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# Whole-Exome sequencing analysis identified *TMSB10/TRABD2A* locus to be associated with carfilzomib-related cardiotoxicity among patients with multiple myeloma

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**Background:** Proteasome inhibitor Carfilzomib (CFZ) is effective in treating patients with refractory or relapsed multiple myeloma (MM) but has been associated with cardiovascular adverse events (CVAE) such as hypertension, cardiomyopathy, and heart failure. This study aimed to investigate the contribution of germline genetic variants in protein-coding genes in CFZ-CVAE among MM patients using whole-exome sequencing (WES) analysis.

**Methods:** Exome-wide single-variant association analysis, gene-based analysis, and rare variant analyses were performed on 603,920 variants in 247 patients with MM who have been treated with CFZ and enrolled in the Oncology Research Information Exchange Network (ORIEN) at the Moffitt Cancer Center. Separate analyses were performed in European Americans and African Americans followed by a trans-ethnic meta-analysis.

**Results:** The most significant variant in the exome-wide single variant analysis was a missense variant rs7148 in the thymosin beta-10/TraB Domain Containing 2A (*TMSB10/TRABD2A*) locus. The effect allele of rs7148 was associated with a higher risk of CVAE [odds ratio (OR) = 9.3 with a 95% confidence interval of 3.9–22.3,  $p = 5.42 \times 10^{-7}$ ]. MM patients with rs7148 AG or AA genotype had a higher risk of CVAE (50%) than those with GG genotype (10%). rs7148 is an expression quantitative trait locus (eQTL) for *TRABD2A* and *TMSB10*. The gene-based analysis also showed *TRABD2A* as the most significant gene associated with CFZ-CVAE ( $p = 1.06 \times 10^{-6}$ ).

**Conclusions:** We identified a missense SNP rs7148 in the *TMSB10/TRABD2A* as associated with CFZ-CVAE in MM patients. More investigation is needed to understand the underlying mechanisms of these associations.

#### KEYWORDS

cardio-oncology, proteasome inhibitors, Multiple Myeloma, carfilzomib, cardiotoxicity, whole exome sequencing

## 1. Introduction

Multiple Myeloma (MM) is a malignancy of the plasma cells. According to yearly incidence rates and prevalence figures, it ranks third among hematologic cancers in the United States (1). Proteasome inhibitors (PIs) are one of the most effective drugs for the treatment of MM and are the backbone therapies for MM treatment (2). Three PIs have been approved by the United States Food and Drug Administration: bortezomib, carfilzomib, and ixazomib (3). Carfilzomib (Kyprolis®) is a second-generation, irreversible PI approved for treating relapsed and refractory MM due to its survival benefit and overall response rate in refractory MM patients (4, 5). The National Comprehensive Cancer Network (NCCN) Guideline recommends carfilzomib-based therapy for newly diagnosed MM patients with pre-existing neuropathy or high-risk patients and patients with relapsed and refractory disease (6). Despite its effectiveness, carfilzomib (CFZ) has been shown to have significant cardiovascular adverse events (CVAE) (20%–25%), including 7.2% incident HF, in the clinical trials that excluded patients with pre-existing cardiovascular disorders (7). In the previous two meta-analyses studies, CFZ was associated with a high incidence of CVAE (8%–18%) (2, 3) including hypertension, heart failure, cardiomyopathy, and arrhythmia (8, 9). The rate of CVAE was even higher (~50%) in an observational study when MM patients with pre-existing cardiovascular conditions were not excluded (10). A severe clinical implication developed from this cardiotoxicity is treatment interruption, which could lead to disease progression (11).

Studies have shown that early detection of and early intervention for cardiotoxicity induced by other therapies (i.e., anthracyclines, Trastuzumab) can improve long-term outcomes (12, 13). Therefore, stratifying patients before the carfilzomib treatment might provide opportunities for early intervention to optimize patient outcomes. Pharmacogenomics, or the identification of genetic determinants of drug response and adverse effects, is a tool that has been useful in individualizing medication therapy (14, 15). This study aims to identify germline genetic variants in protein-coding genes associated with CFZ-CVAE in MM patients using whole-exome sequencing (WES) analysis.

## 2. Material and methods

### 2.1. Patients

Patients included in this study were admitted to the Moffitt Cancer Center's Total Cancer Care (TCC) Protocol with IRB

approval (MCC#14690; Advarra IRB Pro00014441) (16). Patients consented to contribute blood specimens and medical information for research purposes in collaboration with the Oncology Research Information Exchange Network (ORIEN), a network of seventeen cancer centers that have agreed to implement a common TCC biospecimen collection protocol to follow patients throughout their lifetime (17). Clinical and epidemiological data were collected for select TCC-consented patients, and the molecular data were produced as described below. This study included a total of 247 patients who have been diagnosed with MM and treated with carfilzomib at the H. Lee Moffitt Cancer Center and have germline DNA WES data available through ORIEN. This study was also approved by the University of Florida Institutional Review Board (IRB202003031).

### 2.2. Cardiovascular adverse events

We queried the electronic health records data of Moffitt's health research informatics platform to determine if a CVAE had occurred after initiation of carfilzomib treatment. We used the International Classification of Disease (ICD) revisions 9 and 10 to define cardiovascular events. A complete list of the ICD-9 and -10 codes used is listed in the supplementary materials **Supplementary Table S1**. Moffitt's Pentecost Myeloma Research Center clinical database was utilized to identify if the eligible TCC patients receiving carfilzomib as part of their treatment had one of these cardiac events between the start and end date. We performed a manual chart review on 10% of patients to verify the accuracy of the billing records in terms of the definition of the CVAE. All records reviewed for cardiovascular adverse events matched the determination by the billing codes.

### 2.3. Whole-Exome sequencing and quality control

Germline DNA was extracted from peripheral blood samples with buffy coat using the QIAasymphony SP instrument (QIAGEN, Hilden, Germany) following standard protocols implemented across the ORIEN. The WES of germline DNA was performed for each patient using SeqCap EZ Exome Enrichment Kit v3.0 (Roche NimbleGen, Pleasanton, CA) or xGen Exome Hybridization Panel with supplement probes (integrated Data Technologies, Inc., Coralville, IA), with 100× coverage. Capture kits covered variants for limited regions; each captured library was loaded onto Illumina-HiSeq 4,000 (Illumina, San Diego,

CA). Over 26,000 protein-coding genes were sequenced. The raw sequencing data underwent a rigorous analysis pipeline for alignment, variant calling, quality control steps, and annotation algorithms (18–20). Before the genetic association analysis, the WES data underwent additional quality control steps: variant call rate > 95%, sample call rate > 95%, sex check, and Hardy Weinberg Equilibrium analysis. Principle component analysis was performed on a subset of variants after more stringent quality control steps [variant rate and sample call rate > 99% and minor allele frequency (MAF) >10%] to evaluate the genetic ancestry of these patients.

Germline WES data was available on 605,446 variants in 247 patients. Of the 247 patients, 228 were genetically clustered with individuals of European ancestry, and 19 were clustered with African ancestry. A total of 603,920 variants passed the quality control steps and were included in the WES analysis.

## 2.4. Whole-Exome sequencing data analysis

All association analyses were performed in genetically clustered European Americans and African Americans separately, adjusting for age, gender, and principal components for ancestry. Trans-ethnic meta-analyses were then performed to combine the results from both groups.

Exome-wide association analysis of single variants with a MAF  $\geq 1\%$  was performed to estimate the odds ratio (OR) and 95% confidence interval (CI) for each variant on chromosomes 1–22 for the development of CFZ-CVAE using multivariable logistic regression assuming an additive model of inheritance using PLINK (21). All variants with  $p < 5 \times 10^{-8}$  were considered statistical significance. Variants with  $p < 5 \times 10^{-4}$  were considered suggestive (22).

Following the exome-wide association analysis, the summary statistics were functionally annotated using Functional Mapping and Annotation of Genome-Wide Association Studies (FUMA GWAS) for a gene-based and gene set analysis to recognize potential genes of interest (23). The Genotype-Tissue Expression (GTEx) database was queried to identify tissue-specific gene expression and regulation. ANNOVAR (24) was used to annotate the genetic variants appropriately.

Rare variants analysis was performed using the sequence kernel association test (SKAT) using the SKAT package (25, 26) to evaluate the association of the joint effect of multiple rare variants (MAF < 1%) with CFZ-CVAE.

## 2.5. Ingenuity pathway analysis (IPA)

Functional assignment and pathway analysis of the association results was performed on the top variants from the WES analyses ( $p < 0.001$ ). IPA uses a network generation algorithm to create multiple networks and uses hypergeometric distribution to create scores for each network (27). The statistical significance level is generated using Fisher's Exact test. Any pathway enriched by genes more than by chance would be statistically significant.

## 3. Results

### 3.1. Patients characteristics

Overall, 247 MM patients were included in the analysis. The mean age was  $\sim 59$  years, and 57% were men. Thirty-eight (15.5%) developed CVAE after initiation and during carfilzomib-based therapy. Table 1 summarizes the characteristics of the 38 patients who developed CVAE and the 209 who did not. The baseline demographics and medical history were similar between the two groups of patients. A total of 228 (92.3%) patients were genetically clustered with individuals of European ancestry (EA), and 19 clustered with individuals of African ancestry (AA). The baseline characteristics and medical history of the 228 EA MM patients were summarized in Supplementary Table S2.

### 3.2. Exome-wide common variant analysis

The results of the exome-wide association analysis of the common variants in the EA patients are summarized in the Manhattan plot (Figure 1A) and the QQ plot (Figure 1B). None of the variants were genome-wide significant. However, eleven SNPs from five loci reached the suggestive significance level with  $p < 5 \times 10^{-4}$  (Table 2). The top SNP rs7148 is a missense variant in the thymosin beta-10 (*TMSB10*) gene, with OR of 9.33% and 95% CI of 3.90 – 22.35 ( $p = 5.42 \times 10^{-07}$ ) (Table 2, Figure 2). The minor allele frequency rs7148 was  $\sim 7\%$  in EA patients. Amongst patients with the rs7148 variant, 50% (99/198) of patients with the AG or AA genotype developed a CVAE compared with 10.1% (20/198) with the GG genotype ( $p < 0.0001$ ). GTEx analysis revealed that the rs7148 A allele was associated with higher TraB Domain Containing 2A (*TRABD2A*) gene expression in the left ventricle tissue with a  $p$ -value  $1.9 \times 10^{-7}$  (Supplementary Figure S1). The second most significant SNP, rs12471929, is an intronic variant in the *TMSB10* gene that is in

TABLE 1 Demographics and clinical characteristics of patients.

Characteristics	Overall (n = 247)	CVAE (n = 38)	No CVAE (n = 209)	P
Age (years)	58.9 $\pm$ 9.8	60.6 $\pm$ 9.4	58.6 $\pm$ 9.8	0.27
Sex (male)	141 (57.1%)	21 (55.3%)	120 (57.4%)	0.81
Race				
European American	228 (92.3%)	35 (92.1%)	193 (92.3%)	0.96
African American	19 (7.7%)	3 (7.9%)	16 (7.7%)	
Medical History				
Hyperlipidemia	78 (31.6%)	15 (39.4%)	63 (30.1%)	0.26
Hypertension	130 (52.6%)	25 (65.8%)	105 (50.2%)	0.077
Diabetes	22 (8.9%)	6 (15.8%)	16 (7.7%)	0.11
Ischemic Heart Disease	5 (2.0%)	1 (2.6%)	4 (1.9%)	0.77
Myocardial Infarction	12 (4.9%)	3 (7.9%)	9 (4.3%)	0.34

Continuous variables were summarized as mean  $\pm$  standard deviation, and categorical variables were presented as numbers (%).  $P$  values shown were from a  $t$ -test for continuous variables and chi-squared test for categorical variables. CVAE: cardiovascular adverse events.

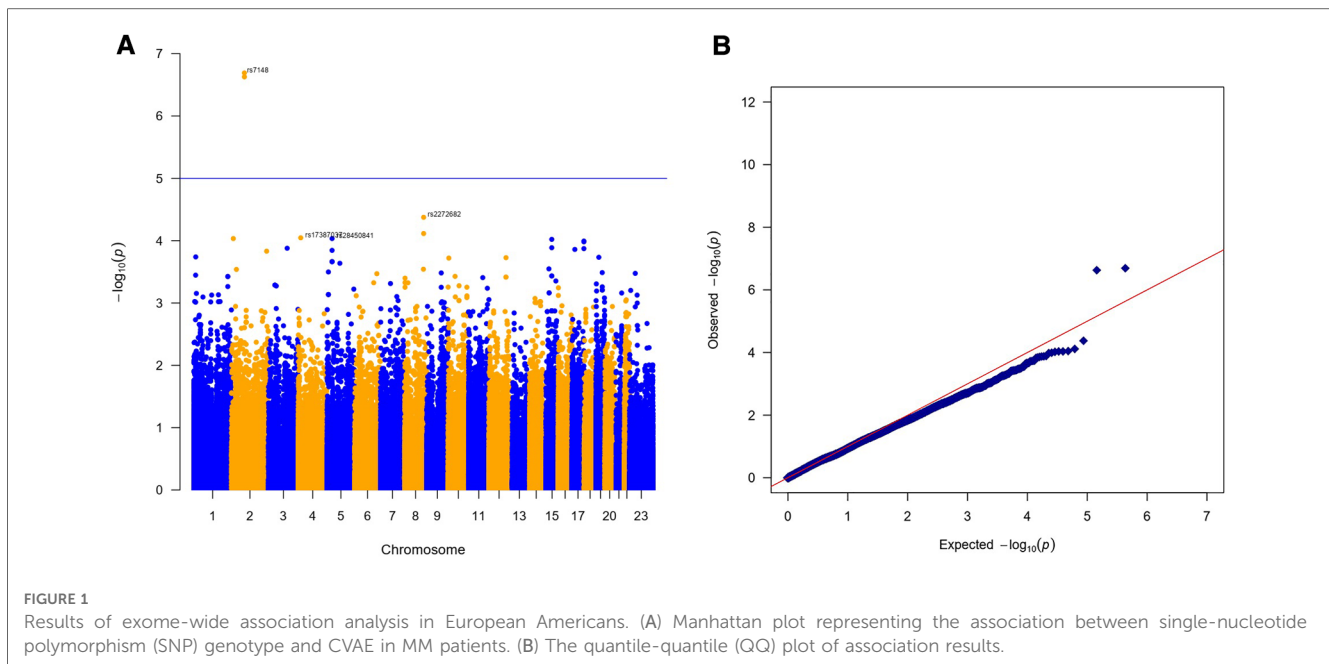


FIGURE 1

Results of exome-wide association analysis in European Americans. (A) Manhattan plot representing the association between single-nucleotide polymorphism (SNP) genotype and CVAE in MM patients. (B) The quantile-quantile (QQ) plot of association results.

perfect linkage disequilibrium (LD) ( $r^2 = 1$ ,  $D' = 1$ ) with rs7148, with OR of 8.95 (3.78–21.18),  $p = 6.17 \times 10^{-7}$  (Table 2, Figure 2). A few other SNPs in LD with rs7148 in the 1,000 genomes database are shown in Supplementary Table S3.

Among the other SNPs with a suggestive level of significance were: five SNPs in the *PDZD2* gene, which encodes PDZ Domain Containing Protein 2; an intronic variant in the Prominin 1, *CD133* (*PROM1*) gene; a synonymous variant on *GREB1* (Growth Regulating Estrogen Receptor Binding 1) gene on Chromosome 2, and two variants in *WASHC5/SQLE* gene on Chromosome 8 (Table 2). The minor allele frequencies, allele counts, and Hardy Weinberg Equilibrium test results by CVAE status are summarized in Supplementary Table S4.

We also performed an exploratory analysis on AA patients. No SNPs reached statistical significance as a result of the small sample size. The four SNPs with nominal significance ( $p < 0.05$ ) are shown in Supplementary Table S5. The two most frequent

SNPs in EA (rs7148, rs1247192) were not observed in AA patients (MAF = 0).

The top SNPs in the trans-ethnic meta-analysis combining EA and AA are listed in Table 3. Only two of these eleven SNPs were observed in AA. Therefore, the results of these SNPs in the meta-analysis were almost identical to those in the EA analysis. The only top EA SNPs observed in AA were the Chromosome 8 SNPs in the *WASHC5/SQLE* locus. While these two SNPs had minor allele frequencies of 6%–9% in EA, the frequencies were much higher in AA (42%–45%). The directions of associations with CVAE were consistent in AA patients compared to those in EA patients (Table 3).

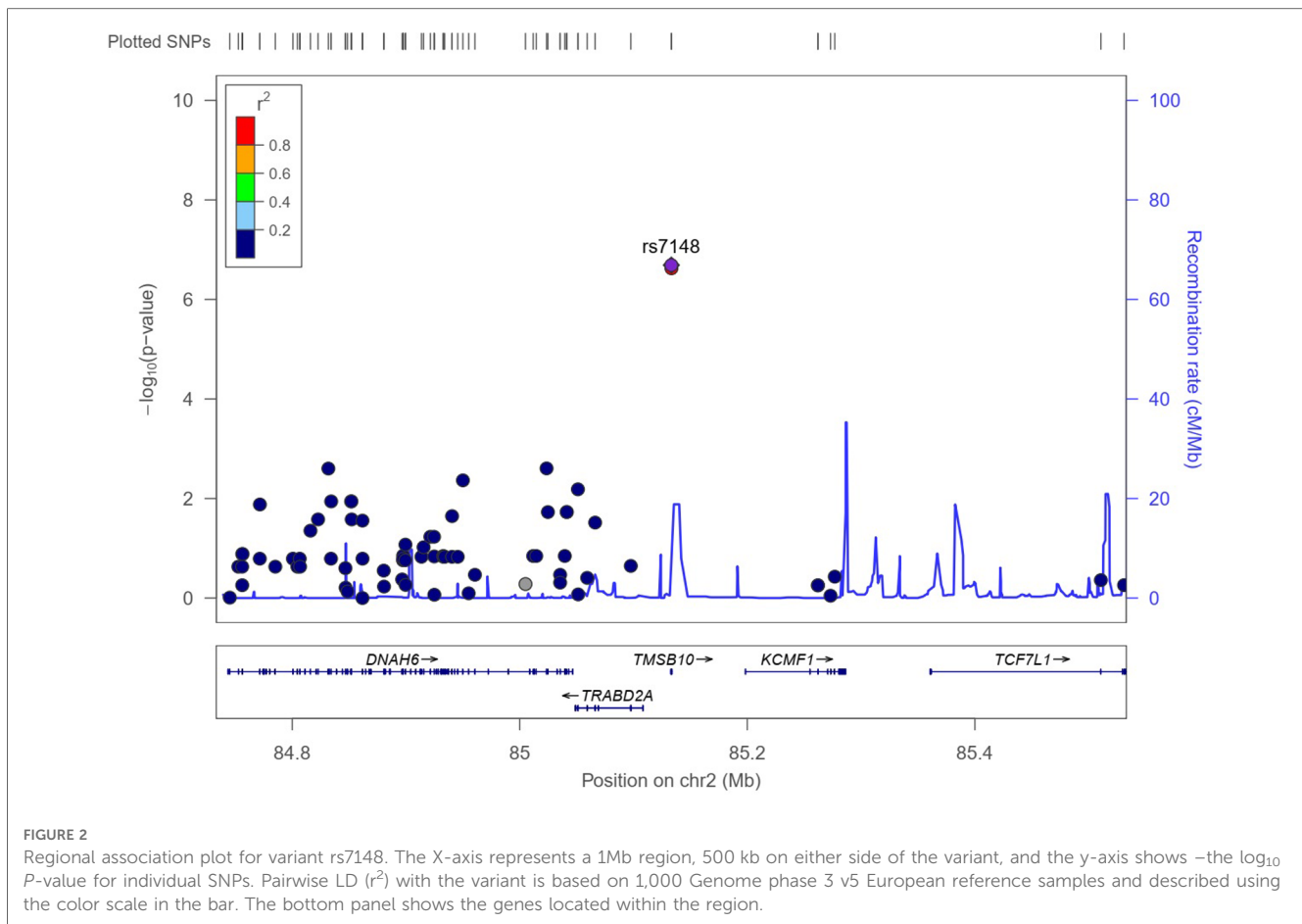
### 3.3. Gene-based analysis

The gene-based analysis in EA patients using FUMA revealed that the *TRABD2A* gene that encodes TraB Domain Containing

TABLE 2 Top SNPs in the WES analysis of CFZ-CVAE in the European American patients.

CHR	SNP	BP	Gene	A1	A2	MAF	OR	95% CI	P	dbSNP functional annotation	MAF CVAE		MAC CVAE	
											No	Yes	No	Yes
2	rs7148	85,13,3216	TMSB10	A	G	0.068	9.33	3.90–22.35	5.42E-07	Missense	0.039	0.229	15	16
2	rs12,47,1929	85,13,3320	TMSB10	T	C	0.070	8.95	3.78–21.18	6.17E-07	Intronic	0.039	0.243	15	17
5	rs28,45,0841	32,09,3210	PDZD2	T	C	0.077	5.25	2.38–11.58	3.91E-05	Intronic	0.052	0.214	20	15
5	rs22,91,113	32,07,4509	PDZD2	A	G	0.057	6.17	2.52–15.15	7.05E-05	Synonymous	0.036	0.171	14	12
4	rs17,38,7037	15,99,2783	PROM1	C	A	0.057	5.84	2.44–139.5	7.32E-05	Intronic	0.036	0.171	14	12
2	rs75,34,8511	11,73,8951	GREB1	T	C	0.037	8.30	2.90–23.77	8.08E-05	Synonymous	0.021	0.129	8	9
8	rs22,72,682	12,60,49443	WASHC5	C	T	0.094	3.83	1.95–7.49	9.26E-05	Intronic	0.065	0.257	25	18
8	rs11,54,2889	12,60,44527	WASHC5/SQLE	T	C	0.059	5.01	2.23–11.24	9.50E-05	Synonymous	0.034	0.200	13	14
5	rs37,33,720	32,08,7808	PDZD2	C	G	0.072	4.90	2.20–10.87	9.55E-05	Synonymous	0.049	0.200	19	14
5	rs10,06,6063	32,09,0294	PDZD2	A	G	0.072	4.90	2.20–10.87	9.55E-05	Missense	0.049	0.200	19	14
5	rs16,88,9442	32,09,3070	PDZD2	A	G	0.072	4.90	2.20–10.87	9.55E-05	Intronic	0.049	0.200	19	14

WES, whole exome sequencing; CFZ-CVAE, carfilzomib-related cardiovascular adverse events; Chr., chromosome; SNP, single nucleotide polymorphism; BP: base pair. A1: minor allele; A2: major allele; MAF, minor allele frequency; OR: odds ratio; CI, confidence interval, MAC, minor allele counts.



2A is significantly associated with CFZ-CVAE ( $p = 1.06 \times 10^{-6}$ ) (Figure 3).

### 3.4. Rare variant analysis performed using SKAT

SKAT analysis was performed on 40,969 gene sets and 620,661 SNPs, and a significant association was determined by comparing CFZ-CVAE after correcting for multiple testing. The genes—Chromosome 1 Open Reading Frame 116 (C1orf116), LOC102724084 (DYNLRB2 antisense RNA1), Diphosphoinositol pentakisphosphate kinase 2 (PPIP5K2 (NM\_0013)), and Transmembrane Protein 183A (TMEM183A) were statistically significantly associated with CFZ-CVAE ( $p$ -value =  $1.1 \times 10^{-5}$ ,  $4.1 \times 10^{-5}$ ,  $6.2 \times 10^{-5}$ , and  $6.2 \times 10^{-5}$ , respectively) (Supplementary Table S6).

### 3.5. IPA analysis of WES results

Using IPA, the pathway enrichment analysis showed that the lowest  $p$ -value and most significant genes overlapped with cardiotoxicity. The functional toxicity annotation of genes related to cardiotoxicity implicated cardiac arteriopathy with three variants from WES results: rs3750765 located in leucine-rich

repeat-containing 20 (LRRC20) gene, rs72713436 in sterile alpha motif domain-containing 4A (SAMD4A) gene, and rs75454001 in CUB-Sushi multiple domains 1 (CSMD1) gene. All of these genes were associated with human coronary artery disease ( $P < 0.001$ ) (28).

## 4. Discussion

In this first genetic association analysis of CFZ-related CVAE in MM patients, we conducted a WES of germline DNA samples from patients who received CFZ in the ORIEN network. In this retrospective cohort study of patients in a real-world clinical setting, we identified a missense SNP rs7148 in the *TMSB10/TRABD2A* gene locus on chromosome 2 to be associated with a higher risk for CVAE in MM patients treated with CFZ.

In order to reflect real-world practices, all MM patients treated with CFZ were included in this study regardless of their cardiovascular disease history. We found that 15.4% of the MM patients treated with CFZ developed CVAE, which is in line with the event rates of 8%–18% reported in prior meta-analyses (2, 3). Due to the retrospective nature of this study and a lack of clinical guidelines on proteome inhibitor monitoring at the time of this study, these patients were not actively monitored for cardiotoxicity by cardiologists. Not surprisingly, the event rate we observed is lower than the CFZ-CVAE event rate of 50% reported in a prospective study, the Prospective Observation of



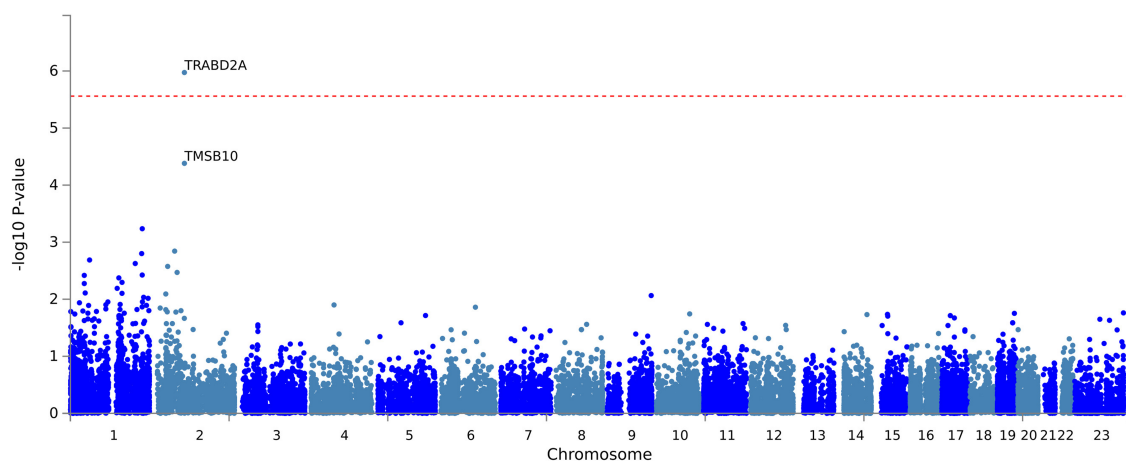


FIGURE 3

The gene-based association testing results from the FUMA analysis of CFZ-CVAE yielded *TRABD2A* as a significant gene ( $p = 1.06 \times 10^{-6}$ ).

dysfunction (42, 43), and *in vivo* studies showed that *TMSB4* increases EPC migration and decreases EPC apoptosis under serum deprivation *via* the (PI3K/Akt/eNOS) signal transduction pathway (44, 45), and several studies showed that the telomerase length and telomerase activity of circulating EPCs and decreased in patients with coronary artery disease (46, 47). In a recent study that included 48 patients with relapsed/refractory MM and received CFZ, the brachial artery flow-mediated dilation (FMD) and 26s proteasome activity were detected to evaluate the endothelial function. This study concluded that patients who received CFZ and with low potential for proteasome activity recovery may suffer from both acute and long-term endothelial dysfunction (48).

Endothelial cell homeostasis depends on the ubiquitin-proteasome system, which induces oxidative stress in the cells and regulates the expression of endothelial nitric oxide synthase (49). The proteasome inhibitor CFZ causes the plasma of cancer cells, cardiomyocytes, and endothelial cells to accumulate with unfolded, dysfunctional proteins, which may lead to impaired vasodilation, excessive oxidative stress, inflammation, cell apoptosis, and autophagy (50, 51). Other studies have shown that the endothelial dysfunction caused by CFZ's inhibition of proteasome activity may result in CVAE or other endothelial dysfunction-related events like hypertension, heart failure, and coronary artery disease (51–53). In light of the literature, our finding of the genetic variants in the *TMSB10/TRABD2A* locus appears to support the role of endothelial dysfunction in CFZ-CVAE.

Among the other SNPs with a suggestive significance level, five are located on the PDZ domain-containing protein 2 (*PDZD2*) gene, which contains six PDZ domains and shares sequence similarity with pro-interleukin-16 (pro-IL-16). SNPs in the *PDZD2* gene have been associated with heart rate in heart failure patients with reduced ejection fraction (54). *PROM1* has a role in cell differentiation, proliferation, and apoptosis. PRIP is a small peptide derived from the extracellular domain of *PROM1*-derived peptide and improves cardiac function following ischemia. *SQLE* encodes squalene epoxidase, which catalyzes the first oxygenation step in sterol biosynthesis and is one of the rate-limiting enzymes in this pathway.

It is important to recognize some limitations of our study. Firstly, the patient population is predominantly European Americans. The sample size of patients of African descent was too small to have enough statistical power for any meaningful discovery. Further investigation is required to explore this phenotype and outcomes in MM patients of African ancestry. Secondly, using WES means we may have missed critical genetic variants outside the coding regions of the genome. Thirdly, using ICD codes to identify CVAE has its limitations. Even though our manual chart review of 10% of patients indicated 100% of CVAE were confirmed, it would have been ideal to review all the charts to confirm CVAE status. Fourthly, due to the small sample size, we had to combine all the CVAEs. Lastly, our study findings need to be replicated in an independent study before these genetic variants can be incorporated into the risk stratification of MM patients.

In summary, in this WES study of MM patients in a real-world clinical setting, we identified a missense variant in the *TMSB10/TRABD2A* locus to be associated with CFZ-CVAE among MM patients. Once validated, this association could provide the basis for a Precision Medicine approach to identify MM patients at high risk for CFZ-CVAE.

## Data availability statement

The data presented in the study are deposited in the database of Genotypes and Phenotypes (dbGaP) repository, accession number phs003308.v1.p1.

## Ethics statement

The studies involving human participants were reviewed and approved by the University of Florida Institutional Review Board (IRB202003031). The patients/participants provided their written informed consent to participate in this study.

## Author contributions

MT and YG wrote the manuscript, and RA extracted the data from ORIEN, RCB manually reviewed the charts to validate the CVAE. GD, EMS, and KHS directed sample identification and contributed to the TCC and ORIEN project. MT, GY, and YG performed statistical analyses. YG secured the fund for this study. All authors contributed to the article and approved the submitted version.

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## References

1. Siegel RL, Miller KD, Fuchs HE, Jemal A. Cancer statistics, 2022. *CA Cancer J Clin.* (2022) 72:7–33. doi: 10.3322/caac.21708
2. Gandolfi S, Laubach JP, Hideshima T, Chauhan D, Anderson KC, Richardson PG. The proteasome and proteasome inhibitors in multiple myeloma. *Cancer Metastasis Rev.* (2017) 36:561–84. doi: 10.1007/s10555-017-9707-8
3. Teicher BA, Tomaszewski JE. Competitive landscape report. *Biochem Pharmacol.* (2015) 96:1–9. doi: 10.1016/j.bcp.2015.04.008
4. Hasinoff BB, Patel D, Wu X. Molecular mechanisms of the cardiotoxicity of the proteasomal-targeted drugs bortezomib and carfilzomib. *Cardiovasc Toxicol.* (2017) 17:237–50. doi: 10.1007/s12012-016-9378-7
5. Dimopoulos MA, Goldschmidt H, Niesvizky R, Joshua D, Chng WJ, Oriol A, et al. Carfilzomib or bortezomib in relapsed or refractory multiple myeloma (ENDEAVOR): an interim overall survival analysis of an open-label, randomised, phase 3 trial. *Lancet Oncol.* (2017) 18:1327–37. doi: 10.1016/S1470-2045(17)30578-8
6. Callander NS, Baljevic M, Adekola K, Anderson LD, Campagnaro E, Castillo JJ, et al. NCCN Guidelines<sup>®</sup> Insights: Multiple Myeloma, Version 3.2022. *J Natl Compr Canc Netw.* (2022) 20(1):8–19. doi: 10.6004/jncn.2022.0002
7. Siegel D, Martin T, Nooka A, Harvey RD, Vij R, Niesvizky R, et al. Integrated safety profile of single-agent carfilzomib: experience from 526 patients enrolled in 4 phase II clinical studies. *Haematologica.* (2013) 98(11):1753–61. doi: 10.3324/haematol.2013.089334
8. Kistler KD, Kalman J, Sahni G, Murphy B, Werther W, Rajangam K, et al. Incidence and risk of cardiac events in patients with previously treated multiple

## Conflict of interest

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The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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## Supplementary material

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fcvm.2023.1181806/full#supplementary-material>

- myeloma versus matched patients without multiple myeloma: an observational, retrospective, cohort study. *Clin Lymphoma Myeloma Leuk.* (2017) 17:89–96.e3. doi: 10.1016/j.clml.2016.11.009
9. Atrash S, Tullos A, Panozzo S, Bhutani M, Van Rhee F, Barlogie B, et al. Cardiac complications in relapsed and refractory multiple myeloma patients treated with carfilzomib. *Blood Cancer J.* (2015) 5:e272. doi: 10.1038/bcj.2014.93
  10. Cornell RF, Ky B, Weiss BM, Dahm CN, Gupta DK, Du L, et al. Prospective study of cardiac events during proteasome inhibitor therapy for relapsed multiple myeloma. *J Clin Oncol.* (2019) 37:1946–55. doi: 10.1200/JCO.19.00231
  11. Yu AF, Yadav NU, Lung BY, Eaton AA, Thaler HT, Hudis CA, et al. Trastuzumab interruption and treatment-induced cardiotoxicity in early HER2-positive breast cancer. *Breast Cancer Res Treat.* (2015) 149:489–95. doi: 10.1007/s10549-014-3253-7
  12. Koutsoukis A, Ntalianis A, Repasos E, Kastiris E, Dimopoulos MA, Paraskevaidis I. Cardio-oncology: a focus on cardiotoxicity. *European Cardiology Review.* (2018) 13:64–9. doi: 10.15420/ecr.2017:17:2
  13. Henry ML, Niu J, Zhang N, Giordano SH, Chavez-MacGregor M. Cardiotoxicity and cardiac monitoring among chemotherapy-treated breast cancer patients. *JACC Cardiovasc Imaging.* (2018) 11:1084–93. doi: 10.1016/j.jcmg.2018.06.005
  14. Shuldiner AR, O'Connell JR, Bliden KP, Gandhi A, Ryan K, Horenstein RB, et al. Association of cytochrome P450 2C19 genotype with the antiplatelet effect and clinical efficacy of clopidogrel therapy. *JAMA.* (2009) 302:849–57. doi: 10.1001/jama.2009.1232



15. Krynetski EY, Tai HL, Yates CR, Fessing MY, Loennechen T, Schuetz JD, et al. Genetic polymorphism of thiopurine S-methyltransferase: clinical importance and molecular mechanisms. *Pharmacogenetics*. (1996) 6:279–90. doi: 10.1097/00008571-199608000-00001
16. Fenstermacher DA, Wenham RM, Rollison DE, Dalton WS. Implementing personalized medicine in a cancer center. *Cancer J*. (2011) 17:528–36. doi: 10.1097/PPO.0b013e318238216e
17. Dalton WS, Sullivan D, Ecsedy J, Caligiuri MA. Patient enrichment for precision-based cancer clinical trials: using prospective cohort surveillance as an approach to improve clinical trials. *Clin Pharmacol Ther*. (2018) 104:23–6. doi: 10.1002/cpt.1051
18. Li H. A statistical framework for SNP calling, mutation discovery, association mapping and population genetical parameter estimation from sequencing data. *Bioinformatics* (2011) 27:2987–93. doi: 10.1093/bioinformatics/btr509
19. Depristo MA, Banks E, Poplin R, Garimella K V, Maguire JR, Hartl C, et al. A framework for variation discovery and genotyping using next-generation DNA sequencing data. *Nat Genet*. (2011) 43:491–501. doi: 10.1038/ng.806
20. Van der Auwera GA, Carneiro MO, Hartl C, Poplin R, Del Angel G, Levy-Moonshine A, et al. From FastQ data to high confidence variant calls: the Genome Analysis Toolkit best practices pipeline. *Curr Protoc Bioinformatics*. (2013) 43(1110):11.10.1–11.10.33. doi: 10.1002/0471250953.b1110s43
21. Purcell S, Neale B, Todd-Brown K, Thomas L, Ferreira MAR, Bender D, et al. PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am J Hum Genet*. (2007) 81:559–75. doi: 10.1086/519795
22. Fadista J, Manning AK, Florez JC, Groop L. The (in)famous GWAS P-value threshold revisited and updated for low-frequency variants. *Eur J Hum Genet*. (2016) 24:1202–5. doi: 10.1038/ejhg.2015.269
23. Watanabe K, Taskesen E, Van Bochoven A, Posthuma D. Functional mapping and annotation of genetic associations with FUMA. *Nat Commun*. (2017) 8:1826. doi: 10.1038/s41467-017-01261-5
24. Wang K, Li M, Hakonarson H. ANNOVAR: functional annotation of genetic variants from high-throughput sequencing data. *Nucleic Acids Res*. (2010) 38:e164. doi: 10.1093/NAR/GKQ603
25. Lee S, Abecasis GR, Boehnke M, Lin X. Rare-variant association analysis: study designs and statistical tests. *Am J Hum Genet*. (2014) 95:5–23. doi: 10.1016/j.ajhg.2014.06.009
26. Wu MC, Lee S, Cai T, Li Y, Boehnke M, Lin X. Rare-variant association testing for sequencing data with the sequence kernel association test. *Am J Hum Genet*. (2011) 89:82–93. doi: 10.1016/j.ajhg.2011.05.029
27. Krämer A, Green J, Pollard J Jr, Tugendreich S. Causal analysis approaches in Ingenuity Pathway Analysis. *Bioinformatics*. (2014) 30(4):523–30. doi: 10.1093/bioinformatics/btt703
28. Burton PR, Clayton DG, Cardon LR, Craddock N, Deloukas P, Duncanson A, et al. Genome-wide association study of 14,000 cases of seven common diseases and 3,000 shared controls. *Nature*. (2007) 447:661–78. doi: 10.1038/nature05911
29. Lyon AR, López-Fernández T, Couch LS, Asteggiano R, Aznar MC, Bergler-Klein J, et al. 2022 ESC guidelines on cardio-oncology developed in collaboration with the European hematology association (EHA), the European society for therapeutic radiology and oncology (ESTRO) and the international cardio-oncology society (IC-OS). *Eur Heart J*. (2022) 43:4229–361. doi: 10.1093/eurheartj/ehac244
30. Carithers LJ, Ardlie K, Barcus M, Branton PA, Britton A, Buia SA, et al. A novel approach to high-quality postmortem tissue procurement: the GTEx project. *Biopreserv Biobank*. (2015) 13:311–7. doi: 10.1089/bio.2015.0032
31. Santelli G, Califano D, Chiappetta G, Vento MT, Bartoli PC, Zullo F, et al. Thymosin beta-10 gene overexpression is a general event in human carcinogenesis. *Am J Pathol*. (1999) 155:799–804. doi: 10.1016/s0002-9440(10)65178-4
32. Chen C, Li M, Yang H, Chai H, Fisher W, Yao Q. Roles of thymosins in cancers and other organ systems. *World J Surg*. (2005) 29:264–70. doi: 10.1007/S00268-004-7817-2
33. Sribenja S, Li M, Wongkham S, Wongkham C, Yao Q, Chen C. Cancer investigation advances in thymosin  $\beta$ 10 research: differential expression, molecular mechanisms, and clinical implications in cancer and other conditions advances in thymosin  $\beta$ 10 research: differential expression, molecular mechanisms, and clinical implications in cancer and other conditions. *Cancer Invest*. (2009) 27:1016–22. doi: 10.3109/07357900902849640
34. Yu FX, Lin SC, Morrison-Bogorad M, Yin HL. Effects of thymosin beta 4 and thymosin beta 10 on actin structures in living cells. *Cell Motil Cytoskeleton*. (1994) 27:13–25. doi: 10.1002/cm.970270103
35. Nukala SB, Regazzoni L, Aldini G, Zodda E, Tura-Ceide O, Mills NL, et al. Differentially expressed proteins in primary endothelial cells derived from patients with acute myocardial infarction. *Hypertension*. (2019) 74:947–56. doi: 10.1161/HYPERTENSIONAHA.119.13472
36. Lee SH, Son MJ, Oh SH, Rho SB, Park K, Kim YJ, Park MS, Lee JH. Thymosin  $\beta$ 10 inhibits angiogenesis and tumor growth by interfering with ras function. *Cancer Res* (2005) 65:137–47. doi: 10.1158/0008-5472.137.65.1
37. Liu M, Dong J, Ouyang J, Zhao L, Liang G, Shang H. Metalloprotease TRABD2A restriction of HIV-1 production in monocyte-derived dendritic cells. *AIDS Res Hum Retroviruses*. (2019) 35:887–9. doi: 10.1089/AID.2019.0140
38. Nasu T, Satoh M, Hachiya T, Sutoh Y, Ohmomo H, Hitomi S, et al. A genome-wide association study for highly sensitive cardiac troponin T levels identified a novel genetic variation near a RBAK-ZNF890P locus in the Japanese general population. *Int J Cardiol*. (2021) 329:186–91. doi: 10.1016/j.ijcard.2020.12.019
39. Welsh P, Preiss D, Hayward C, Shah AS, McAllister D, Briggs A, et al. Cardiac troponin T and troponin I in the general population. *Circulation*. (2019) 139:2754–64. doi: 10.1161/CIRCULATIONAHA.118.038529
40. Saunders JT, Nambi V, de Lemos JA, Chambless LE, Virani SS, Boerwinkle E, et al. Cardiac troponin T measured by a highly sensitive assay predicts coronary heart disease, heart failure, and mortality in the atherosclerosis risk in communities study. *Circulation*. (2011) 123:1367–76. doi: 10.1161/CIRCULATIONAHA.110.005264
41. Efentakis P, Kremastiotis G, Varela A, Nikolou PE, Papanagnou ED, Davos CH, et al. Molecular mechanisms of carfilzomib-induced cardiotoxicity in mice and the emerging cardioprotective role of metformin. *Blood*. (2019) 133:710–23. doi: 10.1182/blood-2018-06-858415
42. Shi Q, Rafii S, Wu Hong-De M, Wijelath ES, Yu C, Ishida A, et al. Evidence for circulating bone marrow-derived endothelial cells. *Blood*. (1998) 92:362–7. doi: 10.1182/blood.v92.2.362
43. Asahara T, Murohara T, Sullivan A, Silver M, Van Der Zee R, Li T, et al. Isolation of putative progenitor endothelial cells for angiogenesis. *Science*. (1997) 275:964–7. doi: 10.1126/science.275.5302.964
44. Zhao Y, Qiu F, Xu S, Yu L, Fu G. Thymosin  $\beta$ 4 activates integrin-linked kinase and decreases endothelial progenitor cells apoptosis under serum deprivation. *J Cell Physiol*. (2011) 226:2798–806. doi: 10.1002/jcp.22624
45. Qiu FY, Song XX, Zheng H, Zhao YB, Fu GS. Thymosin  $\beta$ 4 induces endothelial progenitor cell migration via PI3K/akt/eNOS signal transduction pathway. *J Cardiovasc Pharmacol*. (2009) 53:209–14. doi: 10.1097/FJC.0b013e318199f326
46. Murasawa S, Llevadot J, Silver M, Isner JM, Losordo DW, Asahara T. Constitutive human telomerase reverse transcriptase expression enhances regenerative properties of endothelial progenitor cells. *Circulation*. (2002) 106:1133–9. doi: 10.1161/01.CIR.0000027584.85865.B4
47. Vasa M, Fichtlscherer S, Aicher A, Adler K, Urbich C, Martin H, et al. Number and migratory activity of circulating endothelial progenitor cells inversely correlate with risk factors for coronary artery disease. *Circ Res*. (2001) 89:E1–7. doi: 10.1161/hh1301.093953
48. Kastritis E, Laina A, Georgiopoulos G, Gavriatopoulou M, Papanagnou E-D, Eleutherakis-Papaikovou E, et al. Carfilzomib-induced endothelial dysfunction, recovery of proteasome activity, and prediction of cardiovascular complications: a prospective study. *Leukemia*. (2021) 35:1418–27. doi: 10.1038/s41375-021-01141-4
49. de Carvalho JE R, Verwoert MT, Vogels IMC, Reits EA, van Noorden CJF, Klaassen I, et al. Involvement of the ubiquitin-proteasome system in the expression of extracellular matrix genes in retinal pigment epithelial cells. *Biochem Biophys Rep*. (2018) 13:83–92. doi: 10.1016/j.bbrep.2018.01.005
50. Meiners S, Ludwig A, Stangl V, Stangl K. Proteasome inhibitors: poisons and remedies. *Med Res Rev*. (2008) 28:309–27. doi: 10.1002/med.20111
51. Brandes RP. Endothelial dysfunction and hypertension. *Hypertension*. (2014) 64:924–8. doi: 10.1161/HYPERTENSIONAHA.114.03575
52. Lerman A, Zeiher AM. Endothelial function: cardiac events. *Circulation*. (2005) 111:363–8. doi: 10.1161/01.CIR.0000153339.27064.14
53. Chen-Scarabelli C, Corsetti G, Pasini E, Dioguardi FS, Sahni G, Narula J, et al. Spasmogenic effects of the proteasome inhibitor carfilzomib on coronary resistance, vascular tone and reactivity. *EBioMedicine*. (2017) 21:206–12. doi: 10.1016/j.ebiom.2017.05.024
54. Evans KL, Wirtz HS, Li J, She R, Maya J, Gui H, et al. Genetics of heart rate in heart failure patients (GenHRate). *Hum Genomics*. (2019) 13:22. doi: 10.1186/s40246-019-0206-6
55. Tantawy M, Guang Y, Algebelli RR, Rubinstein SM, Fradley MG, Lu Q, et al. TMSB10/TRABD2A Locus associated with carfilzomib-related cardiotoxicity in multiple myeloma patients: a whole-exome sequencing analysis. *Clin Pharmacol Ther*. (2022) 111:S69-PW-004. doi: 10.1002/cpt.2521