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SPECIALTY SECTION

This article was submitted to
Animal Nutrition and Metabolism,
a section of the journal
Frontiers in Veterinary Science

RECEIVED 08 July 2022

ACCEPTED 15 September 2022

PUBLISHED 13 October 2022

CITATION

Azzaz HH, Kholif AE, Murad HA and
Vargas-Bello-Pérez E (2022) A newly
developed strain of *Enterococcus
faecium* isolated from fresh dairy
products to be used as a probiotic in
lactating Holstein cows.
Front. Vet. Sci. 9:989606.
doi: 10.3389/fvets.2022.989606

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A newly developed strain of *Enterococcus faecium* isolated from fresh dairy products to be used as a probiotic in lactating Holstein cows

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The objective of this study was to determine the ability of an isolated strain (EGY_NRC1) or commercial (NCIMB 11181) *Enterococcus faecium* as a probiotic for lactating cows. Two experiments were conducted: In Experiment 1, the effects of three levels (1, 2, and 3 g/kg diet, DM basis) of isolated and commercial *E. faecium* on *in vitro* ruminal fermentation kinetics, gas, methane (CH₄) and nutrient degradability were determined. In Experiment 2, thirty multiparous Holstein cows (633 ± 25.4 kg body weight) with 7 days in milk, were randomly assigned to 3 treatments in a completely randomized design in a 60-day experiment. Cows were fed without any additives (control treatment) or supplemented with 2 g/kg feed daily of *E. faecium* EGY_NRC1 (contain 1.1 × 10⁹ CFU/g) or commercial *E. faecium* NCIMB 11181 (contain 2 × 10¹² CFU/g). Diets were prepared to meet cow's nutrient requirements according to NRC recommendations. Probiotic doses were based on the *in vitro* Experiment 1. Feed intake, digestibility, blood parameters and lactation performance were evaluated. In Experiment 1, the isolated *E. faecium* linearly and quadratically increased ($P < 0.001$) *in vitro* total gas production (TGP), the degradability of dry matter (dDM) and organic matter (dOM) while decreased ($P < 0.05$) methane (CH₄) percent of TGP, NH₃CH₄ production, and pH. The commercial *E. faecium* increased TGP and decreased ($P < 0.01$) CH₄ production, pH and increased the dDM and dOM, short chain fatty acids and ruminal NH₃-N concentration. In Experiment 2, the isolated *E. faecium* increased ($P < 0.01$) total tract digestibility of DM, neutral and acid detergent fiber, daily milk production and feed efficiency compared to the control treatment without affecting feed intake and milk composition. Moreover, the isolated *E. faecium* increased ($P < 0.05$) the proportion of C18:1 *trans*-9, C18:2 *cis*-9-12 and C18:2 *trans*-10 *cis*-12. Both isolated and commercial *E. faecium* improved ($P < 0.01$) organic matter, crude protein and nonstructural carbohydrates digestibility, increased serum glucose ($P = 0.002$) and decreased serum cholesterol ($P = 0.002$). Additionally, both

E. faecium strains decreased C23:0 ($P = 0.005$) in milk. In conclusion, the use of *E. faecium* (isolated and commercial) at 2 g/kg DM of feed improved feed efficiency and production performance, with superior effects on animal performance from isolated *E. faecium* compared to the commercial one.

KEYWORDS

Enterococcus faecium, digestion, feed utilization, lactic acid bacteria, milk production

Introduction

Probiotics and prebiotics have been administered to animals for several years to enhance their health and production. In animal production, probiotics are now widely accepted as safe and sustainable alternatives to antibiotics (1, 2). For many years, lactic acid bacteria (LAB) have been used in livestock production, as probiotic supplements or as silage preservative by inhibiting pathogenic microorganisms (e.g., fungal and clostridial growth) and increasing lactic acid formation (3, 4). Normally, LAB are predominantly found in the gastrointestinal tract of animals and humans and are also found in dairy products (5).

In livestock, different bacterial and fungal species (i.e., *Bacillus*, *Bifidobacterium*, *Enterococcus*, *Lactobacillus*, *Propionibacterium*, *Megasphaera elsdenii* and *Prevotella bryantii*) have been used as probiotics as well as strains of *Aspergillus* and *Saccharomyces* (1, 6). Strains of LAB including *Lactobacillus*, *Bifidobacterium* and *Streptococcus* are commonly used as probiotics in ruminant feed (1, 7). LAB can alter ruminal fermentation and enhance nutrient digestibility and productive performance (8). LAB reduce oxygen from the rumen environment and prevent excess of ruminal lactate production, inhibit ruminal pathogens, and modulate the microbial balance (9). Previous studies have reported the effect of LAB in ruminant diets, including the increased yield of microbial biomass (10), reduced methane (11) and increased dry matter (DM) digestibility (12). Recently, it has been reported that feeding dairy cows with *L. casei* TH14 can improve feed utilization, rumen fermentation parameters and milk production (13).

The use of commercial LAB can increase total mixed rations costs and finding alternatives that keep similar efficiency to that from commercial products is of great interest for production improvement. Also, research on novel strains of microorganisms with different origins and properties is still needed. Therefore, the objective of this study was to determine the ability of an isolated strain (EGY_NRC1) or commercial (NCIMB 11181) *Enterococcus faecium* as a probiotic for lactating cows. The hypothesis of this study was that the in-feed supplementation of commercial or isolated *Enterococcus faecium* would lead to

similar effects on feed utilization, milk production, milk composition and milk fatty acid profile in early lactating dairy Holstein cows.

Materials and methods

Isolation and identification of *E. faecium*

Lactic acid bacterial strains were isolated from 15 samples of fresh dairy products (homemade 8 samples of yogurt and 5 samples soft white cheese). For each sample, 10 g were added to 90 ml sterile saline solution and homogenized by vortex for 10 min. Decimal dilutions were placed on double layered M17 agar plates then incubated for 48 h at 30°C. Well defined round colonies were selected randomly and only Gram-positive catalase-negative cocci were retained and stored in M17 broth for further experiments.

To characterize the selected isolate, the carbohydrate fermentation pattern (14) of the selected isolate (that possessed antibacterial activity) and its ability to produce ammonia (NH_3) from arginine (15) was examined. After that, the strain was identified by matrix-assisted laser desorption ionization-time-of-flight mass spectrometry (MALDI-TOF MS) and 16S ribosomal DNA (rDNA) sequencing. The isolate was identified (Figure 1) via the analysis of its total proteome in which a score with more than 1.7 indicates genus identification and a score with more than 2 is the confidence value at the species level (16).

For DNA extraction, genomic DNA was used from the selected isolate that was cultivated at 37°C for 24 h. Freshly prepared culture was subjected to 16S rDNA PCR partial amplification by use of Qiagen genomic DNA purification kit. The genomic DNA was used as a template for PCR amplification of a segment of its 16S rRNA gene. The two primers used were previously described by Liu et al. (17), 8f (5'_AGAGTTTGATCCTGGCTCAG-3) and 1495 R (5'_CTACGGCTACCTTGTACGA-3). The PCR products yielded were analyzed on a 1% (w/v) agarose gel after staining with ethidium bromide. The PCR products were separated on an agarose gel, followed by ethidium bromide staining to check for the presence of a unique amplicon. When a gene from a particular species was amplified using a primer initially designed for a different species, the corresponding amplicon was

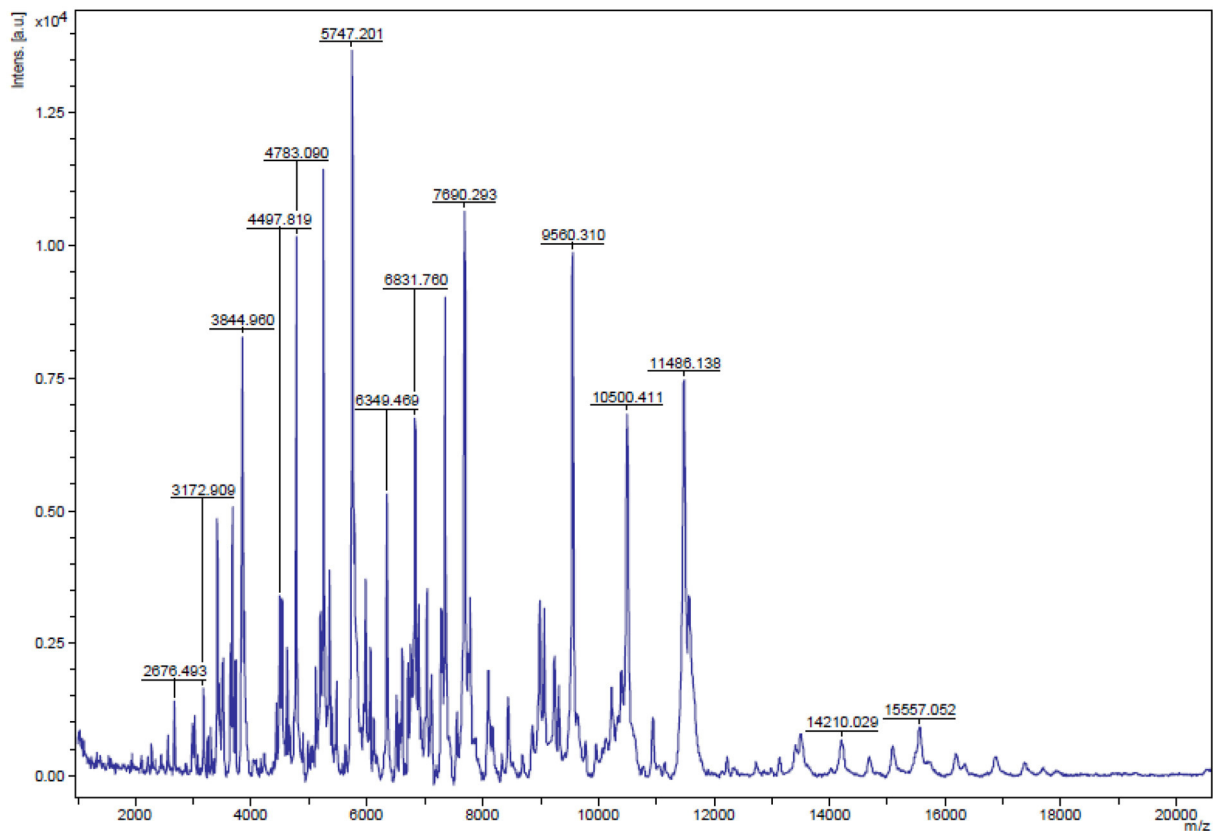


FIGURE 1

Result of the matrix-assisted laser desorption ionization time-of-flight mass spectrometry (MALDI-TOF MS) analysis. The MALDI-TOF MS showed that the isolated strain of *E. faecium* that possess antimicrobial activity was initially identified with a high confidence value of 2.31 that indicated a reliable identification of the isolate up to species level.

purified by Promega Wizard Genomic DNA Purification Kit and sequenced by HVD life science, Germany.

For phylogenetic analysis (data not shown), the 16S rDNA sequence obtained was added to publicly available bacterial 16SrRNA sequences and integrated into the database with the automatic alignment tool. Phylogenetic analysis was inferred using neighbor joining method and the phylogenetic tree was constructed (18). The isolated LAB was identified as *E. faecium* EGY_NRC1 with NCBI accession number MW856655.

The biomass production of the isolated LAB was done on whey permeate media with the following composition (/L permeate): 5 g yeast extract, 5 g peptone, 0.5 g magnesium sulfate, 3 g ammonium chloride and 2.5 g ascorbic acid. The medium was inoculated individually with 5% (v/v) of the isolated LAB culture (24 h old age activated M17 broth culture) and then incubated at 37°C for 48 h. The cultured biomasses were separated by centrifugation at 6,000 rpm for 15 min at 4°C then added to dry permeate as a carrier agent, and dried in oven at 37°C. The un-inoculated media were used as control. Unpublished data showed that the viable count of the strain after pre-incubation at 60°C for 30 min was enumerated and the

result indicated that the remaining count was 3.274 log CFU/mL in compared to 5.477 log CFU/mL without pre-incubation. The reported viable count was determined after drying.

Experiment 1

In vitro evaluation

Using a stomach tube, rumen liquor was obtained from three adult Barki sheep (51 ± 2.6 kg of body weight) fed a fixed amount of concentrate (500 g) and *ad libitum* berseem hay daily. The rumen contents (liquid and solid contents 1:1 v/v) were collected before morning feeding, kept in pre-warmed thermo containers at 39°C under anaerobic conditions. About 500 mL of ruminal fluid was collected from all ewes of each treatment. To avoid saliva contamination, the first 50 mL of the rumen fluid samples were discarded. The rumen fluid was mixed for 10 s, squeezed through four layers of cheese cloth, and maintained in a water bath at 39°C under continuous CO₂ flushing until inoculation (19). Three incubation runs were performed in three different weeks. Rumen contents obtained

from the three sheep were combined for each run. Animal use for this trial was approved by the technical committee of the Science, Technology & Innovation Funding Authority (STDF), Egypt (project STDF 33413).

A total mixed ration composed of (per kg DM) 300 g berseem clover, 300 g corn silage, 150 g soybean meal, and 250 g yellow corn was used as a substrate. The nutrient contents of feed ingredients and basal diet are shown in Table 1. The *in vitro* total gas production (TGP) assay was conducted as described by Theodorou et al. (19) and adapted to the semi-automatic system of Mauricio et al. (20). Ground substrate samples (500 mg of DM) were incubated in 120 mL serum bottles (5 bottles per dose of LAB at each time). LAB (isolated and commercial) was included at 0, 1, 2, and 3 g/kg DM substrate. The isolated bacteria contained 1.1×10^9 CFU/g. Levels of inclusion were based on initial screening of many doses of *E. faecium* on ruminal fermentation (2, 21). After dispensing, bottles were closed with rubber stoppers, shaken manually, and incubated at 39°C in a forced-air oven for 48 h. The bottles were shaken at 1 h intervals during incubation. The amount of TGP was calculated according to the regression equation $\{V = 4.974 12 \times p + 0.171; n = 500, r^2 = 0.98; [V \text{ is gas volume (mL); } p \text{ is measured pressure (psi)}]\}$ obtained in our laboratory under our conditions according to the gas volume vs. pressure. Bottles containing only buffered rumen fluid without substrate were considered as blanks. At each incubation time, 5 mL of headspace gas was taken from each bottle and infused into a Gas-Pro detector (Gas Analyzer CROWCON Model Tetra3, Abingdon, UK) to measure the concentration of CH₄ and carbon dioxide. The control and experimental treatments were tested in 6 bottles (analytical replicates) and three incubation runs in three consecutive weeks with 3 bottles containing inoculum and buffer but no feed (blanks).

At 48 h (the end of incubation), fermentation was terminated by immersing the bottles in ice. For each LAB level, 3 bottles were used to measure the pH, NH₃-N (22) and short chain fatty acids (SCFA) by steam distillation and titration (23), whereas the other 3 bottles were filtered in pre-weighed

crucibles and washed with hot water then acetone, and the residual DM and ash were estimated to determine true DM, organic matter (OM), neutral detergent fiber (NDF) and acid detergent fiber (ADF) degradability (dDM, dOM, dNDF, and dADF, respectively).

Experiment 2

Animals

Cows were managed in accordance with the Guide for the Care and Use of Agricultural Animals in Agricultural Research and Teaching, 3rd edition, 2010 (Federation of Animal Science Societies, Champaign, IL, USA). The protocol was approved by the technical committee of the Science, Technology & Innovation Funding Authority (STDF), Egypt (project STDF 33413).

During the first week of lactation, thirty lactating Holstein cows (633 ± 25.4 kg body weight, 3 ± 1 parity, 7 ± 1 days in milk, with a previous milk production of 24 ± 2.2 kg/d, were assigned randomly to one of three experimental treatments in a complete randomized designed with 10 cows per treatment for 60 days.

Cows were divided into three barns in soil-surfaced free stalls (122×175 cm²/cows), under shade, without any bedding and with free access to water. Cows were fed *ad libitum* a diet containing [per kg DM] 300 g berseem clover, 300 g corn silage, 150 g soybean meal and 250 g grounded corn grain, to meet their nutrient requirements according to NRC (24) recommendations for 650 kg cow with 20 kg DM intake and 35 kg daily milk production. Animals were fed 10% more of the expected dry matter intake to ensure collection of orts. The diet fed to cows was the same for the *in vitro* experiment.

Cows were fed their diets without any additives (control treatment) or supplemented with 2 g/kg feed daily of *E. faecium* EGY_NRC1 (isolated to contain 1.1×10^9 CFU/g) or commercial *E. faecium* NCIMB 11181 (isolated to contain 2×10^{12} CFU/g; ADM Protexin Limited, Lopen Head, Somerset,

TABLE 1 Chemical composition of ingredients and control diet used in the *in vitro* and *in vivo* experiments (g/kg of dry matter).

	Yellow corn ^a	Soybean meal ^a	Corn silage ^a	Berseem clover ^a	Basal diet ^b
Dry matter, wet weight	896	898	891	889	893
Organic matter	985	930	915	870	921
Crude protein	83	422	77	170	158
Ether extract	51	48	21	32	36
Ash	15	70	85	130	79
Non-structural carbohydrates ^c	666	313	286	258	377
Neutral detergent fiber	185	147	531	410	351
Acid detergent fiber	37	65	320	270	196

^aAnalyzed values. ^bCalculated values. Used as a diet for all treatments. ^cNon-structural carbohydrates = 1,000–(neutral detergent fiber + crude protein + ether extract + ash).

TA13 5JH UK). The doses of probiotics were based on the results obtained from Experiment 1 (*in vitro* experiment). Diets were offered twice a day at 08:00 and 16:00 h. The additives were mixed with all feedstuffs using a feeding wagon. Before use, probiotics were kept at 4°C. Samples of feed ingredients were taken daily, composited weekly, dried at 60°C in a forced-air oven for 48 h (method 930.15) (22) and stored for chemical analyses.

The total mixed ration was prepared and distributed using a horizontal mixer system (DeLaval, Ontario, Canada) after mixing for 20 min. Samples of feed were taken daily, composited weekly, dried at 60°C in a forced-air oven for 48 h (22) and stored for chemical analysis. The nutrient contents of the feed ingredients are shown in Table 1. Cows were weighed on a digital multi-purpose platform scale at the beginning and at the end of the experiment.

Nutrient intake and digestibility

Feed intake was recorded daily by weighing the total daily amount of feed offered to each cow and the total daily amounts of weighedorts. On the 4th and 8th week, nutrient digestibility trials were conducted, in which acid insoluble ash was used as an internal indigestibility marker (25). Acid-insoluble ash contents of feeds and feces were determined gravimetrically after drying, burning, boiling of ash in hydrochloric acid, filtering and washing of the hot hydrolysate, and re-burning. Coefficients of digestion were calculated according to Ferret et al. (26). For the digestibility trial, fecal samples were collected from all cows twice daily at 09:00 and 16:00 h, dried at 60°C for 48 h in a forced-air oven and pooled by cow. The nutritive value of diets expressed as total digestible nutrients (TDN), digestible energy (DE), metabolizable energy (ME), net energy for lactation (NEL) were calculated according to NRC (24) equation, while the net energy requirements for lactation equivalent of 1 kg of standard air-dry barley (UFL) was calculated according to INRA (27) equation.

The dried feed, orts and fecal samples were ground to pass a 1-mm screen using a Wiley mill and analyzed for DM (method ID 930.15), ash (method ID 942.05), nitrogen (method ID 954.01) and ether extract (EE; method ID 920.39) according to the AOAC (22) official methods. Neutral detergent fiber was determined by the procedure of Van Soest et al. (28) with the use of alpha amylase and sodium sulphite. Acid detergent fiber was analyzed according to the AOAC (22). Lignin was analyzed according to Van Soest et al. (28).

Sampling and analysis of blood serum

On the last day of the 4th and 8th weeks of the experiment, individual blood samples (10 mL) were taken before morning feeding at 08:00 h from the jugular vein. Blood samples were centrifuged at 4,000 ×g for 20 min at 4°C. The serum was

separated into 2 mL Eppendorf tubes and frozen at −20°C until analysis. By using specific kits (Spinreact, Ctra. Santa Coloma, Girona, Spain) and following the manufacturers' instructions, blood serum samples were analyzed for total protein, albumin, globulin, urea-N, glucose, cholesterol, triglycerides, aspartate aminotransferase (AST), and alanine aminotransferase (ALT). The globulin concentration was calculated by subtracting the albumin values from their corresponding total protein values.

Milk sampling, and milk composition

Cows were milked (DeLaval parallel parlor P2100, SE-147 21 Tumba, Sweden) three times daily at 04:00, 12:00 and 20:00 h, and individual milk samples (30 g/kg of milk yield) were collected at each milking. A mixed sample of milk (proportional to amounts isolated in each milking time) was taken daily every 2 weeks to determine milk composition. Milk samples were analyzed using infrared spectrophotometry (Ekomilk-M ultrasonic milk analyzer, EON Trading 2000, INC, Bulgaria).

The gross energy content was calculated according to Tyrrell and Reid (29). The milk energy output (MJ/d) was calculated as milk energy (MJ/kg) × milk yield (kg/d). The energy corrected milk (ECM) and 4% fat corrected milk (FCM) were calculated according to Sjaunja et al. (30) and Tyrrell and Reid (29), respectively.

Statistical analyses

Data from *in vitro* measurements were analyzed using the GLM procedure of SAS (SAS Inst. Inc. Cary, NC, USA) in a completely randomized design using the following model: $Y_{ij} = \mu + D_i + E_{ij}$, where Y_{ij} represents the measured variable, μ is the overall mean, D_i is the LAB dose, and E_{ij} is the experimental error. Data from each of the three runs within the same sample were averaged prior to the statistical analysis. Polynomial (linear and quadratic) contrasts were used to examine dose responses for increasing levels of LAB.

Data from *in vivo* measurements were analyzed as a completely randomized design with repeated measures using the PROC MIXED procedure of SAS (SAS Institute, Cary, NC, USA), considering sampling time as repeated measures and individual cow as the experimental unit.

Data for variables measured daily for each week were averaged before statistical analyses. The statistical model included the treatment effect, week effect and the treatment × week interaction. Animal nested within treatment was considered the random effect, while treatment was the fixed effect. Two covariance structures were considered in the REPEATED statement in PROC MIXED: compound symmetry (cs) and auto-regressive [AR(1)]. The error structure, with the lowest Akaike information criteria, that fits the statistics was selected for the model. When the *F*-test was significant at

$P < 0.05$, means were compared by applying the probability of difference option of the least squares means statement. Significance was declared at $P < 0.05$.

Results

Experiment 1 (*in vitro* experiment)

Results of TGP and CH₄ proportions differed between the isolated and commercial strains of *E. faecium* (Table 2). The inclusion of isolated *E. faecium* linearly and quadratically increased ($P < 0.001$) TGP, while linearly and quadratically decreased ($P < 0.05$) proportional CH₄, CH₄ production (per g dDM, g dOM, g dNDF, g dADF) and pH, and increased SCFA (linear and quadratic effects, $P < 0.001$). Linear increases in dDM and dOM were observed with the isolated *E. faecium*, with no effect on dNDF or dADF. The inclusion of isolated *E. faecium* did not affect the concentration of ruminal NH₃-N.

Increased TGP and decreased proportional CH₄ (linear and quadratic effects, $P < 0.01$) were observed with the inclusion of the commercial *E. faecium* strain; however, CH₄ production (per g dDM, g dOM, g dNDF, g dADF) and pH linearly decreased ($P < 0.01$), and dDM, dOM, SCFA and ruminal NH₃-N concentration increased, without affecting dNDF and dADF.

Experiment 2 (lactation experiment)

Feed intake, nutrient digestibility, and blood measurements

Feeding *E. faecium* diets did not affect total feed intake (Table 3). Compared to the control, the highest ($P < 0.01$) DM, NDF and ADF degradabilities were observed with the isolated *E. faecium* followed by the commercial *E. faecium*, while both isolated and commercial *E. faecium* improved ($P < 0.01$) OM, CP, EE and NSC digestibility. The isolated *E. faecium* followed by the commercial *E. faecium* showed higher ($P < 0.01$) diet's nutritive value calculated as TDN, DE, ME, NEL, and UFL compared to the control treatment. Both the probiotic strains influenced an increased ($P < 0.05$) intake of digestible OM, TDN, ME and digestible CP compared to control, while between the strains the improvement in TDN, ME intake was superior ($P < 0.05$) in isolated than the commercial strain.

Isolated or commercial *E. faecium* did not affect concentrations of serum total protein, albumin, globulin, albumin: globulin ratio and urea-N (Table 4). Both isolated and commercial *E. faecium* increased serum glucose ($P = 0.002$) and decreased serum cholesterol ($P = 0.002$). The commercial *E. faecium* decreased ($P = 0.002$) serum triglycerides and ALT ($P = 0.038$), while the isolated *E. faecium* decreased serum AST ($P = 0.023$).

Milk production, milk composition, and milk fatty acids

Compared to the control, the isolated *E. faecium* followed by the commercial *E. faecium* increase ($P < 0.001$) daily milk production (actual, ECM and FCM), daily milk component yields and milk energy output (Table 5). Moreover, the isolated *E. faecium* followed by the commercial *E. faecium* improved feed efficiency compared to the control treatment. Treatments did not affect the concentrations of milk components.

Both *E. faecium* strains decreased the proportion of C23:0 ($P = 0.005$) and increased ($P = 0.017$) C18:1 *trans*-9 in milk (Table 6). Compared to the control treatment, the isolated *E. faecium* increased ($P < 0.05$) the proportion of C18:1 *trans*-9, C18:2 *cis*-9-12 and C18:2 *trans*-10 *cis*-12, while the commercial *E. faecium* did not affect them.

Discussion

As shown in Figure 1, the MALDI-TOF MS showed that the isolated strain of *E. faecium* possess antimicrobial activity which was initially identified with a high confidence value of 2.31 that indicated a reliable identification of the isolate up to species level. This result agreed with that of 16S rDNA sequencing data (31). The phylogenetic analysis and the 16S rDNA sequencing assigned all the *E. faecium* EGY NRC1 isolates belonged to *E. faecium*.

Experiment 1 (*in vitro* experiment)

The inclusion of isolated *E. faecium* (both strains) increased TGP. Generally, production of gases depends mainly on the composition and degradability of the incubated substrate and the concentration of the soluble components in the incubated substrates (32–34). In the present experiment, the composition of the diet and soluble components are the same between treatments indicating that the differences are mainly due to the strains of *E. faecium*. Jiao et al. (35) stated that specific LAB strains interact with rumen microorganisms to alter rumen fermentation with different modes of action in the rumen.

One promising area of research for the use of LAB in ruminant nutrition is its potential for reducing CH₄ emissions (36). The isolated and commercial *E. faecium* decreased proportional CH₄ and CH₄ production per unit of degraded DM, OM, NDF and ADF which may be related to the reduced methanogenesis by stimulating the growth of lactate-utilizing bacteria such as *Selenomonas ruminantium*, *Megasphaera elsdenii*, and *Veillonella parvula*, which promotes H₂ and CO₂ sinks during the conversion of lactate to propionate (37). Moreover, LAB stimulates scavenging of hydrogen atoms to form propionate causing a lack of hydrogen as the main substrate for methanogenic bacteria (36). Cao

TABLE 2 *In vitro* fermentation from Experiment 1 (mean values), where a basal diet was supplemented with isolated or commercial *E. faecium* as probiotics at 1, 2 or 3 g/kg DM.

	Control	Isolated probiotics ^a			Commercial probiotics ^a			Pooled SEM	Isolated probiotics		Commercial probiotics		Control vs. others	Isolated vs. Commercial
		1	2	3	1	2	3		Linear	Quadratic	Linear	Quadratic		
TGP, mL/g DM	114	122	126	121	123	115	129	1.1	<0.001	<0.001	0.066	0.090	<0.001	0.001
CH ₄ , %	25.1	20.4	18.5	19.9	20.3	21.2	19.9	0.36	<0.001	<0.001	<0.001	<0.001	<0.001	0.005
CH ₄ /g dDM	59.7	47.9	42.9	45.5	47.7	42.8	45.0	1.26	<0.001	<0.001	<0.001	<0.001	<0.001	0.813
CH ₄ /g dOM	52.3	42.5	38.2	40.4	42.6	39.9	40.5	1.05	<0.001	<0.001	<0.001	<0.001	<0.001	0.473
CH ₄ /g dNDF	79.9	62.5	55.0	58.7	65.8	60.6	61.9	3.53	<0.001	0.005	0.007	0.036	<0.001	0.173
CH ₄ /g dADF	112	86	74	80	91	83	88	7.1	0.002	0.032	0.016	0.080	0.008	0.244
dDM, g/kg	482	519	546	532	521	569	528	13.2	0.006	0.061	0.003	0.004	0.006	0.505
dOM, g/kg	549	585	611	597	583	613	586	14.0	0.010	0.086	0.032	0.040	0.004	0.726
dNDF, g/kg	365	401	427	413	379	407	399	20.4	0.072	0.235	0.162	0.591	0.084	0.280
dADF, g/kg	266	292	318	304	278	297	290	19.5	0.119	0.321	0.308	0.633	0.160	0.321
pH	6.65	6.38	6.33	6.35	6.35	6.30	6.33	0.038	<0.001	0.006	<0.001	<0.001	<0.001	0.377
SCFA, mmol/L	1.18	1.37	1.45	1.45	1.38	1.41	1.41	0.036	<0.001	0.010	<0.001	0.006	<0.001	0.412
NH ₃ -N, mg/dL	12.7	13.1	13.2	13.1	13.0	13.2	13.1	0.18	0.064	0.199	0.111	0.235	0.031	0.710

dADF, acid detergent fiber degradability; CH₄, methane; dDM, dry matter degradability; dNDF, neutral detergent fiber degradability; NH₃-N, Ammonia-N (mg/dL); dOM, organic matter degradability (g/kg); SCFA, short chain fatty acids (mmol/L); TGP, *in vitro* total gas production (mL/g DM). ^aThe isolated product contained *E. faecium* EGY_NRC1 registered in the NCBI with accession number MW856655 and contained 1.1×10^9 CFU/g product evaluated at 1, 2 or 3 g/kg DM. ^bThe isolated product contained *E. faecium* NCIMB 11181 with a total viable count 2×10^{12} CFU/g (ADM Protexin Limited, Lopen Head, Somerset, TA13 5JH UK) evaluated at 1, 2 or 3 g/kg DM.

TABLE 3 Experiment 2: Intake, nutrient digestibility and nutritive value of diet supplemented with isolated and commercial *E. faecium* as probiotics and fed to lactating Holstein cows.

	Diet ^a			SEM	P-value		
	Control	Isolated	Commercial		Diet	Control vs. others	Isolated vs. Commercial
Intake, kg/cow/d	19.0	20.1	19.5	0.43	0.184	0.127	0.300
Digestibility, g absorbed/kg ingested							
Dry matter	586c	643a	632b	2.50	<0.001	<0.001	0.005
Organic matter (OM)	627b	676a	670a	3.00	<0.001	<0.001	0.121
Crude protein (CP)	603b	650a	635a	5.70	<0.001	<0.001	0.069
Ether extract	653b	702a	696a	5.90	0.001	0.003	0.631
Non-structural carbohydrates	684b	732a	723a	4.30	<0.001	<0.001	0.147
Neutral detergent fiber	566c	637a	615b	5.20	<0.001	<0.001	0.006
Acid detergent fiber	511c	582a	559b	5.20	<0.001	<0.001	0.006
Nutritive value							
TDN (g/kg DM) ^b	612c	666a	652b	2.70	<0.001	<0.001	0.001
DE (Mcal/kg DM) ^b	2.70c	2.94a	2.88b	0.01	<0.001	<0.001	0.009
ME (Mcal/kg DM) ^b	2.73c	2.97a	2.91b	0.01	<0.001	<0.001	0.002
NEL (Mcal/kg DM) ^b	1.38c	1.51a	1.48b	0.007	<0.001	<0.001	0.004
UFL (Mcal/kg DM) ^c	2.43c	2.66a	2.60b	0.01	<0.001	<0.001	0.007
Digestible OM intake, kg/cow/d	10.6b	12.1a	11.6a	0.35	0.004	0.002	0.345
TDN intake, kg/cow/d	11.6c	13.4a	12.7b	0.36	<0.001	<0.001	<0.001
ME intake, Mcal/cow/d	51.9c	59.7a	56.7b	1.44	<0.001	<0.001	<0.001
Digestible CP intake, kg/cow/d	1.81b	2.06a	1.95a	0.031	0.005	0.004	0.071

Means in the same row with different letters differ ($P < 0.05$); SEM, standard error of the mean. ^aDiet: Control diet contained 30% berseem clover + 30% corn silage + 15% soybean meal + 25% yellow corn, without additives, or supplemented with *E. faecium* EGY_NRC1 (isolated to contains 1.1×10^9 CFU/g) or commercial *E. faecium* NCIMB 11181 (isolated to contains 2×10^{12} CFU/g) at 2 g/kg feed daily/cow. ^bTDN, total digestible nutrients; DE, Digestible energy; ME, Metabolizable energy; NEL, Net energy for lactation. All have been calculated according to NRC equation. ^cUFL = unité fourragère du lait (net energy requirements for lactation equivalent of 1 kg of standard air-dry barley) calculated according to INRA equation.

TABLE 4 Experiment 2: Blood parameters of Holstein cows fed diets supplemented with isolated or commercial *E. faecium* as probiotics.

	Diet ^a			SEM	P-value		
	Control	Isolated	Commercial		Diet	Control vs. others	Isolated vs. Commercial
Total proteins, g/dL	9.58	9.93	9.96	0.15	0.193	0.073	0.897
Albumin, g/dL	5.35	5.60	5.52	0.14	0.480	0.252	0.715
Globulin, g/dL	4.23	4.33	4.43	0.13	0.589	0.385	0.590
Albumin: globulin ratio	1.29	1.32	1.27	0.06	0.833	0.943	0.553
Urea-N, mg/dL	78.9	76.8	81.3	1.54	0.144	0.917	0.052
Glucose, mg/dL	74.0b	81.5a	80.3a	1.39	0.002	0.004	0.533
Cholesterol, mg/dL	172a	152b	144b	5.20	0.002	0.007	0.313
Triglycerides, mg/dL	109a	101a	97b	1.30	0.002	0.002	0.034
AST, Units/L	35.3a	30.0b	33.0a	1.27	0.023	0.021	0.012
ALT, Units/L	23.5a	22.3a	20.6b	0.82	0.038	0.049	0.016

AST, Aspartate aminotransferase; ALT, Alanine aminotransferase. Means in the same row with different letters differ ($P < 0.05$); SEM, standard error of the mean. ^aDiet: Control diet contained 30% berseem clover + 30% corn silage + 15% soybean meal + 25% yellow corn, without additives, or supplemented with *E. faecium* EGY_NRC1 (isolated to contains 1.1×10^9 CFU/g) or commercial *E. faecium* NCIMB 11181 (isolated to contains 2×10^{12} CFU/g) at 2 g/kg feed daily/cow.

et al. (11) observed a lowered *in vitro* CH₄ production with LAB supplementation to a silage-based diet prepared with whole crop rice. To confirm our findings, further studies should consider analyzing rumen microbiome. The isolated

strain of *E. faecium* increased *in vitro* TGP and decreased CH₄ production compared to the commercial strain, with no clear reason indicating the need for experiments on genome sequence and their ability to produce bacteriocins

TABLE 5 Experiment 2: Milk production and composition, and feed efficiency of Holstein cows fed diets supplemented with isolated or commercial *E. faecium* as probiotics.

	Diet ^a			SEM	P-value		
	Control	Isolated	Commercial		Diet	Control vs. others	Isolated vs. Commercial
Production, kg/d							
Milk	30.6c	35.9a	34.2b	0.31	<0.001	<0.001	0.001
Energy corrected milk (ECM)	28.9c	35.0a	33.1b	0.42	<0.001	<0.001	0.002
4% Fat corrected milk (FCM)	28.5c	34.4a	32.5b	0.41	<0.001	<0.001	0.001
Total solids	3.76c	4.55a	4.33b	0.05	<0.001	<0.001	0.003
Solids non-fat	2.67c	3.21a	3.07b	0.03	<0.001	<0.001	0.006
Protein	1.01b	1.20a	1.15a	0.01	<0.001	<0.001	0.054
Fat	1.08c	1.34a	1.26b	0.02	<0.001	<0.001	0.006
Lactose	1.46c	1.76a	1.65b	0.02	<0.001	<0.001	0.001
Milk energy output, MJ/d	89.8c	109a	103b	1.32	<0.001	<0.001	0.001
Composition, g/kg DM							
Total solids	122	126	126	2.8	0.153		0.941
Solids non-fat	87.1	89.1	89.6	1.56	0.085	0.001	0.570
Protein	32.7	33.2	33.6	0.28	0.114	0.037	0.316
Fat	35.2	37.1	36.7	0.99	0.102	0.053	0.598
Lactose	47.8	48.9	48.3	0.36	0.070	0.444	0.200
Milk energy content, MJ/kg DM	2.92	3.03	3.01	0.22	0.255	0.069	0.589
Feed efficiency							
Milk: intake	1.61c	1.79a	1.75b	0.02	<0.001	<0.001	0.024
ECM: intake	1.52c	1.74a	1.70b	0.01	<0.001	<0.001	0.011
FCM: intake	1.50c	1.71a	1.67b	0.02	<0.001	<0.001	0.003

Means in the same row with different letters differ ($P < 0.05$); SEM, standard error of the mean. ^aDiet: Control diet contained 30% berseem clover + 30% corn silage + 15% soybean meal + 25% yellow corn, without additives, or supplemented with *E. faecium* EGY_NRC1 (isolated to contains 1.1×10^9 CFU/g) or commercial *E. faecium* NCIMB 11181 (isolated to contains 2×10^{12} CFU/g) at 2 g/kg feed daily/cow.

and non-ribosomal synthesized peptides for explaining such effects (36). Such possible differences between strains or their metabolites will produce different abilities to shift rumen fermentation patterns, and to inhibit specific rumen bacteria that produce H₂ or methyl-containing compounds that are the substrates for methanogenesis (36). Increasing TGP is not always advantageous but concurrent reduction in CH₄ is definitely advantageous (38). The improved fiber digestion is the most probable reason for the lowered CH₄ production (39).

The isolated and commercial strains increased dDM and dOM without affecting dNDF or dADF due to the high fermentative activities of LAB-probiotics. LAB can enhance the whole digestive process, the metabolic utilization of nutrients, and improve the feed efficiency by producing digestive enzymes (e.g., amylases, chitinases, lipases, phytases, proteases) or by just generating volatile fatty acids and B-vitamins: riboflavin, biotin, B₁₂ vitamin (40). Cao et al. (11) observed increased *in vitro* DM degradability with LAB administration to total mixed ration silage containing whole crop rice.

The commercial *E. faecium* increased ruminal NH₃-N concentration; however, the observed concentrations were

greater than the optimum level (8.5 to over 30 mg NH₃-N/dL) for rumen microbial proliferation (41). Basso et al. (10) observed no effects on ruminal pH when lambs were fed a diet treated with LAB. However, the isolated and commercial *E. faecium* decreased fermentation pH, which was somehow mirrored by the obtained SCFA. The isolated and commercial *E. faecium* increased SCFA concentrations, and this may be related to the improved DM and OM digestibility. So et al. (13) reported increased total SCFA in cows fed diets supplemented with LAB. The observed fermentation pH values in all treatments were greater than the optimum level (5.6) for ruminal fiber degrading and microbial growth (42), without changing ruminal fibrolytic and amylolytic microbial communities (43).

The quadratic effects of treatments (levels of *E. faecium*) on some parameters are important to emphasize the importance of defining the optimal dose of *E. faecium* that may improve animal performance. Therefore, the medium dose of *E. faecium* (2 g/kg feed) was recommended for the *in vivo* experiment on lactating cows, as this dose showed better effects compared to the low and high doses.

TABLE 6 Experiment 2: Milk fatty acid profile (g/100 g fatty acids) of lactating Holstein cows fed diets supplemented with isolated or commercial *E. faecium* as probiotics.

	Diet ^a			SEM	Diet	P-value	
	Control	Isolated	Commercial			Control vs. others	Isolated vs. Commercial
C4:0	0.81	0.81	0.75	0.017	0.138	0.247	0.089
C6:0	0.97	0.95	0.99	0.016	0.391	0.876	0.206
C8:0	0.93	0.90	0.93	0.017	0.400	0.587	0.238
C10:0	2.68	2.74	2.75	0.128	0.907	0.686	0.960
C11:0	0.20	0.20	0.19	0.010	0.866	0.942	0.625
C12:0	3.33	3.38	3.35	0.193	0.983	0.892	0.920
C13:0	0.46	0.46	0.47	0.015	0.895	0.705	0.826
C14:0	12.3	12.2	12.3	0.180	0.829	0.709	0.664
C15:0	1.07	1.10	1.08	0.015	0.524	0.466	0.408
C16:0	34.4	34.2	33.9	0.240	0.502	0.343	0.536
C17:0	0.66	0.63	0.65	0.078	0.974	0.866	0.900
C18:0	9.36	9.20	9.21	9.205	0.817	0.559	0.974
C20:0	0.12	0.12	0.12	0.010	0.928	0.721	1.000
C22:0	0.05	0.05	0.05	0.002	0.936	0.737	1.000
C23:0	0.04a	0.03b	0.03b	0.002	0.005	0.023	0.610
C24:0	0.04	0.04	0.04	0.009	0.494	0.550	0.329
∑ saturated fatty acids (SFA)	67.4	66.9	66.8	0.480	0.698	0.531	0.868
C14:1 cis-9	0.28	0.28	0.27	0.009	0.740	0.651	0.711
C14:1 trans-9	0.99	0.98	0.99	0.004	0.650	0.885	0.450
C16:1 cis-9	0.34	0.32	0.34	0.026	0.859	0.466	0.624
C16:1 trans-9	1.86	1.83	1.81	0.037	0.668	0.227	0.663
C18:1 cis-9	19.9	20.8	21.1	0.570	0.412	0.410	0.744
C18:1 trans-9	0.24b	0.26a	0.26a	0.007	0.017	0.037	1.000
C18:1 trans 11	0.97b	1.04a	1.00b	0.019	0.025	0.041	0.022
∑ monounsaturated fatty acids	24.5	25.5	25.7	0.570	0.405	0.505	0.781
C18:2 trans-9,12	1.76	1.76	1.72	0.084	0.911	0.938	0.730
C18:2 cis-9-12	0.17b	0.19a	0.18b	0.005	0.040	0.037	0.033
C18:2 cis-9 trans-11	0.40	0.41	0.40	0.024	0.986	0.405	0.892
C18:2 trans-10 cis-12	0.01b	0.02a	0.01b	0.002	0.029	0.036	0.033
C18:3 trans-9 cis 12,15	0.11	0.10	0.10	0.005	0.441	0.669	0.346
C18: 3 cis-9,12,15	0.40	0.39	0.40	0.026	0.954	0.829	0.806
C20:3 cis- 8,11,14	0.09	0.09	0.09	0.002	0.364	1.000	0.201
C20:4 cis-5,8,11,14	0.12	0.13	0.12	0.009	0.898	0.231	0.711
C20:5 cis- 5,8,11,14,17	0.03	0.03	0.03	0.006	0.604	0.442	0.353
C22:5, cis-7,10,13,16,19	0.17	0.16	0.16	0.004	0.354	0.265	0.450
∑ polyunsaturated fatty acids	3.25	3.26	3.20	0.075	0.818	0.220	0.587
∑ unsaturated fatty acids (UFA)	27.8	28.8	28.9	0.630	0.478	0.821	0.853
∑ conjugated linoleic acid ^b	0.41	0.42	0.41	0.024	0.969	0.882	0.860
UFA: SFA	0.41	0.43	0.43	0.012	0.493	0.276	0.848

Means in the same row with different letters differ ($P < 0.05$); SEM, standard error of the mean. ^aDiet: Control diet contained 30% berseem clover + 30% corn silage + 15% soybean meal + 25% yellow corn, without additives, or supplemented with *E. faecium* EGY_NRC1 (isolated to contains 1.1×10^9 CFU/g) or commercial *E. faecium* NCIMB 11181 (isolated to contains 2×10^{12} CFU/g) at 2 g/kg feed daily/cow. ^bC18:2 cis-9 trans-11 + C18:2 trans-10 cis-12.

Experiment 2 (lactation experiment)

It was not possible to obtain ruminal contents from cows because the experiment was done in a commercial farm without access to rumen-fistulated lactating cows. Therefore, the *in vitro* approach shows results that may partially explain and/or support the outcomes of the lactation experiment.

Feed intake, nutrient digestibility, and blood measurements

E. faecium supplementation did not affect feed intake which partly indicates unchanged feed palatability or acceptability. Other studies reported minor effects on feed intake in lambs and ewes fed with probiotics (8, 44), while So et al. (13) observed increased feed intake with LAB supplementation to lactating cows.

The isolated *E. faecium* improved the digestibility of DM, NDF and ADF compared to the commercial *E. faecium*; however, both isolated and commercial strains improved OM, CP, EE and NSC digestibility revealing the potential of *E. faecium* for improving nutrient digestibility. Similarly, So et al. (13) observed improved nutrient digestibility with LAB supplementation in lactating cows. Fiber degradability results are not consistent with those of the *in vitro* experiment (Experiment 1), which may be due to the conditions of both experiments (*in vitro* vs. *in vivo* conditions) and the fact that feeding bacterial direct fed microbial to livestock is based primarily on potential postruminal effects which is not available in the *in vitro* experiments (35). In this regard, probiotics change rumen fermentation rates and patterns (45), with beneficial effects on the gastrointestinal tract and rumen (46). Additionally, the supplement contains LAB which has a strong inhibitory effect on gastrointestinal infection by pathogens *via* the production of antimicrobial agents (46). It is expected that *E. faecium*, especially, the isolated strain, improved growth or activity of ruminal cellulolytic microbial populations and stabilizes the rumen pH (47), leading to improved nutrient digestion (45) and synthesis of microbial proteins (48). As previously noted, the isolated strain improved digestibility of DM, NDF, and ADF, and diet's nutritive value compared to the commercial strain, which confirm our assumption that the genome of both strains differs. The possible differed production of metabolites and bacteriocins may affect the composition of rumen microbiome, especially in those involved in fiber digestion (36). As observed in this study, previous experiments on lambs (49, 50), and lactating ewes (8) reported that probiotics improved nutrient digestibility.

All the measured serum biochemical parameters were within the standard physiological ranges for healthy cows (51). Treatments did not affect the concentrations of serum total protein, albumin, globulin, albumin: globulin ratio and urea-N indicating minimal effects on cow's nutritional status, muscle

protein catabolism and kidney function (52). Both *E. faecium* strains increased serum glucose because of improved apparent OM and NSC. The levels of serum glucose were above the range (40–60 mg/dL), indicating an adequate energy supply for cows without risk of negative energy balance occurring (53). Further studies should follow on these findings as *E. faecium* supplementation could be helpful during the transition period.

Both *E. faecium* strains decreased serum cholesterol while commercial *E. faecium* decrease serum triglycerides showing the ability of *E. faecium* bacteria to deconjugate bile salts by a specific hydrolase causing a reduction in cholesterol and triglycerides absorption at the intestinal level (54). Additionally, the commercial *E. faecium* decreased serum ALT, while isolated *E. faecium* decreased serum AST showing its potential to improve liver activity, function, and health in cows (55).

Milk production, milk composition, and milk fatty acids

In this study, both isolated and commercial *E. faecium* increased daily milk production by 17.1 and 11.7%, ECM by 21.4 and 14.8% and FCM 20.9 and 14.2%, respectively which is similar to other studies (8, 13, 44, 56) that reported a positive relationship between supplementation of ruminant diets with probiotics and animal performance. The improved nutrient digestibility and increased blood glucose with probiotics supplementation can be considered the main reasons for increasing milk production (44). An improvement in digestibility and intake of nutrients (ME and TDN) supported release of important nutrients required for milk components synthesis (41). Moreover, the improved feed efficiency with the additives is another probable reason for the improved performance (41). As previously noted in Experiment 1, *E. faecium* decreased CH₄ production indicating that a possible suppression in CH₄ production would have redistributed energy for improved milk production (57). In this study, the antagonism of pathogenic organisms *via* antimicrobial effects, competition for adhesion sites or nutrients, stimulation of host defines mechanisms and inhibition of bacterial toxins can partially explain the improved milk production (9, 45).

Moreover, LAB increases the release of different endogenous substances, including antibacterial substances, nutrients, antioxidants, growth factors and coagulating agents, enhances performance and reduces the incidence of diarrhea by increasing the number of beneficial microorganisms in the rumen (45, 58) and enhancing animal health (59), which could also explain the increased milk production. As a result of unchanged feed intake and increased daily milk production, the isolated and commercial *E. faecium* improved feed efficiency by 11.2 and 8.7% (milk: intake ratio), 14.5 and 11.8 (ECM: intake ratio), and 14 and 11.3% (FCM: intake ratio), respectively. Frizzo et al. (60) observed that supplementing diets of lactating cows with LAB including *E. faecium* improved feed efficiency.

The weak effect of treatment on the concentrations of milk components is inconsistent with other experiments (8) that reported some changes in the concentrations of total n-3, n-6 fatty acids, polyunsaturated fatty acids and conjugated linoleic acids components when sheep were fed diet supplemented with LAB. However, more experiments are required to explore these effects.

Plasma uptake of fatty acids is responsible for about half of milk fatty acids and the rest amount is synthesized in the mammary gland (61, 62). The improved fiber digestion with the supplementation might be associated with altered milk fatty acid profiles. Both *E. faecium* strains increased the proportion of C18:1 *trans*-9, while the isolated *E. faecium* increased the proportion of C18:1 *trans*-9, C18:2 *cis*-9-12 and C18:2 *trans*-10 *cis*-12. The observed changes in milk fatty acids are a result of biohydrogenation of dietary PUFA (63). Further attention should be paid to the use of *E. faecium* on cow diets as they may increase the formation of bioactive fatty acids such as C18:1 *trans*-11 and C18:2 *cis*-9-12.

Conclusion

Daily supplementation of cows with *E. faecium* (isolated and commercial) at 2 g/kg DM feed improved *in vitro* nutrient degradation and cows feed digestion, milk production and feed efficiency. The isolated strain of *E. faecium* showed better results compared to the commercial strain. Minimal effects were observed with *E. faecium* supplementation on milk fatty acid profile. Our data could be useful for producers looking for probiotics generated from byproducts for improving feed utilization and milk production.

Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue

reservation. The names of the repository/repositories and accession number(s) can be found below: NCBI accession number MW856655.

Ethics statement

The animal study was reviewed and approved by the Technical Committee of the Science, Technology & Innovation Funding Authority (STDF), Egypt (project STDF 33413).

Author contributions

HA, AK, and HM: conceptualization, methodology, validation, formal analysis, visualization, and supervision. AK and EV-B-P: writing—original draft preparation and writing—review and editing. HA and AK: investigation and data curation. All authors have read and agreed to the published version of the manuscript.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

1. Reuben RC, Elghandour MMY, Alqaisi O, Cone JW, Márquez O, Salem AZM. Influence of microbial probiotics on ruminant health and nutrition: sources, mode of action and implications. *J Sci Food Agric.* (2022) 102:1319–40. doi: 10.1002/jsfa.11643
2. Sun WT, Huang Y, Wu CR, Peng C, Zheng YL, Chen C, et al. Addition of lactic acid bacteria can promote the quality and feeding value of *broussonetia papyrifera* (paper mulberry) silage. *Fermentation.* (2022) 8:25. doi: 10.3390/fermentation8010025
3. Kholif AE, Gouda GA, Patra AK. The sustainable mitigation of *in vitro* ruminal biogas emissions by ensiling date palm leaves and rice straw with lactic acid bacteria and *Pleurotus ostreatus* for cleaner livestock production. *J Appl Microbiol.* (2022) 132:2925–39. doi: 10.1111/jam.15432
4. Kaewpila C, Gunun P, Kesorn P, Subepang S, Thip-uten S, Cai Y, et al. Improving ensiling characteristics by adding lactic acid bacteria modifies *in vitro* digestibility and methane production of forage-sorghum mixture silage. *Sci Rep.* (2021) 11:1968. doi: 10.1038/s41598-021-81505-z
5. Miranda C, Contente D, Igrejas G, Câmara SPA, Dapkevicius M de LE, Poeta P. Role of exposure to lactic acid bacteria from foods of animal origin in human health. *Foods.* (2021) 10:2092. doi: 10.3390/foods10092092
6. Ezema. Probiotics in animal production: a review. *J Vet Med Anim Health.* (2013) 5:308–16. doi: 10.5897/JVMAH2013.0201
7. Vibhute VM, Shelke RR, Chavan SD, Nage SP. Effect of probiotics supplementation on the performance of lactating crossbred cows. *Vet World.* (2011) 4:557–61. doi: 10.5455/vetworld.2011.557-561
8. Kholif AE, Hamdon HA, Gouda GA, Kassab AY, Morsy TA, Patra AK. Feeding date-palm leaves ensiled with fibrolytic enzymes or multi-species probiotics to Farafra ewes: intake, digestibility, ruminal fermentation, blood

- chemistry, milk production and milk fatty acid profile. *Animals*. (2022) 12:1107. doi: 10.3390/ani12091107
9. Elghandour MMY, Salem AZM, Castañeda JSM, Camacho LM, Kholif AE, Chagoyán JCV. Direct-fed microbes: a tool for improving the utilization of low quality roughages in ruminants. *J Integr Agric*. (2015) 14:526–33. doi: 10.1016/S2095-3119(14)60834-0
 10. Basso FC, Adesogan AT, Lara EC, Rabelo CHS, Berchielli TT, Teixeira IAMA, et al. Effects of feeding corn silage inoculated with microbial additives on the ruminal fermentation, microbial protein yield, and growth performance of lambs. *J Anim Sci*. (2014) 92:5640–50. doi: 10.2527/jas.2014-8258
 11. Cao Y, Takahashi T, Horiguchi KI, Yoshida N. Effect of adding lactic acid bacteria and molasses on fermentation quality and *in vitro* ruminal digestion of total mixed ration silage prepared with whole crop rice. *Grassl Sci*. (2010) 56:19–25. doi: 10.1111/j.1744-697X.2009.00168.x
 12. Xing L, Chen LJ, Han LJ. The effect of an inoculant and enzymes on fermentation and nutritive value of sorghum straw silages. *Bioresour Technol*. (2009) 100:488–91. doi: 10.1016/j.biortech.2008.06.017
 13. So S, Wanapat M, Cherdtong A. Effect of sugarcane bagasse as industrial by-products treated with *Lactobacillus casei* TH14, cellulase and molasses on feed utilization, ruminal ecology and milk production of mid-lactating Holstein Friesian cows. *J Sci Food Agric*. (2021) 101:4481–9. doi: 10.1002/jsfa.11087
 14. Mac Faddin JF. *Biochemical Tests for Identification of Medical Bacteria*. 3rd ed. Philadelphia, PA: Lippincott Williams & Wilkins (2000).
 15. Niven CF, Smiley KL, Sherman JM. The hydrolysis of arginine by Streptococci. *J Bacteriol*. (1942) 43:651–60. doi: 10.1128/jb.43.6.651-660.1942
 16. Kanak EK, Yilmaz SÖ. Maldi-tof mass spectrometry for the identification and detection of antimicrobial activity of lactic acid bacteria isolated from local cheeses. *Food Sci Technol*. (2019) 39:462–9. doi: 10.1590/fst.19418
 17. Liu W, Sun Z, Zhang J, Gao W, Wang W, Wu L, et al. Analysis of microbial composition in acid whey for dairy fan making in yunnan by conventional method and 16S rRNA sequencing. *Curr Microbiol*. (2009) 59:199–205. doi: 10.1007/s00284-009-9423-x
 18. Tamura K, Stecher G, Peterson D, Filipski A, Kumar S. MEGA6: molecular evolutionary genetics analysis version 6.0. *Mol Biol Evol*. (2013) 30:2725–9. doi: 10.1093/molbev/mst197
 19. Theodorou MK, Williams BA, Dhanoa MS, McAllan AB, France J. A simple gas production method using a pressure transducer to determine the fermentation kinetics of ruminant feeds. *Anim Feed Sci Technol*. (1994) 48:185–97. doi: 10.1016/0377-8401(94)90171-6
 20. Mauricio RM, Mould FL, Dhanoa MS, Owen E, Channa KS, Theodorou MK, et al. semi-automated *in vitro* gas production technique for ruminant feedstuff evaluation. *Anim Feed Sci Technol*. (1999) 79:321–30. doi: 10.1016/S0377-8401(99)00033-4
 21. Huo W, Wang X, Wei Z, Zhang H, Liu Q, Zhang S, et al. Effect of lactic acid bacteria on the ensiling characteristics and *in vitro* ruminal fermentation parameters of alfalfa silage. *Ital J Anim Sci*. (2021) 20:623–31. doi: 10.1080/1828051X.2021.1906167
 22. AOAC. *Official Methods of Analysis of AOAC International*. 18th ed. Washington, DC: AOAC International (2005).
 23. Warner AC. Production of volatile fatty acids in the rumen: methods of measurement. *Nutr Abstr Rev*. (1964) 34:339–52.
 24. NRC. *Nutrient Requirements of Dairy Cattle*. 7th ed. Washington, DC: National Academies Press (2001).
 25. Sales J, Janssens GPJ. Acid-insoluble ash as a marker in digestibility studies: a review. *J Anim Feed Sci*. (2003) 12:383–401. doi: 10.22358/jafs/67718/2003
 26. Ferret A, Plaixats J, Caja G, Gasa J, Prió P. Using markers to estimate apparent dry matter digestibility, faecal output and dry matter intake in dairy ewes fed Italian ryegrass hay or alfalfa hay. *Small Ruminant Research*. (1999) 33:145–52. doi: 10.1016/S0921-4488(99)00015-2
 27. INRA. *INRA Feeding System for Ruminants*. The Netherlands: Wageningen Academic Publishers (2018).
 28. Van Soest PJ, Robertson JB, Lewis BA. Methods for dietary fiber, neutral detergent fiber, and nonstarch polysaccharides in relation to animal nutrition. *J Dairy Sci*. (1991) 74:3583–97. doi: 10.3168/jds.S0022-0302(91)78551-2
 29. Tyrrell HF, Reid JT. Prediction of the energy value of cow's milk. *J Dairy Sci*. (1965) 48:1215–23. doi: 10.3168/jds.S0022-0302(65)88430-2
 30. Sjaunja LO, Baevre L, Junkkarinen L, Pedersen J, Setälä J. A nordic proposal for an energy corrected milk (ECM) formula: performance recording of animals. *State Art EAAP Publication*. (1991) 50:156–7.
 31. Murad HA, Hosseany EN, Abd Elhamid SM, Abu-El Khair AG, Zahran MO, Azzaz HH. Yeast biomass production from milk permeate with enrichment application of dairy animal's diets. *Inter J Dairy Sci*. (2020) 15:134–141. doi: 10.3923/ijds.2020.134.141
 32. Elghandour MMY, Kholif AE, Hernández A, Salem AZM, Mellado M, Odongo NE. Effects of organic acid salts on ruminal biogas production and fermentation kinetics of total mixed rations with different maize silage to concentrate ratios. *J Clean Prod*. (2017) 147:523–30. doi: 10.1016/j.jclepro.2017.01.078
 33. Blümmel M, Makkar HPS, Becker K. *In vitro* gas production: a technique revisited. *J Anim Physiol Anim Nutr*. (1997) 77:24–34. doi: 10.1111/j.1439-0396.1997.tb00734.x
 34. Pal K, Patra AK, Sahoo A, Kumawat PK. Evaluation of several tropical tree leaves for methane production potential, degradability and rumen fermentation *in vitro*. *Livest Sci*. (2015) 180:98–105. doi: 10.1016/j.livsci.2015.07.011
 35. Jiao PX, Liu FZ, Beauchemin KA, Yang WZ. Impact of strain and dose of lactic acid bacteria on *in vitro* ruminal fermentation with varying media pH levels and feed substrates. *Anim Feed Sci Technol*. (2017) 224:1–13. doi: 10.1016/j.anifeeds.2016.11.005
 36. Doyle N, Mbandlwa P, Kelly WJ, Attwood G, Li Y, Ross RP, et al. Use of lactic acid bacteria to reduce methane production in ruminants, a critical review. *Front Microbiol*. (2019) 10:2207. doi: 10.3389/fmicb.2019.02207
 37. Cao Y, Cai Y, Takahashi T, Yoshida N, Tohno M, Uegaki R, et al. Effect of lactic acid bacteria inoculant and beet pulp addition on fermentation characteristics and *in vitro* ruminal digestion of vegetable residue silage. *J Dairy Sci*. (2011) 94:3902–12. doi: 10.3168/jds.2010-3623
 38. Elghandour MMY, Kholif AE, Salem AZM, Olafadehan OA, Kholif AM. Sustainable anaerobic rumen methane and carbon dioxide productions from prickly pear cactus flour by organic acid salts addition. *J Clean Prod*. (2016) 139:1362–9. doi: 10.1016/j.jclepro.2016.08.075
 39. Elghandour MMY, Kholif AE, Salem AZM, Montes de Oca R, Barbabosa A, Mariezcurrera M, et al. Addressing sustainable ruminal methane and carbon dioxide emissions of soybean hulls by organic acid salts. *J Clean Prod*. (2016) 135:194–200. doi: 10.1016/j.jclepro.2016.06.081
 40. Vieco-Saiz N, Belguesmia Y, Raspoet R, Auclair E, Gancel F, Kempf I, et al. Benefits and inputs from lactic acid bacteria and their bacteriocins as alternatives to antibiotic growth promoters during food-animal production. *Front Microbiol*. (2019) 10:57. doi: 10.3389/fmicb.2019.00057
 41. Jones M, Jones G. *Animal Nutrition*. 7th ed. UK: Pearson Education Limited (2012). p. 64–83. doi: 10.1017/CBO9780511862793.008
 42. Ryle M, Ørskov ER. *Energy Nutrition in Ruminants*. Dordrecht: Springer Netherlands (1990). p. 149. doi: 10.1007/978-94-009-0751-5
 43. Tajima K, Aminov RI, Nagamine T, Matsui H, Nakamura M, Benno Y. Diet-dependent shifts in the bacterial population of the rumen revealed with real-time PCR. *Appl Environ Microbiol*. (2001) 67:2766–74. doi: 10.1128/AEM.67.6.2766-2774.2001
 44. Sallam SMA, Kholif AE, Amin KA, El-Din ANMN, Attia MFA, Matloup OH, et al. Effects of microbial feed additives on feed utilization and growth performance in growing Barki lambs fed diet based on peanut hay. *Anim Biotechnol*. (2020) 31:447–54. doi: 10.1080/10495398.2019.1616554
 45. Kassa SR. Role of probiotics in rumen fermentation and animal performance: a review. *Int J Livest Prod*. (2016) 7:24–32. doi: 10.5897/IJLP2016.0285
 46. Seo JK, Kim SW, Kim MH, Upadhaya SD, Kam DK, Ha JK. Direct-fed microbials for ruminant animals. *Asian-Australas J Anim Sci*. (2010) 23:1657–67. doi: 10.5713/ajas.2010.r.08
 47. Ghazanfar S, Anjum MI, Azim A, Ahmed I. Effects of dietary supplementation of yeast (*Saccharomyces cerevisiae*) culture on growth performance, blood parameters, nutrient digestibility and fecal flora of dairy heifers. *J Anim Plant Sci*. (2015) 25:53–9.
 48. Uyeno Y, Shigemori S, Shimamoto T. Effect of probiotics/prebiotics on cattle health and productivity. *Microbes Environ*. (2015) 30:126–32. doi: 10.1264/jsme2.ME14176
 49. Saleem AM, Zanouny AI, Singer AM. Growth performance, nutrients digestibility, and blood metabolites of lambs fed diets supplemented with probiotics during pre- and post-weaning period. *Asian-Australas J Anim Sci*. (2017) 30:523–30. doi: 10.5713/ajas.16.0691
 50. Hamdon HA, Kholif AE, Mahmoud GB, Khalifa AMA, Abdel Ati MNM. Enhancing the utilization of palm leaf hay using *Bacillus subtilis* and *Phanerochaete chrysosporium* in the diet of lambs under desert conditions. *Ann Anim Sci*. (2020) 20:1395–409. doi: 10.2478/aoas-2020-0052

51. Jerry Kaneko J, Harvey JJ, Bruss ML. *Clinical Biochemistry of Domestic Animals*. Elsevier (2008). doi: 10.1016/B978-0-12-370491-7.X0001-3
52. Hosten AO. BUN and Creatinine. In: Walker HK, Hall WD, Hurst JW, editors. *Clinical Methods: The History, Physical, and Laboratory Examinations*. Boston, MA: Butterworths (1990). p. 874–8.
53. Mair B, Drillich M, Klein-Jöbstl D, Kanz P, Borchardt S, Meyer L, et al. Glucose concentration in capillary blood of dairy cows obtained by a minimally invasive lancet technique and determined with three different hand-held devices. *BMC Vet Res*. (2016) 12: Article number: 34. doi: 10.1186/s12917-016-0662-3
54. Kumar M, Nagpal R, Kumar R, Hemalatha R, Verma V, et al. Cholesterol-lowering probiotics as potential biotherapeutics for metabolic diseases. *Exp Diabetes Res*. (2012). 2012:902917. doi: 10.1155/2012/902917
55. Pettersson J, Hindorf U, Persson P, Bengtsson T, Malmqvist U, Werkström V, et al. Muscular exercise can cause highly pathological liver function tests in healthy men. *Br J Clin Pharmacol*. (2008) 65:253–9. doi: 10.1111/j.1365-2125.2007.03001.x
56. Abd El Tawab AM, Kholif AE, Hassan AM, Matloup OH, El-Nor SAA, Olafadehan OA, et al. Feed utilization and lactational performance of Friesian cows fed beet tops silage treated with lactic acid bacteria as a replacement for corn silage. *Anim Biotechnol*. (2020) 31:473–82. doi: 10.1080/10495398.2019.1622556
57. Tseten T, Sanjorjo RA, Kwon M, Kim SW. Strategies to Mitigate Enteric Methane Emissions from Ruminant Animals. *J Microbiol Biotechnol*. (2022) 32:269–277. doi: 10.4014/jmb.2202.02019
58. Pant B, Lohani V, Mishra A, Trakroo MD, Tewari H. Effect of Probiotic Supplementation on Growth of Carp Fingerlings. *National Academy Science Letters*. (2019) 42:215–220. doi: 10.1007/s40009-018-0733-z
59. Bielke LR, Elwood AL, Donoghue DJ, Donoghue AM, Newberry LA, Neighbor NK, Hargis BM. Approach for selection of individual enteric bacteria for competitive exclusion in Turkey poults. *Poult Sci*. (2003) 82:1378–1382. doi: 10.1093/ps/82.9.1378
60. Frizzo LS, Zbrun M V., Soto LP, Signorini ML. Effects of probiotics on growth performance in young calves: A meta-analysis of randomized controlled trials. *Anim Feed Sci Technol*. (2011) 169:147–156. doi: 10.1016/j.anifeedsci.2011.06.009
61. Chilliard Y, Ferlay A. Dietary lipids and forages interactions on cow and goat milk fatty acid composition and sensory properties. *Reprod Nutr Dev*. (2004) 44:467–492. doi: 10.1051/rnd:2004052
62. Bionaz M, Vargas-Bello-Pérez E, Busato S. Advances in fatty acids nutrition in dairy cows: from gut to cells and effects on performance. *J Anim Sci Biotechnol*. (2020) 11. doi: 10.1186/s40104-020-00512-8
63. Kholif AE, Olafadehan OA. Dietary strategies to enrich milk with healthy fatty acids – A review. *Annals of Animal Science*. (2022) 22:523–536. doi: 10.2478/aoas-2021-0058