Check for updates

OPEN ACCESS

EDITED BY Marian Tsanov, Independent Researcher, Bochum, Germany

REVIEWED BY

Phillip R. Kramer, Texas A&M University, United States Juan Francisco Rodríguez-Landa, Universidad Veracruzana, Mexico Jose R. Eguibar, Meritorious Autonomous University of Puebla, Mexico

*CORRESPONDENCE

Amanda Ribeiro de Oliveira aroliveira@ufscar.br

SPECIALTY SECTION

This article was submitted to Learning and Memory, a section of the journal Frontiers in Behavioral Neuroscience

RECEIVED 31 August 2022 ACCEPTED 08 November 2022 PUBLISHED 28 November 2022

CITATION

Alves CO, Reimer AE and de Oliveira AR (2022) Involvement of D2-like dopaminergic receptors in contextual fear conditioning in female rats: influence of estrous cycle. Front. Behav. Neurosci. 16:1033649. [doi: 10.3389/fnbeh.2022.1033649](https://doi.org/10.3389/fnbeh.2022.1033649)

COPYRIGHT

© 2022 Alves, Reimer and de Oliveira. This is an open-access article distributed under the terms of the [Creative Commons Attribution License](http://creativecommons.org/licenses/by/4.0/)

[\(CC BY\).](http://creativecommons.org/licenses/by/4.0/) The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Involvement of D2-like dopaminergic receptors in contextual fear conditioning in [female rats: influence of estrous](https://www.frontiersin.org/articles/10.3389/fnbeh.2022.1033649/full) cycle

Camila de Oliveira Alves^{1,2}, Adriano Edgar Reimer^{1,2} and Amanda Ribeiro de Oliveira^{1,2*}

¹Department of Psychology, Center of Education and Human Sciences, Federal University of São Carlos (UFSCar), São Carlos, Brazil, ²Institute of Neuroscience and Behavior (INeC), Ribeirão Preto, Brazil

Introduction: Dopamine has been increasingly recognized as a key neurotransmitter regulating fear/anxiety states. Nevertheless, the influence of sex and estrous cycle differences on the role of dopamine in fear responses needs further investigation. We aimed to evaluate the effects of sulpiride (a dopaminergic D2-like receptor antagonist) on contextual fear conditioning in females while exploring the influence of the estrous cycle.

Methods: First, using a contextual fear conditioning paradigm, we assessed potential differences in acquisition, expression, and extinction of the conditioned freezing response in male and female (split in proestrus/estrus and metestrus/diestrus) Wistar rats. In a second cohort, we evaluated the effects of sulpiride (20 and 40 mg/kg) on contextual conditioned fear in females during proestrus/estrus and metestrus/diestrus. Potential nonspecific effects were assessed in motor activity assays (catalepsy and open-field tests).

Results: No sex differences nor estrous cycle effects on freezing behavior were observed during the fear conditioning phases. Sulpiride reduced freezing expression in female rats. Moreover, females during the proestrus/estrus phases of the estrous cycle were more sensitive to the effects of sulpiride than females in metestrus/diestrus. Sulpiride did not cause motor impairments.

Discussion: Although no sex or estrous cycle differences were observed in basal conditioned fear expression and extinction, the estrous cycle seems to influence the effects of D2-like antagonists on contextual fear conditioning.

KEYWORDS

conditioned fear, freezing, sulpiride, dopamine, proestrus/estrus, metestrus/diestrus, mesolimbic

Introduction

Anxiety-related disorders are complex heterogeneous mental disorders and important causes of health-related burden worldwide [\(American Psychiatric Association,](#page-9-0) [2013\)](#page-9-0). A frequent observation is that the prevalence of these disorders is substantially higher in women than in men [\(McLean et al.,](#page-11-0) [2011;](#page-11-0) [Bandelow and Michaelis,](#page-9-1) [2015;](#page-9-1) [Craske et al.,](#page-9-2) [2017\)](#page-9-2). While several biological processes are thought to contribute to sexual dimorphism in anxiety disorders, past and recent evidence suggests that sex steroids, as well as their fluctuations in brain systems, are key in mediating these pathological states [\(Maeng](#page-10-0) [and Milad,](#page-10-0) [2015;](#page-10-0) [Li and Graham,](#page-10-1) [2017;](#page-10-1) [Nouri et al.,](#page-11-1) [2022\)](#page-11-1).

Likewise, animal model studies have revealed that females respond to stress and/or fear differently from males [\(Johnston](#page-10-2) [and File,](#page-10-2) [1991;](#page-10-2) [Imhof et al.,](#page-10-3) [1993;](#page-10-3) [Scholl et al.,](#page-11-2) [2019;](#page-11-2) [Knight](#page-10-4) [et al.,](#page-10-4) [2021\)](#page-10-4) and dependent on circulating levels of sex steroids [\(Frye et al.,](#page-10-5) [2000;](#page-10-5) [Marcondes et al.,](#page-10-6) [2001;](#page-10-6) [Gouveia](#page-10-7) [et al.,](#page-10-7) [2004;](#page-10-7) [Lovick,](#page-10-8) [2014\)](#page-10-8). Therefore, sex steroids may play a role in modulating neurotransmitter systems involved in anxiety-related pathological states. Nevertheless, most studies with animal models are carried out predominantly with males, and their results are generalized to females [\(Bangasser and Cuarenta,](#page-9-3) [2021;](#page-9-3) [Shansky and Murphy,](#page-11-3) [2021\)](#page-11-3). Thus, potential differences between the sexes remain largely unexplored.

Aversive conditioning is one of the most used behavioral paradigms to study fear in rodents [\(LeDoux,](#page-10-9) [2014;](#page-10-9) [Fanselow](#page-10-10) [and Wassum,](#page-10-10) [2015;](#page-10-10) [Izquierdo et al.,](#page-10-11) [2016;](#page-10-11) [Haaker et al.,](#page-10-12) [2019\)](#page-10-12). Regarding sex differences in fear conditioning, conflicting results have been reported. Several studies have shown no sex differences in fear conditioning [\(Milad et al.,](#page-11-4) [2009a;](#page-11-4) [Cossio et al.,](#page-9-4) [2016;](#page-9-4) [Machado Figueiredo et al.,](#page-10-13) [2019;](#page-10-13) [Carvalho et al.,](#page-9-5) [2021\)](#page-9-5). In contrast, higher conditioned freezing in male compared to female rats [\(Graham et al.,](#page-10-14) [2009;](#page-10-14) [Daviu et al.,](#page-9-6) [2014;](#page-9-6) [Urien](#page-11-5) [and Bauer,](#page-11-5) [2022\)](#page-11-5) and increased freezing in females [\(Baran](#page-9-7) [et al.,](#page-9-7) [2009;](#page-9-7) [Blume et al.,](#page-9-8) [2017\)](#page-9-8) have also been found. When considering the estrous cycle, some studies have indicated low estrogen phases in females to be associated with heightened sensitivity to stress and impaired consolidation of extinction memory [\(Markus and Zecevic,](#page-10-15) [1997;](#page-10-15) [Milad et al.,](#page-11-4) [2009a;](#page-11-4) [Zeidan](#page-11-6) [et al.,](#page-11-6) [2011\)](#page-11-6). Others, however, have observed no effects of the estrous cycle on fear conditioning [\(Cossio et al.,](#page-9-4) [2016;](#page-9-4) [Machado Figueiredo et al.,](#page-10-13) [2019;](#page-10-13) [Carvalho et al.,](#page-9-5) [2021\)](#page-9-5). In fact, we found no sex differences or estrous cycle influence on tone-cued fear conditioning, but female rats displayed lower extinction retention when compared to males in contextual fear conditioning independently of the estrous cycle phase [\(Reimer et al.,](#page-11-7) [2018\)](#page-11-7).

Several lines of evidence have indicated that dopamine is a key neurotransmitter in fear conditioning [\(Pezze and Feldon,](#page-11-8) [2004;](#page-11-8) [De la Mora et al.,](#page-9-9) [2010;](#page-9-9) [Lee et al.,](#page-10-16) [2017;](#page-10-16) [Stubbendorff and](#page-11-9) [Stevenson,](#page-11-9) [2021\)](#page-11-9). Our group has demonstrated an important involvement of signaling at D2-like dopaminergic receptors in the expression of conditioned fear in rats [\(De Oliveira](#page-10-17) [et al.,](#page-10-17) [2006,](#page-10-17) [2009;](#page-10-18) [De Souza Caetano et al.,](#page-10-19) [2013;](#page-10-19) [De Vita](#page-10-20) [et al.,](#page-10-20) [2021\)](#page-10-20). Additionally, the fear response to conditioned stimuli appeared to depend on the activation of the ventral tegmental area–basolateral amygdala dopaminergic connections [\(De Oliveira et al.,](#page-10-21) [2011,](#page-10-21) [2013,](#page-10-22) [2014,](#page-10-23) [2017;](#page-10-24) [De Souza Caetano](#page-10-19) [et al.,](#page-10-19) [2013\)](#page-10-19). Other studies, on the other hand, have pointed to the importance of D2-like receptors in the nucleus accumbens and infralimbic cortex for fear extinction [\(Holtzman-Assif et al.,](#page-10-25) [2010;](#page-10-25) [Mueller et al.,](#page-11-10) [2010;](#page-11-10) [Zbukvic et al.,](#page-11-11) [2017\)](#page-11-11). All these studies, however, have overlooked the involvement of dopaminergic mechanisms in conditioned fear in females.

Since there are sex and estrous cycle-dependent fluctuations in dopamine concentration and dopaminergic receptor expression in fear conditioning-relevant brain regions [\(Xiao and](#page-11-12) [Becker,](#page-11-12) [1994;](#page-11-12) [Staiti et al.,](#page-11-13) [2011;](#page-11-13) [Locklear et al.,](#page-10-26) [2016;](#page-10-26) [Kokras](#page-10-27) [et al.,](#page-10-27) [2018;](#page-10-27) [Cullity et al.,](#page-9-10) [2019\)](#page-9-10), the present study aimed to explore the involvement of dopamine in conditioned fear in females. For this, we evaluated the effects of blocking D2-like dopaminergic receptors on the expression and extinction of contextual conditioned fear in female rats at different phases of the estrous cycle.

Materials and methods

Animals

Twenty-four male (310–440 g) and 126 female (220–310 g) Wistar rats (8–12 weeks old) from the animal facility of the Federal University of São Carlos (UFSCar) were used. Rats were housed in groups of four per cage (polypropylene boxes, $40 \times 33 \times 26$ cm), under a 12/12 h dark/light cycle (lights on at 07:00 h), at 23 \pm 2°C, and given *ad libitum* access to water and rat chow. Experiments were carried out during the light phase of the cycle, between 9:00 and 12:00 h. All procedures were performed in accordance with the National Council for Animal Experimentation Control and were approved by the Committee for Animal Care and Use of the Federal University of São Carlos (Protocol No. 5717160919).

Assessment of the estrous cycle phase

To assess female rats' estrous cycle, vaginal smears were taken daily in the morning (09:00 h), starting 10 days before the beginning of the behavioral assays. Vaginal epithelial cell samples were obtained with an inoculation loop, which was initially sterilized in a flame, dipped in 0.9% saline, and then gently inserted into the rat's vagina. Samples were then smeared onto a microscope glass slide and stained with a staining set (Panótico Rápido, Laborclin, PR, Brazil). After

drying, slides were examined under a light microscope. Each phase was identified based on the proportion of epithelial cells, cornified cells, and leukocytes observed in the vaginal smear [\(Cora et al.,](#page-9-11) [2015;](#page-9-11) [Maeng et al.,](#page-10-28) [2015\)](#page-10-28). Proestrus is characterized by the presence of nucleated cells; in estrus, there is a higher prevalence of cornified, non-nucleated cells; metestrus has equivalent portions of leukocytes, cornified cells, and nucleated epithelial cells; diestrus is characterized by the predominance of leukocytes. The experiments were performed on animals that had completed at least two regular cycles. Rats were grouped by follicular (proestrus/estrus) or luteal (metestrus/diestrus) phases [\(Reimer et al.,](#page-11-7) [2018\)](#page-11-7), based on the test day of the conditioned fear protocol or the day of the motor evaluation for the catalepsy and open field tests. Sex hormone concentrations were not measured in the present study.

Drugs

The dopaminergic D2-like antagonist sulpiride (Tocris Bioscience, Bristol, UK) was first mixed in 2% Tween 80 and then dissolved in physiological saline (0.9%) to obtain concentrations of 20 and 40 mg/ml. Physiological saline containing 2% Tween 80 served as vehicle control. Injections were administered in a constant volume of 1 ml/kg, intraperitoneally (i.p.), 15 min before the beginning of the tests. The doses (20 and 40 mg/kg) and injection times were based on previous studies [\(De Oliveira et al.,](#page-10-17) [2006;](#page-10-17) [Carvalho et al.,](#page-9-12) [2009;](#page-9-12) [De Souza Caetano et al.,](#page-10-19) [2013;](#page-10-19) [De Vita et al.,](#page-10-20) [2021\)](#page-10-20). The investigator was blind to the treatment condition of each rat; drugs were prepared and stored in vials labeled with codes by a different investigator than the one who performed the behavioral experiments and data analysis.

Contextual fear conditioning

The experimental protocol for contextual fear conditioning was based on [Reimer et al.](#page-11-7) [\(2018\)](#page-11-7) and [De Vita et al.](#page-10-20) [\(2021\)](#page-10-20). Contextual fear conditioning procedures took place in standard conditioning chambers (Insight, SP, Brazil). Chamber-A (dimensions: $26 \times 20 \times 20$ cm) was made with white metal walls and ceiling, and a rod floor (5 mm in diameter, spaced 1.5 cm apart) connected to a shock generator (Eltrones, SC, Brazil). Chamber-B ($32 \times 30 \times 30$ cm) consisted of stainless steel sides with a clear Plexiglas front, back, and top, with an opaque white polypropylene floor. Conditioning chambers were enclosed in sound-attenuation boxes (66 \times 43 \times 45 cm; Grason-Stadler, MN, USA). Chamber-A was equipped with a fan providing constant background noise. Alcohol (20%) was used as a mild scent in Chamber-A; acetic acid (2%) odor was added to ChamberB. Animals' behavior was recorded with digital video cameras positioned in front of the chambers. Videos were evaluated by a blind rater after the tests were completed. Freezing behavior was scored as an index of the contextual fear conditioning and it was operationally defined as the total absence of movements, except those required for respiration, for at least 6 s per episode.

Catalepsy bar test

The apparatus and testing procedures used for the catalepsy test have been described in detail elsewhere [\(Colombo et al.,](#page-9-13) [2013;](#page-9-13) [Barroca et al.,](#page-9-14) [2019;](#page-9-14) [Waku et al.,](#page-11-14) [2022\)](#page-11-14). Briefly, a horizontal acrylic bar (30 cm in length and 1 cm in diameter) was positioned 8 cm above the floor of a standard polypropylene box (40 \times 33 \times 26 cm). The animal's forepaws were carefully positioned on the bar, while their hind paws remained on the ground. The latency to step down from the horizontal bar was measured 15 and 45 min after drug administration.

Open-field test

During the interval between the catalepsy bar tests, the animals underwent the open-field test. The experimental protocol for the open-field test was based on [Reimer et al.](#page-11-15) [\(2012\)](#page-11-15) and [De Vita et al.](#page-10-20) [\(2021\)](#page-10-20). Locomotor activity was assessed in a circular arena consisting of a transparent acrylic enclosure (60 cm in diameter, 50 cm in height, floor divided into 12 sections). Rats were placed in the middle of the arena and left for a 15-min period of free exploration. The session was recorded by a camera positioned above the openfield. At the end of the test, the total number of crossings (number of floor sections crossed) and rearings (standing with forepaws raised in the middle of the arena or against the walls) were analyzed.

Experiment 1: contextual fear as a function of sex and estrous cycle in female rats

Initially, we aimed at validating our protocol while exploring the influence of sex and estrous cycle on contextual fear conditioning (**[Figure 1A](#page-4-0)**). For that, rats were divided into two groups: Same Context and Different Context groups. Both groups underwent training in Chamber-A. During the training session, animals were placed individually into the experimental chamber and, after a habituation period of 5 min, 10 1-s unsignaled footshocks (0.6 mA, inter-trial interval 30–90 s) were presented. Each animal was removed 2 min after the

Alves et al. [10.3389/fnbeh.2022.1033649](https://doi.org/10.3389/fnbeh.2022.1033649)

last footshock and returned to its home cage. The training session lasted approximately 15 min. After 24 h, animals were submitted to the test session, without shock presentation; the Same Context group was reexposed to Chamber-A while the Different Context group was placed into Chamber-B. Animals remained in the behavioral chambers for 10 min. The retest session was performed 24 h after the test session. Animals were again re-exposed to the same chambers used during the test session for a total of 10 min.

Experiment 2: involvement of D2-like dopaminergic receptors in the expression and extinction of conditioned fear in female rats

Experiment 2's fear conditioning protocol followed the same procedures described for Experiment 1, in which rats were submitted to three consecutive sessions (training, test, and retest) spaced by 24 h each, but no different context group was used this time (**[Figure 2A](#page-5-0)**). Briefly, animals were submitted to training in Chamber-A for aversive conditioning. On the second day (test session), animals received i.p. injections of vehicle or drug treatment and, after 15 min, they were reexposed to the Chamber-A for 10 min. Twenty-four hours after the test, rats were once again placed in Chamber-A for 10 min (retest session), for evaluation of extinction recall. Potential nonspecific effects of sulpiride on motor behavior were evaluated in the same animals. For this, 48 h after the end of the fear conditioning retest session, animals were subjected to the open-field and catalepsy bar tests (**[Figure 3A](#page-6-0)**).

Analysis of results

All analyses were conducted using R version 4.0.5 [\(R](#page-11-16) [Core Team,](#page-11-16) [2020\)](#page-11-16) and Rstudio version 1.4.1717 [\(RStudio](#page-11-17) [Team,](#page-11-17) [2020\)](#page-11-17). Prior to model fitting, we used the fitdistrplus package version 1.1-8 [\(Delignette-Muller and Dutang,](#page-10-29) [2015\)](#page-10-29) to assess the distribution of each response variable by plotting a Cullen-Frey diagram with 1,000 bootstraps. Additionally, using the fitdistrplus's fitdist plot function, we drew four classical goodness-of-fit plots (i.e., density plot, CDF plot, Q-Q plot, and P-P) to compare the fit of other candidate distributions that are often used in fear conditioning paradigm and motor function assessment analyses. Based on this, for the fear conditioning experiments, beta regression analysis was selected because it was a more appropriate fit to the data distribution than the commonly used repeated measures ANOVA. Behavioral scores were rescaled to between 0 and 1 using a min-max normalization. Generalized linear mixed models (GLMMs) with logit link function and beta distribution were used. For Experiment 1, rats' identities were used as

random factors while Group (Same-Context and Different-Context), Session (Test and Retest), and Sex (Male and Female) or Phase (Proestrus/Estrus and Metestrus/Diestrus) were used as fixed factors. For Experiment 2, rats' identities were used as random factors, Phase (Proestrus/Estrus and Metestrus/Diestrus), Session (Test and Retest), and Treatment (Saline, Sulpiride-20, Sulpiride-40) were used as fixed factors. For the assessment of motor function, generalized linear models (GLMs) with log link function and negative binomial distribution were used to evaluate the frequency of crossings and rearings. Phase (Proestrus/Estrus and Metestrus/Diestrus) and Treatment (Saline, Sulpiride-20, Sulpiride-40) were used as fixed factors. GLMMs with logit link function and beta distributions were used to evaluate step-down latency in the catalepsy test. Rats' identities were used as random factors, Phase (Proestrus/Estrus and Metestrus/Diestrus), Time (15 and 45-min), and Treatment (Saline, Sulpiride-20, Sulpiride-40) were used as fixed factors. Model fitting was performed with the glmmTMB package version 1.1.3 [\(Brooks et al.,](#page-9-15) [2017\)](#page-9-15). Likelihood ratio tests were conducted using car package version 3.1-0 [\(Fox and Weisberg,](#page-10-30) [2019\)](#page-10-30). Planned a priori pairwise comparisons were tested using the emmeans package 1.8.1-1 [\(Lenth,](#page-10-31) [2021\)](#page-10-31) and Holm's sequential Bonferroni procedure was used to adjust the significance level due to the multiple comparisons.

Results

Experiment 1: contextual fear as a function of sex and estrous cycle in female rats

Male and female rats submitted to the training and testing procedures in the same cage (Same Context) spent more time freezing than rats submitted to the test session in a cage different from the one used for training (Different Context; **[Figures 1B,C](#page-4-0)**; **[Supplementary Figure 1](#page-9-16)**). An analysis of variance based on mixed beta regression indicated statistically significant effects for Group (Different \times Same-Context: χ^2 = 35.57, $p \, < \, 0.05$), Session (Test × Retest: $\chi^2 = 19.27, p \, < \, 0.05$), and Sex (Male \times Female: χ^2 = 7.58, p < 0.05). Pairwise comparisons using t-tests, corrected with Holm's sequential Bonferroni procedure, indicated that the same-context group froze more than the different-context at the test session in both males ($t = -2.66$, $p < 0.05$) and females ($t = -7.52$, $p < 0.05$). During the retest, the same-context continued to freeze more than the different-context group in females ($t = -5.96, p < 0.05$), but not in males ($t = 0.17$, $p > 0.05$). Decreased freezing during retest was observed for same-context males ($t = -6.11, p < 0.05$) and females ($t = -8.07$, $p < 0.05$). Males and females did not differ in the freezing response during the test session either

for the same-context ($t = -1.86$, $p > 0.05$) or different-context groups ($t = 1.27$, $p > 0.05$). During the retest, females of the different-context group froze less than males ($t = 2.75$, $p < 0.05$), but males and females of the same-context groups did not differ from each other ($t = -2.43$, $p > 0.05$). No significant difference between sexes was observed for the fear extinction index (FEI; Male × Female: $\chi^2 = 0.22, p > 0.05$).

There was no difference in freezing between phases of the estrous cycle for the same-context or different context groups (**[Figures 1D,E](#page-4-0)**). An analysis of variance based on mixed beta regression indicated statistically significant effects for Group (Different \times Same-Context: $\chi^2 = 17.10, p < 0.05$), Session (Test \times Retest: χ^2 = 10.76, p < 0.05), and Cycle-Phase (Proestrus/Estrus \times Metestrus/Diestrus: χ^2 = 4.57,

 $p < 0.05$). Pairwise comparisons using t-tests, corrected with Holm's sequential Bonferroni procedure, indicated that the same-context group froze more than the different-context at the test and retest sessions in both proestrus/estrus (test: $t = -6.12, p < 0.05$; retest: $t = -4.87, p < 0.05$) and metestrus/diestrus groups (test: $t = -5.23$, $p < 0.05$; retest: $t = -4.14$, $p < 0.05$). Decreased freezing during retest was observed for same-context proestrus/estrus ($t = -6.28$, $p < 0.05$) and metestrus/diestrus females ($t = -5.46$, $p < 0.05$). Proestrus/estrus and metestrus/diestrus females did not differ in the freezing response during the test or retest sessions either for the same-context (test: $t = 0.95$, $p > 0.05$; retest: $t = 1.26$, $p >$ 0.05) or different-context groups (test: $t = 1.97$, $p > 0.05$; retest: $t = 2.14$, $p > 0.05$). No significant difference between phases of

the estrous cycle was observed for the fear extinction index (FEI; Proestrus/Estrus × Metestrus/Diestrus: $\chi^2 = 0.20$, $p > 0.05$).

Experiment 2: involvement of D2-like dopaminergic receptors in the expression and extinction of conditioned fear in female rats

Sulpiride decreased contextual conditioned freezing response in female rats (**[Figure 2](#page-5-0)**; **[Supplementary Figure 2](#page-9-16)**). An analysis of variance based on mixed beta regression indicated statistically significant effects for Treatment (Saline \times Sulpiride-20 \times Sulpiride-40: χ^2 = 13.30, p < 0.05), Session (Test \times Retest: χ^2 = 84.61, p < 0.05), but not for Cycle-Phase (Proestrus/Estrus × Metestrus/Diestrus: $\chi^2 = 2.97$, $p >$ 0.05). Pairwise comparisons using t-tests, corrected with Holm's sequential Bonferroni procedure, indicated decreased freezing response for sul20 and sul40 compared to the control group during the test session ($t = 3.42$, $p < 0.05$ and $t = 3.50$, $p < 0.05$, respectively). Freezing response did not differ for sul20 and sul40 compared to the control group during the retest session

 $(t = 2.65, p > 0.05 \text{ and } t = 1.62, p > 0.05, respectively)$. Pairwise comparisons also indicated decreased freezing during retest for control (t = -7.00 , $p < 0.05$), sul20 (t = -5.29 , $p < 0.05$), and sul40 ($t = -3.91$, $p < 0.05$) compared to the test session.

When considering the estrous cycle phases, pairwise comparisons using t-tests, corrected with Holm's sequential Bonferroni procedure, indicated decreased freezing response for sul40 in proestrus/estrus compared to the control group during the test session ($t = 2.92$, $p < 0.05$), but not for sul20 in proestrus/estrus ($t = 2.32$, $p > 0.05$) or sul20 and sul40 in metestrus/diestrus ($t = 2.51$, $p > 0.05$ and $t = 2.04$, $p > 0.05$, respectively). Freezing response did not differ for sul20 and sul40 compared to the control group during the retest session for proestrus/estrus (sul20: $t = 2.66$, $p > 0.05$; sul40: $t = 1.95$, $p > 0.05$) or metestrus/diestrus (sul20: $t = 1.08$, $p > 0.05$; sul40: $t = 0.33, p > 0.05$). Pairwise comparisons also indicated decreased freezing during retest compared to the test session for control in proestrus/estrus ($t = -5.10, p < 0.05$) and metestrus/diestrus $(t = -4.83, p < 0.05)$, and for sul20 and sul40 in proestrus/estrus $(t = -4.82, p < 0.05 \text{ and } t = -3.19, p < 0.05, \text{ respectively}),$ but not for sul20 and sul40 in metestrus/diestrus ($t = -2.55$, $p > 0.05$ and $t = -2.30$, $p > 0.05$, respectively). No significant

differences were observed for the fear extinction index between sul20 or sul40 and the control group during proestrus/estrus $(t = 0.46, p > 0.05; t = -1.40, p > 0.05, respectively)$ or metestrus/diestrus (t = -2.48, $p > 0.05$; t = -1.15, $p > 0.05$, respectively).

Sulpiride did not affect motor performance evaluated with the open-field and catalepsy tests (**[Figure 3](#page-6-0)**). For the open-field test, analysis of variance based on mixed negative binomial regression revealed no statistically significant for crossings (Treatment—Saline \times Sulpiride-20 \times Sulpiride-40: χ^2 = 3.15, $p > 0.05$; Cycle-Phase—Proestrus/Estrus × Metestrus/Diestrus: $χ²$ 0.08, $p > 0.05$ or rearings (Treatment—Saline \times Sulpiride-20 \times Sulpiride-40: χ^2 = 0.78, $p > 0.05$; Cycle-Phase—Proestrus/Estrus × Metestrus/Diestrus:

 χ^2 = 0.34, $p > 0.05$). Pairwise comparisons using t-tests, corrected with Holm's sequential Bonferroni procedure, indicated no differences for crossings between sul20 or sul40 and the control group during proestrus/estrus ($t = 0.66$, $p > 0.05$; $t = 0.04$, $p > 0.05$, respectively) or metestrus/diestrus ($t = 0.84$, p) > 0.05 ; $t = -1.18$, $p > 0.05$, respectively). Pairwise comparisons also indicated no differences for rearings between sul20 or sul40 and the control group during proestrus/estrus ($t = 1.17$, $p > 0.05$; $t = 0.76$, $p > 0.05$, respectively) or metestrus/diestrus $(t = -0.62, p > 0.05; t = -1.64, p > 0.05, respectively)$. For the catalepsy test, an analysis of variance based on mixed beta regression indicated statistically significant effects for Time (15 \times 45 min: χ^2 = 7.80, p < 0.05), but not for Treatment (Saline \times Sulpiride-20 \times Sulpiride-40: $\chi^2 = 1.30, p > 0.05$) or

Cycle-Phase (Proestrus/Estrus × Metestrus/Diestrus: χ^2 = 0.23, $p > 0.05$). Pairwise comparisons using t-tests, corrected with Holm's sequential Bonferroni procedure, indicated no differences between sul20 and the control group during proestrus/estrus (15-min: $t = 1.16$, $p > 0.05$; 45-min: $t = 0.33$, $p > 0.05$) or metestrus/diestrus (15-min: $t = 0.16, p > 0.05$; 45-min: $t = 1.17$, $p > 0.05$). Pairwise comparisons also indicated no differences between sul40 and the control group during proestrus/estrus (15-min: $t = -1.09$, $p > 0.05$; 45-min: $t = -1.05$, $p > 0.05$) or metestrus/diestrus (15-min: $t = 0.72$, $p > 0.05$; 45-min: $t = -1.56, p > 0.05$).

Discussion

The present study aimed to evaluate the involvement of dopamine D2-like receptors on contextual fear conditioning in female rats. Blockade of D2-like receptors significantly decreased the expression of freezing response in female rats. Moreover, females in proestrus/estrus phases were more sensitive to the effects of sulpiride than females in metestrus/diestrus. These findings suggest a functional interplay between the estrous cycle phase and dopaminergic neurotransmission in fear conditioning. The effect of blocking D2-like receptors was specific to the fear response, with no motor impairment observed during the different phases of the estrous cycle.

The protocol used for acquisition, expression, and extinction of contextual conditioned fear was effective for both males and females. This result is in line with other studies from our group obtained using a similar protocol in male rats only [\(De](#page-10-19) [Souza Caetano et al.,](#page-10-19) [2013;](#page-10-19) [De Vita et al.,](#page-10-20) [2021\)](#page-10-20) and extends the validity of our protocol to female rats. We did not verify differences between the sexes or estrous cycle phases for the expression and extinction of conditioned freezing, although same-context females continue to freeze more in the retest than different-context females. No differences were observed in the fear extinction index. Literature has been reporting mixed results regarding sex/estrous cycle differences in fear conditioning paradigms. Previous studies have reported either higher freezing expression in male rats [\(Graham et al.,](#page-10-14) [2009;](#page-10-14) [Daviu et al.,](#page-9-6) [2014;](#page-9-6) [Urien and Bauer,](#page-11-5) [2022\)](#page-11-5), female rats [\(Baran](#page-9-7) [et al.,](#page-9-7) [2009;](#page-9-7) [Blume et al.,](#page-9-8) [2017\)](#page-9-8), or no difference [\(Cossio](#page-9-4) [et al.,](#page-9-4) [2016;](#page-9-4) [Zhao et al.,](#page-11-18) [2018;](#page-11-18) [Machado Figueiredo et al.,](#page-10-13) [2019;](#page-10-13) [Carvalho et al.,](#page-9-5) [2021\)](#page-9-5). There may be several explanations for this, particularly regarding differences in methods, subject strain, determination of gonadal hormonal status, and fear conditioning protocol (e.g., cued or contextual CS) used in those studies.

These potential inconsistencies are raising increasingly important questions regarding the participation of different neurotransmitter/neuromodulator/neurosteroid systems influencing aversive learning and memory. Several pieces of

evidence point to different mechanisms by which fear processing could diverge in male and female subjects. Neuronal activity in the basolateral amygdala shows faster neuron firing rates and greater excitatory synaptic input in females [\(Blume et al.,](#page-9-8) [2017\)](#page-9-8). Different activation patterns in the hippocampus, amygdala, and bed nucleus of the stria terminalis during fear conditioning paradigms are found between male and female rats [\(Maren](#page-10-32) [et al.,](#page-10-32) [1994;](#page-10-32) [Blume et al.,](#page-9-8) [2017;](#page-9-8) [Keiser et al.,](#page-10-33) [2017;](#page-10-33) [Urien and](#page-11-5) [Bauer,](#page-11-5) [2022\)](#page-11-5). Also, the distinct participation of neurotransmitter systems regulating fear response, particularly endocannabinoid [\(Morena et al.,](#page-11-19) [2021\)](#page-11-19), serotonergic and dopaminergic systems [\(Mitsushima et al.,](#page-11-20) [2006;](#page-11-20) [Rey et al.,](#page-11-21) [2014\)](#page-11-21) have been described.

Previous works from our group have demonstrated that the blockade of dopamine D2-like receptors with sulpiride impairs fear expression in male rats [\(De Oliveira et al.,](#page-10-21) [2011,](#page-10-21) [2013,](#page-10-22) [2017;](#page-10-24) [De Souza Caetano et al.,](#page-10-19) [2013;](#page-10-19) [De Vita et al.,](#page-10-20) [2021\)](#page-10-20). Here, sulpiride significantly reduced freezing response in female rats in a similar fashion. Some studies reported that systemically administration of the D2-like antagonists' haloperidol and raclopride to males have, however, increased freezing behavior [\(Holtzman-Assif et al.,](#page-10-25) [2010;](#page-10-25) [Mueller et al.,](#page-11-10) [2010\)](#page-11-10). These studies did not assess the motor side effects of these drugs or could not dissociate the effects on freezing from impaired motor behavior. Sulpiride's effect on the freezing response observed in females in the present study cannot be attributed to nonspecific motor effects, as it neither affected animals' performance in catalepsy nor open-field tests. Likewise, we did not observe motor effects in males using similar doses [\(De Vita et al.,](#page-10-20) [2021\)](#page-10-20). Sulpiride's effect on conditioned fear without adversely affecting motor function suggests a stronger action of sulpiride on the mesolimbic than in the nigrostriatal dopaminergic pathway. In this direction, the atypical neuroleptic amisulpride, which bears a close chemical and clinical resemblance to sulpiride, showed greater selectivity for limbic than striatal dopaminergic projections [\(Schoemaker et al.,](#page-11-22) [1997\)](#page-11-22).

In addition to demonstrating that sulpiride significantly reduced freezing expression in female rats, we found that the females' responsiveness to sulpiride appears to be dependent on the estrous cycle phase. A significant reduction in the expression of contextual conditioned freezing behavior after sulpiride administration was observed in females in proestrus/estrus, but not in metestrus/diestrus. Sulpiride's lack of effect in metestrus/diestrus suggests that the brain's hormonal profile during proestrus/estrus favors the action of the D2-like antagonist. We also observed estrous cycledependent effects on contextual fear in a previous study in which deficits induced by meta-chlorophenylpiperazine (mCPP)—a serotonergic agonist—prevailed in metestrus/diestrus but not proestrus/estrus or in males, suggesting that fluctuations in sex hormones may also influence the action of serotonergic drugs [\(Reimer et al.,](#page-11-7) [2018\)](#page-11-7).

Hormonal fluctuations throughout the cycle could lead to differences in the response to pharmacological manipulations. It is possible, for example, that susceptibility to the effects of sulpiride may result from differences in dopamine availability or the expression of D2-like receptors [\(Xiao and Becker,](#page-11-12) [1994;](#page-11-12) [Thompson and Moss,](#page-11-23) [1997;](#page-11-23) [Le Saux et al.,](#page-10-34) [2006;](#page-10-34) [Diekhof and](#page-10-35) [Ratnayake,](#page-10-35) [2016;](#page-10-35) [Petersen et al.,](#page-11-24) [2021\)](#page-11-24). Our findings suggest that D2-like receptors play an important role in the expression of conditioned fear in females in proestrus/estrus in a manner similar to that observed in previous studies with males [\(De](#page-10-17) [Oliveira et al.,](#page-10-17) [2006,](#page-10-17) [2013;](#page-10-22) [De Souza Caetano et al.,](#page-10-19) [2013;](#page-10-19) [De](#page-10-20) [Vita et al.,](#page-10-20) [2021\)](#page-10-20). The fact that female rats tested during higher hormone levels respond to the pharmacological manipulation similarly to males might be attributed to the role of estradiol, in both sexes [\(Lebron-Milad and Milad,](#page-10-36) [2012;](#page-10-36) [Graham and](#page-10-37) [Milad,](#page-10-37) [2013,](#page-10-37) [2014\)](#page-10-38). Although it seems counterintuitive given that estradiol levels in male rats would be comparable to those found in low-estradiol females, many effects of testosterone actually seem to be mediated by estradiol [\(Purves-Tyson](#page-11-25) [et al.,](#page-11-25) [2007;](#page-11-25) [Hammes and Levin,](#page-10-39) [2019;](#page-10-39) [Zuloaga et al.,](#page-11-26) [2020\)](#page-11-26). Estradiol in males can be synthesized via aromatization of circulating testosterone as well as from testosterone produced de novo by neurons and glia [\(Gillies and McArthur,](#page-10-40) [2010\)](#page-10-40). This would explain why sex differences arose in females tested in metestrus/diestrus. It is important to highlight, however, that hormones were not measured in the present study.

In male rats, besides recently confirming the involvement of D2-like receptor-mediated mechanisms in the expression of conditioned fear, our previous results have suggested that D2-like receptors are not involved in conditioned freezing extinction [\(De Vita et al.,](#page-10-20) [2021\)](#page-10-20). In the present study, sulpiride administered to females in proestrus/estrus during the test session did not affect the freezing behavior exhibited 24 h later, during the retest session. When administered to metestrus/diestrus females during the test session, sulpiride did not alter freezing in the retest when compared to the vehicle control group, but it prevented the expected decrease in freezing in the retest compared to the test session. The analysis of the fear extinction index, however, did not indicate an effect of sulpiride on extinction. The effects of systemically administered drugs acting at D2-like receptors on contextual fear extinction are, in fact, not well understood [\(Stubbendorff](#page-11-9) [and Stevenson,](#page-11-9) [2021\)](#page-11-9). In studies using cued-fear conditioning, systemic haloperidol and raclopride administration have been shown to decrease extinction recall [\(Holtzman-Assif et al.,](#page-10-25) [2010;](#page-10-25) [Mueller et al.,](#page-11-10) [2010\)](#page-11-10). Systemic injections of sulpiride in male mice have been shown to facilitate the extinction of cued fear conditioning only in a protocol that did not promote extinction [\(Ponnusamy et al.,](#page-11-27) [2005\)](#page-11-27). Altogether, data point to a modulatory rather than an essential role for D2-like receptors in fear extinction.

Although our results implicate dopaminergic D2-like receptors in conditioned fear in females, the role of specific dopaminergic areas remains to be demonstrated. The present results agree with those reported by others showing decreased conditioned freezing in males with sulpiride locally administered to the amygdala and prefrontal cortex [\(De Oliveira et al.,](#page-10-21) [2011,](#page-10-21) [2017;](#page-10-24) [De Souza Caetano et al.,](#page-10-19) [2013;](#page-10-19) [Dadkhah et al.,](#page-9-17) [2021\)](#page-9-17). In addition, exposure to aversive conditioned stimuli has been shown to increase dopamine levels in the basolateral amygdala, nucleus accumbens, and prefrontal cortex in male rats [\(Pezze](#page-11-28) [et al.,](#page-11-28) [2001;](#page-11-28) [Matsumoto et al.,](#page-11-29) [2005;](#page-11-29) [Martinez et al.,](#page-11-30) [2008;](#page-11-30) [De Oliveira et al.,](#page-10-21) [2011,](#page-10-21) [2013,](#page-10-22) [2014\)](#page-10-23). It will be important to determine whether the effects of sulpiride in females are also observed after pharmacological manipulations of dopaminergic neurotransmission in regions of the mesocorticolimbic system. Ongoing work in our laboratory is assessing the effects of intrabasolateral amygdala dopaminergic compounds in female rats submitted to contextual fear conditioning. In addition to the role of dopamine, we have previously shown that contextual conditioned freezing was inhibited by GABA-A receptor agonist muscimol injections into the basolateral amygdala in males [\(Martinez et al.,](#page-10-41) [2006\)](#page-10-41). Future studies should explore the GABA-dopamine relationship in contextual conditioned fear in males and females at different phases of the estrous cycle.

The study of fear conditioning in rodents has clinical relevance, contributing to a better understanding of the pathophysiology and treatment of human mental disorders such as post-traumatic stress disorder and obsessive-compulsive disorder [\(Milad et al.,](#page-11-31) [2009b,](#page-11-31) [2013;](#page-11-32) [Indovina et al.,](#page-10-42) [2011;](#page-10-42) [Izquierdo et al.,](#page-10-11) [2016;](#page-10-11) [Dougherty et al.,](#page-10-43) [2018;](#page-10-43) [Sevenster](#page-11-33) [et al.,](#page-11-33) [2018;](#page-11-33) [Bienvenu et al.,](#page-9-18) [2021;](#page-9-18) [Cooper and Dunsmoor,](#page-9-19) [2021\)](#page-9-19). The neglect of using female subjects in preclinical neuroscience research, however, contributes to an incomplete understanding of the pathophysiology and treatment of anxiety-related mental disorders in women [\(Beery and](#page-9-20) [Zucker,](#page-9-20) [2011;](#page-9-20) [Shansky and Murphy,](#page-11-3) [2021\)](#page-11-3). The present results reinforce the importance of dopaminergic mechanisms in fear conditioning by demonstrating the involvement of D2-like receptors during the contextual conditioned freezing response in female rats. The present results also indicate that D2-like receptors in females are mainly involved in the expression of fear states during proestrus/estrus rather than in metestrus/diestrus.

Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

Ethics statement

The animal study was reviewed and approved by Committee for Animal Care and Use of the Federal University of São Carlos (protocol no. 5717160919).

Author contributions

AO designed the study and supervised the project. CA performed the research. AR performed the analysis and compiled the figures. AO, AR, and CA wrote the manuscript. All authors contributed to the article and approved the submitted version.

Funding

This study was supported by the Brazilian National Council for Scientific and Technological Development (CNPq Proc. No. 401032/2016-7), the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior–Brasil (CAPES Finance Code 001), and the São Paulo Research Foundation (FAPESP Proc. No. 2016/04620-1). CA was supported by a Master's fellowship (Proc. No. 2019/20274-4) and holds a Doctoral fellowship (Proc. No. 2021/04949-1) both from FAPESP. AR holds a Postdoctoral Young Talents fellowship from CAPES (88887.576069/2020-00).

References

American Psychiatric Association (2013). Diagnostic and Statistical Manual of Mental Disorders, fifth edition. Arlington, VA: American Psychiatric Association.

Bandelow, B., and Michaelis, S. (2015). Epidemiology of anxiety disorders in the 21st century. Dialogues Clin. Neurosci. 17, 327–335. [doi: 10.31887/DCNS.2015.17.](https://doi.org/10.31887/DCNS.2015.17.3/bbandelow) 3/[bbandelow](https://doi.org/10.31887/DCNS.2015.17.3/bbandelow)

Bangasser, D. A., and Cuarenta, A. (2021). Sex differences in anxiety and depression: circuits and mechanisms. Nat. Rev. Neurosci. 22, 674–684. [doi: 10.1038/s41583-021-00513-0](https://doi.org/10.1038/s41583-021-00513-0)

Baran, S. E., Armstrong, C. E., Niren, D. C., Hanna, J. J., and Conrad, C. D. (2009). Chronic stress and sex differences on the recall of fear conditioning and extinction. Neurobiol. Learn. Mem. 91, 323–332. [doi: 10.1016/j.nlm.2008.11.005](https://doi.org/10.1016/j.nlm.2008.11.005)

Barroca, N. C. B., Guarda, M. D., Da Silva, N. T., Colombo, A. C., Reimer, A. E., Brandão, M. L., et al. (2019). Influence of aversive stimulation on haloperidolinduced catalepsy in rats. Behav. Pharmacol. 30, 229–238. [doi: 10.1097/FBP.](https://doi.org/10.1097/FBP.0000000000000462) 0[000000000000462](https://doi.org/10.1097/FBP.0000000000000462)

Beery, A. K., and Zucker, I. (2011). Sex bias in neuroscience and biomedical research. Neurosci. Biobehav. Rev. 35, 565–572. [doi: 10.1016/j.neubiorev.2010.](https://doi.org/10.1016/j.neubiorev.2010.07.002) 0[7.002](https://doi.org/10.1016/j.neubiorev.2010.07.002)

Bienvenu, T. C. M., Dejean, C., Jercog, D., Aouizerate, B., Lemoine, M., and Herry, C. (2021). The advent of fear conditioning as an animal model of post-traumatic stress disorder: learning from the past to shape the future of PTSD research. Neuron 109, 2380–2397. [doi: 10.1016/j.neuron.2021.05.017](https://doi.org/10.1016/j.neuron.2021.05.017)

Blume, S. R., Freedberg, M., Vantrease, J. E., Chan, R., Padival, M., Matthew, J., et al. (2017). Sex and estrus-dependent differences in rat basolateral amygdala. J. Neurosci. 37, 10567–10586. [doi: 10.1523/JNEUROSCI.0758-17.2017](https://doi.org/10.1523/JNEUROSCI.0758-17.2017)

Brooks, M. E., Kristensen, K., Van Benthem, K. J., Magnusson, A., Berg, C. W., Nielsen, A., et al. (2017). glmmTMB balances speed and flexibility among packages for zero-inflated generalized linear mixed modeling. R J. 9, 378–400. [doi: 10.32614/RJ-2017-066](https://doi.org/10.32614/RJ-2017-066)

Carvalho, J. D., De Oliveira, A. R., Da Silva, R. C., and Brandão, M. L. (2009). A comparative study on the effects of the benzodiazepine midazolam and the dopamine agents, apomorphine and sulpiride, on rat behavior in the two-way avoidance test. Pharmacol. Biochem. Behav. 92, 351–356. [doi: 10.1016/j.pbb.2009.](https://doi.org/10.1016/j.pbb.2009.01.001) 0[1.001](https://doi.org/10.1016/j.pbb.2009.01.001)

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: [https://www.frontiersin.org/articles/10.3389/](https://www.frontiersin.org/articles/10.3389/fnbeh.2022.1033649/full#supplementary-material) [fnbeh.2022.1033649/full#supplementary-material.](https://www.frontiersin.org/articles/10.3389/fnbeh.2022.1033649/full#supplementary-material)

Carvalho, M. C., Genaro, K., Leite-Panissi, C. R. A., and Lovick, T. A. (2021). Influence of estrous cycle stage on acquisition and expression of fear conditioning in female rats. Physiol. Behav. 234:e113372. [doi: 10.1016/j.physbeh.2021.113372](https://doi.org/10.1016/j.physbeh.2021.113372)

Colombo, A. C., De Oliveira, A. R., Reimer, A. E., and Brandão, M. L. (2013). Dopaminergic mechanisms underlying catalepsy, fear and anxiety: do they interact? Behav. Brain Res. 257, 201–207. [doi: 10.1016/j.bbr.2013.10.002](https://doi.org/10.1016/j.bbr.2013.10.002)

Cooper, S. E., and Dunsmoor, J. E. (2021). Fear conditioning and extinction in obsessive-compulsive disorder: a systematic review. Neurosci. Biobehav. Rev. 129, 75–94. [doi: 10.1016/j.neubiorev.2021.07.026](https://doi.org/10.1016/j.neubiorev.2021.07.026)

Cora, M. C., Kooistra, L., and Travlos, G. (2015). Vaginal cytology of the laboratory rat and mouse: review and criteria for the staging of the estrous cycle using stained vaginal smears. Toxicol. Pathol. 43, 776–793. [doi: 10.1177/0192623315570339](https://doi.org/10.1177/0192623315570339)

Cossio, R., Carreira, M. B., Vásquez, C. E., and Britton, G. B. (2016). Sex differences and estrous cycle effects on foreground contextual fear conditioning. Physiol. Behav. 163, 305–311. [doi: 10.1016/j.physbeh.2016.05.026](https://doi.org/10.1016/j.physbeh.2016.05.026)

Craske, M., Stein, M., Eley, T., Milad, M. R., Holmes, A., Rapee, R. M., et al. (2017). Anxiety disorders. Nat. Rev. Dis. Primers 3:17024. [doi: 10.1038/nrdp.](https://doi.org/10.1038/nrdp.2017.24) 2[017.24](https://doi.org/10.1038/nrdp.2017.24)

Cullity, E. R., Madsen, H. B., Perry, C. J., and Kim, J. H. (2019). Postnatal developmental trajectory of dopamine receptor 1 and 2 expression in cortical and striatal brain regions. J. Comp. Neurol. 527, 1039–1055. [doi: 10.1002/cne.24574](https://doi.org/10.1002/cne.24574)

Dadkhah, M., Rashidy-Pour, A., and Vafaei, A. A. (2021). Temporary inactivation of the infralimbic cortex impairs while the blockade of its dopamine D2 receptors enhances auditory fear extinction in rats. Pharmacol. Biochem. Behav. 203:173131. [doi: 10.1016/j.pbb.2021.173131](https://doi.org/10.1016/j.pbb.2021.173131)

Daviu, N., Andero, R., Armario, A., and Nadal, R. (2014). Sex differences in the behavioural and hypothalamic-pituitary-adrenal response to contextual fear conditioning in rats. Horm. Behav. 66, 713–723. [doi: 10.1016/j.yhbeh.2014.](https://doi.org/10.1016/j.yhbeh.2014.09.015) 0[9.015](https://doi.org/10.1016/j.yhbeh.2014.09.015)

De la Mora, M. P., Gallegos-Cari, A., Arizmendi-García, Y., Marcellino, D., and Fuxe, K. (2010). Role of dopamine receptor mechanisms in the amygdaloid modulation of fear and anxiety: structural and functional analysis. Prog. Neurobiol. 90, 198–216. [doi: 10.1016/j.pneurobio.2009.10.010](https://doi.org/10.1016/j.pneurobio.2009.10.010)

De Oliveira, A. R., Reimer, A. E., and Brandão, M. L. (2006). Dopamine D2 receptor mechanisms in the expression of conditioned fear. Pharmacol. Biochem. Behav. 84, 102–111. [doi: 10.1016/j.pbb.2006.04.012](https://doi.org/10.1016/j.pbb.2006.04.012)

De Oliveira, A. R., Reimer, A. E., and Brandão, M. L. (2009). Role of dopamine receptors in the ventral tegmental area in conditioned fear. Behav. Brain Res. 199, 271–277. [doi: 10.1016/j.bbr.2008.12.004](https://doi.org/10.1016/j.bbr.2008.12.004)

De Oliveira, A. R., Reimer, A. E., and Brandão, M. L. (2014). Mineralocorticoid receptors in the ventral tegmental area regulate dopamine efflux in the basolateral amygdala during the expression of conditioned fear. Psychoneuroendocrinology 43, 114–125. [doi: 10.1016/j.psyneuen.2014.02.010](https://doi.org/10.1016/j.psyneuen.2014.02.010)

De Oliveira, A. R., Reimer, A. E., De Macedo, C. E., De Carvalho, M. C., Silva, M. A., and Brandão, M. L. (2011). Conditioned fear is modulated by D2 receptor pathway connecting the ventral tegmental area and basolateral amygdala. Neurobiol. Learn. Mem. 95, 37–45. [doi: 10.1016/j.nlm.2010.](https://doi.org/10.1016/j.nlm.2010.10.005) 1[0.005](https://doi.org/10.1016/j.nlm.2010.10.005)

De Oliveira, A. R., Reimer, A. E., Reis, F. M., and Brandão, M. L. (2013). Conditioned fear response is modulated by a combined action of the hypothalamic -pituitary-adrenal axis and dopamine activity in the basolateral amygdala. Eur. Neuropsychopharmacol. 23, 379–389. [doi: 10.1016/j.euroneuro.2012.](https://doi.org/10.1016/j.euroneuro.2012.05.007) 0[5.007](https://doi.org/10.1016/j.euroneuro.2012.05.007)

De Oliveira, A. R., Reimer, A. E., Reis, F. M., and Brandão, M. L. (2017). Dopamine D2-like receptors modulate freezing response, but not the activation of HPA axis, during the expression of conditioned fear. Exp. Brain Res. 235, 429–436. [doi: 10.1007/s00221-016-4805-3](https://doi.org/10.1007/s00221-016-4805-3)

De Souza Caetano, K. A., De Oliveira, A. R., and Brandão, M. L. (2013). Dopamine D2 receptors modulate the expression of contextual conditioned fear:
role of the ventral tegmental area and the basolateral amygdala. *Behav. Pharmacol*. 24, 264–274. [doi: 10.1097/FBP.0b013e32836356c4](https://doi.org/10.1097/FBP.0b013e32836356c4)

De Vita, V. M., Zapparoli, H. R., Reimer, A. E., Brandão, M. L., and De Oliveira, A. R. (2021). Dopamine D2 receptors in the expression and extinction of contextual and cued conditioned fear in rats. Exp. Brain Res. 239, 1963–1974. [doi: 10.1007/s00221-021-06116-6](https://doi.org/10.1007/s00221-021-06116-6)

Delignette-Muller, M. L., and Dutang, C. (2015). fitdistrplus: an R package for fitting distributions. J. Statist. Softw. 64, 1–34. [doi: 10.18637/jss.v064.i04](https://doi.org/10.18637/jss.v064.i04)

Diekhof, E. K., and Ratnayake, M. (2016). Menstrual cycle phase modulates reward sensitivity and performance monitoring in young women: preliminary fMRI evidence. Neuropsychologia 84, 70–80. [doi: 10.1016/j.neuropsychologia.2015.](https://doi.org/10.1016/j.neuropsychologia.2015.10.016) 1[0.016](https://doi.org/10.1016/j.neuropsychologia.2015.10.016)

Dougherty, D. D., Brennan, B. P., Stewart, S. E., Wilhelm, S., Widge, A. S., and Rauch, S. L. (2018). Neuroscientifically informed formulation and treatment planning for patients with obsessive-compulsive disorder: a review. JAMA Psychiatry 75, 1081–1087. [doi: 10.1001/jamapsychiatry.2018.0930](https://doi.org/10.1001/jamapsychiatry.2018.0930)

Fanselow, M. S., and Wassum, K. M. (2015). The origins and organization of vertebrate pavlovian conditioning. Cold Spring Harb. Perspect. Biol. 8:a021717. [doi: 10.1101/cshperspect.a021717](https://doi.org/10.1101/cshperspect.a021717)

Fox, J., and Weisberg, S. (2019). An R Companion to Applied Regression, Third edition. Thousand Oaks CA: Sage.

Frye, C. A., Petralia, S. M., and Rhodes, M. E. (2000). Estrous cycle and sex differences in performance on anxiety tasks coincide with increases in hippocampal progesterone and 3alpha,5alpha-THP. Pharmacol. Biochem. Behav. 67, 587–596. [doi: 10.1016/s0091-3057\(00\)00392-0](https://doi.org/10.1016/s0091-3057(00)00392-0)

Gillies, G. E., and McArthur, S. (2010). Estrogen actions in the brain and the basis for differential action in men and women: a case for sex-specific medicines. Pharmacol. Rev. 62, 155–198. [doi: 10.1124/pr.109.002071](https://doi.org/10.1124/pr.109.002071)

Gouveia, A., Jr., dos Santos, U. D., Felisbino, F. E., de Afonseca, T. L., Antunes, G., and Morato, S. (2004). Influence of the estrous cycle on the behavior of rats in the elevated T-maze. Behav. Process. 67, 167–171. [doi: 10.1016/j.beproc.2004.](https://doi.org/10.1016/j.beproc.2004.03.018) 0[3.018](https://doi.org/10.1016/j.beproc.2004.03.018)

Graham, B. M., and Milad, M. R. (2013). Blockade of estrogen by hormonal contraceptives impairs fear extinction in female rats and women. Biol. Psychiatry. 73, 371–378. [doi: 10.1016/j.biopsych.2012.09.018](https://doi.org/10.1016/j.biopsych.2012.09.018)

Graham, B. M., and Milad, M. R. (2014). Inhibition of estradiol synthesis impairs fear extinction in male rats. Learn. Mem. 21, 347–350. [doi: 10.1101/lm.034926.114](https://doi.org/10.1101/lm.034926.114)

Graham, L. K., Yoon, T., Lee, H. J., and Kim, J. J. (2009). Strain and sex differences in fear conditioning: 22 kHz ultrasonic vocalizations and freezing in rats. Psychol. Neurosci. 2, 219–225. [doi: 10.3922/j.psns.2009.2.015](https://doi.org/10.3922/j.psns.2009.2.015)

Haaker, J., Maren, S., Andreatta, M., Merz, C. J., Richter, J., Richter, S. H., et al. (2019). Making translation work: harmonizing cross-species methodology in the behavioural neuroscience of Pavlovian fear conditioning. Neurosci. Biobehav. Rev. 107, 329–345. [doi: 10.1016/j.neubiorev.2019.09.020](https://doi.org/10.1016/j.neubiorev.2019.09.020)

Hammes, S. R., and Levin, E. R. (2019). Impact of estrogens in males and androgens in females. J. Clin. Invest. 129, 1818–1826. [doi: 10.1172/JCI125755](https://doi.org/10.1172/JCI125755)

Holtzman-Assif, O., Laurent, V., and Westbrook, R. F. (2010). Blockade of dopamine activity in the nucleus accumbens impairs learning extinction of conditioned fear. Learn. Mem. 17, 71–75. [doi: 10.1101/lm.1668310](https://doi.org/10.1101/lm.1668310)

Imhof, J. T., Coelho, Z. M., Schmitt, M. L., Morato, G. S., and Carobrez, A. P. (1993). Influence of gender and age on performance of rats in the elevated plus maze apparatus. Behav. Brain Res. 56, 177–180. [doi: 10.1016/0166-4328\(93\)](https://doi.org/10.1016/0166-4328(93)90036-p) 9[0036-p](https://doi.org/10.1016/0166-4328(93)90036-p)

Indovina, I., Robbins, T. W., Núñez-Elizalde, A. O., Dunn, B. D., and Bishop, S. J. (2011). Fear-conditioning mechanisms associated with trait vulnerability to anxiety in humans. Neuron 69, 563–571. [doi: 10.1016/j.neuron.2010.12.034](https://doi.org/10.1016/j.neuron.2010.12.034)

Izquierdo, I., Furini, C. R. G., and Myskiw, J. C. (2016). Fear Memory. Physiol. Rev. 96, 695–750. [doi: 10.1152/physrev.00018.2015](https://doi.org/10.1152/physrev.00018.2015)

Johnston, A. L., and File, S. E. (1991). Sex differences in animal tests of anxiety. Physiol. Behav. 49, 245–250. [doi: 10.1016/0031-9384\(91\)90039-q](https://doi.org/10.1016/0031-9384(91)90039-q)

Keiser, A., Turnbull, L., Darian, M., Feldman, D. E., Song, I., and Tronson, N. C. (2017). Sex differences in context fear generalization and recruitment of hippocampus and amygdala during retrieval. Neuropsychopharmacology 42, 397–407. [doi: 10.1038/npp.2016.174](https://doi.org/10.1038/npp.2016.174)

Knight, P., Chellian, R., Wilson, R., Behnood-Rod, A., Panunzio, S., and Bruijnzeel, A. W. (2021). Sex differences in the elevated plus-maze test and large open field test in adult Wistar rats. Pharmacol. Biochem. Behav. 204:e173168. [doi: 10.1016/j.pbb.2021.173168](https://doi.org/10.1016/j.pbb.2021.173168)

Kokras, N., Pastromas, N., Papasava, D., de Bournonville, C., Cornil, C. A., and Dalla, C. (2018). Sex differences in behavioral and neurochemical effects of gonadectomy and aromatase inhibition in rats. Psychoneuroendocrinology 87, 93–107. [doi: 10.1016/j.psyneuen.2017.10.007](https://doi.org/10.1016/j.psyneuen.2017.10.007)

Le Saux, M., Morissette, M., and Di Paolo, T. (2006). ERbeta mediates the estradiol increase of D2 receptors in rat striatum and nucleus accumbens.
Neuropharmacology 50, 451–457. [doi: 10.1016/j.neuropharm.2005.10.004](https://doi.org/10.1016/j.neuropharm.2005.10.004)

Lebron-Milad, K., and Milad, M. R. (2012). Sex differences, gonadal hormones and the fear extinction network: implications for anxiety disorders. *Biol. Mood* Anxiety Disord. 2:3. [doi: 10.1186/2045-5380-2-3](https://doi.org/10.1186/2045-5380-2-3)

LeDoux, J. E. (2014). Coming to terms with fear. Proc. Natl. Acad. Sci. U S A 111, 2871–2878. [doi: 10.1073/pnas.1400335111](https://doi.org/10.1073/pnas.1400335111)

Lee, J. H., Lee, S., and Kim, J. H. (2017). Amygdala circuits for fear memory: a key role for dopamine regulation. Neuroscientist 23, 542–553. [doi: 10.1177/1073858416679936](https://doi.org/10.1177/1073858416679936)

Lenth, R. V. (2021). emmeans: estimated marginal means, aka leastsquares means. R package version 1.7.0. Available online at: [https://CRAN.R](https://CRAN.R-project.org/package=emmeans)project.org/packa

Li, S. H., and Graham, B. M. (2017). Why are women so vulnerable to anxiety, trauma-related and stress-related disorders? The potential role of sex hormones. Lancet Psychiatry 4, 73–82. [doi: 10.1016/S2215-0366\(16\)30358-3](https://doi.org/10.1016/S2215-0366(16)30358-3)

Locklear, M. N., Cohen, A. B., Jone, A., and Kritzer, M. F. (2016). Sex differences distinguish intracortical glutamate receptor-mediated regulation of extracellular dopamine levels in the prefrontal cortex of adult rats. Cereb. Cortex 26, 599–610. [doi: 10.1093/cercor/bhu222](https://doi.org/10.1093/cercor/bhu222)

Lovick, T. A. (2014). Sex determinants of experimental panic attacks. Neurosci. Biobehav. Rev. 46, 465–471. [doi: 10.1016/j.neubiorev.2014.03.006](https://doi.org/10.1016/j.neubiorev.2014.03.006)

Machado Figueiredo, R., Carvalho, M. C., Brandão, M. L., and Lovick, T. A. (2019). Short-term, low-dose fluoxetine prevents oestrous cycle linked increase in anxiety-like behaviour in female rats. J. Psychopharmacol. 33, 548–557. [doi: 10.1177/0269881119841833](https://doi.org/10.1177/0269881119841833)

Maeng, L. Y., Cover, K. K., Landau, A. J., Milad, M. R., and Lebron-Milad, K. (2015). Protocol for studying extinction of conditioned fear in naturally cycling female rats. J. Vis. Exp. 96:e52202. [doi: 10.3791/52202](https://doi.org/10.3791/52202)

Maeng, L. Y., and Milad, M. R. (2015). Sex differences in anxiety disorders: interactions between fear, stress and gonadal hormones. Horm. Behav. 76, 106–117. [doi: 10.1016/j.yhbeh.2015.04.002](https://doi.org/10.1016/j.yhbeh.2015.04.002)

Marcondes, F. K., Miguel, K. J., Melo, L. L., and Spadari-Bratfisch, R. C. (2001). Estrous cycle influences the response of female rats in the elevated plus-maze test. Physiol. Behav. 74, 435–440. [doi: 10.1016/s0031-9384\(01\)00593-5](https://doi.org/10.1016/s0031-9384(01)00593-5)

Maren, S., De Oca, B., and Fanselow, M. S. (1994). Sex differences in hippocampal long-term potentiation (LTP) and Pavlovian fear conditioning in rats: positive correlation between LTP and contextual learning. Brain Res. 661, 25–34. [doi: 10.1016/0006-8993\(94\)91176-2](https://doi.org/10.1016/0006-8993(94)91176-2)

Markus, E. J., and Zecevic, M. (1997). Sex differences and estrous cycle changes in hippocampus-dependent fear conditioning. Psychobiology 25, 246-252. [doi: 10.3758/BF03331934](https://doi.org/10.3758/BF03331934)

Martinez, R. C. R., de Oliveira, A. R., and Brandão, M. L. (2006). Conditioned and unconditioned fear organized in the periaqueductal gray are differentially

sensitive to injections of muscimol into amygdaloid nuclei. Neurobiol. Learn. Mem. 85, 58–65. [doi: 10.1016/j.nlm.2005.08.007](https://doi.org/10.1016/j.nlm.2005.08.007)

Martinez, R. C. R., Oliveira, A. R., Macedo, C. E., Molina, V. A., and Brandão, M. L. (2008). Involvement of dopaminergic mechanisms in the nucleus accumbens core and shell subregions in the expression of fear conditioning. Neurosci. Lett. 446, 112–116. [doi: 10.1016/j.neulet.2008.09.057](https://doi.org/10.1016/j.neulet.2008.09.057)

Matsumoto, M., Togashi, H., Kaku, A., Kanno, M., Tahara, K., and Yoshioka, M. (2005). Cortical GABAergic regulation of dopaminergic responses to psychological stress in the rat dorsolateral striatum. Synapse 56, 117–121. [doi: 10.1002/syn.20131](https://doi.org/10.1002/syn.20131)

McLean, C. P., Asnaani, A., Litz, B. T., and Hofmann, S. G. (2011). Gender differences in anxiety disorders: prevalence, course of illness, comorbidity and burden of illness. J. Psychiatr. Res. 45, 1027–1035. [doi: 10.1016/j.jpsychires.2011.](https://doi.org/10.1016/j.jpsychires.2011.03.006) 0[3.006](https://doi.org/10.1016/j.jpsychires.2011.03.006)

Milad, M. R., Furtak, S. C., Greenberg, J. L., Keshaviah, A., Im J. J., Falkenstein M. J. et al. (2013). Deficits in conditioned fear extinction in obsessive-compulsive disorder and neurobiological changes in the fear circuit. JAMA Psychiatry 70, 608–618. [doi: 10.1001/jamapsychiatry.2013.914](https://doi.org/10.1001/jamapsychiatry.2013.914)

Milad, M. R., Igoe, S. A., Lebron-Milad, K., and Novales, J. E. (2009a). Estrous cycle phase and gonadal hormones influence conditioned fear extinction. Neuroscience 164, 887–895. [doi: 10.1016/j.neuroscience.2009.09.011](https://doi.org/10.1016/j.neuroscience.2009.09.011)

Milad, M. R., Pitman, R. K., Ellis, C. B., Gold, A. L., Shin L. M., Lasko N. B. et al. (2009b). Neurobiological basis of failure to recall extinction memory in posttraumatic stress disorder. Biol. Psychiatry 66, 1075–1082. [doi: 10.1016/j.biopsych.2009.06.026](https://doi.org/10.1016/j.biopsych.2009.06.026)

Mitsushima, D., Yamada, K., Takase, K., Funabashi, T., and Kimura, F. (2006). Sex differences in the basolateral amygdala: the extracellular levels of serotonin and dopamine and their responses to restraint stress in rats. Eur. J. Neurosci. 24, 3245–3254. [doi: 10.1111/j.1460-9568.2006.05214.x](https://doi.org/10.1111/j.1460-9568.2006.05214.x)

Morena, M., Nastase, A. S., Santori, A., Cravatt, B. F., Shansky, R. M., and Hill, M. N. (2021). Sex-dependent effects of endocannabinoid modulation of conditioned fear extinction in rats. Br. J. Pharmacol. 178, 983–996. [doi: 10.1111/bph.15341](https://doi.org/10.1111/bph.15341)

Mueller, D., Bravo-Rivera, C., and Quirk, G. J. (2010). Infralimbic D2 receptors are necessary for fear extinction and extinction-related tone responses. Biol. Psychiatry 68, 1055–1060. [doi: 10.1016/j.biopsych.2010.08.014](https://doi.org/10.1016/j.biopsych.2010.08.014)

Nouri, S., Biedermann, S. V., Joue, G., Auer, M. K., Sommer, T., and Fuss, J. (2022). Effects of circulating estradiol on physiological, behavioural and subjective correlates of anxiety: a double-blind, randomized, placebo-controlled trial. Psychoneuroendocrinology 138:e105682. [doi: 10.1016/j.psyneuen.2022.105682](https://doi.org/10.1016/j.psyneuen.2022.105682)

Petersen, N., Rapkin, A. J., Okita, K., Kinney, K. R., Mizuno, T., Mandelkern, M. A., et al. (2021). Striatal dopamine D 2-type receptor availability
and peripheral 17β-estradiol. *Mol. Psychiatry* 26, 2038–2047. [doi: 10.1038/s41380-](https://doi.org/10.1038/s41380-020-01000-1) 0[20-01000-1](https://doi.org/10.1038/s41380-020-01000-1)

Pezze, M. A., and Feldon, J. (2004). Mesolimbic dopaminergic pathways in fear conditioning. Prog. Neurobiol. 74, 301–320. [doi: 10.1016/j.pneurobio.2004.09.004](https://doi.org/10.1016/j.pneurobio.2004.09.004)

Pezze, M. A., Heidbreder, C. A., Feldon, J., and Murphy, C. A. (2001). Selective responding of nucleus accumbens core and shell dopamine to aversively conditioned contextual and discrete stimuli. Neuroscience 108, 91-102. [doi: 10.1016/s0306-4522\(01\)00403-1](https://doi.org/10.1016/s0306-4522(01)00403-1)

Ponnusamy, R., Nissim, H. A., and Barad, M. (2005). Systemic blockade of D2-like dopamine receptors facilitates extinction of conditioned fear in mice. Learn. Mem. 12, 399–406. [doi: 10.1101/lm.96605](https://doi.org/10.1101/lm.96605)

Purves-Tyson, T. D., Arshi, M. S., Handelsman, D. J., Cheng, Y., and Keast, J. R. (2007). Androgen and estrogen receptor-mediated mechanisms of
testosterone action in male rat pelvic autonomic ganglia. N*euroscience* 148,92–104. [doi: 10.1016/j.neuroscience.2007.05.043](https://doi.org/10.1016/j.neuroscience.2007.05.043)

R Core Team (2020). R: a language and environment for statistical computing. R Foundation for Statistical Computing. Vienna, Austria. Available online at: [https://www.R-project.org/.](https://www.R-project.org/)

Reimer, A. E., De Oliveira, A. R., and Brandão, M. L. (2012). Glutamatergic mechanisms of the dorsal periaqueductal gray matter modulate the expression

of conditioned freezing and fear-potentiated startle. Neuroscience 219, 72–81. [doi: 10.1016/j.neuroscience.2012.06.005](https://doi.org/10.1016/j.neuroscience.2012.06.005)

Reimer, A. E., de Oliveira, A. R., Diniz, J. B., Hoexter, M. Q., Miguel, E. C., Milad, M. R., et al. (2018). Fear extinction in an obsessive-compulsive disorder animal model: influence of sex and estrous cycle. Neuropharmacology 131, 104–115. [doi: 10.1016/j.neuropharm.2017.12.015](https://doi.org/10.1016/j.neuropharm.2017.12.015)

Rey, C. D., Lipps, J., and Shansky, R. M. (2014). Dopamine D1 receptor activation rescues extinction impairments in low-estrogen female rats and induces cortical layer-specific activation changes in prefrontal-amygdala circuits. Neuropsychopharmacology 39, 1282–1289. [doi: 10.1038/npp.](https://doi.org/10.1038/npp.2013.338) 2[013.338](https://doi.org/10.1038/npp.2013.338)

RStudio Team (2020). RStudio: Integrated Development for R. Boston, MA: RStudio, PBC. Available online at: [http://www.rstudio.com/.](http://www.rstudio.com/)

Schoemaker, H., Claustre, Y., Fage, D., Rouquier, L., Chergui, K., Curet, O., et al. (1997). Neurochemical characteristics of amisulpride, an atypical dopamine D2/D3 receptor antagonist with both presynaptic and limbic selectivity. J. Pharmacol. Exp. Ther. 280, 83–97.

Scholl, J. L., Afzal, A., Fox, L. C., Watt, M. J., and Forster, G. L. (2019). Sex differences in anxiety-like behaviors in rats. Physiol. Behav. 211:e112670. [doi: 10.1016/j.physbeh.2019.112670](https://doi.org/10.1016/j.physbeh.2019.112670)

Sevenster, D., Visser, R. M., and D'Hooge, R. (2018). A translational perspective on neural circuits of fear extinction: current promises and challenges. Neurobiol. Learn. Mem. 155, 113–126. [doi: 10.1016/j.nlm.2018.07.002](https://doi.org/10.1016/j.nlm.2018.07.002)

Shansky, R. M., and Murphy, A. Z. (2021). Considering sex as a biological variable will require a global shift in science culture. Nat. Neurosci. 24, 457-464. [doi: 10.1038/s41593-021-00806-8](https://doi.org/10.1038/s41593-021-00806-8)

Staiti, A. M., Morgane, P. J., Galler, J. R., Grivetti, J. Y., Bass, D. C., and Mokler, D. J. (2011). A microdialysis study of the medial prefrontal cortex of adolescent and adult rats. Neuropharmacology 61, 544–549. [doi: 10.1016/j.](https://doi.org/10.1016/j.neuropharm.2011.04.005) n[europharm.2011.04.005](https://doi.org/10.1016/j.neuropharm.2011.04.005)

Stubbendorff, C., and Stevenson, C. W. (2021). Dopamine regulation of contextual fear and associated neural circuit function. Eur. J. Neurosci. 54, 6933–6947. [doi: 10.1111/ejn.14772](https://doi.org/10.1111/ejn.14772)

Thompson, T. L., and Moss, R. L. (1997). Modulation of mesolimbic dopaminergic activity over the rat estrous cycle. Neurosci. Lett. 229, 145–148. [doi: 10.1016/s0304-3940\(97\)00450-3](https://doi.org/10.1016/s0304-3940(97)00450-3)

Urien, L., and Bauer, E. P. (2022). Sex differences in BNST and amygdala activation by contextual, cued and unpredictable threats. eNeuro 9:ENEURO.0233-21.2021. [doi: 10.1523/ENEURO.0233-21.2021](https://doi.org/10.1523/ENEURO.0233-21.2021)

Waku, I., Reimer, A. E., and De Oliveira, A. R. (2022). Effects of immediate aversive stimulation on haloperidol-induced catalepsy in rats. Front. Behav. Neurosci. 16:e867180. [doi: 10.3389/fnbeh.2022.867180](https://doi.org/10.3389/fnbeh.2022.867180)

Xiao, L., and Becker, J. B. (1994). Quantitative microdialysis determination of extracellular striatal dopamine concentration in male and female rats: effects of estrous cycle and gonadectomy. *Neurosci. Lett.* 180, 155–158. [doi: 10.1016/0304-](https://doi.org/10.1016/0304-3940(94)90510-x) 3[940\(94\)90510-x](https://doi.org/10.1016/0304-3940(94)90510-x)

Zbukvic, I. C., Park, C. H., Ganella, D. E., Lawrence, A. J., and Kim, J. H. (2017). Prefrontal dopaminergic mechanisms of extinction in adolescence compared to adulthood in rats. Front. Behav. Neurosci. 11:32. [doi: 10.3389/fnbeh.2017.](https://doi.org/10.3389/fnbeh.2017.00032) 0[0032](https://doi.org/10.3389/fnbeh.2017.00032)

Zeidan, M. A., Igoe, S. A., Linnman, C., Vitalo, A., Levine, J. B., Klibanski, A., et al. (2011). Estradiol modulates medial prefrontal cortex and amygdala activity during fear extinction in women and female rats. Biol. Psychiatry 70, 920–927. [doi: 10.1016/j.biopsych.2011.05.016](https://doi.org/10.1016/j.biopsych.2011.05.016)

Zhao, Y., Bijlsma, E. Y., Verdouw, M. P., and Groenink, L. (2018). No effect of sex and estrous cycle on the fear potentiated startle response in rats. Behav. Brain Res. 1, 24–33. [doi: 10.1016/j.bbr.2018.05.022](https://doi.org/10.1016/j.bbr.2018.05.022)

Zuloaga, D. G., Heck, A. L., De Guzman, R. M., and Handa, R. J. (2020). Roles for androgens in mediating the sex differences of neuroendocrine and behavioral stress responses. Biol. Sex Differ. 11:44. [doi: 10.1186/s13293-020](https://doi.org/10.1186/s13293-020-00319-2) -0[0319-2](https://doi.org/10.1186/s13293-020-00319-2)