



Liver Dysfunction as a Novel Player in Alzheimer's Progression: Looking Outside the Brain

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Alzheimer's disease (AD) afflicts an estimated 20 million people worldwide and is the fourth-leading cause of death in the developed world. The most common cause of dementia in older individuals, AD is characterized by neuropathologies including synaptic and neuronal degeneration, amyloid plaques, and neurofibrillary tangles (NTFs). Amyloid plaques are primarily composed of amyloid-beta peptide (A β), which accumulates in the brains of patients with AD. Further, small aggregates termed A β oligomers are implicated in the synaptic loss and neuronal degeneration underlying early cognitive impairments. Whether A β accumulates in part because of dysregulated clearance from the brain remains unclear. The flow of substances (e.g., nutrients, drugs, toxins) in and out of the brain is mediated by the blood-brain-barrier (BBB). The BBB exhibits impairment in AD patients and animal models. The effect of BBB impairment on A β , and whether BBB function is affected by non-neurological pathologies that impair peripheral clearance requires further investigation. In particular, impaired peripheral clearance is a feature of nonalcoholic fatty liver disease (NAFLD), a spectrum of liver disorders characterized by accumulation of fat in the liver accompanied by varying degrees of inflammation and hepatocyte injury. NAFLD has reached epidemic proportions, with an estimated prevalence between 20% and 30% of the general population. This chronic condition may influence AD pathogenesis. This review article summarizes the current state of the literature linking NAFLD and AD, highlighting the role of the major A β efflux and clearance protein, the LRP-1 receptor, which is abundantly expressed in liver, brain, and vasculature.

Keywords: amyloid beta, NAFLD, LRP-1, BBB, Alzheimer's

AMYLOID BETA ROLE IN ALZHEIMER'S DISEASE

Alzheimer's disease (AD) belongs to a large group of neurodegenerative diseases characterized by the pathophysiological brain changes related to the accumulation of misfolded proteins. Specifically, extracellular peptide variants of the amyloid- β (A β) accumulate in the form of amyloid plaques or senile plaques, and the intracellular accumulation

of neurofibrillary tangles (NTFs) composed by phosphorylated Tau protein (pTau; Bloom, 2014; Héraud et al., 2014; He et al., 2018).

Both are reported to underlie progressive synaptic dysfunction in the AD brain, loss of dendritic spines, and neuronal death (Serrano-Pozo et al., 2011; Busche et al., 2019). Although AD was first described 100 years ago, its precise etiology remains unknown. Efforts to better understand AD have resulted in multiple hypotheses to explain events in AD pathogenesis, for example, the amyloid cascade theory that describes the imbalance between A β production and clearance (Selkoe and Hardy, 2016). Here, we provide an overview of the etiology of AD, and the principal concepts that support the critical role of the brain-blood barrier (BBB) and liver in AD development and progression.

In neurons under physiological conditions, A β is secreted to maintain normal synaptic function, morphology, and plasticity (Wang et al., 2012; Gouras et al., 2015; Klevanski et al., 2015). A β is a by-product generated from cleavage of the amyloid protein precursor (APP). APP plays an important physiological role in regulating γ -aminobutyric acid type B receptor (GABA_BR) and modulating synaptic transmission and plasticity (Chen et al., 2017; Doshina et al., 2017; Rice et al., 2019). In primary cortical neurons, APP modulates frequency and amplitude of calcium oscillations essential for synaptic transmission (Octave et al., 2013). A mouse model deficient for APP demonstrated that APP is necessary for the synapsis and maintenance of dendritic integrity in the hippocampus (Tyan et al., 2012). Likewise, hippocampal neurons in culture derived from APP knockout mice showed APP is critical for synaptogenesis and dendritic and axonal growth process and regulates substrate adhesion (Southam et al., 2019).

On the other hand, in the amyloidogenic (i.e., disease-causing) pathway, APP is cleaved by β - and γ -secretase to generate A β , which accumulates as senile plaques (Hardy and Selkoe, 2002; Konietzko, 2011). AD-related plaques are associated with high levels of soluble oligomeric forms of A β (A β Os; Esparza et al., 2013). A β Os comprise soluble dimers and trimers of low molecular weight and soluble oligomeric forms of 12–14 monomers (Mroczko et al., 2018). In addition, these oligomers have been identified as the toxic conformers of A β plaques in AD (Jin et al., 2011; Verma et al., 2015). A β Os can diffuse across synaptic membranes (Hong et al., 2014) and trigger a cascade of injurious events in neurons, causing synaptic failure and memory loss (Morris et al., 2014; Brito-Moreira et al., 2017). Moreover, A β Os are associated with dystrophic neurites, reactive astrocytes, and aberrant activation of glutamatergic neurotransmission; the consequence of these changes is neuronal death by excessive neuronal influx of sodium and calcium (Ziegler-Waldkirch and Meyer-Luehmann, 2018). Postsynaptic protein disruption (Lésne et al., 2013) and hippocampal synaptic plasticity impairment by A β Os contributes to memory loss (Müller-Schiffmann et al., 2016). Intracellular A β Os are detectable in cholinergic neurons, suggesting that they play a critical role in cholinergic deficiency (Baker-Nigh et al., 2015). These devastating events not only lead to memory loss and learning impairment in AD patients, but also

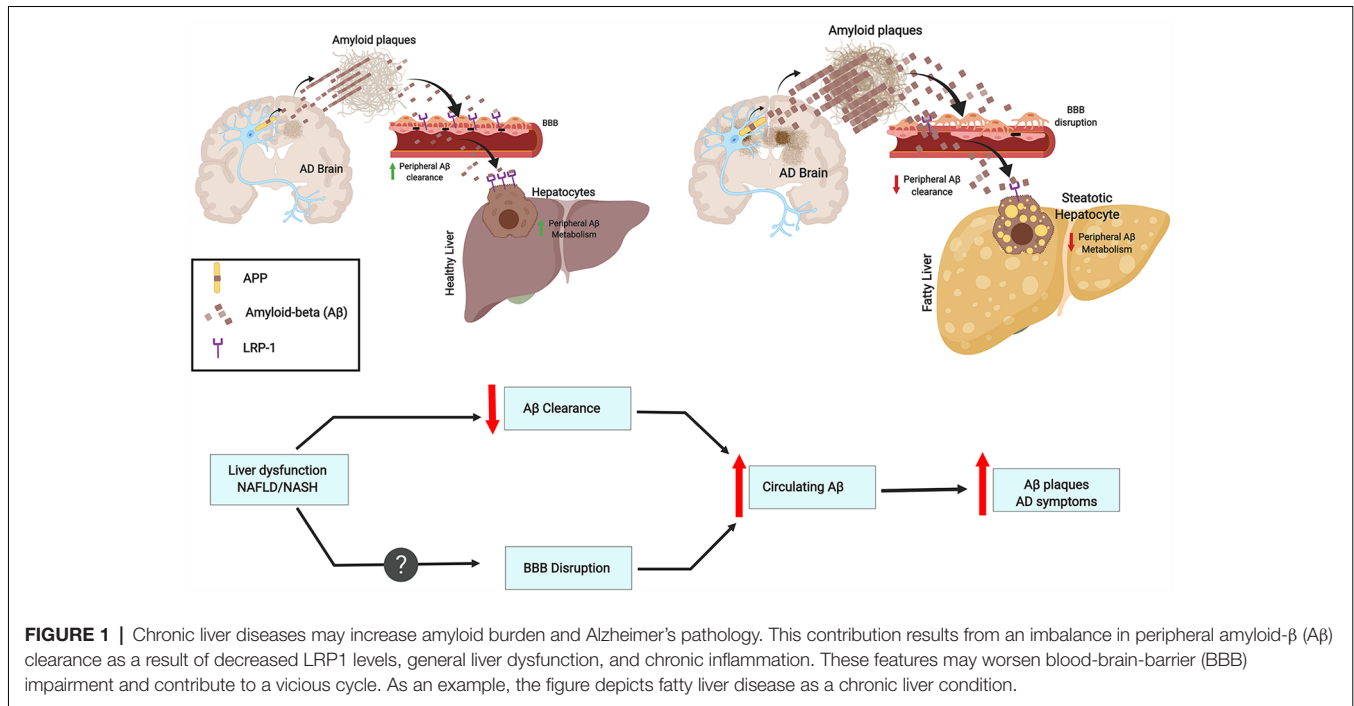
affect the capacities of reasoning, abstraction, and language (Duyckaerts et al., 2009).

BLOOD-BRAIN BARRIER BREAKDOWN AND ROLE OF LRP-1 IN ALZHEIMER'S DISEASE

The blood-brain barrier (BBB) is a specialized structure that supports brain function. This structure supports the brain by regulating electrolyte flux, cerebral blood flow (CBF) and efficient oxygen and metabolite delivery, and restricting entry of potentially toxic and even some therapeutic agents into the brain (Provias and Jeynes, 2014; Andreone et al., 2015; Di Marco et al., 2015). BBB function is mediated by neurovascular units (NVU) comprising neurons, glial cells, pericytes, and brain endothelial cells, which maintain homeostasis of the cerebral microenvironment (Armulik et al., 2011). Brain endothelial cells are an important component mediating the flow between brain and blood by cell-to-cell communications called tight junctions and adherent junctions; these junctions connect cell networks (Deli et al., 2005; Van de Haar et al., 2015) and regulate the paracellular permeability of substances across the BBB (Bowman and Quinn, 2008; Viggars et al., 2011; Kook et al., 2013; Chow and Gu, 2015; Ulrich et al., 2015). Tight junction proteins ZO-1, Occludin and CLN-5 are key to maintaining BBB integrity (Jiao et al., 2011). ZO-1 joins tight junctions with the actin cytoskeleton, working as accessory proteins (Xiao et al., 2017). Occludin and CLN-5 are transmembrane tight junction proteins involved in signal transduction of cytokines (Haseloff et al., 2015). The high expression of these proteins on brain endothelial cells regulates the transport of essential molecules through the BBB, such as the free and rapid diffusion of oxygen and carbon dioxide (Lin et al., 2015; Pardridge, 2015). Hydrophobic molecules permeate the BBB faster and more easily than hydrophilic molecules, while molecules that are larger than 180 kDa or water-soluble do not penetrate the BBB (Kroll and Neuwelt, 1998; Zlokovic, 2005; Masserini, 2013). For example, the BBB restricts the passage of albumin and immunoglobulins, high-molecular-weight proteins from the peripheral blood circulation (Xiao and Gan, 2013).

Another important component of brain endothelial cells is a complex and specific transport-receptor protein system that also contributes to BBB permeability (Zlokovic, 2011). The luminal side of the BBB contains transporters for specific classes of nutrients, such as glucose and vitamins, and receptors for peptides, proteins, and hormones. These mediators facilitate transport across the BBB from circulating blood into the brain (Deane and Zlokovic, 2007; Simpson et al., 2007). In contrast, the transport system of the abluminal side of the BBB eliminates neurotoxic molecules and metabolic waste (Begley and Brightman, 2003).

Dysfunction of the BBB, therefore, could result in altered permeability. Indeed, age-dependent BBB breakdown at the hippocampus is associated with mild cognitive impairment and correlates with pericytes injury. This finding suggests that the cerebrovascular integrity loss that begins at the hippocampus



may contribute to early stages of dementia associated with AD (Montagne et al., 2015). Similarly, early cognitive dysfunction has been associated with capillary damage and BBB breakdown in older adults (Nation et al., 2019).

This breakdown of BBB function may be related to alterations in specific components of the BBB structure. Low-density lipoprotein receptor-related protein 1 (LRP-1) is a membrane receptor that mediates the cellular internalization of multiple ligands. Further, LRP-1 regulates several tight junction proteins in endothelial cells of the BBB (Zhao et al., 2016). Functional LRP-1 is expressed in liver sinusoidal endothelial cells (LECs), highly specialized scavenger cells, and LRP-1 expression contributes to the rapid removal of its blood ligands (Öie et al., 2011). Cell surface LRP-1 and circulating sLRP-1 are needed for brain and systemic clearance of A β ; however, in AD, both cell surface LRP-1 and circulating sLRP-1 concentrations are dramatically reduced (Sagare et al., 2012). Importantly, these alterations may begin as early as two decades before the manifestation of cognitive impairment symptoms (Beason-Held et al., 2013; Jack et al., 2013; De Strooper, 2014). Clearance of A β may also be affected by other pathologies, however.

CLEARANCE OF A β AT THE PERIPHERY: ROLE OF THE LIVER

Peripheral organs, including the kidney and the liver, play an essential role in the clearance of circulating A β . Elimination of A β from the circulation may contribute to AD progression, by helping to displace the dynamic equilibrium from A β deposited in the senile plaques toward soluble A β . This hypothesis is supported by evidence that peritoneal dialysis reduces the circulating levels of A β in humans and diminishes AD features

in an animal model (Jin et al., 2017). Insufficient clearance of brain A β also contributes to the progression of sporadic AD (Wang et al., 2006). As brain A β equilibrates with A β in plasma, peripheral clearance of A β provides a potential approach to facilitate efflux of A β from the brain (Liu et al., 2015). Peripheral organs and tissues are key in clearing brain-derived A β under physiological conditions (Xiang et al., 2015).

The liver has many functions, one of which is metabolic detoxification. When the liver is under constant injury, as is found in metabolic diseases, it exhibits decreased detoxification capacity. Indeed, the expression of metabolic enzymes decreases in conditions such as obesity, diabetes, and cirrhosis (Rolle et al., 2018). Hepatocytes can act directly on circulating A β , promoting its clearance by degradation or through bile excretion. Further, A β uptake from circulation can be mediated through LRP-1, which is highly expressed in hepatocytes (Kanekiyo and Bu, 2014). Interestingly, liver dysfunction is accompanied by low LRP-1 hepatic expression and high levels of circulating A β . This correlation suggests that A β clearance decreases due to low hepatic LRP-1 activity (Wang et al., 2017; see **Figure 1**).

AD pathophysiology has not been evaluated from a hepatic point of view; yet, the evidence points to a critical role for liver in AD pathogenesis. A β levels found in liver samples from AD patients are lower when compared to neurologically healthy controls, raising the possibility that the liver is not properly eliminating circulating A β (Roher et al., 2009). This observation is supported by studies where insulin promotes LRP-1 translocation to the cell membrane in hepatocytes, favoring A β clearance (Tamaki et al., 2007). The stimulation of LRP-1-mediated liver uptake improves cognitive impairment and decreases A β aggregation in the brain in AD transgenic mice (Sehgal et al., 2012).

NAFLD/NASH AFFECTS A β CLEARANCE

Non-alcoholic fatty liver disease (NAFLD) encompasses a spectrum of liver disorders characterized by excessive fat deposition in hepatocytes from individuals who drink little or no alcohol. NAFLD is an umbrella term for several subtypes ranging from isolated hepatic steatosis, or fatty liver, to nonalcoholic steatohepatitis (NASH). NASH is defined by the presence of fatty changes with inflammation and several degrees of hepatocellular injury or fibrosis. Thus, NASH is the aggressive form of NAFLD and can progress to advanced fibrosis and cirrhosis.

NAFLD/NASH is the leading cause of chronic liver disease worldwide and has reached epidemic proportions. Interestingly, most of the deaths in NAFLD patients are not restricted to liver-related morbidity or mortality; rather, cardiovascular disease (CVD) and cancer predominate (Armstrong et al., 2014). Therefore, the presence of fatty liver is not a benign pathology as was historically considered by most clinicians. Indeed, extensive evidence in recent years shows that NAFLD also increases the risk of end-stage liver disease, hepatocellular carcinoma (HCC), liver-related mortality, and all-cause mortality. These observations prompted the idea that NAFLD/NASH, either independently or concomitantly with other metabolic risk factors, determines or even drives extra-hepatic diseases such as CVD, chronic kidney disease, colorectal cancer, endocrine disorders like type 2 diabetes mellitus, osteoporosis, and, indeed, AD. Recent studies have linked insulin-resistance (the key pathophysiological feature of NAFLD) to several of the neurodegenerative mechanisms of AD including oxidative stress, mitochondrial dysfunction, and inflammation, via dysregulated insulin/IGF-1 signaling with attendant impairments in signal transduction and gene expression (de la Monte and Tong, 2014; de la Monte, 2017; Kim et al., 2016).

A network clustering analysis conducted by Karbalaee et al. (2018) indicated that there are 189 genes shared between NAFLD and AD. Further, three main groups of pathways are candidates for contributing to both AD and NAFLD: carbohydrate metabolism, long fatty acid metabolism, and IL-17 signaling pathways (Karbalaee et al., 2018). This suggests that diabetes and obesity might be considered as a risk factor for AD and NAFLD.

One study showed that NAFLD promotes AD in mice (Kim et al., 2016). This study evaluated whether NAFLD induction, through a dietary approach (high-fat diet), promotes the development of AD signs. Brains of HFD-fed mice showed increased levels of neuro-inflammation, characterized by higher levels of cytokines, toll-like receptors, and microgliosis. These features were accompanied by increased plaque formation in a transgenic mouse model of AD. In addition, intense and frequent signs of cerebral amyloid angiopathy (CAA)—a condition characterized by the A β deposition in the media and adventitia of small and mid-sized arteries—were observed in mice fed with HFD.

An abnormal lipid metabolism is linked with increased risk for AD development, and the liver plays a crucial role since is the main peripheral organ responsible for lipid metabolism

(Fukumoto et al., 2002; Hooijmans and Kiliaan, 2008). A β is able to bind Apolipoprotein E (ApoE) and can be cleared from the brain together with cholesterol (Mahley, 1988). Interestingly, ApoE is a ligand of LRP-1 and both are genetically associated with AD and plasma A β levels (Kang et al., 2000). This link is intriguing since LRP-1 is suggested to facilitate A β clearance from the brain across the BBB (Deane et al., 2004; Sagare et al., 2012; see **Figure 1**).

LIVER INFLAMMATION AND A β LEVELS

Hepatitis B is a liver infection that can become chronic and severe. Interestingly, Hepatitis B Virus (HBV) carriers have significantly higher plasma A β levels than non-carriers. Moreover, HBV carrier status is associated with plasma A β levels (Jin et al., 2017). Overall infectious burden including cytomegalovirus (CMV), herpes simplex virus type 1 (HSV-1), *Borrelia burgdorferi*, *Chlamydia pneumoniae* and *Helicobacter pylori* was found to significantly contribute to AD pathogenesis (Bu et al., 2015). However, currently, no epidemiological study has been designed to understand the association between HBV infection and the risk for AD. The effect of chronic inflammation on A β clearance is lesser than the effects of HBV infection or liver dysfunction (Liu et al., 2013). Further, although plasma concentrations of cytokines IL-1 β and IL-6 are significantly increased in cirrhosis patients and plasma IL-6 levels are correlated with A β 40 levels (a 40 amino acid proteolytic product of APP cleavage that has gained attention as a biomarker correlating with AD), no association is observed by linear regression between IL-6 and A β 40 levels. On the other hand, the ratio of AST/ALT, which is an indicator of liver functional impairment (Giannini et al., 1999), is significantly associated with circulating A β 40 levels (Wang et al., 2017). Furthermore, hepatic dysfunction may lead to a plethora of systemic changes. Approximately 95% of A β in the blood is bound to serum albumin (Stanyon and Viles, 2012). The serum albumin pool represents an important reservoir for peripheral clearance of A β . Thus, a diminution in blood albumin in cirrhotic patients might contribute to the increase in plasma A β levels (see **Figure 1**).

CONCLUDING REMARKS

AD is a degenerative condition that will afflict an increasing number of people as the global population ages. Unfortunately, current treatments have only transient or modest effects. This article reviews evidence that supports the involvement of liver diseases, a growing health concern, in AD pathogenesis. The liver is the major player in the clearance of A β at the periphery, and an impairment of this clearance may shift the delicate A β equilibrium toward brain accumulation.

As to the possible role that the liver plays in brain-derived A β clearance, the impaired clearance of serum A β might contribute to the high A β levels in NAFLD patients. This effect is likely due to an intensification of the BBB disruption and drop in LRP-1

levels, the major receptor for A β efflux and important effector of clearance.

It is possible that hepatic malfunction contributes to AD in a plethora of non-excluding pathways, including: (i) the failure to maintain A β homeostasis at the periphery; (ii) acting as a source of pro-inflammatory cytokines when chronic inflammation follows different types of injury (like virus infection, drug-induced injury, and metabolic diseases); and (iii) through metabolic impairment.

AUTHOR CONTRIBUTIONS

LE wrote and edited the manuscript. PA participated in manuscript writing. DC wrote the manuscript and designed figures. JA participated in manuscript writing and editing.

REFERENCES

- Andreone, B. J., Lacoste, B., and Gu, C. (2015). Neuronal and vascular interactions. *Annu. Rev. Neurosci.* 38, 25–46. doi: 10.1146/annurev-neuro-071714-033835
- Armstrong, M. J., Adams, L. A., Canbay, A., and Syn, W.-K. (2014). Extrahepatic complications of nonalcoholic fatty liver disease. *Hepatology* 59, 1174–1197. doi: 10.1002/hep.26717
- Armulik, A., Genové, G., and Betsholtz, C. (2011). Pericytes: developmental, physiological and pathological perspectives, problems and promises. *Dev. Cell* 21, 193–215. doi: 10.1016/j.devcel.2011.07.001
- Baker-Nigh, A., Vahedi, S., Davis, E.-G., Weintraub, S., Bigio, E. H., Klein, W. L., et al. (2015). Neuronal amyloid- β accumulation within cholinergic basal forebrain in ageing and Alzheimer's disease. *Brain* 138, 1722–1737. doi: 10.1093/brain/awv024
- Beason-Held, L., Goh, J. O., An, Y., Kraut, M. A., O'Brien, R. J., Ferrucci, L., et al. (2013). Changes in brain function occurs years before the onset of cognitive impairment. *J. Neurosci.* 33, 18008–18014. doi: 10.1523/JNEUROSCI.1402-13.2013
- Begley, D. J., and Brightman, M. W. (2003). Structural and functional aspects of the blood-brain barrier. *Prog. Drug Res.* 61, 39–78. doi: 10.1007/978-3-0348-8049-7_2
- Bloom, G. S. (2014). Amyloid- β and tau: the trigger and bullet in Alzheimer disease pathogenesis. *JAMA Neurol.* 71, 505–508. doi: 10.1001/jamaneurol.2013.5847
- Bowman, G. L., and Quinn, J. F. (2008). Alzheimer's disease and the blood-brain barrier: past, present and future. *Aging Health* 4, 47–55. doi: 10.2217/1745509X.4.1.47
- Brito-Moreira, J., Lourenco, M. V., Oliveira, M. M., Ribeiro, F. C., Ledo, J. H., Diniz, L. P., et al. (2017). Interaction of amyloid- β (A β) oligomers with neurexin 2 α and neuroligin 1 mediates synapse damage and memory loss in mice. *J. Biol. Chem.* 292, 7327–7337. doi: 10.1074/jbc.M116.761189
- Bu, X.-L., Yao, X.-Q., Jiao, S.-S., Zeng, F., Liu, Y.-H., Xiang, Y., et al. (2015). A study on the association between infectious burden and Alzheimer's disease. *Eur. J. Neurol.* 22, 1519–1525. doi: 10.1111/ene.12477
- Busche, M. A., Wegmann, S., Dujardin, S., Commins, C., Schiantarelli, J., Klickstein, N., et al. (2019). Tau impairs neural circuits, dominating amyloid- β effects, in Alzheimer models *in vivo*. *Nat. Neurosci.* 22, 57–64. doi: 10.1038/s41593-018-0289-8
- Chen, M., Wang, J., Jiang, J., Zheng, X., Justice, N. J., Wang, K., et al. (2017). APP modulates KCC2 expression and function in hippocampal GABAergic inhibition. *eLife* 6:e20142. doi: 10.7554/eLife.20142
- Chow, B. W., and Gu, C. (2015). The molecular constituents of the blood-brain barrier. *Trends Neurosci.* 38, 598–608. doi: 10.1016/j.tins.2015.08.003
- Deane, R., Wu, Z., and Zlokovic, B. V. (2004). RAGE (Yin) versus LRP (Yang) balance regulates Alzheimer amyloid-peptide clearance through transport across the blood-brain barrier. *Stroke* 35, 2628–2631. doi: 10.1161/01.str.0000143452.85382.d1
- Deane, R., and Zlokovic, B. V. (2007). Role of the blood-brain barrier in the pathogenesis of Alzheimer's disease. *Curr. Alzheimer Res.* 4, 191–197. doi: 10.2174/156720507780362245
- de la Monte, S. M. (2017). Insulin resistance and neurodegeneration: progress towards the development of new therapeutics for Alzheimer's disease. *Drugs* 77, 47–65. doi: 10.1007/s40265-016-0674-0
- de la Monte, S. M., and Tong, M. (2014). Brain metabolic dysfunction at the core of Alzheimer's disease. *Biochem. Pharmacol.* 88, 548–559. doi: 10.1016/j.bcp.2013.12.012
- Deli, M. A., Abrahám, C. S., Kataoka, Y., and Niwa, M. (2005). Permeability studies on *in vitro* blood-brain barrier models: physiology, pathology and pharmacology. *Cell. Mol. Neurobiol.* 25, 59–127. doi: 10.1007/s10571-004-1377-8
- De Strooper, B. (2014). Lessons from a failed γ -secretase Alzheimer trial. *Cell* 159, 721–726. doi: 10.1016/j.cell.2014.10.016
- Di Marco, L. Y., Venneri, A., Farkas, E., Evans, P. C., Marzo, A., and Frangi, A. F. (2015). Vascular dysfunction in the pathogenesis of Alzheimer's disease—a review of endothelium-mediated mechanisms and ensuing vicious circles. *Neurobiol. Dis.* 82, 593–606. doi: 10.1016/j.nbd.2015.08.014
- Doshina, A., Gourgue, F., Onizuka, M., Opsomer, R., Wang, P., Ando, K., et al. (2017). Cortical cells reveal APP as a new player in the regulation of GABAergic neurotransmission. *Sci. Rep.* 7:370. doi: 10.1038/s41598-017-00325-2
- Duyckaerts, C., Delatour, B., and Potier, M. C. (2009). Classification and basic pathology of Alzheimer disease. *Acta Neuropathol.* 118, 5–36. doi: 10.1007/s00401-009-0532-1
- Esparza, T. J., Zhao, H., Cirrito, J. R., Cairns, N. J., Bateman, R. J., Holtzman, D. M., et al. (2013). Amyloid- β oligomerization in Alzheimer dementia versus highpathology controls. *Ann. Neurol.* 73, 104–119. doi: 10.1002/ana.23748
- Fukumoto, H., Deng, A., Irizarry, M. C., Fitzgerald, M. L., and Rebeck, G. W. (2002). Induction of the cholesterol transporter ABCA1 in central nervous system cells by liver X receptor agonists increases secreted A β levels. *J. Biol. Chem.* 277, 48508–48513. doi: 10.1074/jbc.M209085200
- Giannini, E., Botta, F., Fasoli, A., Ceppa, P., Risso, D., Lantieri, P. B., et al. (1999). Progressive liver functional impairment is associated with an increase in AST/ALT ratio. *Dig. Dis. Sci.* 44, 1249–1253. doi: 10.1023/A:1026609231094
- Gouras, G. K., Olsson, T. T., and Hansson, O. (2015). β -amyloid peptide and amyloid plaques in Alzheimer's disease. *Neurotherapeutics* 12, 3–11. doi: 10.1007/s13311-014-0313-y
- Hardy, J., and Selkoe, D. J. (2002). The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *Science* 297, 353–356. doi: 10.1126/science.1072994
- Haseloff, R. F., Dithmer, S., Winkler, L., Wolburg, H., and Blasig, I. E. (2015). Transmembrane proteins of the tight junctions at the blood-brain barrier: structural and functional aspects. *Semin. Cell Dev. Biol.* 38, 16–25. doi: 10.1016/j.semcdb.2014.11.004

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- He, Z., Guo, J. L., McBride, J. D., Narashimhan, S., Kim, H., Changolkar, L., et al. (2018). Amyloid- β plaques enhance Alzheimer's brain tau-seeded pathologies by facilitating neuritic plaque and aggregation. *Nat. Med.* 24, 29–38. doi: 10.1038/nm.4443
- Héraud, C., Goufak, D., Ando, K., Leroy, K., Suain, V., Yilmaz, Z., et al. (2014). Increased misfolding and truncation of tau in APP/PS1/tau transgenic mice compared to mutant tau mice. *Neurobiol. Dis.* 62, 100–112. doi: 10.1016/j.nbd.2013.09.010
- Hong, S., Ostaszewski, B. L., Yang, T., O'Malley, T. T., Jin, M., Yanagisawa, K., et al. (2014). Soluble A β oligomers are rapidly sequestered from brain ISF *in vivo* and bind GM1 ganglioside on cellular membranes. *Neuron* 82, 308–319. doi: 10.1016/j.neuron.2014.02.027
- Hooijmans, C. R., and Kiliaan, A. J. (2008). Fatty acids, lipid metabolism and Alzheimer pathology. *Eur. J. Pharmacol.* 585, 176–196. doi: 10.1016/j.ejphar.2007.11.081
- Jack, C. R., Knopman, D. S., Jagust, W. J., Petersen, R. C., Weiner, M. W., Aisen, P. S., et al. (2013). Tracking pathophysiological processes in Alzheimer's disease: an updated hypothetical model of dynamic biomarkers. *Lancet Neurol.* 12, 207–216. doi: 10.1016/S1474-4422(12)70291-0
- Jiao, H., Wang, Y. Z., Liu, P., and Wang, Y. X. (2011). Specific role of tight junction proteins claudin-5, occludin and ZO-1 of the blood-brain barrier in a focal cerebral ischemic insult. *J. Mol. Neurosci.* 44, 130–139. doi: 10.1007/s12031-011-9496-4
- Jin, W. S., Shen, L. L., Bu, X. L., Zhang, W. W., Chen, S. H., Huang, Z. L., et al. (2017). Peritoneal dialysis reduces amyloid- β plasma levels in humans and attenuates Alzheimer-associated phenotypes in an APP/PS1 mouse model. *Acta Neuropathol.* 134, 207–220. doi: 10.1007/s00401-017-1721-y
- Jin, M., Shepardson, N., Yang, T., Chen, G., Walsh, D., and Selkoe, D. J. (2011). Soluble amyloid β -protein dimers isolated from Alzheimer cortex directly induce Tau hyperphosphorylation and neuritic degeneration. *Proc. Natl. Acad. Sci. U S A* 108, 5819–5824. doi: 10.1073/pnas.1017033108
- Kanekiyo, T., and Bu, G. (2014). The low-density lipoprotein receptor-related protein 1 and amyloid- β clearance in Alzheimer's disease. *Front Aging Neurosci.* 6:93. doi: 10.3389/fnagi.2014.00093
- Kang, D. E., Pietrzik, C. U., Baum, L., Chevallier, N., Merriam, D. E., Kounnas, M. Z., et al. (2000). Modulation of amyloid β -protein clearance and Alzheimer's disease susceptibility by the LDL receptor-related protein pathway. *J. Clin. Invest.* 106, 1159–1166. doi: 10.1172/JCI11013
- Karbalaei, R., Allahyari, M., Rezaei-Tavirani, M., Asadzadeh-Aghdaei, H., and Zali, M. R. (2018). Protein-protein interaction analysis of Alzheimer's disease and NAFLD based on systems biology methods unhide common ancestor pathways. *Gastroenterol. Hepatol. Bed Bench* 1, 27–33. doi: 10.22037/ghfbb.v0i0.1327
- Kim, D. G., Krenz, A., Toussaint, L. E., Maurer, K. J., Robinson, S. A., Yan, A., et al. (2016). Non-alcoholic fatty liver disease induces signs of Alzheimer's disease (AD) in wild-type mice and accelerates pathological signs of AD in an AD model. *J. Neuroinflammation* 13:1. doi: 10.1186/s12974-015-0467-5
- Klevanski, M., Herrmann, U., Weyer, S. W., Fol, R., Cartier, N., Wolfer, D. P., et al. (2015). The APP intracellular domain is required for normal synaptic morphology, plasticity and hippocampus-dependent behavior. *J. Neurosci.* 35, 16018–16033. doi: 10.1523/JNEUROSCI.2009-15.2015
- Konietzko, U. (2011). AICD nuclear signaling and its possible contribution to Alzheimer's disease. *Curr. Alzheimer Res.* 9, 200–216. doi: 10.2174/156720512799361673
- Kook, S. Y., Seok Hong, H., Moon, M., and Mook-Jung, I. (2013). Disruption of blood-brain barrier in Alzheimer disease pathogenesis. *Tissue Barriers* 1:e23993. doi: 10.4161/tisb.23993
- Kroll, R. A., and Neuwelt, E. A. (1998). Outwitting the blood-brain barrier for therapeutic purposes: osmotic opening and other means. *Neurosurgery* 42, 1083–1099; discussion 1099–1100. doi: 10.1097/00006123-199805000-00082
- Lésne, S. E., Sherman, M. A., Grant, M., Kuskowski, M., Schneider, J. A., Bannet, D. A., et al. (2013). Brain amyloid- β oligomers in ageing and Alzheimer's disease. *Brain* 136, 1383–1398. doi: 10.1093/brain/awt062
- Lin, L., Yee, S. W., Kim, R. B., and Giacomini, K. M. (2015). SLC transporters as therapeutic targets: emerging opportunities. *Nat. Rev. Drug Discov.* 14, 543–560. doi: 10.1038/nrd4626
- Liu, Y. H., Wang, Y. R., Xiang, Y., Zhou, H. D., Giunta, B., Mañucat-Tan, N. B., et al. (2015). Clearance of amyloid- β in Alzheimer's disease: shifting the action site from center to periphery. *Mol. Neurobiol.* 51, 1–7. doi: 10.1007/s12035-014-8694-9
- Liu, Y. H., Zeng, F., Wang, Y. R., Zhou, H. D., Giunta, B., Tan, J., et al. (2013). Immunity and Alzheimer's disease: immunological perspectives on the development of novel therapies. *Drug Discov. Today* 18, 1212–1220. doi: 10.1016/j.drudis.2013.07.020
- Mahley, R. W. (1988). Apolipoprotein E: cholesterol transport protein with expanding role in cell biology. *Science* 240, 622–630. doi: 10.1126/science.3283935
- Masserini, M. (2013). Nanoparticles for brain drug delivery. *ISRN Biochem.* 2013:238428. doi: 10.1155/2013/238428
- Montagne, A., Barnes, S. R., Sweeney, M. D., Halliday, M. R., Sagare, A. P., Zhao, Z., et al. (2015). Blood-brain barrier breakdown in the aging human hippocampus. *Neuron* 85, 296–302. doi: 10.1016/j.neuron.2014.12.032
- Morris, G. P., Clark, I. A., and Vissel, B. (2014). Inconsistencies and controversies surrounding the amyloid hypothesis of Alzheimer's disease. *Acta Neuropathol. Commun.* 2:135. doi: 10.1186/s40478-014-0135-5
- Mroczkow, B., Groblewska, M., Litman-Zawadzka, A., Kornhuber, J., and Lewczuk, P. (2018). Cellular receptors of amyloid β oligomers (A β Os) in Alzheimer's disease. *Int. J. Mol. Sci.* 19:1884. doi: 10.3390/ijms19071884
- Müller-Schiffmann, A., Herring, A., Abdel-Hafiz, L., Chepkova, A. N., Schäble, S., Wedel, D., et al. (2016). Amyloid- β dimers in the absence of plaque pathology impair learning and synaptic plasticity. *Brain* 139, 509–525. doi: 10.1093/brain/awv355
- Nation, D. A., Sweeney, M. D., Montagne, A., Sagare, A. P., D'Orazio, L. M., Pachicano, M., et al. (2019). Blood-brain barrier breakdown is an early biomarker of human cognitive dysfunction. *Nat. Med.* 25, 270–276. doi: 10.1038/s41591-018-0297-y
- Octave, J. N., Pierrot, N., Ferao Santos, S., Nalivaeva, N. N., and Turner, A. (2013). From synaptic spines to nuclear signaling: nuclear and synaptic actions of the amyloid precursor protein. *J. Neurochem.* 126, 183–190. doi: 10.1111/jnc.12239
- Øie, C. I., Appa, R. S., Hilden, I., Petersen, H. H., Gruhler, A., Smedsrød, B., et al. (2011). Rat liver sinusoidal endothelial cells (LSECs) express functional low density lipoprotein receptor-related protein-1 (LRP-1). *J. Hepatol.* 55, 1346–1352. doi: 10.1016/j.jhep.2011.03.013
- Pardridge, W. M. (2015). Blood-brain barrier endogenous transporters as therapeutic targets: a new model for small molecule CNS drug discovery. *Expert Opin. Ther. Targets* 19, 1059–1072. doi: 10.1517/14728222.2015.1042364
- Provias, J., and Jaynes, B. (2014). The role of the blood-brain barrier in the pathogenesis of senile plaques in Alzheimer's disease. *Int. J. Alzheimers Dis.* 2014:191863. doi: 10.1155/2014/191863
- Rice, H. C., de Malmazet, D., Schreurs, A., Frere, S., Van Molle, I., Volkov, A. N., et al. (2019). Secreted amyloid- β precursor protein functions as a GABABR1a ligand to modulate synaptic transmission. *Science* 363:eaa04827. doi: 10.1126/science.aa04827
- Roher, A. E., Esh, C. L., Kokjohn, T. A., Castaño, E. M., Van Vickle, G. D., Kalback, W. M., et al. (2009). Amyloid beta peptides in human plasma and tissues and their significance for Alzheimer's disease. *Alzheimers Dement.* 5, 18–29. doi: 10.1016/j.jalz.2008.10.004
- Rolle, A., Paredes, S., Cortínez, L. I., Anderson, B.-J., Quezada, N., Solari, S., et al. (2018). Dexmedetomidine metabolic clearance is not affected by fat mass in obese patients. *Br. J. Anaesth.* 120, 969–977. doi: 10.1016/j.bja.2018.01.040
- Sagare, A. P., Deane, R., and Zlokovic, B. V. (2012). Low-density lipoprotein receptor-related protein 1, A physiological A β homeostatic mechanism with multiple therapeutic opportunities. *Pharmacol. Ther.* 136, 94–105. doi: 10.1016/j.pharmthera.2012.07.008
- Sehgal, N., Gupta, A., Valli, R. K., Joshi, S. D., Mills, J. T., Hamel, E., et al. (2012). Withania somnifera reverses Alzheimer's disease pathology by enhancing low-density lipoprotein receptor-related protein in liver. *Proc. Natl. Acad. Sci. U S A* 109, 3510–3515. doi: 10.1073/pnas.1112209109
- Selkoe, D. J., and Hardy, J. (2016). The amyloid hypothesis of Alzheimer's disease at 25 years. *EMBO Mol. Med.* 8, 595–608. doi: 10.15252/emmm.201606210

- Serrano-Pozo, A., Frosch, M. P., Masliah, E., and Hyman, B. T. (2011). Neuropathological alterations in Alzheimer disease. *Cold Spring Harb. Perspect. Med.* 1:a006189. doi: 10.1101/cshperspect.a006189
- Simpson, I. A., Carruthers, A., and Vannucci, S. J. (2007). Supply and demand in cerebral energy metabolism: the role of nutrient transporters. *J. Cereb. Blood Flow Metab.* 27, 1766–1791. doi: 10.1038/sj.jcbfm.9600521
- Southam, K. A., Stennard, F., Pavez, C., and Small, D. H. (2019). Knockout of amyloid- β protein precursor (APP) expression alters synaptogenesis, neurite branching and axonal morphology of hippocampal neurons. *Neurochem. Res.* 44, 1346–1355. doi: 10.1007/s11064-018-2512-0
- Stanyon, H. F., and Viles, J. H. (2012). Human serum albumin can regulate amyloid- β peptide fiber growth in the brain interstitium. *J. Biol. Chem.* 287, 28163–28168. doi: 10.1074/jbc.C112.360800
- Tamaki, C., Ohtsuki, S., and Terasaki, T. (2007). Insulin facilitates the hepatic clearance of plasma amyloid β -peptide (1–40) by intracellular translocation of low-density lipoprotein receptor-related protein 1 (LRP-1) to the plasma membrane in hepatocytes. *Mol. Pharmacol.* 72, 850–855. doi: 10.1124/mol.107.036913
- Tyan, S. H., Shih, A., Walsh, J., Maruyama, H., Sarsoza, F., Ku, L., et al. (2012). Amyloid precursor protein (APP) regulates synaptic structure and function. *Mol. Cell. Neurosci.* 51, 43–52. doi: 10.1016/j.mcn.2012.07.009
- Ulrich, J. D., Huynh, T. P., and Holtzman, D. M. (2015). Re-evaluation of the blood-brain barrier in the presence of Alzheimer's disease pathology. *Neuron* 88, 237–239. doi: 10.1016/j.neuron.2015.10.008
- Van de Haar, H. J., Burgmans, S., Hofman, P. A., Verhey, F. R., Jansen, J. F., and Backes, W. H. (2015). Blood-brain barrier impairment in dementia: current and future *in vivo* assessments. *Neurosci. Biobehav. Rev.* 49, 71–81. doi: 10.1016/j.neubiorev.2014.11.022
- Verma, M., Vats, A., and Taneja, V. (2015). Toxic species in amyloid disorders: oligomers or mature fibrils. *Ann. Indian Acad. Neurol.* 18, 138–145. doi: 10.4103/0972-2327.144284
- Viggars, A. P., Wharton, S. B., Simpson, J. E., Matthews, F. E., Brayne, C., Savva, G. M., et al. (2011). Alterations in the blood brain barrier in ageing cerebral cortex in relationship to Alzheimer-type pathology: a study in the MRC-CFAS population neuropathology cohort. *Neurosci. Lett.* 505, 25–30. doi: 10.1016/j.neulet.2011.09.049
- Wang, Y. R., Wang, Q. H., Zhang, T., Liu, Y. H., Yao, X. Q., Zeng, F., et al. (2017). Associations between hepatic functions and plasma amyloid- β levels—implications for the capacity of liver in peripheral amyloid- β clearance. *Mol. Neurobiol.* 54, 2338–2344. doi: 10.1007/s12035-016-9826-1
- Wang, Z., Yang, L., and Zheng, H. (2012). Role of APP and A β in synaptic physiology. *Curr. Alzheimer Res.* 9, 217–226. doi: 10.2174/156720512799361691
- Wang, Y. J., Zhou, H. D., and Zhou, X. F. (2006). Clearance of amyloid-beta in Alzheimer's disease: progress, problems and perspectives. *Drug Discov. Today* 11, 931–938. doi: 10.1016/j.drudis.2006.08.004
- Xiang, Y., Bu, X. L., Liu, Y. H., Zhu, C., Shen, L. L., Jiao, S. S., et al. (2015). Physiological amyloid-beta clearance in the periphery and its therapeutic potential for Alzheimer's disease. *Acta Neuropathol.* 130, 487–499. doi: 10.1007/s00401-015-1477-1
- Xiao, H., Deng, M., Yang, B., Tang, J., and Hu, Z. (2017). Role of glycogen synthase kinase 3 in ischemia-induced blood-brain barrier disruption in aged female rats. *J. Neurochem.* 142, 194–203. doi: 10.1111/jnc.14051
- Xiao, G., and Gan, L. S. (2013). Receptor-mediated endocytosis and brain delivery of therapeutic biologics. *Int. J. Cell Biol.* 2013:703545. doi: 10.1155/2013/703545
- Zhao, Y., Li, D., Zhao, J., Song, J., and Zhao, Y. (2016). The role of the low-density lipoprotein receptor-related protein 1 (LRP-1) in regulating blood-brain barrier integrity. *Rev. Neurosci.* 27, 623–634. doi: 10.1515/revneuro-2015-0069
- Ziegler-Waldkirch, S., and Meyer-Luehmann, M. (2018). The role of glial cells and synapse loss in mouse models of Alzheimer's disease. *Front. Cell. Neurosci.* 12:473. doi: 10.3389/fncel.2018.00473
- Zlokovic, B. V. (2005). Neurovascular mechanisms of Alzheimer's neurodegeneration. *Trends Neurosci.* 28, 202–208. doi: 10.1016/j.tins.2005.02.001
- Zlokovic, B. V. (2011). Neurovascular pathways to neurodegeneration in Alzheimer's disease and other disorders. *Nat. Rev. Neurosci.* 12, 723–738. doi: 10.1038/nrn3114

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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