



# Mitochondrial Dynamics and Mitophagy in Cardiometabolic Disease

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### Specialty section:

This article was submitted to  
Cardiovascular Metabolism,  
a section of the journal  
Frontiers in Cardiovascular Medicine

**Received:** 10 April 2022

**Accepted:** 20 May 2022

**Published:** 17 June 2022

### Citation:

Lin J, Duan J, Wang Q, Xu S, Zhou S  
and Yao K (2022) Mitochondrial  
Dynamics and Mitophagy in  
Cardiometabolic Disease.  
*Front. Cardiovasc. Med.* 9:917135.  
doi: 10.3389/fcvm.2022.917135

Mitochondria play a key role in cellular metabolism. Mitochondrial dynamics (fusion and fission) and mitophagy, are critical to mitochondrial function. Fusion allows organelles to share metabolites, proteins, and mitochondrial DNA, promoting complementarity between damaged mitochondria. Fission increases the number of mitochondria to ensure that they are passed on to their offspring during mitosis. Mitophagy is a process of selective removal of excess or damaged mitochondria that helps improve energy metabolism. Cardiometabolic disease is characterized by mitochondrial dysfunction, high production of reactive oxygen species, increased inflammatory response, and low levels of ATP. Cardiometabolic disease is closely related to mitochondrial dynamics and mitophagy. This paper reviewed the mechanisms of mitochondrial dynamics and mitophagy (focus on MFN1, MFN2, OPA1, DRP1, and PINK1 proteins) and their roles in diabetic cardiomyopathy, myocardial infarction, cardiac hypertrophy, heart failure, atherosclerosis, and obesity.

**Keywords:** mitochondrial dynamics, cardiometabolic disease, mitophagy, mitochondrial fission, mitochondrial fusion, diabetic cardiomyopathy, myocardial infarction

## INTRODUCTION

Cardiometabolic disease (CMD) is a clinical syndrome caused by genes (heredity, environment, behavior) and metabolic drivers (abnormal obesity, abnormal blood glucose, metabolic syndrome characteristics), including hypertension, diabetes, dyslipidemia, coronary heart disease, stroke, and other diseases (1, 2). The incidence of CMD increases gradually with the increase of age, which is the primary cause of death and disease burden of the global population. Unreasonable diet, lack of exercise, smoking, and excessive drinking are the main risk factors for the sharp increase in CMD (3). At present, pathogenesis, risk assessment, and treatment of CMD remain unclear. One of the most promising treatments for CMD is to improve cardiac metabolism and energy supply (4, 5).

The heart has a very high energy requirement, which must continuously produce large amounts of ATP through the metabolism of various fuels (fatty acids, glucose, lactic acid, pyruvate, and amino acids) to maintain systolic function. Mitochondrial oxidative phosphorylation (OXPHOS) contributes 95% of cardiac ATP requirements and glycolysis provides the remaining 5% (6). Cardiac energy disturbance is an important cause of most CMD (7). The ability of the heart to switch between different energy substrates is known as metabolic flexibility (8). As insulin resistance develops, the metabolic flexibility of the heart gradually decreases, making myocardial

energy production largely dependent on fatty acid oxidation. This shift leads to increased uptake and accumulation of lipids in the heart, resulting in lipotoxicity. At the same time, glycolytic intermediates accumulate in the heart due to the unavailability of glucose, producing glucotoxicity (9). By improving mitochondrial homeostasis and converting metabolic pathways, it is beneficial to support the homeostasis of the body environment and promote the change of cell phenotype, thereby improving the pathological mechanism of CMD (10, 11).

The endosymbiosis theory suggests that mitochondria are bacteria that are engulfed by primitive eukaryotes. This bacterium was symbiotic with eukaryotes and became mitochondria through evolution during the long symbiosis (12). Mitochondria are the two membrane-encapsulated organelles present in most cells and are the main sites for the aerobic respiration of cells. Mitochondria make up about one-third of the volume of adult cardiomyocytes and are the primary energy source for the coupling of excitatory contractions in the heart (13). Mitochondria play an important role in sustaining cardiology and physiology as metabolic and signal transduction centers, involved in many important biological processes such as OXPHOS production of ATP, fatty acid oxidation, calcium homeostasis, phospholipid synthesis, production, and maintenance of reactive oxygen species (ROS), and iron-sulfur cluster biosynthesis (14). In recent years, more and more evidence showed that mitochondria and CMD are closely related. Mitochondrial dysfunction can induce ROS production, activate DNA damage responses, lead to cardiac cell cycle arrest, and ultimately lead to fatal cardiomyopathy (15). Mitochondrial  $Ca^{2+}$  is the basic substance that activates the mitochondrial respiratory chain complex and ATP production and regulates the key mitochondrial dehydrogenase activity. When mitochondrial calcium homeostasis is unbalanced, mitochondrial bioenergetics are damaged, leading to the occurrence of diabetic heart disease (16).

Given the unique and highly dynamic structure of mitochondria in the heart and intimate links between mitochondria and CMD homeostasis in physiology and pathology. In this review, we focus on the mitochondrial fusion, fission, and mitophagy in CMD (with a focus on diabetic cardiomyopathy, myocardial infarction, heart failure, atherosclerosis, and obesity), and the potential drugs that target mitochondria to treat CMD.

## INTRODUCTION TO MITOCHONDRIA

### Mitochondrial Structure

Mitochondria can be divided into four functional regions from outside to inside: outer mitochondrial membrane (OMM), intermembrane space (IMS), inner mitochondrial membrane (IMM), and mitochondrial matrix (Figure 1).

The OMM is the outermost unit membrane of mitochondria with a thickness of about 6–7 nm. The OMM acts as a diffusion barrier and mediates mitochondrial signaling. The OMM is smooth and usually permeable, restricting diffusion only to molecules greater than ~5,000 Da. Molecules with a molecular weight greater than the above limit require a specific signal

sequence for recognition and active transport of mitochondria by translocase of the outer membrane (TOM) (17, 18). Importantly, the OMM is crucial to mitochondrial dynamics because it carries all the molecules involved in mitochondrial fusion and fission.

The IMM is a unit membrane located inside the OMM and surrounding the mitochondrial matrix. The mass ratio of protein to phospholipid in the IMM is about 0.7:0.3 and contains a large amount of cardiolipin. The IMM includes two main sub compartments: the inner boundary membrane (IBM) and mitochondrial cristae. The IBM is the part of the IMM that does not protrude into the matrix but runs parallel to the OMM. The cristae and IBM are connected by narrow tubular or slit structures called cristae junctions (CJS) (19).

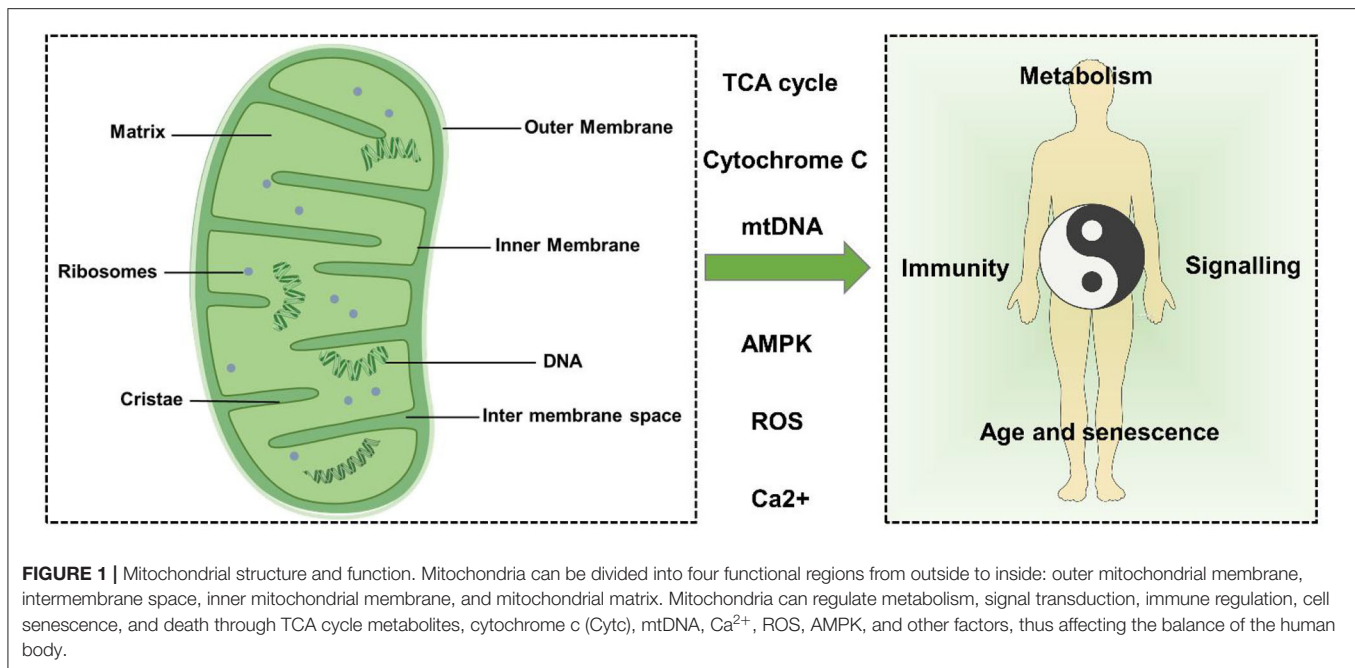
Cristae are characteristic folds of the inner membrane that penetrate the matrix. The IBM and cristae are functionally differentiated and have a distinct protein composition. Respiratory chain complexes and proteins involved in iron-sulfur cluster assembly are enriched in cristae membranes, while protein translocation and membrane fusion mechanisms are mainly present in IBM (20–23).

The mitochondrial lumen (named “matrix”) is the inner space enclosed by IMM, which contains many proteins such as enzymes involved in biochemical reactions such as tricarboxylic acid cycle, fatty acid oxidation, and amino acid degradation (24). In addition, the matrix also contains mitochondrial DNA (mtDNA), RNA, and mitochondrial ribosomes (25).

Human mtDNA is a double-stranded circular molecule of 16,569 base pairs with a molecular mass of 107 Da (26). Maintenance of mtDNA stability and integrity is critical for cellular energetics. mtDNA is particularly susceptible to damage, whether by respiration or exogenous contact, which can lead to damage to the base of the DNA. mtDNA is closely related to mitochondrial dynamics (25). When cells undergo apoptosis, mtDNA can be released into the cytoplasm, causing an inflammatory response (27).

### Mitochondrial Fusion and Fission

Homeostasis of mitochondrial dynamics is critical to maintaining cardiac structure and function. Mitochondria are highly dynamic organelles that maintain their shape, distribution, and size through fusion, fission, and mitophagy when cells undergo metabolic or environmental stress (Figure 2). The coordinated cycle of mitochondrial fission and fusion is known as mitochondrial dynamics (28, 29). Mitochondrial fusion allows organelles to share metabolites, proteins, and mtDNA, promoting complementarity between damaged mitochondria (30, 31). Fission increases the number of mitochondria to ensure that they are passed on to their offspring during mitosis. In addition, fission can signal to cells that mitochondria are damaged and need to be removed through mitosis to maintain a healthy mitochondrial network (32–34). The balance of fusion and fission affects the cardiac phenotype. An imbalance between fusion and fission is more detrimental than stopping both processes at the same time. Recently, a study showed that compared to MFN1/MFN2 cardiac knockout or DRP1 cardiac knockout mice, MFN1/MFN2/DRP1 cardiac triple knockout mice survived



longer and manifested a unique pathological form of cardiac hypertrophy (35).

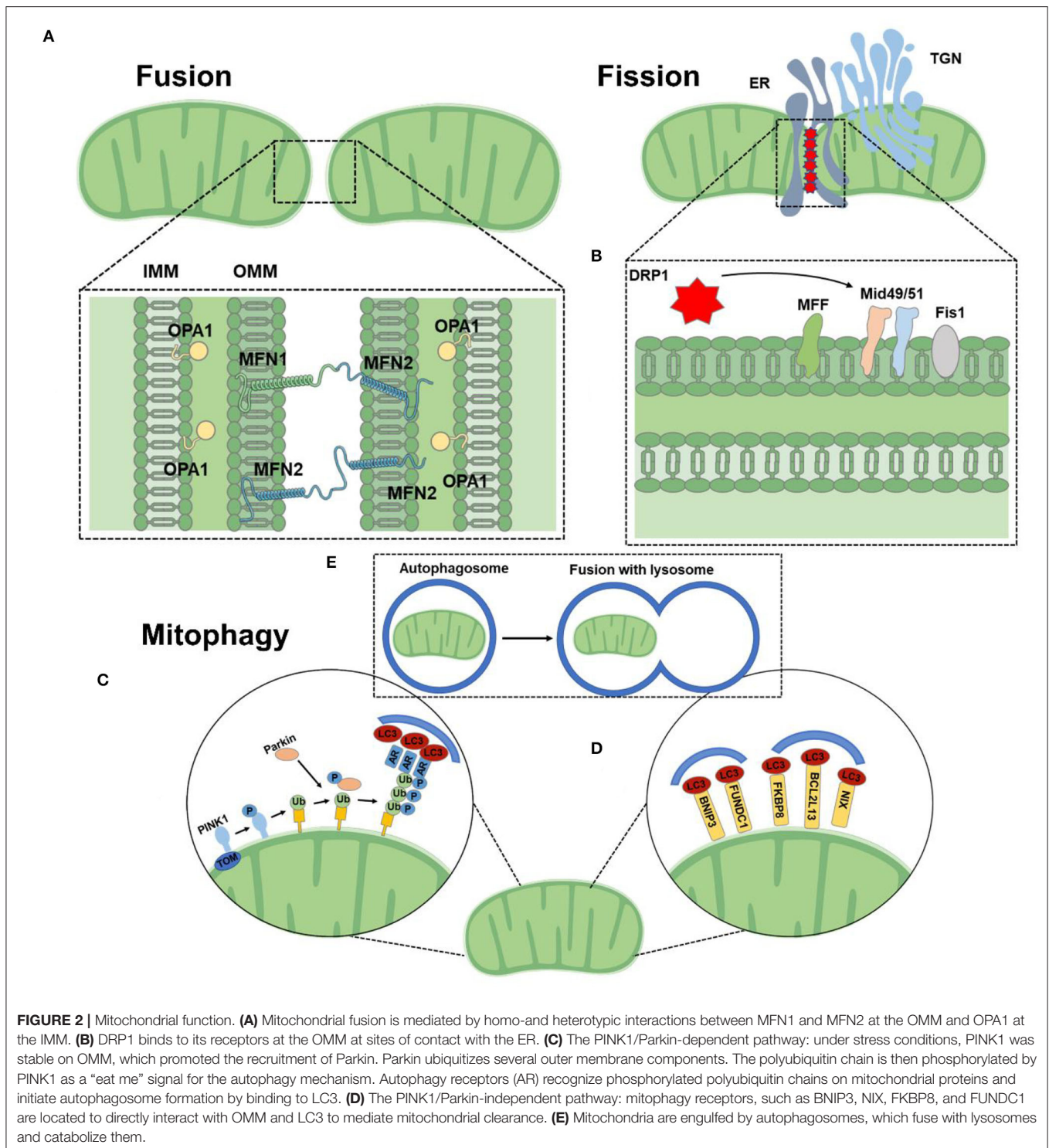
The core mechanism of mitochondrial dynamics is regulated by a group of GTPases related to the dynamin family. These proteins can oligomerize and change conformation, driving the remodeling, contraction, shearing, and fusion of mitochondrial membranes (36). Mitochondria are double-membrane organelles and complete fusion requires the merging of the outer and inner membranes. Mitochondrial fusion is a two-step mechanism. The OMM located GTPases mitofusin (MFN) 1 and 2 ensure the OMM fusion, and the IMM located optic atrophy protein 1 (OPA1) is responsible for IMM fusion (37, 38). OMM fusion is mediated by the MFN. MFN form both homo-oligomeric (MFN1-MFN1 or MFN2-MFN2) and hetero-oligomeric (MFN1-MFN2) complexes in trans between apposing mitochondria (39). Mitochondrial fusion is crucial for embryonic development, mice deficient in either MFN1 or MFN2 die in midgestation (40). OPA1 is a large GTPase tethered to the IMM facing the intermembrane space (39). OPA1 is dependent on MFN1 for fusion function, but not MFN2 (37).

Fission is mainly dominated by dynamin-related protein 1 (DRP1) which translocates from the cytoplasm to mitochondria and binds to its OMM partners [mitochondrial fission factor (MFF), mitochondrial dynamics protein of 49 kDa (MID49), MID51, and mitochondrial fission 1 protein (FIS1)] at sites of contact with the endoplasmic reticulum (ER) and actin (14, 41, 42). Following this binding, DRP1 oligomerizes and drives scission. Recently, DRP2 has been found to collaborate to drive mitochondrial fission (43). Loss of DRP1 results in highly elongated mitochondria and peroxisome (44). Kageyama et al. found that DRP1 and Parkin synergistically control the biogenesis and degradation of mitochondria. In the absence of mitochondrial fission mediated by DRP1, mitochondria

became defective in mitophagy. This mitophagy defect led to the accumulation of the mitophagy adaptor protein p62 and ubiquitinated proteins on mitochondria in a Parkin-independent manner, increases in cardiac defects, and loss of mitochondrial respiratory competence (45).

## Mitochondrial Biogenesis

Mitochondrial biogenesis is as important as other mitochondrial dynamics (46, 47), but it's almost neglected. Mitochondria especially injured ones actively regulate cell death, which is critical for maintaining cardiac homeostasis (48, 49). Mitochondrial biogenesis is the process by which cells increase mitochondrial mass. Mitochondrial biogenesis requires extensive coordination of both mitochondrial and nuclear genomes. Mitochondrial biogenesis is influenced by a variety of exogenous and endogenous factors such as exercise, caloric restriction, low temperature, oxidative stress, cell division, renewal, and differentiation (50). Mitochondrial biogenesis increases the copy number of mtDNA, the protein subunits of metabolic enzymes, and ultimately leads to greater metabolic capacity. In mammals, mitochondrial biogenesis is primarily regulated by the transcriptional coactivator PGC-1 $\alpha$  (46). A large body of evidence suggested that CMD is associated with mitochondrial biogenesis. By regulating the PGC-1 $\alpha$  signaling pathway, it is possible to treat diabetic cardiomyopathy, heart failure, obesity, and other diseases (51, 52). There is evidence that the genes involved in mitochondrial fusion (MFN1, OPA1) and fission (DRP1, Fis1) were altered expression in the hearts of PGC-1 $\alpha$ / $\beta$ -deficient mice. Significant mitochondrial structural dysregulation, including breakage and elongation, was observed in the hearts of PGC-1 $\alpha$ / $\beta$ - mice, associated with the development of fatal cardiomyopathy (53). In addition, PGC-1 $\alpha$ / $\beta$ - mice died shortly after birth with small hearts,

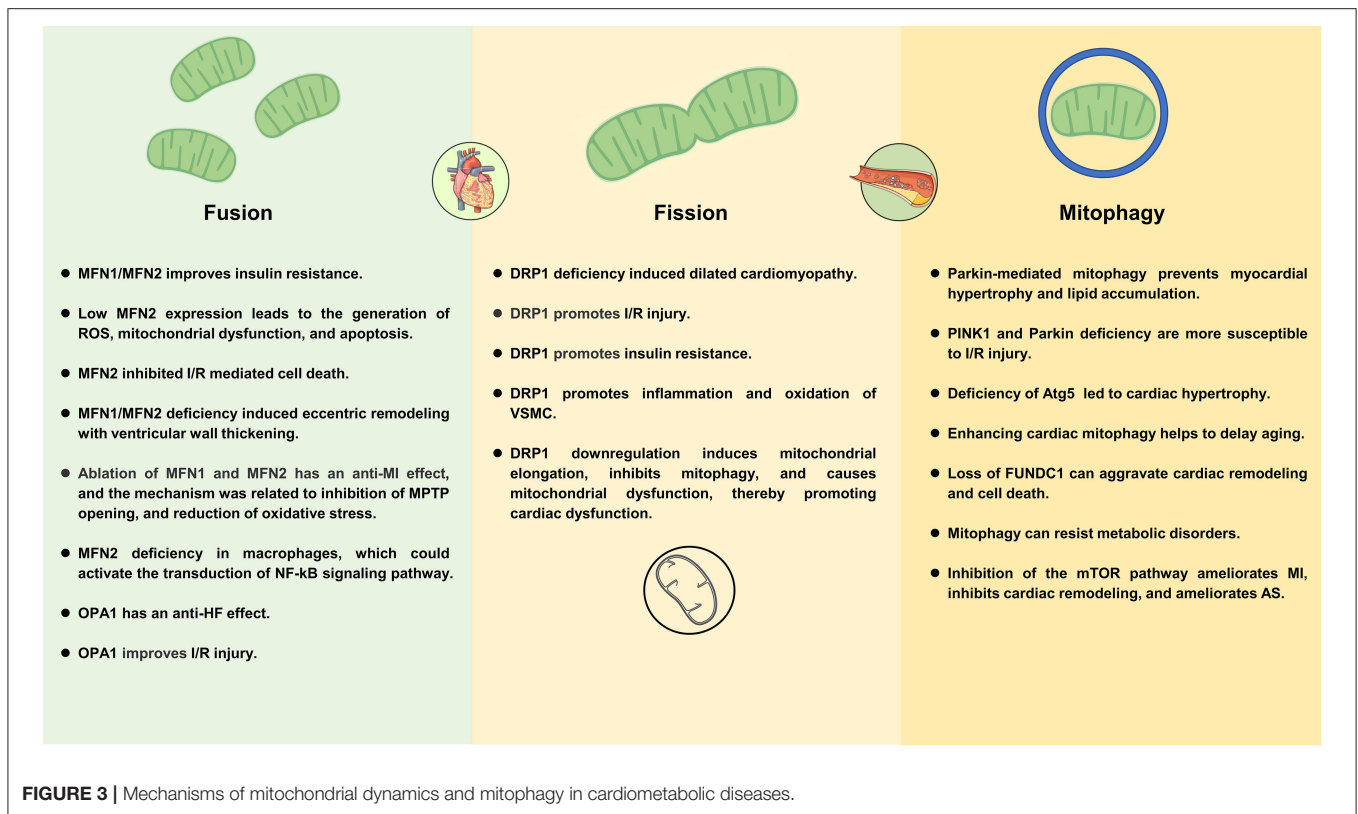


bradycardia, intermittent heart block, and a markedly reduced cardiac output (54).

## Mitophagy

Autophagy is an evolutionarily conserved mechanism that segregates superfluous, aging, or damaged cytoplasmic material

and deliver it to lysosomes for degradation (55). The main physiological role of autophagy may be to maintain cellular homeostasis in the context of reduced nutrient supply and other metabolic disturbances (56). Autophagy occurs in almost all types of cardiovascular cells, including myocytes, vascular smooth muscle cells (VSMCs), fibroblasts, macrophages, and



endothelial cells (57). It has been shown that a measured level of constructive autophagy is beneficial in restoring cardiac homeostasis in the CMD settings; whereas both autophagy impairment and excessive activation can lead to structural and functional dysfunction of the heart (58).

Mitophagy is the selective degradation of mitochondria by autophagy. Mitophagy can promote the turnover of mitochondria, maintain mitochondrial quality, and prevent the accumulation of dysfunctional mitochondria (59). Mitophagy is generally divided into two major functional groups based on the requirements for the kinase PINK1 and the Ub E3 ligase Parkin, often referred to as PINK1/Parkin-dependent (initiating by a loss of mitochondrial membrane potential) and PINK1/Parkin-independent mitophagy (not require loss of the mitochondrial membrane potential) (14). Mitochondrial autophagy is involved in metabolic activity, cell differentiation, apoptosis, and other physiological processes associated with major phenotypic changes, which is an important target for the treatment of CMD (60). Cardiac stress-induced mitophagy helps to remove damaged and dysfunctional mitochondria, thus preventing oxidative damage that could in turn initiate apoptosis and ultimately lead to heart failure (61). Autophagy related 7 (Atg7)-and Parkin-dependent mitophagy plays an essential role in the maintenance of mitochondrial function and protects the heart during the early development of diabetic cardiomyopathy (62). In addition, Atg5 deficiency-mediated mitophagy increases ROS production and NF- $\kappa$ B activity in macrophages, thereby aggravating cardiac inflammation and

injury (63). Thus, improving mitophagy may be a novel therapeutic strategy to ameliorate CMD.

## MITOCHONDRIAL DYNAMICS AND MITOPHAGY IN CARDIOMETABOLIC DISEASES

### Diabetic Cardiomyopathy

Diabetic cardiomyopathy (DC) refers to a cardiac disease that occurs in diabetic patients and cannot be explained by hypertensive heart disease, coronary atherosclerotic heart disease, or other cardiac lesions. Diabetic hearts utilize fatty acids as their main source of energy, producing high levels of oxidative stress that can lead to mitochondrial dysfunction (64). More and more evidence suggested that cardiovascular complications of diabetes are concentrated in the mitochondria, which are central to cardiomyocyte damage (65, 66).

Exposure to excess nutrients promotes the growth of the mitochondrial fission and reduces mitochondrial fusion, which is associated with uncoupled respiration (67). In agreement with this view, in hyperglycemic conditions, mitochondria can induce rapid division through DRP1 signaling, resulting in excessive production of ROS (68). Low MFN2 expression leads to the generation of ROS, mitochondrial dysfunction, and mitochondria-dependent apoptosis, which leading DC (69) (Figure 3). Along this line, a study showed that ablation of MFN2 leads to the development of impaired glucose tolerance, hyperinsulinemia, and insulin resistance (70). Montaigne et

al. found that the deterioration of endogenous myocardial contraction during the transition from obesity to diabetes may be related to the deterioration of myocardial mitochondrial function. Furthermore, they indicated diabetes mellitus was associated with cardiac mitochondrial network fragmentation and myocardial MFN1 content was inversely proportional to hemoglobin A1C (71).

A growing number of studies have demonstrated that balancing mitochondrial biogenesis and mitophagy is essential for maintaining cellular metabolism in the DC (72, 73). Mitophagy dysregulation exacerbate high-fat diet (HFD)-induced DC. Tong et al. (62) showed that Parkin-mediated mitophagy protects the heart against HFD-induced cardiac hypertrophy, and diastolic dysfunction and lipid accumulation, furthermore, Tat-Beclin 1 (inducer of autophagy) therapy alleviates cardiac dysfunction induced by HFD. Similarly, Wu et al. (74) found that deficiency of mitophagy receptor FUNDC1 impairs mitochondrial quality and aggravates dietary-induced obesity and metabolic syndrome.

Inflammation plays a key role in the pathogenesis of diabetes mellitus, and inflammatory injury is usually associated with pancreatic  $\beta$  cell dysfunction (75). Mitochondria are negatively affected by pancreatic  $\beta$  cell inflammatory signals, which can lead to impaired mitochondrial dynamics (76, 77). Therefore, strategies that block inflammation and/or protect mitochondrial function are potential treatments for diabetes. Tanajak et al. (78) showed that impaired  $\beta$  cell mitophagy exacerbates hyperglycemia and mitochondrial fragmentation *in vivo* following inflammatory stimuli, and overexpression of CLEC16A (mitophagy regulator whose expression in islets is protective against T1D) can mediate mitophagy and protect  $\beta$  cells from cytokine-mediated death.

Diabetic cardiomyopathy is closely related to diet, exercise, and metabolic changes. Caloric restriction and exercise may be a strategy to treat DC. A study found that caloric restriction is effective in improving metabolic regulation, and attenuating cardiac mitochondrial dysfunction in obese-insulin-resistant rats (79). Furthermore, clinical trials found that exercise improved fat oxidation and insulin sensitivity in obese insulin-resistant adults, and the mechanism was related to the decrease of DRP1 and the increase of MFN1 and MFN2 (80).

Based on the above evidence, we can suggest that inhibition of mitochondrial fission, promotion of mitochondrial fusion, and mitophagy are potential strategies for the treatment of DC. It is worth noting that effective exercise and diet are also a measure to modulate mitochondria.

## Myocardial Infarction

Myocardial infarction (MI) is an irreversible disease of the myocardium, which is based on ischemic hypoxic necrosis of cardiac myocytes (81). Ischemia is generally accompanied by overproduction of ROS, mitochondrial dysfunction, the translocation of Cyt c from the mitochondria to the cytoplasm, the opening of the mitochondrial permeability transition pore (mPTP) and inflammatory cascade (82, 83). In this setting, effective mitochondrial homeostasis is important.

Studies demonstrated that ischemia induces mitochondrial fission, inhibition of DRP1 protected cardiomyocytes against ischemia/reperfusion (I/R), inhibited mPTP opening, and reduced the size of MI (84–86). Although it is widely believed that the fused mitochondria function better, however, Ikeda et al. (87) suggested DRP1 plays an essential role in mediating mitophagy in cardiomyocytes, they found that DRP1 downregulation induces mitochondrial elongation, inhibits mitophagy, and causes mitochondrial dysfunction, thereby promoting cardiac dysfunction and increased susceptibility to I/R.

Mdivi-1 pharmacologically inhibits mitochondrial fission by blocking the binding of DRP1 to its receptor. Mdivi-1 given before ischemia exerts cardioprotective effects by reducing the incidence of arrhythmias, decreasing infarct size, improving cardiac mitochondrial function and fragmentation, and reducing cardiac apoptosis (88). Interestingly, another study found negative results, they found that treatment with Mdivi-1 at the start of reperfusion did not reduce MI size or preserve left ventricular function in pig MI model (89). Mitochondrial fission is dependent on ER-mitochondria contact. BAP31, an ER protein, regulates intracellular calcium homeostasis and ER stress (90). Cheng et al. (91) found that silencing Fis1/ BAP31 reduced mitochondrial fission and inhibited JNK activation, which led to a reduction in ROS and promoted cardiomyocyte survival.

Mitochondrial fusion protects stressed cells through two separate mechanisms. First, fusion counteracts the effects of excess mitochondrial fission, thereby limiting fission-induced mitochondrial apoptosis. Second, fusion enhances the timely detection of damaged parts of mitochondria and balances mitochondrial proteins, lipids, metabolites, and mitochondrial DNA (92, 93). Mitochondrial fusion has a protective effect in physiological conditions, but the role of mitochondrial fusion-related proteins in I/R injury remains a hot topic of debate. A study found that MFN2 overexpression significantly inhibited I/R mediated cell death by promoting mitochondrial fusion, regulating glucose metabolism, and oxidative stress (94). Interestingly, another study reported the opposite observation, they showed that ablation of MFN1 and MFN2 genes had a protective effect on myocardial infarction, and the mechanism was related to inhibition of mPTP opening, reduction of oxidative stress, and attenuating mitochondrial  $\text{Ca}^{2+}$  overload (95). Analogously, MFN1 KO heart mitochondria displayed a normal respiratory repertoire and were not affected by mitochondrial depolarization and their viability was enhanced when attacked by ROS in the form of hydrogen peroxide (96).

Optic atrophy protein 1-related mitochondrial fusion and mitophagy are vital to sustaining mitochondrial homeostasis under stress conditions (97, 98). Le et al. (99) showed that increase in cardiac I/R injury, impairment of dynamic mitochondrial  $\text{Ca}^{2+}$  uptake, and increased incidence of arrhythmias in Opa1 $^{+/-}$  mouse models. Similarly, Zhang et al. (100) demonstrated that manipulation of the AMPK-OPA1-mitochondrial fusion/mitophagy axis *via* melatonin can block cardiomyocyte caspase-9-involved mitochondrial apoptosis, which attenuates I/R injury.

Mitochondrial damage and ROS produced by mitochondrial oxidative stress can act as substrates to induce mitophagy (101).

During ischemia, upregulation of mitophagy is considered beneficial (102, 103). For instance, a study showed that Parkin ablation caused a decrease in a severe decrease in mitochondrial respiration, mitochondrial uncoupling, and increased susceptibility to the opening of the permeability transition pore (104). Further, Parkin (-/-) mice were much more sensitive to MI, and Parkin (-/-) myocytes had reduced mitophagy and accumulated swollen, dysfunctional mitochondria after the infarction (105). PINK1 protein is highly expressed in the myocardium. PINK1-/- mice had larger MI and were more susceptible to I/R injury, which is associated with impaired mitochondrial function (106). Therefore, induction of autophagy through activation of the Pink1/Parkin pathway could exert a protective effect against MI (107).

The mTOR pathway is a well-known upstream node that acts to inhibit autophagy. It was found that rapamycin improves MI and inhibits cardiac remodeling by inhibiting mTORC1 and ER stress pathways, preventing angiotensin II-induced apoptosis in H9c2 cells, and promoting autophagy (108).

FUNDC1 was first reported in 2012 as a new hypoxia-induced mitophagy receptor (109). Zhou et al. showed that the pathogenesis of cardiac I/R injury is related to the disruption of mitochondrial homeostasis by CK2 $\alpha$  through the inhibition of FUNDC1-related mitophagy. They indicated that CK2 $\alpha$  inactivation of FUNDC1, thus effectively inhibiting mitophagy. Defective mitophagy fails to clear damaged mitochondria induced by I/R injury, leading to mitochondrial genome collapse, electron transport chain complex inhibition, mitochondrial biogenesis arrest, mPTP turn-on, and ultimately mitochondrial apoptosis (103). Zhang et al. (110) demonstrated that FUNDC1-mediated mitophagy regulates both mitochondrial quality and quantity *in vivo* under I/R conditions, and plays a role in mitochondrial quality control and functional integrity in platelet activation. Similarly, Mao et al. (111) found that the enhancement of the p-AMPK/FUNDC1 axis can alleviate the hypoxia and reoxygenation induced apoptosis of H9c2 cells and alleviate injury in the I/R mouse model. In addition, Zhang et al. showed that mitophagy in MI is mediated by Rab9-associated autophagosomes, rather than relying on the Atg7 conjugating system and LC3. And they indicated that the Rab9-associated autophagosome pathway is mediated by the synergistic action of Ulk1, RAB9, Rip1, and DRP1 (112).

In conclusion, the role of proteins regulating mitochondrial fusion and fission on MI is divergent, and there seems to be a bidirectional regulation, and the beneficial and detrimental effects still need further validation. Notably, promoting mitophagy is beneficial for MI.

## Cardiac Hypertrophy and Heart Failure

Heart failure (HF) is a syndrome in which the pumping function of the heart is impaired for various reasons and the output of the heart is unable to meet the basic metabolic demands of the tissues of the body (113). It is known that prolonged or high levels of cardiac stress can cause mitochondrial damage and dysfunction in cardiac myocytes. Throughout, mitochondrial metabolism is essential for adequate myocardial pump function, as cardiomyocytes in this state require large amounts of energy

to maintain contractile performance, Ca<sup>2+</sup> homeostasis, and ion transport (114). Based on this by improving mitochondrial function, clearance has the potential to be a strategy for the treatment of HF.

Mitochondrial fusion and fission are associated with myocardial hypertrophy and the development of HF. A study showed that MFN1/MFN2 deficiency-induced eccentric ventricular remodeling with ventricular wall thickening and DRP1 deficiency-induced dilated cardiomyopathy. Further, this study showed that inhibition of the mitochondrial permeability transition alleviates cardiomyocyte death and mitochondrial loss induced by cardiac DRP1 deficiency (115). Another study showed that MFN2 was downregulated in a rat model of myocardial hypertrophy, depending on the etiology and time course of myocardial hypertrophy (116). Therefore, myocardial hypertrophy can be inhibited by up-regulation of MFN2 expression (117, 118). Along similar lines, a clinical study found reduced mitochondrial content, oxidative capacity, and MFN2 expression in skeletal muscle of patients with HF, which is related to the accumulation of dysfunctional organelles and decreased OXPHOS ability in the mitochondrial network due to the low expression of MFN2 (119).

Optic atrophy protein 1 plays an important role in preventing the release of Cyt c from the cristae (120). Apoptotic cell death *via* reduction of OPA1 and mitochondrial fusion may contribute to HF progression. Chen et al. (121) found that OPA1 reduction in HF models leads to increased mitochondrial fragmentation, increased apoptosis, increased sensitivity to ischemia. In addition, OPA1 also has a significant effect on energy utilization, OPA1 can increase the utilization of cardiac fatty acids, thereby reducing ROS production and maintaining mitochondrial morphology during HF (122). Two mitochondrial proteases (OMA1 and the AAA protease YME1L) cleave OPA1 from long (L-OPA1) to short (S-OPA1) forms. L-OPA1 is required for mitochondrial fusion, but S-OPA1 is not (123). Wai et al. found that deletion of YME1L activates OMA1, accelerates the proteolytic processing of OPA1, and causes mitochondrial fission of the heart, which changes the metabolism of the heart and further causes dilated cardiomyopathy and HF. However, if deletion of OMA1, OPA1 processing can be avoided and cardiac function and mitochondrial morphology can be restored (124).

Mitophagy in the heart is a homeostatic mechanism for maintaining cardiomyocyte size and global cardiac structure and function, and the upregulation of mitophagy in failing hearts is an adaptive response for protecting cells from hemodynamic stress. For instance, in adult mice, deficiency of Atg5, a protein required for autophagy, led to cardiac hypertrophy, a disorganized sarcomere structure, left ventricular dilatation, and contractile dysfunction, accompanied by increased levels of ubiquitination (125, 126).

Mitophagy is transiently activated and then downregulated in cardiac tissues during the early phase of HF, restoration of mitophagy attenuates dysfunction in the heart during pressure overload. In the presence of pressure overload, treatment with Tat-Beclin 1 can attenuate the progression of HF (127). For instance, a study demonstrated that AMPK $\alpha$ 2 protects against the development of HF by enhancing mitophagy *via* PINK1

Phosphorylation (128). Similarly, Beak et al. (129) demonstrated that deficiency of the nuclear receptor ROR $\alpha$  in the mouse exacerbates angiotensin II-induced cardiac hypertrophy and compromises cardiomyocyte mitochondrial function, which is associated with reduced mitophagy. Along similar lines, Nah et al. (130) showed that Ulk1-cKO mice had suffered from impaired mitochondrial quality control and more severe cardiac dysfunction, hypertrophy, and myocardial fibrosis, the mechanism of which is related to Ulk1-dependent alternative mitophagy. Lysosomal-associated membrane protein-2 (LAMP-2) is a highly glycosylated protein that is an essential component of the lysosomal membrane. This protein is critical for the fusion of lysosomes and autophagosomes, leading to the degradation of their contents, and LAMP-2 deficiency can lead to premature death in mice and inhibit myocardial systolic function (131). Therefore, overexpression of either autophagy gene and thus increased autophagy levels could improve CMD.

Current evidence overwhelmingly suggests that caloric restriction and fasting induce mitophagy and mitophagy-related markers (132). Fasting or caloric restriction can as a new and practical treatment for CMD. For instance, a study found that caloric restriction can enhance cardiac autophagy and delay cardiac aging by inhibiting the mTOR pathway. Meanwhile, caloric restriction can also reduce the accumulation of senescence-associated  $\beta$ -galactosidase and lipofuscin and reduced myocyte apoptosis (133). Weir et al. (134) found that caloric restriction increased fatty acid oxidation by maintaining mitochondrial network homeostasis and functional coordination with the peroxisome to promote longevity in *C. elegans*. Therefore, it is practical to use non-pharmacological therapies like exercise and diet to treat metabolic diseases.

## Atherosclerosis

Atherosclerosis (AS) is a chronic inflammatory disease of large and medium-sized arteries that causes ischemic heart disease, strokes, and peripheral vascular disease (135). The pathogenesis of AS begins with the accumulation and retention of apolipoprotein B-containing lipoproteins in the vascular sub endothelium which initiates the recruitment and entrance of inflammatory monocytes into the vessel wall. There is increasing evidence that mitochondria play a key role in the development of inflammatory responses and the maintenance of chronic inflammation (136).

Recently, studies have found that age-related mitochondrial dysfunction promotes AS, which is associated with elevated inflammatory factors (137). Mitochondrial damage results in the release of mitochondrial components (mtDNA, cardiolipin), which is a potent DAMP recognized by the immune cells that can trigger the inflammatory response. Exposure to these cellular debris allows the recruitment of adaptor molecules/receptors that triggers an innate immune response (138). It is worth noting that a study has found that mtDNA damage can promote AS independently of ROS through effects on smooth muscle cells and monocytes and correlates with higher-risk plaques in humans (139). Khodzhaeva et al. (140) suggested pro-inflammatory effects of MFN2 deficiency in human macrophages, which could enhance the expression of IL- $\beta$

and TNF- $\alpha$ , and activate the transduction of the NF- $\kappa$ B signaling pathway. Thus, by correcting mitochondrial dysfunction, it may be possible to reduce the inflammatory state in AS.

It is well-known that mitochondrial generation of ROS is closely related to the occurrence and development of AS. Under normal conditions, mitochondrial antioxidant and repair systems counteract the harmful effects of excess ROS. In addition, mitochondria can counteract the effects of ROS by regulating fusion and fission (141). When mitochondrial dysfunction occurs, ROS is overproduced, leading to the oxidation of lipids, nucleic acids, and proteins, which eventually leads to severe cell damage. Excessive ROS production can lead to endothelial dysfunction, inflammation of blood vessels, and the accumulation of oxLDL in arterial walls (142). By inhibiting DRP-mediated mitochondrial fission, oxidative stress is reduced, endothelial dysfunction is improved, and inflammation is suppressed, thereby reducing the progression of AS (143).

Vascular smooth muscle cells are the main components of the vessel wall and plaques. Abnormal proliferation of VSMCs promotes plaque formation, but VSMCs are beneficial in advanced plaques (preventing rupture of fibrous caps) (144). The pathogenesis of AS is closely related to VSMCs mitochondrial dysfunction (145). DRP1 and mitochondrial fission could promote inflammation and oxidation of VSMCs, and Mdivi1 can significantly inhibit the inflammatory response and oxygen consumption in VSMCs (146). The relationship between MFN2 and AS has been studied for a long time. In 2004, Chen et al. (147) found that MFN2 was significantly reduced in hyperproliferative VSMCs of AS plaques or balloon lesions, and that increasing MFN2 prevented neointimal VSMCs proliferation after balloon injury and prevented restenosis caused by balloon injury. Along this line, another study found that MFN2 constituted a primary determinant of VSMCs apoptosis, and the mechanism is associated with MFN2 mediated Akt signaling pathway as well as a mitochondrial apoptotic pathway, specifically reflected in increasing Bax/Bcl-2 ratio, promoting Cyt c release, activating Caspase-9 and Caspase-3 activation (148).

Lack of autophagy genes may severely disrupt intravascular homeostasis (149). Progressive AS has features of dysfunctional autophagy, which is related to plaque macrophages (150). Recently, a study suggested that a high protein diet increases AS progression by activating macrophage mTOR to inhibit mitophagy (151). Macrophages lacking the key autophagic protein ATG5 enhance atherosclerotic plaque formation. Inclusion bodies rich in P62 and polyubiquitinated proteins in macrophages prevent AS (152). Analogously, VSMC-specific ATG7 knockout mice were found to have increased total collagen deposition, nuclear hypertrophy, up-regulation of CDKN2A, reduced phosphorylation of RB, and enhanced GLB1 activity, which is closely related to the promotion of neointima formation and atherosclerosis formation (153). Endothelial autophagy is necessary to maintain vascular lipid homeostasis. By both confocal and electron microscopy, Torisu et al. found that autophagy in endothelial cells may represent an important mechanism for regulating excess exogenous lipids. Their study showed that excess LDL appeared to be engulfed within autophagic structures and that temporary knockdown



of ATG7 resulted in increased intracellular levels of I-LDL and oxLDL (154).

Overall, the above evidence demonstrates that mitochondrial dynamic proteins play an important role in inflammation and oxidative stress in AS. Effective mitophagy can play an anti-AS role.

## Obesity

Obesity is a major risk factor for several other diseases, such as diabetes, cardiovascular disease, and metabolic disease. High levels of fat mass accelerate the development of dyslipidemia, blood pressure, inflammation, and glucose/insulin resistance (155, 156). Long-term, high-concentration substrate supplies deplete NAD<sup>+</sup>, leading to the accumulation of metabolic intermediates, such as acetyl-CoA, which promotes the production of ROS and makes mitochondria vulnerable to damage. Glucose and lipid metabolism are largely dependent on mitochondrial metabolism, and mitochondrial function is most affected in primary tissues, such as bone and heart muscle, liver, and adipose tissue.

It has been found that hyperglycemia and high free fatty acids can modulate insulin sensitivity and lead to mitochondrial dysfunction (157, 158). Several mitochondrial genes critical to mitochondrial function and OXPHOS were down-regulated in obese, HFD fed, insulin-resistant mice (159, 160). Mitochondrial dynamics are influenced by metabolic demands, changes, and alterations. Specific MFN2 mutations lead to tissue-selective mitochondrial dysfunction and increase adipocyte proliferation and survival (161). MFN2 in fat is important for regulating metabolism and has been found to be lower in adipose tissue of mice and obese humans on a HFD (162). Bach et al. showed that weight loss upregulated MFN2 expression in skeletal muscle and type 2 diabetes downregulated MFN2 expression in skeletal muscle in the obese population. Further, MFN2 expression in skeletal muscle was positively associated with insulin sensitivity, but inversely associated with BMI, TNF- $\alpha$  and IL-6 (163). Along this line, Pich et al. (164) found that decreasing MFN2 inhibited the oxidation of pyruvate, glucose and fatty acids and decreased the mitochondrial membrane potential, while increasing MFN2 increased glucose oxidation and mitochondrial membrane potential.

Excessive lipid uptake in the heart affects dynamin and induces mitochondrial fission and dysfunction (165). Lipid overload increased DRP1 level and activity in mouse hearts, and overactivated DRP1 increases mitochondrial fission and interacts with VDAC1 to lead to myocardial cell death, and targeting DRP1 helps reduce cardiac hypertrophy and dysfunction caused by lipid overload. Similarly, it has been reported that blocking DRP-1 on mitochondrial fission can improve white adipose tissue abnormalities in obesity and diabetes in ob/ob mice by short-term leptin and Mdivi-1 treatment (166).

Mitophagy plays an important role in maintaining cardiac function in obesity (167). Defective mitophagy is causally linked to obesity complications. Loss of FUNDC1 aggravates HFD-induced cardiac remodeling, functional and mitochondrial abnormalities, cell death, and Ca<sup>2+</sup> overload (168). In contrast, a study found that mice lacking muscle FUNDC1 were found

to have a protective effect against HFD-induced obesity, with improved insulin sensitivity and glucose tolerance despite reduced muscle mitochondrial energy. The mechanism may be related to FUNDC1 deficiency leading to muscle degeneration and up-regulation of FGF21 expression, thus promoting thermogenic remodeling of adipose tissue (169).

In 2021, Cho et al. reported that serine/threonine-protein kinase 3 (STK3) and STK4 are key physiological suppressors of mitochondrial capacity in brown, beige, and white adipose tissues. They found that STK3 and STK4 increase adipocyte mitophagy in part by regulating the phosphorylation and dimerization status of the mitophagy receptor BNIP3, which increased resistance to metabolic disorders induced by a HFD (170). Increased fatty acid oxidation has been implicated in the development of cardiomyopathy induced by obesity/diabetes. However, Shao D et al. (171) found that increased fatty acid oxidation by regulating Parkin-mediated mitophagy can prevent HFD-induced cardiomyopathy.

## THERAPIES FOR CARDIOMETABOLIC DISEASES

### Mitochondrial Targeted Drug Therapy

Targeting mitochondrial dynamics and mitophagy is increasingly becoming a research direction in the treatment of CMD. DRP1-mediated mitochondrial fission is an attractive drug target. By inhibiting DRP1, excessive mitochondrial fission can be reduced, mitochondrial fusion activity can be enhanced, and loss of mitochondrial membrane potential and release of Cyt c can be prevented, thereby preventing cell death (172). Deng et al. (173) found that treatment with Mdivi-1 (20 mg/kg/day) significantly reduced angiotensin II-induced hypertension, arterial remodeling, and cardiac hypertrophy by a mechanism that may be related to preventing phenotypic transformation of VSMCs. Aishwarya et al. (174) found several novel pleiotropy effects of Mdivi-1 in cardiomyocytes, including decreased expression of OXPHOS complex protein and superoxide production, impaired macroautophagy flux, and altered mitochondrial serine protease expression during L-OPA1 proteolysis. Mdivi-1 has been shown to have a cellular protective effect on I/R injury (84, 175). Interestingly, a study found that Mdivi-1 attenuates oxidative stress and exerts vascular protection in I/R injury through a mechanism unrelated to DRP1 activity, possibly due to elevated levels of antioxidant enzymes, SOD1, and catalase, as well as Nrf2 expression (176). Insulin resistance co-occurs with mitochondrial dysfunction in skeletal muscle, and Mdivi-1 can reduce insulin resistance by enhancing mitochondrial function (177, 178).

Melatonin belongs to an indole heterocyclic compound with numerous receptor-mediated and receptor-independent actions. It has been found that melatonin can reduce the size of MI, inhibit myocardial cell death, and maintain myocardial function by promoting OPA1-related mitochondrial fusion (179). Further, it was found that melatonin can promote mitochondrial fusion/mitophagy through the AMPK/OPA1 pathway and reduce calcium deposition in VSMCs (180). Zhou et al. (181) showed

that melatonin protects cardiac microcirculation from I/R injury by inhibiting mitophagy in PINK1/Parkin pathway and weakening mitochondrial fission. In addition, it was found that melatonin can also protect against diabetic heart disease by inhibiting DRP1 expression through SIRT1-PGC-1 $\alpha$  (182). In 2021, Bai et al. (183) demonstrated that melatonin regulates mitophagy and mitochondrial dynamics in a SIRT3-dependent manner to ameliorate hypoxia/reoxygenation damage.

## Mitochondrial Targeted Natural Medicines Therapy

Natural medicines are characterized by multi-target and multi-pathway synergies, and their potential in mitochondrial dynamics is gradually recognized (184). *Panax ginseng* is a traditional herbal medicine that can improve human immunity and ginsenosides are the major bioactive constituents in ginseng root. Yang et al. showed that ginsenoside Rg5 can attenuate I/R injury in cardiomyocytes by inhibiting the opening of mPTP and increasing ATP production. In addition, ginsenoside Rg5 can also inhibit the activation of DRP1 through the AKT pathway, thereby protecting cells from I/R injury (185). Recently, Jiang et al. (186) discovered through proteomics that the mechanism by which ginsenosides Rb1 alleviate I/R injury is that ginsenoside Rb1 reduces the activity of NADH dehydrogenase, making mitochondrial complex I in a deactivated form upon reperfusion, thereby reducing the burst of ROS. Berberine is the extract of *Coptis chinensis* and is widely used. A study showed that berberine improved myocardial hypertrophy and apoptosis induced by transverse aortic contraction in mice by upregulating PINK1/Parkin-mediated mitophagy (187). Gypenosides is an extraction product of *Gynostemma pentaphyllum* (Thunb) Makino. A study found that gypenosides improve AS levels in ApoE $^{-/-}$  mice through the regulation of mitochondrial fission proteins and fusion proteins *via* the PI3K/Akt/Bad pathway (188). Quercetin, widely distributed in the plant kingdom, has a variety of bioactive flavonol compounds. Quercetin can improve vascular calcification (189), insulin sensitivity (190), and I/R injury (191) by inhibiting DRP1-mediated mitochondrial fission. In addition, a study found that quercetin inhibited excessive mitochondrial fission by activating mitophagy *via* SIRT1/TMBIM6, inhibits endoplasmic reticulum stress, and improves human cardiomyocyte activity (192). Related studies on DC also found that quercetin can regulate mitochondrial fusion and fission mechanisms by regulating SIRT5 and IDH2-related succinylation, thereby protecting the pathological damage of myocardial cells stimulated by high glucose (193).

In addition to the active ingredients of traditional Chinese medicine, the compound preparation of traditional Chinese medicine can also have a certain regulatory effect on the interaction mechanism of mitochondrial dynamics and mitophagy. Tongyang Huoxue Recipe, a traditional Chinese medicine compound, can protect sinoatrial node cells under stress through PINK1/parkin-mediated mitophagy, and can also inhibit DRP1-mediated mitochondrial fusion/fission dysfunction, increase mitochondrial membrane potential,

maintain calcium homeostasis, and inhibits oxidative stress damage (194). Zishen Huoxue Decoction is an effective compound for the treatment of patients with coronary heart disease. Liu et al. (195) found that Zishen Huoxue Decoction has the effects of activating mTORC1 signaling pathway, inhibiting the overexpression of 4E-BP1, inhibiting fatty acid oxidation, protecting the respiratory function of mitochondria, and thus protecting myocardial cells from injury. It is worth noting that natural medicines are subject to many uncertainties and require extensive clinical and basic research for validation.

## CONCLUSIONS

Mitochondrial dynamics and mitophagy play an important role in the physiology and pathology of CMD. CMD is closely related to energy metabolism, and effective mitochondrial homeostasis is an important defense mechanism for the heart to deal with various stress and injury. In CMD, mitochondrial dynamics and mitophagy play an important role in improving insulin resistance, improving metabolite utilization, inhibiting inflammatory response, reducing apoptosis, protecting endothelial cells, and reducing ventricular remodeling.

Current evidence overwhelmingly suggested that mitophagy has a positive effect on CMD. Mitochondrial fusion and fission, on the other hand, need to be viewed dialectically. Many studies have shown that MFN1 and MFN2 are beneficial and DRP1 is detrimental, but there are also studies that take the opposite attitude. Therefore, more *in vivo* and *in vitro* studies are still needed for further validation. In terms of targeted mitochondrial therapy, exercise and diet therapy are promising roles without side effects and should be recommended. In addition, natural medicines are a great treasure with great potential from which we can discover better targeted mitochondrial drugs. With the continuous development of modern science and technology and the deepening of research on mitochondrial dynamics and mitophagy drugs, mitochondrial-targeted therapy will provide more ideas for the treatment of CMD, and with the development of new microscopy, biotechnology, and computer technology, the mystery of mitochondria will be gradually explored by human beings.

## AUTHOR CONTRIBUTIONS

JL and KY designed the manuscript. JL and JD edited the manuscript. SZ, SX, and QW revised the manuscript. All authors contributed to manuscript revision, read, and approved the submitted version.

## FUNDING

This study was funded by grants from the General Program of National Natural Science Foundation of China (81873173) and the Science and Technology Innovation Project of China Academy of Chinese Medical Sciences (CI2021A00906).

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