (2021) 276:119440. doi: 10.1016/j.lfs.2021.119440
- 16. Wolfes J, Ellermann C, Kirchner LM, Willy K, Rath B, Leitz PR, et al. Electrophysiological safety profile of antiestrogenic therapies in the isolated rabbit heart. *Pharmacology.* (2022) 107(11–12):608–14. doi: 10.1159/000526612
- 17. Milberg P, Reinsch N, Osada N, Wasmer K, Monnig G, Stypmann J, et al. Verapamil prevents torsade de pointes by reduction of transmural dispersion of

repolarization and suppression of early afterdepolarizations in an intact heart model of LQT3. *Basic Res Cardiol.* (2005) 100(4):365–71. doi: 10.1007/s00395-005-0533-8

- 18. Frommeyer G, Milberg P, Witte P, Stypmann J, Koopmann M, Lucke M, et al. A new mechanism preventing proarrhythmia in chronic heart failure: rapid phase-III repolarization explains the low proarrhythmic potential of amiodarone in contrast to sotalol in a model of pacing-induced heart failure. *Eur J Heart Fail.* (2011) 13 (10):1060–9. doi: 10.1093/eurjhf/hfr107
- 19. Milberg P, Hilker E, Ramtin S, Cakir Y, Stypmann J, Engelen MA, et al. Proarrhythmia as a class effect of quinolones: increased dispersion of repolarization and triangulation of action potential predict torsades de pointes. *J Cardiovasc Electrophysiol.* (2007) 18(6):647–54. doi: 10.1111/j.1540-8167.2007.00793.x
- 20. Frommeyer G, Schmidt M, Clauss C, Kaese S, Stypmann J, Pott C, et al. Further insights into the underlying electrophysiological mechanisms for reduction of atrial fibrillation by ranolazine in an experimental model of chronic heart failure. *Eur J Heart Fail.* (2012) 14(12):1322–31. doi: 10.1093/eurjhf/hfs163
- 21. Sicouri S, Gianetti B, Zygmunt AC, Cordeiro JM, Antzelevitch C. Antiarrhythmic effects of simvastatin in canine pulmonary vein sleeve preparations. *J Am Coll Cardiol.* (2011) 57(8):986–93. doi: 10.1016/j.jacc.2010.08.649
- 22. Kasichayanula S, Liu X, Lacreta F, Griffen SC, Boulton DW. Clinical pharmacokinetics and pharmacodynamics of dapagliflozin, a selective inhibitor of sodium-glucose co-transporter type 2. Clin Pharmacokinet. (2014) 53(1):17–27. doi: 10.1007/s40262-013-0104-3
- 23. Scheen AJ. Pharmacokinetic and pharmacodynamic profile of empagliflozin, a sodium glucose co-transporter 2 inhibitor. *Clin Pharmacokinet*. (2014) 53 (3):213–25. doi: 10.1007/s40262-013-0126-x
- 24. Qin T, Kong B, Dai C, Xiao Z, Fang J, Shuai W, et al. Protective effects of dapagliflozin on the vulnerability of ventricular arrhythmia in rats with pulmonary artery hypertension induced by monocrotaline. *Bioengineered.* (2022) 13 (2):2697–709. doi: 10.1080/21655979.2021.2017652
- 25. Milberg P, Tegelkamp R, Osada N, Schimpf R, Wolpert C, Breithardt G, et al. Reduction of dispersion of repolarization and prolongation of postrepolarization refractoriness explain the antiarrhythmic effects of quinidine in a model of short QT syndrome. *J Cardiovasc Electrophysiol.* (2007) 18(6):658–64. doi: 10.1111/j.1540-8167.2007.00813.x
- 26. Frommeyer G, Ellermann C, Dechering DG, Kochhauser S, Bogeholz N, Guner F, et al. Ranolazine and vernakalant prevent ventricular arrhythmias in an experimental whole-heart model of short QT syndrome. *J Cardiovasc Electrophysiol.* (2016) 27(10):1214–9. doi: 10.1111/jce.13029
- 27. Frommeyer G, Garthmann J, Ellermann C, Dechering DG, Kochhauser S, Reinke F, et al. Broad antiarrhythmic effect of mexiletine in different arrhythmia models. *Europace*. (2018) 20(8):1375–81. doi: 10.1093/europace/eux221
- 28. Philippaert K, Kalyaanamoorthy S, Fatehi M, Long W, Soni S, Byrne NJ, et al. Cardiac late sodium channel current is a molecular target for the sodium/glucose cotransporter 2 inhibitor empagliflozin. *Circulation*. (2021) 143(22):2188–204. doi: 10.1161/CIRCULATIONAHA.121.053350
- 29. Lee TI, Chen YC, Lin YK, Chung CC, Lu YY, Kao YH, et al. Empagliflozin attenuates myocardial sodium and calcium dysregulation and reverses cardiac remodeling in streptozotocin-induced diabetic rats. *Int J Mol Sci.* (2019) 20(7):1680. doi: 10.3390/ijms20071680
- 30. Baartscheer A, Schumacher CA, Wust RC, Fiolet JW, Stienen GJ, Coronel R, et al. Empagliflozin decreases myocardial cytoplasmic na(+) through inhibition of the cardiac na(+)/H(+) exchanger in rats and rabbits. *Diabetologia*. (2017) 60 (3):568–73. doi: 10.1007/s00125-016-4134-x
- 31. Makielski JC. Late sodium current: a mechanism for angina, heart failure, and arrhythmia. *Trends Cardiovasc Med.* (2016) 26(2):115–22. doi: 10.1016/j.tcm.2015. 05.006
- 32. Cho JH, Zhang R, Kilfoil PJ, Gallet R, de Couto G, Bresee C, et al. Delayed repolarization underlies ventricular arrhythmias in rats with heart failure and preserved ejection fraction. *Circulation*. (2017) 136(21):2037–50. doi: 10.1161/CIRCULATIONAHA.117.028202

- 33. Gaita F, Giustetto C, Bianchi F, Wolpert C, Schimpf R, Riccardi R, et al. Short QT syndrome: a familial cause of sudden death. *Circulation*. (2003) 108(8):965–70. doi: 10.1161/01.CIR.0000085071.28695.C4
- 34. Okunrintemi V, Mishriky BM, Powell JR, Cummings DM. Sodium-glucose co-transporter-2 inhibitors and atrial fibrillation in the cardiovascular and renal outcome trials. *Diabetes Obes Metab.* (2021) 23(1):276–80. doi: 10.1111/dom.14211
- 35. Ellermann C, Wolfes J, Eckardt L, Frommeyer G. Role of the rabbit whole-heart model for electrophysiologic safety pharmacology of non-cardiovascular drugs. *Europace*. (2021) 23(6):828–36. doi: 10.1093/europace/euaa288
- 36. Hegyi B, Mira Hernandez J, Shen EY, Habibi NR, Bossuyt J, Bers DM. Empagliflozin reverses late na(+) current enhancement and cardiomyocyte proarrhythmia in a translational murine model of heart failure with preserved ejection fraction. *Circulation*. (2022) 145(13):1029–31. doi: 10.1161/CIRCULATIONAHA.121.057237