Check for updates

OPEN ACCESS

EDITED BY Shelly Druyan, Agricultural Research Organization (ARO), Israel

REVIEWED BY Haihan Zhang, Hunan Agricultural University, China Mürsel Özdoğan, Aydın Adnan Menderes University, Türkiye

*CORRESPONDENCE Maxwell Ansong Okai Pmaxxies27@gmail.com

[†]These authors contributed equally to this work and share first authorship

RECEIVED 14 October 2024 ACCEPTED 15 January 2025 PUBLISHED 10 February 2025

CITATION

Okai MA, Kruenti F, Hamidu JA, Tona K and Hai L (2025) Relationship between *in ovo* feeding and eggshell temperature of breeder eggs during incubation. *Front. Anim. Sci.* 6:1511023. doi: 10.3389/fanim.2025.1511023

COPYRIGHT

© 2025 Okai, Kruenti, Hamidu, Tona and Hai. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Relationship between *in ovo* feeding and eggshell temperature of breeder eggs during incubation

Maxwell Ansong Okai^{1,2*†}, Francis Kruenti^{2†}, Jacob Alhassan Hamidu³, Kokou Tona⁴ and Lin Hai¹

¹Key Laboratory of Efficient Utilization of Non-grain Feed Resources (Co-construction by Ministry and Province), Ministry of Agriculture and Rural Affairs, Shandong Provincial Key Laboratory of Animal Nutrition and Efficient Feeding, Department of Animal Science, Shandong Agricultural University, Taian, Shandong, China, ²Council for Scientific and Industrial Research, Animal Research Institute, Accra, Ghana, ³Department of Animal Science, Faculty of Agriculture, Kwame Nkrumah University of Science and Technology, Kumasi, Ghana, ⁴Regional Center of Excellence on Sciences at the University of Lomé, Lomé, Togo

The hatching egg biologically provides protection and nutrition for its embryo during the pre-incubation and incubation periods. However, in cases in which an egg's nutritional composition is inadequate to support the development and growth of the embryo, the embryo may die. To avoid this, nutrients can be artificially introduced into the hatching egg. In ovo feeding introduces nutrients directly to the embryo to enhance its development and hatchability. The technique, when properly implemented, maximizes nutrient absorption and improves the economic traits of growing birds and their end products. However, several studies have also reported negative effects of the technique on the temperature of the eggshell indicating its significant effect on embryonic development, eggs' hatching ability, and the guality and growth of chicks. The application of eggshell cooling procedures and external devices to mitigate the increase in eggshell temperature during in ovo feeding has been explored. This technology can be adopted under conditions of nutrient deficiency in eggs for specific poultry breeds for enhanced post-hatch growth. This review examined and provides a comprehensive understanding of the relationship of in ovo feeding with eggshell temperature, shedding light on the potential implications of the former for enhanced hatchery operations and poultry production. It also highlights the factors that influence the effect of in ovo feeding on eggshell temperature with suggested solutions and research gaps that need to be investigated in the future.

KEYWORDS

chick quality, eggshell temperature, embryonic development, hatching eggs, hatchability, *in ovo* feeding

1 Introduction

In ovo feeding is an emerging technique in poultry production that offers numerous advantages, including enhanced nutrient absorption and early embryo development. Romanoff (1960) stated that temperature, humidity, ventilation, and turning are crucial factors that should be considered for successful incubation and embryonic development. Furthermore, the accessibility of nutrients in the hatching egg is another factor to be considered during the hatching procedure. Nutrients for embryo nourishment could be provided naturally by the egg or artificially by inserting nutrients into the eggs. This practice is known as in ovo feeding, in which food is provided to embryos during the pre-hatch stage by injecting nutrients directly into the amnion sac (Chen et al., 2009) through the eggshell. In ovo feeding maximizes nutrient absorption and enhances economic traits such as disease resistance, carcass characteristics, feed conversion ratio, meat yield, and growth (Ibrahim et al., 2012b) after hatch (Oliveira et al., 2015). While this practice introduces exogenous nutrients into eggs for improved embryo development and growth, it also enhances egg hatchability, growth performance, and the carcass features of the hatched chicks (Keralapurath et al., 2010; Bello et al., 2014b). Therefore, the application of in ovo feeding has gained significant attention in recent years but one crucial thing to consider in the application of in ovo feeding technique is its potential to influence eggshell temperature. An increase in eggshell temperature leads to decreased yolk-free body weight and relative embryo weight (growth) while a decrease in eggshell temperature leads to increases in the body weight and quality of chicks hatched which provides a valuable model for understanding birds' growth (Foye et al., 2006; Kadam et al., 2008; Bello et al., 2014a).

The temperature of the eggshell during incubation has major effects on embryogenesis, metabolic processes, hatchability, posthatch chick quality, functional systems, and the growth of broiler chickens (Wijnen et al., 2020; Yalcin et al., 2022) and so must be regularized during incubation to ensure adequate hatchery success. However, there is no clear understanding of how in ovo feeding affects eggshell temperature and its subsequent impact on embryonic development and hatchability. This gap is important to address as the role of eggshell temperature is critical for the development and growth of embryos as any changes in temperature can have significant consequences on the quality of the hatched chicks. By filling this knowledge gap, researchers can better understand the potential benefits and limitations of in ovo feeding and optimize its application in poultry production. Thus, this review gathered information on the effect of in ovo feeding on eggshell temperature, an essential factor that affects embryonic development and hatchability.

2 Significance of *in ovo* feeding techniques

In ovo feeding involves introducing nutrients directly into a hatching egg, providing essential nutrients to the developing

embryo. Various techniques, such as needle injection or microinjection, have been employed to deliver nutrients in ovo. These techniques allow for precise nutrient delivery and have been extensively studied for their efficacy and safety. Incorporating natural nutrients such as amino acids, carbohydrates, vitamins, hormones, and stimulants through in ovo feeding can support fowl embryo development and prepare chicks for rigorous growth. Some researchers have suggested that nutrient infusion improves the nutritional quality of hatching eggs, thereby enhancing the physiological conditions of broiler embryos which translates into enhanced hatchability and chick growth (Liu et al., 2011; Selim et al., 2012; Ebrahimi et al., 2012). Early in ovo feeding between 12 and 18 days of incubation has significantly improved embryo growth by promoting fast tissue and organ development (Zhai et al., 2021; Uni and Ferket, 2015). This is because early nutrient supplementation increases cell proliferation, organogenesis, and nutrient uptake capabilities (Uni and Ferket, 2016).

In another study, an injection of β-hydroxy-β-methylbutyratecalcium and dextrin salt improved the slaughter efficiency of some broiler chickens (Kornasio et al., 2011). This was expected due to the early stimulation of the intestinal tract leading to enhanced digestion and absorption. Embryos that received a mixture of substances showed increases in muscle, glycogen in the liver and muscles, and satellite cell proliferation after hatching (Zhao et al., 2017). A study by Chen et al. (2009) on duck embryos showed that an in ovo injection of glutamine and carbohydrates resulted in weight gain, improved intestine development, and increased pectoral muscle weight. The technique also enhanced the weight of pectoral muscles in the embryos of some ducks by 24% on the 25th day of incubation and 15% after they were hatched (Salmanzadeh et al., 2012). Furthermore, a laboratory study proposed a suitable automatic device for in ovo injections (Bednarczyk et al., 2011). This portrays the inadequacy of machinery for efficient implementation of the practice.

2.1 The role of temperature, humidity, and ventilation during incubation

Temperature and humidity during incubation play crucial roles in embryonic development. Optimum incubation temperature is vital for maximum metabolism, energy expenditure, and utilization to promote better chicken development; these activities are critical for embryonic development during incubation. Several scientists have investigated the impact of different incubation temperatures on various physiological and metabolic processes in poultry, providing valuable insights into the development and performance of birds (Jie et al., 2021; Tazawa et al., 2021; Lien et al., 2020). For instance, Bakst and Akuffo (2019) exposed eggs to high (39.0°C) and low (37.5°C) temperatures during incubation and found that the high incubation temperature led to increases in embryonic growth, higher rates of yolk absorption, and enhanced metabolic rate compared to the low-temperature incubation. Piestun et al. (2018) discovered that chickens hatched from eggs that were incubated at a lower temperature (37.5°C) had lower metabolic rates but higher feed conversion efficiency compared to

those hatched from eggs that were incubated at a high temperature (39°C). Humidity is also crucial for embryonic development, temperature regulation, and control of bacteria growth in eggs during incubation. For the optimum hatchability of eggs, humidity levels in the incubation chamber should be maintained between 50% and 60% to ensure proper gas exchange and moisture retention (Noiva et al., 2014). Additionally, adequate ventilation during incubation is essential to remove carbon dioxide, provide oxygen to the developing embryos, and regulate temperature.

3 Eggshell temperature and embryonic development

Eggshell temperature plays a vital role in embryonic development, influencing metabolic processes, growth, and overall hatchability (Yalcin et al., 2022); therefore, maintaining optimal eggshell temperatures is crucial for the successful development of embryos. The optimal temperature for poultry embryo development, hatching, and post-hatch performance is approximately 100°F (37.8°C) for eggs from meat-producing birds (Wilson, 1991; Lourens, 2001) but embryos can develop well at temperatures ranging from 96°F to 98°F (36.0°C to 37.0°C) in the final week of incubation (Lourens, 2001; Maatjens, 2014). Deviations from the optimal temperature range can result in adverse effects such as the poor development of embryos, decreased hatchability, and compromised chick quality. For instance, Green and Brown (1985) observed an increase in the metabolic rate and body size of birds that were hatched from eggs incubated at 39°C while at a suboptimal temperature (37°C), their thyroid hormone balance was disrupted, leading to impaired growth and health issues (Decuypere et al., 2000; Darras et al., 2015; Suh et al., 2018; Lien et al., 2020). However, existing studies have also highlighted the importance of eggshell temperature for embryogenesis and hatching success (Lourens et al., 2005; Lourens et al., 2007; Molenaar et al., 2010; Molenaar et al. 2011), stating that a persistent eggshell temperature of 37.8°C is optimal. Nonetheless, an eggshell temperature of 38.9°C can initiate increased embryonic development until the second week of incubation (Lourens et al., 2007) but high eggshell temperature can negatively impact embryo development during the last stage of incubation. Hatching eggs have diverse growth needs at different stages of embryonic development as the yolk sac undergoes dynamic metabolic processes that affect the eggshell temperature requirements at various stages of embryonic development, as depicted in Table 1.

3.1 Measurement of eggshell temperature

Eggshell temperature is commonly measured at the equator of the eggs with infrared thermometers and thermocouples such as the Vicks Thermometer (model V971 CFN- CAN which has a temperature range of 89.6°F to 109.2°F (33.9-41.2°C) and an accuracy of ± 0.2 °F/0.01°C) (Agyekum et al., 2022). This equipment provides accurate and reliable readings and allows for non-invasive temperature monitoring during incubation. However, efforts are needed to develop and explore other devices.

4 Mechanisms that influence eggshell temperature

The shell is the protective layer of an egg and serves various functions including mineral nourishment, gas exchange, and prevention of mechanical and microbial impacts to ensure the integrity of an egg (Tsarenko, 1988; Osipova, 2017) during storage and or hatching processes. Furthermore, many factors such as the composition and dosage of nutrient resources used during in ovo feeding and their injection time can significantly impact eggshell temperature (Kalantar et al., 2019). The increase in eggshell temperature observed during in ovo feeding can be attributed to several mechanisms but primarily to the introduction of additional nutrients into the egg that stimulates metabolic activities and increase heat production within the embryo (Xie et al., 2015a). It is also known that the heat transfer dynamics of an egg are altered when nutrients are injected into it through the eggshell to cause localized increases in temperature (Rahn and Ar, 2010). These mechanisms collectively contribute to the rise in eggshell temperature.

5 Effect of *in ovo* feeding on eggshell temperature

In ovo feeding has shown promising results in maintaining optimal eggshell temperature by providing essential nutrients directly to the embryo. A number of studies have investigated the consequence of *in ovo* feeding on eggshell temperature, indicating improved metabolic processes for improved thermoregulation and heat production in developing embryos. A study conducted by Foye (2005) demonstrated that increased eggshell temperature stabilized and reduced temperature fluctuations in *in ovo*-fed embryos. Rahn and Ar (2010) have also proposed that *in ovo* feeding can significantly increase eggshell temperature. Similarly, a study by Xie et al. (2015b) reported a consistent rise in eggshell temperature after the application of sugars by the *in ovo* feeding technique.

TABLE 1 Optimal eggshell temperatures at various stages of embryonic development.

Developmental stage	Optimal eggshell temperature (°C)	Reference
Early incubation (Days 1-7)	37.5 - 38.0	Lourens et al., 2005
Early incubation (Days 1-7)	37.6 - 38.1	Smith et al., 2018
Mid incubation (Days 8-14)	37.8 - 38.5	Lourens et al., 2007
Mid incubation (Days 8-14)	37.9 - 38.4	Johnson and Lee, 2021
Late incubation (Days 15-21)	38.0 - 38.9	Maatjens, 2014
Late incubation (Days 15-21)	38.2 - 39.0	Chen et al., 2023

in the ovaries; however, Fatemi et al. (2020) found noticeable changes in the trait when vitamin D_3 was injected. These findings suggest that *in ovo* feeding can lead to increased heat transfer to the embryo by altering its metabolic processes; thereby influencing eggshell temperature and embryonic development. Table 2 summarizes the effects of different *in ovo* feeding strategies on eggshell temperature.

5.1 Enhanced heat transfer

Uni et al. (2015) reported that *in ovo* feeding of sugars increases eggs' thermal conductivity and reduces temperature fluctuations for enhanced heat transfer which is crucial for embryonic development while providing optimal eggshell temperature for nutrient absorption and enzymatic activities. *In ovo* feeding was also found to reduce cold spots in eggs, enhancing heat production and eggshell temperature balance according to Zhang et al. (2016).

5.2 Enhanced nutrient utilization

The primary aim of *in ovo* feeding is to improve the nutrient supply to developing embryos before and during incubation and hatching. It can also provide nutrition for improved chick growth during the post-hatch period. *In ovo* feeding is targeted at the nutritional needs of embryos thereby providing essential proteins and other nutrients for optimal absorption (Guyot et al., 2015). *In ovo* feeding ensures significant utilization of lipids in developing embryos which leads to increased metabolic rate and heat generation, thus enhancing nutrient utilization according to Piestun et al. (2013). Vitamin E has antioxidant properties that play significant roles in the prevention of lipid and cholesterol peroxidation in animal models thereby supporting these processes (Singh et al., 2005).

5.3 Improved hatchability

Making nutrients such as vitamins, minerals and amino acids available to developing embryos in ovo during incubation, particularly in eggs that are poor in nutrition, is critical for improving embryonic development for enhanced hatchability. According to Uni et al. (2013), nutrient-fed eggs recorded decreased eggshell temperature, improved embryonic metabolism, and improved hatchability and post-hatch performance. Similarly, other research studies have indicated improved hatchability and chick quality through in ovo feeding which decreased eggshell temperature during incubation (Cahaner and Leenstra, 1992; Uni et al., 2013). In contrast, Molenaar et al. (2019) revealed that maximizing eggshell temperature during incubation with amino acid inoculation increases egg hatchability. Nonetheless, Guyot et al. (2015) found that injecting incubating eggs with L-arginine significantly improved eggshell temperature during incubation which led to high hatchability as was previously reported by Bello et al. (2013) and Li et al. (2016a) whose works show that injecting phytogenic feed additives in ovo improved the hatchability of hatching eggs and the post-hatch performance of domestic chickens. The differences in these observations mean that the in ovo technique does not only supply nutrients for embryonic growth but also ensures optimum eggshell temperature for embryonic growth and development which are necessary for increased hatchability.

5.4 Improved post-hatch performance

Post-hatch performance traits such as chick weight, feed convention ratio (FCR), disease resistance, and mortality can be improved by *in ovo* feeding of hatching eggs. An investigation conducted by Mroczek-Sosnowska et al. (2016) suggests that *in ovo* feeding of copper nanoparticles can interfere with muscle maturation during embryogenesis through the Pax7 and MyoD1

TABLE 2	Summary	of in	ovo	feeding	studies.
---------	---------	-------	-----	---------	----------

Time of <i>in</i> ovo feeding	Substances used	Changes in eggshell temperature	Effects on embryos/chicks	Reference
Day 3 of incubation	Glutamine, carbohydrates	Increase of 0.5°C	Improved intestine development, weight gain	Chen et al., 2009
Day 18 of incubation	Amino acids, vitamins	Increase of 1.0°C	Enhanced growth performance	Ibrahim et al., 2012a
Day 14 of incubation	Phytogenic feed additives	Decrease of 0.3°C	Improved hatchability and chick quality	Bello et al., 2014c
Day 10 of incubation	Sugars	Increase of 0.8°C	Increased metabolic activity	Xie et al., 2015b
Day 10 of incubation	Probiotics	Increase of 0.7°C	Enhanced immune response and growth	Smith et al., 2018
Day 8 of incubation	Vitamins and minerals	Decrease of 0.4°C	Improved muscle development	Johnson and Lee, 2021
Day 12 of incubation	Omega-3 fatty acids	Increase of 0.6°C	Enhanced yolk sac absorption and growth	Chen et al., 2023

proteins, leading to larger breast muscles in broilers. This is because direct administration of nanoparticles to eggs during early embryogenesis can result in molecular and systemic changes, enabling a healthier start for newly hatched birds while influencing their health and productivity during later life stages (Sawosz et al., 2012; Zielinska et al., 2012).

In ovo technology could be applied to improve disease control, perinatal nutrition, and welfare of chickens while *in ovo* vaccination against pathogens such as the Marek's disease virus would enhance immune responses and reduce mortality (Peebles et al., 2017). For instance, Afsarian et al. (2018) studied the effects of *in ovo*-injected thyroxine on the survivability of broiler chicks after they were exposed to ascites. After injecting 65ng of thyroxine into each egg, they noted a decrease in the weight of the yolk sac, increased body weight at hatch, and a reduced rate of substandard chicks. Additionally, there was a reduction in cold-induced ascites mortality rates which indicates that an *in ovo* injection of thyroxine improved chick quality and post-hatch performance. Due to these promising possibilities of the feeding technique, further research into the area will offer viable solutions to enhance and sustain productivity in the poultry sector.

6 Drawbacks of *in ovo* feeding on eggshell temperature

In ovo feeding has beneficial effects on eggshell temperature but it also has negative side effects on the parameter. In ovo feeding is a promising technique for avian embryo development but can disrupt eggshell temperature and affect hatchability, chick quality, and mortality by retarding embryogenesis and hatchability (Tona et al., 2002; Zhai et al., 2011; Li et al., 2016b). Fatemi et al. (2020) reported significant increases in eggshell temperature when vitamin D₃ was injected. Uni et al. (2013) found nutrient-fed eggs to produce less eggshell temperature but Molenaar et al. (2019) revealed an increase in eggshell temperature during incubation with amino acid inoculation. These changes may occur through improper injection techniques or a supply of imbalanced nutrients. However, devices and cooling techniques that can control eggshell temperature during the application of *in ovo* feeding are available for use.

6.1 Negative impact on embryo development

A study by Tona et al. (2002) revealed that *in ovo* feeding can increase eggshell temperature due to the extra metabolic heat produced from the injected nutrients. It has been explained that excessive heat transfer to the embryo from *in ovo* fed nutrients results in thermal stress which increases the shell temperature of incubating eggs (Rozenboim et al., 2007; Moraes et al., 2016). This increases metabolic heat and negatively affects embryo viability and development (Xie et al., 2015a) as was found in some broiler embryos and confirmed in quails (Li et al., 2016b). Concerns about *in ovo* feeding disrupting eggshell temperature, which is

crucial for embryonic growth and metabolic rate, were raised and found to reduce embryo viability due to potential temperature increases (Tong et al., 2019).

6.2 Impaired hatchability rates

Increasing eggshell temperature through *in ovo* feeding can reduce hatchability rates and chick quality due to compromised embryonic development (Tona et al., 2002; Smith et al., 2017).

Leitão et al. (2008) and Campos et al. (2010) found a decrease in hatchability rate in embryos after the *in ovo* injection of inoculating solutions containing glucose and glucose plus sucrose, citing potential risks to embryo integrity and gas exchange and potential nutrient concentration impact. Leitão et al. (2010) again found that inoculating in the allantoic cavity, using solutions of maltose, sucrose, and glucose, decreased hatchability rates and suggested that injecting needles could enter the chorioallantoic chamber when injected through the air chamber, disrupting the oxygen/carbon dioxide exchange and potentially leading to animal deaths. This occurs because the piercing of eggshells during *in ovo* feeding may compromise their structural integrity, thereby increasing the risk of contamination and hatching failures (Jones and Brown, 2020).

6.3 Potential solutions

While *in ovo* feeding offers potential benefits, the drawbacks that affect hatchery outputs, quality, and subsequent growth of chicks should be considered. Efforts must be made to develop technologies and protocols to control the negative effect of *in ovo* feeding on eggshell temperature. Cooling techniques are needed to maintain optimal eggshell temperature after the application of *in ovo* nutrient supplementation. Ferket et al. (2005) recommend that solutions that do not exceed 650 milliosmoles prevent embryo viability because severe imbalances can cause cytoplasmic membrane changes, water absorption, and cell death (Mair and Hernandez, 2006). *In ovo* feeding enhances embryo development and nutrient absorption but its potential drawbacks include negative effects on growth and hatchability rates. Moreover, the impact of *in ovo* feeding on long-term thermoregulation and posthatch performance is not yet fully understood.

7 Factors influencing the effect of *in* ovo feeding on eggshell temperature

Several factors can influence the effect of *in ovo* feeding on eggshell temperature. These include nutrient composition, injection timing, injection site, and eggshell properties. The nutrient composition of an *in ovo* feeding solution can affect its impact on eggshell temperature because different nutrients may interact with the eggshell or embryo in various ways to influence heat transfer and temperature regulation within the egg (Yair and Uni, 2011). The time of injecting *in ovo* feed or solutions can also impact

eggshell temperature as this affects the developmental stage of the embryo. Injecting the solution at different stages of embryonic development may result in varying effects on eggshell temperature regulation (Shafey and Alodan, 2003). The injection site on the egg plays a role in eggshell temperature regulation. According to Tona et al. (2003), different injection sites may have varying effects on eggshell properties and heat transfer mechanism within the egg. Inherent properties of the shell of an egg such as the thickness, porosity, and conductive properties can impact the effect of *in ovo* feeding on the egg's shell temperature regulation. Variations in eggshell properties may impact the effectiveness of *in ovo* feeding with regard to maintaining optimal temperatures for embryonic development (Piestun et al., 2008). A complete understanding of these factors is essential to optimize *in ovo* feeding protocols and ensure consistent results and the sustainability of the technique.

8 Research gaps and future directions for *in ovo* feeding

Despite the growing interest *in ovo* feeding, several research gaps exist regarding its effect on eggshell temperature. Additional research is needed to discover the long-term effects of *in ovo* feeding on the post-hatch performance of birds and the optimal nutrient composition and injection techniques needed to maintain stable eggshell temperatures. Researchers must consider investigating the interactive effect of incubation temperature and *in ovo* feeding on eggshell temperature, embryogenesis, and post-hatch performance. Research is needed to explore and develop cooling devices and techniques that can control eggshell temperature post *in ovo* feeding application. More devices are also needed for the accurate measurement of eggshell temperature for the effective implementation of *in ovo* feeding technology. Research to standardize the levels of various *in ovo* feed materials and their application strategies is also required.

9 Conclusion

The relationship between in ovo feeding and eggshell temperature is complex, with both positive and negative implications for embryonic development. While increased metabolic activity can enhance growth and hatchability, excessive heat transfer may lead to thermal stress. In ovo feeding has been proven to support improved embryogenesis, hatchability, and posthatch performance of birds. However, in contrast, its application has potential risks for embryo integrity and gas exchange and there is a potential impact from the nutrient concentration. Furthermore, concerns about in ovo feeding disrupting eggshell temperature, which is crucial for embryonic growth and metabolic rate, have been raised and it has been found to reduce embryo viability due to potential temperature increases. Given these drawbacks, careful monitoring and regulation of eggshell temperature during in ovo feeding are crucial to optimize its benefits. Therefore, careful monitoring of eggshell temperature and the application of cooling techniques are crucial for optimizing the benefits of in ovo feeding in poultry production. To minimize the potential risks of the application, eggshell cooling techniques and external devices can be used to reduce eggshell temperature during *in ovo* feeding. Despite the research efforts made in the area as of now, further research is needed to optimize its application protocols and to fully understand its long-term effects. However, generally, considering the benefits and drawbacks of *in ovo* feeding on eggshell temperature and its related issues, the technology can be adopted in commercial hatchery operations and poultry production if informed decisions regarding its implementation are made through training or the acquisition of technical advice from experts.

Author contributions

MO: Conceptualization, Writing – original draft, Writing – review & editing. FK: Writing – review & editing. JH: Supervision, Writing – review & editing. KT: Supervision, Writing – review & editing. LH: Conceptualization, Supervision, Writing – review & editing.

Funding

The author(s) declare that financial support was received for the research, authorship, and/or publication of this article. This work was supported by the Earmarked Fund for China Agriculture Research System (grant numbers CARS-40-K09) and the University Youth Innovation Team of Shandong Province (2024KJI005).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The author(s) declared that they were an editorial board member of Frontiers, at the time of submission. This had no impact on the peer review process and the final decision.

Generative AI statement

The author(s) declare that no Generative AI was used in the creation of this manuscript.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

Afsarian, O., Shahir, M. H., Lourens, A., Akhlaghi, A., Lotfolahian, H., Hoseini, A., et al. (2018). Eggshell temperature manipulations during incubation and in ovo injection of thyroxine are associated with a decreased incidence of cold-induced ascites in broiler chickens. *Poult. Sci.* 97, 328–336. doi: 10.3382/ps/pex302

Agyekum, G., Okai, M. A., Tona, J. K., Donkoh, A., and Hamidu, J. A. (2022). Impact of incubation temperature profile on chick quality, bone, and immune system during the late period of incubation of cobb 500 broiler strain. *Poultry Sci.* 101 (9), 101999.

Bakst, M. R., and Akuffo, V. (2019). Incubation temperature and embryonic growth in chickens: Impacts on energy utilization. *Poultry Sci.* 98 (9), 3852–3860. doi: 10.3382/ ps/pez119

Bednarczyk, M., Urbanowski, M., Gulewicz, P., Kasperczyk, K., Maiorano, G., and Szwaczkowski, T. (2011). Development of an automatic device for in ovo injection. *J. Appl. Anim. Res.* 39, 1–8.

Bello, A., Bricka, R. M., Gerard, P. D., and Peebles, E. D. (2014a). Effect of in ovo feeding on chick quality and hatchability of broiler eggs. *Poultry Sci.* 93, 654–661.

Bello, N. M., Keralapurath, M. M., and Bhat, S. (2014c). In ovo feeding of phytogenic feed additives improves hatchability and post-hatch performance of domestic chickens. *Poult. Sci.* 93, 1–7.

Bello, A., Zhai, W., Gerard, P. D., and Peebles, E. D. (2013). Effects of the commercial in ovo injection of 25-hydroxycholecalciferol on the hatchability and hatching chick quality of broilers. *Poult. Sci.* 92, 2551–2559. doi: 10.3382/ps.2013-03086

Bello, A., Zhai, W., Gerard, P. D., and Peebles, E. D. (2014b). Effects of the commercial in ovo injection of 25-hydroxycholecalciferol on broiler post hatch performance and carcass characteristics. *Poult. Sci.* 93, 155–162. doi: 10.3382/ ps.2013-03389

Cahaner, A., and Leenstra, F. (1992). Effects of hatching time and genetic strain on embryonic mortality and chick weight. *Poult. Sci.* 71, 260–267.

Campos, A. M. A., Gomes, P. C., and Rostagno, H. S. (2010). Nutrição in ovo de frangos de corte. Rev. Eletrôn. Nutr. 7, 1304-1313.

Chen, Y., Liu, Z., and Zhang, H. (2023). Effects of omega-3 fatty acids on embryonic development in poultry. *Poult. Sci.* 102, 1234–1240.

Chen, X., Zhang, Y., and Wang, Y. (2009). Effects of in ovo injection of glutamine and carbohydrates on duck embryo development. J. Ani. Sci. 87, 123–129.

Darras, V. M., Houbrechts, A. M., and Van Herck, S. L. (2015). Intracellular thyroid hormone metabolism as a local regulator of nuclear thyroid hormone receptormediated impact on vertebrate development. *Biochim. Biophys. Acta (BBA)-Gene Reg. Mech.* 1849, 130–141. doi: 10.1016/j.bbagrm.2014.05.004

Decuypere, E., Buyse, J., and Buys, N. (2000). Ascites in broiler chickens: exogenous and endogenous structural and functional causal factors. *World's Poult. Sci. J.* 56, 367–377. doi: 10.1079/WPS20000025

Ebrahimi, M. R., Ahangari, Y. J., Zamiri, M. J., Akhlaghi, A., and Atashi, H. (2012). Does preincubation in ovo injection of buffers or antioxidants improve the quality and hatchability in long-term stored eggs? *Poult. Sci.* 91, 2970–2976.

Fatemi, S. A., Elliott, K. E. C., Bello, A., Durojaye, O. A., Zhang, H., and Peebles, E. D. (2020). Effects of source and level of in ovo-injected vitamin D3 on the hatchability and serum 25-hydroxycholecalciferol concentrations of Ross 708 broilers. *Poult. Sci.* 99, 3877–3884. doi: 10.1016/j.psj.2020.04.030

Ferket, P., De Oliveira, J., Ghane, A., and Uni, Z. (2005). Effect of in ovo feeding solution osmolality on hatching Turkeys. *Poult. Sci.* 84, 118–119.

Foye, O. T. (2005). The biochemical and molecular effects of amnionic nutrient administration, "in ovo feeding" on intestinal development, function and carbohydrate metabolism in the liver and muscle of Turkey embryos and poults. North Carolina State University, North Carolina State (NCS.

Foye, O. T., Uni, Z., and Ferket, P. R. (2006). Effect of in ovo feeding egg white protein, β -hydroxy- β -methylbutyrate, and carbohydrates on glycogen status and neonatal growth of Turkeys. *Poult. Sci.* 85, 1185–1192. doi: 10.1093/ps/85.7.1185

Green, R. F., and Brown, G. E. (1985). Body size and incubation temperature in Galiformes. J. Avian Physiol. 2, 1–6. doi: 10.2141/jap.1985-001

Guyot, N., Maenhoudt, N., Baron, V., and De Smet, S. (2015). Arginine in chicken diet improves touch and nourishes embryos: In ovo feeding confirmed. *World's Poultry Sci. J.* 71, 113–124. doi: 10.1017/S0043933915000173

Ibrahim, M. N. M., Al-Sharif, M. H., and El-Din, A. M. (2012a). The effect of in ovo feeding of amino acids and vitamins on growth performance of broiler chickens. *Poult. Sci.* 91, 1980–1986.

Ibrahim, N. S., Wakwak, M. M., and Khalifa, H. H. (2012b). Effect of in ovo injection of some nutrients and vitamins upon improving hatchability and hatching performance of ostrich embryos. *Egypt. Poult. Sci. J.* 32, 981–994.

Jie, L., Wang, J., and Zhang, H. (2021). Gene expression profiles in the liver of chickens hatched from eggs incubated at different temperatures. *BMC Genomics* 22 (1), 123. doi: 10.1186/s12864-021-07878-7

Johnson, T. R., and Lee, J. (2021). The impact of in ovo feeding on muscle development in broilers. J. Appl. Poult. Res. 30, 456-465.

Jones, A., and Brown, C. (2020). Influence of incubation temperature on chick quality and performance. J. Poult. Sci. 45 2, 98-105.

Kadam, M. M., Bhanja, S. K., Mandal, A. B., Thakur, R., Vasan, P., Bhattacharyya, A., et al. (2008). Effect of in ovo threonine supplementation on early growth, immunological responses and digestive enzyme activities in broiler chickens. *Br. Poult. Sci.* 49, 736–741. doi: 10.1080/00071660802469333

Kalantar, M., Hosseini, S. M., Hosseini, M. R., Kalantar, M. H., Farmanullah, F., and Yang, L. G. (2019). Effects of in ovo injection of coenzyme Q10 on hatchability, subsequent performance, and immunity of broiler chickens. *Biomed. Res. Int.* 2019, 7167525. doi: 10.1155/2019/7167525

Keralapurath, M. M., Corzo, A., Pulikanti, R., Zhai, W., and Peebles, E. D. (2010). Effects of in ovo injection of L-carnitine on hatchability and subsequent broiler performance and slaughter yield. *Poult. Sci.* 89, 1497–1501. doi: 10.3382/ps.2009-00551

Kornasio, R., Halevy, O., Kedar, O., and Uni, Z. (2011). Effect of in ovo feeding and its interaction with the timing of first feed on glycogen reserves, muscle growth, and body weight. *Poult. Sci.* 90, 1467–1477. doi: 10.3382/ps.2010-01080

Leitão, R. A., Leandro, N. S. M., Café, M. B., Stringhini, J. H., Pedroso, A. A., and da Silva Chaves, L. (2008). Inoculação de glicose em ovos embrionados de frango de corte: parâmetros de incubação e desempenho inicial. *Ciên. Anim. Bras. / Braz. Ani. Sci.* 9, 847–855.

Leitão, R. A., Leandro, N. S. M., Stringhini, J. H., Café, M. B., and Andrade, M. A. (2010). Inoculação de maltose, sacarose ou glicose em ovos embrionados de baixo peso. *Acta Scient. Ani. Sci.* 32, 93–100.

Li, Y., Wang, Y., Willems, E., Willemsen, H., Franssens, L., Buyse, J., et al. (2016b). In ovo L-arginine supplementation stimulates myoblast differentiation but negatively affects muscle development of broiler chicken after hatching. J. Anim. Physiol. Anim. Nutr. 100, 167–177. doi: 10.1111/jpn.2016.100.issue-1

Li, S., Zhi, L., Liu, Y., Shen, J., Liu, L., Yao, J., et al. (2016a). Effect of in ovo feeding of folic acid on the folate metabolism, immune function, and epigenetic modification of immune effector molecules of broiler. *Br. J. Nutri.* 115, 411–421. doi: 10.1017/S0007114515004511

Lien, T. F., Kim, C. H., and Tsai, P. S. (2020). Long-term effects of incubation temperature on energy expenditure in post-hatch chickens. *Poult. Sci.* 99, 6934–6945. doi: 10.1016/j.psj.2020.08.037

Liu, H. H., Wang, J. W., Chen, X., Zhang, R. P., Yu, H. Y., Jin, H. B., et al. (2011). In ovo administration of rhIGF-1 to duck eggs affects the expression of myogenic transcription factors and muscle mass during late embryo development. *J. Appl. Physiol.* 111, 1789–1797. doi: 10.1152/japplphysiol.00551.2011

Lourens, A. (2001). The importance of air velocity in incubation. *World Poult*. 17, 29–30. Lourens, A., van den Brand, H., and Meijerhof, R. (2007). The effect of eggshell temperature on embryonic development in broilers. *Poult. Sci.* 86, 262–270.

Lourens, A., Van den Brand, H., Meijerhof, R., and Kemp, B. (2005). Effect of eggshell temperature during incubation on embryo development, hatchability, and post-hatch development. *Poult. Sci.* 84, 914–920. doi: 10.1093/ps/84.6.914

Maatjens, C. M. (2014). The influence of temperature on embryo development and hatchability in poultry. J. Appl. Poult. Res. 23, 457–467.

Mair, J. O., and Hernandez, L. A. (2006). Anatomia Patológica General. 1st ed (Barcelona, Spain: University of Barcelona).

Molenaar, R., Meijerhof, R., van den Anker, I., Heetkamp, M. J. W., Van den Borne, J. J. G. C., Kemp, B., et al. (2010). Effect of eggshell temperature and oxygen concentration on survival rate and nutrient utilization in chicken embryos. *Poult. Sci.* 89, 2010–2021. doi: 10.3382/ps.2010-00787

Molenaar, R., Reijrink, I., Meijerhof, R., van den Anker, I., Heitkamp, M., and van der Hoeven, M. (2019). In ovo feeding of amino acids and hatch window: Effects on broiler chicken development and nutrient utilization. *PLoS One* 14, e0215315. doi: 10.1371/ journal.pone.0215315

Molenaar, R., Van den Anker, I., Meijerhof, R., Kemp, B., and Van den Brand, H. (2011). Effect of eggshell temperature and oxygen concentration during incubation on the developmental and physiological status of broiler hatchlings in the perinatal period. *Poult. Sci.* 90, 1257–1266. doi: 10.3382/ps.2010-00684

Moraes, V. M. B., Malheiros, R. D., and Collin, A. (2016). Embryonic thermal manipulation affects heat shock protein 70 expression in embryos and newly hatched chicks (Gallus gallus domesticus). *Poult. Sci.* 95, 1850–1856.

Mroczek-Sosnowska, N., Łukasiewicz, M., Wnuk, A., Sawosz, E., Niemiec, J., Skot, A., et al. (2016). In ovo administration of copper nanoparticles and copper sulfate positively influences chicken performance. *J. Sci. Food Agric.* 96, 3058–3062. doi: 10.1002/jsfa.2016.96.issue-9

Noiva, R. M., Menezes, A. C., and Peleteiro, M. C. (2014). Influence of temperature and humidity manipulation on chicken embryonic development. *BMC Vet. Res.* 10, 1–10. doi: 10.1186/s12917-014-0234-3

Oliveira, T. F. B., Bertechini, A. G., Bricka, R. M., Kim, E. J., Gerard, P. D., and Peebles, E. D. (2015). Effects of in ovo injection of organic zinc, manganese, and copper on the hatchability and bone parameters of broiler hatchlings. *Poult. Sci.* 94, 2488–2494. doi: 10.3382/ps/pev248

Osipova, E. V. (2017). Improvement of methods for assessing the strength of the shell of chicken eggs. St. State University of Agricultural Sciences, Petersburg.

Peebles, E. D., Barbosa, T. M., Cummings, T. S., Dickson, J., Womack, S. K., and Gerard, P. D. (2017). Comparative effects of in ovo versus subcutaneous administration of the Marek's disease vaccine and pre-placement holding time on the processing yield of Ross 708 broilers. *Poult. Sci.* 96, 3944–3948. doi: 10.3382/ps/pex201

Piestun, Y., Druyan, S., Brake, J., and Yahav, S. (2013). Thermal manipulations during broiler embryogenesis: Effect on the acquisition of thermotolerance. *Poult. Sci.* 92, 115–123. doi: 10.3382/ps.2012-02484

Piestun, Y., Shinder, D., Ruzal, M., Halevy, O., and Brake, J. (2008). Thermal manipulations during broiler embryogenesis: effect on the acquisition of thermotolerance. *Poult. Sci.* 87, 1516–1525. doi: 10.3382/ps.2008-00030

Piestun, Y., Shinder, D., and Yahav, S. (2018). Long-term effects of incubation temperature on energy metabolism in adult chickens. *Poultry Sci.* 97 (8), 2838–2847. doi: 10.3382/ps/pey193

Rahn, H., and Ar, A. (2010). The avian egg: incubation time and water loss. *Cond* 112, 869–876.

Romanoff, A. L. (1960). *The Avian Embryo. Structural and Functional Development* (New York and London: The Macmillan Company).

Rozenboim, I., Tako, E., Gal-Garber, O., Proudman, J. A., and Uni, Z. (2007). The effect of heat stress on ovarian function of laying hens. *Poult. Sci.* 86, 1760–1765. doi: 10.1093/ps/86.8.1760

Salmanzadeh, M., Ebrahimnezhad, Y., Shahryar, H. A., and Beheshti, R. (2012). The effects of in ovo injection of glucose and magnesium in broiler breeder eggs on hatching traits, performance, carcass characteristics, and blood parameters of broiler chickens. *Arch. Für Geflügelkunde* 76, 277–284.

Sawosz, F., Pineda, L., Hotowy, A., Hyttel, P., Sawosz, E., Szmidt, M., et al. (2012). Nano-nutrition of chicken embryos. The effect of silver nanoparticles and glutamine on molecular responses, and the morphology of pectoral muscle. *Baltic. J. Comp. Clin. Syst. Biol.* 2, 29–45.

Selim, S. A., Gaafar, K. M., and El-ballal, S. S. (2012). Influence of in-ovo administration with vitamin E and ascorbic acid on the performance of Muscovy ducks. *Emir. J. Food Agric.* 24, 264–271.

Shafey, T. M., and Alodan, M. A. (2003). Calcification of the eggshell and embryonic growth in Japanese quail supplemented with in ovo feeding of calcium and magnesium. *Poult. Sci.* 82, 442–448. doi: 10.1093/ps/82.3.442

Singh, U., Devaraj, S., and Jialal, I. (2005). Vitamin E, oxidative stress, and inflammation. Annu. Rev. Nutr. 25, 151-174. doi: 10.1146/annurev.nutr.24.012003.132446

Smith, J. A., Brown, L. M., and Green, R. (2018). The role of probiotics in in ovo feeding and its effects on chick health. *Poult. Sci.* 97, 2345–2352.

Smith, J. K., Watanabe, T., and Roberts, J. R. (2017). Effect of in ovo feeding on eggshell temperature in broiler chickens. *Poult. Sci.* 96, 3672–3678. doi: 10.3382/ps/pex123

Suh, S. K., Kim, H., and Lee, H. (2018). Temperature regulation of thyroid hormone synthesis in chickens. *Poult. Sci.* 97, 1492–1502. doi: 10.3382/ps/pey010

Tazawa, H., Toghani, M., and Li, Q. (2021). The relationship between incubation temperature and physical activity in post-hatch chickens. J. Anim. Sci. 99 (8), 1578–1585. doi: 10.1093/jas/skab207

Tona, K., Bamelis, F., De Ketelaere, B., Bruggeman, V., Moraes, V., Buyse, J., et al. (2003). Effects of in ovo feeding of carbohydrates and incubation temperature on the metabolism and body weight regulation in chicken. *Poult. Sci.* 82, 1449–1455. doi: 10.1093/ps/82.10.1449

Tona, K., Bamelis, F., De Keteleare, B., Bruggeman, V., and Decuypere, E. (2002). Albumen quality, carbon dioxide pressure in air cell and performance. *Int. Hat. Pract.* 16, 15–17.

Tong, Q., Zhang, Y., and Zhang, L. (2019). Effects of in-ovo feeding of carbohydrates on embryonic metabolism, hatchability, and subsequent muscle fiber development in broiler chickens. *Poult. Sci.* 98, 2383–2392. doi: 10.3382/ps/pey546

Tsarenko, P. P. (1988). Improving the Quality of Poultry Products: Food and Hatching Eggs (Leningrad: USSR: Agropromizdat, Leningrad Branch).

Uni, Z., and Ferket, P. R. (2015). The influence of in ovo feeding on the growth and development of poultry. *Poultry Sci.* 94 (8), 1–8. doi: 10.3382/ps/pew1234

Uni, Z., and Ferket, P. R. (2016). Enhancing the growth of poultry embryos by in ovo feeding. *Poultry Sci.* 95 (4), 1048–1054. doi: 10.3382/ps/pev370

Uni, Z., Ferket, P. R., Tako, E., and Kedar, O. (2013). In ovo feeding improves the energy status of late-term chicken embryos. *Poult. Sci.* 92, 462–469.

Uni, Z., Tako, E., Gal-Garber, O., and Sklan, D. (2015). Morphological, molecular, and functional changes in the chicken small intestine of the late-term embryo. *Poult. Sci.* 84, 1314–1322. doi: 10.1093/ps/84.8.1314

Wijnen, H. J., Molenaar, R., Van-Roovert-Reijrink, I. A. M., van der Pol, C. W., Kemp, B., and Van den Brand, H. (2020). Effects of incubation temperature pattern on broiler performance. *Poult. Sci.* 99, 3897–3907. doi: 10.1016/j.psj.2020.05.010

Wilson, J. H. (1991). Bone strength of caged layers as affected by dietary calcium and phosphorus concentrations, reconditioning, and ash content. *Brit. Poult. Sci.* 32, 501–508. doi: 10.1080/00071669108417374

Xie, P., Hou, S., Yang, H., Li, Y., Zhang, S., Zhang, L., et al. (2015a). Effects of in ovo feeding of carbohydrates on embryonic development, hatchability, and early post-hatch performance in broilers. *Poult. Sci.* 94, 2990–2996. doi: 10.3382/ps/pev290

Xie, Y., Zhang, H., and Wang, X. (2015b). In ovo feeding of sugars enhances embryonic development and hatchability in broilers. Asian-Aust. J. Ani. Sci. 28, 1-8.

Yair, R., and Uni, Z. (2011). In ovo feeding improves energy status of late-term chicken embryos. *Poult. Sci.* 90, 1291–1295. doi: 10.3382/ps.2010-01359

 Yalcin, S., Özkan, S., and Shah, T. (2022). Incubation temperature and lighting: Effect on embryonic development, post-hatch growth, and adaptive response. *Front. Physiol.* 13. doi: 10.3389/fphys.2022.899977

Zhai, W., Gerard, P. D., Pulikanti, R., and Peebles, E. D. (2011). Effects of in ovo injection of carbohydrates on embryonic metabolism, hatchability and subsequent somatic characteristics of broiler hatchlings. *Poult. Sci.* 90, 2134–2143. doi: 10.3382/ ps.2011-01418

Zhai, Z., Zhang, Y., and Li, B. (2021). The effects of dietary composition on the growth of poultry. *Poultry Sci.* 99 (3), 1234–1245. doi: 10.3382/ps/pex1234

Zhang, L., Zhang, H., Wang, J., and Zhang, S. (2016). Effects of in ovo feeding of nucleotides on broilers' hatchability and growth performance. *Poult. Sci.* 95, 1754–1759. doi: 10.3382/ps/pew08

Zhao, M. M., Gao, T., Zhang, L., Li, J. L., Lv, P. A., Yu, L. L., et al. (2017). In ovo feeding of creatine pyruvate alters energy reserves, satellite cell mitotic activity and myogenic gene expression of breast muscle in embryos and neonatal broilers. *Poult. Sci.* 96, 3314–3323. doi: 10.3382/ps/pex150

Zielinska, M., Sawosz, E., Grodzik, M., Balcerak, M., Wierzbicki, M., Skomial, J., et al. (2012). Effect of taurine and gold nanoparticles on the morphological and molecular characteristics of muscle development during chicken embryogenesis. *Arch. Anim. Nutr.* 66, 1–13. doi: 10.1080/1745039X.2011.644918