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Inflammation as the nexus: exploring the link between acute myocardial infarction and chronic obstructive pulmonary disease

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Chronic obstructive pulmonary disease (COPD), particularly following acute exacerbations (AE-COPD), significantly heightens the risks and mortality associated with acute myocardial infarction (AMI). The intersection of COPD and AMI is characterised by a considerable overlap in inflammatory mechanisms, which play a crucial role in the development of both conditions. Although extensive research has been conducted on individual inflammatory pathways in AMI and COPD, the understanding of thrombo-inflammatory crosstalk in comorbid settings remains limited. The effectiveness of various inflammatory components in reducing AMI infarct size or slowing COPD progression has shown promise, yet their efficacy in the context of comorbidity with COPD and AMI is not established. This review focuses on the critical importance of both local and systemic inflammation, highlighting it as a key pathophysiological connection between AMI and COPD/AE-COPD.

KEYWORDS

chronic obstructive pulmonary disease, acute myocardial infarction, inflammation, antiinflammatory, interleukins, S100A8/A9

Introduction

Chronic obstructive pulmonary disease (COPD) is the third leading cause of death worldwide, with cardiovascular diseases (CVDs) ultimately contributing for approximately 50% of these deaths. Individuals with COPD have a significantly higher risk of acute myocardial infarction (AMI) and increased mortality from AMI (1), particularly during the first month following an acute exacerbation (AE)-COPD event. Indeed, AE-COPD patients are at an eightfold increased risk of suffering from an AMI in the first 3 days following hospitalisation (2). This is largely due to inflammation, both local and systemic, which is a major pathophysiological mechanistic link between AE-COPD/COPD and AMI incidence/mortality. While the contribution of various inflammatory pathways has been studied separately in each of these vascular beds, there is a lack of understanding around the thrombo-inflammatory crosstalk between these organs in health and disease. In this review we will explore: (i) the inflammatory factors involved in the interactions in and between COPD/AE-COPD and AMI; (ii) antiinflammatories used in COPD/AE-COPD and AMI separately; and (iii) the unmet clinical needs in the treatment of patients with COPD with AMI.

Inflammatory mechanisms in acute myocardial infarction

The onset of an ischaemic event during an AMI, and reperfusion through primary percutaneous coronary intervention (PCI), can each induce inflammation because of cellular injury and death to cardiac myocytes, endothelial cells, and fibroblasts. This inflammation initiates through various processes, including the release of necrotic cell contents, reactive oxygen species (ROS) production, and activation of the complement system. Biomarkers such as extracellular DNA, mtDNA, RNA, and HMGB1, known to be elevated post-AMI, play dual roles in inflammation and cardioprotection, as indicated by the effectiveness of DNAase, RNAase1, or HMGB1 inhibitors in reducing MI severity (3, 4). The inflammatory cascade involves the production of chemokines, cytokines, and adhesion molecules at the infarct site (Table 1), followed by the recruitment of neutrophils and other leukocytes, aiding in wound healing and scar formation (22). However, a prolonged and excessive inflammatory response can worsen the condition, leading to larger infarct size, left ventricular (LV) remodeling, and heart failure (Figure 1). The inflammatory response following myocardial ischemia-reperfusion (IR) injury is mediated through the coordinated activity of several types of cells, both within and distant to the infarcted area. A hallmark among patients with AMI is leucocytosis which has been widely used as a predictor of mortality within this group of patients. Neutrophils are thought to be the first type of cells to be recruited during myocardial IR injury, followed by monocytes, mast cells, and lymphocytes. Each of these cells has a distinct role within the inflammatory process during myocardial IR injury (23, 24).

Neutrophils

In an AMI, neutrophil counts are found to be significantly raised as a result of mobilization from the bone marrow, and this is used as a predictor of adverse outcomes and mortality (25). Significant neutrophil recruitment occurs in which their phagocytic functions serve to contribute to the clearance of debris. However, the vast array of hydrolytic, oxidative, and poreforming molecules released by an overabundance of phagocytic neutrophils, alongside their delayed removal from the infarct site, end up causing significant collateral heart tissue destruction (7, 26). Circulating neutrophil counts peak within 1-3 days and drop after day 4 (Figure 1). Neutrophils are recruited to the infarcted area within hours following myocardial IR injury. In 2016, Ma et al. further showed that neutrophils harvested from the heart on day 1 post-MI were N1 neutrophils (proinflammatory), that express highly pro-inflammatory mediators, whereas those isolated between days 5-7 were N2 (antiinflammatory) neutrophils (8, 27). Strategies such as neutrophil CD11/CD18 inhibition have been effective in reducing MI size in animal models (28). Recent studies highlighted neutrophils as a source of S100A8/A9, and its timely pharmacological inhibition was shown to improve cardiac function (18, 29).

Monocytes & macrophages

Following an AMI, monocytes from the bone marrow and spleen are mobilised into the circulation and drawn to the injury site by chemotaxis. Peripheral monocytosis serves as an indicator of potential adverse LV remodeling (30). Murine studies showed that pro-inflammatory classical, or Ly6C^{high} monocytes, peak in numbers between days 3 and 4. These monocytes then differentiate into pro-inflammatory M1 macrophages, responsible for producing proteases (MMPs),

TABLE 1 Inflammatory mediators in chronic obstructive pulmonary disease and acute myocardial infarction.

Inflammatory mediator	Chronic obstructive pulmonary disease	Acute myocardial infarction	References
Targets which hav	ve undergone clinical trials		
IL-1	Promotes emphysema onset and airway remodeling. Upregulated in AE-COPD	Upregulates pro-inflammatory mediators, adhesion molecules and leucocyte recruitment.	(5-8)
IL-6	Promotes emphysema onset, pulmonary fibrosis and increases epithelial permeability. Upregulated in AE-COPD. Pre-clinical.	Upregulates adhesion molecules and leucocyte integrins. Endothelial injury and vascular dysfunction. Activates T-cells and is important for infarct healing	(9, 10)
TNF-α	Promotes upregulation of neutrophilic cytokines and MMPs. Upregulated in AE-COPD	Promotes immune cell extravasation	(7, 11, 12)
Targets which have	ve undergone preclinical studies		
IL-8	Induces lung inflammation, promotes chemotaxis, mucus secretion and airway obstruction. Upregulated in AE-COPD	Promotes neutrophil and monocyte recruitment	(7, 8, 12)
IL-17	Pro-inflammatory, induces fibrosis, stimulates mast cells, promotes neutrophil recruitment, protease release and mucus secretion. Upregulated in AE-COPD	Promotes cardiomyocyte apoptosis and neutrophil recruitment	(13, 14)
IL-36	Pro-inflammatory, promotes neutrophil recruitment, mucus hypersecretion, emphysema, and small airway remodeling	Pro-inflammatory, promotes chemotaxis, endothelial and cardiomyocytes oxidative stress, cardiac fibrosis, and VCAM-1 upregulation	(15–17)
S100A8/9	Activates inflammasomes, induce release of pro-inflammatory mediator. Upregulated in AE-COPD	Activates inflammasomes, promotes IL-1 β production, immune cell recruitment and apoptosis	(18-20)
TGF-β	Induces emphysema and airway remodeling. Promotes ECM secretion and proliferation of fibroblast and smooth muscle cells	Influences post-infarct remodeling	(7, 21)

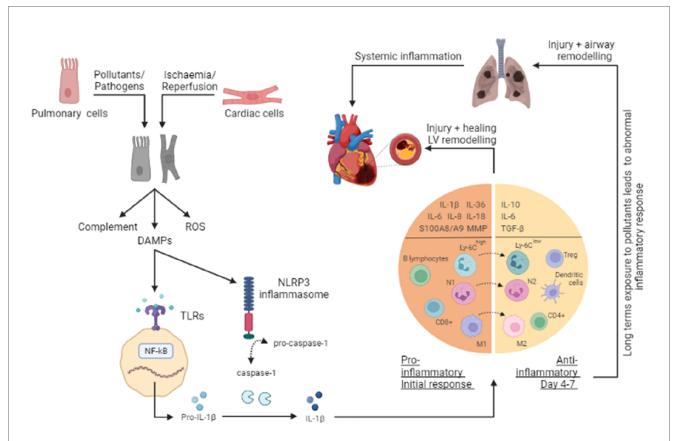


FIGURE 1

Overview of the inflammatory response in a COPD and AMI. The onset of either acute myocardial infarction (AMI) or chronic obstructive pulmonary disease (COPD) leads to cellular injury and necrosis of cardiac cells and pulmonary cells respectively, resulting in the release of intracellular content such as damage-associated molecular patterns (DAMPs), production of reactive oxygen species (ROS) and activation of the complement cascade. Once a DAMP (such as DNA, mitochondrial DNA (mtDNA), RNA, and the nuclear factor - high mobility group box 1 (HMGB1), and ATP) is recognised by toll-like receptors (TLRs), cytoplasmic myeloid differentiation factor 88 (MyD88) is activated. This in turn, stimulates the NF- κ B pathway and through a signaling cascade and transcription, leads to the release of several inflammatory mediators such as pro-interleukin (IL)- 1β . Additionally, in response to the release of DAMPs, inflammasomes (such as nucleotide-binding oligomerization domain receptor family protein 3, NLRP3) are activated to promote the cleavage of pro-Caspase-1 into Caspase-1, which in turn processes the pro-IL- 1β into the active IL- 1β form. As a result, pro-inflammatory mediators (IL- 1β , IL- 1α , IL-6, IL-8, IL-18, IL-16, IL-1

cytokines (IL-1, IL-6, TNF-α, IFN-γ), chemokines (CCL2), and growth factors, aiding in debris clearance and initiating wound healing (31). Subsequently, the recruitment of anti-inflammatory non-classical, or Ly6C^{low} monocytes, takes place, peaking around day seven (8, 24). As M1 macrophage numbers diminish, M2 anti-inflammatory macrophages take over, orchestrating wound healing in the subsequent reparative phase (Figure 1). However, a prolonged presence of M1 macrophages can intensify the pro-inflammatory phase, aggravate damage beyond the initially infarcted area, and lead to adverse LV remodeling. This underlines the potential cardioprotective benefits of pharmacologically inhibiting M1 macrophages (8, 32). Furthermore, monocytes can differentiate into antigenpresenting dendritic cells, initiating the adaptive immune system through the production of chemokines and cytokines (IFN-7, IL-6), which in turn activate T cells (9, 33).

Lymphocytes

Lymphocytes are key players in the cell-mediated cytotoxic innate immune response, with various subsets actively involved. In animal models, both T and B lymphocyte subsets have been observed infiltrating the injury site. Following an AMI, patients display an increase in pro-inflammatory CD4⁺ Th1 and cytotoxic CD8⁺ T cells and a reduction in anti-inflammatory and protective CD4⁺ Th2 cells. The prolonged presence of these detrimental lymphocytes is linked to poorer patient prognosis (34, 35). Moreover, an increase in peripheral B cells has been noted in AMI cases, with mature B cell infiltration peaking around day five. Zouggari et al., (36), demonstrated that B cells release CCL7, which mobilizes bone marrow Ly6C^{high} monocytes to the injury site, exacerbating tissue damage. They also demonstrated a potential therapeutic role following B cell genetic depletion, as evidenced by reduced systemic inflammation, LV remodeling and MI size (36). Additionally,

natural killer (NK) cells and their receptor expressions are significantly reduced following an MI, with recent studies indicating a protective role for these cells in atherosclerosis (37).

Platelets

Platelets are well-known for their critical role in thrombosis. Recently, they have also been identified as key players in inflammation, contributing to the innate immune response. They aggregate in injured areas, localising the inflammatory response and aiding in the formation of a provisional matrix. During inflammatory events like AMI or post-reperfusion, platelets release inflammatory mediators such as cytokines, chemokines, and platelet-derived growth factor (PDGF). Their pro-inflammatory activities include upregulating ICAM-1, VCAM-1, and selectins on leukocytes, triggering ROS production, activating macrophages and cytotoxic lymphocytes, and enhancing circulating microparticle production (38, 39). Studies also shows a link between innate immunity, platelet activation, and coagulation, known as immunothrombosis. Its abnormal activation in cardiovascular diseases increases the risk of cardiovascular events, including AMI (40). Extracellular DNA, a trigger for immunothrombosis in AMI, has been identified, with studies highlighting the potential of DNase I treatment to reduce infarct size in myocardial ischemia models (41). Moreover, platelets can influence leukocyte functions by forming platelet-leukocyte (P-L) aggregates through P-selectin/ PSGL-1 interactions. An increase in P-L aggregates in peripheral blood has been noted post-AMI, suggesting their potential as early biomarkers for MI (42). Additionally, targeting these P-L aggregates with anti-platelet therapies has shown effectiveness in reducing inflammation and subsequent complications in MI patients.

Inflammatory mechanisms in chronic obstructive pulmonary disease

Harmful particles and gases, notably in cigarette smoke, trigger inflammation and damage lung cells. Prolonged exposure leads to an abnormal and enhanced inflammatory response, hallmark features of COPD. This altered response contributes to tissue destruction (emphysema), increased mucus production (chronic bronchitis), and fibrosis (bronchiolitis) (43). Imbalances between oxidants and antioxidants, as well as proteases and antiproteases, are central to COPD development, exacerbating inflammation and lung tissue damage (44, 45). Even after the cessation of harmful exposure, the inflammatory process in COPD remains persistent and progressive, especially once symptomatic (46). This is attributed to airway remodeling, impaired macrophage clearance, oxidative stress, and hypoxia (47). Moreover, COPD is linked to a systemic inflammatory response, particularly notable in advanced stages or during AE-COPD, heightening the risk of associated comorbidities (48).

Neutrophils

In COPD patients, high neutrophil counts correlate with poor outcomes and increased mortality (49, 50). However, these

neutrophils exhibit compromised chemotaxis and release inflammatory mediators that contribute to alveolar damage (Figure 1) (51, 52). Both neutrophils and macrophages are key sources of MMPs, which are implicated in the breakdown of the extracellular matrix (ECM), airway obstruction, remodeling, and the development of emphysema (53, 54). MMP-12, in particular, is crucial for emphysema formation, as evidenced by MMP-12 deficient mice not developing emphysema post-cigarette smoke (CS) exposure (55). In COPD patients, MMP-2 and MMP-9 are potential biomarkers, with the highest expression levels observed in severe COPD cases (56). Specifically, the MMP-9 to tissue inhibitors of MMP-1 (TIMP-1) ratio has been linked to an increased mortality risk, indicating that targeting MMP-9 may help reduce this risk (57). Additionally, MMP-2 is implicated in ECM remodelling (58). Neutrophil elastase (NE), a neutrophilderived serine protease, also plays a significant role in ECM degradation, influencing mucus secretion and fibroblast proliferation, which further contribute to airway obstruction and bronchial fibrosis (59, 60). Mice deficient in NE showed resistance to emphysema development in a CS exposure model (61). Moreover, excessive formation of neutrophil extracellular traps (NETs) has been associated with alveolar destruction (62).

Monocytes & macrophages

In COPD, heightened macrophage recruitment within airways and lungs is a predictor of alveolar damage and disease progression (63). Pro-inflammatory M1 macrophages, prevalent in emphysematous regions, release a variety of inflammatory mediators. These mediators drive alveolar destruction via autocrine and paracrine pathways (Figure 1) (64-66). IL-1β and TNF-α intensify inflammation and lung tissue damage by upregulating neutrophilic cytokines and MMPs (Table 1) (5, 11). Mice lacking the IL-1 receptor exhibited resistance to emphysema and small airway remodeling in a CS exposure model (6). However, a clinical trial using an anti-IL-1R1 monoclonal antibody did not enhance lung function or quality of life in COPD patients (67). Recent studies have also delved into the role of the IL-36 sub-family in COPD (68, 69). IL-36 cytokines, primarily expressed in small airway cells, macrophages, and fibroblasts, are implicated in neutrophil recruitment, mucus hypersecretion, emphysema, and airway remodeling. Notably, COPD patients exhibit exacerbated symptoms due to a reduction in endogenous IL-36Ra in macrophages (15). IL-8, produced by macrophages, small airway cells, and endothelial cells, is also involved in mucus secretion and airway obstruction (70). Mice deficient in the IL-8 receptor, CXCR2, demonstrated protection against lung inflammation and DNA damage in a CS exposure model (71). IL-18 is notably elevated in the lungs of COPD patients, particularly in alveolar macrophages and epithelia, and its serum levels are higher compared to smokers and nonsmokers (72). This increase negatively correlates with pulmonary function and is linked to enhanced IFNy production in severe COPD, contributing to remodeling responses (73). Similarly, IL-32 levels correlate positively with COPD severity, suggesting its potential as a biomarker for disease severity (74).

Lymphocytes

COPD activates the adaptive immune system, characterised by the presence of B, T, and T-helper type 17 (Th17) cells, and a decrease in regulatory T-cells (Tregs) in the airways (Figure 1) (75). The progression of the disease sees the formation of lymphoid follicles, comprising B and T-cells, in the walls of small airways (76). Mice deficient in T and B-cells exhibited significantly reduced airway remodeling (77). A recent study noted an increased ratio of CD8⁺ to CD4⁺ cells in COPD patients, indicating a shift towards Th1/Tc1 dominance, which was associated with COPD exacerbation (78). Th17 cells, prominent producers of pro-inflammatory IL-17, are elevated in advanced COPD stages and contribute to CS-induced lymphoid neogenesis (13). Importantly, mice lacking the IL-17 receptor were protected from airway inflammation and fibrosis (14).

Platelets

Platelets contribute to COPD-related inflammation through lung elastin breakdown and the formation of platelet aggregates (79). Elevated platelet counts and reactivity are associated with increased lung inflammation, reduced lung function, and greater disease severity in COPD patients (80). COPD is linked to a hypercoagulable state marked by raised coagulation factor levels, leading to a prothrombotic effect. This state features increased fibrinogen and coagulation factors, alongside decreased coagulation inhibitors. Systemic and neutrophilic inflammation contribute to these prothrombotic changes (81). While the exact role of immunothrombosis in COPD is still under investigation, it's believed to contribute to pathogenesis, potentially through NETs (82).

Other inflammatory cells

In COPD, mast cells and eosinophils play significant roles in inflammation. Changes in mast cell populations in COPD patients include an increase in connective tissue mast cells (MC_{TC}) and a decrease in mucosal mast cells (MC_{T}), which are associated with airway remodeling and reduced lung function (83). The molecular mechanisms of mast cell involvement in COPD are still under investigation, but current hypotheses include their role in neutrophil recruitment, mucus hypersecretion through mast cell chymase, and tissue damage via enzymes like elastase, tryptase, and chymase (84). Eosinophil counts and related proteins may rise in certain COPD patients, particularly those with concurrent asthma (85, 86), although other studies report no significant increase or activation of eosinophils (86).

Inflammatory mechanisms in acute exacerbations of chronic obstructive pulmonary disease

A hallmark of COPD is the occurrence of acute exacerbations, typically triggered by infections (bacterial or viral) or environmental factors (pollutants). These exacerbations are characterised by a sudden worsening of respiratory symptoms and an elevated inflammatory response. Consequences include aggravated mucus hypersecretion, bronchoconstriction, impaired gas exchange, respiratory muscle fatigue, hypoxemia, hypercapnia, respiratory acidosis, and compromised vascular perfusion. The frequency of AE-COPD is closely associated with declining lung function and increased mortality (87). Neutrophils predominantly mediate the inflammatory response in AE-COPD, although eosinophilia has also been linked to AE-COPD and its risk (88, 89). Elevated neutrophil counts have been documented in bronchoalveolar lavage fluid (BALF), bronchial walls, pulmonary microcirculation, and airways (90). During AE-COPD, various inflammatory mediators show increased levels compared to stable COPD, including serum CXCL8, e-selectin, TNF-α, IL-6, and more recently, MMP-9, IL-1β, IL-8, IL-17, IL-18, and S100A8/A9 (Table 1) (91-93). Serum IL-8 has proven more sensitive than TNF-α or IL-6 in predicting AE-COPD (12). Additionally, a clinical correlation has been established between increased NK cell presence and AE-COPD (94). In a mouse model of AE-COPD (CS exposure and viral infection), NK cells were found to produce more IFN-7 (95). Furthermore, evidence suggests that inflammatory mediators from the lungs can spillover into systemic circulation, causing chronic, non-resolving, low-grade systemic inflammation. This condition potentially leads to cardiovascular comorbidities, muscle weakness, and other systemic issues (21). The mechanism of this spillover is under debate. Currently, it is hypothesized to involve local activation of inflammatory pathways, elevating various inflammatory markers such as IL-6, IL-1β, TNF-α, CRP, and inflammatory cells in the peripheral blood (21, 96). Furthermore, the mobilization of inflammatory cells from the bone marrow, driven by factors like granulocyte-colony-stimulating factor (G-CSF), is believed to contribute to the increased presence of these cells in peripheral blood (97, 98). AE-COPD are also linked to an inflammationdriven prothrombotic state, featuring elevated platelet-monocyte aggregates, increased platelet count, endothelial activation, and heightened plasmatic prothrombotic markers (46).

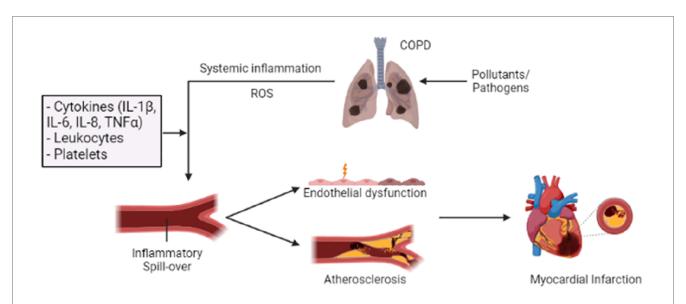
Inflammatory mechanisms in AMI and COPD

Over recent decades, a decline in the incidence and mortality of AMI, particularly ST-elevation myocardial infarction (STEMI), has been observed. Contrastingly, non-ST-elevation myocardial infarction (NSTEMI) cases are increasingly prevalent. Patients with NSTEMI often present with comorbidities or are of advanced age. Although significant focus has been placed on high-risk groups like diabetic patients, other equally at-risk populations, such as COPD patients, have received less attention (99). The prevalence of COPD in AMI populations, possibly underestimated due to general COPD underdiagnosis, ranges from 7% to 30% (43). The risk of AMI and its related mortality is markedly higher in COPD patients, especially soon after an AE-COPD (1). Cardiovascular diseases, predominantly AMI, account for around 50% of deaths in COPD cases. Notably, AMI is closely associated with COPD due to factors like systemic inflammation, thrombosis, and oxidative stress linked to COPD (43). The increased risk is also linked with the

unique physiological crosstalk between the lungs and heart, which is unlike that in other organs. Indeed, research indicates that lung inflammation, as seen in conditions like SARS-CoV-2 infection, significantly impacts the heart due to the interconnectedness of the cardiopulmonary system (100). A substantial portion of the COPD disease burden relates to the management of cardiovascular comorbidities and infectious AE-COPD. AMI and COPD share common risk factors, including smoking, aging, and male sex, however the underlying mechanisms connecting them remain elusive. Systemic inflammation and hypoxia, stemming from COPD, are identified as independent risk factors for AMI (101). Despite extensive epidemiological research on both COPD and AMI, the pathophysiological link between them is less explored, due to the complexity of the multimorbidity involved.

The inflammatory response in COPD can lead to endothelial damage and vascular dysfunction, though the underlying mechanisms warrant further investigation. Inflammatory mediators (such as IL-1, IL-6, IL-8, TNF-α, IL-36, and MMPs) released during COPD, can directly harm endothelial cells lining blood vessels, promoting atherosclerosis, the underlying cause of most AMI (102-104). Chronic lung inflammation promotes systemic inflammation and oxidative stress, which are key in atherosclerotic plaque formation (Figure 2). Prolonged inflammation and oxidative stress are known to elevate the risk of plaque development and rupture, leading to thrombosis and ischemic injury. COPD and cigarette smoke (CS) exposure further exacerbate this risk by increasing thrombosis through the secretion of procoagulant factors. COPD patients typically exhibit higher platelet counts and reactivity, along with increased coagulation factors, resulting in elevated thrombin levels and a greater risk of thrombotic events (103). A recent study focusing on the neutrophil-to-lymphocyte ratio (NLR), an indicator of systemic inflammation, found that a high NLR in COPD patients is associated with an increased risk of AMI, stroke, and mortality, suggesting systemic inflammation's role in cardiovascular disease development within this group (105). Moreover, elevated troponin levels, particularly during AE-COPD and linked to increased neutrophil recruitment, have been noted in COPD patients (106). Patel et al. demonstrated that frequent AE-COPD result in increased arterial stiffness and elevated troponin and BNP levels, correlating these disturbances with the inflammatory response marked by raised IL-6 and IL-8 during AE-COPD (107).

The survival rate of patients with COPD who are subject to an AMI is significantly worse than non-COPD patients. This increased vulnerability to myocardial injury in COPD patients is multifactorial, attributed to airway remodeling driven by inflammation-induced vascular changes and deteriorating lung function. Notably, pulmonary artery intima thickening has been observed not only in mild to moderate COPD patients but also in smokers, indicating that such vascular alterations can manifest early in disease progression (108). Furthermore, pulmonary hypertension (PH), a common complication of severe COPD, significantly contributes to AMI and heart failure. COPD leads to PH through sustained hypoxia, hypoxic vasoconstriction, inflammation in the lungs and systemic inflammation, as well as endothelial dysfunction. These factors result in pulmonary arteriole remodeling and narrowing, exacerbating hypoxia, and elevating pulmonary blood pressure (109). Hypoxia heightens ROS production and oxidative stress, impacting vascular function, cardiac output, and blood pressure. Recent studies have highlighted the role of hypoxia-inducible factor (HIF)- α in promoting vascular remodeling and atherosclerosis via inflammatory pathways



Poerview of how inflammation in COPD can "spill over" into the systemic circulation, increasing the risk of AMI. Long-term exposure to pollutants and frequent acute exacerbations of chronic obstructive pulmonary disease (AE-COPD) can enhance and alter the inflammatory response in the lungs, resulting in a progressive and persistent inflammatory response. This COPD-related inflammatory response can promote systemic inflammation and oxidative stress through several inflammatory mediators (IL-1 β , IL-6, IL-8, and TNF- α). Their actions can induce endothelial injury and vascular dysfunction through increased endothelial permeability and enhanced oxidative stress. Additionally, prolonged inflammation and oxidative stress can contribute to the development of atherosclerotic plaque and thrombosis, thereby increasing the risk of acute myocardial infarction (AMI). Patients with COPD exhibit a heightened platelet count and reactivity, ultimately increasing the risk of thrombotic events.

(110). Current research underscores the inflammatory response as a pivotal element in the COPD-AMI connection, suggesting that targeting inflammation could be a promising approach in treating cardiovascular disease among COPD patients.

Therapeutic strategies for targeting inflammation in AMI and COPD

A potential therapeutic target for limiting COPD progression, infarct size and adverse remodeling while enhancing prognosis could be to target the pro-inflammatory responses observed during COPD and myocardial IR injury (8, 111, 112). The current standard anti-inflammatory therapy for COPD is corticosteroids, often combined with a long-acting bronchodilator to enhance lung function and reduce exacerbations. The impact of corticosteroids on AMI incidence and outcomes in both COPD and non-COPD patients is subject to conflicting findings. Inhaled corticosteroids may potentially reduce AMI risk in COPD patients, although some studies show no significant effect (113, 114). In contrast, oral corticosteroid use increases AMI risk in COPD patients (115). In non-COPD patients, corticosteroid use for AMI treatment raises concerns about adverse effects like impaired wound healing and remodelling (116). The role of corticosteroids in AMI treatment remains a topic of research and clinical debate, necessitating further investigation into their potential benefits and risks. Aspirin, a standard therapy for AMI patients to prevent blood clots and reduce the risk of new MI or stroke, may also provide pulmonary benefits and decelerate COPD progression (117). A recent trail suggests its potential in reducing cardiovascular complications in COPD patients, warranting further investigation (118).

Several other studies and clinical trials have focused on individual components of the inflammatory adhesion cascade, showing some benefits for COPD and AMI independently (Table 1). However, elevated levels of inflammatory mediators in these diseases do not necessarily suggest that their inhibition will be an effective antiinflammatory strategy. For instance, while TNFα levels are high in both AMI and COPD, inhibiting it has not demonstrated clinical benefits in either of these conditions (119). The pleiotropic effects of inflammatory cytokines on the immune system make them attractive targets for reducing inflammation. Several clinical trials have been undertaken to assess therapies targeting key inflammatory cytokines, especially IL-1, IL-6, and TNF- α (Table 1). However, there has been a lack of studies examining the impact of anti-inflammatory treatments in animal models or patients with both COPD and AMI. Therefore, this review will focus only on targets that have demonstrated benefits in both COPD and AMI independently, recognizing that potential dual therapy targets warrant further research.

Therapeutic targets in the interleukin-1 super family

The IL-1 superfamily (IL-1F), encompassing eleven cytokines, plays a pivotal role in modulating the innate immune response.

This family includes both pro-inflammatory and antiinflammatory cytokines that influence integrin expression on target cells and stimulate cytokine release from stromal cells. The pro-inflammatory members are IL-1α, IL-1β, IL-18, IL-36α, IL-36β, IL-36γ, and IL-33. The anti-inflammatory cytokines are divided into immunosuppressive (IL-37 and IL-38) and antagonist cytokines (IL-1Ra, which antagonizes IL-1α and IL-1β, and IL-36Ra, targeting IL-36α, IL-36β, and IL-36γ). Except for IL-1Ra, all members of the IL-1 family are initially produced as precursor proteins, requiring proteolytic cleavage to become active, mature proteins (120). IL-1 cytokines (IL-1 α and IL-1β) have garnered significant attention; these agonists, located in the cytoplasm, are expressed in various cells, including neutrophils, monocytes, and macrophages. However, some members, like the IL-36 cytokines, have been recently discovered, and research on their functions remains limited (121). IL-1F cytokines are often the earliest and most upstream responders to injury, making them promising targets for managing inflammatory disorders (122). Given their critical role in mediating inflammation, IL-1F members could be key to understanding and addressing myocardial and pulmonary disturbances.

Targeting IL-1 in AMI and COPD

Experimental models that target IL-1 in myocardial IR injury have shown promise. Toldo et al. (123) reported that anakinra, a recombinant human IL-1 receptor antagonist (IL-1Ra), significantly reduced infarct size and improved LV ejection fraction in mice following myocardial IR injury (123). Likewise, a study in rat hearts indicated that IL-1Ra overexpression protected against IR injury by attenuating cell death and reducing infarct size through its anti-inflammatory properties on neutrophils (124). Clinical studies using anakinra or canakinumab, an IL-1β monoclonal antibody, in STEMI or NSTEMI patients have been mixed. Morton et al. (125) conducted a double-blinded randomized phase II trial with anakinra in NSTEMI patients, finding decreased CRP levels but increased adverse outcomes (recurrent MI, stroke, and death) over one year (8, 125). Conversely, the CANTOS trial by Ridker et al. (126) demonstrated that canakinumab significantly reduced nonfatal MI, stroke, or cardiovascular death in highrisk patients over 3.7 years (126). This was the first major clinical trial to show that targeting inflammation can confer modest cardiovascular benefits in very high-risk patients and has been credited with providing an exciting glimpse at the potential for using anti-inflammatory therapies for treating cardiovascular diseases.

In CS exposure models, IL-1 inhibition also showed effectiveness. Castro et al. (127) found that an anti-IL-1 β antibody significantly reduced CS-induced lung inflammation (127). Churg et al. (6) reported protection against emphysema and airway remodeling in IL-1R knockout mice exposed to CS (6). A phase I clinical study by Hernandez et al. (128) using anakinra in healthy volunteers exposed to inhaled

lipopolysaccharide (LPS) showed reduced airway neutrophilia and cytokine levels (IL-1β, IL-6, and IL-8) (128). In a phase II trial, Calverley et al. (67) tested MEDI8968, an IL-1R1 monoclonal antibody, in moderate-to-severe COPD patients, finding initial inflammation reduction but no significant changes in AE-COPD rate, lung function, or quality of life compared to placebo (67). Overall, this trial suggested potential benefits for COPD patients with high neutrophil counts, though current serum biomarkers might not adequately identify these patients.

Targeting IL-36 in AMI and COPD

IL-36, a relatively recent member of the IL-1 family discovered about two decades ago, plays a key role in pro-inflammatory mediator production, immune cell activation, and antigen presentation (129). As well as amplifying IL-1 effects, IL-36 is also a mediator of inflammation in its own right. Indeed, its critical role in psoriasis, equalling if not surpassing that of IL-1, is well established with emerging roles in Crohn disease and rheumatoid arthritis recently identified (130–132). Research into IL-36 as a pro-inflammatory cytokine in various inflammatory diseases is extensive, but studies in AMI or COPD are more recent.

In experimental models of myocardial IR injury, inhibiting IL-36R has shown success. In 2019, Wei et al. found that administering recombinant mouse IL-38 in mice post-IR injury reduced inflammation, cardiac fibrosis, and myocardial injury, suggesting IL-38 could benefit cardiac remodeling after MI (133). More recently, we demonstrated that treatment with IL-36Ra in mice with myocardial IR injury significantly reduced infarct size and neutrophil recruitment, and improved blood flow. The mechanism involved attenuating endothelial and cardiac myocyte oxidative damage and VCAM-1 expression (16, 134). In the context of COPD, while the IL-36/IL36R pathway's pro-inflammatory role in the lungs is well recognized, its protective potential is still emerging. Baker et al. (15) reported reduced virally induced IL-36-mediated inflammation in COPD using IL-36Ra (Spesolimab) or an IL-36R blocking antibody. This was mechanistically explained by blocking the crosstalk between virally stimulated COPD small airway epithelial cells and small airway fibroblasts (15). Additionally, their 2021 study showed that IL-36R knockout mice had diminished lung inflammation and neutrophil recruitment in a CS exposure model (17). These findings suggest a novel role for IL-36 in both myocardial IR injury and COPD and necessitate further exploration in a comorbidity model.

Therapeutic targets beyond IL-1F members

Targeting interleukin-6

IL-6, released from immune cells after myocardial IR injury and during COPD, exhibits both anti- and pro-inflammatory effects, notably as a potent inducer of CRP secretion. Vanfleteren

et al. (135) identified elevated IL-6 plasma levels exclusively in COPD patients with cardiovascular comorbidities (135). AMI studies in mice with IL-6 genetic depletion or pharmacological inhibition show mixed outcomes on myocardial infarction size and LV remodeling, with some studies indicating worsened outcomes post-inhibition (136–138). Broch et al. (139) reported a phase II trial testing tocilizumab, an IL-6R monoclonal antibody, in STEMI patients, finding increased myocardial salvage and decreased microvascular obstruction, although infarct size was unchanged after 7 days (139). Inhibition of IL-6 also demonstrated to be beneficial in COPD. Indeed, Wei et al. (10) isolated human and mouse primary bronchial epithelial cells from COPD patients and mice. They showed IL-6 neutralizing antibody (IL-6 Ab) to attenuate airway mucus hypersecretion in COPD (10).

Targeting S100A8/A9

S100A8/A9, or calprotectin, part of the S100 DAMP family, is a prominent neutrophil and monocyte protein secreted during inflammation and infection. It plays a role in proinflammatory mediator production but can also exhibit antiinflammatory effects under specific conditions. In myocardial IR injury models, inhibition of S100A8/A9 has been effective. Marinković et al. (29) observed reduced inflammatory cell recruitment and improved cardiac function following short term treatment with ABR-238901, an S100A8/A9 inhibitor (29). Similar improvements in cardiac function were seen in a study of mice transplanted with bone marrow from S100A9 knockout mice (18). However, prolonged ABR-238901 treatment worsened cardiac function and accelerated LV remodeling (140). These findings underline the need for timely interventions. Clinically, S100A8/A9 levels have been linked with recurrence risk and poorer prognosis (141). Despite recent increases in our understanding of how S100A8/A9 is involved in several lung diseases, we are still at an early stage in our current understanding of its potential protective role in COPD. Zhao et al. (20) found that ABR-238901 prevented LPS-induced lung injury in mice, primarily through NLRP3 pathway suppression (20). Conversely, recombinant S100A8/A9 reduced acute lung injury in LPS-challenged mice in another study (142). These varied outcomes, potentially influenced by differences in mouse species, treatment timing, and experimental setups, may underscore the dual pro- and antiinflammatory nature of S100A8/A9. Additionally, S100A8 was found to protect human alveolar cells against emphysema and injury (143).

Optimizing anti-inflammatory therapies for AMI and COPD

While various components of inflammatory process have shown promise in reducing infarct size and slowing COPD progression, their efficacy in a comorbidity context involving AMI in COPD patients remains untested. Consequently, the applicability of these strategies to such patients is uncertain.

Additionally, several potential targets not covered in this review failed in clinical translation, possibly due to discrepancies in experimental design, therapeutic intervention timing, dosing, and the distinct pathophysiological mechanisms in animal models vs. humans. Experimental models often do not fully mirror human pathogenesis, especially considering that most COPD patients have comorbidities like atherosclerosis and AMI, are typically older, and frequently use multiple medications. Many animal studies also neglect sex differences in outcomes (8).

Non-resolving inflammation, which fails to naturally subside, plays a pivotal role in the pathophysiology of both COPD and AMI. In AMI, it contributes to adverse remodeling and advanced heart failure, leading to a poor prognosis, while in COPD, it triggers systemic effects, contributing to comorbidities like cardiovascular diseases, including AMI (21, 144). The timing of therapeutic intervention significantly influences inflammation resolution. Clinical trials administering anti-inflammatories days or months after AMI aim to target resolution but often miss the initial inflammatory peak, resulting in limited cardiovascular benefits (126). Recent research explored the benefits of very early anti-inflammatory therapy in AMI patients, showcasing its potential for salvaging myocardial tissue (139). Nevertheless, tackling persistent inflammation remains challenging, potentially necessitating sustained or additional therapeutic approaches. Similarly, in COPD patients, commonly used anti-inflammatories like corticosteroids have exhibited limitations in modulating nonresolving inflammation, even after smoking cessation (145). Unfortunately, no highly effective anti-inflammatory treatment successfully targets persistent inflammation. Therefore, understanding the molecular mechanisms governing resolution onset is imperative, offering the potential for developing resolution-based strategies, expanding research horizons, and potential therapeutic breakthroughs.

Enhancing clinical success may hinge on patient stratification and selection, particularly in COPD, a heterogeneous disease with various distinct phenotypes beyond genetic factors. This diversity also applies to AMI. COPD progression varies widely among patients, and factors like the degree of airway obstruction, pulmonary emphysema, chronic bronchitis, and the frequency of acute exacerbations do not fully capture this heterogeneity. Understanding clinical phenotypes and linking them to underlying pathophysiological mechanisms via inflammatory biomarkers is crucial. This approach allows for targeted therapy against specific anti-inflammatory markers predominantly expressed in individual patients. Moreover, developing new antiinflammatory targets necessitates testing in animal models that more accurately represent clinical scenarios and through meticulous patient selection. Pursuing an inflammatory target that optimizes perfusion in pulmonary and coronary microcirculation, minimizes damage, and enhances patient prognosis remains a valuable endeavour.

Concluding remarks

Over recent decades, substantial progress has been achieved in understanding the pathophysiological intricacies of COPD and AMI, with a particular emphasis on the pivotal role of inflammation as a connecting thread between these conditions. Despite these advances, the understanding of inflammatory interactions in a comorbidity context remains elusive. Furthermore, while specific inflammatory targets have been identified for therapeutic intervention, their efficacy in treating concurrent COPD and AMI remains unclear. This gap underscores an essential direction for future research, particularly given the variability in clinical manifestations and patient responses. Looking ahead, the adoption of personalized medicine, finely tuned to the unique inflammatory profiles of individuals, promises to yield more effective and precise treatment modalities. Hence, a deeper and more nuanced comprehension of the shared inflammatory pathways in COPD and AMI is imperative to enhance patient care and clinical outcomes.

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References

- 1. Kunisaki KM, Dransfield MT, Anderson JA, Brook RD, Calverley PMA, Celli BR, et al. Exacerbations of chronic obstructive pulmonary disease and cardiac events. A post hoc cohort analysis from the SUMMIT randomized clinical trial. *Am J Respir Crit Care Med.* (2018) 198(1):51–7. doi: 10.1164/rccm.201711-2239OC
- 2. Rothnie KJ, Connell O, Müllerová H, Smeeth L, Pearce N, Douglas I, et al. Myocardial infarction and ischemic stroke after exacerbations of chronic obstructive pulmonary disease. *Ann Am Thorac Soc.* (2018) 15(8):935–46. doi: 10.1513/AnnalsATS.201710-815OC
- Stieger P, Daniel J-M, Thölen C, Dutzmann J, Knöpp K, Gündüz D, et al. Targeting of extracellular RNA reduces edema formation and infarct size and improves survival after myocardial infarction in mice. J Am Heart Assoc. (2017) 6 (6):e004541. doi: 10.1161/JAHA.116.004541
- 4. Hu G, Zhang Y, Jiang H, Hu X. Exendin-4 attenuates myocardial ischemia and reperfusion injury by inhibiting high mobility group box 1 protein expression. Cardiol J. (2013) 20(6):600–4. doi: 10.5603/CJ.2013.0159
- 5. Frangogiannis NG. The immune system and cardiac repair. Pharmacol Res. $(2008)\ 58(2):88-111$. doi: 10.1016/j.phrs.2008.06.007
- 6. Núñez J, Núñez E, Bodí V, Sanchis J, Miñana G, Mainar L, et al. Usefulness of the neutrophil to lymphocyte ratio in predicting long-term mortality in ST segment elevation myocardial infarction. *Am J Cardiol.* (2008) 101(6):747–52. doi: 10.1016/j. amjcard.2007.11.004
- 7. Fang L, Moore X-L, Dart AM, Wang L-M. Systemic inflammatory response following acute myocardial infarction. *J Geriatr Cardiol.* (2015) 12(3):305–12. doi: 10.11909/j.issn.1671-5411.2015.03.020
- 8. Guasti L, Dentali F, Castiglioni L, Maroni L, Marino F, Squizzato A, et al. Neutrophils and clinical outcomes in patients with acute coronary syndromes and/or cardiac revascularisation. A systematic review on more than 34,000 subjects. *Thromb Haemost.* (2011) 106(4):591–9. doi: 10.1160/TH11-02-0096
- 9. Prabhu SD, Frangogiannis NG. The biological basis for cardiac repair after myocardial infarction: from inflammation to fibrosis. *Circ Res.* (2016) 119 (1):91–112. doi: 10.1161/CIRCRESAHA.116.303577
- 10. Ong S-B, Hernández-Reséndiz S, Crespo-Avilan GE, Mukhametshina RT, Kwek X-Y, Cabrera-Fuentes HA, et al. Inflammation following acute myocardial infarction: multiple players, dynamic roles, and novel therapeutic opportunities. *Pharmacol Ther*. (2018) 186:73–87. doi: 10.1016/j.pharmthera.2018.01.001
- 11. McDonald B, Pittman K, Menezes GB, Hirota SA, Slaba I, Waterhouse CC, et al. Intravascular danger signals guide neutrophils to sites of sterile inflammation. *Science*. (2010) 330(6002):362–6. doi: 10.1126/science.1195491
- 12. Ma Y, Yabluchanskiy A, Iyer RP, Cannon PL, Flynn ER, Jung M, et al. Temporal neutrophil polarization following myocardial infarction. *Cardiovasc Res.* (2016) 110 (1):51–61. doi: 10.1093/cvr/cvw024
- 13. Arai M, Lefer DJ, So T, DiPaula A, Aversano T, Becker LC. An anti-CD18 antibody limits infarct size and preserves left ventricular function in dogs with ischemia and 48-hour reperfusion. *J Am Coll Cardiol.* (1996) 27(5):1278–85. doi: 10.1016/0735-1097(95)00578-1
- 14. Sreejit G, Abdel-Latif A, Athmanathan B, Annabathula R, Dhyani A, Noothi SK, et al. Neutrophil-Derived \$100A8/A9 amplify granulopoiesis after myocardial infarction. *Circulation*. (2020) 141(13):1080–94. doi: 10.1161/CIRCULATIONAHA. 119.043833
- 15. Marinković G, Grauen Larsen H, Yndigegn T, Szabo IA, Mares RG, de Camp L, et al. Inhibition of pro-inflammatory myeloid cell responses by short-term S100A9 blockade improves cardiac function after myocardial infarction. *Eur Heart J.* (2019) 40(32):2713–23. doi: 10.1093/eurheartj/ehz461
- 16. Mariani M, Fetiveau R, Rossetti E, Poli A, Poletti F, Vandoni P, et al. Significance of total and differential leucocyte count in patients with acute myocardial infarction treated with primary coronary angioplasty. *Eur Heart J.* (2006) 27(21):2511–5. doi: 10.1093/eurheartj/ehl191
- 17. Frangogiannis NG. The inflammatory response in myocardial injury, repair, and remodelling. *Nat Rev Cardiol.* (2014) 11(5):255-65. doi: 10.1038/nrcardio.2014.28
- 18. Harel-Adar T, Ben Mordechai T, Amsalem Y, Feinberg MS, Leor J, Cohen S. Modulation of cardiac macrophages by phosphatidylserine-presenting liposomes improves infarct repair. *Proc Natl Acad Sci U S A.* (2011) 108(5):1827–32. doi: 10. 1073/pnas.1015623108
- 19. Yilmaz A, Weber J, Cicha I, Stumpf C, Klein M, Raithel D, et al. Decrease in circulating myeloid dendritic cell precursors in coronary artery disease. *J Am Coll Cardiol.* (2006) 48(1):70–80. doi: 10.1016/j.jacc.2006.01.078
- 20. Yan X, Anzai A, Katsumata Y, Matsuhashi T, Ito K, Endo J, et al. Temporal dynamics of cardiac immune cell accumulation following acute myocardial infarction. *J Mol Cell Cardiol.* (2013) 62:24–35. doi: 10.1016/j.yjmcc.2013.04.023
- 21. Blum A, Yeganeh S. The role of T-lymphocyte subpopulations in acute myocardial infarction. *Eur J Intern Med.* (2003) 14(7):407–10. doi: 10.1016/j.ejim. 2003.09.002

- 22. Al-Ahmad RS, Mahafzah AM, Al-Mousa EN. Immunological changes in acute myocardial infarction. Saudi Med I. (2004) 25(7):923–8.
- 23. Zouggari Y, Ait-Oufella H, Bonnin P, Simon T, Sage AP, Guérin C, et al. B lymphocytes trigger monocyte mobilization and impair heart function after acute myocardial infarction. *Nat Med.* (2013) 19(10):1273–80. doi: 10.1038/nm.3284
- 24. Backteman K, Ernerudh J, Jonasson L. Natural killer (NK) cell deficit in coronary artery disease: no aberrations in phenotype but sustained reduction of NK cells is associated with low-grade inflammation. *Clin Exp Immunol.* (2014) 175 (1):104–12. doi: 10.1111/cei.12210
- 25. Chen Y, Zhong H, Zhao Y, Luo X, Gao W. Role of platelet biomarkers in inflammatory response. *Biomark Res.* (2020) 8(1):28. doi: 10.1186/s40364-020-00207-2
- 26. Gros A, Ollivier V, Ho-Tin-Noé B. Platelets in inflammation: regulation of leukocyte activities and vascular repair. *Front Immunol.* (2015) 5:678. doi: 10.3389/fimmu 2014 00678
- 27. Stark K, Massberg S. Interplay between inflammation and thrombosis in cardiovascular pathology. Nat Rev Cardiol. (2021) 18(9):666-82. doi: 10.1038/s41569-021-00552-1
- 28. Ge L, Zhou X, Ji WJ, Lu RY, Zhang Y, Zhang YD, et al. Neutrophil extracellular traps in ischemia-reperfusion injury-induced myocardial no-reflow: therapeutic potential of DNase-based reperfusion strategy. *Am J Physiol Heart Circ Physiol.* (2015) 308(5):H500–9. doi: 10.1152/ajpheart.00381.2014
- 29. Furman MI, Barnard MR, Krueger LA, Fox ML, Shilale EA, Lessard DM, et al. Circulating monocyte-platelet aggregates are an early marker of acute myocardial infarction. *J Am Coll Cardiol.* (2001) 38(4):1002–6. doi: 10.1016/S0735-1097(01) 01485-1
- 30. Goedemans L, Bax JJ, Delgado V. COPD and acute myocardial infarction. Eur Respir Rev. (2020) 29(156):190139. doi: 10.1183/16000617.0139-2019
- 31. Brode SK, Ling SC, Chapman KR. Alpha-1 antitrypsin deficiency: a commonly overlooked cause of lung disease. *CMAJ*. (2012) 184(12):1365–71. doi: 10.1503/cmaj. 111749
- 32. Fischer BM, Voynow JA, Ghio AJ. COPD: balancing oxidants and antioxidants. Int J Chron Obstruct Pulmon Dis. (2015) 10:261–76. doi: 10.2147/COPD.S42414
- 33. van der Vorm LN, Li L, Huskens D, Hulstein JJJ, Roest M, de Groot PG, et al. Acute exacerbations of COPD are associated with a prothrombotic state through platelet-monocyte complexes, endothelial activation and increased thrombin generation. *Respir Med.* (2020) 171:106094. doi: 10.1016/j.rmed.2020.106094
- 34. Hogg JC, Chu F, Utokaparch S, Woods R, Elliott WM, Buzatu L, et al. The nature of small-airway obstruction in chronic obstructive pulmonary disease. N Engl J Med. (2004) 350(26):2645–53. doi: 10.1056/NEJMoa032158
- 35. Choudhury G, Rabinovich R, MacNee W. Comorbidities and systemic effects of chronic obstructive pulmonary disease. *Clin Chest Med.* (2014) 35(1):101–30. doi: 10. 1016/j.ccm.2013.10.007
- 36. Mendy A, Forno E, Niyonsenga T, Gasana J. Blood biomarkers as predictors of long-term mortality in COPD. *Clin Respir J.* (2018) 12(5):1891–9. doi: 10.1111/crj. 12752
- 37. Singh D, Edwards L, Tal-Singer R, Rennard S. Sputum neutrophils as a biomarker in COPD: findings from the ECLIPSE study. *Respir Res.* (2010) 11(1):77. doi: 10.1186/1465-9921-11-77
- 38. Sapey E, Patel JM, Greenwood HL, Walton GM, Hazeldine J, Sadhra C, et al. Pulmonary infections in the elderly lead to impaired neutrophil targeting, which is improved by simvastatin. *Am J Respir Crit Care Med.* (2017) 196(10):1325–36. doi: 10.1164/rccm.201704-0814OC
- 39. Wang Y, Jia M, Yan X, Cao L, Barnes PJ, Adcock IM, et al. Increased neutrophil gelatinase-associated lipocalin (NGAL) promotes airway remodelling in chronic obstructive pulmonary disease. *Clin Sci (Lond)*. (2017) 131(11):1147–59. doi: 10.1042/CS20170096
- 40. Donnelly LE, Barnes PJ. Chemokine receptors as therapeutic targets in chronic obstructive pulmonary disease. *Trends Pharmacol Sci.* (2006) 27(10):546–53. doi: 10. 1016/j.tips.2006.08.001
- 41. Gharib SA, Manicone AM, Parks WC. Matrix metalloproteinases in emphysema. *Matrix Biol.* (2018) 73:34–51. doi: 10.1016/j.matbio.2018.01.018
- 42. Hautamaki RD, Kobayashi DK, Senior RM, Shapiro SD. Requirement for macrophage elastase for cigarette smoke-induced emphysema in mice. *Science*. (1997) 277(5334):2002–4. doi: 10.1126/science.277.5334.2002
- 43. Christopoulou M-E, Papakonstantinou E, Stolz D. Matrix metalloproteinases in chronic obstructive pulmonary disease. *Int J Mol Sci.* (2023) 24(4):3786. doi: 10.3390/ijms24043786
- 44. Linder R, Rönmark E, Pourazar J, Behndig AF, Blomberg A, Lindberg A. Proteolytic biomarkers are related to prognosis in COPD- report from a population-based cohort. *Respir Res.* (2018) 19(1):64. doi: 10.1186/s12931-018-0772-5

- 45. Deb N, Mallick S, Jaiswal A, Manna A, Mabalirajan U, Swarnakar S. Role of MMPs and oxidants in lung diseases. In: Chakraborti S, Chakraborti T, Das SK, Chattopadhyay D, editors. Oxidative Stress in Lung Diseases: Volume 1. Singapore: Springer Singapore (2019). p. 149–69.
- 46. Ding L, Yang J, Zhang C, Zhang X, Gao P. Neutrophils modulate fibrogenesis in chronic pulmonary diseases. *Front Med (Lausanne)*. (2021) 8:616200. doi: 10.3389/fmed.2021.616200
- 47. Arai N, Kondo M, Izumo T, Tamaoki J, Nagai A. Inhibition of neutrophil elastase-induced goblet cell metaplasia by tiotropium in mice. *Eur Respir J.* (2010) 35(5):1164–71. doi: 10.1183/09031936.00040709
- 48. Guyot N, Wartelle J, Malleret L, Todorov AA, Devouassoux G, Pacheco Y, et al. Unopposed cathepsin G, neutrophil elastase, and proteinase 3 cause severe lung damage and emphysema. *Am J Pathol.* (2014) 184(8):2197–210. doi: 10.1016/j. ajpath.2014.04.015
- 49. Qiu SL, Zhang H, Tang QY, Bai J, He ZY, Zhang JQ, et al. Neutrophil extracellular traps induced by cigarette smoke activate plasmacytoid dendritic cells. *Thorax.* (2017) 72(12):1084–93. doi: 10.1136/thoraxjnl-2016-209887
- 50. Arora S, Dev K, Agarwal B, Das P, Syed MA. Macrophages: their role, activation and polarization in pulmonary diseases. *Immunobiology.* (2018) 223(4–5):383–96. doi: 10.1016/j.imbio.2017.11.001
- 51. Russell RE, Thorley A, Culpitt SV, Dodd S, Donnelly LE, Demattos C, et al. Alveolar macrophage-mediated elastolysis: roles of matrix metalloproteinases, cysteine, and serine proteases. *Am J Physiol Lung Cell Mol Physiol.* (2002) 283(4): L867–73. doi: 10.1152/ajplung.00020.2002
- 52. Bazzan E, Turato G, Tinè M, Radu CM, Balestro E, Rigobello C, et al. Dual polarization of human alveolar macrophages progressively increases with smoking and COPD severity. *Respir Res.* (2017) 18(1):40. doi: 10.1186/s12931-017-0522-0
- 53. Barnes PJ. Cellular and molecular mechanisms of chronic obstructive pulmonary disease. *Clin Chest Med.* (2014) 35(1):71–86. doi: 10.1016/j.ccm.2013.10.004
- 54. Lappalainen U, Whitsett JA, Wert SE, Tichelaar JW, Bry K. Interleukin-1beta causes pulmonary inflammation, emphysema, and airway remodeling in the adult murine lung. *Am J Respir Cell Mol Biol.* (2005) 32(4):311–8. doi: 10.1165/rcmb.2004-0309OC
- 55. Reynaert N, Eurlings I, Mercken E, Cabo RD, Aesif S, Van der Velden J, et al. Involvement of JNK in TNF α driven remodelling. *Eur Respir J.* (2015) 46(Suppl 59):PA5058. doi: 10.1183/13993003.congress-2015.PA5058
- 56. Churg A, Zhou S, Wang X, Wang R, Wright JL. The role of interleukin-1beta in murine cigarette smoke-induced emphysema and small airway remodeling. Am J Respir Cell Mol Biol. (2009) 40(4):482–90. doi: 10.1165/rcmb.2008-0038OC
- 57. Calverley PMA, Sethi S, Dawson M, Ward CK, Finch DK, Penney M, et al. A randomised, placebo-controlled trial of anti-interleukin-1 receptor 1 monoclonal antibody MEDI8968 in chronic obstructive pulmonary disease. *Respir Res.* (2017) 18(1):153. doi: 10.1186/s12931-017-0633-7
- 58. McCombs JE, Kolls JK. Walking down the "IL": the newfound marriage between IL-36 and chronic obstructive pulmonary disease. *Am J Respir Cell Mol Biol.* (2021) 64 (2):153–4. doi: 10.1165/rcmb.2020-0461ED
- 59. Kovach MA, Che K, Brundin B, Andersson A, Asgeirsdottir H, Padra M, et al. IL-36 Cytokines promote inflammation in the lungs of long-term smokers. *Am J Respir Cell Mol Biol.* (2021) 64(2):173–82. doi: 10.1165/rcmb.2020-0035OC
- 60. Baker JR, Fenwick PS, Koss CK, Owles HB, Elkin SL, Fine J, et al. IL-36 receptor agonist and antagonist imbalance drives neutrophilic inflammation in COPD. *JCI Insight.* (2022) 7(15):e155581. doi: 10.1172/jci.insight.155581
- $61.\,Li$ J, Ye Z. The potential role and regulatory mechanisms of MUC5AC in chronic obstructive pulmonary disease. Molecules. (2020) 25(19):4437. doi: 10.3390/molecules25194437
- 62. Lerner CA, Lei W, Sundar IK, Rahman I. Genetic ablation of CXCR2 protects against cigarette smoke-induced lung inflammation and injury. *Front Pharmacol.* (2016) 7:391. doi: 10.3389/fphar.2016.00391
- 63. Imaoka H, Hoshino T, Takei S, Kinoshita T, Okamoto M, Kawayama T, et al. Interleukin-18 production and pulmonary function in COPD. *Eur Respir J.* (2008) 31(2):287–97. doi: 10.1183/09031936.00019207
- 64. Briend E, Ferguson GJ, Mori M, Damera G, Stephenson K, Karp NA, et al. IL-18 associated with lung lymphoid aggregates drives IFN γ production in severe COPD. Respir Res. (2017) 18(1):159. doi: 10.1186/s12931-017-0641-7
- 65. Rong B, Fu T, Rong C, Liu W, Li K, Liu H. Correlation between serum IL-32 concentration and clinical parameters of stable COPD: a retrospective clinical analysis. *Sci Rep.* (2020) 10(1):12092. doi: 10.1038/s41598-020-69000-3
- $66.\ Brusselle$ GG, Joos GF, Bracke KR. New insights into the immunology of chronic obstructive pulmonary disease. *Lancet.* (2011) 378(9795):1015–26. doi: 10.1016/S0140-6736(11)60988-4
- 67. Tam A, Tanabe N, Churg A, Wright JL, Hogg JC, Sin DD. Sex differences in lymphoid follicles in COPD airways. *Respir Res.* (2020) 21(1):46. doi: 10.1186/s12931-020-1311-8
- 68. Donovan C, Starkey MR, Kim RY, Rana BMJ, Barlow JL, Jones B, et al. Roles for T/B lymphocytes and ILC2s in experimental chronic obstructive pulmonary disease. *J Leukoc Biol.* (2019) 105(1):143–50. doi: 10.1002/JLB.3AB0518-178R

69. Xue W, Ma J, Li Y, Xie C. Role of CD(4) (+) T and CD(8) (+) T lymphocytes-mediated cellular immunity in pathogenesis of chronic obstructive pulmonary disease. *J Immunol Res.* (2022) 2022:1429213. doi: 10.1155/2022/1429213

- 70. Roos AB, Sandén C, Mori M, Bjermer L, Stampfli MR, Erjefält JS. IL-17A is elevated in End-stage chronic obstructive pulmonary disease and contributes to cigarette smoke-induced lymphoid neogenesis. *Am J Respir Crit Care Med.* (2015) 191(11):1232–41. doi: 10.1164/rccm.201410-1861OC
- 71. Yanagisawa H, Hashimoto M, Minagawa S, Takasaka N, Ma R, Moermans C, et al. Role of IL-17A in murine models of COPD airway disease. *Am J Physiol Lung Cell Mol Physiol.* (2017) 312(1):L122–l30. doi: 10.1152/ajplung.00301.2016
- 72. Lonky SA, Wohl H. Stimulation of human leukocyte elastase by platelet factor 4. Physiologic, morphologic, and biochemical effects on hamster lungs in vitro. *J Clin Invest.* (1981) 67(3):817–26. doi: 10.1172/JCI110099
- 73. Mallah H, Ball S, Sekhon J, Parmar K, Nugent K. Platelets in chronic obstructive pulmonary disease: an update on pathophysiology and implications for antiplatelet therapy. *Respir Med.* (2020) 171:106098. doi: 10.1016/j.rmed.2020.106098
- 74. Kyriakopoulos C, Chronis C, Papapetrou E, Tatsioni A, Gartzonika K, Tsaousi C, et al. Prothrombotic state in patients with stable COPD: an observational study. *ERJ Open Res.* (2021) 7(4):00297-2021. doi: 10.1183/23120541.00297-2021
- 75. Trivedi A, Khan MA, Bade G, Talwar A. Orchestration of neutrophil extracellular traps (nets), a unique innate immune function during chronic obstructive pulmonary disease (COPD) development. *Biomedicines*. (2021) 9(1):53. doi: 10.3390/biomedicines9010053
- 76. Andersson CK, Mori M, Bjermer L, Löfdahl CG, Erjefält JS. Alterations in lung mast cell populations in patients with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med.* (2010) 181(3):206–17. doi: 10.1164/rccm.200906-0932OC
- 77. Sommerhoff CP, Caughey GH, Finkbeiner WE, Lazarus SC, Basbaum CB, Nadel JA. Mast cell chymase. A potent secretagogue for airway gland serous cells. *J Immunol.* (1989) 142(7):2450–6. doi: 10.4049/jimmunol.142.7.2450
- 78. Eltboli O, Brightling CE. Eosinophils as diagnostic tools in chronic lung disease. Expert Rev Respir Med. (2013) 7(1):33–42. doi: 10.1586/ers.12.81
- 79. Tashkin DP, Wechsler ME. Role of eosinophils in airway inflammation of chronic obstructive pulmonary disease. *Int J Chron Obstruct Pulmon Dis.* (2018) 13:335–49. doi: 10.2147/COPD.S152291
- 80. Perng DW, Chen PK. The relationship between airway inflammation and exacerbation in chronic obstructive pulmonary disease. *Tuberc Respir Dis* (Seoul). (2017) 80(4):325–35. doi: 10.4046/trd.2017.0085
- 81. Vedel-Krogh S, Nielsen SF, Lange P, Vestbo J, Nordestgaard BG. Blood eosinophils and exacerbations in chronic obstructive pulmonary disease. The Copenhagen general population study. *Am J Respir Crit Care Med.* (2016) 193 (9):965–74. doi: 10.1164/rccm.201509-1869OC
- 82. Saha S, Brightling CE. Eosinophilic airway inflammation in COPD. Int J Chron Obstruct Pulmon Dis. (2006) 1(1):39–47. doi: 10.2147/copd.2006.1.1.39
- 83. Tsoumakidou M, Tzanakis N, Chrysofakis G, Kyriakou D, Siafakas NM. Changes in sputum T-lymphocyte subpopulations at the onset of severe exacerbations of chronic obstructive pulmonary disease. *Respir Med.* (2005) 99 (5):572–9. doi: 10.1016/j.rmed.2004.10.005
- 84. Gerritsen WB, Asin J, Zanen P, van den Bosch JM, Haas FJ. Markers of inflammation and oxidative stress in exacerbated chronic obstructive pulmonary disease patients. *Respir Med.* (2005) 99(1):84–90. doi: 10.1016/j.rmed.2004.04.017
- 85. Huang SJ, Ding ZN, Xiang HX, Fu L, Fei J. Association between Serum S100A8/S100A9 heterodimer and pulmonary function in patients with acute exacerbation of chronic obstructive pulmonary disease. *Lung.* (2020) 198(4):645–52. doi: 10.1007/s00408-020-00376-9
- 86. Zhang J, Bai C. The significance of Serum interleukin-8 in acute exacerbations of chronic obstructive pulmonary disease. Tanaffos. (2018) 17(1):13–21.
- 87. Wells JM, Parker MM, Oster RA, Bowler RP, Dransfield MT, Bhatt SP, et al. Elevated circulating MMP-9 is linked to increased COPD exacerbation risk in SPIROMICS and COPDGene. *JCI Insight.* (2018) 3(22):e123614. doi: 10.1172/jci. insight.123614
- 88. Malo O, Sauleda J, Busquets X, Miralles C, Agustí AG, Noguera A. Systemic inflammation during exacerbations of chronic obstructive pulmonary disease. *Arch Bronconeumol.* (2002) 38(4):172–6. doi: 10.1016/S0300-2896(02)75184-3
- 89. Pascual-Guardia S, Ataya M, Ramírez-Martínez I, Yélamos J, Chalela R, Bellido S, et al. Adaptive NKG2C+ natural killer cells are related to exacerbations and nutritional abnormalities in COPD patients. *Respir Res.* (2020) 21(1):63. doi: 10. 1186/s12931-020-1323-4
- 90. Marin A, Monsó E, Garcia-Nuñez M, Sauleda J, Noguera A, Pons J, et al. Variability and effects of bronchial colonisation in patients with moderate COPD. *Eur Respir J.* (2010) 35(2):295–302. doi: 10.1183/09031936.00126808
- 91. Barnes PJ. Chronic obstructive pulmonary disease: effects beyond the lungs. *PLoS Med.* (2010) 7(3):e1000220. doi: 10.1371/journal.pmed.1000220
- 92. Agustí AG, Noguera A, Sauleda J, Sala E, Pons J, Busquets X. Systemic effects of chronic obstructive pulmonary disease. *Eur Respir J.* (2003) 21(2):347–60. doi: 10. 1183/09031936.03.00405703

- 93. Evrard M, Kwok IWH, Chong SZ, Teng KWW, Becht E, Chen J, et al. Developmental analysis of bone marrow neutrophils reveals populations specialized in expansion, trafficking, and effector functions. *Immunity.* (2018) 48(2):364–79.e8. doi: 10.1016/j.immuni.2018.02.002
- 94. Naik D, Joshi A, Paul TV, Thomas N. Chronic obstructive pulmonary disease and the metabolic syndrome: consequences of a dual threat. *Indian J Endocrinol Metab.* (2014) 18(5):608–16. doi: 10.4103/2230-8210.139212
- 95. Halbert RJ, Isonaka S, George D, Iqbal A. Interpreting COPD prevalence estimates: what is the true burden of disease? *Chest.* (2003) 123(5):1684–92. doi: 10. 1378/chest.123.5.1684
- 96. Bose P, Sunita P, Pattanayak SP. Molecular insights into the crosstalk between immune inflammation nexus and SARS-CoV-2 virus. *Curr Microbiol.* (2021) 78 (11):3813–28. doi: 10.1007/s00284-021-02657-9
- 97. Finkelstein J, Cha E, Scharf SM. Chronic obstructive pulmonary disease as an independent risk factor for cardiovascular morbidity. *Int J Chron Obstruct Pulmon Dis.* (2009) 4:337–49. doi: 10.2147/COPD.S6400
- 98. Roversi S, Roversi P, Spadafora G, Rossi R, Fabbri LM. Coronary artery disease concomitant with chronic obstructive pulmonary disease. *Eur J Clin Invest.* (2014) 44 (1):93–102. doi: 10.1111/eci.12181
- 99. Wedzicha JA, Seemungal TAR, MacCallum PK, Paul EA, Donaldson GC, Bhowmik A, et al. Acute exacerbations of chronic obstructive pulmonary disease are accompanied by elevations of plasma fibrinogen and Serum IL-6 levels. *Thromb Haemost.* (2000) 84(08):210–5. doi: 10.1055/s-0037-1613998
- 100. Pike D, Lindenmaier T, Sin D, Parraga G. Imaging evidence of the relationship between atherosclerosis and chronic obstructive pulmonary disease. *Imaging Med.* (2014) 6:53–73. doi: 10.2217/iim.13.70
- 101. Zinellu A, Zinellu E, Mangoni AA, Pau MC, Carru C, Pirina P, et al. Clinical significance of the neutrophil-to-lymphocyte ratio and platelet-to-lymphocyte ratio in acute exacerbations of COPD: present and future. *Eur Respir Rev.* (2022) 31 (166):220095. doi: 10.1183/16000617.0095-2022
- 102. Brekke PH, Omland T, Holmedal SH, Smith P, Søyseth V. Determinants of cardiac troponin T elevation in COPD exacerbation—a cross-sectional study. *BMC Pulm Med.* (2009) 9:35. doi: 10.1186/1471-2466-9-35
- 103. Patel AR, Kowlessar BS, Donaldson GC, Mackay AJ, Singh R, George SN, et al. Cardiovascular risk, myocardial injury, and exacerbations of chronic obstructive pulmonary disease. *Am J Respir Crit Care Med.* (2013) 188(9):1091–9. doi: 10.1164/rccm.201306-1170OC
- 104. Santos S, Peinado VI, Ramírez J, Melgosa T, Roca J, Rodriguez-Roisin R, et al. Characterization of pulmonary vascular remodelling in smokers and patients with mild COPD. *Eur Respir J.* (2002) 19(4):632–8. doi: 10.1183/09031936.02.00245902
- 105. Ball MK, Waypa GB, Mungai PT, Nielsen JM, Czech L, Dudley VJ, et al. Regulation of hypoxia-induced pulmonary hypertension by vascular smooth muscle hypoxia-inducible factor-1α. Am J Respir Crit Care Med. (2014) 189(3):314–24. doi: 10.1164/rccm.201302-0302OC
- 106. Abe H, Semba H, Takeda N. The roles of hypoxia signaling in the pathogenesis of cardiovascular diseases. *J Atheroscler Thromb*. (2017) 24(9):884–94. doi: 10.5551/jat.RV17009
- 107. Huang S, Frangogiannis NG. Anti-inflammatory the rapies in myocardial infarction: failures, hopes and challenges. $\it Br~J~Pharmacol.~(2018)~175(9):1377-400.$ doi: 10.1111/bph.14155
- 108. Chen ZH, Kim HP, Ryter SW, Choi AM. Identifying targets for COPD treatment through gene expression analyses. *Int J Chron Obstruct Pulmon Dis.* (2008) 3(3):359–70. doi: 10.2147/copd.s1758
- 109. Huiart L, Ernst P, Ranouil X, Suissa S. Low-dose inhaled corticosteroids and the risk of acute myocardial infarction in COPD. *Eur Respir J.* (2005) 25(4):634–9. doi: 10. 1183/09031936.05.00079004
- 110. Loke YK, Kwok CS, Singh S. Risk of myocardial infarction and cardiovascular death associated with inhaled corticosteroids in COPD. *Eur Respir J.* (2010) 35 (5):1003–21. doi: 10.1183/09031936.00095909
- 111. Huiart L, Ernst P, Ranouil X, Suissa S. Oral corticosteroid use and the risk of acute myocardial infarction in chronic obstructive pulmonary disease. *Can Respir J.* (2006) 13(3):134–8. doi: 10.1155/2006/935718
- 112. Giugliano GR, Giugliano RP, Gibson CM, Kuntz RE. Meta-analysis of corticosteroid treatment in acute myocardial infarction. *Am J Cardiol.* (2003) 91 (9):1055–9. doi: 10.1016/S0002-9149(03)00148-6
- 113. Aaron CP, Schwartz JE, Hoffman EA, Angelini E, Austin JHM, Cushman M, et al. A longitudinal cohort study of aspirin use and progression of emphysema-like lung characteristics on CT imaging: the MESA lung study. *Chest.* (2018) 154 (1):41–50. doi: 10.1016/j.chest.2017.11.031
- 114. Kunadian V, Wilson N, Stocken DD, Ali H, McColl E, Burns G, et al. Antiplatelet therapy in the primary prevention of cardiovascular disease in patients with chronic obstructive pulmonary disease: a randomised controlled proof-of-concept trial. *ERJ Open Res.* (2019) 5(3):00110–2019. doi: 10.1183/23120541.00110-2019
- 115. Rennard SI, Fogarty C, Kelsen S, Long W, Ramsdell J, Allison J, et al. The safety and efficacy of infliximab in moderate to severe chronic obstructive pulmonary

disease. Am J Respir Crit Care Med. (2007) 175(9):926–34. doi: 10.1164/rccm.

- 116. Weber A, Wasiliew P, Kracht M. Interleukin-1 (IL-1) pathway. Sci Signal. (2010) 3(105):cm1. doi: 10.1126/scisignal.3105cm1
- 117. Krumm B, Xiang Y, Deng J. Structural biology of the IL-1 superfamily: key cytokines in the regulation of immune and inflammatory responses. *Protein Sci.* (2014) 23(5):526–38. doi: 10.1002/pro.2441
- 118. Ridker PM. From C-reactive protein to interleukin-6 to interleukin-1: moving upstream to identify novel targets for atheroprotection. *Circ Res.* (2016) 118 (1):145–56. doi: 10.1161/CIRCRESAHA.115.306656
- 119. Toldo S, Schatz AM, Mezzaroma E, Chawla R, Stallard TW, Stallard WC, et al. Recombinant human interleukin-1 receptor antagonist provides cardioprotection during myocardial ischemia reperfusion in the mouse. *Cardiovasc Drugs Ther.* (2012) 26(3):273–6. doi: 10.1007/s10557-012-6389-x
- 120. Suzuki K, Murtuza B, Smolenski RT, Sammut IA, Suzuki N, Kaneda Y, et al. Overexpression of interleukin-1 receptor antagonist provides cardioprotection against ischemia-reperfusion injury associated with reduction in apoptosis. *Circulation*. (2001) 104(Suppl_1):I-308-I-13. doi: 10.1161/circ.104.suppl_1.I-308
- 121. Morton AC, Rothman AMK, Greenwood JP, Gunn J, Chase A, Clarke B, et al. The effect of interleukin-1 receptor antagonist therapy on markers of inflammation in non-ST elevation acute coronary syndromes: the MRC-ILA heart study. *Eur Heart J.* (2015) 36(6):377–84. doi: 10.1093/eurheartj/ehu272
- 122. Ridker PM, Everett BM, Thuren T, MacFadyen JG, Chang WH, Ballantyne C, et al. Antiinflammatory therapy with canakinumab for atherosclerotic disease. *N Engl J Med.* (2017) 377(12):1119–31. doi: 10.1056/NEJMoa1707914
- 123. Castro P, Legora-Machado A, Cardilo-Reis L, Valença S, Porto LC, Walker C, et al. Inhibition of interleukin-1β reduces mouse lung inflammation induced by exposure to cigarette smoke. *Eur J Pharmacol.* (2004) 498(1):279–86. doi: 10.1016/j.ejphar.2004.07.047
- 124. Hernandez ML, Mills K, Almond M, Todoric K, Aleman MM, Zhang H, et al. IL-1 receptor antagonist reduces endotoxin-induced airway inflammation in healthy volunteers. *J Allergy Clin Immunol.* (2015) 135(2):379–85. doi: 10.1016/j.jaci.2014.
- 125. Buhl A-L, Wenzel J. Interleukin-36 in infectious and inflammatory skin diseases. Front Immunol. (2019) 10:1162. doi: 10.3389/fimmu.2019.01162
- 126. Ainscough JS, Macleod T, McGonagle D, Brakefield R, Baron JM, Alase A, et al. Cathepsin S is the major activator of the psoriasis-associated proinflammatory cytokine IL-36*y. Proc Natl Acad Sci U S A.* (2017) 114(13):E2748–57. doi: 10.1073/pnas.1620954114
- 127. Hahn M, Frey S, Hueber AJ. The novel interleukin-1 cytokine family members in inflammatory diseases. *Curr Opin Rheumatol.* (2017) 29(2):208–13. doi: 10.1097/BOR.000000000000361
- 128. Johnston A, Xing X, Wolterink L, Barnes DH, Yin Z, Reingold L, et al. IL-1 and IL-36 are dominant cytokines in generalized pustular psoriasis. *J Allergy Clin Immunol.* (2017) 140(1):109–20. doi: 10.1016/j.jaci.2016.08.056
- 129. Wei Y, Lan Y, Zhong Y, Yu K, Xu W, Zhu R, et al. Interleukin-38 alleviates cardiac remodelling after myocardial infarction. *J Cell Mol Med.* (2020) 24 (1):371–84. doi: 10.1111/jcmm.14741
- 130. El-Awaisi J, Kavanagh DPJ, Rink MR, Weston CJ, Drury NE, Kalia N. Targeting IL-36 improves age-related coronary microcirculatory dysfunction and attenuates myocardial ischemia/reperfusion injury in mice. *JCI Insight*. (2022) 7(5): e155236. doi: 10.1172/jci.insight.155236
- 131. El-Awaisi J, Mitchell JL, Ranasinghe A, Kalia N. Interleukin-36 is vasculoprotective in both sexes despite sex-specific changes in the coronary microcirculation response to IR injury. *Front Cardiovasc Med.* (2023) 10:1227499. doi: 10.3389/fcvm.2023.1227499
- 132. Koss CK, Wohnhaas CT, Baker JR, Tilp C, Przibilla M, Lerner C, et al. IL36 Is a critical upstream amplifier of neutrophilic lung inflammation in mice. *Communications Biology.* (2021) 4(1):172. doi: 10.1038/s42003-021-01703-3
- 133. Vanfleteren LE, Spruit MA, Groenen M, Gaffron S, van Empel VP, Bruijnzeel PL, et al. Clusters of comorbidities based on validated objective measurements and systemic inflammation in patients with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med.* (2013) 187(7):728–35. doi: 10.1164/rccm.201209-1665OC
- 134. Kobara M, Noda K, Kitamura M, Okamoto A, Shiraishi T, Toba H, et al. Antibody against interleukin-6 receptor attenuates left ventricular remodelling after myocardial infarction in mice. *Cardiovasc Res.* (2010) 87(3):424–30. doi: 10.1093/cvr/cvq078
- 135. Müller J, Gorressen S, Grandoch M, Feldmann K, Kretschmer I, Lehr S, et al. Interleukin-6-dependent phenotypic modulation of cardiac fibroblasts after acute myocardial infarction. *Basic Res Cardiol.* (2014) 109(6):440. doi: 10.1007/s00395-014-0440.x
- 136. Anstensrud AK, Woxholt S, Sharma K, Broch K, Bendz B, Aakhus S, et al. Rationale for the ASSAIL-MI-trial: a randomised controlled trial designed to assess the effect of tocilizumab on myocardial salvage in patients with acute ST-elevation myocardial infarction (STEMI). *Open Heart.* (2019) 6(2):e001108. doi: 10.1136/openhrt-2019-001108

- 137. Broch K, Anstensrud AK, Woxholt S, Sharma K, Tøllefsen IM, Bendz B, et al. Randomized trial of interleukin-6 receptor inhibition in patients with acute ST-segment elevation myocardial infarction. *J Am Coll Cardiol.* (2021) 77(15):1845–55. doi: 10.1016/j.jacc.2021.02.049
- 138. Wei Y-Y, Zhang D-W, Ye J-J, Lan Q-X, Ji S, Sun L, et al. Interleukin-6 neutralizing antibody attenuates the hypersecretion of airway mucus via inducing the nuclear translocation of Nrf2 in chronic obstructive pulmonary disease. *Biomed Pharmacother*. (2022) 152:113244. doi: 10.1016/j.biopha.2022.
- 139. Marinković G, Koenis DS, de Camp L, Jablonowski R, Graber N, de Waard V, et al. S100a9 links inflammation and repair in myocardial infarction. *Circ Res.* (2020) 127(5):664–76. doi: 10.1161/CIRCRESAHA.120.315865
- 140. Cai Z, Xie Q, Hu T, Yao Q, Zhao J, Wu Q, et al. S100a8/A9 in myocardial infarction: a promising biomarker and therapeutic target. Front Cell Dev Biol. (2020) 8:603902. doi: 10.3389/fcell.2020.603902

- 141. Zhao B, Lu R, Chen J, Xie M, Zhao X, Kong L. S100a9 blockade prevents lipopolysaccharide-induced lung injury via suppressing the NLRP3 pathway. *Respir Res.* (2021) 22(1):45. doi: 10.1186/s12931-021-01641-y
- $142.\ Hiroshima\ Y,\ Hsu\ K,\ Tedla\ N,\ Wong\ SW,\ Chow\ S,\ Kawaguchi\ N,\ et\ al.\ S100a8/$ A9 and S100A9 reduce acute lung injury. $Immunol\ Cell\ Biol.\ (2017)\ 95(5):461-72.$ doi: 10.1038/icb.2017.2
- 143. Lin CR, Bahmed K, Criner GJ, Marchetti N, Tuder RM, Kelsen S, et al. S100a8 protects human primary alveolar type II cells against injury and emphysema. *Am J Respir Cell Mol Biol.* (2019) 60(3):299–307. doi: 10.1165/rcmb.2018-0144OC
- 144. Matter MA, Paneni F, Libby P, Frantz S, Stähli BE, Templin C, et al. Inflammation in acute myocardial infarction: the good, the bad and the ugly. *Eur Heart J.* (2023) 45(2):89–103. doi: 10.1093/eurheartj/ehad486
- 145. King PT. Inflammation in chronic obstructive pulmonary disease and its role in cardiovascular disease and lung cancer. *Clin Transl Med.* (2015) 4(1):68. doi: 10.1186/s40169-015-0068-z