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Aging is a non-modifiable risk factor for stroke and the global burden of stroke is continuing to increase due to the aging society. Muscle dysfunction, common sequela of stroke, has long been of research interests. Therefore, how to accurately assess muscle function is particularly important. Electrical impedance myography (EIM) has proven to be feasible to assess muscle impairment in patients with stroke in terms of micro structures, such as muscle membrane integrity, extracellular and intracellular fluids. However, EIM alone is not sufficient to assess muscle function comprehensively given the complex contributors to paretic muscle after an insult. This article discusses the potential to combine EIM and other common quantitative methods as ways to improve the assessment of muscle function in stroke survivors. Clinically, these combined assessments provide not only a distinct advantage for greater accuracy of muscle assessment through cross-validation, but also the physiological explanation on muscle dysfunction at the micro level. Different combinations of assessments are discussed with insights for different purposes. The assessments of morphological, mechanical and contractile properties combined with EIM are focused since changes in muscle structures, tone and strength directly reflect the muscle function of stroke survivors. With advances in computational technology, finite element model and machine learning model that incorporate multi-modal evaluation parameters to enable the establishment of predictive or diagnostic model will be the next step forward to assess muscle function for individual with stroke.

KEYWORDS

electrical impedance myography, stroke, skeletal muscle, spasticity, ultrasonography

Introduction

Population aging is an urgent global issue, accompanied with a substantial burden for age-related diseases. It was reported that about 1,000 among every hundred thousand people aged 70 years or older suffer stroke in the world (Feigin et al., 2021). As one of the leading causes of death and disability in the elderly, stroke has long been of interest, particularly in motor function recovery (Lo, 2014; Feigin et al., 2022). Approximately 80% of stroke survivors suffer from motor dysfunction, including hemiparesis, incoordination, and spasticity, which severely hinder their daily life (Lora-Millan et al., 2022). How to improve motor control and movement quality are challenging clinicians and researchers (Patten et al., 2004). The smooth control of movement relies not only on the integrity of neural pathways, but also on the function of the musculoskeletal system. Therefore, assessing muscle function is essential to understand factors underlying motor dysfunction after stroke. Typically, muscle function is affected by its structural changes (Power et al., 2013). For stroke survivors, disrupted synaptic inputs to the motor neurons commonly resulted in muscle fiber loss, intramuscular fat infiltration and connective tissue growth (Scherbakov et al., 2013; Li et al., 2016d). All of these changes could influence the intrinsic muscle properties (Li et al., 2017c). In turn, the evaluation of muscle properties may reflect the muscle function and improve current knowledge on the mechanisms of muscle dysfunction in stroke survivors.

Electrical impedance myography (EIM), a non-invasive evaluation instrument, aims at quantifying the intrinsic passive electrical properties of muscle (Rutkove, 2009). Alterations in the voltage caused by changes of diseased muscle conductivity and permittivity are the basis of EIM (Foster and Schwan, 1989). It has been studied that EIM parameters, resistance, reactance and phase angle, are able to reflect specific physiological information on the composition and micro structures of muscle tissue respectively, e.g., myofiber membrane, extracellular and intracellular fluids (Castizo-Olier et al., 2018; Fu and Freeborn, 2020). In clinical setting, electrical impedance variables have been verified as reliable biomarkers for neuromuscular diseases, such as amyotrophic lateral sclerosis and muscular dystrophy (Rutkove and Sanchez, 2019). Clark et al. (2021) reviewed the feasibility of EIM in age-related diseases and affirmed its potential clinical value in the assessment of muscle health concerning aging. In elderly with stroke, we reported significant changes in the passive electrical behavior of muscle (Li et al., 2017c). Similar results were also observed in patients with spinal cord injury (Li et al., 2017a,b). How to interpret the changes recorded by EIM has always been the focus of clinical application. With the help of animal experiments, electrical impedance data have been proved to be involved with the structural degeneration of muscle, especially in the number of muscle fibers, state of the cell membranes and the proportion of fat and connective tissue infiltration (Pandeya et al., 2021b,a). From a functional perspective, Li et al. (2016a) observed a significant relationship between EIM parameters and force production in mice. It follows that EIM has the potential to establish the relationship between muscle function and microstructural properties, providing physiological evidence for muscle evaluation. Furthermore, our team also demonstrated excellent sensitivity of EIM technique in the measurement of alterations on muscle composition in stroke patients (Li et al., 2017d). Coupled with excellent robustness and convenience, EIM holds a high potential for clinical assessment of muscle function (Martinez-Gonzalez et al., 2020; Rutkove et al., 2021). However, there have been various contributors to muscle dysfunction in persons after stroke and the passive electrical property measured by EIM may not fully reflect the paretic muscle changes.

Given that the factors leading to muscle dysfunction are uncertain and the sensitivity of techniques varies at different pathological features of muscle, there is a need for combined assessments of muscle function in clinical settings. Apart from muscle properties itself, its dysfunction tends to initially derive from the denervation of muscle after stroke (Scherbakov et al., 2013) and follows that nerve-related factors are indispensable for muscle function evaluation. Thus, EIM alone is not sufficient for assessment of muscle function comprehensively. Additionally, the clinical value of EIM remains controversial (Rutkove et al., 2021; Sanchez et al., 2021). Sanchez et al. (2021) pointed out that EIM measurements is likely to be influenced by the accuracy of measurement localization and the specificity of the target tissue (i.e., subcutaneous fat tissues and skin tissues affect the measurements). Although previous work has drawn links between EIM values and electrophysiological, histological, imaging and functional outcomes (Rutkove et al., 2014a; Hamel et al., 2020), the interpretation of EIM values is suggested to be limited and EIM cannot substitute for other assessments in many cases. It is noted that EIM is only capable of assessing superficial muscles but not the deep muscle layer, owing to the limited penetrative ability of electrical current (Hu et al., 2021c). Therefore, the EIM findings need to be interpreted with caution in clinical decision-making.

Over the past decades, quantitative assessment of muscle function gained popularity due to their precision and objectivity. These assessment methods evaluate various aspects influencing muscle function, including morphological, mechanical and contractile properties. Here, we discuss the potential utility for combined assessments of EIM with other quantitative techniques and provide the insights and challenges for evaluating muscle function in patients with stroke (Figure 1). The main findings are displayed in the Table 1.

Combination of EIM and morphological assessment

The loss of muscle function is typically accompanied with changes of muscle morphology after stroke (Metoki et al., 2003). Common morphological features found in patients with stroke include a reduction of muscle cross-sectional area, mass, volume, pennation angle and fascicle length. Sions et al. (2012) proposed that the decrease in the muscle cross-sectional area would eventually lead to the loss of muscle strength. Moreover, intramuscular fat is also found associated with greater muscle torque (Goodpaster et al., 2001). It is possible that muscle morphological changes reflect its dysfunction indirectly. Assessments of muscle morphology adopted in clinical practice involve magnetic resonance imaging (MRI), computed tomography (CT), dual-energy x-ray absorptiometry (DXA) and ultrasound imaging. Although these tools could provide reliable and accurate information on the morphology (Heymsfield et al., 2014; Clark et al., 2018), their clinical utility is limited by complex operation procedure, data analysis and expenses (Bazzocchi et al., 2016; Tavoian et al., 2019; Clark et al., 2021). Further, these are also challenges to incorporate with other kinetic instruments such as torque dynamometers simultaneously. Morphological



measurement alone does not give the overall muscle status qualification.

Faced with above limitations, increasing number of researchers turned their sights on EIM to search for alternative verifiable biomarkers of muscle status. There is evidence that the application of EIM can estimate well for the muscle cross-sectional area (Pandeya et al., 2021a), myofiber size (Arnold et al., 2017) and arrangement of myofibers (Garmirian et al., 2009). In subacute stroke survivors, we have demonstrated significant associations of electrical impedance data with ultrasonography parameters (Hu et al., 2019). Muscle fascicle length, thickness and pennation angle recorded by ultrasound have been recognized to effectively illustrate the muscle status in poststroke impairments (Li et al., 2007). According to previous reports, the EIM and ultrasound could be applied jointly to depict muscle architecture (Rutkove et al., 2010; Rutkove and Darras, 2013; Johnson et al., 2018; Hu et al., 2019; Longo et al., 2021). With the development of quantitative ultrasound, its relationship with EIM is playing an increasing important role in improving disease diagnosis (Rutkove et al., 2014b; Shklyar et al., 2015; Hobson-Webb et al., 2018, 2021; Roy et al., 2019). Shklyar et al. (2015) reported that composite data from EIM and quantitative ultrasound can better predict the muscle function compared with individual measurement data. Furthermore, several studies have established the relationship between EIM variables and structural features of muscles, as measured by MRI, CT, and DXA. Those studies also further supported the potential value of EIM in muscle structure assessment (Anderson et al., 2014; McLester et al., 2018; van Rassel et al., 2019; Czeck et al., 2020; Hamel et al., 2020; Albano et al., 2022b,a). While aforementioned correlation was validated, not all findings are fully consistent, since different techniques are sensitive to different pathological features of muscle (Roy et al., 2019). There is a need for combined assessment to assist in clinical muscle evaluation or diagnosis. The correspondence of variables between EIM and other quantitative techniques provide a foundation for cross-validation in muscle assessment and facilitates the accurate evaluation of muscle function.

In addition to accuracy, combined assessments could improve the understanding of mechanisms of muscle dysfunction. Parameters from EIM, including resistance, reactance and phase angle, are reported to be closely associated with muscle's components, extra- and intra-cellular fluids, tissue interfaces and the integrity of cell membrane (Esper et al., 2006). EIM data could be an important compliment to the morphological assessment. It is these relationships that help clinicians to understand the physiological significance behind morphological variables, further driving the development of precision therapy.

The combination with other assessment methods would advance the application of EIM. As the electrical impedance data is susceptible to subcutaneous fat tissue, our team investigate the effect of subcutaneous fat on EIM with ultrasound assistance and found appropriate electrode configuration to reduce the impact (Li et al., 2016b). Sung et al. (2013) found that the reactance, one of the EIM parameters, was least affected by subcutaneous fat when the EIM was applied with the aid of ultrasound. Conventional muscle morphological assessment may guide the application of EIM to improve accuracy and stability. If clinicians performed conventional assessments (e.g., ultrasound) and identify the amount of subcutaneous fat thickness in advance, they would select the appropriate electrode configuration to improve the accuracy and stability of measurement.

Combination of EIM and mechanical property assessment

Dystonia is a common muscle dysfunction for stroke patients, characterized by reduced or increased muscle tension (Tsai et al., 2020). These deficits could be attributed to inherent properties of contractile elements, viscoelastic properties of musculotendinous units and abnormalities of reflex excitability (Chuang et al., 2012). Thus, the assessment of both neural and mechanical properties is critical to provide a full picture of muscle function. Muscle strength and joint range of motion are often adopted as indirect measurements of muscle mechanical properties (Brandenburg et al., 2014). However, these indirect methods can only evaluate mechanical properties across the joint instead of individual muscles, potentially limiting precision assessment.

Myotonometry was proposed as a way to quantify individual muscle mechanical properties including stiffness, elasticity, and muscle tone (Chuang et al., 2013). The principle behind myotonometry is to apply brief impulses over the muscles to induce oscillations, which are recorded to calculate the parameters of tone, creep, stiffness and decrement (Lo et al., 2017). There have been several studies which reported the reliability and sensitivity of myotonometric measurement in stroke rehabilitation (Chuang et al., 2012; Li et al., 2017e). Previous work highlighted the potential benefit of applying myotonometry to quantify muscle mechanical properties. However, there is uncertainty regarding the interpretation of myotonometric data since there is little information regarding the normative values that could be used for reference. There are also uncertainties in the physiological contributing factors to alteration of muscle mechanical properties (Lo and Li, 2020). Thus, myotonometry is not recommended to be applied alone. Establishing the relationship between physiological structures and muscle mechanical properties remains necessary for researchers or clinicians to optimize therapeutic regimens. Unfortunately, it is not realistic to widely use muscle biopsy for assessing muscle structures in the clinic. According to our previous studies, reactance recorded by EIM was significantly associated with the creep and relaxation time from myotonometer in patients who suffered cervical spinal cord injury (Hu et al., 2021b). The former was reported to be related to myocyte atrophy (Li et al., 2012), which is also likely to lead to the reduction of muscle contractile ability. This change often reflects as higher relaxation time and creep in myotonometry. Similar relationships can also be observed in patients with stroke, where muscle viscosity was significantly associated with phase angle (Leng et al., 2021). These findings offer us an opportunity to interpret the mechanical property alterations from a physiological structural point. Whether or not the combined assessments may improve assessment of muscle function warrant further investigation.

Spastic hypertonia stems from the neural and non-neural factors is frequently observed in stroke survivors (Luo et al., 2020). Conventionally, the neural components are defined as the intrinsic excitability of motoneurons themselves, while nonneural components can be represented as the resistance caused by inertia, elasticity, and viscosity of the moved body part (Lindberg et al., 2011). Distinguishing neural and non-neural components is critically important for developing targeted intervention regimens. Spastic muscle mainly caused by neural contribution is suited to treatment against nerve-pathway (e.g., nerve block) (Bovend'Eerdt et al., 2008). While varieties of approaches proposed to distinguish the neural factors (Lee, 2002; Kamper et al., 2003; Musampa et al., 2007; Alibiglou et al., 2008), it is hard for these methods to truly distinguish neural components and their clinical utilities are limited by operational complexity (Wood et al., 2005). Lindberg et al. (2011) built a biomechanical model, Neuroflexor model, to quantify the neural and non-neural contributions by measuring the resisting force during passive extension at slow and fast speeds. Our previous studies reported the higher neural, elastic and viscosity components of the Neuroflexor method on the affected side compared to non-affected side in stroke patients (Leng et al., 2021). Combined with EIM, we found EIM parameters of reactance was inversely proportional to neural component, while phase angle was proportional to viscosity component in spastic muscle, potentially supporting the feasibility of combined assessment based on EIM and Neuroflexor in stroke patients. This relationship also provides the basis to explore the effect of nervous factors on peripheral muscular structures.

Combination of EIM and contractile property assessment

Motor function deficits, those caused by muscle weakness, can only be observed during voluntary muscle contraction. Thus, understanding of muscle contractile property (i.e., shortening velocity and maximal force) is of great interest (Gao et al., 2005). Active movement tests, such as the handgrip test, are easily performed but their results are susceptible to be affected by the integrity of the nervous system and subjective factors (Carson, 2018). That is, these tests fail to accurately distinguish the true cause of muscle dysfunction (nerve or muscle related). Prior studies have found that muscle weakness was associated with the loss of muscle fiber and increase in intramuscular adipose tissue, caused by muscle denervation, disuse, and atrophy (Scherbakov et al., 2015; Berenpas et al., 2017). Numerous researches have reported the association of muscle strength with various macrostructures clinically (Petterson et al., 2008; Kellis et al., 2021). However, these studies cannot explore muscle dysfunction at a micro level. Building on strength testing, the addition of EIM, which reflects indirectly the properties of cell membranes and the content of extracellular and intracellular water (Faes et al., 1999; Shiffman and Rutkove, 2013), probably provide advantages for the exploration of mechanism behind muscle dysfunction. Several studies have

TABLE 1 The main findings of each section.

Combining EIM with	Assessment	Study	Main findings
Morphological Property Assessment	Ultrasound	Rutkove et al., 2010; Rutkove and Darras, 2013; Johnson et al., 2018; Hu et al., 2019; Longo et al., 2021	 Clinical measurements from EIM and ultrasound were verified mutually. Both methods could be applied applied to depict muscle architecture and improve disease diagnosis accuracy.
		Sung et al., 2013; Li et al., 2016b	 The application of ultrasound could guide the utilization of EIM for improved accuracy and stability. The application of ultrasound could evaluate the influence of subcutaneous fat layer thickness on EIM measurements.
		Rutkove et al., 2014b; Shklyar et al., 2015; Hobson-Webb et al., 2018, 2021; Roy et al., 2019	 Significant associations were observed between measurements obtained from EIM and those obtained from quantitative ultrasound. Combining the data from these two techniques has been shown to yield more accurate muscle function measurements.
	MRI	Hamel et al., 2020; Albano et al., 2022a,b	 Establishing the correlation of the EIM with structural MRI features. Comparing EIM and MRI measurements in assessing muscle composition.
	СТ	Anderson et al., 2014	• Paraspinal electrical impedance myography phase was positively correlated with paraspinal attenuation based on CT, which established the relationship between EIM and CT.
	DXA	McLester et al., 2018; van Rassel et al., 2019; Czeck et al., 2020	• EIM has been proved to be a relatively reliable method to estimate body fat percent compared to DXA.
Mechanical Property Assessment	NeuroFlexor	Leng et al., 2021	 Neural component was found significantly associated with reactance recorded by EIM. Viscosity component was significantly associated with phase angle.
	Myotonometer	Hu et al., 2021b	• The creep and relaxation time obtained from myotonometer was significantly associated with reactance recorded by EIM.
Contractile Property Assessment	Strength test	Rutkove et al., 2014a; Shellikeri et al., 2015; Zaidman et al., 2015	• The observed relationship between EIM and strength tests suggested that EIM had the potential as a meaningful measure in the diagnosis of neuromuscular diseases.
		Li et al., 2016a,c; Coutinho et al., 2020	• The combination of EIM and strength test has the potential to directly reveal the changes in the force generation properties of the target muscle.
	EMG	Li et al., 2013, 2014, 2017f; Arnold et al., 2016, 2017; de Carvalho and Swash, 2016; Kolb et al., 2016; Zong et al., 2018	 EIM was closely associated with various electrophysiological measures, especially compound muscle action potential from EMG. EIM is a promising biomarker in assessment of muscle function in neuromuscular diseases and its relationships with other electrophysiological measures have been well-established in various neuromuscular diseases.
		Hu et al., 2021a; Ngo et al., 2022	 Applying EIM and EMG simultaneously could provide more comprehensive information of the inherent properties and muscle activation alteration.
Computational Model	Finite element model	Ahad and Rutkove, 2009; Wang et al., 2011; Jafarpoor et al., 2011; Pacheck et al., 2016	• Finite element models have been demonstrated as a capable method to investigate the biophysical mechanisms of EIM in various diseases.
		Jafarpoor et al., 2013; Baidya and Ahad, 2016; Rutkove et al., 2017; de Cardoner et al., 2021; Schooling et al., 2020; Luo et al., 2022; Schrunder et al., 2022	• Finite element analysis provides a basic methodology to optimize electrode configuration for EIM
	Machine learning model	Srivastava et al., 2012; Pandeya et al., 2022	• Combining biomarkers from EIM or other assessments could enhance the diagnostic performance of machine learning models.
		Kapur et al., 2018a,b; Pandeya et al., 2021a,b; Cheng et al., 2022	• Combining EIM and other biomarkers through machine learning can improve diagnostic accuracy for estimating muscle function parameters.

explored the relationship between the EIM and strength tests in neuromuscular diseases, which suggested that EIM may serve as a valuable measure for diagnosing neuromuscular conditions (Rutkove et al., 2014a; Shellikeri et al., 2015; Zaidman et al., 2015). Our findings indicated that this combination could also reveal the changes in properties of target muscle directly during force generation (Li et al., 2016c). Likewise, the relationship between the EIM data and force production was verified in animal experiments (Li et al., 2016a; Coutinho et al., 2020). These insights might be beneficial when further extended to clinical assessments of alterations in force production after stroke.

Actually, the generation of force is inseparable from active muscle contraction, depending on the number and size of active motor units, as well as the rate and timing of their discharge (Herzog et al., 2015; Duchateau and Enoka, 2016). In the past decades, EMG assessment has been recognized as a gold standard for evaluating the recruitment and activation patterns of muscle (Schuermans et al., 2014). Compared with strength testing, EMG records electrophysiological data to explain the potential reasons for the impairment of muscle contractile function. However, EMG is limited to the evaluation of active electrophysiological property without concerning passive property. From a mechanistic perspective, the relationships between EIM data and EMG parameters, especially compound muscle action potentials, have been demonstrated in persons with various neuromuscular disease over the years (Li et al., 2013, 2014, 2017f; Arnold et al., 2016, 2017; de Carvalho and Swash, 2016; Kolb et al., 2016; Zong et al., 2018). Our team has demonstrated the potential of the EIM and EMG to jointly evaluate lower-extremity muscle function and to assess the impact of rehabilitation training in a clinical setting (Hu et al., 2021a; Li et al., 2022). Both active and passive muscle properties were assessed at the same time, which could advance the understanding of muscle dysfunction. Ngo et al. (2022) proposed a novel device that can combine EIM and EMG measurements simultaneously, offering a more robust method for muscle status assessment even in the presence of artifacts.

Combination of EIM and computational model

Standards for EIM parameters in clinical applications are still lacking, especially when the comparison of data across individuals is considered. As this technique is sensitive to muscle morphological, electrical properties and the electrode configuration (Schrunder et al., 2022), it is then necessary to identify the impact of these factors and to explore approaches to minimize their effects. Computational modeling that incorporates detailed finite element models has been proven capable in studying the biophysical mechanisms of EIM and optimize the design of electrode arrangements (Ahad and Rutkove, 2009; Jafarpoor et al., 2011, 2013; Wang et al., 2011; Baidya and Ahad, 2016; Pacheck et al., 2016; Rutkove et al., 2017; Schooling et al., 2020; de Cardoner et al., 2021; Luo et al., 2022; Schrunder et al., 2022). Wang et al. (2011) utilized a finite element model to investigate the correlation between changes in surface impedance and electrical properties of the muscle, offering valuable insights into the biophysical mechanisms underlying the EIM. Schrunder et al. (2022) simulated muscle tissues and impacts from anthropometric variations and EIM electrode placements. Baidya and Ahad (2016) applied a finite element model to determine an optimized EIM electrode configuration which could balance both stability and sensitivity. However, previous finite element studies seldom focus on the application of EIM in stroke survivors. This work needs to be studied based on the actual data from individuals with stroke in the future.

As the data of muscle assessment becomes increasingly diverse, how to identify and integrate valuable information are attracting more and more attention. Multimodal learning has grown rapidly with the continuous progress of multimodal information input and algorithm (Baltrusaitis et al., 2019). Drawing on a muscle perspective, the multi-modality fusion of different assessments is considered to substantially drive the clinical assessment advancement with the help of machine learning. Srivastava et al. (2012) refined biomarkers from EIM and quantitative muscle ultrasound on the basis of machine learning to successfully classify the muscle affected by spinal muscular atrophy. Pandeya et al. (2022) demonstrated that using multifrequency EIM values instead of single-frequency values can improve classification performance of machine learning. Similarly, Cheng et al. (2022) reported that machine learning could estimate the total mass of muscles with the EIM and anthropometric parameters. The use of machine learning approaches has allowed for the prediction of myofiber size, cross-sectional area, and connective tissue deposition using the EIM measurement (Kapur et al., 2018b,a; Pandeya et al., 2021b,a; Cheng et al., 2022). Addressing different requirements, specific quantitative assessment parameters are selected to be combined with electrical impedance data. Currently, the combined assessments based on machine learning are rarely employed to detect muscle changes in stroke patients. Establishing a prediction or diagnostic model based on machine learning to assess muscle function for patients with stroke will be the next step forward.

Conclusions and future directions

Overall, the application of combined assessments based on EIM with quantitative techniques is a promising direction to assess muscle function in stroke patients. In view of different purposes, distinct combinations are selected to evaluate specific aspect of muscle function. Combining different techniques could improve the accuracy and comprehensiveness of muscle assessment, and also advance the understanding of physiological mechanism that underpins muscle dysfunction.

Despite the potential of combined assessments in clinical application, several challenges remain at both the mechanistic and clinical levels.

At the mechanistic level, there were lacks of robustness in identifying the relationship between EIM and standard parameters of other assessments, thereby hindering the application of EIM in clinical settings. Establishing this relationship requires a deeper understanding of the biophysical mechanisms of EIM, which requires further investigation. Secondly, most studies cannot provide a causal interpretation, and the sequence of

alterations in variables remains unclear. To address this issue, more longitudinal studies with longer period of follow-ups are required. Thirdly, research efforts in the field of muscle function assessment have been uneven, with the combination of EIM and mechanical property assessment receiving less attention and requiring further exploration (see Table 1). Furthermore, enabling clinical application also raises unique challenges. On the one hand, selecting effective combination of assessments for a specific purpose remains nontrivial for clinicians and researchers. As sensitivity varies across different assessments, selecting appropriate combination and extract essential features from multiple types of data are particularly crucial. The advancement of various computational techniques can assist in identifying, extracting, and analyzing multimodal information of muscle function, leading to the standardization in the clinical practice. On the other hand, the estimation accuracy of EIM has been a concern for years. Good accuracy relies on proper electrode configuration where finite element modeling could inform clinicians in an appropriate electrode configuration and placement. However, performing modeling for each patient based on individual MRI or CT scans is impractical. Therefore, building prediction models based on multimodal information is necessary. In summary, both opportunities and challenges exist in mechanistic studies and clinical applications for EIM.

Author contributions

LL and ZG: conceptualization. ZG, WL, RW, and LL: methodology. ZG and WL: writing – original draft preparation. WL, RW, and LL: writing – review and editing. All authors contributed to the article and approved the submitted version.

References

Ahad, M. A., and Rutkove, S. B. (2009). "Finite element analysis of electrical impedance myography in the rat hind limb," in *Proceedings of the 2009 Annual International Conference of the IEEE Engineering in Medicine and Biology Society*, (Piscataway, NJ: IEEE), 630–633. doi: 10.1109/IEMBS.2009.53 34069

Albano, D., Gitto, S., Vitale, J., Bernareggi, S., Aliprandi, A., Sconfienza, L. M., et al. (2022a). Comparison between magnetic resonance imaging and electrical impedance myography for evaluating lumbar skeletal muscle composition. *BMC Musculoskelet. Disord.* 23:970. doi: 10.1186/s12891-022-05902-9

Albano, D., Gitto, S., Vitale, J., Bernareggi, S., Lamorte, S., Aliprandi, A., et al. (2022b). Knee muscles composition using electrical impedance myography and magnetic resonance imaging. *Diagnostics* 12:2217. doi: 10.3390/diagnostics12092217

Alibiglou, L., Rymer, W. Z., Harvey, R. L., and Mirbagheri, M. M. (2008). The relation between Ashworth scores and neuromechanical measurements of spasticity following stroke. *J. Neuroeng. Rehabil.* 5:18. doi: 10.1186/1743-0003-5-18

Anderson, D. E., Bean, J. F., Holt, N. E., Keel, J. C., and Bouxsein, M. L. (2014). Computed tomography-based muscle attenuation and electrical impedance myography as indicators of trunk muscle strength independent of muscle size in older adults. *Am. J. Phys. Med. Rehabil.* 93, 553–561. doi: 10.1097/PHM.00000000000059

Arnold, W., McGovern, V. L., Sanchez, B., Li, J., Corlett, K. M., Kolb, S. J., et al. (2016). The neuromuscular impact of symptomatic SMN restoration in a mouse model of spinal muscular atrophy. *Neurobiol. Dis.* 87, 116–123. doi: 10.1016/j.nbd.20 15.12.014

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Conflict of interest

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Supplementary material

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Arnold, W. D., Taylor, R. S., Li, J., Nagy, J. A., Sanchez, B., and Rutkove, S. B. (2017). Electrical impedance myography detects age-related muscle change in mice. *PLoS One* 12:e0185614. doi: 10.1371/journal.pone.0185614

Baidya, S., and Ahad, M. A. (2016). Assessment of optimized electrode configuration for electrical impedance myography using genetic algorithm via finite element model. *J. Med. Eng.* 2016;9123464. doi: 10.1155/2016/9123464

Baltrusaitis, T., Ahuja, C., and Morency, L.-P. (2019). Multimodal machine learning: a survey and taxonomy. *IEEE Trans. Pattern Anal. Mach. Intell.* 41, 423–443. doi: 10.1109/TPAMI.2018.2798607

Bazzocchi, A., Filonzi, G., Ponti, F., Albisinni, U., Guglielmi, G., and Battista, G. (2016). Ultrasound: which role in body composition? *Eur. J. Radiol.* 85, 1469–1480. doi: 10.1016/j.ejrad.2016.04.005

Berenpas, F., Martens, A.-M., Weerdesteyn, V., Geurts, A. C., and van Alfen, N. (2017). Bilateral changes in muscle architecture of physically active people with chronic stroke: a quantitative muscle ultrasound study. *Clin. Neurophysiol.* 128, 115–122. doi: 10.1016/j.clinph.2016.10.096

Bovend'Eerdt, T. J., Newman, M., Barker, K., Dawes, H., Minelli, C., and Wade, D. T. (2008). The effects of stretching in spasticity: a systematic review. *Arch. Phys. Med. Rehabil.* 89, 1395–1406. doi: 10.1016/j.apmr.2008.02.015

Brandenburg, J. E., Eby, S. F., Song, P., Zhao, H., Brault, J. S., Chen, S., et al. (2014). Ultrasound elastography: the new frontier in direct measurement of muscle stiffness. *Arch. Phys. Med. Rehabil.* 95, 2207–2219. doi: 10.1016/j.apmr.2014. 07.007

Carson, R. G. (2018). Get a grip: individual variations in grip strength are a marker of brain health. *Neurobiol. Aging* 71, 189–222. doi: 10.1016/j.neurobiolaging.2018.07.023

Castizo-Olier, J., Irurtia, A., Jemni, M., Carrasco-Marginet, M., Fernández-García, R., and Rodríguez, F. A. (2018). Bioelectrical impedance vector analysis (BIVA) in sport and exercise: systematic review and future perspectives. *PLoS One* 13:e0197957. doi: 10.1371/journal.pone.0197957

Cheng, K.-S., Su, Y.-L., Kuo, L.-C., Yang, T.-H., Lee, C.-L., Chen, W., et al. (2022). Muscle mass measurement using machine learning algorithms with electrical impedance myography. *Sensors* 22:3087. doi: 10.3390/s22083087

Chuang, L., Wu, C., and Lin, K. (2012). Reliability, validity, and responsiveness of myotonometric measurement of muscle tone, elasticity, and stiffness in patients with stroke. *Arch. Phys. Med. Rehabil.* 93, 532–540. doi: 10.1016/j.apmr.2011.09.014

Chuang, L.-L., Lin, K.-C., Wu, C.-Y., Chang, C.-W., Chen, H.-C., Yin, H.-P., et al. (2013). Relative and absolute reliabilities of the myotonometric measurements of hemiparetic arms in patients with stroke. *Arch. Phys. Med. Rehabil.* 94, 459–466. doi: 10.1016/j.apmr.2012.08.212

Clark, B. C., Rutkove, S., Lupton, E. C., Padilla, C. J., and Arnold, W. D. (2021). Potential utility of electrical impedance myography in evaluating age-related skeletal muscle function deficits. *Front. Physiol.* 12:666964. doi: 10.3389/fphys.2021.666964

Clark, B. C., Tavoian, D., Goodpaster, B. H., Cawthon, P. M., Hansen, R. D., and Manini, T. M. (2018). Comment on: "Pitfalls in the measurement of muscle mass: a need for a reference standard" by Buckinx et al. *J. Cachexia Sarcopenia Muscle* 9, 1269–1271. doi: 10.1002/jcsm.12372

Coutinho, A. B. B., Jotta, B., Werneck-de-Castro, J. P., Pino, A. V., and Souza, M. N. (2020). Invasive electrical impedance myography at different levels of contraction of gastrocnemius muscle of rat. *Rev. Sci. Instruments* 91:084103. doi: 10.1063/1.5131631

Czeck, M. A., Raymond-Pope, C. J., Prescott, E., Bisch, K. L., and Dengel, D. R. (2020). Body fat percent assessment between electrical impedance myography and dual X-ray absorptiometry. *Am. J. Hum. Biol.* 32:e23330. doi: 10.1002/ajhb.23330

de Cardoner, M. M. M., Kwon, H., Pulido, H. V. G., Nagy, J., Rutkove, S., and Sanchez, B. (2021). Modeling and reproducibility of twin concentric electrical impedance myography. *IEEE Trans. Biomed. Eng.* 68, 3068–3077. doi: 10.1109/TBME. 2021.3063724

de Carvalho, M., and Swash, M. (2016). Lower motor neuron dysfunction in ALS. Clin. Neurophysiol. 127, 2670–2681. doi: 10.1016/j.clinph.2016.03.024

Duchateau, J., and Enoka, R. M. (2016). Neural control of lengthening contractions. J. Exp. Biol. 219, 197–204. doi: 10.1242/jeb.123158

Esper, G. J., Shiffman, C. A., Aaron, R., Lee, K. S., and Rutkove, S. B. (2006). Assessing neuromuscular disease with multifrequency electrical impedance myography. *Muscle Nerve* 34, 595–602. doi: 10.1002/mus.20626

Faes, T. J. C., van der Meij, H. A., de Munck, J. C., and Heethaar, R. M. (1999). The electric resistivity of human tissues (100 Hz-10 MHz): a meta-analysis of review studies. *Physiol. Meas.* 20, R1–R10. doi: 10.1088/0967-3334/20/4/201

Feigin, V. L., Brainin, M., Norrving, B., Martins, S., Sacco, R. L., Hacke, W., et al. (2022). World Stroke Organization (WSO): global stroke fact sheet 2022. *Int. J. Stroke* 17, 18–29. doi: 10.1177/17474930211065917

Feigin, V. L., Stark, B. A., Johnson, C. O., Roth, G. A., Bisignano, C., Abady, G. G., et al. (2021). Global, regional, and national burden of stroke and its risk factors, 1990– 2019: a systematic analysis for the Global Burden of Disease Study 2019. *Lancet Neurol*. 20, 795–820. doi: 10.1016/S1474-4422(21)00252-0

Foster, K. R., and Schwan, H. P. (1989). Dielectric properties of tissues and biological materials: a critical review. *Crit. Rev. Biomed. Eng.* 17, 25–104.

Fu, B., and Freeborn, T. J. (2020). Cole-impedance parameters representing biceps tissue bioimpedance in healthy adults and their alterations following eccentric exercise. J. Adv. Res. 25, 285–293. doi: 10.1016/j.jare.2020.05.016

Gao, W., Reiser, P. J., Coss, C. C., Phelps, M. A., Kearbey, J. D., Miller, D. D., et al. (2005). Selective androgen receptor modulator treatment improves muscle strength and body composition and prevents bone loss in orchidectomized rats. *Endocrinology* 146, 4887–4897. doi: 10.1210/en.2005-0572

Garmirian, L. P., Chin, A. B., and Rutkove, S. B. (2009). Discriminating neurogenic from myopathic disease via measurement of muscle anisotropy. *Muscle Nerve* 39, 16–24. doi: 10.1002/mus.21115

Goodpaster, B. H., Carlson, C. L., Visser, M., Kelley, D. E., Scherzinger, A., Harris, T. B., et al. (2001). Attenuation of skeletal muscle and strength in the elderly: the health ABC study. *J. Appl. Physiol.* 90, 2157–2165. doi: 10.1152/jappl.2001.90.6.2157

Hamel, J., Lee, P., Glenn, M. D., Burka, T., Choi, I., Friedman, S. D., et al. (2020). Magnetic resonance imaging correlates with electrical impedance myography in facioscapulohumeral muscular dystrophy. *Muscle Nerve* 61, 644–649. doi: 10.1002/ mus.26792

Herzog, W., Powers, K., Johnston, K., and Duvall, M. (2015). A new paradigm for muscle contraction. *Front. Physiol.* 6:174. doi: 10.3389/fphys.2015.00174

Heymsfield, S. B., Adamek, M., Gonzalez, M. C., Jia, G., and Thomas, D. M. (2014). Assessing skeletal muscle mass: historical overview and state of the art. J. Cachexia Sarcopenia Muscle 5, 9–18. doi: 10.1007/s13539-014-0130-5 Hobson-Webb, L., Zwelling, P., Pifer, A., Killelea, C., Faherty, M., Sell, T., et al. (2018). Point of care quantitative assessment of muscle health in older individuals: an investigation of quantitative muscle ultrasound and electrical impedance myography techniques. *Geriatrics* 3:92. doi: 10.3390/geriatrics3040092

Hobson-Webb, L. D., Zwelling, P. J., Raja, S. S., Pifer, A. N., and Kishnani, P. S. (2021). Quantitative muscle ultrasound and electrical impedance myography in late onset pompe disease: a pilot study of reliability, longitudinal change and correlation with function. *Mol. Genet. Metab. Rep.* 28:100785. doi: 10.1016/j.ymgmr.2021.100785

Hu, C., Hu, H., Mai, X., Lo, W. L. A., and Li, L. (2019). Correlation between muscle structures and electrical properties of the tibialis anterior in subacute stroke survivors: a pilot study. *Front. Neurosci.* 13:1270. doi: 10.3389/fnins.2019.01270

Hu, C., Wang, T., Leung, K. W. C., Li, L., and Tong, R. K.-Y. (2021a). Muscle electrical impedance properties and activation alteration after functional electrical stimulation-assisted cycling training for chronic stroke survivors: a longitudinal pilot study. *Front. Neurol.* 12:746263. doi: 10.3389/fneur.2021.746263

Hu, H., Chen, Y., Wang, X., Lo, W. L. A., and Li, L. (2021b). Quantifying the changes of mechanical and electrical properties of paralyzed muscle in survivors with cervical spinal cord injury. *Front. Neurol.* 12:720901. doi: 10.3389/fneur.2021.720901

Hu, H., Lo, W., Wang, X., Li, L., and Zhou, P. (2021c). Interrater and intrarater reliability of electrical impedance myography: a comparison between large and small handheld electrode arrays. *J. Healthc. Eng.* 2021, 7296322. doi: 10.1155/2021/7296322

Jafarpoor, M., Jia, L., White, J. K., and Rutkove, S. B. (2013). Optimizing electrode configuration for electrical impedance measurements of muscle via the finite element method. *IEEE Trans. Biomed. Eng.* 60, 1446–1452. doi: 10.1109/TBME.2012.2237030

Jafarpoor, M., Spieker, A. J., Jia, L., Minhee, S., Darras, B. T., et al. (2011). "Assessing electrical impedance alterations in spinal muscular atrophy via the finite element method," in *Proceedings of the 2011 Annual International Conference of the IEEE Engineering in Medicine and Biology Society*, (Piscataway, NJ: IEEE), 1871–1874. doi: 10.1109/IEMBS.2011.6090531

Johnson, R. W., Ng, K. W. P., Dietz, A. R., Hartman, M. E., Baty, J. D., Hasan, N., et al. (2018). Muscle atrophy in mechanically-ventilated critically ill children. *PLoS One* 13:e0207720. doi: 10.1371/journal.pone.0207720

Kamper, D. G., Harvey, R. L., Suresh, S., and Rymer, W. Z. (2003). Relative contributions of neural mechanisms versus muscle mechanics in promoting finger extension deficits following stroke. *Muscle Nerve* 28, 309–318. doi: 10.1002/mus.10443

Kapur, K., Nagy, J. A., Taylor, R. S., Sanchez, B., and Rutkove, S. B. (2018a). Estimating myofiber size with electrical impedance myography: a study in amyotrophic lateral sclerosis MICE. *Muscle Nerve* 58, 713–717. doi: 10.1002/mus. 26187

Kapur, K., Taylor, R. S., Qi, K., Nagy, J. A., Li, J., Sanchez, B., et al. (2018b). Predicting myofiber size with electrical impedance myography: a study in immature mice. *Muscle Nerve* 58, 106–113. doi: 10.1002/mus.26111

Kellis, E., Sahinis, C., Dafkou, K., Ellinoudis, A., and Galanis, N. (2021). Hamstring to quadriceps strength ratio and cross-sectional area of the quadriceps and hamstrings muscles assessed using extended field-of-view ultrasonography. *Res. Sports Med.* 29, 25–42. doi: 10.1080/15438627.2020.1770250

Kolb, S. J., Coffey, C. S., Yankey, J. W., Krosschell, K., Arnold, W. D., Rutkove, S. B., et al. (2016). Baseline results of the Neuro NEXT spinal muscular atrophy infant biomarker study. *Ann. Clin. Transl. Neurol.* 3, 132–145. doi: 10.1002/acn3.283

Lee, H.-M. (2002). Quantitative analysis of the velocity related pathophysiology of spasticity and rigidity in the elbow flexors. J. Neurol. Neurosurg. Psychiatry 72, 621-629. doi: 10.1136/jnnp.72.5.621

Leng, Y., Lo, W. L. A., Hu, C., Bian, R., Xu, Z., Shan, X., et al. (2021). The effects of extracorporeal shock wave therapy on spastic muscle of the wrist joint in stroke survivors: evidence from neuromechanical analysis. *Front. Neurosci.* 14:580762. doi: 10.3389/fnins.2020.580762

Li, J., Geisbush, T. R., Arnold, W. D., Rosen, G. D., Zaworski, P. G., and Rutkove, S. B. (2014). A comparison of three electrophysiological methods for the assessment of disease status in a mild spinal muscular atrophy mouse model. *PLoS One* 9:e111428. doi: 10.1371/journal.pone.0111428

Li, J., Pacheck, A., Sanchez, B., and Rutkove, S. B. (2016a). Single and modeled multifrequency electrical impedance myography parameters and their relationship to force production in the ALS SOD1G93A mouse. *Amyotroph Lateral Scler Frontotemporal. Degener.* 17, 397–403. doi: 10.3109/21678421.2016.116 5258

Li, L., Li, X., Hu, H., Shin, H., and Zhou, P. (2016b). The effect of subcutaneous fat on electrical impedance myography: electrode configuration and multi-frequency analyses. *PLoS One* 11:e0156154. doi: 10.1371/journal.pone.015 6154

Li, L., Shin, H., Li, X., Li, S., and Zhou, P. (2016c). Localized electrical impedance myography of the biceps brachii muscle during different levels of isometric contraction and fatigue. *Sensors* 16:581. doi: 10.3390/s16040581

Li, X., Fisher, M., Rymer, W. Z., and Zhou, P. (2016d). Application of the F-Response for estimating motor unit number and amplitude distribution in hand muscles of stroke survivors. *IEEE Trans. Neural Syst. Rehabil. Eng.* 24, 674–681. doi: 10.1109/TNSRE.2015.2453274

Li, J., Staats, W. L., Spieker, A., Sung, M., and Rutkove, S. B. (2012). A technique for performing electrical impedance myography in the mouse hind limb: data in normal and ALS SOD1 G93A animals. *PLoS One* 7:e45004. doi: 10.1371/journal.pone.0045004

Li, J., Sung, M., and Rutkove, S. B. (2013). Electrophysiologic biomarkers for assessing disease progression and the effect of riluzole in SOD1 G93A ALS mice. *PLoS One* 8:e65976. doi: 10.1371/journal.pone.0065976

Li, L., Hu, C., Leung, K. W. C., and Tong, R. K. Y. (2022). Immediate effects of functional electrical stimulation-assisted cycling on the paretic muscles of patients with hemiparesis after stroke: evidence from electrical impedance myography. *Front. Aging Neurosci.* 14:880221. doi: 10.3389/fnagi.2022.880221

Li, L., Shin, H., Stampas, A., Li, X., and Zhou, P. (2017a). Electrical impedance myography changes after incomplete cervical spinal cord injury: an examination of hand muscles. *Clin. Neurophysiol.* 128, 2242–2247. doi: 10.1016/j.clinph.2017.08.027

Li, L., Stampas, A., Shin, H., Li, X., and Zhou, P. (2017b). Alterations in localized electrical impedance myography of biceps brachii muscles paralyzed by spinal cord injury. *Front. Neurol.* 8:253. doi: 10.3389/fneur.2017.00253

Li, X., Li, L., Shin, H., Li, S., and Zhou, P. (2017c). Electrical impedance myography for evaluating paretic muscle changes after stroke. *IEEE Trans. Neural Systems Rehabil. Eng.* 25, 2113–2121. doi: 10.1109/TNSRE.2017.2707403

Li, X., Shin, H., Li, L., Magat, E., Li, S., and Zhou, P. (2017d). Assessing the immediate impact of botulinum toxin injection on impedance of spastic muscle. *Med. Eng. Phys.* 43, 97–102. doi: 10.1016/j.medengphy.2017.01.018

Li, X., Shin, H., Zong, Y., Li, S., and Zhou, P. (2017e). Assessing muscle compliance in stroke with the Myotonometer. *Clin. Biomechan.* 50, 110–113. doi: 10.1016/j. clinbiomech.2017.10.013

Li, Z., Tian, D., Chen, L., Wang, X., Jiang, L., and Yu, Y. (2017f). Electrical impedance myography for discriminating traumatic peripheral nerve injury in the upper extremity. *Clin. Neurophysiol.* 128, 384–390. doi: 10.1016/j.clinph.2016.11.016

Li, L., Tong, K. Y., and Hu, X. (2007). The effect of poststroke impairments on brachialis muscle architecture as measured by ultrasound. *Arch. Phys. Med. Rehabil.* 88, 243–250. doi: 10.1016/j.apmr.2006.11.013

Lindberg, P. G., Gäverth, J., Islam, M., Fagergren, A., Borg, J., and Forssberg, H. (2011). Validation of a new biomechanical model to measure muscle tone in spastic muscles. *Neurorehabil. Neural Repair.* 25, 617–625. doi: 10.1177/1545968311403494

Lo, E. H. (2014). 2013 Thomas Willis award lecture: causation and collaboration for stroke research. *Stroke* 45, 305–308. doi: 10.1161/STROKEAHA.113.001269

Lo, W. L. A., Zhao, J. L., Li, L., Mao, Y. R., and Huang, D. F. (2017). Relative and absolute interrater reliabilities of a hand-held myotonometer to quantify mechanical muscle properties in patients with acute stroke in an inpatient ward. *Biomed. Res. Int.* 2017;4294028. doi: 10.1155/2017/4294028

Lo, W. L. A., and Li, L. (2020). "Quantitative evaluation," in *Intelligent Biomechatronics in Neurorehabilitation*, X. Hu (Amsterdam: Elsevier), 193–207. doi: 10.1016/B978-0-12-814942-3.00012-X

Longo, S., Coratella, G., Rampichini, S., Borrelli, M., Scurati, R., Limonta, E., et al. (2021). Local fat content and muscle quality measured by a new electrical impedance myography device: correlations with ultrasound variables. *Eur. J. Sport Sci.* 21, 388–399. doi: 10.1080/17461391.2020.1751306

Lora-Millan, J. S., Sanchez-Cuesta, F. J., Romero, J. P., Moreno, J. C., and Rocon, E. (2022). A unilateral robotic knee exoskeleton to assess the role of natural gait assistance in hemiparetic patients. *J. Neuroeng. Rehabil.* 19:109. doi: 10.1186/s12984-02 2-01088-2

Luo, X., Wang, S., and Terrones, B. S. (2022). Modeling and simulation of needle electrical impedance myography in nonhomogeneous isotropic skeletal muscle. *IEEE J. Electromagn. RF Microw. Med. Biol.* 6, 103–110. doi: 10.1109/JERM.2021.3091515

Luo, Z., Lo, W. L. A., Bian, R., Wong, S., and Li, L. (2020). Advanced quantitative estimation methods for spasticity: a literature review. *J. Int. Med. Res.* 48:030006051988842. doi: 10.1177/0300060519888425

Martinez-Gonzalez, M., Montilla-Herrador, J., García-Vidal, J. A., Escolar-Reina, P., Gacto-Sánchez, M., and Medina-Mirapeix, F. (2020). Intra- and inter-rater reliability of electrical impedance myography using adhesive electrodes in healthy volunteers. *J. Electromyogr. Kinesiol.* 55:102456. doi: 10.1016/j.jelekin.2020.102456

McLester, C. N., Dewitt, A. D., Rooks, R., and McLester, J. R. (2018). An investigation of the accuracy and reliability of body composition assessed with a handheld electrical impedance myography device. *Eur. J. Sport Sci.* 18, 763–771. doi: 10.1080/17461391. 2018.148458

Metoki, N., Sato, Y., Satoh, K., Okumura, K., and Iwamoto, J. (2003). Muscular atrophy in the hemiplegic thigh in patients after stroke. *Am. J. Phys. Med. Rehabil.* 82, 862–865. doi: 10.1097/01.PHM.0000091988.20916.EF

Musampa, N. K., Mathieu, P. A., and Levin, M. F. (2007). Relationship between stretch reflex thresholds and voluntary arm muscle activation in patients with spasticity. *Exp. Brain Res.* 181, 579–593. doi: 10.1007/s00221-007-0956-6

Ngo, C., Munoz, C., Lueken, M., Hülkenberg, A., Bollheimer, C., Briko, A., et al. (2022). A wearable, multi-frequency device to measure muscle activity

combining simultaneous electromyography and electrical impedance myography. Sensors 22:1941. doi: 10.3390/s22051941

Pacheck, A., Mijailovic, A., Yim, S., Li, J., Green, J. R., McIlduff, C. E., et al. (2016). Tongue electrical impedance in amyotrophic lateral sclerosis modeled using the finite element method. *Clin. Neurophysiol.* 127, 1886–1890. doi: 10.1016/j.clinph.20 15.11.046

Pandeya, S. R., Nagy, J. A., Riveros, D., Semple, C., Taylor, R. S., Hu, A., et al. (2022). Using machine learning algorithms to enhance the diagnostic performance of electrical impedance myography. *Muscle Nerve* 66, 354–361. doi: 10.1002/mus.27664

Pandeya, S. R., Nagy, J. A., Riveros, D., Semple, C., Taylor, R. S., Mortreux, M., et al. (2021a). Estimating myofiber cross-sectional area and connective tissue deposition with electrical impedance myography: a study in D2 - mdx mice. *Muscle Nerve* 63, 941–950. doi: 10.1002/mus.27240

Pandeya, S. R., Nagy, J. A., Riveros, D., Semple, C., Taylor, R. S., Mortreux, M., et al. (2021b). Predicting myofiber cross-sectional area and triglyceride content with electrical impedance myography: a study in db/db mice. *Muscle Nerve* 63, 127–140. doi: 10.1002/mus.27095

Patten, C., Lexell, J., and Brown, H. E. (2004). Weakness and strength training in persons with poststroke hemiplegia: rationale, method, and efficacy. *J. Rehabil. Res. Dev.* 41, 293–312. doi: 10.1682/jrrd.2004.03.0293

Petterson, S. C., Barrance, P., Buchanan, T., Binder-Macleod, S., and Snyder-Mackler, L. (2008). Mechanisms underlying quadriceps weakness in knee osteoarthritis. *Med. Sci. Sports Exerc.* 40, 422–427. doi: 10.1249/MSS. 0b013e31815ef285

Power, G. A., Dalton, B. H., and Rice, C. L. (2013). Human neuromuscular structure and function in old age: a brief review. *J. Sport Health Sci.* 2, 215–226. doi: 10.1016/j. jshs.2013.07.001

Roy, B., Darras, B. T., Zaidman, C. M., Wu, J. S., Kapur, K., and Rutkove, S. B. (2019). Exploring the relationship between electrical impedance myography and quantitative ultrasound parameters in Duchenne muscular dystrophy. *Clin. Neurophysiol.* 130, 515–520. doi: 10.1016/j.clinph.2019.01.018

Rutkove, S. B. (2009). Electrical impedance myography: background, current state, and future directions. *Muscle Nerve* 40, 936–946. doi: 10.1002/mus.21362

Rutkove, S. B., Caress, J. B., Cartwright, M. S., Burns, T. M., Warder, J., David, W. S., et al. (2014a). Electrical impedance myography correlates with standard measures of ALS severity. *Muscle Nerve* 49, 441–443. doi: 10.1002/mus.24128

Rutkove, S. B., Geisbush, T. R., Mijailovic, A., Shklyar, I., Pasternak, A., Visyak, N., et al. (2014b). Cross-sectional evaluation of electrical impedance myography and quantitative ultrasound for the assessment of duchenne muscular dystrophy in a clinical trial setting. *Pediatr. Neurol.* 51, 88–92. doi: 10.1016/j.pediatrneurol.20 14.02.015

Rutkove, S. B., and Darras, B. T. (2013). Electrical impedance myography for the assessment of children with muscular dystrophy: a preliminary study. *J. Phys. Conf. Ser.* 434:012069. doi: 10.1088/1742-6596/434/1/012069

Rutkove, S. B., Narayanaswami, P., David Arnold, W., Kolb, S. J., Clark, B. C., Darras, B. T., et al. (2021). Putting the patient first: the validity and value of surface-based electrical impedance myography techniques. *Clin. Neurophysiol.* 132, 1752–1753. doi: 10.1016/j.clinph.2021.03.020

Rutkove, S. B., Pacheck, A., and Sanchez, B. (2017). Sensitivity distribution simulations of surface electrode configurations for electrical impedance myography. *Muscle Nerve* 56, 887–895. doi: 10.1002/mus.25561

Rutkove, S. B., and Sanchez, B. (2019). Electrical impedance methods in neuromuscular assessment: an overview. *Cold Spring Harb. Perspect. Med.* 9:a034405. doi: 10.1101/cshperspect.a034405

Rutkove, S. B., Shefner, J. M., Gregas, M., Butler, H., Caracciolo, J., Lin, C., et al. (2010). Characterizing spinal muscular atrophy with electrical impedance myography. *Muscle Nerve* 42, 915–921. doi: 10.1002/mus.21784

Sanchez, B., Martinsen, O. G., Freeborn, T. J., and Furse, C. M. (2021). Electrical impedance myography: a critical review and outlook. *Clin. Neurophysiol.* 132, 338–344. doi: 10.1016/j.clinph.2020.11.014

Scherbakov, N., Sandek, A., and Doehner, W. (2015). Stroke-Related sarcopenia: specific characteristics. J. Am. Med. Dir. Assoc. 16, 272–276. doi: 10.1016/j.jamda.2014. 12.007

Scherbakov, N., von Haehling, S., Anker, S. D., Dirnagl, U., and Doehner, W. (2013). Stroke induced sarcopenia: muscle wasting and disability after stroke. *Int. J. Cardiol.* 170, 89–94. doi: 10.1016/j.ijcard.2013.10.031

Schooling, C. N., Jamie Healey, T., McDonough, H. E., French, S. J., McDermott, C. J., Shaw, P. J., et al. (2020). Modelling and analysis of electrical impedance myography of the lateral tongue. *Physiol. Meas.* 41:125008. doi: 10.1088/1361-6579/ abcb9b

Schrunder, A. F., Rodriguez, S., and Rusu, A. (2022). A finite element analysis and circuit modelling methodology for studying electrical impedance myography of human limbs. *IEEE Trans. Biomed. Eng.* 69, 244–255. doi: 10.1109/TBME.2021. 3091884 Schuermans, J., van Tiggelen, D., Danneels, L., and Witvrouw, E. (2014). Biceps femoris and semitendinosus-teammates or competitors? new insights into hamstring injury mechanisms in male football players: a muscle functional MRI study. *Br. J. Sports Med.* 48, 1599–1606. doi: 10.1136/bjsports-2014-094017

Shellikeri, S., Yunusova, Y., Green, J. R., Pattee, G. L., Berry, J. D., Rutkove, S. B., et al. (2015). Electrical impedance myography in the evaluation of the tongue musculature in amyotrophic lateral sclerosis. *Muscle Nerve* 52, 584–591. doi: 10.1002/mus. 24565

Shiffman, C. A., and Rutkove, S. B. (2013). Circuit modeling of the electrical impedance: I. neuromuscular disease. *Physiol. Meas.* 34, 203–221. doi: 10.1088/0967-3334/34/2/203

Shklyar, I., Pasternak, A., Kapur, K., Darras, B. T., and Rutkove, S. B. (2015). Composite biomarkers for assessing duchenne muscular dystrophy: an initial assessment. *Pediatr. Neurol.* 52, 202–205. doi: 10.1016/j.pediatrneurol.2014.09.014

Sions, J. M., Tyrell, C. M., Knarr, B. A., Jancosko, A., and Binder-Macleod, S. A. (2012). Age- and stroke-related skeletal muscle changes. *J. Geriatric Phys. Therapy* 35, 155–161. doi: 10.1519/JPT.0b013e318236db92

Srivastava, T., Darras, B. T., Wu, J. S., and Rutkove, S. B. (2012). Machine learning algorithms to classify spinal muscular atrophy subtypes. *Neurology* 79, 358–364. doi: 10.1212/WNL.0b013e3182604395

Sung, M., Spieker, A. J., Narayanaswami, P., and Rutkove, S. B. (2013). The effect of subcutaneous fat on electrical impedance myography when using a handheld electrode array: the case for measuring reactance. *Clin. Neurophysiol.* 124, 400–404. doi: 10.1016/j.clinph.2012.07.013

Tavoian, D., Ampomah, K., Amano, S., Law, T. D., and Clark, B. C. (2019). Changes in DXA-derived lean mass and MRI-derived cross-sectional area of the thigh are modestly associated. *Sci. Rep.* 9:10028. doi: 10.1038/s41598-019-46428-w

Tsai, L.-C., Wu, Y.-N., Liu, S. Q., and Zhang, L.-Q. (2020). Changes in muscle stress and sarcomere adaptation in mice following ischemic stroke. *Front. Physiol.* 11:581846. doi: 10.3389/fphys.2020.581846

van Rassel, C. R., Bewski, N. A., O'loughlin, E. K., Wright, A., Scheel, D. P., Puig, L., et al. (2019). Validity of electrical impedance myography to estimate percent body fat: comparison to bio-electrical impedance and dual-energy X-ray absorptiometry. *J. Sports Med. Phys. Fitness* 59, 632–639. doi: 10.23736/S0022-4707.18.08505-5

Wang, L. L., Ahad, M., McEwan, A., Jia, Li, Jafarpoor, M., et al. (2011). Assessment of alterations in the electrical impedance of muscle after experimental nerve injury via finite-element analysis. *IEEE Trans. Biomed. Eng.* 58, 1585–1591. doi: 10.1109/TBME. 2011.2104957

Wood, D., Burridge, J., van Wijck, F., McFadden, C., Hitchcock, R., Pandyan, A., et al. (2005). Biomechanical approaches applied to the lower and upper limb for the measurement of spasticity: a systematic review of the literature. *Disabil. Rehabil.* 27, 19–33. doi: 10.1080/09638280400014683

Zaidman, C. M., Wang, L. L., Connolly, A. M., Florence, J., Wong, B. L., Parsons, J. A., et al. (2015). Electrical impedance myography in duchenne muscular dystrophy and healthy controls: a multicenter study of reliability and validity. *Muscle Nerve* 52, 592–597. doi: 10.1002/mus.24611

Zong, Y., Shin, H. H., Wang, Y.-C., Li, S., Zhou, P., and Li, X. (2018). Assessing hand muscle structural modifications in chronic stroke. *Front. Neurol.* 9:296. doi: 10.3389/fneur.2018.00296