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Selection of microhabitats, plants, and plant parts eaten by a threatened tortoise: observations during a superbloom

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Populations of the threatened desert tortoise (Gopherus agassizii) continue to decline throughout the geographic range, in part because of degraded and fragmented habitats in the Mojave and western Sonoran deserts. The species is herbivorous and highly selective in choice of plant species. To increase options for recovery, we analyzed behaviors, patterns of movements while foraging, and parts of plants consumed during a superbloom. We characterized foraging routes and the habitat strata and microhabitats where tortoises traveled to eat preferred wildflower species. Tortoises walked one foraging route per day in early spring, often switched to two routes per day in middle and late spring with rise of midday temperatures. They chose habitat strata (primarily hills and ephemeral stream channels) and three of seven microhabitats for foraging on preferred food plants. Preferred microhabitats were intershrub open space and small (1-2 m wide) ephemeral stream channels. They rarely took bites of forbs growing under and in the dripline of shrubs or nonnative forbs and grasses. Tortoises typically did not select specific plant parts to eat but important exceptions occurred. For example, they usually ignored the inflorescences of the annual Eremothera boothii and, when eating the non-native annual Erodium cicutarium, tended to focus on fruits. All such information aids recovery efforts to restore declining tortoise populations.

KEYWORDS

foraging, Gopherus agassizii, microhabitats, movements, plant parts

1 Introduction

Agassiz's desert tortoise (*Gopherus agassizii*, herein tortoise) is a federally threatened species under the Endangered Species Act (U.S. Fish and Wildlife Service [USFWS], 1990) and is a globally critically endangered species (Berry et al., 2021). Despite recovery efforts, most populations are no longer viable (U.S. Fish and Wildlife Service [USFWS], 2015; Berry and Murphy, 2019). Allison and McLuckie (2018) noted that the desert tortoise was on the path to extinction under current conditions. The status is primarily the result of

habitat degradation, fragmentation, and loss; disease and hyperpredation are also contributing to declines (Berry and Murphy, 2019). The sources of habitat degradation and loss include urban and agricultural development, roads and utility corridors, mining, livestock grazing, military activities, authorized and unauthorized off-road vehicle use, and many other anthropogenic activities. Disturbances to soil and native plant communities contributed to invasion and establishment of non-native forbs and combustible grasses, which created a new fire cycle in the Mojave Desert (Brooks and Matchett, 2006; Klinger et al., 2021). Recovery of tortoise populations and restoration of deteriorated land in the Mojave and western Sonoran deserts where desert tortoises live present major challenges (Abella et al., 2023). Tortoises are long-lived and slow to reproduce; mortality rates are high for young individuals (Woodbury and Hardy, 1948; Hardy, 1976; Turner et al., 1987; Medica et al., 2012). Additionally, they are herbivorous, highly selective in choice of plant foods, and rely on rains for free water to drink (Burge and Bradley, 1976; Jennings and Fontenot, 1993; Henen et al., 1998; Jennings and Berry, 2015). The amounts and timing of precipitation during fall and winter months drive germination, growth, and flowering of ephemeral plants during the subsequent spring in the western Mojave Desert (Rowlands et al., 1982; Jennings, 2001). In years of low or no rainfall, little or no food is available, whereas in years of high precipitation, a superbloom may result with abundant wildflowers (Jennings, 2001; Brooks and Berry, 2006; Jennings and Berry, 2015).

After the federal listing of the tortoise as threatened in 1990 (U.S. Fish and Wildlife Service [USFWS], 1990), several observational and experimental research projects were undertaken to determine the plants tortoises consumed and avoided, because of threats to habitat from such disturbances as livestock grazing (Webb and Stielstra, 1979; U.S. Fish and Wildlife Service [USFWS], 1990; U.S. Fish and Wildlife Service [USFWS], 1994). Research on the digestibility and nutritional characteristics of potential plants available to wild desert tortoises provided answers to why tortoises selected and preferred certain species in altered habitats (e.g., Brooks and Berry, 2006). Barboza (1995), in experimental studies of feeding different diets to tortoises, reported loss of body mass from the grass diet, a finding later supported by feeding trials with both native and non-native annuals (forbs and grasses) for juvenile tortoises (Hazard et al., 2009; Hazard et al., 2010; Drake et al., 2016). Further, young tortoises gained weight and grew when eating forbs but did not thrive on either native or nonnative grasses (Hazard et al., 2009; Hazard et al., 2010). Jennings (2002) and Jennings and Berry (2015) detailed the preferences of wild tortoises for selected species of forbs and herbaceous perennial plants using observations of bite counts by species during a wet year. Oftedal et al., (1994, Oftedal, 2002; Oftedal et al., 2002) explained the selectivity for a few species of forbs by content of potassium, ubiquitous in desert plants and difficult to metabolize without free water. Wild tortoises needed to select ephemeral plants high in water and nitrogen but low in potassium to maintain a healthy diet, grow, and reproduce (Oftedal et al., 1994). Oftedal (2002) developed the Potassium Excretion Potential (PEP) index of a plant: a positive PEP index occurs when a plant contains more nitrogen and water than is needed to excrete the ingested potassium, whereas a negative PEP signified that a plant contained insufficient protein and water for a tortoise to dispose of ingested potassium (Oftedal, 2002). Observational studies indicated that tortoises were highly selective in choices of annual and herbaceous plant species (Jennings, 2002; Jennings and Berry, 2015) and and favored species low in potassium (Oftedal, 2002).

Our objectives were to build off previous work on tortoise foraging by characterizing preferences as plant resource availability and phenology changed throughout the spring in the western Mojave Desert (Jennings and Berry, 2015). Here we address foraging behaviors and choices of plant parts consumed by the tortoises during a year with a superbloom and when confronted with a suite of choices of plant foods. We asked three questions about foraging in spring during a superbloom: 1) do tortoises walk among habitat strata in a random manner to forage, 2) do distances walked within each habitat stratum change as phenological stages of plants change, and 3) do tortoises randomly select microhabitats to forage? In addition, for each plant species, we estimated the percentage of the plants consumed and relative proportions eaten of different plant parts (stems, leaves, flowers/buds, and fruits). We evaluated occurrences and proportions of years of high and low precipitation before and after the study. Our findings will contribute to the knowledge of tortoise movements and foraging behaviors, important elements in critical recovery efforts to build viable populations (U.S. Fish and Wildlife Service [USFWS], 2011).

2 Study area

The study was conducted on a ~15 km² site within the Desert Tortoise Research Natural Area (DTRNA; 35°14'34", -117°51'45"), at elevations of 800–915 m, in the western Mojave Desert, California, USA. The ~101 km² area received designation in 1980 by U.S. Congress and was protected in 1978–1979 with a fence to exclude recreational vehicle use and sheep grazing (Berry et al., 2014). The vegetation alliance of perennial shrubs was diverse with 11 species, including creosote bush (*Larrea tridentata*), white bursage (*Ambrosia dumosa*), Nevada ephedra (*Ephedra nevadensis*), goldenhead (*Acamptopappus sphaerocephalus*), Mojave indigo bush (*Psorothamnus arborescens*), Mojave California buckwheat (*Eriogonum fasciculatum*), Anderson boxthorn (*Lycium andersonii*), winterfat (*Krascheninnikovia lanata*), hop-sage (*Grayia spinosa*), and Mojave aster (*Xylorhiza tortifolia*).

3 Materials and methods

3.1 Data collection

3.1.1 Precipitation

The study area is in the western region of the Mojave Desert, where most rainfall occurs during fall and winter and can produce, depending on amounts of rain, annual plants in late winter and spring (Rowlands et al., 1982; Rowlands, 1995). We used highresolution gridded data on climate that were geographically and elevation-weighted from PRISM to summarize monthly estimates of precipitation from 1961 to the present. PRISM is a gridded data product (Daly et al., 2008) using elevation, latitude, and longitude of the study area (2) at 4-km resolution from the Prism Climate Group (https://www.prism.oregonstate.edu, accessed 5 April and 10 October 2023). We recognize that several variables affect climate in the American Southwest—precipitation, temperature, and soil moisture are probably the most important (Ault et al., 2016; Cook et al., 2021; Dannenberg et al., 2022). Because amounts of winter and summer precipitation are prime drivers of almost every aspect in the lives of tortoises, we used precipitation here instead of other variables (e.g., Henen, 1997; Henen et al., 1998; Christopher et al., 1999; Duda et al, 1999; Henen, 2002; Berry et al., 2023).

3.1.2 Desert tortoises

Field work occurred between 26 March and 23 June 1992. One person (WBJ) recorded all daily movements and foraging behaviors for 16 adult tortoises with an average (±SE) carapace length (mm) at the midline for eight males of 230 ±14.42 mm (range 179–280 mm) and eight females, 222 ±4.14 mm (range 210–239 mm). Twelve of the study animals were part of another study and carried radio transmitters (Model 2B-2 AVM Instrument Co., Livermore, California, powered by lithium C-cells, Tadiran Co., Israel) with 48-cm copper antennae affixed to the shells (Christopher et al., 1999). Using a receiver and a 2-element antenna (Telonics, Mesa, Arizona), selected individuals were located each day (see details below). Data were also collected opportunistically on movements and foraging for four incidentally encountered adults without radio transmitters.

Throughout the field season, a focal animal was selected to observe and record foraging and movements for each field day. When possible, observations of foraging were recorded for tortoises over the entire course of the daily or bi-daily foraging route, i.e., a complete foraging route = from emergence from a cover site (pallet or burrow) to retreat to a cover site after foraging ended. Typically, tortoises emerged from cover sites during morning hours. If the selected focal animal did not emerge, an alternate was selected, usually an individual inside or outside a cover site. The footpath (m) distances walked by each tortoise during foraging routes were also measured for each of three habitat strata (% of occurrence within the study area): 1) broad and sandy alluvial fans (48%); 2) low, rocky hills (42%); and 3) dry, sandy, ephemeral stream channels (10%) (Jennings and Berry, 2015). Tortoises used two patterns of movement on foraging routes: 1) round trips, with returns to the same cover site occupied earlier in the day and generally remaining within the same habitat stratum to forage; and 2) non-round trips in which a tortoise left a cover site to forage, traveled among >1 habitat strata and ended foraging at a different cover site. On several occasions, observations were recorded for multiple tortoises during the same day, e.g., observations of foraging activities of one individual in the morning and another during the afternoon-thus recording data for the foraging route of each tortoise. Observations were also recorded simultaneously for two to three individuals foraging <10 m apart when all were feeding on the same plant species. Focal individuals were followed at distances of 4–8 m, frequently using binoculars, which minimized disturbance to the tortoise.

While watching a tortoise, data were recorded onto a microcassette recorder (Sony M560V, China): start and end times of observations; times for each foraging bout (1 bout = all bites taken from a single food item); species of plant eaten; number of bites taken (Jennings and Berry, 2015); the estimated percentage of the plant eaten (above-ground biomass only); parts of the plant consumed (leaves, stems, flowers, fruits); condition of the plant (succulent or dry); and the microhabitat where the plant grew. Note that our estimates of the percentage of plant eaten are based on simple visual estimates of the portion of a plant eaten (i.e., visual comparisons of a plant before and after a tortoise fed from it). These estimates provide data on whether tortoises ate a plant in its entirety, ate part of it, or just took a few bites, etc. The microhabitats were: 1) beneath creosote bush; 2) at or under the dripline of a creosote bush; 3) intershrub space between shrubs; 4) outer edge of species of non-creosote bushes; 5) margin of small (1-2 m wide) ephemeral stream channels, 6) center of small stream channels; and 7) sandy areas within large ephemeral stream channels (>2 m wide). The recognition of two distinct microhabitat categories for the small stream channels was warranted because tortoises showed a strong tendency to walk along the edges of small stream channels where some preferred plants grew (see Jennings and Fontenot, 1993). Plant nomenclature was from the Jepson Flora Project (2023). Data for plants available for forage are in Jennings and Berry (2015).

3.2 Analyses of data

3.2.1 Precipitation

We evaluated precipitation data monthly for winter rain (October 1 - March 31) and for the hydrologic year (October 1 -September 30). We identified three blocks of years for analysis of annual and winter rainfall averages: the 30 years prior to the study (1961-1990) as a benchmark; the eight years, 1991-1998, bracketing when the study occurred and before the start of the megadrought; and from the start of the megadrought in 1999 through the potential end point in 2022 (Ault et al., 2016; Williams et al., 2020; Cook et al., 2022). According to Ault et al. (2016: abstract), "Megadroughts are comparable in severity to the worst droughts of the 20th century but are of much longer duration." We identified numbers of years with: 1) unusually high winter precipitation, when superblooms occurred, which we defined as winter rainfall exceeding the long-term averages by $\geq 70\%$; 2) above average, average, and slightly below average precipitation; and 3) dry years or droughts for the tortoise, when winter precipitation fell well below long-term averages.

3.2.2 Phenological periods

To account for seasonal variation in availability of food plants and plant parts, we analyzed foraging behaviors and movement patterns within each of three successive phenological periods in spring, defining them as: early spring, characterized by early-spring species of blooming plants in March and April; middle spring with the middle-spring bloomers in May; and late spring, the late-spring bloomers in the final phase of the spring wildflower season in June (Jennings and Berry, 2015).

3.2.3 Selection of habitat strata by tortoises

Observations from the pilot study of tortoise foraging behavior conducted the previous year at the same site suggested that the tortoises walk among different habitat strata during their daily or bidaily foraging walks or travels (Jennings and Fontenot, 1993). We therefore tested the null hypothesis that tortoises randomly walked among habitat strata while they were foraging. To perform this test, we compared the distances (m) walked by tortoises within each of three strata to the randomly expected values (derived using the relative coverages of each stratum) using a χ^2 test. Data for each phenological period were tested separately. We addressed the question whether distances walked by tortoises within each habitat stratum varied across phenological periods using a Kruskal-Wallis test; we tested each habitat stratum with phenological period as the independent variable. We addressed the question whether lengths of foraging routes differed among phenological periods by conducting a Kruskal-Wallis test with phenological period as the independent variable. Tests yielding significant P-values were followed with post hoc pairwise comparisons using Wilcoxon rank sum tests to determine which group(s) were statistically different from other.

3.2.4 Selection of microhabitats by tortoises

We addressed the question whether tortoises randomly selected microhabitats for foraging by comparing the expected frequencies of plants consumed (assumed to be equal among the seven microhabitats) vs. observed frequencies using a χ^2 test. We analyzed each phenological period separately. All statistical tests were conducted using the R statistical package (version 4.3.0, R Development Core Team, 2023; https://cran.r-project.org/ accessed June 15, 2023). For χ^2 and Kruskal-Wallis tests, we designated $\alpha = 0.05$ as the level of significance. In all *post hoc* Wilcoxon rank sum tests, we adjusted α using a Bonferroni correction.

3.2.5 Patterns of plant consumption

We estimated the relative percentage of each anatomical part consumed (i.e., flowers, leaves, stems, and fruits) by plant species using the equation:

% plant part consumed =
$$\left[\frac{\text{No. plants for which a particular part was eaten}}{\text{Total no. of plants of that species eaten by tortoises}}\right] \times 100$$
 (1)

Each species was analyzed separately for each phenological period. We did not test these data statistically because of interindividual variation in the phenological states of the plants.

4 Results

4.1 Precipitation

During the study in the spring of 1992, the fall-winter rainfall was 240.5 mm, 96.2% above the long-term average of 122.59 mm for winter, calculated for the previous 30 years. The result was a superbloom of wildflowers. In the 30 previous years, three superblooms occurred, an average of one per decade (Table 1). In comparison, in the 1990s, eight years prior to the beginning of the megadrought, five high rainfall years and superblooms occurred. These eight years were unusually wet compared to the years prior to the study. During the ensuing megadrought (1999–2023 and possibly ongoing), five of 22 winters of high rainfall produced superblooms.

4.2 Tortoises

Tortoise foraging and movement data were obtained for 52 observation days: 24 in early spring, 20 in middle spring, and eight in late spring. The average (\pm SE) number of foraging bouts was 244.1 \pm 170.8 (range 2–1,415) for the eight male tortoises and 133.3 \pm 65.5 (range 2–568) for the eight females. We obtained data for 40 foraging routes among the three phenological periods.

Years	No. years	Average precipitation, mm (range)		% fall- winter rain	Superbloom frequency/ span of years	Drought frequency/ span of years		
		Annual	Fall-winter					
1961– 1990	30	148.6 (38.9–379.9)	122.6 (35.6–313.5)	82.5	3/30	12/30		
1991– 1998	8	214.8 (81.6-336.1)	191.7 (67.0–313.5)	89.2	5/8	1/8		
1999– 2023	22	142.5 (45.3–347.0	130.9 (43.0-316.9)	91.9	5/22	7/22		

TABLE 1 Frequency of years with high fall-winter precipitation and associated superblooms at the Desert Tortoise Research Natural Area, western Mojave Desert, California, USA, in the 30 years before and after the study, including the 1999–2023 megadrought.

We defined drought as a dry year below the fall-winter average of 131 mm, based on available preferred forbs for the tortoises to eat.

4.3 Foraging among habitat strata and movements

During early spring, 16% of foraging routes were in alluvial fans, 68% were in hills, and 16% were in ephemeral stream channels. These values differed from the random expected percentages of 48%, 42%, and 10%, respectively ($\chi^2 = 2054.8$, df = 2, *P* = 0.001). The patterns of movements for foraging routes differed during the three phenological periods. In early spring, tortoises tended to embark on foraging routes from a cover site in one stratum and continued through >1 strata-usually hills, where they fed on solitary Mirabilis laevis, to small ephemeral stream channels, where they ate small groups of Astragalus layneae-before terminating at a second cover site. In middle spring, tortoises again selected foraging routes, because 88% were in hills, and 12% were in ephemeral stream channels ($\chi^2 = 2199.9$, df = 2, P = 0.001). They departed from and returned to the same cover sites, usually within the hill stratum, where they foraged on Acmispon brachycarpus or clusters of Prenanthella exigua. Eleven of the 12 recorded foraging routes only occurred in the hill stratum, whereas one was in a small ephemeral stream channel. In late spring, tortoises took roundtrips from the same cover site to forage on patches of Chamaesyce albomarginata in the ephemeral stream channel stratum (37% more than expected), while they walked in alluvial fans (41%) and hills (22%) less than expected ($\chi^2 = 841.7$, df = 2, P = 0.001). Although the tortoises frequently walked along stream channels while they were foraging, there was no standing surface water in these channels-or anywhere else on the study site -during the entire spring activity season.

4.4 Within seasonal variations in movements

The average length of foraging routes was 268 ± 45.7 m (n = 18) in early spring, 185 ± 39.9 m (n = 12), and 161 ± 37.8 m (n = 6) in middle and late spring periods, respectively. The lengths of foraging routes did not vary during the three periods (Kruskal-Wallis test, P = 0.424).

Distances of foraging routes differed by habitat strata and phenological period. Within the alluvial fan stratum, distances varied among spring periods (Kruskal-Wallis test, P = 0.035). During middle spring, tortoises completely avoided alluvial fan areas, whereas they foraged in this stratum during the early and late spring periods (*post hoc* Wilcoxon rank sum tests, P = 0.025, P = 0.012, respectively). The distances walked in the alluvial fan stratum between early and late spring did not differ (*post hoc* Wilcoxon rank sum tests, P = 0.710). In contrast, tortoises showed no difference in the distances of foraging routes within hill and stream channel strata over the entire spring season (Kruskal-Wallis tests, P = 0.078 and P = 0.070, respectively).

4.5 Selective use of microhabitats

Foraging in microhabitats varied depending on the phenological period. During early spring, tortoises fed on plants

growing in the intershrub space microhabitat and mostly ignored plants in other microhabitats ($\chi^2 = 1720.5$, df = 6, *P* = 0.001). Nearly 60% of the plants eaten were in the intershrub space, including some preferred species, i.e., Mirabilis laevis and Astragalus didymocarpus. Other preferred forage species, particularly A. layneae and Eremothera boothii, were in small and large sandy ephemeral stream channel microhabitats, respectively. These microhabitats provided open spaces for foraging. In middle spring, nearly 90% of plants consumed were in the intershrub space microhabitat ($\chi^2 = 6363$, df = 6, *P* = 0.001). Comparable to the early spring period, tortoises fed extensively on the herbaceous perennial A. layneae along the margins of small stream channel microhabitats. They rarely foraged in dripline and canopy microhabitats. During the late spring period, tortoises continued to feed on plants primarily located in the intershrub space (~60%) and small stream channel (~37%) microhabitats (χ^2 = 1041.8, df = 6, P = 0.001). The most frequently eaten species was the herbaceous perennial Chamaesyce albomarginata, which appeared restricted to intershrub space and small stream channel microhabitats.

4.6 Plant parts consumed by tortoises

Tortoises clipped off bite-sized pieces of plants before swallowing them. The tortoises only fed on above-ground plant parts and never pulled plants from the ground or ate roots. In early spring, tortoises, on average, consumed 46% of aboveground biomass from each plant encountered (Table 2). On average, tortoises ate ~60% (range, 59-63%) of flowers, leaves, and stems on each plant across all forage species (Table 2). In contrast, they only consumed fruits on ~9% of each plant across all forage species (Table 2). Though tortoises appeared to generally eat food plants indiscriminately, close inspection of the data suggested that they were selective about the plant parts they consumed for some forage plants. For example, tortoises ate fruits from: 36 of the 40 (90%) Erodium cicutarium plants, 9 of 11 Pectocarya spp. individuals, and both individuals of Tropidocarpum gracile (Table 2). We repeatedly observed an apparent avoidance of flowers, leaves, and stems of E. cicutarium, a non-native species (Table 2). When tortoises encountered Eremothera boothii, the fourth favored forage species by bite counts, they bit off the entire inflorescence, which dropped to the ground, before eating other parts of the plant (Table 2). This behavior was not observed for other forage species. It is important to add that none of the preferred food plants were so large that they were inaccessible to the tortoises (e.g., fruits).

Throughout middle spring, tortoises maintained a pattern of consuming nearly half (41%) of each forage plant (Table 3); that is, they almost always consumed the leaves, stems, and fruits of *Acmispon brachycarpus*, a species that largely finished flowering but remained green and succulent (Table 3). Similarly, whenever tortoises fed on the annual *Prenanthella exigua*, the second most-preferred plant species by bite counts, they generally only ate flowers and stems, the only available parts (Table 3). Some foraging behaviors observed in early spring were repeated during middle spring (e.g., the apparent preference for the fruits of *E. cicutarium* and avoidance of flowers of *E. boothii*; Table 3). Overall,

TABLE 2 Characteristics of winter annual and herbaceous perennial species of plants and plant parts consumed by adult desert tortoises (*Gopherus agassizii*) during early spring (the first phenological period: March and April) at the Desert Tortoise Research Natural Area, western Mojave Desert, California, USA, in 1992.

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Immediate leaderTotalTotalSolutionSolutionSolutionSolutionSolutionPlanckia kanacetifika6682	Astragalus layneae ¹	1,704	35	37	23 (66)	32 (91)	19 (54)	0 (0)	
Dencilia namerifoliaGeneGeneGeneGeneMarcialia app.GeneGeneGeneGeneGeneGeneAnsinckia tessellataGeneGeneGeneGeneGeneGeneGeneGlia minorGeneGeneGeneGeneGeneGeneGeneGeneGeneGlia minorGeneGeneGeneGeneGeneGeneGeneGeneGeneGeneGlia minorGeneGeneGeneGeneGeneGeneGeneGeneGeneGeneGene mathematicGeneGeneGeneGeneGeneGeneGeneGeneGeneGeneSylackar pathematicGeneGeneGeneGeneGeneGeneGeneGeneGeneGeneAllacotrix contrarGeneGeneGeneGeneGeneGeneGeneGeneGeneGeneSylackar pathematicGeneGeneGeneGeneGeneGeneGeneGeneGeneGeneSylackar pathematicGeneGeneGeneGeneGeneGeneGeneGeneGeneGeneGeneSylackar pathematicGene<	Astragalus didymocarpus	1,509	123*	64	123 (100)	120 (98)	120 (98)	0 (0)	
Manzala app.155257711(%)17(%)14(%)00(0)Aminakia tasadiati1751082(2)16(2)82(3)0.0(0)Gila minor13421227251(9)46(8)51(9)11(2)Endum cicatarian ² 12814074.0(0)10.02382(3)36(60)Tringterm palmeri126126730.0026(00)11(4)6(23)Spicine palmeri1261269550(8)40(8)40(8)0(0)Malacothric coulteri12131366417(10)11(10)10(10)0(0)Spicine palmeri12131489991(3)11(10)11(10)0(0)Spicine palmeri12131213122131(10)21(13)14(1)0(0)Spicine palmeri1119121313820(0)13(10)14(1)0(0)Spicine palmeri1119121313820(0)13(10)14(1)0(0)Spicine palmeri1119121313820(0)13(10)14(1)0(0)0(0)Spicine palmeri1119121313820(0)13(10)14(10)14(10)14(10)Spicine palmeri1119121313820(0)36(10)14(10)14(10)14(10)Spicine palmeri1119121314810(0)36(10)14(10)14(10)14(10)14(10)Spicine palmeri1119141149141	Eremothera boothii	708	96	65	0 (0)	95 (99)	44 (46)	0 (0)	
Anisckia scalaar173129108(2)26(0)8(28)0(0)Glia minor13421527251(98)14(88)51(98)11(3)Endian cicatarian?1281140744(00)101(3)8(20)36(00)Tripprem paineri1266126730.0026(100)11(4)6(23)Splochin paineri12051569550(89)49(88)49(88)0(0)Malcothris conteri12111386417(19)16(169)11(10)0(0)Lapina odoratus12111386417(10)11(10)11(10)0(0)Anisoph Inducycarpa201113899(50)18(100)11(10)0(0)Splanoneris parys*13131221310021(15)14(19)0(0)Glanastir fremonti111912131820(95)11(52)14(19)0(0)Caselastrun schotti7791331221310026(100)26(10)16(10)0(0)Lacelastrun schotti77913414914(3)86(73)16(10)0(0)0(0)Caselastrun schotti775140144140014(10)16(10)16(10)Lanasha dichotmas668878810(10)10(10)10(10)10(10)Splanoneris park75140144140014(10)10(10)10(10)Splanoneris park675140140014(10)10(10)10(Phacelia tanacetifolia	682	34	51	29 (85)	29 (85)	30 (88)	0 (0)	
Idia minor34237251(88)44(88)51(98)11(1)Evolum cicitariun'2814074(10)10(5)8(20)3(6)0)Terapteron palmeri226266730(0)26(10)11(4)6(23)Sylocline paloarpholder22555695960(9)49(88)49(88)0(0)Malacadrix coulteri21171186411(10)11(00)11(10)0(0)Aninjon bradycarpus20101183630(0)3(100)0(0)3(100)0(0)Suphanoniria paryt21312113(8)9(10)3(10)11(10)0(0)Suphanoniria paryt21412182213(10)21(1)3(10)0(0)Malacubrix gloiruia21751182213(10)21(1)3(10)0(0)Cheenactis fremoriti2192826(0)3(10)10(0)0(0)0(0)Cryptartha circumcisa1792890(0)8(10)0(0)0(0)Potodarpus gocille27514890(0)8(10)10(0)0(0)Cryptartha circumcisa17928290(0)8(10)0(0)0(0)Potodarpus gocille275216090(0)10(0)10(0)10(0)Cryptartha circumcisa17928290(0)8(10)10(0)10(0)Potodarpus gocille27521600(0)0(0)0(0)0(0)0(0)	Mentzelia spp.	558	25	37	21 (84)	17 (68)	14 (56)	0 (0)	
Inclusion28840074(10)10(25)8(20)3(30)Tricipieron palmeri22625300025(100)1(4)6(23)Splocine paloeartheide2231669550.08944.08844.0880.000Malacobrix conteri217111864117(40)16.09915.0390.000Amipon bradycarpas22011380.00131.00010.0000.000Siphanomeia parry111311889.90.60011.00011.0000.000Malacobrix gabrata211321213.00021.01014.0000.000Chanactis frementi111921418.020.00020.0000.000Coptandae circumcisa177918.00.008.0000.0000.000Potadage variat77018.00.0008.0000.0000.000Potadage variat77521.021.014.014.0014.000.000Potadage variat77514.014.014.0014.0014.000.000Potadage variat77521.021.00021.00021.00021.00014.0014.00Potadage variat77521.021.021.00021.00021.00021.00021.000Potadage variat77521.021.021.00021.00021.00021.00021.000Potadage variat77521.021.021.00021.00021.00021.000	Amsinckia tessellata	373	29	10	8 (28)	26 (90)	8 (28)	0 (0)	
Trapteron palmeri26666730.1026.0001.016.03Sylechine palsearphoides225569550.08949.08949.0890.00Malacohrix coulteri2171.18641.7 0401.6 033.0000.00Lapinus odoratus2111.3630.003.1 0001.0 000.000Acnispon brachycarpus2.0001.114401.1 1001.1 0001.0 00Stephanomeria parryi1.1131.183.99.5031.8 (100)1.8 (100)Malacohrix galaratu1.1251.132.21.3 1002.6 (100)2.6 (100)Charaactis ferenonti1.192.111.82.0 0531.1 (20)4.0 (100)Charaactis ferenonti1.1072.6 (100)2.6 (100)2.6 (100)0.0 (100)Charagin spp.7.81.114.94.4 G38.6 (100)0.0 (100)Perioary spp.7.81.114.94.6 (10, 100)2.1 (100)2.1 (100)Trajendur arcine arcial1.751.003.4 (10, 100)2.1 (100)2.1 (100)Partago ontat7.61.61.81.0 (100)2.1 (100)1.1 (100)Charanter galar1.751.0 (100)1.0 (100)0.0 (100)1.1 (100)Partago ontat1.41.51.0 (100)1.0 (100)0.0 (100)Charanter galar1.41.41.0 (100)1.0 (100)1.0 (100)Partago ontat1.41.41.0 (100)	Gilia minor	342	52	72	51 (98)	46 (88)	51 (98)	1 (2)	
Syloline pikacarhoids23556959694949490Malacathric coulteri2171864171615100000Lupins odoratus21036300031000000000Acnispon brachycarpus200111*4401110001110011100000Stephanomeria parry111311183995051810018100000Malacathriz glabrata1125113221310021151418000Chaenacis fremonti1192111820195111534419000Coglanha circurcisa101726826100261002569000Lacedrary app.781114944688873101099600Patago ovat7510046101008100000000Trajdago neta7512880108100010011001100Inanthus dichatoma668187880100100110011000100Shimus barkut215100100110010001000000000000Charanthrigida121490110001000000000000Polosoma membranacum12149001001000000000Freigonum guillinm12149010001000000000Polosoma membranacu	<i>Erodium cicutarium</i> ²	281	40	7	4 (10)	10 (25)	8 (20)	36 (90)	
Malacadrix coulteri1217186417 (90)16 (80)15 (80)Lupius adoratus121136.30.003.1000.000.00Acrispon brachycarpus2001.111.110.110.001.11 (100)1.11 (100)0.000Stephanomeria parry11.131.189.99.9(50)1.81(50)1.81(50)0.000Malcodhrix glabrata1.121.182.151.13 (100)2.1(55)1.180.000Cheanactis fremonti1.112.211.31(0)2.2(50)1.4(5)0.000Cryptantha circumcissa0.1012.662.61(00)2.5 (100)2.5 (100)Cocarya sp.7.751.184.94.4(3)8.6(3)1.01(0)4.0(0)Parlago orata7.751.164.41.01(10)8.8(0)0.0(0)2.1(10)2.1(10)Linardius dichotomus6.657.80.002.1(10)2.1(10)1.1(10)1.1(10)1.1(10)Shismus barbatus ² 7.754.41.613.431.1(14)7.1(10)1.1(10)1.1(10)Shismus barbatus ² 7.557.70.000.000.000.0(0)0.0(0)0.0(0)Chorizanthe rigida1.121.415.1.1(10)1.1(10)1.1(10)0.0(0)Chorizanthe rigida1.121.250.0(0)0.0(0)0.0(0)0.0(0)0.0(0)Chorizanthe rigida1.121.411.430.0(0)0.1(10)0.0(0)<	Tetrapteron palmeri	266	26	73	0 (0)	26 (100)	1 (4)	6 (23)	
Lapius odoratus2113630.100.100.00Acnispon brachycarpus20011*4011(100)11(100)11(100)000Stephanomeria paryał131138399(50)18(100)18(100)000Malacohris glabrata1125132213(100)2(15)14(8)000Chaenaciis fremonti119211820(95)11(52)41(9)000Cryptantha circuncissa107265626(100)26(100)25(96)000Loesclustrum schotti7718900(0)8(100)0(0)9(82)Partago ovata751014610(100)8(80)10(10)9(82)Pantago ovata751014610(100)8(80)10(10)21(10)Choinauts dichotomus668187(88)0(0)0(0)11(10)Schimus barbata ² 75700(0)10(10)10(10)10(10)10(10)Pholisoma membranacum1411511(10)11(10)10(10)0(0)0(0)Friegorum grafilman101101250(0)0(0)0(0)0(0)0(0)Points madriteris ² 795321(10)10(10)10(10)0(0)0(0)Friegorum grafilman11911521(10)0(0)0(0)0(0)Points madriteris ² 795321(10)10(10)0(10)0(0) <td< td=""><td>Stylocline psilocarphoides</td><td>235</td><td>56</td><td>95</td><td>50 (89)</td><td>49 (88)</td><td>49 (88)</td><td>0 (0)</td></td<>	Stylocline psilocarphoides	235	56	95	50 (89)	49 (88)	49 (88)	0 (0)	
Actingion branchycarpus200111440111(100111(100111(100111(100010Stephanomeria parryi*113118399 (50)18 (100)18 (100)0 (0)Malacohrix glabrata1125132213 100)2 (15)1 (8)0 (0)Chaenactis fremanti119211820 (5)11 (52)4 (19)0 (0)Cryptantha circumcissa1070265626 (100)25 (60)25 (60)0 (0)Losciastrum schotti779108900 (0)8 (100)0 (0)9 (20)Perdocaryu spp.778110494 (63)8 (73)10 (9)9 (21)Pantago ovata7751004610 (100)8 (80)10 (10)2 (100)Tropidocaryu spracile7751004610 (100)8 (80)10 (10)2 (100)Schismus barbatus*765700 (0)0 (0)1 (10)1 (10)1 (10)Allum finbriatum725700 (0)1 (10)1 (10)0 (0)0 (0)Polistoma mebranaceum114551 (100)1 (100)1 (100)0 (0)0 (0)Eriognum gracilium112114751 (100)1 (100)1 (100)0 (0)0 (0)Chorizanthe rigida11211250 (0)1 (10)1 (10)0 (0)0 (0)Eriognum gracilium11211151 (100)1 (10)1 (10)0 (0)0 (0	Malacothrix coulteri	217	18	64	17 (94)	16 (89)	15 (83)	0 (0)	
Action Action Action Action Action Stephanomeria party1 113 118 39 9 (50) 18 (100) 18 (100) 0 (0) Malacothrix glabrata 1125 113 22 13 (100) 2 (15) 14 (8) 0 (0) Chanactis fremonti 119 211 18 20 (95) 11 (52) 4 (19) 0 (0) Cryptantha circumcisa 0107 266 26 (100) 26 (100) 25 (96) 0 (0) Loescliastrum schotti 79 8 90 0 (0) 8 (100) 0 (0) 9 (82) Planago ovata 775 101 46 10 (100) 8 (80) 10 (100) 2 (100) 2 (100) Linanthus dichotomus 066 8 18 7 (88) 0 (0) 0 (10) 1 (14) Allium finibriatum 205 70 0 (0) 0 (0) 1 (14) Allium finibriatum 125 10 (0) 1 (100) 1 (10) 0 (0) Foloonum membranaceum	Lupinus odoratus	211	3	63	0 (0)	3 (100)	0 (0)	0 (0)	
Malacothris glabrata125132213 (100)2 (15)1 (8)0 (0)Chanactis fremonti119211820 (95)11 (52)4 (19)0 (0)Cryptantha circumcissa107265626 (100)26 (100)25 (96)0 (0)Losseliastrum schotti7988900 (0)8 (100)0 (0)9 (82)Partago ovata775101444 (36)8 (73)10 (9)9 (82)Plantago ovata77520880 (0)2 (100)2 (100)2 (100)Linanthus dichotomus66618187 (88)0 (0)0 (0)1 (13)Schismus barbatus ² 505700 (0)0 (0)0 (0)0 (0)Pholistoma membranaceum1014151 (100)1 (100)0 (0)0 (0)Chorizanthe rigida101221901 (100)1 (100)0 (0)Eriogonum pusillum101214250 (0)0 (0)0 (0)0 (0)Frenanthelia exigua881X0 (0)0 (0)0 (0)0 (0)Chorizanthe brevicornu2021532 (100)0 (0)0 (0)0 (0)Chorizanthe brevicornu2021532 (100)0 (0)0 (0)0 (0)Chorizanthe rigida1214250 (0)0 (0)0 (0)0 (0)Eriogonum pusillum212332 (100)1 (100)0 (0)0 (0)0 (0)<	Acmispon brachycarpus	200	11*	40	11 (100)	11 (100)	11 (100)	0 (0)	
Chanactis fremonti11111111820 (95)11 (52)4 (19)0 (0)Cryptantha circumcissa10072665626 (100)25 (100)25 (96)0 (0)Loescliastrum schotti1709188900 (0)8 (100)8 (100)0 (0)Pactocarya spp.17081718494 (36)8 (73)10 (9)9 (82)Plantago ovata170517094610 (100)8 (80)10 (100)0 (0)Trojelocarpus gracile1705170880 (0)2 (100)2 (100)Linanthus dichotomus6668187 (8)0 (0)0 (0)1 (10)1 (10)Schimus barbatus ² 1705700 (0)0 (0)0 (0)1 (10)0 (0)Phoistoma membranaceum1014171151 (100)1 (100)0 (0)0 (0)Chorizanthe rigida1012141250 (0)1 (100)1 (100)0 (0)Eriogonum gracilimum10121411250 (0)1 (100)0 (0)0 (0)Freandentierisis ² 1991332 (100)1 (100)0 (0)0 (0)0 (0)Calvaseris parryi174151 (100)0 (0)0 (0)0 (0)Chorizanthe breicormu174151 (100)0 (0)0 (0)0 (0)Chorizanthe breicormu174151 (100)0 (0)0 (0)0 (0)Chorizanthe breicormu164151 (100)0 (0) <t< td=""><td>Stephanomeria parryi¹</td><td>131</td><td>18</td><td>39</td><td>9 (50)</td><td>18 (100)</td><td>18 (100)</td><td>0 (0)</td></t<>	Stephanomeria parryi ¹	131	18	39	9 (50)	18 (100)	18 (100)	0 (0)	
Cryptantha circumcissa107265626 (100)26 (100)25 (96)0 (0)Loeseliastrum schotti79890008 (100)600000000Pectocarya spp.78111494 (36)8 (73)10 (10)9 (82)Plantago ovata751004610 (100)8 (80)10 (100)0 (0)Tropidocarpus gracile752880002 (100)2 (100)2 (100)Linanthus dichotomus6668187 (88)0 (0)0 (0)1 (14)Alium finibriatum25700000 (0)5 (100)0 (0)Pholistoma membranaceum1121451 (100)1 (100)0 (0)Eriogonum gracillimun11211250 (0)1 (100)0 (0)Friogonum gracillimun1211901 (100)1 (100)0 (0)Bromus madritensis ³ 192532 (100)1 (100)1 (100)0 (0)Chorizanthe brevicornu172632 (100)1 (100)1 (100)0 (0)Chorizanthe brevicornu121151 (100)0 (0)0 (0)0 (0) <td>Malacothrix glabrata</td> <td>125</td> <td>13</td> <td>22</td> <td>13 (100)</td> <td>2 (15)</td> <td>1 (8)</td> <td>0 (0)</td>	Malacothrix glabrata	125	13	22	13 (100)	2 (15)	1 (8)	0 (0)	
Lock Model Model <th< td=""><td>Chaenactis fremonti</td><td>119</td><td>21</td><td>18</td><td>20 (95)</td><td>11 (52)</td><td>4 (19)</td><td>0 (0)</td></th<>	Chaenactis fremonti	119	21	18	20 (95)	11 (52)	4 (19)	0 (0)	
Pectocarya spp. 1 4 1	Cryptantha circumcissa	107	26	56	26 (100)	26 (100)	25 (96)	0 (0)	
Plantago ovata $\overline{1}$	Loeseliastrum schotti	79	8	90	0 (0)	8 (100)	0 (0)	0 (0)	
Trojidocarpus gracile 170 170 170 170 170 170 1100 1000 1000 1000 1000 1000 1000 1000 1000 1000 1000 1000 000 000 000 000 000 000 000 000 000 000 000 000 000 000 0000 <td>Pectocarya spp.</td> <td>78</td> <td>11</td> <td>49</td> <td>4 (36)</td> <td>8 (73)</td> <td>10 (91)</td> <td>9 (82)</td>	Pectocarya spp.	78	11	49	4 (36)	8 (73)	10 (91)	9 (82)	
Linanthus dichotomus 66 8 18 7 (8) 0 (0) 0 (0) 1 (13) Schismus barbatus ² 50 77 40 3 (43) 1 (14) 7 (100) 1 (14) Allium fimbriatum 25 55 70 0 (0) 0 (0) 5 (100) 0 (0) Pholistoma membranaceum 14 1 5 1 (100) 1 (100) 1 (100) 0 (0) Chorizanthe rigida 112 44 43 0 (0) 4 (100) 0 (0) 0 (0) Eriogonum gracillinum 112 14 25 0 (0) 0 (0) 1 (100) 0 (0) 0 (0) 0 (0) Bromus madritensis ² 9 2 53 2 (100) 0 (0)	Plantago ovata	75	10	46	10 (100)	8 (80)	10 (100)	0 (0)	
Schismus barbatus ² 100 100 100 100 100 100 Allium fimbriatum 25 5 70 000 000 5100 000 Pholistoma membranaceum 114 15 1100 1100 1100 000 Chorizanthe rigida 112 44 43 000 $4(100)$ 000 000 Eriogonum gracillinum 112 11 25 000 000 1100 000 Eriogonum pusillum 112 11 90 1100 1100 010 000 Bromus madritensis ² 99 253 2100 000 0100 000 Prenanthella exigua 183 $1X$ 000 0100 000 000 Chorizanthe brevicornu 22 63 2100 1000 1000 000 Lomatium mohavens ¹ 12 11 5 1100 000 000 0	Tropidocarpus gracile	75	2	88	0 (0)	2 (100)	2 (100)	2 (100)	
Allium fimbriatum 25 70 000 000 5100 000 Pholistoma membranaceum 14 1 5 1100 1100 1100 000 Chorizanthe rigida 12 44 43 000 $4(100)$ 000 000 Eriogonum gracillimum 12 1 25 000 000 1100 000 Eriogonum gracillimum 12 1 25 000 000 1100 000 Bromus madritensis ² 9 2 53 $2(100)$ 000 000 000 Chorizanthe brevicornu 17 2 63 $2(100)$ 000 1000 000 Chorizanthe brevicornu 12 1 X 000 1000 000 000 Lomatium mohavense ¹ 12 1 X $1(100)$ 000 000 000 Lomatium mohavense ¹ 10 1 X $1(100)$ 00	Linanthus dichotomus	66	8	18	7 (88)	0 (0)	0 (0)	1 (13)	
Pholistoma membranaceum 14 1 5 1 (100) 1 (100) 1 (100) 0 (0) Chorizanthe rigida 12 4 43 0 (0) 4 (100) 0 (0) 0 (0) Eriogonum gracillimum 12 1 25 0 (0) 0 (0) 1 (100) 1 (100) 0 (0) Eriogonum pusillum 12 1 90 1 (100) 1 (100) 1 (100) 0 (0) Bromus madritensis ² 99 2 53 2 (100) 0 (0) 0 (0) 0 (0) Chorizanthe brevicornu 77 2 63 2 (100) 0 (0) 1 (100) 0 (0) 0 (0) Lomatium mohavense ¹ 2 1 X 0 (0) 0 (0) 0 (0) 0 (0) 0 (0) Chaenactis carphoclinia 1 X 1 (100) 0 (0) 0 (0) 0 (0) 0 (0)	Schismus barbatus ²	50	7	40	3 (43)	1 (14)	7 (100)	1 (14)	
Chorizanthe rigida Image: Chorizanthe rigida <thimage: chorizanthe="" rigida<="" th=""> Image: Cho</thimage:>	Allium fimbriatum	25	5	70	0 (0)	0 (0)	5 (100)	0 (0)	
Eriogonum gracillimum 12 1 25 0 (0) 0 (0) 1 (100) 0 (0) Eriogonum pusillum 12 1 90 1 (100) 1 (100) 1 (100) 0 (0) Bromus madritensis ² 9 2 53 2 (100) 0 (0) 0 (0) 0 (0) Prenanthella exigua 8 1 X 0 (0) 0 (0) 1 (100) 0 (0) Calycoseris parryi 77 22 63 2 (100) 1 (50) 0 (0) 0 (0) Chorizanthe brevicornu 2 1 5 1 (100) 0 (0) 1 (100) 0 (0) Chaenactis carphoclinia 11 5 1 (100) 0 (0) 0 (0) 0 (0)	Pholistoma membranaceum	14	1	5	1 (100)	1 (100)	1 (100)	0 (0)	
Eriogonum pusillum 12 1 90 1 (100) 1 (100) 1 (100) 0 (0) Bromus madritensis ² 9 2 53 2 (100) 0 (0) 0 (0) 0 (0) Prenanthella exigua 8 1 X 0 (0) 0 (0) 1 (100) 0 (0) Calycoseris parryi 77 22 63 2 (100) 1 (50) 0 (0) 0 (0) Chorizanthe brevicornu 2 1 5 1 (100) 0 (0) 1 (100) 0 (0) Lomatium mohavense ¹ 2 1 X 1 (100) 0 (0) 0 (0) 0 (0)	Chorizanthe rigida	12	4	43	0 (0)	4 (100)	0 (0)	0 (0)	
Bromus madritensis ² 9 2 53 2 (100) 0 (0) 0 (0) 0 (0) Prenanthella exigua 8 1 X 0 (0) 0 (0) 1 (100) 0 (0) Calycoseris parryi 7 2 63 2 (100) 1 (50) 0 (0) 0 (0) Chorizanthe brevicornu 2 1 5 1 (100) 0 (0) 1 (100) 0 (0) Lomatium mohavense ¹ 2 1 X 1 (100) 0 (0) 0 (0) 0 (0)	Eriogonum gracillimum	12	1	25	0 (0)	0 (0)	1 (100)	0 (0)	
Prenanthella exigua 8 1 X 0 (0) 0 (0) 1 (100) 0 (0) Calycoseris parryi 7 2 63 2 (100) 1 (50) 0 (0) 0 (0) Chorizanthe brevicornu 2 1 5 1 (100) 0 (0) 1 (100) 0 (0) Lomatium mohavense ¹ 2 1 X 1 (100) 0 (0) 0 (0) 0 (0)	Eriogonum pusillum	12	1	90	1 (100)	1 (100)	1 (100)	0 (0)	
Calycoseris parryi 7 2 63 2 (100) 1 (50) 0 (0) 0 (0) Chorizanthe brevicornu 2 1 5 1 (100) 0 (0) 1 (100) 0 (0) Lomatium mohavense ¹ 2 1 X 1 (100) 0 (0) 0 (0) 0 (0) Chaenactis carphoclinia 1 1 5 1 (100) 0 (0) 0 (0) 0 (0)	Bromus madritensis ²	9	2	53	2 (100)	0 (0)	0 (0)	0 (0)	
Chorizanthe brevicornu 2 1 5 1 (100) 0 (0) 1 (100) 0 (0) Lomatium mohavense ¹ 2 1 X 1 (100) 0 (0) 0 (0) 0 (0) Chaenactis carphoclinia 1 5 1 (100) 0 (0) 0 (0) 0 (0)	Prenanthella exigua	8	1	Х	0 (0)	0 (0)	1 (100)	0 (0)	
Lomatium mohavense ¹ 2 1 X 1 (100) 0 (0) 0 (0) 0 (0) Chaenactis carphoclinia 1 5 1 (100) 0 (0) 0 (0) 0 (0)	Calycoseris parryi	7	2	63	2 (100)	1 (50)	0 (0)	0 (0)	
Chaenactis carphoclinia 1 1 5 1 (100) 0 (0) 0 (0) 0 (0)	Chorizanthe brevicornu	2	1	5	1 (100)	0 (0)	1 (100)	0 (0)	
	Lomatium mohavense ¹	2	1	Х	1 (100)	0 (0)	0 (0)	0 (0)	
Total count (average %) 11,933 723 (46) (60) (63) (59) (9)	Chaenactis carphoclinia	1	1	5	1 (100)	0 (0)	0 (0)	0 (0)	
	Total count (average %)	11,933	723	(46)	(60)	(63)	(59)	(9)	

See Methods for calculations of parts of plants eaten (% consumed). ¹ Herbaceous perennials. ² Non-native species. Data on bite count and plant numbers consumed are from Jennings and Berry (2015, Table 6) except for values with an asterisk, which are the correct numbers. Parts of plants eaten (% consumed), see Methods for calculations. X = data not estimated.

tortoises appeared to focus more on eating the vegetative parts of plants; on average they consumed leaves and stems 73% and 76% of the time, respectively (Table 3). The frequency of eating flowers (43%) was reduced, and consumption of fruits (32%) increased (Table 3).

In late spring, tortoise foraging tactics differed from the other time periods. On average, they consumed a smaller percentage of each forage plant (31%) compared to earlier periods. For the preferred forage species, Chamaesyce albomarginata, tortoises tended to eat ~30% of each plant (Table 4). They ate a larger fraction of the green and succulent E. cicutarium (20%) compared with early and middle spring periods (7% and 5%, respectively). The leaves, stems, and fruits of succulent E. cicutarium were consumed in a largely equal manner compared to early spring (Table 4). The tortoises also were less selective about parts to eat on succulent E. boothii; they ate flowers more frequently than leaves and stems (Table 4). Another change in foraging habits was the substantial inclusion of dried annual plants-annuals that had flowered earlier in the year but had flowered and dried a year earlier (Table 4). The tortoises appeared to eat many dried plants comparable to when the plants were succulent (Table 4).

5 Discussion

5.1 Frequency of superblooms in a warming climate

The proportions of high winter rains that produce superblooms varied in the last 60 years in the vicinity of the study site from approximately one every 10 years between 1961 and 1990, to a cluster of five in eight years between 1991 and 1998. Since 1999, the return frequency of superblooms averaged less than five years during the megadrought. The increase in extreme precipitation events was predicted to occur with increases in global temperatures as part of anthropogenic climate warming (Williams et al., 2020; Cook et al., 2022; Williams et al., 2022). The superblooms during the megadrought years and beyond may not carry the same high productivity of annual blooms or length of the spring blooming season, because of decreases in soil moisture and increasing heat waves (Williams et al., 2020; Cook et al., 2021; Cook et al., 2022; Dannenberg et al., 2022; Thompson et al., 2022; Williams et al., 2022). Annual and herbaceous perennial plants may dry up earlier, limiting the foraging time for tortoises.

TABLE 3 Characteristics of winter annual and herbaceous perennial species of plants and plant parts consumed by adult desert tortoises (*Gopherus agassizii*) during middle spring (the second phenological period: May) at the Desert Tortoise Research Natural Area, western Mojave Desert, California, USA, in 1992.

Plant species eaten	Number of		%	Parts of plants eaten (% consumed)			
	bites taken	plants eaten	of plant eaten	flowers	leaves	stems	fruits
Acmispon brachycarpus	10,312	999	58	157 (16)	989 (99)	989 (99)	989 (99)
Prenanthella exigua	1,969	133	81	121 (91)	8 (6)	112 (84)	63 (47)
Astragalus layneae ¹	1,191	38*	21	0 (0)	38 (100)	38 (100)	0 (0)
Chorizanthe brevicornu	541	32	31	31 (97)	31 (97)	31 (97)	3 (9)
Cryptantha circumcissa	420	30	43	30 (100)	30 (100)	30 (100)	0 (0)
Erodium cicutarium ²	391	63	5	1 (2)	4 (6)	3 (5)	63 (100)
Plantago ovata	352	56	66	53 (95)	1 2)	18 (32)	3 (5)
Eremothera boothii	287	27	32	3 (11)	27 (100)	18 (67)	0(0)
Chamaesyce albomarginata ¹	200	4	80	4 (100)	4 (100)	4 (100)	0 (0)
Amsinckia tessellata	194	16	15	12 (75)	16 (100)	1 (6)	1 (6)
Astragalus didymocarpus	114	5	83	5 (100)	5 (100)	5 (100)	0 (0)
Mirabilis laevis ¹	99	7	2	0 (0)	7 (100)	5 (71)	1 (14)
Mentzelia spp.	18	1	50	0 (0)	1 (100)	1 (100)	1 (100)
Pectocarya spp.	13	3	50	0 (0)	1 (33)	1 (33)	2 (67)
Caulanthus inflatus	8	1	5	0 (0)	0 (0)	1 (100)	0 (0)
Astragalus acutirostris	5	1	80	0 (0)	1 (100)	1 (100)	1 (100)
Eriastrum eremicum	5	2*	1	1 (50)	2 (100)	2 (100)	0 (0)
Total count (average %)	16,119	1,418	(41)	(43)	(73)	(76)	(32)

See Methods for calculations of parts of plants eaten (% consumed). ¹ Herbaceous perennials. ² Non-native species. Data on bite count and plant numbers consumed are from Jennings and Berry (2015, Table 8) except for values with an asterisk, which are the correct numbers. % plant part eaten = average % of above-ground consumed for each species.

TABLE 4 Characteristics of winter annual and herbaceous perennial species of plants and plant parts consumed by adult desert tortoises (*Gopherus agassizii*) during late spring (the third phenological period: June) at the Desert Tortoise Research Natural Area, western Mojave Desert, California, USA, in 1992.

Plant species eaten	Number of		%	Parts of plants eaten (% consumed)			
	bites taken	plants eaten	of plant eaten	flowers	leaves	stems	fruits
Chamaesyce albomarginata ¹	3,601	98*	29	98 (100)	98 (100)	98 (100)	1(1)
<i>Erodium cicutarium</i> ²	491*	66*	20	24 (36)	56 (85)	55 (83)	59 (89)
Eriastrum eremicum ³	376*	36	31	0 (0)	27 (75)	32 (89)	30 (83)
Eremothera boothii	366	35	31	29 (83)	24 (69)	25 (71)	18 (51)
Chorizanthe brevicornu	257	8	33	8 (100)	8 (100)	8 (100)	8 (100)
Erodium cicutarium ^{2,4}	218*	35*	28	0 (0)	35 (100)	35 (100)	0(0)
Schismus barbatus ^{2,3}	194	26	65	0 (0)	26 (100)	26 (100)	26 (100)
Eriastrum eremicum ⁴	116*	16	39	0 (0)	16 (100)	16 (100)	0 (0)
Amsinckia tessellata ⁴	106	8	35	0 (0)	0 (0)	8 (100)	0 (0)
Stylocline psilocarphoides ³	99	12	86	0 (0)	12 (100)	12 (100)	12 (100)
Chorizanthe brevicornu ⁴	88	11	33	0 (0)	11 (100)	11 (100)	11 (100)
Mirabilis laevis ¹	83	2	5	0 (0)	2 (100)	2 (100)	0 (0)
Cryptantha circumcissa ³	34	5	54	0 (0)	5 (100)	5 (100)	5 (100)
<i>Chorizanthe brevicornu</i> ³	31	4	51	0 (0)	4 (100)	4 (100)	3 (75)
Phacelia tanacetifolia ³	28	1	15	0 (0)	0 (0)	0 (0)	1 (100)
Eriastrum eremicum	26	6*	22	6 (100)	4 (67)	6 (100)	3 (50)
Oxytheca perfoliata ³	25	3	7	0 (0)	3 (100)	3 (100)	3 (100)
Amsinckia tessellata ³	10	4	5	0 (0)	0 (0)	1 (25)	4 (100)
Astragalus layneae ¹	7	1	15	0 (0)	1 (100)	1 (100)	0 (0)
Eremothera boothii ³	6	3	3	0 (0)	1 (33)	3 (100)	3 (100)
Linanthus parryae ⁴	4	1	50	0 (0)	1 (100)	1 (100)	0 (0)
Ambrosia salsola	4	1	Х	1 (100)	0 (0)	0 (0)	0 (0)
Total count (average %)	6,170	382	(31)	(24)	(74)	(85)	(57)

See Methods for calculations of parts of plants eaten (% consumed). ¹ Herbaceous perennial species. ² Non-native species. ³ Dried annual plant from spring. ⁴ Dried annual plant from previous year. Data on bite count and plant numbers consumed are from Jennings and Berry (2015, Table 10) except for values with an asterisk, which are the correct numbers. *Ambrosia salsola* is a perennial shrub.

Superblooms and years of average and above rainfall are times for tortoises of all sizes to consume succulent green plants, develop fat bodies, fill bladders with dilute fluid, and grow (Henen et al., 1998; Berry et al., 2023). This is especially important for hatchling, juvenile, and immature tortoises, e.g., Berry et al. (2023). Although our findings pertain to a single site during a superbloom, we note that the preferred forage plants observed during a superbloom at the DTRNA were also present and consumed by tortoises at a site in the Soda Mountains in the central Mojave Desert in other years where tortoises fed on Acmispon brachycarpus and Prenanthella exigua in a year of above average rainfall (Berry et al., 2006; KHB, personal observations). The same forage plant species also occurred during years of average and above average rainfall, but not at the high levels of abundance (data not shown). Depending on the timing of winter precipitation, not all species bloom in such years, even during superblooms. Notably, in a multi-year study of juvenile, immature, and young adult tortoises at a site south of the DTRNA, immature and young adult tortoises grew during a dry year (winter rain, 54.01 mm, water year 71.07 mm) (Table 1 in Berry et al., 2023).

Dry or drought years associated with decadal changes in precipitation between 1961 and 2023 at the study site and elsewhere deserve consideration too, because tortoises cannot live for long periods without free water and food (Henen et al., 1998). Increased mortality from dehydration and starvation occurred when one dry year followed another in wild populations, with small, young tortoises especially vulnerable (Peterson, 1996; Berry et al., 2002; Longshore et al., 2003). From 1961 to 1990, droughts occurred in 12 out of 30 years, whereas during 1991–1998, only one drought occurred in eight years (Table 1). From the start of the megadrought through winter rains in 2022–2023, dry years occurred in seven out of 22 years, an increase of 25% from the 1961 to 1990 period. Except during 1991–1998, two years of drought occasionally occurred sequentially.

Summer and winter rains are important to tortoises for drinking free water throughout the Mojave Desert (Henen et al., 1998). Prior to this study, from 1961–1990, summer rains averaged 18% of annual rainfall, whereas during the megadrought, summer rains had declined to 8% of annual rain (Table 1). The western Mojave Desert rarely receives the extreme rainfall events typical of monsoons experienced in the eastern and northeastern Mojave Desert (e.g., Bhattacharya et al., 2023), although greater portions of annual rainfall occur in summer in these regions of the Mojave Desert than in the western Mojave Desert (Rowlands, 1995).

5.2 Regional and local differences in annual and herbaceous perennial plant species

Species of available annual and herbaceous perennial native and non-native plants may differ depending on desert region, precipitation patterns, surficial geology, topography, elevation, and history of anthropogenic uses (e.g., Burge and Bradley, 1976; Avery and Neibergs, 1997; Drake et al., 2015). For example, Burge and Bradley (1976) described bite counts and frequency of use of plant species eaten by tortoises at a site where homes were under construction in the northeastern Mojave Desert of Nevada. Nineteen species of annuals, 13 species of shrubs, herbaceous perennials, and cacti occurred onsite; of these, tortoises ate parts of three species of herbaceous perennials, four species of cacti, and 10 species of forbs and grasses during spring and summer. Of the 100 bites observed, 73% were on Sphaeralcea ambigua, Plantago insularis, and the native annual grass, Festuca octoflora. Drake et al. (2015) also reported use of S. ambigua by tortoises in a burned area in Nevada. At a site in Ivanpah Valley, California, also in the northeastern Mojave Desert, observers reported dietary overlap between tortoises and cattle (Avery and Neibergs, 1997). The site had been continually grazed between the 1870s and mid-1990s. Seven major forage species and 12 minor species were noted as part of the tortoise diet, but no bite counts were reported (Avery and Neibergs, 1997). Of the 19 species, two were common non-natives. In the studies by Burge and Bradley (1976) and Avery and Neibergs (1997), some species eaten by the tortoises were on the plant lists described in Tables 2-4, some were new, and some were summer annuals.

5.3 Composition and productivity of plants during superblooms and other years

During the superbloom of 1992, annual and herbaceous perennial plants began to grow in the weeks following the arrival of substantial late winter precipitation. Overall species richness and biomass of these ephemeral plants peaked in April before the plants gradually dried out and became senescent during middle to late spring periods (Jennings, 2001; SI-1). Native annuals formed most of the annual biomass, especially throughout the early spring period (SI-1). Non-native grasses in the genera *Bromus* spp. and *Schismus* spp. and forbs can be successful competitors of native annual plants in the Mojave Desert (Brooks, 2000). The effects of competition can vary by year and level of precipitation. Because the non-native grasses and forb (*Erodium cicutarium*) are so successful, they have negative effects on biomass of native forbs eaten by tortoises (Jennings and Berry, 2015). Results differ by year (Jennings and Berry, 2015).

Estimates of native and non-native annual biomass varied between wet years, between wet and dry years, and between protected and unprotected sites, following the variations in patterns of precipitation and levels of disturbance (SI-1). During dry years (1989, 1994, 1999, 2012), non-native annual species dominated the annual plant biomass (91–100%) at unprotected sites in the western, southern, and central Mojave Desert but were lower in protected habitats (SI-1). In contrast, non-native annual plant biomass tended to be lower during wet years (1993, 1995, 1997); estimates were 56–69% on protected sites and 30–86% on unprotected sites (SI-1).

Nonetheless, because production of richness and biomass of winter-annual species depends on the amounts of fall-winter precipitation, the predicted swings between extreme dry and wet years as the climate warms are likely to present tortoises with varying nutritional challenges (Rowlands et al., 1982; Rowlands, 1995; Brooks and Berry, 2006; Williams et al., 2020; Cook et al., 2021; Cook et al., 2022). During years with high, above average precipitation, a high diversity of native annual species is likely to bloom and thus be available as potential forage for tortoises (SI-1). In contrast, in extremely dry years—such as occurred in 1989 and 2012—most annuals did not germinate, leaving tortoises with little or no available food—or only with poor quality non-native annual grasses (Hazard et al., 2009; Hazard et al., 2010; SI-1).

Climate warming may alter the flowering phenologies of preferred tortoise forage species comparable to observations for other plant species (e.g., Anderson et al., 2012). Changes in availability of food plants may have severe consequences for tortoises in the Mojave Desert and elsewhere because tortoises track the flowering phenologies of preferred species during a superbloom and probably in other years with differing precipitation levels (Jennings, 2002; Jennings and Berry, 2015).

Another determinant of annual plant production is the level of anthropogenic disturbance to the land (e.g., livestock grazing and off-highway vehicle use). Inside protected areas such as the DTRNA, the biomass of native annuals tended to be higher than non-protected sites in both dry and wet years (Brooks, 2000; Brooks and Berry, 2006; Berry et al., 2020). Thus, production of native annual plants is likely to be limited on disturbed lands during drought years (e.g., Brooks and Berry, 2006).

5.4 Other factors potentially affecting forage preferences

The presence of contaminants and toxicants in some food plants may affect nutritional qualities and taste. Chaffee and Berry (2006) analyzed samples of soils and ephemeral stream bed sediments, and plants known or thought to be eaten by tortoises in six desert regions for potential contaminants or toxicity. Elevated levels of some elements were identified locally in some plant species, and sources were associated with contamination from mines, mine wastes, roads, railroads, and other disturbances (Chaffee and Berry, 2006). For example, elevated concentrations of arsenic, potentially toxic to tortoises, were identified in regions (and plants) associated with two mining districts (Chaffee and Berry, 2006). Elevated levels of arsenic were evident in an ill, necropsied tortoise from a mining district (Homer et al., 1998; Homer et al., 2001; Selzer and Berry, 2005). Dust from human disturbances settling on plants may contribute to uptake of such toxicants, or by contact with or breathing contaminated soils (Chaffee and Berry, 2006).

5.5 Forage routes and preferred microhabitats

The foraging routes taken by adult tortoises offer valuable information for ongoing and future recovery actions, e.g., projects for restoration of degraded and burned habitats and microhabitats that could benefit from protection from human disturbance. Tortoises more frequently used habitat strata in hills and ephemeral stream channels than on alluvial fans; microhabitats selected for foraging were the intershrub spaces and ephemeral stream channels. These microhabitats were open spaces in the environment and where most bites were taken. Importantly, the ephemeral stream channels were small, not the often wide, axial valley stream channels or washes with large shrubs and trees, but the third, fourth, or higher-level ephemeral stream channels upslope and draining into secondary channels to the axial valley ephemeral stream channel.

Vegetation in general and microhabitats favored by tortoises are vulnerable to disturbance from anthropogenic uses: livestock grazing, vehicle use, whether by recreationists or other users, and other sources. Livestock grazing occurred in desert tortoise habitats from the 1860s and continues in some critical habitat units (e.g., U.S. Bureau of Land Management, 1980; Muhn and Stuart, 1988; U.S. Fish and Wildlife Service [USFWS], 1994; U.S. Bureau of Land Management, 2019). Livestock grazing contributed to alteration of composition of perennial and annual plants and invasion of nonnative species in some designated critical habitat units for desert tortoises (e.g., Webb and Stielstra, 1979; Fleischner, 1994; Brooks and Berry, 2006; Brooks et al., 2006; Berry and Murphy, 2019). Livestock walk in ephemeral stream channels and intershrub spaces to feed, feed on annual plants and within shrubs, and seek shade within shrubs or under trees. Recreational vehicle use increased in intensity in the 1970s, with continuing expansion (e.g., U.S. Bureau of Land Management, 2019). This use remains a major factor in deterioration and fragmentation of tortoise habitats within the geographic range (e.g., U.S. Bureau of Land Management, 1973; U.S. Bureau of Land Management, 1980; Webb and Wilshire, 1983; U.S. Fish and Wildlife Service [USFWS], 1990; Brooks and Lair, 2009; U.S. Bureau of Land Management, 2019). Both authorized and unauthorized travel continues to occur in ephemeral stream channels and cross-country in intershrub spaces, and results in damaging shrubs (Berry et al., 2014; Berry et al., 2020).

6 Conclusions

In the American West where tortoises occur, climate warming and droughts are expected to bring greater aridity, prolonged and severe dry spells (Overpeck and Udall, 2020), more extreme heat waves (Dannenberg et al., 2022), and reductions in soil moisture (Cook et al., 2021; Cook et al., 2022). The frequency of superblooms and production of annual herbaceous perennial plants described here are likely to change as the century progresses. Superblooms were previously important to desert tortoises because they provided an abundance of succulent, green annual forbs, and herbaceous perennial plants, some of which are low in potassium and edible to tortoises. Years with superblooms were important in providing tortoises with sufficient resources to recover from droughts and other years of low rainfall, to develop fat stores, and to grow. New data on parts of plants utilized by tortoises provide a greater understanding of how tortoises forage. Similarly, the movements and travels focused on foraging added clarity to the value of different parts of the habitat and specific microhabitats essential for feeding. Maintenance and protection of these microhabitats from anthropogenic disturbances can contribute to recovery efforts for the species.

Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material. Further inquiries can be directed to the corresponding authors.

Ethics statement

Ethical approval was not required for the study involving animals in accordance with the local legislation and institutional requirements because the study was observational, and thus did not require handling of desert tortoises. This study did receive reviews prior to its commencement from J. Campbell at the University of Texas, Arlington and the U.S. Bureau of Land Management.

Author contributions

WBJ: Conceptualization, Data curation, Formal Analysis, Investigation, Methodology, Project administration, Resources, Validation, Visualization, Writing – original draft, Writing – review & editing. KHB: Conceptualization, Funding acquisition, Methodology, Project administration, Resources, Supervision, Visualization, Writing – original draft, Writing – review & editing.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

References

Abella, S. R., Berry, K. H., and Ferrazzano, S. (2023). Techniques for restoring damaged Mojave and western Sonoran habitats, including those for threatened desert tortoises and Joshua trees. *Desert Plants* 38, 4–52.

Allison, L. J., and McLuckie, A. M. (2018). Population trends in desert tortoises (Gopherus agassizii). Herpetol. Conserv. Biol. 13, 433-452.

Anderson, J. T., Inouye, D. W., McKinney, A. M., Colautti, R. I., and Mitchell-Olds, T. (2012). Phenotypic plasticity and adaptive evolution contribute to advancing flowering phenology in response to climate change. *Proc. Royal Soc. B* 279.1743, 3843–3852. doi: 10.1098/rspb.2012.1051

Ault, T. R., Mankin, J. S., Cook, B. I., and Smerdon, J. E. (2016). Relative impacts of mitigation, temperature, and precipitation on 21st century megadrought risk in the American Southwest. *Sci. Adv.* 2, 1–8. doi: 10.1126/sciadv.1600873

Avery, H. W., and Neibergs, A. (1997). "Effects of cattle grazing on the desert tortoise." J. Van Abbema, In *Proceedings: Conservation, Restoration, and Management of Tortoises and Turtles—An International Conference*, (New York: New York Turtle and Tortoise Society and WCS Turtle Recovery Program), 13–19.

Barboza, P. S. (1995). Nutrient balances and maintenance requirements for nitrogen and energy in desert tortoises (*Xerobates agassizii*) consuming forages. *Comp. Biochem. Physiol.* 112A, 537–545. doi: 10.1016/0300-9629(95)02023-3

Berry, K. H., Allison, L. J., McLuckie, A. M., Vaughn, M., and Murphy, R. W. (2021) *Gopherus agassizii, Mojave Desert tortoise* (The International Union for Conservation of Nature Red List of Threatened Species[™] (IUCN 2021). Available at: https://www. iucnredlist.org/species/97246272/3150871 (Accessed 14, 2023).

Berry, K. H., Bailey, T. Y., and Anderson, K. M. (2006). Attributes of desert tortoise populations at the National Training Center, Central Mojave Desert, California, USA. J. Arid Environ. 67 Suppl, 165–191. doi: 10.1016/j.jaridenv.2006.09.026

Berry, K. H., Lyren, L. M., Yee, J. L., and Bailey, T. Y. (2014). Protection benefits desert tortoise (*Gopherus agassizii*) abundance: the influence of three management strategies on a threatened species. *Herpetological Monogr.* 28, 66–92. doi: 10.1655/Herpmonographs-D-14-00002

Berry, K. H., Mack, J. S., and Anderson, K. M. (2023). Variations in climate drive behavior and survival of small desert tortoises. *Front. Ecol. Evol.* doi: 10.3389/fevo.2023.1164050

Berry, K. H., and Murphy, R. W. (2019). *Gopherus agassizii* (Cooper 1861) – mojave desert tortoise, agassiz's desert tortoise. *Chelonian Res. Monogr.* 5, 1–45. doi: 10.3854/crm.5.109.agassizii.v1.2019

Berry, K. H., Spangenberg, E. K., Homer, B. L., and Jacobson, E. R. (2002). Deaths of desert tortoises following periods of drought and research manipulations. *Chelonian Conserv. Bio.* 4, 436–448.

Berry, K. H., Yee, J. L., Shields, T. A., and Stockton, L. (2020). The catastrophic decline of tortoises at a fenced Natural Area. *Wildl. Monogr.* 205, 1–53. doi: 10.1002/wmon.1052

Bhattacharya, T., Feng, R., Maupin, C. R., Coats, S., Brennan, P. R., and Carter, E. (2023). California margin temperatures modulate extreme summer precipitation in the desert Southwest. *Environ. Res. Lett.* 18, 104048. doi: 10.1088/1748-9326/acfd43

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Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/famrs.2023.1283255/full#supplementary-material

Brooks, M. L. (2000). Competition between alien annual grasses and native annual plants in the Mojave Desert. *Am. Midl. Nat.* 144, 92–108. doi: 10.1674/0003-0031(2000) 144[0092:CBAAGA]2.0.CO;2

Brooks, M. L., and Berry, K. H. (2006). Dominance and environmental correlates of alien annual plants in the Mojave Desert, USA. *J. Arid Environ.* 67 (Supplement), 100–124. doi: 10.1016/j.aridenv.2006.09.021

Brooks, M. L., and Lair, B. M. (2009). "Ecological effects of vehicular routes in a desert ecosystem," in *The Mojave Desert*. Eds. R. H. Webb, L. F. Festermaker, J. S. Heaton, D. L. Hughson, E. V. McDonald and D. M. Miller (Reno, NV: University of Nevada Press), 168–195.

Brooks, M. L., and Matchett, J. R. (2006). Spatial and temporal patterns of wildfires in the Mojave Desert 1980–2004. *J. Arid Environ*. Supplement), 67, 148–164. doi: 10.1016/j.jaridenv.2006.09.027

Brooks, M. L., Matchett, J. R., and Berry, K. H. (2006). Effects of livestock watering sites on alien and native plants in the Mojave Desert, USA. *J. Arid Environ.* Supplement) 67, 125–147. doi: 10.1016/j.aridenv.2006.09.027

Burge, B. L., and Bradley, W. G. (1976). "Population density, structure and feeding habits of the desert tortoise, *Gopherus agassizi*, in a low desert study area in southern Nevada,". In: *Proc. 1st Ann. Desert Tortoise Council Symp* (Las Vegas, Nevada, USA: The Desert Tortoise Council). Available at: https://deserttortoise.org (Accessed 2, 2023).

Chaffee, M. A., and Berry, K. H. (2006). Abundance and distribution of selected elements in soils, stream sediments, and selected forage plants from desert tortoise habitat in the Mojave and Colorado deserts, USA. *J. Arid Environ*. Supplement) 67, 35–87. doi: 10.1016/j.jaridenv.2006.09.018

Christopher, M. M., Berry, K. H., Wallis, I. R., Nagy, K. A., Henen, B. T., and Peterson, C. C. (1999). Reference intervals and physiologic alterations in hematologic and biochemical values of free ranging desert tortoises in the Mojave Desert. *J. Wildl. Dis.* 35, 212–238. doi: 10.7589/0090-3558-35.2.212

Cook, B. I., Mankin, J. S., Williams, A. P., Marvel, K. D., Smerdon, J. E., and Liu, H. (2021). Uncertainties, limits, and benefits of climate change mitigation for soil moisture drought in southwestern North America. *Earth's Future* 9, e2021ER002013. doi: 10.1029/2021EF002014

Cook, B. I., Smerdon, J. E., Cook, E. R., Williams, A. P., Anchukaitis, K. J., Mankin, J. S., et al. (2022). Megadroughts in the common era and the anthropocene. *Nat. Rev. Earth Environ.* 3, 741–757. doi: 10.1038/s43017-022-00329-1

Daly, C., Halbleib, M., Smith, J. I., Gibson, W. P., Doggett, M. K., Taylor, G. H., et al. (2008). Physiographically sensitive mapping of climatological temperature and precipitation across the conterminous United States. *Int. J. Climatol.* 28, 2031–2064. doi: 10.1002/joc.1688

Dannenberg, M. P., Yan, D., Barnes, M. L., Smith, W. K., Johnston, M. R., Scott, R. L., et al. (2022). Exceptional heat and atmospheric dryness amplified losses of primary production during the 2020 U.S. Southwest hot drought. *Global Change Biol.* 22, 4794–4806. doi: 10.111/gcb.16214

Drake, K. K., Bowen, L., Nussear, K. E., Esque, T. C., Berger, A. J., Custer, N. A., et al. (2016). Negative impacts of invasive plants on conservation of sensitive desert wildlife. *Ecosphere* 7, e01531. doi: 10.1002/ecs2.1531

Drake, K. K., Esque, T. C., Nussear, K. E., DeFalco, L. A., Scoles-Sciulla, S. J., Modlin, A. T., et al. (2015). Desert tortoise use of burned habitat in the eastern Mojave Desert. *J. Wildlife Manage*. 79, 618–629. doi: 10.1002/ecs2.1531

Duda, J. J., Krzysik, A. J., and Freilich, J. E. (1999). Effects of drought on desert tortoise movement and activity. J. Wildl. Manage. 63, 1181–1192. doi: 10.2307/3802836

Fleischner, T. L. (1994). Ecological costs of livestock grazing in western North America. Conserv. Biol. 8, 629–644. doi: 10.1046/j.1523-1739.1994.08030629.x

Hardy, R. (1976). The Utah population—a look in the 1970's. In: *Proc. 1st Ann. Desert Tortoise Council Symp* (Nevada, USA: The Desert Tortoise Council). Available at: https://www.deserttortoise.org (Accessed 10, 2023).

Hazard, L. C., Shemanski, D. R., and Nagy, K. A. (2009). Nutritional quality of natural foods of juvenile desert tortoises (*Gopherus agassizii*): energy, nitrogen, and fiber digestibility. J. Herpetol. 43, 38–48. doi: 10.1670/07-160R1.1

Hazard, L. C., Shemanski, D. R., and Nagy, K. A. (2010). Nutritional quality of natural foods of juvenile and adult desert tortoises (*Gopherus agassizii*): calcium, phosphorus, and magnesium digestibility. *J. Herpetol.* 44, 135–147. doi: 10.1670/08-123.1

Henen, B. T. (1997). Seasonal and annual energy budgets of female desert tortoises (*Gopherus agassizii*). Ecology 78, 283–296. doi: 10.1890/0012-9658(1997)078[0283: SAAEBO]2.0.CO;2

Henen, B. T., Peterson, C. C., Wallis, I. R., Berry, K. H., and Nagy, K. A. (1998). Effects of climatic variation on field metabolism and water relations of desert tortoises. *Oecologia* 117, 365–373. doi: 10.1007/s004420050669

Henen, B. T. (2002). Energy and water balance, diet, and reproduction of female desert tortoises (*Gopherus agassizii*). Chelonian Conserv. Bio. 4, 319–328.

Homer, B. L., Berry, K. H., Brown, M. B., Ellis, G., and Jacobson, E. R. (1998). Pathology of diseases in wild desert tortoises from California. *J. Wildlife Dis.* 35, 508–523. doi: 10.7589/0090-3558-34.3.508

Homer, B. L., Li, C., Berry, K. H., Denslow, N. D., Jacobson, E. R., Sawyer, R. H., et al. (2001). Soluble scute proteins of healthy and ill desert tortoises (*Gopherus agassizii*). *Am. J. Veterinary Res.* 62, 104–110. doi: 10.2460/ajvr.2001.62.104

Jennings, W. B. (2001). Comparative flowering phenology of plants in the western Mojave Desert. *Madroño* 48, 162–171.

Jennings, W. B. (2002). Diet selection by the desert tortoise in relation to the flowering phenology of ephemeral plants. *Chelonian Conserv. Biol.* 4, 353–335.

Jennings, W. B., and Berry, K. H. (2015). Desert tortoises (*Gopherus agassizii*) are selective herbivores that track the flowering phenology of their preferred food plants. *PloS One* 10, e0116716. doi: 10.371/journal.pone.0116716

Jennings, W. B., and Fontenot, C. L. (1993). Observations on the feeding behavior of desert tortoises (*Gopherus agassizii*) at the Desert Tortoise Research Natural Area, Kern County, California. In: *Proc. Desert Tortoise Counc. Symp* (Nevada, USA: The Desert Tortoise Council). Available at: https://www.deserttortoise.org (Accessed 20, 2023).

Jepson Flora Project (2023) Jepson eFlora. Available at: https://ucjeps.berkeley.edu/ eflora/ (Accessed 6, 2023).

Klinger, R., Underwood, E. C., McKinley, R., and Brooks, M. L. (2021). Contrasting geographic patterns of ignition probability and burn severity in the Mojave Desert. *Front. Ecol. Evol.* 9. doi: 10.3389/fevo.2021.593167

Longshore, K. M., Jaeger, J. R., and Sappington, J. M. (2003). Desert tortoise (*Gopherus agassizii*) survival at two eastern Mojave Desert sites: death by short-term drought? *J. Herpetology* 37, 169–177. doi: 10.1670/0022-1511(2003)037[0169:DTGASA]2.0.CO;2

Medica, P. A., Nussear, K. E., Esque, T. C., and Saethre, M. B. (2012). Long-term growth of desert tortoises (*Gopherus agassizii*) in a southern Nevada population. J. Herpetol. 46, 213–220. doi: 10.1670/11-327

Muhn, J., and Stuart, H. R. (1988). *Opportunity and Challenge, The Story of BLM* (Washington, D.C: U.S. Dept. of the Interior, Bureau of Land Management, U.S. Government Printing Office).

Oftedal, O. T. (2002). "Nutritional ecology of the desert tortoise in the Mohave and Sonoran deserts," in *The Sonoran Desert Tortoise: Natural History, Biology, and* *Conservation.* Ed. T. R. Van Devender (Tucson, AZ: University of Arizona Press and the Arizona-Sonora Desert Museum), 194–241.

Oftedal, O. T., Allen, M. E., Chung, A. L., Reed, R. C., and Ullrey, D. E. (1994). Nutrition, urates, and desert survival: Potassium and the desert tortoise (*Gopherus agassizii*). Proc. Am. Assoc. Zoo Veterinarians Ann. Meeting 1994 308–313.

Oftedal, O. T., Hillard, S., and Morafka, D. J. (2002). Selective spring foraging by juvenile desert tortoises (*Gopherus agassizii*) in the Mojave Desert: evidence of an adaptive nutritional strategy. *Chelonian Conserv. Biol.* 4, 341–352.

Overpeck, J. T., and Udall, B. (2020). Climate change and the aridification of North America. Proc. Natl. Acad. Sci. U.S.A. 117, 11856–11858. doi: 10.1073/pnas.2006323117

Peterson, C. C. (1996). Anhomeostasis: seasonal water and solute relations in two populations of the desert tortoise (*Gopherus agassizii*) during chronic drought. *Physiol. Zool* 69, 1324–1358. doi: 10.1086/physzool.69.6.30164263

R Development Core Team (2023) R: A Language and Environment for Statistical Computing. Available at: https://cran.r-project.org/ (Accessed 15, 2023).

Rowlands, P. (1995). "Regional bioclimatology of the California Desert," in *The California Desert: An Introduction to Natural Resources and Man's Impact.* Eds. J. Latting and P. G. Rowlands (Riverside, CA: June Latting Books), 95–134.

Rowlands, P., Johnson, H., Ritter, E., and Endo, A. (1982). "The Mojave Desert," in *Reference Handbook on the Deserts of North America*. Ed. G. Bender (Westport, CT: Greenwood Press), 103–162.

Selzer, M. D., and Berry, K. H. (2005). Laser ablation ICP-MS profiling and semiquantitative determination of trace element concentrations in desert tortoise shells: documenting the uptake of elemental toxicants. *Sci. Total Environ.* 339, 253–265. doi: 10.1016/j.scitotenv.2004.07.027

Thompson, V., Kennedy-Asser, A. T., Vosper, E., Lo, Y. T. E., Huntingford, C., Andrews, O., et al. (2022). The 2021 western North America heat wave among the most extreme events ever recorded globally. *Sci. Adv.* 8, eabm6860. doi: 10.1126/sciadv.abm6860

Turner, F. B., Berry, K. H., Randall, D. C., and White, G. C. (1987). *Population ecology of the desert tortoise at Goffs, California 1983–1986* (Rosemead, California, USA: Report to Southern California Edison).

U.S. Bureau of Land Management (1973). Interim Critical management Plan for Recreational Vehicle Use in the California Desert (U.S. Dept. of the Interior, Bureau of Land Management, California Desert Program, USA).

U.S. Bureau of Land Management (1980). *The California Desert Conservation Area Pla* (California, USA: U.S. Department of the Interior, Bureau of Land Management, Desert District).

U.S. Bureau of Land Management (2019). West Mojave route network project. Final supplemental environmental impact statement for the California Desert District. BLM/ CA/DOI-BLM-CA-2018-008-EIS (Moreno Valley, California, USA: USBLM).

U.S. Fish and Wildlife Service [USFWS] (1990). Endangered and threatened wildlife and plants; determination of threatened status for the Mojave population of the desert tortoise. *Fed. Regist.* 55, 12178–12191.

U.S. Fish and Wildlife Service [USFWS] (1994). *Desert tortoise (Mojave population) recovery plan* (Portland, Oregon. USA: USFWS).

U.S. Fish and Wildlife Service [USFWS] (2011). Revised Recovery Plan for the Mojave Population of the desert tortoise (Gopherus agassizii) (Reno, Nevada, USA: USFWS).

U.S. Fish and Wildlife Service [USFWS] (2015). Range-wide Monitoring of the Mojave Desert Tortoise (Gopherus agassizii): 2013–2014. Annual Report (Reno, NV: U.S. Department of the Interior, Fish and Wildlife Service).

Webb, R. H., and Stielstra, S. S. (1979). Sheep grazing effects on Mojave Desert vegetation and soils. *Environ. Manage.* 3, 517–529. doi: 10.1007/BF01866321

Webb, R. H., and Wilshire, H. G. (1983). Environmental Effects of Off-Road Vehicles (New York, New York: Springer Verlag).

Williams, A. P., Cook, B. I., and Smerdon, J. E. (2022). Rapid intensification of the merging southwestern North American megadrought in 2021. *Nat. Clim. Change* 12, 232–234. doi: 10.1038/s41558-022-01290-z

Williams, A. P., Cook, E. R., Smerdon, J. E., Cook, B. L., Abatzoglou, J. T., Bolles, K., et al. (2020). Large contribution from anthropogenic warming to an emerging North American megadrought. *Science* 368, 314–318. doi: 10.1126/science.aaz9600

Woodbury, A. M., and Hardy, R. (1948). Studies of the desert tortoise, *Gopherus agassizii. Ecol. Monogr.* 18, 145–200. doi: 10.2307/1948638