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Advances in innovative seed potato production systems in India

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India is the second largest producer of potatoes in the world. Seed is the single most important input in potato cultivation. High seed rate (2.5–3.0 tons/ha), low rate of multiplication, progressive viral degeneration, storage, and transportation are major issues of potato seed production in the country. Potato seed alone accounts for 40%–50% of the total potato production cost, and huge quantities of potentially edible food is put back into the soil as potato seed. The delayed penetration of new improved potato/seed varieties into farmers' fields due to the slow multiplication rate and frequent seed replacement because of degeneration are associated issues. To circumvent these issues, continuous efforts are being made by potato researchers to develop innovative technologies for quick multiplication of initial healthy breeder's seed of the released varieties in sufficient quantities to meet the demand in our country. A paradigm shift in potato seed production methods has taken place globally since the early 1900s. Major potato producers of the world have shifted from conventional to hi-tech seed production systems to improve the seed quality and enhance seed multiplication rate. New innovations can overcome many of the problems associated with potato seed production, particularly in tropical and sub-tropical countries. Recent advances in potato seed production systems in India and challenges ahead for seed production are described here.

KEYWORDS

solanum tuberosum, micropropagation, aeroponics, ARC, TPS, potato diploids

Introduction

Potato (*Solanum tuberosum* L) is emerging as one of the important food crops in India accounting for 11.3% of the total global potato area and contributing 12.5% to the global potato production. In 2019–2020, India harvested more than 51.3 million tons of potatoes from 2.16 million hectares of cropped area with an average production of 23.77 tons/ha (Food and Agriculture Organization (FAO), 2022). Supported by technological innovations and the development of improved potato varieties suitable for growing under sub-tropical conditions, and driven by the growing demands, potato productivity and production in India have increased 3.28- and 18.87-fold, respectively during the period of 1961–2020. Nonetheless, potato production in India is still constrained by scarcity of varieties having diverse economic attributes that give better options to the potato growers; inadequacy of quality seed, and shortage of storage infrastructure. Most important among these is the inadequacy of quality planting material, as it has a direct bearing on crop productivity (Singh and Sharma, 2018). Being bulky and having high seed rate and slow rate of multiplication (five to six times) pose a unique challenge to seed potato production and its supply chain (Young, 1990). Since the crop is multiplied vegetatively using the tuber as seed, it gets degenerated very fast, necessitating the replacement of the seed every year (ideally, or after every 2 years) (Singh et al., 2018). Furthermore, the seed is either short in supply or out of reach of farmers owing to high price. The seed-related issues are further aggravated because of the restricted availability of aphid-free, seed-producing regions in the country. It has been demonstrated that quality seeds alone can increase productivity by 15%–20% (Shaheb et al., 2016).

There is a huge gap between the requirement and supply of certified seed potatoes in India [National Academy of Agricultural Sciences (NAAS), 2021]. The Central Potato Research Institute (CPRI) of the Indian Council of Agricultural Research (ICAR) produces about 2,400 tons of breeder's seed (basic seed) every year and supplies 80% of it to the states and other agencies for its multiplication ((ICAR-CPRI Annual Report, 2020). If this stock has to multiply in three stages, i.e., foundation 1, foundation 2, and certified grades (as per norms), we can produce only about 0.5 million tons of certified seed. On the assumption of a 100% seed replacement rate (SRR), it meets only 10% of the total seed requirement, leaving a deficit of about 4.9 million tons (Buckseth et al., 2020). It is virtually impossible to produce such a huge quantity of certified seed using traditional methods. The traditional system of seed potato production in India consists of producing breeders' seed (basic seed) after four field multiplications of nucleus seed on research farms of ICAR-CPRI followed by three more field multiplications as foundation 1, foundation 2, and

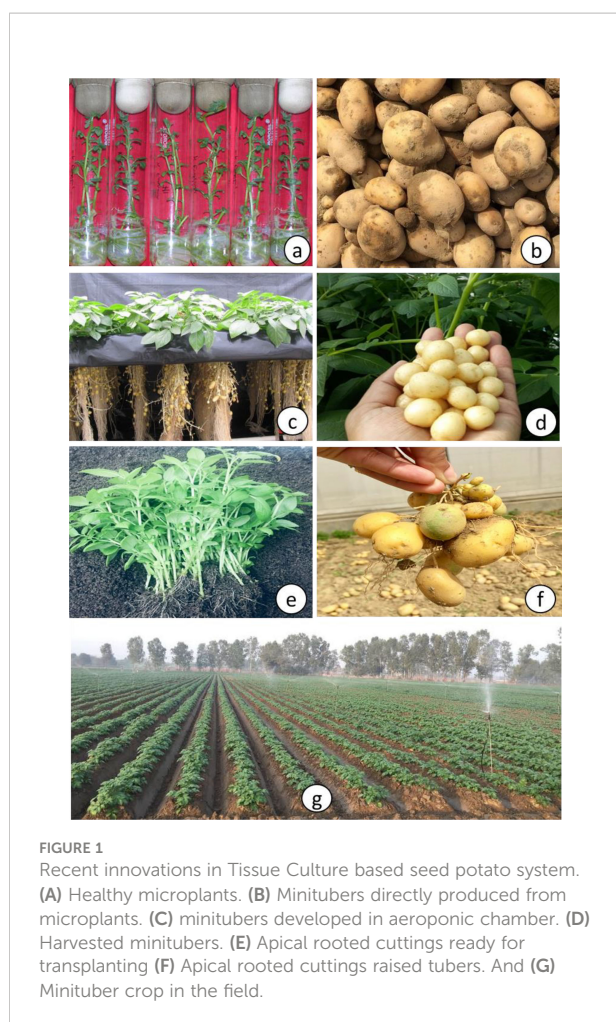
certified seed by respective states of the country (Venkataslam et al., 2017; Singh et al., 2019a). In addition to the low rate of field multiplications, the traditional system suffers from several other constraints like the requirement of a huge number of disease-free propagules in the initial stage and progressive accumulation of degenerative viral diseases with each field exposure (Venkataslam et al., 2017; Naik and Buckseth, 2018). The situation is further aggravated by the fact that the breeder's seed supplied by ICAR-CPRI is seldom multiplied in three recommended generations by the states. About 1.2 million tons of correctly labeled potato seed are sold by private seed producers, especially those from Punjab, western Uttar Pradesh, West Bengal, and Haryana, without any mechanisms and infrastructure for monitoring the seed quality [National Academy of Agricultural Sciences (NAAS), 2021]. It is, therefore, imperative to evolve a seed production system, encompassing innovative techniques to improve the quality of seed and reduce field exposures, along with a robust system of certification and quality assurance of the seed produced and supplied by the private seed growers. This mini-review highlights the recent innovations in seed potato production in India.

Current innovations in seed potato production

Tissue culture-based seed potato production

The use of micropropagation for commercial seed production has moved potatoes from the test tube to the field (Wang and Hu, 1982). The first establishment of tissue culture from potato tubers was attempted as early as 1951 (Steward and Caplin, 1951), and since then, a variety of tissues from different plant organs, such as leaves, petioles, internode segments, ovaries, anthers, stems, roots, and shoot tips have been successfully demonstrated (reviewed in Wang and Hu, 1985; Bajaj, 1987; Wenzel, 1994; Naik et al., 2000; Naik and Sarkar, 2001). Seed potato production involving micropropagation techniques can overcome many of the problems associated with the traditional seed production system (Singh et al., 2019b). Figure 1 illustrates tissue culture-based commercial seed production systems.

Meristem culture for virus elimination. The meristem culture procedure is combined with thermotherapy and/or chemotherapy to increase the likelihood of obtaining virus-free plants (Sarkar et al., 2011). Even after taking all precautions for virus elimination, only a few virus-free mericlones are obtained. It is, therefore, essential that each mericlone is tested for viruses using enzyme-linked immunosorbent assay (ELISA),



immunosorbent electron microscopy (ISEM), and polymerase chain reaction (PCR) techniques before it is used as a source plant in a large-scale micropropagation program (Naik and Buckseth, 2018). The virus-indexed and pathogen-free mericlones are subjected to a rapid tissue culture method to generate abundant quantities of pathogen-free cultures (Naik and Karihaloo, 2007).

Micropropagation. Single-node cuttings of virus-free potato mericlones are grown in semi-solid or liquid culture media under aseptic conditions to obtain new microplants (Naik et al., 2000). Hormone-free Murashige and Skoog's (MS) medium supplemented with 2.0 mg L⁻¹ D-calcium pantothenate and 30 g L⁻¹ sucrose is ideally suited for large-scale *in vitro* micropropagation of potatoes (Buckseth et al., 2016). Cultures are incubated under a 16-h photoperiod (60 μmol m⁻² s⁻¹ PFD) at 20 ± 2°C. Usually, three single-node cuttings with one or two leaves are planted into each culture tube (25 × 150 mm). Within 3 weeks, the axillary/apical buds of these cuttings develop into full plants. These microplants are further sub-cultured up to 10 times on fresh culture media at an

interval of 3 weeks. Therefore, it has the potential to produce about 59,049 microplants from a single culture tube within 7–8 months under controlled conditions [National Academy of Agricultural Sciences (NAAS), 2021]. Micropropagation now underpins many seed potato production systems and specifically provides the nuclear stock material, in the form of microplants or microtubers (*in vitro* produced tubers) for their subsequent use in potato seed production channel (Millam and Sharma, 2007; Chindi et al., 2014).

Microtuber production from microplants. Microtubers are miniature tubers produced *in vitro* under tuber-inducing conditions (Naik and Karihaloo, 2007). These small dormant tubers are particularly convenient for handling, storage, transportation, and conserving germplasm (Gopal et al., 2004). In general, 15–20 microtubers with an average size of about 100–200 mg, are obtained from each tissue culture flask. Microtubers are usually not used for raising commercial crops but utilized for the production of minitubers in greenhouses (Naik and Sarkar, 2000). The microtubers are planted in nursery beds under aphid-proof net houses (50 microtubers m²) to produce minitubers. This technology, however, has not become very popular because of the poor survival of microtubers (60–65%) and the poor crop emergence (50%–60%) in nursery beds (Venkataslam et al., 2017). Bioreactors have also been tried for the mass production of microtubers (Ankita and Takayama, 1994; Piao et al., 2003); however, bioreactors are expensive and add to the cost of microtuber production (Mamiya et al., 2020).

Minituber production in soil. Minitubers are small seed potato tubers that can be produced in glasshouses or aphid-proof nethouses from *in vitro* propagated plantlets planted at a high density (Buckseth et al., 2020). Minitubers can be produced using either microplants as planting material or microtubers (Naik, 2005). In this system, the hardened virus-free microplants or microtubers are transplanted inside a nethouse for the production of minitubers. The minitubers so obtained are multiplied usually in two subsequent field generations (generations 1 and 2) before supplying to farmers as seeds, thereby reducing the number of field exposures of initial disease-free material.

Soil-less aeroponic system. Aeroponics is a more recent technology that has made inroads into the potato sector in the last few decades. This technology has been developed for the production of minitubers by utilizing healthy *in vitro* plants (Otazu, 2010; Singh et al., 2012; Pandey and Singh, 2014; Buckseth et al., 2016). This system facilitates year-round production and adoption of phytosanitary standards (Singh et al., 2012; Tierno et al., 2014; Singh et al., 2016). In this system, microplants are planted on top of the growth chamber, and the developing root zone inside the chamber is fogged with nutrient solution (Tierno et al., 2014; Buckseth et al., 2016). The chambers are installed inside an insect-proof nethouse. The aeroponic chamber has a removable opening at the top with

holes for holding the potato plants. The front of the aeroponic chamber is set with pivots and can be opened to harvest minitubers of ideal size at different time intervals. Picking of tubers begins after 45–50 days of planting when they attain a size of 3–10g. Picking of minitubers is done every week, and around 10–12 harvests are taken during the whole crop season of 4–5 months (Tiwari et al., 2019). Normally, 40–50 minitubers can be harvested from a single *in vitro* plant depending upon the variety as against 8–10 minitubers under the nethouse in nursery beds (Buckseth et al., 2020). The harvested minitubers are stored at 2° C–4°C to be utilized for planting in the next crop season. This system requires a high level of specialization in planning, operational expenses, and standardization of genotype-responsive nutrient solutions, but it offers quality seeds and has revolutionized seed potato production in India (Buckseth et al., 2022). Variation for root morphology and yield traits of all Indian varieties under aeroponics has also been identified (Tiwari et al., 2022).

Apical rooted cuttings technology. Rapidly growing young vegetative part of the potato plant can be cut and rooted in a number of ways (Bryan et al., 1981). Apical rooted cuttings have long been used in Southeast Asia (Vander Zaag and Escobar, 1990), particularly in Vietnam (Tran et al., 1990). The approach could be duplicated elsewhere in the developing world. On the same note, this technology has been successfully integrated into the potato breeders' seed production program in India (Buckseth et al., 2019b). Apical rooted cuttings (ARCs) is simple, effective, easy to implement, and has a small production cycle; this has an edge over aeroponic technology, wherein the large capital investment and long production cycle required are limiting factors (Buckseth et al., 2019a). In ARC, the healthy buffer stock of microplants acts as initial planting material on nursery beds (400 microplants m²). The first round of cutting starts after 15–20 days of planting of the mother stock. The apical portion (1.5–2.0 cm) of the growing microplant is cut and planted in pro-trays containing cocopeat/other soilless media. Once the apical dominance in the mother plant is removed, the growth of lateral buds is promoted, and these buds grow into new branches, which are further used to increase the multiplication rate of potato plants. Sequential cuttings of laterals from mother plants are followed at an interval of 7–10 days, and it continues depending on the growth of new shoots in the variety for over 35–45 days. To follow the seed plot technique (i.e., growing potato seed crop under an aphid-free period of the season) and to fit into the seed production window, only three to four cuttings are recommended in the northwestern central plains of India. Thus, one microplant can produce six to eight rooted cuttings (Buckseth et al., 2022). Around 7–10-day-old rooted cuttings are planted under insect-proof nethouse for tuber production. Batch-wise pedigree of the cuttings is maintained so as even if any plant out of a cutting is found

infected with a virus during testing, all the counterparts of the cutting/sister counterpart tubers can be rejected. Based on the available data, 7–10 tubers weighing 10–70 g can be harvested from a single cutting, thereby achieving a multiplication rate of more than 40 depending upon the variety. This technology has immense potential if it can be fitted in the seed production window following standard operating procedures to ensure seed health.

Botanical true potato seed

The idea of true potato seed (TPS) was conceived by Dr. S. Ramanujam as early as 1949. Like all other botanical seeds, TPS has the potential to grow into a full plant, but every plant is genetically different, thus making TPS population heterogeneous (Gaur and Pandey, 1993). Efforts were made at ICAR-CPRI to develop uniform TPS populations to get uniform crop stand, disease resistance, and tuber yields. TPS has many advantages over potato seed tubers. The major ones are disease- and pest-free planting material, easy storage and transportation, and a highly reduced seed rate (about 150 g/ha as against 2.5–3.0 t/ha of seed tubers). Three TPS populations, namely, HPS-I/13, TPS-C-3, and 92-PT-27, were developed by ICAR-CPRI during the 1980s. Quality control parameters for TPS production have been well prescribed in the Indian Minimum Seed Certification Standards (Indian Minimum Seed Certification Standards, 2013). However, TPS technology remains to be seen as a ready-to-use technology with the economic comparison of TPS versus seed tubers in the dynamics of the cropping system for potato production (Tiwari et al., 2017). The use of TPS appears to be the technology to solve the problem of a shortage of good quality potato seed tubers in developing countries (Muthoni et al., 2019). The obstacles that have prevented the adoption and widespread use of TPS are the late maturity of a TPS crop, unreliable germination, and non-uniformity of the produce. However, there has been a renewed enthusiasm for TPS technology since the advent of diploid F1 hybrid breeding in 2008 (which was declared International Potato Year by the UN).

Diploid hybrid TPS

Being an autotetraploid and highly heterozygous crop, the selfing of potatoes results in severe inbreeding depression. Bringing down the ploidy to a diploid level and selfing thereafter through the incorporation of a self-compatibility gene can result in homozygosity at a much faster rate than at the tetraploid level. This approach has been demonstrated by

Lindhout et al. (2011). Since the offspring of a sexual cross would be a pristine true seed, which would be completely free of diseases and therefore make excellent seed material for potato growers around the world (Sood et al., 2021). Diploid potatoes can be converted from an outbreeding species, in which self-pollination is prevented by a gametophytic self-incompatibility system, into one where self-pollination is possible, either through a dominant self-incompatibility inhibitor gene (Sli) or knockout mutations in the incompatibility locus. As a result, diploid F1 hybrid breeding can be used to produce genetically uniform potato cultivars for propagation from true potato seeds by crossing two near-homozygous inbred lines, derived from a number of generations of self-pollination despite inbreeding depression (Bradshaw, 2022). Later, the developed F1 hybrid planting material can be delivered to farmers as true seeds or young plants and minitubers derived from true seeds. Many public and private organizations, including ICAR-CPRI, are working on the development of diploid inbred lines and exploiting heterosis in potatoes at the diploid level. Although the methodology of F1 diploid hybrid TPS-based potato breeding has several advantages over the conventional tetraploid tuber-based approach, a number of challenges need to be overcome to bring this technology to the forefront in the farmer's field (Sood et al., 2021).

Concluding remarks

In mitigating the scarcity of good quality seeds, strategies to produce good quality potato seeds using tissue culture in conjunction with aeroponic systems and apical rooted cuttings have been tried. Currently, about 30% of potato breeders' seed (basic seed) in India is being produced using these hi-tech methods. The merits and demerits of conventional and hi-tech seed potato production systems are highlighted in Table 1. These technologies need to be given serious thought and should be promoted in developing countries so as to increase potato productivity. In areas having high disease pressure, the hi-tech system of seed potato production has the advantage of better health status of seed stocks due to the reduced number of field multiplications over the conventional (clonal multiplication) system. In terms of the need for greater efficiency of seed potato production and for reduced energy input, research on soil-free techniques will continue to be the subject of focus in both established and developing potato-producing areas in the near and distant future. Advances in engineering technology will also assist in the development of more automated and controlled seed propagation systems. However, there are also options for simplifying the seed potato production systems for adaptation to low-cost technology situations for resource-poor potato growers,

TABLE 1 Conventional *vis-à-vis* hi-tech system of seed potato production.

Parameters	Conventional system	Hi-tech system
Type of multiplication	Mainly based on the clonal selection of indexed tubers and their multiplication in subsequent stages in the field.	This system is based on tissue culture, thus ensuring 100% healthy material as a seed source.
Procedure	Involves successive multiplication of nucleus seed tubers through super-elite, elite tubers.	Involves successive multiplication of tissue culture material either directly through minituber production or through aeroponics or through apical rooted cuttings.
Field exposure	Higher field exposure of 7-8 years.	Reduces the field exposure and thereby minimizes the accumulation of viruses in the seed stocks.
Uniformity	Less uniform.	More uniform production of the initial generation of potatoes.
Speed of the multiplication	Slow	Fast
Seed multiplication rate	Multiplication rate is low, varying from 1:4 to 1:15 (one tuber yields 4–15 tubers) depending upon variety, agro-climatic conditions, and crop management practices.	Higher multiplication rate (avg. 1:40-50) depending upon variety.
Period	Seed window specific particularly in tropical and sub-tropical regions where potato is a winter crop.	Allowed off-season multiplication of the pre-nucleus seed.
Disease incidence/ degeneration of stock	Higher degeneration of seed stocks from one generation to another due to accumulation of bacteria, fungi, viruses, and viroids.	Degeneration of stock is less due to less field exposure
Requirement of Land	Require more land due to bulkiness and more spacing in early seed multiplication.	Require less land due to small and uniform planting material and less spacing in early seed multiplication.
Cost	Usually, tubers produced in the conventional system are of large size, and the whole seed tubers have to be planted for a good quality crop. Therefore, the seed cost is high.	The initial establishment cost is very high, which can be recovered after 3-4 crop seasons.
Benefit	In India, presently, 70% of seed production is done through the conventional system, which meets the seed requirement of the country on a yearly basis.	Clean seed tubers, free from all diseases obtained in a very less span of time.

which has greater scope and relevance for the expansion of potato production in developing countries.

Author contributions

TB conceived the idea and wrote the manuscript. JT, AS, DD, VB, SS, MS, CC, and SN performed research and literature collection. VK, RS, MK, and NP edited the manuscript. All authors approved the manuscript.

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