Check for updates

OPEN ACCESS

EDITED BY Feilong Zhang, Nanyang Technological University, Singapore

REVIEWED BY Benhui Hu, Nanjing Medical University, China Jing Chen, Shandong University, China

*CORRESPONDENCE Zhangping Li, ≅ wzlizhangping@126.com Jian Xiao, ≅ xfxj2000@126.com Chenyao Nie, ≅ niechenyao@wmu.edu.cn

RECEIVED 01 November 2023 ACCEPTED 13 November 2023 PUBLISHED 22 November 2023

CITATION

Li J, Li Z, Xiao J and Nie C (2023), Conformable electrochemical devices for closed-loop wound management. *Front. Bioeng. Biotechnol.* 11:1331567. doi: 10.3389/fbioe.2023.1331567

COPYRIGHT

© 2023 Li, Li, Xiao and Nie. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Conformable electrochemical devices for closed-loop wound management

Jie Li¹, Zhangping Li^{1,2}*, Jian Xiao¹* and Chenyao Nie¹*

¹School of Pharmaceutical Sciences, Cixi Biomedical Research Institute, Wenzhou Medical University, Wenzhou, Zhejiang, China, ²The Quzhou Affiliated Hospital of Wenzhou Medical University, Quzhou People's Hospital, Quzhou, Zhejiang, China

Chronic wounds arising from accidents, surgeries, or diseases impose a significant clinical and economic burden, underscoring the need for effective solutions to prevent severe complications. Recent advancements in materials science and electrochemical technology have facilitated the development of conformable electrochemical platforms for detection and management, incorporating monitoring, diagnosis, and treatment. Nevertheless, current wound detection and therapy systems face challenges related to the stability and specificity of sensor monitoring, as well as the need for on-site and comprehensive evaluation criteria to offer timely treatment guidance and follow-up care. This review provides a comprehensive overview of the closed-loop management system, emphasizing wound biomarker detection, wound assessment, and on-demand treatment, ultimately culminating in an integrated wound management approach by conformable electrochemical devices. Additionally, we explore the challenges, opportunities, and future prospects of soft and stretchable electrochemical biosensors, with the aim of enhancing the efficiency and timeliness of wound management.

KEYWORDS

wound management, conformable device, closed-loop, electrochemical biosensing, ondemand treatment

1 Introduction

Wounds resulting from accidents, surgeries, or chronic conditions pose a significant clinical challenge and impose a substantial economic burden on global healthcare systems. Effective wound management is not only about alleviating physical discomfort but also stands as a crucial healthcare necessity. Neglected or inadequately treated wounds have the potential to escalate into severe medical issues, including infections, sepsis, amputations, and, at times, even fatalities (Sen, 2021). According to a survey conducted by the World Health Organization, the issue of wounds has affected more than 305 million people around the world. It is estimated that by 2024, global expenditures related to wound care will reach \$80 billion (Yao et al., 2022). Traditional wound treatment, often involving dressing removal or sample examination, is time-consuming, expensive, and uncomfortable. A patient-friendly tool is needed to monitor disease progression, guide treatment, and reduce costs (Romanelli et al., 2013).

Recent advancements in materials fabrication, sensing techniques, and device integration strategies have opened up numerous opportunities in various healthcare management systems (Mathew et al., 2020; Sun et al., 2021). In particular, conformable electrochemical devices stand out due to their flexibility, breathability, biocompatibility, and

their capability to continuously analyze biomarkers in personalized wound management (Mirvakili and Langer, 2021; Tang et al., 2021). Electrochemical biosensors are cost-effective, portable, and seamlessly integration into electronic acquisition modules, making them promising for comprehensive evaluation of the wound progress and treatment in the intricate wound microenvironment. Furthermore, to enhance patient outcomes and achieve greater efficiency and precision in wound management, electrochemical devices are undergoing continuous evolution. They are shifting from single-function devices that primarily focus on biomarker sensing or treatment to more interconnected wound management systems (Mathew et al., 2020). This evolution is leading to the development of interconnected wound management systems, which seamlessly coordinate various facets of the wound healing process. These systems cover everything from biomarker detection, wound evaluation, and on-demand treatment, ultimately culminating in the creation of a closed-loop wound management system that seamlessly integrates all these modules. This logical progression in technology promises a more holistic and efficient approach to wound care, benefiting both patients and healthcare providers (Mathew et al., 2020; Mirvakili and Langer, 2021; Tang et al., 2021; Yang et al., 2023).

In this review, we provide an overview of recent advancements in closed-loop wound management systems utilizing conformable electrochemical devices. As illustrated in Figure 1, we begin by introducing the wound biomarkers and the associated electrochemical detection techniques. These techniques are designed to collect essential information through real-time monitoring or periodic detection, tailored to the unique characteristics of different biomarkers. Next, a comprehensive wound condition evaluation criteria and strategies were



Schematic illustration of conformable electrochemical device for closed-loop wound management.

introduced. Furthermore, we explore electrochemical-based wound treatment approaches, encompassing physical treatments and diverse drug delivery strategies. Our primary aim is to provide an extensive and in-depth perspective on the use of cutting-edge adaptable electrochemical devices in the field of wound management. This review advocates for an integrated closed-loop wound management platform that combines biomarker detection, holistic evaluation, and on-demand treatment, ultimately creating an adaptive and efficient approach to wound care.

2 Biomarkers detection

Wound environments encompass a multitude of measurable indicators crucial for comprehensive wound assessment and treatment guidance. These biomarkers provide valuable insights into the wound's condition and the healing process (Tang et al., 2021). To address this complexity, we provide a concise overview of current electrochemical detection techniques, categorizing them into two modes: real-time monitoring and regular assessment. Real-time monitoring is applied to holistic wound indicators like temperature, pH, and humidity, and small molecule metabolites (such as uric acid, nitric oxide, etc.), while regular detection is employed for biomarkers recognized by bioreceptors that can be detected by disposable devices, such as metabolites, cytokines, and enzymes. This classification allows for a targeted and tailored strategy in wound management, ensuring a cohesive approach to assessing and addressing wound conditions.

2.1 Brief introduction to electrochemical detection techniques

Electrochemical detection is a valuable tool for precisely converting wound markers into electrical signals in complex wound environments, ensuring high sensitivity and specificity. Electrochemical sensors stand out for their cost-effectiveness, user-friendliness, and portability compared to alternative methods, making them an ideal choice for in vitro diagnosis (Li et al., 2021). These sensors employ various techniques, including amperometric, voltammetric, potentiometric, and impedance methods. In amperometric methods, analyte concentration directly correlates with the measured current. Enzyme sensors, such as uricase-modified sensors for uric acid detection, are commonly used. Voltammetry, the most prevalent technique, captures redox currents in relation to analyte concentration. Potentiometric sensors detect potential shifts resulting from ion concentration changes, while impedance spectroscopy quantifies analytes based on their impedance response (Piro and Reisberg, 2017; Liu et al., 2021; Pollard et al., 2021). Wound related biomarkers and corresponding electrochemical detection methods are summarized in Table 1.

Electrochemical sensors can provide a versatile and efficient approach to monitor wound biomarkers through real-time monitoring as well as regular detection modes. These methods significantly contribute to enhancing wound management by offering precise insights into wound conditions and facilitating well-informed treatment decisions.

	Biomarkers	Method	Sensing elements	Detection range	Data transfer	Reference
Biomarkers for real- time detection	pH	Potentiometry	PEDOT	4-8	Bluetooth	Shirzaei Sani et al. (2023)
		Potentiometry	PANI	5–9	Bluetooth	Yang et al. (2022)
		Potentiometry	PANI	3–9	NFC	Xu et al. (2021)
		Voltammetry	PEDOT:PSS	4–10	Bluetooth	Liu et al. (2021)
		Potentiometry	PANI	4-9	Bluetooth	Gao et al. (2021)
	Humidity	Impedance	PEDOT: PSS		NFC	Tessarolo et al. (2021)
		Capacitance	IFRep gel		Bluetooth	Yang et al. (2022)
	Uric acid	Voltammetry	UOx	0-300 μΜ	Bluetooth	Pal et al. (2018), RoyChoudhury et al. (2018)
		Voltammetry	rGO/AuNPs	100–800 μM	NFC	Xu et al. (2021)
		Voltammetry	chitosan/PB	0-1000 μM	Bluetooth	Liu et al. (2021)
		Voltammetry	РВ	0-100 μΜ	Bluetooth	Shirzaei Sani et al. (2023)
		Voltammetry	PA/CNT	100–1000 μM	None	Jarošová et al. (2019), Simoska et al. (2020)
		Voltammetry	CUAs	100–700 µM	None	Simoska et al. (2020)
	NO	Voltammetry	CUAs	1–100 µM	None	Simoska et al. (2020)
	РҮО	Voltammetry	CUAs	1–250 µM	None	Simoska et al. (2020)
		Voltammetry	PA/CNT	1–118 µM	None	Jarošová et al. (2019)
		Voltammetry	MXene/AuNPs/ peptides	1–100 μΜ	NFC	Shi et al. (2023)
Biomarkers for regular detection	Cytokines (TNF-α, TGF- β1, IL-6, IL-8)	Voltammetry	MB	TNF-α:0–2 ng/mL; TGF-β1: 0–150 pg/mL; IL-6: 0–30 ng/mL; IL-8: 0–30 ng/mL	Bluetooth	Gao et al. (2021)
	Cathepsin, Deoxyribonuclease	Capacitance	IFRep gel	0–500 ng/mL	NFC	Yang et al. (2022)
	Sortase A	Voltammetry	MXene/AuNPs/ peptides	1 pg/mL to 100 ng/mL	NFC	Shi et al. (2023)

TABLE 1 Summary of wound related biomarkers and corresponding electrochemical detection methods.

PEDOT- poly(3,4-ethylenedioxythiophene); PANI- polyaniline; PEDOT: PSS- poly(3,4-ethylenedioxythiophene): Poly (styrenesulfonate); IFRep gel-Inflammation-responsive gel (IFRep gel); UOx- urate oxidase; PB- prussian blue; PA/CNT- Polyacrylamide-coated, carbon nanotube; CUAs- carbon ultramicroelectrode arrays; MB- methylene blue.

2.2 Biomarkers for real-time detection

Wound management is a multifaceted process involving various factors such as temperature, pH, and moisture levels, which serve as holistic biomarkers for real-time sensing. It's important to note that most holistic biomarkers are monitored in conjunction with other biomarkers to provide a comprehensive view of wound conditions from various perspectives. Additionally, small molecule metabolites, closely associated with inflammatory and infectious processes, can be continuously monitored as well. These metabolites are usually detected by using enzyme-based biosensors employing amperometric methods (e.g., glucose) or voltammetric methods.

When the integrity of skin is destroyed, many factors will lead to the change of temperature in and around the wound. It is found that the temperature is the highest in the unhealed and infected wounds, and it decreases with the healing of the wounds (Power et al., 2017; Gethin et al., 2021). Lou and colleagues, by monitoring the temperature of the wound in real time, gets the specific situation of the wound (Lou et al., 2020). For instance, a sudden temperature increase or prolonged high temperature may indicate infection or delayed healing (Gianino et al., 2018; Prasad Mahindrakar et al., 2023). Wound pH is another critical element in the wound healing equation, significantly influencing essential processes like collagen formation, inflammation, angiogenesis, and tissue oxygen utilization (Romanelli et al., 2013; Pang et al., 2023; Derwin et al., 2023). Among the pH sensing materials, polyaniline (PANI) is reported as a type of widely used conductive polymer that can sensing pH change through the protonation-deprotonation pathway (Figure 2A). And, because of their reversible property of protonation-deprotonation reaction, PANI based sensors can continuously monitoring pH value in a wide pH range (Figure 2B). In the wound environment, moisture levels are equally vital. An adequately humid setting can promote tissue growth and collagen proliferation (Weller, 2009; Kruse et al.,



FIGURE 2

Real-time monitoring (A–E) and regular detection (F–H) of electrochemical devices for various wound biomarkers. (A) The images depict a printed potentiometric sensor on an adhesive bandage, and illustration of the polyaniline protonation-deprotonation pathway from base conditions (top structure) to acid conditions (bottom structure) (Cascioferro et al., 2014). (Copyright 2014 *Electroanalysis*) (B) The potentiometric time-trace of a pH bandage sensor ranges from pH 8.51 to 2.69, with an inset showing the EMF dependence versus pH for the PANI conducting polymer ISE (Jayaseelan et al., 2014). (Copyright 2014 *Electroanalysis*) Additionally, (C–E) display square wave voltammetric current responses (background subtracted) for several biologically relevant concentrations of (C) PYO, (D) NO, and (E) UA in simulated wound exudates (Simoska et al., 2020). (Copyright 2020 *ACS Sensors*) (F–H) Aptamer-based sensors for regular detection of cytokines and bacteria, (F) presents a schematic of the sensing mechanisms, and (G–H) show variations in relative peak height based on voltammetric spectra of the TNF-α and *S. aureus* sensors (Gao et al., 2021). (Copyright 2021 *Science Advances*).

2015), while facilitating the autolysis of necrotic tissue (Nuutila and Eriksson, 2021). However, excessive exudate may not necessarily be beneficial for wound healing, as it can hinder the process (Zhang et al., 2021; Zhang et al., 2022). Striking the right moisture balance is critical because both excessive and insufficient moisture can impede wound healing (Okan et al., 2007). Notably, pH is often monitored by potentiometry and Voltammetric technology, and humidity is often monitored by impendence and capacity.

The wound-environment interface contains electroactive small molecule metabolites, such as glucose, uric acid, and nitric oxide, closely linked to inflammation and infection. These metabolites significantly affect wound healing. Detecting and monitoring these metabolites offers valuable insights into wound conditions and informs appropriate treatments. For example, high glucose levels can harm blood vessels, impair nerve function, reduce circulation, and weaken the immune system, all vital for wound healing. Prolonged uncontrolled blood sugar can result in chronic wounds, which are more susceptible to infection due to impaired healing (Zhu et al., 2019). Uric acid (UA) serves as a critical indicator of wound severity, with its

concentration increasing as wounds become more severe. Excess uric acid levels can hinder healing by promoting inflammation, especially in chronically hypoxic wounds (Fernandez et al., 2012; Fernandez et al., 2014; Faruk Hossain and Slaughter, 2021). Nitric oxide (NO) has antiinflammatory and antibacterial properties, regulates angiogenesis, and influences collagen deposition, playing a crucial role in various cell activities during wound healing. Wounds in the inflammatory stage and those infected by bacteria produce significant NO amounts (Schwentker et al., 2002; Schwentker and Billiar, 2003; Luo and Chen, 2005; Cinelli et al., 2019; Wu et al., 2021; Li et al., 2022). Bacterial metabolites with redox activity such as pyocyanin (PYO) serve as an infection indicator, which is also applied as biomarker for real-time detection. As shown in Figures 2C-E, UA, NO, and PYO with redox activity are able to be simultaneously detected in wound samples or simulators using a flexible carbon microelectrode array (CUAs) sensor. Square wave voltammetry (SWV) was employed to evaluate the sensor's response, determining the limits of detection (LODs) and linear dynamic range (LDR) in simulated wound fluid. It was found that the characteristic SWV current response of PYO and uric acid increased with the increase of analyte concentration in the micromolar range determined by *in vitro* and *in vivo* concentrations (Simoska et al., 2020). As Figures 2F–H shows, a multiplex immunosensor can realize the in-situ analysis of wound microenvironment, inflammation and infection status through real-time quantitative evaluation of pro-inflammatory factor TNF- α and bacterial load. The concentration of TNF- α in wound fluid of unhealed ulcer is higher than that of healed ulcer. *Staphylococcus aureus* is a dominant species in all types of chronic wound samples. The peak current height of each adaptive sensor decreases with the increase of target concentration, in which the monitoring range of TNF- α is 0–2 ng/mL, and the range of *Staphylococcus aureus* is 0–1 × 10⁹ colony forming unit (CFU)/ml, and it has been proved that it has good selectivity (Gao et al., 2021).

2.3 Biomarkers for regular detection

In contrast to the holistic biomarkers and some small molecule metabolites, there are specific biomarkers present in the wound interfaces that are related to both external microbes and internal physiological conditions and inflammatory responses. However, the recognition of these biomarkers is not reversible, and they can only be detected with disposable biosensors. Specific detection techniques are necessary to target these biomarkers and provide a precise evaluation of the wound conditions.

Cytokines are small molecular proteins, which are involved in the regulation and coordination of various cell types (Borena et al., 2015) and are very important in cell signal transduction. Its role may be promoting inflammation or inhibiting inflammation (Pilvenyte et al., 2023), strictly control the wound healing process (Efron et al., 2000; Werner and Grose, 2003). Therefore, it is very meaningful to monitor the change of cytokine expression level (Filik and Avan, 2020). Legrand and coworkers reported that tumor necrosis factoralpha (TNF-a), interleukin-6 (IL-6), interleukin-8 (IL-8) and transforming growth factor- β 1 (TGF- β 1) are used for wound assessment. Pro-inflammatory cytokines interleukin-1 (IL-1), IL-6 and TNF-a are upregulated in the inflammatory stage of wound healing (Legrand and Martino, 2022), and their concentrations in chronic wounds are higher than those in normal healing wounds (Gianino et al., 2018). TNF-a and IL-1 are necessary in the early stage of wound healing, but the persistent over-expression may lead to the aggravation of tissue damage through the over-activation of immune cells and their protease products (Efron and Moldawer, 2004). TNF-α has pleiotropic, growth regulating and differentiation effects on various cell types. It is a key regulator of inflammatory reaction and plays an important role in inflammation and infection (Pilvenyte et al., 2023). Three days after skin injury, the level of TNFa in wound fluid reached its peak. IL-1 exists in two forms: IL-1a and IL-1 β . IL-1 β is a pro-inflammatory cytokine, which can be induced by infections such as Staphylococcus aureus and Pseudomonas aeruginosa. It can be detected in the wound environment within 24 h after injury, and its concentration reaches its peak within 24-72 h. Infections such as Staphylococcus aureus and Pseudomonas aeruginosa can induce IL-1ß secretion (Efron and Moldawer, 2004; Barrientos et al., 2008; Pilvenyte et al., 2023). TGF-β plays an important role in inflammation, angiogenesis, epithelial re-formation and connective tissue regeneration. With the occurrence of traumatic injury, its expression also increased. For these wound markers, electrochemical device such as aptamer modified.

Matrix metalloproteinases (MMPs) are the main proteases involved in wound healing, among which MMP-9, MMP-2 and tissue inhibitor of matrix metalloproteinases (TIMPs) are the most common components in wound exudates (Power et al., 2017). The contents of MMP-9 and MMP-2 decreased during wound healing (Tarlton et al.,1997). The balance of protease is very important for wound healing. Excessive protease activity will cause damage to extracellular matrix (ECM) and newly formed tissues, thus delaying the healing process. MMP-9 is produced by many kinds of cells and induced by cytokines, growth factors, stress or inflammation (Rayment et al., 2008). TIMPs is a tissue inhibitor of matrix metalloproteinases, which regulates the activity of MMPs (Behm et al., 2012; Lindley et al., 2016). According to the research, the long-term increase of matrix protease level and the decrease of TIMPs level or the abnormal ratio of matrix protease level to TIMPs level are related to wound nonunion, especially the decrease of MMP/TIMP ratio is a good prognostic indicator of wound behavior (Ladwig et al., 2002). Taking advantage of the proteinase activity, MMPs can be detected using substrate peptide-based biosensors. Using MMP-2 as an example, the peptide unit consists of two regions: one for self-assembly and one for biorecognition. Ion nanochannels can then generate an electrochemical response in response to MMP activities (Wang et al., 2022).

Many pathogenic species, including *Pseudomonas aeruginosa*, *Enterococcus faecalis, Escherichia coli* and *Staphylococcus aureus*, are usually distributed in chronic wounds. Therefore, microbiological methods for pathogen identification can be used as qualitative indicators of infection (Simoska et al., 2020). The current research progress, aiming at the detection of bacteria, Enzymes such as lipase, hyaluronidase, antimicrobial peptides, antibacterial peptides, substances on the outer membrane of bacteria (such as lipopolysaccharide (LPS), peptidoglycan and lipopeptide), bacterial flagellin, bacterial DNA, etc., are used to monitor bacteria (Khoshroo et al., 2022; Wang et al.,2020).

3 Evaluation of the wound status

Despite the presence of a wide array of strategies and technologies in wearable chemical sensors for wound interfaces and non-invasive biofluids, the parameterization and calibration of these sensor modes have not received enough attention (Ronkainen et al., 2010). It is crucial to emphasize the collection and integration of data from multiplexed and multimodal sensors in the context of wound information. This is imperative for robust post-processing, particularly when considering the physiological (inflammation) and pathogenic (infection) processes at the wound interface (Sempionatto et al., 2022). The comprehensive interpretation of this multiplexed and multimodal data, whether derived from real-time monitoring or regular sensor readings, occupies an important position in the closeloop wound management systems.

3.1 Inflammation at the wound interfaces

The body's immune system plays a crucial role in initiating an inflammatory response as a protective mechanism at the wound



FIGURE 3

Evaluation of inflammatory status of wound. (A–C) Inflammatory Evaluation with a Real-Time NO Monitoring Device in the Puncture Cavity of New Zealand Rabbits (Li et al., 2020). (Copyright 2020 *Nature Communications*) (A) X-ray image and (B) the implanted NO sensor, wire connections, and wireless module. A photograph of the rabbit after NO sensor implantation with the wireless circuit secured on the thigh. (C) High NO concentration correlated with IL-1 β -induced inflammation (blue), compared to the untreated (black) and penicillin-treated (red) groups. (D–G) Interpretation of Multiplexed Immunosensors with Wound Parameters (Gao et al., 2021). (Copyright 2021 *Science Advances*) (D) Illustration of the microenvironment of venous ulcers and *in situ* assessment of pH, temperature, mouse TNF- α , and *S. aureus* using Immunosensors. (E) Radar map depicting the integration of different inflammatory-related biomarkers, including TNF- α , IL-6, IL-8, TGF- β J, *S. aureus*, and pH sensors. (F, G) Interpretation of sensor-derived data analysis of wound exudate from a patient with a venous ulcer over a 5-week period (F) and the resulting correlation matrices (G).

interface. This response involves the participation of various signaling molecules, including metabolites, cytokines, and macrophages, which work in concert to regulate the immune response and coordinate the repair process (Raziyeva et al., 2021; Zhao et al., 2016).

The assessment of inflammation holds paramount importance in effective wound management, with numerous biomarkers intricately involved in this process. NO plays a pivotal role in the regulation of the immune response by influencing key factors such as chemokines, IL-8, TGF- β 1, monocytes, and neutrophils, etc., (Luo and Chen, 2005; Cinelli et al., 2019; Wu et al., 2021). In the proliferative stage of wound healing, NO promotes the proliferation of endothelial cells and mediates the production of vascular endothelial growth factor (VEGF) (Luo and Chen, 2005). A study by Li R. et al. illustrated a strong correlation between NO and cytokines, utilizing a real-time wireless NO monitoring platform (Li et al., 2020) (Figures 3A, B). Moreover, NO sensors placed in the puncture cavity of New Zealand rabbits exhibited a robust correlation with the inflammatory cytokine IL-1 β and proved adaptable for wound assessment (Figure 3C).

Considering the complex nature of conditions at the wound interface, the need for versatile and integrated in-situ multi-channel inflammatory cytokine detection platforms, along with holistic pH sensors, becomes evident for comprehensive wound condition assessment (Figure 3D). The Lim group's work showcases the integration of various types of inflammatory data using a flexible multiplexed immunosensor, allowing quantitative interpretation of cytokines like TNF- α , IL-6, IL-8, and the growth factor TGF- β 1 through a radar map (Figure 3E) (Gao et al., 2021). This approach also revealed changes in these biomarkers closely correlated with different stages of wound recovery over a period of 5 weeks. The correlation of this inflammatory information with wound conditions offers valuable insights into the healing process



Advances) (C) Schematics of the infection status. (A) itidstrates a soft sensor parcin equipped with pir and temperature (Shirzaei Sani et al., 2023). (Copyright 2023 Science Advances) (C) Schematics of the infection sensing mechanism for DNAgel-based infectious evaluation. DNAgel degrades upon exposure to DNase, resulting in a change in the capacitance of the sensor. (D) Signal changes recorded by a smartphone for *S. aureus* load at the wound site established by wound culture 24 h after wounding (Xiong et al., 2021). (Copyright 2021 *Sci Adv.*) (E, F) Schematics of the electrochemical detection of pyocyanin (E) based on MXene-modified electrodes and detection of softase A (F) with MXene/AuNPs-modified electrode in the presence of a 100 µM ferrocenylacetic acid solution in PBS solution (Shi et al., 2023). (Copyright 2023 *Sensors and Actuators B: Chemical*.)

(Figures 3F, G). Ultimately, this integrated immune monitoring system serves as a powerful tool for real-time assessment of wound conditions, offering invaluable support for optimizing wound management and making informed treatment decisions.

3.2 Infection at the wound interfaces

In the context of chronic wounds, infections present significant challenges as external microorganisms release virulence metabolites. These metabolites cause changes in the overall wound environment and specific biomarker alterations. These alterations hold vital information for early warning and diagnostics, bridging the crucial link between infection and wound assessment to the on-demand treatment.

Holistic biomarkers such as temperature, pH, and other vital parameters can be utilized to evaluate infectious status. For temperature, distinct temperature changes were observed in wounds infected with various bacterial strains. Prolonged elevated temperatures may indicate infections, such as those caused by *Staphylococcus aureus*. A Flexible Wound Healing System (FWHS) incorporates a visual indicator and an alarm for automatically detecting abnormal temperature changes exceeding the rectal temperature. This system, tested on both early and late infected wounds, demonstrated varying temperature fluctuations at different stages, confirming its early warning capabilities (Lou et al., 2020). In Gao group, wearable patches with pH and temperature sensor arrays were used to monitor the critical size of full-thickness infected wound defects in diabetic rats (Figure 4A). They observed that the infected wound showed a more uniform pH and temperature in regions 2 and 3 days after infection due to the formation of a uniform biofilm after infection. After treatment on the fourth day, the pH value and temperature value decreased significantly and returned to the level before infection on the seventh day, which indicated that the biofilm was destroyed and finally eliminated after treatment (Figure 4B) (Shirzaei Sani et al., 2023).

The virulence factors associated with conditional pathogens can serve as reliable criteria for assessing infection status (Clatworthy et al., 2007; Mota et al., 2021). For instance, the secretion of deoxyribonuclease (DNase) by bacteria such as *Staphylococcus aureus*, *Pseudomonas aeruginosa*, and *Streptococcus* pyogenes reflects the occurrence of infection (Figure 4C). In this method, a reactive DNAgel with wound extrudes is employed. Elevated levels of DNase trigger the degradation of the DNA hydrogel, resulting in a change in the dielectric constant. This change can be utilized as an indicator to evaluate the infection status. When infection occurs, the bacterial reactive DNAgel contacts with DNase, and the bacterial reactive DNA hydrogel degrades, which leads to the change of dielectric constant in the area above the staggered electrodes, thus triggering the infection alarm on the smart phone. The capability to indicate wound infection *in vivo* was validated with an acute wound model in mice. The widely accepted clinical threshold for laboratory infection diagnosis, ranging from 10^5 to 10^6 colony-forming units (CFU), was employed. In the control group, minimal changes were observed.

However, in wounds inoculated with live *Staphylococcus aureus* (10⁵ and 10⁶ CFU), a signal change of 0.4 V was detected after 24 h (Xiong et al., 2021) (Figure 4D). In another study, two typical biomarkers, sortase A and pyocyanin,

were employed to indicate infections caused by Gram-positive *Staphylococcus aureus* and Gram-negative *Pseudomonas aeruginosa*. Electrochemical differential pulse voltammetry was utilized for timely evaluation of wound infections and to distinguish between infection types. Results from animal experiments demonstrate the effectiveness of the smart bandage in conducting multi-biomarker analysis of wounds, enabling the early warning of wound infections (Figures 4E, F) (Shi et al., 2023).

The wound evaluation approaches mentioned above are based on specific biomarkers closely related to the status of wound conditions. However, the variation in these wound-related biomarkers is influenced by multiple factors within the complex wound environment. In such cases, artificial intelligence can analyze complex data, offering potential applications in early detection, diagnosis, treatment, prognosis prediction, and prognosis evaluation of wounds, contributing significantly to the field of wound care. Research has successfully integrated artificial intelligence technology into wound management, particularly in the classification of chronic wounds, including diabetic ulcers, lymphatic vessel wounds, surgical wounds, and pressure injuries (Sarp et al., 2021). This integration has facilitated accurate evaluations of wound area and the percentage of granulation tissue (PGT) (Howell et al., 2021), prediction of the healing stage of inflammatory wounds (Kalasin et al., 2022), and overall assessment and management of wounds (Barakat-Johnson et al., 2022). These advancements not only optimize the diagnosis and treatment workflow but also improve the efficiency of the entire wound care process.

4 On-demand wound management systems

The progress in the management of wounds has resulted in the emergence of precise, instantaneous, and dynamic on-demand treatment systems that cater to individual requirements (Verdolino et al., 2021; Vivcharenko et al., 2023). Traditional flexible electrochemical devices with singular monitoring functions are no longer adequate. To effectively address the needs of wound management, integrated closed-loop systems that amalgamate monitoring and controllable treatment systems are emerging as a superior option (Pollard et al., 2021).

These integrated closed-loop systems hold immense potential, encompassing diagnosis, early infection warning, treatment effect assessment, and long-term wound management (Liu et al., 2023). Recent years the application of physical treatment (i.g. electrical stimulation), controlled drug release, or combined treatment have been reported in closed-loop wound treatment (Bagherifard et al., 2016; Amjadi et al., 2018; Jiang et al., 2022).

4.1 Physical treatment

Electrical stimulation (ES) has exhibited its efficacy in regulating cellular proliferation and migration, mitigating inflammation, and expediting wound closure by emulating or amplifying the inherent electromagnetic field's influence induced by potassium (K⁺) and sodium (Na⁺) ions (Luo et al., 2021). Electrical stimulation application was found to accelerate wound healing and skin remodelling, with increased wound impedance and improved histological characteristics such as skin thickness and the number of skin appendages. Single-cell RNA sequencing (SCRNA-SEQ) analysis confirmed the activation of regeneration-promoting genes in monocytes and macrophages and followed by enhancing tissue regeneration, neovascularization, and skin recovery (Jiang et al., 2022). Yang and coworkers proposed a medic-free wound manage approach by a wireless electrochemical device (Figure 5A) that contains a unit for electrical/optical stimulations (Figure 5B). Figure 5C suggests the electronic dressing devices induced wound healing acceleration successfully with only physical treatment (Yang et al., 2022).

Furthermore, phototherapy, encompassing photodynamic therapy (PDT) and photothermal therapy (PTT), has been demonstrated to accelerate wound healing by impacting macrophage phenotypic metamorphosis, fostering tissue proliferation, and impeding bacterial functionalities, thereby restraining biofilm growth and microbial infections (Xu et al., 2022; Chen et al., 2023).

4.2 Controlled drug release treatment

In certain situations, physical treatments alone may not suffice for managing chronic wound conditions, particularly when dealing with infectious microbes. There is a clear need for specific drugs and controlled drug release strategies that can respond actively based on the information and feedback obtained through monitoring and simulation results. This integrated approach offers a more comprehensive and effective way to address the complex and evolving nature of chronic wounds, ensuring that treatment is tailored to individual needs (Luo et al., 2021; Jiang et al., 2022).

One common strategy is thermal-induced drug release, known for its ease of control. Gong et al., integrated a thermally responsive nanomesh film: Screen printing crosslinked poly (N- isopropylacrylamide -N- methacrylamide), with loaded moxifloxacin hydrochloride (MOX) and a dual-functional electrochemical device for wound management. As shown in Figure 5D, this device can monitor temperature changes when double-sided switch connects to site A, and the signal can be wirelessly transmitted to a mobile phone for infection determination. If an increase in temperature was confirmed by



FIGURE 5

Conformable electrochemical device for wound therapy. (A) Photograph of an all-in-one wireless electronic wound dressing system with two units as shown in (B) Unit I for sensing and optical/electrical stimulations, and Unit II for functional control. (C) Images of wound closure on day 0 and day 14 without or with electric stimulation by the device in A (Yang et al., 2022). (Copyright 2022 *Nano Today*) (D) Schematic illustration of temperature monitoring (double-sided switch connecting to site (A) and thermal-induced MOX drug release from C-PNHM (double-sided switch connecting to site (B) for on-demand wound management. (E) Temperature responsive MOX release from C-PNHM during several temperature change cycle between 25°C and 45°C; inset: viability of bacteria after 1 h or 6 h incubation with C-PNHM fiber at 25°C or 45°C (Gong et al., 2019) (Copyright 2019 *Advanced Functional Material*) (F) Schematic diagram of simultaneously exudate removal and drug release in thermal response. (G) Cefazolin release under cyclic thermal stimulation (5 min in 42°C and 5 min in 25°C) (Ge et al., 2023). (H) Schematic diagram of voltage-controlled drug release in a battery-free wound management system (Xu et al., 2021). (Copyright 2021 *Advanced Functional Material*).

infection, turning double-sided switch to site B can active the heater function of device. The heater's temperature can be precisely controlled by adjusting the applied voltage, ensuring accurate MOX drug release from nanofibers (Figure 5E) for effective bacterial infection control (Gong et al., 2019).

Ge Z and coworkers added an exudate management module to optimize wound management. Self-pumping microfluidic channel, temperature and humidity sensor, liquid metal (LM) heating module and antibiotics-loaded thermosensitive hydrogel are integrated into wearable wound patch (Figure 5F). When the pulse voltage was applied to the LM heating coil, the exudate was removed through microfluidic channel, and the broad-spectrum antibiotic cefazolin was released from the temperature-sensitive hydrogel. The drug release rate at 42°C is significantly higher than that at 25°C, showing controllable and on-demand thermal response release characteristics (Figure 5G). Under the combined action of exudate removal and drug treatment, the wound temperature returned to normal level quickly due to a synergistic effect (Ge et al., 2023). Xu and coworkers adopted conductive polymer and prepared a battery free electronically controlled drug release system offers low power consumption, costeffectiveness, and rapid response (Figure 5H) (Xu et al., 2021).

Light serves as an optimal stimulus for precise drug delivery owing to its cleanliness, accuracy, and remote controllability. A comprehensive system comprising a temperature sensor, UVresponsive antibacterial hydrogel, and UV-LED enables the monitoring of wound temperature in real-time and the release of antibiotics as needed. Once the presence of infection was confirmed, the UV-LED (365 nm) is applied to activate the release of gentamicin (GS) and the release rate of GS escalates with prolonged exposure to UV irradiation. Upon the release of antibiotic, a gradual decrease in temperature can be observed. Subsequent cycles of UV irradiation maintained this pattern. Animal experiments validated the system's capability for realtime wound monitoring, infection detection, and effective treatment (Pang et al., 2020).

4.3 Combined wound treatment

The combination of electrical stimulation and drugs offers a comprehensive approach to wound management. A multichannel electrochemical biosensor array is employed to monitor wound biomarkers and regulate drug release via electrodes. The positively charged chondroitin 4-sulfate (CS) hydrogel facilitates the controlled release of dual-function anti-inflammatory and antibacterial peptides (AMP). *In vitro* experiments validated the precise release of AMP and its potent antibacterial activity against diverse pathogens. After treatment, pH and temperature normalized by the fifth and sixth days. The combined treatment group exhibited the highest rate of wound closure, as evidenced by histopathological analysis revealing collagen deposition and the formation of granulation tissue (Shirzaei Sani et al., 2023).

Above all these treatment strategies are effective in different scenarios. Thermal stimulation drug release is common in wound management but comes with drawbacks like high energy consumption and the need for external components. In contrast, electric stimulation release is cost-effective and has low power consumption, although it's limited to charged drugs. Light, as a clean energy source, offers high temporal and spatial resolution and remote control for drug release. The choice of treatment methods is closely tied to specific wound conditions, underscoring the importance of closed-loop wound management (Armstrong et al., 2020; Jiang et al., 2022; Pang et al., 2023).

5 Discussion and perspectives

In summary, our review has provided an overview of closed-loop wound management systems based on conformable electrochemical devices. These systems encompass biomarker detection, wound evaluation, and on-demand treatment, offering a comprehensive and efficient approach to wound management. Wound healing is a complex process that demands a thorough understanding of its status. Utilizing various electrochemical techniques to assess multiple wound biomarkers from different perspectives is essential, forming a solid foundation for diagnosis and treatment. Different wound types present distinct microenvironments, requiring adjustments in monitoring parameters for more precise and detailed assessments. The choice of electrochemical strategies depends on the types of biomarkers, ranging from holistic environmental biomarkers requiring continuous monitoring to specific biomarkers necessitating regular detection. The integration of multi-channel electrochemical devices allows for comprehensive monitoring, including wound microflora and their metabolites, by combining various detection modes. This comprehensive evaluation of wound conditions enables the precise identification of infectious or inflammatory states, facilitating timely treatment. Implementing measures to prevent biological contamination of interfaces, enhance sensitivity and specificity, perform active biosensor calibration, reduce response times, lower detection limits, and advance equipment miniaturization, networking, and intelligence is crucial for optimizing electrochemical sensors.

Furthermore, these closed-loop systems can address the challenges posed by multidrug resistance associated with traditional antibiotics. Alternative therapies such as growth factors, NO, metal ions, antimicrobial peptides, and physical modalities like electrical stimulation, light therapy, ultrasound, and heat have emerged as promising options for promoting skin regeneration. Recent developments have seen the application of drug-controlled release, electrical stimulation, photobiological regulation, and other technical methods in closed-loop wound treatment. These advancements underscore the increasing need for innovative design and manufacturing technologies for wearable sensors.

The future of wound management is heading towards intelligent platforms that seamlessly integrate monitoring and treatment, creating a closed-loop for wound detection and treatment. This evolution not only depends on the development of sensing materials but also the incorporation of artificial intelligence algorithms capable of integrating the uncovered bioinformation at the sensor-wound interfaces. With this capability, precise wound models can be reconstructed into comprehensive profiles, enabling much more efficient wound management steps based on personalized situations. A promising approach involves integrating monitoring systems with controllable treatment systems to establish comprehensive closed-loop systems for detection, diagnosis, and treatment.

Author contributions

JL: Writing-original draft. ZL: Writing-review and editing. JX: Writing-review and editing. CN: Conceptualization, Funding acquisition, Writing-original draft, Writing-review and editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This research was funded by the Zhejiang Provincial Natural Science Foundation (LQ22H300004) and Ningbo Natural Science Foundation (2022J045).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

Amjadi, M., Sheykhansari, S., Nelson, B. J., and Sitti, M. (2018). Recent advances in wearable transdermal delivery systems. *Adv. Mater* 30 (7), 8. doi:10.1002/adma. 201704530

Armstrong, D. G., Bauer, K., Bohn, G., Carter, M., Snyder, R., and Serena, T. E. (2020). Principles of best diagnostic practice in tissue repair and wound healing: an expert consensus. *Diagnostics* 11 (1), 50. doi:10.3390/diagnostics11010050

Bagherifard, S., Tamayol, A., Mostafalu, P., Akbari, M., Comotto, M., Annabi, N., et al. (2016). Dermal patch with integrated flexible heater for on demand drug delivery. *Adv. Healthc. Mater* 5 (1), 175–184. doi:10.1002/adhm.201500357

Barakat - Johnson, M., Jones, A., Burger, M., Leong, T., Frotjold, A., Randall, S., et al. (2022). Reshaping wound care: evaluation of an artificial intelligence app to improve wound assessment and management amid the COVID-19 pandemic. *Int. Wound J.* 19 (6), 1561–1577. doi:10.1111/iwj.13755

Barrientos, S., Stojadinovic, O., Golinko, M. S., Brem, H., and Tomic-Canic, M. (2008). PERSPECTIVE ARTICLE: growth factors and cytokines in wound healing. *Wound Repair Regen.* 16 (5), 585–601. doi:10.1111/j.1524-475X.2008.00410.x

Behm, B., Babilas, P., Landthaler, and M., and Schreml, S. (2012). Cytokines, chemokines and growth factors in wound healing. *J. Eur. Acad. Dermatol Venereol.* 26 (7), 812–820. doi:10.1111/j.1468-3083.2011.04415.x

Borena, B. M., Martens, A., Broeckx, S. Y., Meyer, E., Chiers, K., Duchateau, L., et al. (2015). Regenerative skin wound healing in mammals: state-of-the-art on growth factor and stem cell based treatments. *Cell Physiol. Biochem.* 36 (1), 1–23. doi:10.1159/ 000374049

Cascioferro, S., Totsika, M., and Schillaci, D. (2014). Sortase A: an ideal target for antivirulence drug development. *Microb. Pathog.* 77, 105–112. doi:10.1016/j.micpath.2014. 10.007

Chen, H. Y., Wu, L. J., Wang, T. Y., Zhang, F. L., Song, J. Y., Fu, J., et al. (2023). PTT/ PDT-induced microbial apoptosis and wound healing depend on immune activation and macrophage phenotype transformation. *Acta Biomater*. 167, 489–505. doi:10.1016/ j.actbio.2023.06.025

Cinelli, M. A., Do, H. T., Miley, G. P., and Silverman, R. B. (2019). Inducible nitric oxide synthase: regulation, structure, and inhibition. *Med. Res. Rev.* 40 (1), 158–189. doi:10.1002/med.21599

Clatworthy, A. E., Pierson, E., and Hung, D. T. (2007). Targeting virulence: a new paradigm for antimicrobial therapy. *Nat. Chem. Biol.* 3 (9), 541–548. doi:10.1038/ nchembio.2007.24

Derwin, R., Patton, D., Strapp, H., and Moore, Z. (2023). Wound pH and temperature as predictors of healing: an observational study. *J. Wound Care* 32 (5), 302–310. doi:10. 12968/jowc.2023.32.5.302

Efron, D. T., Most, D., and Barbul, A. (2000). Role of nitric oxide in wound healing. *Curr. Opin. Clin. Nutr. Metab. Care* 3 (3), 197–204. doi:10.1097/00075197-200005000-00006

Efron, P. A., and Moldawer, L. L. (2004). Cytokines and wound healing: the role of cytokine and anticytokine therapy in the repair response. *J. Burn Care Rehabil.* 25 (2), 149–160. doi:10.1097/01.bcr.0000111766.97335.34

Faruk Hossain, M., and Slaughter, G. (2021). Flexible electrochemical uric acid and glucosebiosensor. *Bioelectrochemistry* 141, 6, 107870. doi:10.1016/j.bioelechem.2021. 107870

Fernandez, M. L., Upton, Z., Edwards, H., Finlayson, K., and Shooter, G. K. (2012). Elevated uric acid correlates with wound severity. *Int. Wound J.* 9 (2), 139–149. doi:10. 1111/j.1742-481X.2011.00870.x

Fernandez, M. L., Upton, Z., and Shooter, G. K. (2014). Uric acid and xanthine oxidoreductase in wound healing. *Curr. Rheumatol. Rep.* 16 (2), 396. doi:10.1007/s11926-013-0396-1

Filik, H., and Avan, A. A. (2020). Electrochemical immunosensors for the detection of cytokine tumor necrosis factor alpha: a review. *Talanta* 211, 17, 120758. doi:10.1016/j. talanta.2020.120758

Gao, Y., Nguyen, D., Yeo, T., Lim, S. B., Tan, W. X., Madden, L. E., et al. (2021). A flexible multiplexed immunosensor for point-of-care *in situ* wound monitoring. *Sci. Adv.* 7 (21), eabg9614. doi:10.1126/sciadv.abg9614

Ge, Z., Guo, W., Tao, Y., Sun, H., Meng, X., Cao, L., et al. (2023). Wireless and closedloop smart dressing for exudate management and on-demand treatment of chronic wounds. *Adv. Mater* 2023, e2304005. doi:10.1002/adma.202304005

Gethin, G., Ivory, J. D., Sezgin, D., Muller, H., O'Connor, G., and Vellinga, A. (2021). What is the "normal" wound bed temperature? A scoping review and new hypothesis. *Wound Repair Regen.* 29 (5), 843–847. doi:10.1111/wrr.12930

Gianino, E., Miller, C., and Gilmore, J. (2018). Smart wound dressings for diabetic chronic wounds. *Bioengineering* 5 (3), 5030051. doi:10.3390/ bioengineering5030051

Gong, M., Wan, P., Ma, D., Zhong, M., Liao, M., Ye, J., et al. (2019). Flexible breathable nanomesh electronic devices for on-demand therapy. *Adv. Funct. Mater* 29 (26). doi:10.1002/adfm.201902127

Howell, R. S., Liu, H. H., Khan, A. A., Woods, J. S., Lin, L. J., Saxena, M., et al. (2021). Development of a method for clinical evaluation of artificial intelligence-based digital wound assessment tools. *JAMA Netw. Open* 4 (5), e217234. doi:10.1001/jamanetworkopen.2021.7234

Jarošová, R., McClure, S. E., Gajda, M., Jović, M., Girault, H. H., Lesch, A., et al. (2019). Inkjet-printed carbon nanotube electrodes for measuring pyocyanin and uric acid in a wound fluid simulant and culture media. *Anal. Chem.* 91 (14), 8835–8844. doi:10.1021/acs.analchem.8b05591

Jayaseelan, S., Ramaswamy, D., and Dharmaraj, S. (2014). Pyocyanin: production, applications, challenges and new insights. *World J. Microbiol. Biotechnol.* 30 (4), 1159–1168. doi:10.1007/s11274-013-1552-5

Jiang, Y., Trotsyuk, A. A., Niu, S., Henn, D., Chen, K., Shih, C.-C., et al. (2022). Wireless, closed-loop, smart bandage with integrated sensors and stimulators for advanced wound care and accelerated healing. *Nat. Biotechnol.* 41 (5), 652–662. doi:10.1038/s41587-022-01528-3

Kalasin, S., Sangnuang, P., and Surareungchai, W. (2022). Intelligent wearable sensors interconnected with advanced wound dressing bandages for contactless chronic skin monitoring: artificial intelligence for predicting tissue regeneration. *Anal. Chem.* 94 (18), 6842–6852. doi:10.1021/acs.analchem.2c00782

Khoshroo, A., Mavaei, M., Rostami, M., Valinezhad-Saghezi, B., and Fattahi, A. (2022). Recent advances in electrochemical strategies for bacteria detection. *BioImpacts* 12 (6), 567–588. doi:10.34172/bi.2022.23616

Kruse, C. R., Nuutila, K., Lee, C. C. Y., Kiwanuka, E., Singh, M., Caterson, E. J., et al. (2015). The external microenvironment of healing skin wounds. *Wound Repair Regen.* 23 (4), 456–464. doi:10.1111/wrr.12303

Ladwig, G. P., Robson, M. C., Liu, R. A. N., Kuhn, M. A., Muir, D. F., and Schultz, G. S. (2002). Ratios of activated matrix metalloproteinase-9 to tissue inhibitor of matrix metalloproteinase-1 in wound fluids are inversely correlated with healing of pressure ulcers. *Wound Repair Regen*. 10 (1), 26–37. doi:10.1046/j.1524-475x.2002.10903.x

Legrand, J. M. D., and Martino, M. M. (2022). Growth factor and cytokine delivery systems for wound healing. *Cold Spring Harb. Perspect. Biol.* 14 (8), a041234. doi:10. 1101/cshperspect.a041234

Li, D., Wu, C., Tang, X. H., Zhang, Y., and abd Wang, T. (2021). Electrochemical sensors applied for *in vitro* diagnosis. *Chem. Res. Chin. U* 37 (4), 803–822. doi:10.1007/ s40242-021-0387-0

Li, R., Qi, H., Ma, Y., Deng, Y., Liu, S., Jie, Y., et al. (2020). A flexible and physically transient electrochemical sensor for real-time wireless nitric oxide monitoring. *Nat. Commun.* 11 (1), 3207. doi:10.1038/s41467-020-17008-8

Li, X., Lu, Y., and Hu, Y. (2022). A wireless and battery-free DNA hydrogel biosensor for wound infection monitoring. *Matter* 5 (8), 2473–2475. doi:10.1016/j.matt.2022. 06.021

Lindley, L. E., Stojadinovic, O., Pastar, I., and Tomic-Canic, M. (2016). Biology and biomarkers for wound healing. *Plast. Reconstr. Surg.* 138, 185–285. doi:10.1097/PRS. 00000000002682

Liu, Y., Li, J., Xiao, S., Liu, Y., Bai, M., Gong, L., et al. (2023). Revolutionizing precision medicine: exploring wearable sensors for therapeutic drug monitoring and personalized therapy. *Biosensors* 13 (7), 726. doi:10.3390/bios13070726

Liu, Z., Liu, J., Sun, T., Zeng, D., Yang, C., Wang, H., et al. (2021). Integrated multiplex sensing bandage for *in situ* monitoring of early infected wounds. *ACS Sens.* 6 (8), 3112–3124. doi:10.1021/acssensors.1c01279

Lou, D., Pang, Q., Pei, X., Dong, S., Li, S., Tan, W. q., et al. (2020). Flexible wound healing system for pro-regeneration, temperature monitoring and infection early warning. *Biosens. Bioelectron.* 162, 112275. doi:10.1016/j.bios.2020.112275

Luo, J. d., and Chen, A. F. (2005). Nitric oxide: a newly discovered function on wound healing. *Acta Pharmacol. Sin.* 26 (3), 259–264. doi:10.1111/j.1745-7254.2005.00058.x

Luo, R., Dai, J., Zhang, J., and Li, Z. (2021). Accelerated skin wound healing by electrical stimulation. *Adv. Healthc. Mater* 10 (16), e2100557. doi:10.1002/adhm. 202100557

Mathew, M., Radhakrishnan, S., Vaidyanathan, A., Chakraborty, B., and Rout, C. S. (2020). Flexible and wearable electrochemical biosensors based on two-dimensional materials: recent developments. *Anal. Bioanal. Chem.* 413 (3), 727–762. doi:10.1007/s00216-020-03002-y

Mirvakili, S. M., and Langer, R. (2021). Wireless on-demand drug delivery. Nat. Electron. 4 (7), 464-477. doi:10.1038/s41928-021-00614-9

Mota, F., Pereira, S., Araújo, A., Passos, M., and Saraiva, M. (2021). Biomarkers in the diagnosis of wounds infection: an analytical perspective. *TrAC Trends Anal. Chem.* 143, 116405. doi:10.1016/j.trac.2021.116405

Nuutila, K., and Eriksson, E. (2021). Moist wound healing with commonly available dressings. *Adv. Wound Care* 10 (12), 685–698. doi:10.1089/wound.2020.1232

Okan, D., Woo, K., Ayello, E. A., and Sibbald, G. (2007). The role of moisture balance in wound healing. *Adv. Skin. Wound Care* 20 (1), 39–53. doi:10.1097/00129334-200701000-00013 Pal, A., Goswami, D., Cuellar, H. E., Castro, B., Kuang, S., and Martinez, R. V. (2018). Early detection and monitoring of chronic wounds using low-cost, omniphobic paperbased smart bandages. *Biosens. Bioelectron.* 117, 696–705. doi:10.1016/j.bios.2018. 06.060

Pang, Q., Lou, D., Li, S., Wang, G., Qiao, B., Dong, S., et al. (2020). Smart flexible electronics-integrated wound dressing for real-time monitoring and on- demand treatment of infected wounds. *Adv. Sci.* 7 (6), 1902673. doi:10.1002/advs.201902673

Pang, Q., Yang, F., Jiang, Z., Wu, K., Hou, R., and Zhu, Y. (2023). Smart wound dressing for advanced wound management: real-time monitoring and on-demand treatment. *Mater. & Design* 229, 111917. doi:10.1016/j.matdes.2023.111917

Pilvenyte, G., Ratautaite, V., Boguzaite, R., Ramanavicius, S., Chen, C. F., Viter, R., et al. (2023). Molecularly imprinted polymer-based electrochemical sensors for the diagnosis of infectious diseases. *Biosensors* 13 (6), 620. doi:10.3390/bios13060620

Piro, B., and Reisberg, S. (2017). Recent advances in electrochemical immunosensors. Sensors (Basel) 17 (4), 794. doi:10.3390/s17040794

Pollard, T. D., Ong, J. J., Goyanes, A., Orlu, M., Gaisford, S., Elbadawi, M., et al. (2021). Electrochemical biosensors: a nexus for precision medicine. *Drug Discov. Today* 26 (1), 69–79. doi:10.1016/j.drudis.2020.10.021

Power, G., Moore, Z., and O'Connor, T. (2017). Measurement of pH, exudate composition and temperature in wound healing: a systematic review. *J. Wound Care* 26 (7), 381–397. doi:10.12968/jowc.2017.26.7.381

Prasad Mahindrakar, B., Goswami, A. G., Huda, F., Naithani, M., and Basu, S. (2023). Wound pH and surface temperature as a predictive biomarker of healing in diabetic foot ulcers. *Int. J. Low. Extrem Wounds* 23, 15347346231156962. doi:10.1177/ 15347346231156962

Rayment, E. A., Upton, Z., and Shooter, G. K. (2008). Increased matrix metalloproteinase-9 (MMP-9) activity observed in chronic wound fluid is related to the clinical severity of the ulcer. *Br. J. Dermatol* 158 (5), 951–961. doi:10.1111/j.1365-2133.2008.08462.x

Raziyeva, K., Kim, Y., Zharkinbekov, Z., Kassymbek, K., Jimi, S., and Saparov, A. (2021). Immunology of acute and chronic wound healing. *Biomolecules* 11 (5), 700. doi:10.3390/biom11050700

Romanelli, M., Miteva, M., Romanelli, P., Barbanera, S., and Dini, V. (2013). Use of diagnostics in wound management. *Curr. Opin. Support Palliat. Care* 7 (1), 106–110. doi:10.1097/SPC.0b013e32835dc0fc

Ronkainen, N. J., Halsall, H. B., and Heineman, W. R. (2010). Electrochemical biosensors. *Chem. Soc. Rev.* 39 (5), 1747–1763. doi:10.1039/b714449k

RoyChoudhury, S., Umasankar, Y., Jaller, J., Herskovitz, I., Mervis, J., Darwin, E., et al. (2018). Continuous monitoring of wound healing using a wearable enzymatic uric acid biosensor. *J. Electrochem. Soc.* 165 (8), B3168–B3175. doi:10.1149/2. 0231808jes

Sarp, S., Kuzlu, M., Wilson, E., Cali, U., and Guler, O. (2021). The enlightening role of explainable artificial intelligence in chronic wound classification. *Electronics* 10 (12), 1406. doi:10.3390/electronics10121406

Schilrreff, P. A., and Alexiev, U. (2022). Chronic inflammation in non-healing skin wounds and promising natural bioactive compounds treatment. *Int. J. Mol. Sci.* 23 (9), 4928. doi:10.3390/ijms23094928

Schwentker, A., and Billiar, T. R. (2003). Nitric oxide and wound repair. Surg. Clin. North Am. 83 (3), 521-530. doi:10.1016/S0039-6109(02)00207-4

Schwentker, A., Vodovotz, Y., Weller, R., and Billiar, T. R. (2002). Nitric oxide and wound repair: role of cytokines? *Nitric Oxide* 7 (1), 1–10. doi:10.1016/s1089-8603(02) 00002-2

Sempionatto, J. R., Lasalde-Ramírez, J. A., Mahato, K., Wang, J., and Gao, W. (2022). Wearable chemical sensors for biomarker discovery in the omics era. *Nat. Rev. Chem.* 6 (12), 899–915. doi:10.1038/s41570-022-00439-w

Sen, C. K. (2021). Human wound and its burden: updated 2020 compendium of estimates. Adv. Wound Care 10 (5), 281-292. doi:10.1089/wound.2021.0026

Shi, Z. H., Dai, C. B., Deng, P. X., Li, X., Wu, Y., Lv, J. J., et al. (2023). Wearable battery-free smart bandage with peptide functionalized biosensors based on MXene for bacterial wound infection detection. *Sensors Actuators B Chem. Sens* 23, 133598. doi:10. 1016/j.snb.2023.133598

Shirzaei Sani, E. A., Xu, C., Wang, C., Song, Y., Min, J., Tu, J., et al. (2023). A stretchable wireless wearable bioelectronic system for multiplexed monitoring and combination treatment of infected chronic wounds. *Sci. Adv.* 9 (12), eadf7388. doi:10. 1126/sciadv.adf7388

Simoska, O., Duay, J., and Stevenson, K. J. (2020). Electrochemical detection of multianalyte biomarkers in wound healing efficacy. *ACS Sens.* 5 (11), 3547–3557. doi:10. 1021/acssensors.0c01697

Sun, X., Zhang, Y., Ma, C., Yuan, Q., Wang, X., Wan, H., et al. (2021). A review of recent advances in flexible wearable sensors for wound detection based on optical and electrical sensing. *Biosensors* 12 (1), 10. doi:10.3390/bios12010010

Tang, N., Zheng, Y., Cui, D., and Haick, H. (2021). Multifunctional dressing for wound diagnosis and rehabilitation. *Adv. Healthc. Mater* 10 (22), e2101292. doi:10. 1002/adhm.202101292

Tarlton, J. F., Vickery Cj Leaper, D. J., Leaper Dj Bailey, A. J., and Bailey, A. J. (1997). Postsurgical wound progression monitored by temporal changes in the expression of matrix metalloproteinase-9. *Br. J. Dermatol* 137 (4), 506–516. doi:10.1111/j.1365-2133. 1997.tb03779.x

Tessarolo, M., Possanzini, L., Gualandi, I., Mariani, F., Torchia, L. D., Arcangeli, D., et al. (2021). Wireless textile moisture sensor for wound care. *Front. Phys.* 9. doi:10. 3389/fphy.2021.722173

Verdolino, D. V., Thomason, H. A., Fotticchia, A., and Cartmell, S. (2021). Wound dressings: curbing inflammation in chronic wound healing. *Emerg. Top. Life Sci.* 5 (4), 523–537. doi:10.1042/ETLS20200346

Vivcharenko, V., Trzaskowska, M., and Przekora, A. (2023). Wound dressing modifications for accelerated healing of infected wounds. *Int. J. Mol. Sci.* 24 (8), 7193. doi:10.3390/ijms24087193

Wang, L., Li, H., Shi, L., Li, L., Jia, F., Gao, T., et al. (2022). *In situ* peptide self-assembly on ionic nanochannel for dynamic monitoring of MMPs in extracellular matrix. *Biosens. Bioelectron.* 195, 113671. doi:10.1016/j.bios.2021.113671

Wang, X., Chen, X., Song, L., Zhou, R., and Luan, S. A. (2020). An enzyme-responsive and photoactivatable carbon-monoxide releasing molecule for bacterial infection theranostics. *J. Mater Chem. B* 8 (40), 9325–9334. doi:10.1039/d0tb01761b

Weller, C. (2009). "Interactive dressings and their role in moist wound management," in Advanced textiles for wound care (Berlin, Germany: Springer), 97-113.

Werner, S., and Grose, R. (2003). Regulation of wound healing by growth factors and cytokines. *Physiol. Rev.* 83 (3), 835–870. doi:10.1152/physrev.2003.83.3.835

Wu, M., Lu, Z., Wu, K., Nam, C., Zhang, L., and Guo, J. (2021). Recent advances in the development of nitric oxide-releasing biomaterials and their application potentials in chronic wound healing. *J. Mater Chem. B* 9 (35), 7063–7075. doi:10.1039/d1tb00847a

Xiong, Z. A., Achavananthadith, S., Lian, S., Madden, L., Ong, Z., Chua, W., et al. (2021). A wireless and battery-free wound infection sensor based on DNA hydrogel. *Sci. Adv.* 7 (47), eabj1617. doi:10.1126/sciadv.abj1617

Xu, G., Lu, Y., Cheng, C., Li, X., Xu, J., Liu, Z., et al. (2021). Battery-free and wireless smart wound dressing for wound infection monitoring and electrically controlled ondemand drug delivery. *Adv. Funct. Mat.* 31 (26). doi:10.1002/adfm.202100852

Xu, Y., Chen, H., Fang, Y., and Wu, J. (2022). Hydrogel combined with phototherapy in wound healing. *Adv. Healthc. Mater* 11 (16), e2200494. doi:10.1002/adhm.202200494

Yang, C., Yang, C., Chen, Y., Liu, J., Liu, Z., and Chen, H. J. (2023). The trends in wound management: sensing, therapeutic treatment, and "theranostics". J. Sci-Adv Mater Dev. 8 (4), 100619. doi:10.1016/j.jsamd.2023.100619

Yang, S. M., Kim, H., Ko, G.-J., Choe, J. C., Lee, J. H., Rajaram, K., et al. (2022). Soft, wireless electronic dressing system for wound analysis and biophysical therapy. *Nano Today* 47, 101685. doi:10.1016/j.nantod.2022.101685

Yao, G., Mo, X., Yin, C., Lou, W., Wang, Q., Huang, S., et al. (2022). A programmable and skin temperature-activated electromechanical synergistic dressing for effective wound healing. *Sci. Adv.* 8 (4), eabl8379. doi:10.1126/sciadv.abl8379

Zhang, Y., Lin, B., Huang, R., Lin, Z., Li, Y., Li, J., et al. (2021). Flexible integrated sensing platform for monitoring wound temperature and predicting infection. *Microb. Biotechnol.* 14 (4), 1566–1579. doi:10.1111/1751-7915.13821

Zhang, Z., Su, R., Han, F., Zheng, Z., Liu, Y., Zhou, X., et al. (2022). A soft intelligent dressing with pH and temperature sensors for early detection of wound infection. *RSC Adv.* 12 (6), 3243–3252. doi:10.1039/d1ra08375a

Zhao, R., Liang, H., Clarke, E., Jackson, C., and Xue, M. (2016). Inflammation in chronic wounds. Int. J. Mol. Sci. 17 (12), 2085. doi:10.3390/ijms17122085

Zhu, Y., Zhang, J., Song, J., Yang, J., Du, Z., Zhao, W., et al. (2019). A multifunctional pro-healing zwitterionic hydrogel for simultaneous optical monitoring of pH and glucose in diabetic wound treatment. *Adv. Funct. Mat.* 30 (6). doi:10.1002/adfm. 201905493