



Moringa oleifera Leaf Powder Dietary Inclusion Differentially Modulates the Antioxidant, Inflammatory, and Histopathological Responses of Normal and *Aeromonas hydrophila*-Infected Mono-Sex Nile Tilapia (*Oreochromis niloticus*)

OPEN ACCESS

Edited by:

Guillermo Tellez-Isaias,
University of Arkansas, United States

Reviewed by:

Abraham Mendez Albores,
National Autonomous University of
Mexico, Mexico
Alma Vazquez-Duran,
Universidad Nacional Autónoma de
México, Mexico

*Correspondence:

Seham El-Kassas
seham.elkassas@vet.kfs.edu.eg
Manal E. Shafi
meshafi@kau.edu.sa
Mohamed T. El-Saadony
m.talaatelsadony@gmail.com

Specialty section:

This article was submitted to
Veterinary Infectious Diseases,
a section of the journal
Frontiers in Veterinary Science

Received: 12 April 2022

Accepted: 18 May 2022

Published: 23 June 2022

Citation:

El-Kassas S, Aljhdali N, Abdo SE,
Alaryani FS, Moustafa EM,
Mohamed R, Abosheashaa W,
Abdulraouf E, Helal MA, Shafi ME,
El-Saadony MT, El-Naggar K and
Conte-Junior CA (2022)
Moringa oleifera Leaf Powder Dietary
Inclusion Differentially Modulates the
Antioxidant, Inflammatory, and
Histopathological Responses of
Normal and
Aeromonas hydrophila-Infected
Mono-Sex Nile Tilapia
(*Oreochromis niloticus*).
Front. Vet. Sci. 9:918933.
doi: 10.3389/fvets.2022.918933

Seham El-Kassas^{1*}, Nesreen Aljhdali², Safaa E. Abdo³, Fatima S. Alaryani⁴,
Eman M. Moustafa⁵, Radi Mohamed⁶, Wesam Abosheashaa⁷, Esraa Abdulraouf⁷,
Mohamed Atef Helal⁷, Manal E. Shafi^{8*}, Mohamed T. El-Saadony^{9*}, Karima El-Naggar¹⁰
and Carlos Adam Conte-Junior¹¹

¹ Animal, Poultry and Fish Breeding and Production, Department of Animal Wealth Development, Faculty of Veterinary Medicine, Kafrelsheikh University, Kafrelsheikh, Egypt, ² Department of Biological Science, College of Science, King Abdulaziz University, Jeddah, Saudi Arabia, ³ Genetics and Genetic Engineering, Department of Animal Wealth Development, Faculty of Veterinary Medicine, Kafrelsheikh University, Kafrelsheikh, Egypt, ⁴ Biology Department, Faculty of Sciences, University of Jeddah, Jeddah, Saudi Arabia, ⁵ Department of Fish Diseases and Management, Faculty of Veterinary Medicine, Kafrelsheikh University, Kafrelsheikh, Egypt, ⁶ Department of Aquaculture, Faculty of Aquatic and Fisheries Sciences, Kafrelsheikh University, Kafrelsheikh, Egypt, ⁷ Department of Animal Wealth Development, Faculty of Veterinary Medicine, Kafrelsheikh University, Kafrelsheikh, Egypt, ⁸ Department of Biological Science, Zoology, Faculty of Science, King Abdulaziz University, Jeddah, Saudi Arabia, ⁹ Department of Agricultural Microbiology, Faculty of Agriculture, Zagazig University, Zagazig, Egypt, ¹⁰ Department of Nutrition and Veterinary Clinical Nutrition, Faculty of Veterinary Medicine, Alexandria University, Alexandria, Egypt, ¹¹ Center for Food Analysis (NAL), Technological Development Support Laboratory (LADETEC), Federal University of Rio de Janeiro (UFRJ), Cidade Universitária, Rio de Janeiro, Brazil

This study aimed to detect the impact of *Moringa oleifera* leaf powder dietary inclusion on the antioxidant and innate immune responses of mono-sex Nile tilapia fingerlings. A total of 180 fingerlings were allocated in a random method into three groups with triplicate each. One group (1st group) received the control diet (basal diet (BD) free of moringa) and the other groups (2nd and 3rd) fed BD containing *M. oleifera* leaf powder at 5 and 10% of the diet, respectively. After 6 weeks of feeding, fish were randomly redistributed into four replicates and rested for 24 h. Then, each fish in the first two replicates was injected with 0.2 mL of PBS, while the others were injected with 0.2 mL of *A. hydrophila* suspension (1.8×10^6 CFU/mL). Healthy fish fed on *M. oleifera* leaf powder showed enhanced immune response manifested by significant increases in phagocytic and lysozyme activities with a marked H/L ratio ($P < 0.05$). In addition, significant alterations of the lymphocytic and heterophilic population in circulation with increasing infiltration in tissue such as the spleen were noticed. Also, *M. oleifera* significantly upregulated the antioxidants, CAT and GPx, proinflammatory cytokines, IL1- β , IL-8, and IFN- γ relative mRNA levels. On the other hand, following *A. hydrophila* challenging

conditions, *M. oleifera* caused downregulations of *IL1- β* , *IL-8*, and *IFN- γ* transcription levels, and also lowered the *CAT* and *GPx* mRNA levels. In addition, a marked reduction of leukocytic infiltration plus a significant improvement of the degenerative changes in intestinal architecture has occurred. So, *M. oleifera* leaf powder can be included in the fish diet to enhance immune response under normal health conditions and lower the infection-associated inflammatory response.

Keywords: *Moringa oleifera*, dietary additives, phagocytosis, lysozyme level, inflammatory response, antioxidant activities, *A. hydrophila* infection

INTRODUCTION

Intensive aquaculture is commonly associated with increasing exposure to pathogens (1), causing serious economic losses and zoonotic threats (2, 3). Most of the fish pathogens, especially the bacterial ones, are normal inhabitants. Under stressful conditions such as sudden water temperature changes and increasing stocking density produce diseases and mass mortalities (4). Competing for these diseases depends on several traditional strategies, such as antibiotics or vaccines. However, antibiotic use is now dwindling due to antibiotic resistance (5–7) and the health risk to human beings (8). Besides, using vaccines is neither cost-effective nor practically applicable (2).

Consequently, recent research focuses on finding new alternative strategies that could reduce or replace antibiotics, prevent and control fish diseases, enhance environmental protection, and are safe for human and animal health (9–11). One of these alternatives is phytochemical immune-stimulant products that improve fish resistance by boosting their innate defense and antioxidant mechanisms (12). One of these products is the medicinal plants (13), which are cheap and have effective compounds that enhance fish immune response (14). These plants can be utilized in whole, in sections (roots, leaves, or seeds), or as extracts through inclusion in diet or water (13). *Moringa oleifera* (*M. oleifera*) is one of these plants cultivated in tropical and subtropical regions with multifunctional applications in agriculture, medicine, industry, and human and animal nutrition (15).

Different parts of *M. oleifera*, especially its leaves, are of high nutritional value, rich in poly-unsaturated fatty acids (16), essential amino acids, vitamins, and carotenoids, as well as have a low content of antinutrients like phytates, oxalates, saponins, and tannins (17, 18) besides they induce several biological activities such as antioxidant, hypolipidemic and immunomodulatory properties (19). *M. oleifera* leaf powder or its extract can be supplemented at 0.5, 1, or 1.5% without modulating fish's growth performance despite maintaining fish health under normal and in case of infection or toxicity (20–24). Moreover, its supplementation can be safely increased to 10% of the *Clarias gariepinus* (25) and *Oreochromis niloticus* (26) feed and 25% of a *Tilapia rendalli* feed (27) without harming fish health, and the most growth-enhancing level was 5% with a hypolipidemic effect (28).

The anti-stress properties of *M. oleifera* to alleviate stress either due to infection (22, 29) or toxicity (23, 24) in Nile tilapia

have been proved to be mainly through modulating antioxidants response. Further, most studies were done on young fish (larvae) (22) and only investigated the antioxidant response. However, to our knowledge, the mechanism of how *M. oleifera* leaf powder modifies innate fish immunity at the transcriptomic level has not been investigated. So, the study aimed to explore how the dietary inclusion of *M. oleifera* leaf powder at 5 and 10% to mono-sex Nile tilapia fingerlings will modify its innate immune response under normal conditions and in case of *A. hydrophila* infection by examining phagocytic activity, lysozyme level, and transcriptional levels of some antioxidant and proinflammatory cytokines genes.

MATERIALS AND METHODS

Fish Management and Experimental Design

The present trial was adopted on mono-sex Nile tilapia (*Oreochromis niloticus*) following the standard operating steps approved by the animal care committee (KFS-2019/9).

O. niloticus fingerlings, with an average body weight of 27.98 \pm 0.75 g, $n = 180$, were acquired from a private farm. Fish was acclimatized for 14 days by feeding a commercial tilapia diet containing 30% Crude Protein with digestible energy of 4070 MJ/kg diet in glass aquaria (70 \times 30 \times 40 cm). An aerator and a multifunction internal filter (Shark SH-1000 Multi-Function Filter, China) were used for each glass aquarium. Following the acclimatization period, fish were allotted at random method into three different groups with triplicates each (20 fish/replicate). Groups' summary and descriptions are illustrated in **Table 1**. A control group (1st group) was fed a basal diet (BD) free of *Moringa oleifera* leaf powder. While the 2nd and 3rd groups were fed BD containing 5% and 10% of *M. oleifera* leaf powder, respectively.

M. oleifera leaves were thoroughly cleaned with water to get rid of dirt and dust, then air-dried at room temperature for 7 days or till complete drying. After that, a mechanical grinder ground moringa leaves into fine particles and stored them at 4°C until used in the diet formulation.

Diets (**Table 2**) were formulated and adjusted as indicated by (28). This feeding trial continued for 2 months. In the beginning, fish were supplied at a ratio equal to 4% of the body's weight, then decreased over the experiment until they reached 2.5 to 3% at the end of the feeding trial. The food was offered three times

TABLE 1 | Experimental design.

Experimental design		G1 (Fed basal diet)			G2 (Fed basal diet contained 5% Moringa oleifera leaf powder)			G3 (Fed basal diet contained 10% Moringa oleifera leaf powder)		
Groups	Replicates	R1	R2	R3	R1	R2	R3	R1	R2	R3
Feeding period										
Challenging with <i>A. hydrophila</i>		R1	R2	R3	R1	R2	R3	R1	R2	R3
Sampling		no	no	✓	no	no	✓	no	no	✓
		Samples were collected after 24 and 48 h post <i>A. hydrophila</i> challenging								

TABLE 2 | Ingredient's structure of the used trial feed.

Ingredients (%)	Diet 1 (C)	Diet 2 (T)	Diet 3 (M)
Corn oil	2.60	2.00	2.00
Wheat middling	8.25	7.30	7.30
Yellow corn	36.0	34.85	31.19
Soybean meal (44%)	28.0	26.0	27.0
Corn gluten meal (60%)	7.40	8.00	9.50
Carboxy methyl cellulose (CMC)	2.0	2.0	2.0
Moringa leaf powder		5.0	10.0
Fish meal(60%)	14.70	13.80	9.70
Mineral and vitamin premix (Premix)	0.30	0.30	0.30
CaHPO ₄	0.40	0.40	0.50
Methionine	0.05	0.05	0.10
Lysine -HCL	0.05	0.05	0.16
Choline chloride	0.05	0.05	0.05
Common salt	0.20	0.20	0.20
Calculated analysis (%)			
Moisture	10.22	10.08	10.04
Ether extract	6.10	5.90	5.82
Crude fiber	3.80	4.10	4.19
Crude protein	30.0	30.05	30.04
Ash	6.20	6.10	5.90
Lysine	1.76	1.73	1.75
Methionine	0.68	0.67	0.68
Calcium	1.00	1.07	1.02
Phosphorus	0.88	0.85	0.77
NFE*	43.62	43.77	44.01
Digestible energy (DE Kcal/kg)**	2927.85	2934.90	2957.79

CaHPO₄, contain Ca 21% and P 18%. Mineral and vitamin premix obtained from Allegaevet Co., Egypt, each kg includes: Fat soluble vitamins; vit K3 (3.333 mg), vit E (16.666 mg), vit D (666.666 IU), vit. A (3.333.333 IU); water soluble vitamins, vit B1 (3.333 mg), vit B2 (6.666 mg), vit B6 (5000 mg), vit B12 (6.6 mg), nicotinic acid (20.000 mg), pantothenic acid (13.333 mg), folic acid (1.666 mg), biotin (16.6 mg), elements, Fe (33.333 mg), Zn (16.666 mg), Mn (8.333 mg), Cu (5000 mg, l (100 mg), Se (100 mg), Co (66.6 mg), and CaCO₃ as a carrier. Nitrogen-free extract (NFE), ** Digestible energy, Carbohydrate (3.48 Kcal/g), Ether extract (8.5 kcal/g), Protein (4.49 Kcal/g), as (30). Tilapia diet supplemented with Moringa powder at 0, 5, and 10% levels were C, T, and M, diets, respectively.

daily and the amount was changed every 2 weeks depending on the biomass in each aquarium. The aquarium water was partially (about 25–30%) changed with overnight stored water (dechlorinated water) 2 to 3 periods per week. Water quality parameters such as water temperature and dissolved oxygen (DO), total ammonia (NH₃), total dissolved salts (TDS), pH, and electrical conductivity (EC) were determined based on methods previously used by El-Kassas et al. (28). Growth performance parameters and body indices were measured (Table 3) and published previously (28). However, they were included again in this study depending on a Copywrite agreement from Elsevier (License Number: 4983960289771).

Challenging With *Aeromonas hydrophila*

At the end of the feeding trial, fish in every group was redistributed in a random method and equally into four replicates (after confirming the non-statistical significance of replicating

TABLE 3 | A summary of growth and water quality parameters (28).

	C	T	M	P-value
Initial body weight (g)	27.68 ^a ± 0.74	27.84 ^a ± 0.77	28.42 ^a ± 0.80	0.7739
Final body weight (g)	50.53 ^b ± 2.74	61.00 ^a ± 2.52	48.67 ^b ± 2.59	0.0013
Final body weight gain (g)	20.13 ^b ± 2.57	33.16 ^a ± 2.47	21.38 ^b ± 2.62	0.0009
Feed intake/fish (g)	60.70 ^a ± 1.73	56.74 ^a ± 2.29	53.15 ^b ± 1.22	0.0497
Feed conversion ratio (FCR)	2.59 ^a ± 0.34	1.81 ^b ± 0.11	2.84 ^a ± 0.20	0.0021
Specific growth rate (SGR)	3.03 ^b ± 0.11	3.35 ^a ± 0.06	2.83 ^b ± 0.09	0.0006
Body length (cm)	14.69 ^b ± 0.30	15.53 ^a ± 0.21	14.62 ^b ± 0.25	0.0210
Body width (cm)	4.93 ^b ± 1.48	7.58 ^a ± 0.66	6.00 ^a ± 0.87	0.0411
Ammonia (mg/L)	0.052 ^a ± 0.001	0.048 ^b ± 0.002	0.0515 ^a ± 0.006	<0.0001
DO (mg/L)	4.76 ^b ± 0.10	5.24 ^a ± 0.20	5.34 ^a ± 0.14	0.042
pH	7.87 ^a ± 0.029	7.87 ^a ± 0.036	7.95 ^a ± 0.008	0.703
Temperature (°C)	27.13 ^a ± 0.118	27.20 ^a ± 0.107	27.06 ^a ± 0.230	0.824
TDS (mg/L)	7.87 ^a ± 0.029	7.87 ^a ± 0.036	7.95 ^a ± 0.008	0.703
EC (µS/L)	778.7 ^a ± 16.24	754.5 ^a ± 2.91	763.1 ^a ± 10.71	0.372

C, T, and M, refer to diets used in this feeding trial. Moringa leaf powder was included at 0, 5, and 10% of C, T, and M diets, respectively. TDS, Total dissolved salts; EC, Electric conductivity; DO, dissolved oxygen. The results are expressed as means ± SEM (n = 3). Different superscript letters within the same row denote significant difference at p < 0.05 using one-way ANOVA.

the effect, $P > 0.05$) and rested for 24 h to get rid of distribution's stress. Thus, fish in each treatment group with first and second replicates was injected with phosphate buffer saline (PBS). In contrast, the 3rd and 4th replicates infected with *Aeromonas hydrophila* strain were obtained. The severity of *A. hydrophila* was monitored routinely (31). *A. hydrophila* was cultured on tryptic soy broth (TSB) at 37 °C for 24 h with constant shaking (250 rpm) to obtain an inoculum of 1.8×10^6 CFU/mL. The bacterial count was enumerated by standard dilution and plating assay. The median lethal dose (LD50) that kills 50% of injected fish was evaluated before the terminal injection challenge following (32). The challenge was conducted by intra-peritoneal injection of each fish with 0.2 mL fresh growth of *A. hydrophila* inoculum following (33). However, fishes in control negative ones were intra-peritoneally injected with 0.2 mL/fish of PBS. Mortalities were recorded daily.

Sample Collection

Random fish samples (6 fish per aquarium) from each group after 24 and 48 h post-challenge were subjected to blood and tissue analysis. Fish were first mildly anesthetized with 100 mg/L of tricaine methanesulfonate (MS-222). Then, two blood samples (heparinized and non-heparinized) were collected through venipuncture of the caudal vein. The blood samples collected in heparinized syringes were used for measuring the differential leukocytic count and phagocytic activity. While blood collected without anticoagulant was used for serum separation (28) and was kept at -20°C for further analysis. Following blood collection, two spleen samples were collected from dissected fish. One sample was employed for qRT-PCR assay where it was kept in a sterile microcentrifuge tube, rapidly frozen in liquid nitrogen, and stored at -80°C for total RNA isolation. While the 2nd spleen specimen was used for histopathological examination; thus, it was collected in a 10% neutral-buffered formalin

fixative solution. In addition, samples from the mid-gut were collected, fixed overnight in a Bouin's solution, and employed for histopathological evaluation of intestinal morphometry. Lesion scoring analysis of lesions in the spleen and mid-gut was done according to (34, 35) with some modification in the scoring values. In addition, the number of goblet cells as an indicator of mucous production in the intestine was counted in five microscopic fields using a light microscope at a higher magnification. The goblet cells were recognized depending on whether they appeared white or clear when stained with H&E and are distributed in the epithelia of the intestine (36).

Immunological Parameter Assessment

The differential leukocytic count was assessed following the method of Abo-Al-Ela et al. (37). Blood smears were briefly fixed with methanol and stained by rapid field stains (polychrome methylene blue and eosin). Then, it was rinsed with distilled water and subjected to air-dry. The blood film was examined using a computer-assisted light microscope under an oil immersion lens (×100). The number of heterophils and lymphocytes per every 100 leucocytes was counted and used to calculate the heterophils to lymphocytes (H/L) ratio.

Leucocyte phagocytic activity to *Candida albicans* was adopted *in vitro* (37). An aliquot of 100 µl of fresh blood sample was mixed with 100 µl of both *C. albicans* inoculum (1×10^6 CFU/mL) and fetal bovine serum, then kept at 37°C for 30 min, afterward centrifuged at 1,500 rpm for 10 min. The blood smear was prepared with 5 µL of the resuspended cells. The phagocytic activity (PA) was estimated as the percentage of phagocytic cells which overwhelmed yeast cells. At the same time, the phagocytic index (PI) equaled the total number of phagocytized yeast cells classified by the count of phagocytic cells.

Measuring serum lysozyme activity depended on the agarose plate diffusion assay previously mentioned by Lie et al. (38). In

TABLE 4 | Primer sequences.

Gene	Primer	Reference
β -actin	F: CAGCAAGCAGGAGTACGATGAG R: TGTGTGGTGTGTGGTTGTTTTG	Dawood et al. (39)
CAT	F: CCCAGCTCTTCATCCAGAAAC R: GCCTCCGCATTGTACTTCTT	
GPx	F: CCAAGAGAAGTCAAGAACGA R: CAGGACACGTCATTCTACAC	
IL-1 β	F: CAAGGATGACGACAAGCCAACC R: AGCGGACAGACATGAGAGTGC	
IL-8	F: GCACTGCCGCTGCATTAAG R: GCAGTGGGAGTTGGGAAGAA	Abo-Al-Ela et al. (37)
IFN- γ	F: AAGAATCGCAGCTCTGCACCAT R: GTGTCGTATTGCTGTGGCTTCC	

brief, this method aimed to measure the diameter of clear lysed zones developed by different serum samples on 1% agarose gel containing a suspension of *Micrococcus lysodeikticus* cells (Sigma-Aldrich, 500 mg/L) and then compared to those formed by the standard (hen egg-white lysozyme solution, 20 mg/mL). In this regard, serum samples (25 μ L) were inoculated in the agarose gel plate (1%) in a 50-mM phosphate buffer (pH 6.3). The plates were then closed tightly and kept at 37°C for 18 h. Serum lysozyme activity was analyzed using a logarithmic regression analysis according to this formula: $Y = A + \text{Blog } X$, where Y = diameter of the lysed zone and X = the lysozyme activity expressed in μ g/mL.

qRT-PCR Analysis of Some Antioxidant and Proinflammatory Cytokine Genes

According to the manufacturer's guidelines, total RNA was extracted from 50 mg of spleen tissues by Trizol (iNtRON Biotechnology, Inc.). The RNA integrity was confirmed by ethidium bromide-stained 2% agarose gel electrophoresis, while the concentration was estimated by Nanodrop (Uv-Vis spectrophotometer Q5000/Quawell, USA). Two micrograms of each RNA sample were utilized for cDNA synthesis by SensiFAST™ kit (Bioline, UK) according to the manufacturer's manual.

Real-time PCR was then performed to evaluate the relative mRNA transcriptomic profile for some antioxidant genes; catalase (CAT) and glutathione peroxidase (GPX) and some proinflammatory cytokines genes; interleukin 8 (IL-8), interleukin 1 β (IL-1 β), and interferon-gamma (IFN- γ), using specific primers (Table 4), in a Stratagene MX3000 system (Agilent Technologies), and SensiFast SYBR Low-Rox kit (Bioline, UK). PCR mixture preparations and the amplification conditions were done for each gene according to Dawood et al. (39). All genes tests were done in duplicates. CT values for each sample were estimated and combined in fold change ($2^{-\Delta\Delta CT}$) calculation according to Livak and Schmittgen method (40). In this regard, the relative transcription levels were normalized against the housekeeping gene (β -actin) and the corresponding values of the control negative group (fed BD free of moringa leaf powder and injected with PBS).

Statistical Analysis

The results were statistically analyzed by GLM procedure in IBM SPSS Statistics for Windows (SPSS version 22, SPSS Inc., IL, USA) at $P < 0.05$. Before analysis, data were confirmed for normality and homogeneity of variance among variables by the Shapiro-Wilk and Levene test, respectively. Then, the results were subjected to three-way ANOVA to test the impacts of *M. oleifera* feed inclusion, challenge with *A. hydrophila*, and time of samples collected and their interactions. Using Tukey's HSD test, multiple comparisons were performed to determine the statistical significance. The results were expressed as means \pm SEM. GraphPad Prism 9 statistical software (GraphPrism Software, La Jolla, California, USA) was employed to draw figures.

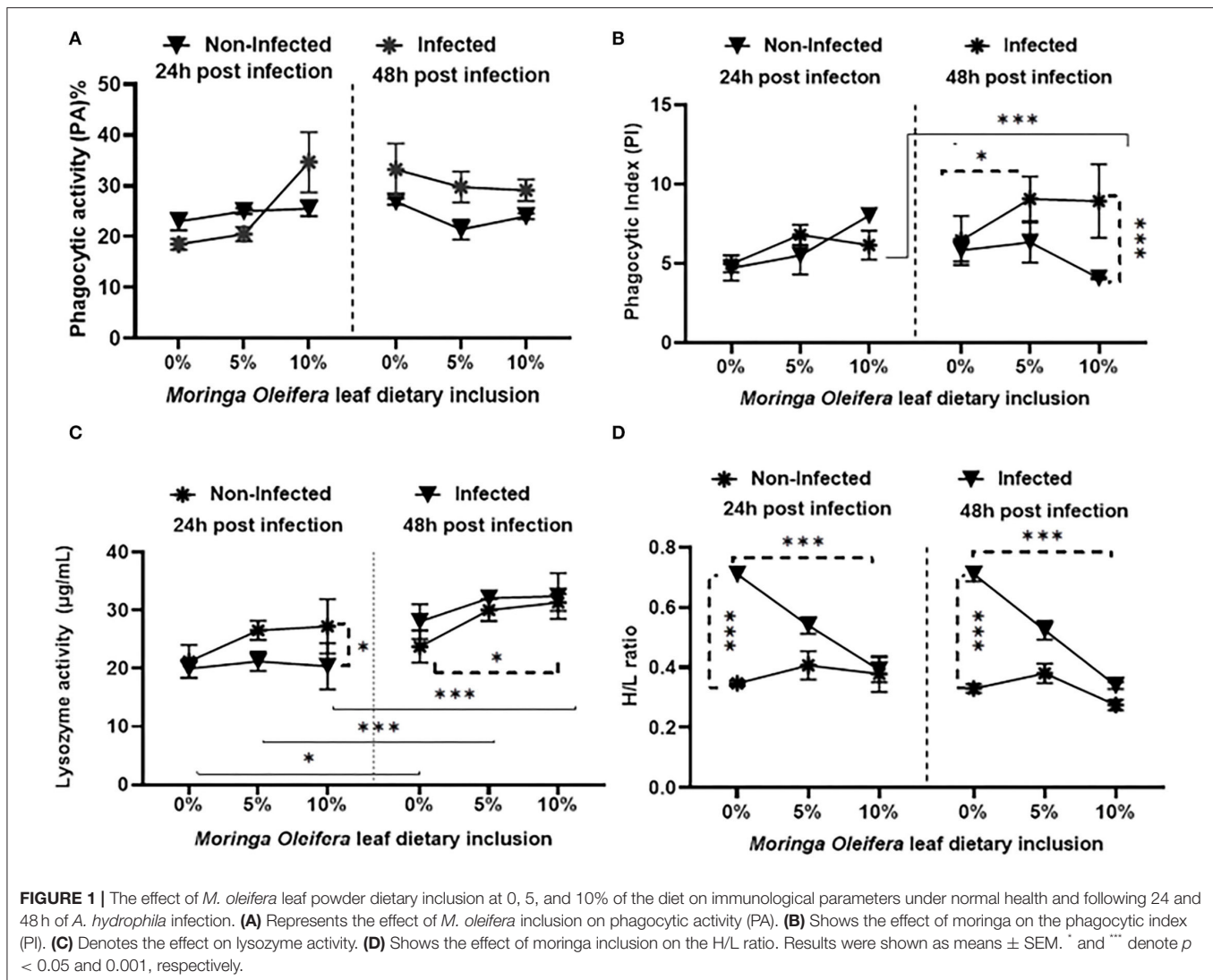
RESULTS

Impact of *M. oleifera* Leaf Powder Dietary Inclusion on Immunological Parameters

Feeding on *M. oleifera* contained a diet that did not modify phagocytic activity (PA) (Figure 1A) either under normal or following *A. hydrophila* challenging conditions ($P > 0.05$). Only slight non-significant increases and decreases were measured in the case of 5% and 10% *M. oleifera* containing diet after 24 and 48 h post-challenge, respectively. On the other side, dietary feeding of *M. oleifera* to mono-sex Nile tilapia, challenging with *A. hydrophila*, time of sampling post challenge, and their interactions resulted in significant changes in the phagocytic index (PI) (Figure 1B) ($P < 0.05$). After 24 h post-challenge, no changes were noticed ($P > 0.05$). However, after 48 h, healthy fish fed on *M. oleifera*-contained diet (especially at 10%) showed a significant reduction of PI compared to those infected with *A. hydrophila* and fed on *M. oleifera*-contained diet ($P < 0.05$). In addition, among infected fish, a significant increase of PI was noticed in the case of those fed on a 5% *M. oleifera* compared to moringa-free diet ($p < 0.05$). Besides, increasing exposure to *A. hydrophila* resulted in a marked increase in PI ($p < 0.05$).

Feeding *M. oleifera* and exposure time to *A. hydrophila* resulted in a significant change in lysozyme activity (LZ) (Figure 1C) of mono-sex Nile tilapia ($p < 0.05$). Where, under normal health conditions, dietary inclusion of *M. oleifera*, especially at 10%, induced a significant increase in LZ activity ($p < 0.05$). LZ activity was significantly higher in infected fish than in the non-infected, especially 24 h after the challenge ($p < 0.05$). In addition, increasing the exposure time of mono-sex Nile tilapia to *A. hydrophila* significantly stimulated higher LZ activities in *M. oleifera* fed and non-fed groups ($p < 0.05$).

The effect of *M. oleifera* leaf powder dietary inclusion to mono-sex Nile tilapia on its differential leukocytic count and heterophil to lymphocyte (H/L) ratio is illustrated in Table 5 and Figure 1D, respectively. Healthy tilapias that were fed a moringa-containing diet exhibited significant increases in the lymphocytic count, with the highest count observed in the 10% moringa fed group ($p < 0.05$). In contrast, significant reductions in heterophils' count were recorded ($p < 0.05$). These changes in heterophils and lymphocyte count were associated with a significant reduction in the H/L ratio, which elevated with increasing moringa inclusion level in the diet ($p < 0.05$). In



addition, Nile tilapias supplied with an *M. oleifera* leaf powder containing diet showed changes in the differential leukocytic count and H/L ratio following *A. hydrophila* infection. No changes were detected after 24 h of tilapia's challenge with *A. hydrophila* ($p > 0.05$). However, after 48 h of *A. hydrophila* exposure, a significant elevation was documented for the lymphocytic count in the case of the 10% moringa fed group ($p < 0.05$). At the same time, the heterophils count increased in the case of 5% moringa-fed fish ($p < 0.05$). These changes were also linked with lowering the H/L ratio.

CAT and GPx MRNA Transcripts Following *M. oleifera* Leaf Powder Dietary Inclusion and *A. hydrophila* Challenge

Feeding moringa having a diet to mono-sex Nile tilapia modified the relative expression levels of some antioxidant genes such as CAT and GPx under normal health conditions

and the infection with *A. hydrophila* after 24 and 48 h of exposure (Figure 2). Under normal health status, moringa leaf powder inclusion at 5 and 10% enhanced a significant upregulation of CAT relative mRNA level ($P < 0.05$). For GPx mRNA transcripts, feeding moringa leaf powder to healthy fish also stimulated higher levels and increased time ($p < 0.05$). On the other side, following infection with *A. hydrophila*, significant upregulations of both CAT and GPx were measured in the tilapias supplied with a moringa-free diet (0% moringa) ($p < 0.05$). The infection-induced upregulations were interestingly decreased in the case of moringa-fed groups, with the maximum reduction noticed in the 10% moringa and 48 h post-infection except for CAT which showed a slight increase after 48 h ($p < 0.05$). Therefore, *M. oleifera* feeding to mono-sex Nile tilapia stimulated a higher antioxidant response under normal health conditions, which decreased in case of infection.

TABLE 5 | Differential leukocytic counts (%) of mono-sex Nile tilapia supplied with *Moringa oleifera* leaf including diet.

	24 h post infection					24 h non-infected				
	Lymphocytes	Heterophils	Monocytes	Eosinophils	Basophils	Lymphocytes	Heterophils	Monocytes	Eosinophils	Basophils
C	69.6 ± 0.81 ^{Aa}	24.1 ± 0.79 ^{Ba}	3.9 ± 0.06	1.6 ± 0.06	0.8 ± 0.09	55.0 ± 1.45 ^{Bb}	39.0 ± 1.35 ^{Aa}	3.7 ± 0.10	1.6 ± 0.05	0.8 ± 0.05
T	66.8 ± 2.33 ^{Aa}	26.6 ± 2.28 ^{Ba}	4.3 ± 0.05	1.6 ± 0.04	0.7 ± 0.07	60.9 ± 0.96 ^{Ba}	32.8 ± 1.16 ^{Ab}	3.9 ± 0.03	1.4 ± 0.06	0.9 ± 0.20
M	68.3 ± 2.60 ^{Aa}	25.0 ± 2.59 ^{Aa}	4.5 ± 0.04	1.7 ± 0.03	0.6 ± 0.04	67.5 ± 1.86 ^{Aa}	26.3 ± 1.99 ^{Ac}	3.9 ± 0.03	1.5 ± 0.03	0.8 ± 0.08
	48 h post infection					48 h non-infected				
	Lymphocytes	Heterophils	Monocytes	Eosinophils	Basophils	Lymphocytes	Heterophils	Monocytes	Eosinophils	Basophils
C	69.7 ± 1.18 ^{Ab}	22.9 ± 1.16 ^{Bb}	4.8 ± 0.09	1.7 ± 0.08	0.9 ± 0.09	54.8 ± 1.40 ^{Bc}	38.9 ± 1.40 ^{Aa}	3.9 ± 0.05	1.6 ± 0.00	0.8 ± 0.05
T	67.5 ± 1.52 ^{Ab}	25.4 ± 1.56 ^{Ba}	4.8 ± 0.08	1.7 ± 0.04	0.7 ± 0.05	61.5 ± 1.08 ^{Bb}	32.1 ± 1.33 ^{Ab}	3.9 ± 0.03	1.5 ± 0.03	1.0 ± 0.24
M	72.7 ± 1.01 ^{Aa}	19.9 ± 1.00 ^{Bb}	5.1 ± 0.06	1.7 ± 0.03	0.6 ± 0.04	70.0 ± 0.62 ^{Aa}	23.7 ± 0.62 ^{Ac}	3.9 ± 0.06	1.6 ± 0.07	0.8 ± 0.03

C, T, and M refer to diets used in this feeding trail. Moringa leaf powder was included at 0, 5, and 10% of C, T, and M diets, respectively. Data were analyzed using three-way ANOVA test and the results are expressed as means ± SEM (n = 6 fishes/replicate). Different lowercase letters in the same column indicate the differences between various *Moringa oleifera* inclusion levels at $p < 0.05$. Different uppercase letters within the same row indicate statistical significance between infected and non-infected treatments at $P < 0.05$.

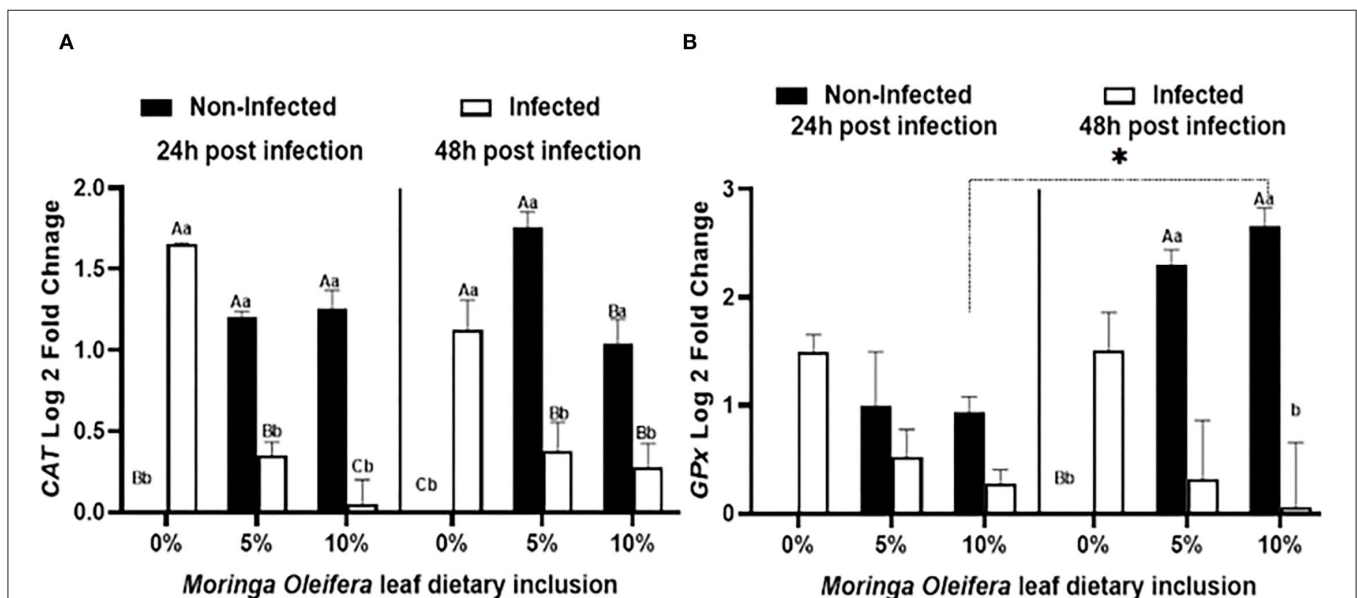
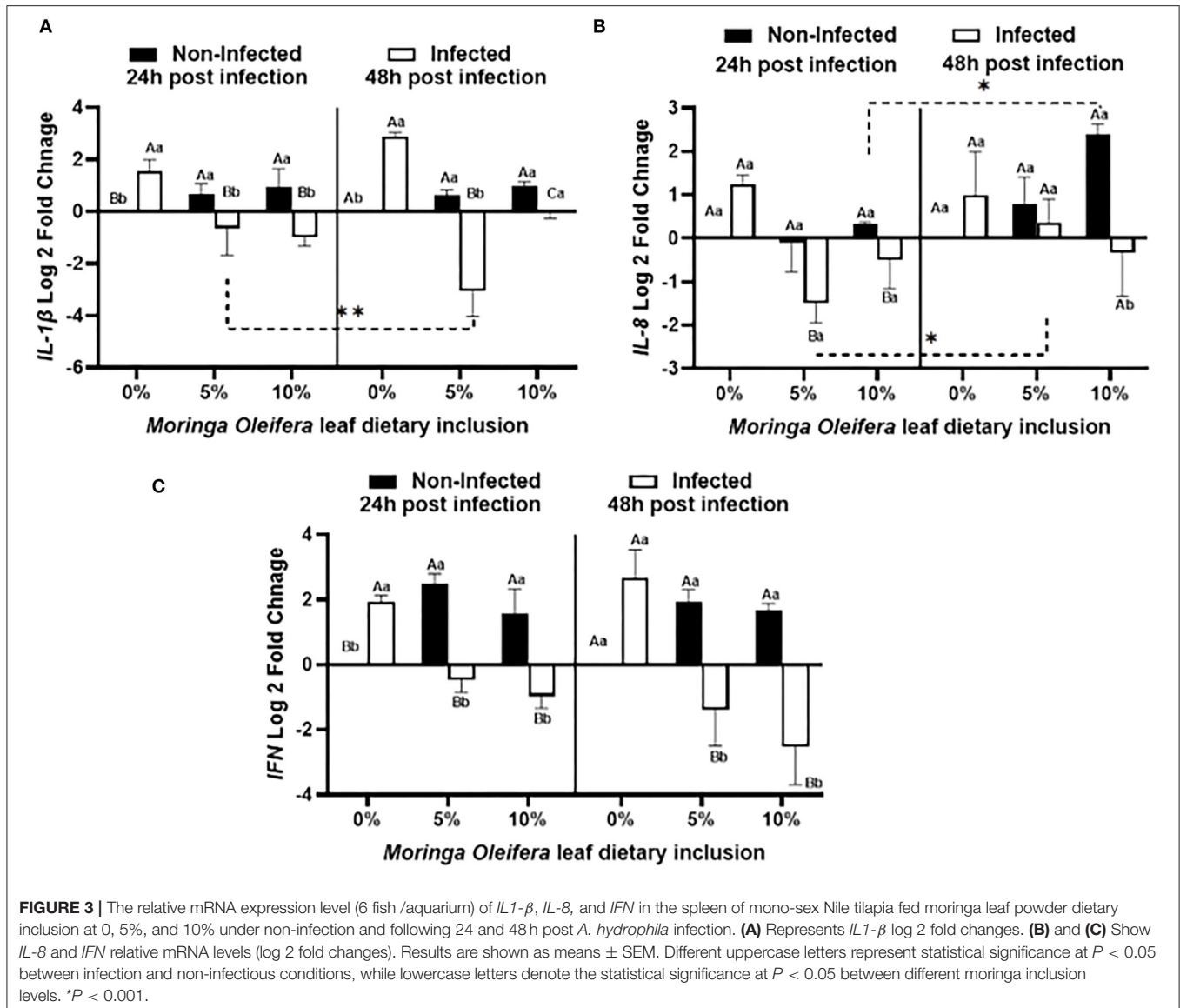


FIGURE 2 | The relative mRNA expression level (6 fish/aquarium) of *CAT* and *GPx* in the spleen of mono-sex Nile tilapia fed dietary inclusion of moringa leaf powder at 0, 5, and 10% under non-infection and following 24 and 48 h post *A. hydrophila* infection. **(A)** Represents *CAT* log 2 fold changes and **(B)** shows *GPx* relative mRNA level (log 2 fold changes). Results are shown as means ± SEM. Different uppercase letters represent statistical significance at $P < 0.05$ between infection and non-infection conditions, while lowercase letters denote the statistical significance at $P < 0.05$ between different moringa inclusion levels. * $P < 0.05$.

Proinflammatory Cytokines mRNA Transcripts' Response Following *M. oleifera* Dietary Inclusion and *A. hydrophila* Infection

Dietary inclusion of *Moringa oleifera* induced significantly differential modulations of the relative mRNA expression levels of proinflammatory cytokines such as *IL-1 β* , *IL-8*, and *IFN- γ* between normal health and infection conditions (Figure 3). For *IL-1 β* (Figure 3A), normal healthy tilapias fed *M. oleifera* at 5% and 10% inclusion levels exhibited significant upregulations

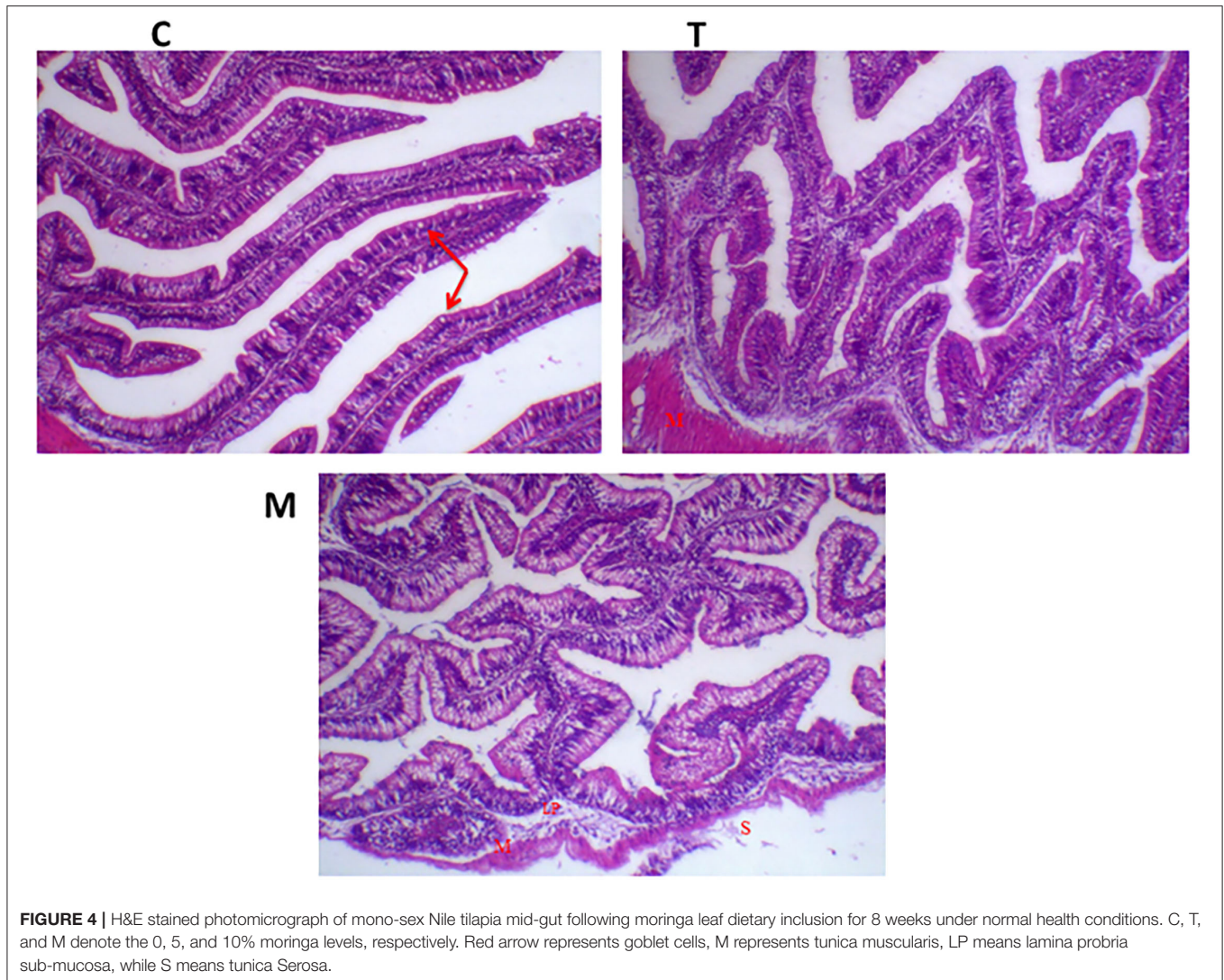
of *IL-1 β* in comparison with those supplied the BD ($P < 0.05$). However, in the case of *A. hydrophila* infection, fish fed moringa free diet showed a marked increased level of *IL-1 β* . This effect was diversely changed in the case of fish fed on a moringa-containing diet and infected with *A. hydrophila* ($p < 0.05$). This downregulation increased with increasing the exposure time to *A. hydrophila*, especially in the case of the moringa 5% fed tilapias ($P < 0.05$). In addition, *IL-8* mRNA transcripts (Figure 3B) were significantly modified where, under normal conditions, moringa inclusion in the diet caused only slight increases of *IL-8* mRNA copies, which increased over



time, especially in the case of the 10% moringa-fed group ($P < 0.05$). *A. hydrophila* infection caused a significant upregulation of *IL-8* expression levels in the absence of *M. oleifera* in the diet. This response was significantly downregulated in the case of moringa-fed groups ($P < 0.05$). In addition, this down-regulatory effect decreased over time, especially in the case of 5% moringa ($P < 0.05$). Likewise, *INF-γ* expression level (Figure 3C) was distinctly changed. Under normal health conditions, significant upregulations were reported in the case of moringa-fed groups compared to fish fed on BD free of moringa ($P < 0.05$). However, when fish fed on moringa-containing diet and infected with *A. hydrophila*, they displayed a significant downregulation to *IFN-g* relative mRNA level, which was distinctly increased following infection in the fish supplied with BD ($p < 0.05$).

Histopathological Response of Mono-Sex Nile Tilapia Following *M. oleifera* Dietary Inclusion and *A. hydrophila* Infection

Histopathological changes in mid-gut from different groups were illustrated in Figures 4, 5 and Tables 6, 7. Under normal health conditions (Figure 4), fish fed the basal diet with 0% moringa showed normal and thin villi with intact four layers of mid-gut; tunica mucosa, lamina propria sub-mucosa, tunica muscularis, and tunica serosa. Dietary inclusion of *M. oleifera* increased villi length and branches of mid-gut with normal villi architecture (Table 6). In this regard, a significant dose-dependent increase in villi length and width was noticed, with the highest values reported 10% of moringa leaf powder. However, moringa leaf powder inclusion did not alter the inter villi space, with no pathological lesions noticed (0 scores). Infection



with *A. hydrophila* caused marked degenerative changes in the mid-gut (**Figure 5**). After 24 h post-infection, clear blunting of intestinal villi (yellow arrow) was seen. In addition, increasing exposure to *A. hydrophila* (48 h post-infection) decreased villi length and caused detach of the apical part of enterocytes (yellow arrowhead) with leukocytic infiltration (star). These findings were scored over 2. On the other side, tilapias fed on *M. oleifera* at 5% and 10% showed intact and increased villi length with an increasing number of goblet cells (red arrow), especially with increasing the exposure time to infection (**Table 8**).

Spleen also displayed marked changes in its architecture which varied between normal and following infection conditions (**Figures 6, 7**). Under normal health conditions (**Figure 6**), tilapias fed on the basal diet free of moringa showed normal splenic tissue with normal red and white pulps with melanomacrophage cells located within white pulp (black arrowhead) and lymphocytic infiltration (black arrow). Interestingly, moringa inclusion at 5% and 10% of the diet caused marked hyperplasia of both melanomacrophage cells

(black arrowhead) and increased lymphocytic infiltration within white pulp (black arrow). Since no pathological lesions were noticed, a 0 score was considered. On the other hand, after 24 and 48 h post *A. hydrophila* infection (**Figure 7**), the histological findings in the control groups showed signs of inflammation, including congestion of blood sinusoids and perivascular edema to degenerative changes in both red and white bulbs. The melanomacrophage center was present mainly near the blood sinusoids. These findings were confirmed with a significant increase in lesion scores (**Table 7**), where the highest score (around 3) was reported following *A. hydrophila* challenging and increased with increasing exposure time ($P < 0.05$).

Interestingly, the signs of inflammation improved with moringa inclusion, which caused decreased degeneration with very clear melanomacrophage centers and slight vascular congestion. The lymphocytic infiltration was revealed particularly after 48 h post-infection. Again, this effect was confirmed with significantly decreased lesion scores in moringa-fed groups and exposed to *A. hydrophila*.

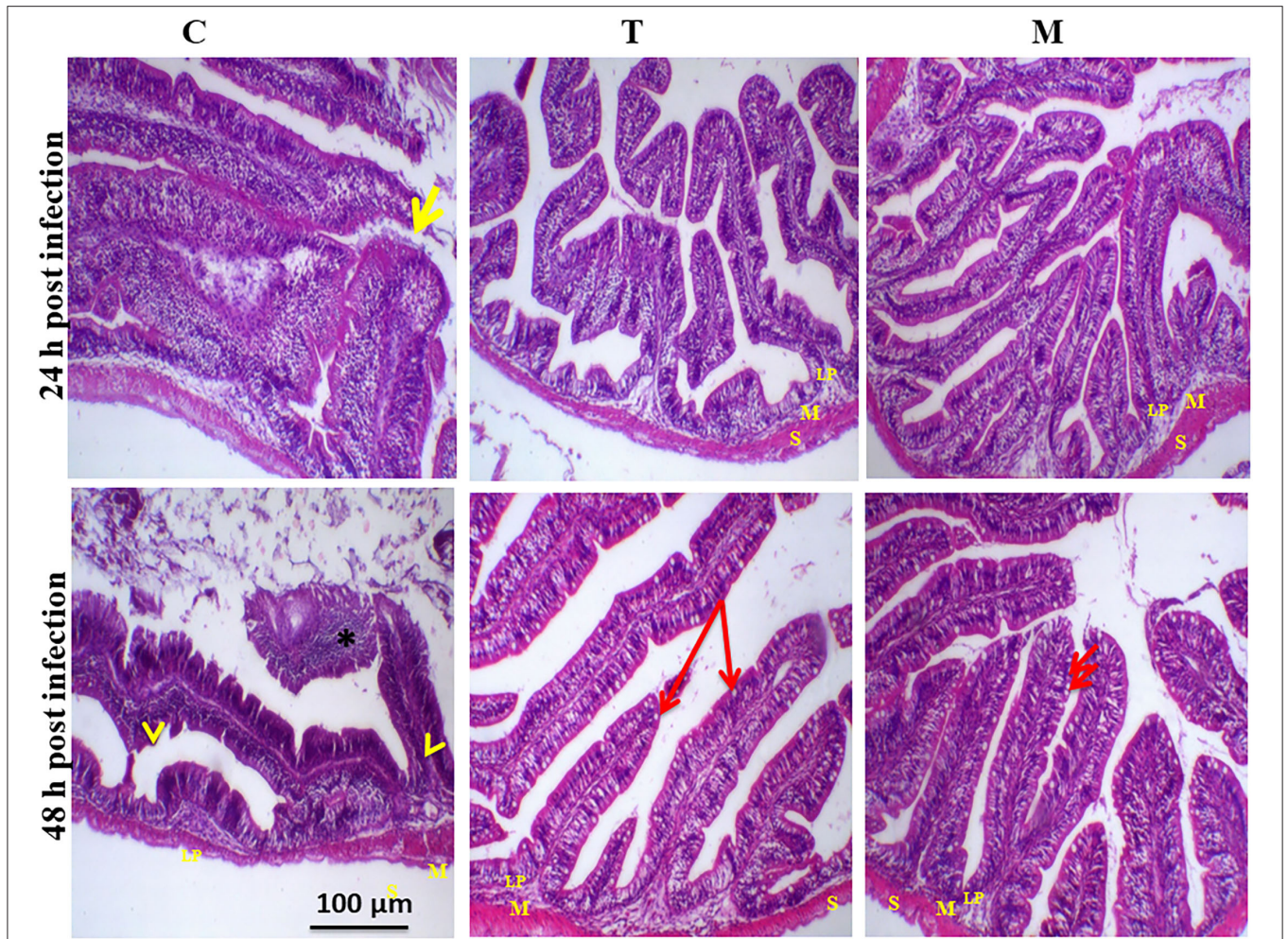


FIGURE 5 | H&E stained photomicrograph of mono-sex Nile tilapia mid-gut following 24 and 48 h of *A. hydrophila* infection with and without moringa dietary inclusion. C, T, and M denote the 0, 5, and 10% moringa levels, respectively. The yellow arrow represents intestinal villi, yellow arrowhead indicates detachment of the apical part of enterocytes with leukocytic infiltration (star), while the red arrow represents goblet cells. M represents tunica muscularis, LP means lamina propria sub-mucosa, while S means tunica Serosa.

TABLE 6 | Mid-gut morphometry of mono-sex Nile tilapia fed *M. oleifera* leaf powder containing diet.

	24 h non-infected			24 h infected		
	Length (µ.M)	Width (µ.M)	Inter-villi space (µ.M)	Length (µ.M)	Width (µ.M)	Inter-villi space (µ.M)
C	585.16 ± 32.05 ^{Ac*}	90.19 ± 3.33 ^{Ab*}	61.21 ± 3.69	466.90 ± 53.80 ^{Ab}	60.69 ± 3.38 ^{Bc}	53.63 ± 9.25
T	678.18 ± 21.88 ^{Bb}	109.93 ± 4.95 ^{Ab*}	73.39 ± 8.93	620.75 ± 22.95 ^{Ba}	82.69 ± 9.78 ^{Bb}	72.71 ± 4.32
M	830.67 ± 41.61 ^{Aa*}	131.84 ± 15.17 ^{Aa}	81.43 ± 16.58	616.66 ± 12.30 ^{Aa}	111.59 ± 2.70 ^{Aa}	69.36 ± 3.55
	48 h non-infected			48 h infected		
	Length (µ.M)	Width (µ.M)	Inter-villi space (µ.M)	Length (µ.M)	Width (µ.M)	Inter-villi space (µ.M)
C	568.95 ± 49.14 ^{Ac*}	96.34 ± 4.26 ^{Ac}	66.24 ± 4.48	474.89 ± 27.88 ^{Ab}	93.49 ± 7.04 ^{Ab}	62.13 ± 2.39
T	743.41 ± 18.08 ^{Ab*}	118.58 ± 4.44 ^{Ab}	80.51 ± 4.53	508.56 ± 23.11 ^{Ba}	123.40 ± 3.51 ^{Aa}	75.92 ± 2.39
M	851.39 ± 25.24 ^{Aa*}	164.79 ± 25.32 ^{Aa}	77.64 ± 4.85	558.87 ± 5.14 ^{Ba}	169.53 ± 4.83 ^{Aa}	69.88 ± 6.53

C, control group fed diet with 0% *M. oleifera*; T, fishes fed *M. Oleifera* leaf dietary inclusion at 5% while M, fishes fed *M. Oleifera* leaf dietary inclusion at 10%. Data are presented as means ± SEM. Different letters represent statistical differences between different treatments at p-value < 0.05. *P < 0.01

TABLE 7 | A summary of lesion scores in spleen and mid-gut of mono-sex Nile tilapia fed *M. oleifera* leaf powder containing diet.

Moringa levels	Normal health condition		24 h post infection		48 h post infection	
	Spleen	Mid-gut	Spleen	Mid-gut	Spleen	Mid-gut
C	00.00 ± 00 ^{ab}	00.00 ± 00 ^{ab}	2.83 ± 0.12 ^{ab}	2.75 ± 0.14 ^{ab}	2.87 ± 0.13 ^{ab}	2.92 ± 0.08 ^{ab}
T	00.00 ± 00 ^{ab}	00.00 ± 00 ^{ab}	1.90 ± 0.20 ^{ba}	1.83 ± 0.22 ^{ba}	1.60 ± 0.17 ^{ba}	1.50 ± 0.14 ^{ba}
M	0.00 ± 00 ^{ab}	0.00 ± 00 ^{ab}	1.30 ± 0.17 ^{ba}	1.25 ± 0.14 ^{ba}	1.30 ± 0.16 ^{ba}	1.00 ± 0.63 ^{ba}
P-values	Spleen		Mid-gut			
Moringa effect	0.009		0.001			
Time effect	0.399		0.184			
Infection	0.026		0.004			
Moringa x time x infection	0.395		0.510			

C, T, and M denote the 0, 5, and 10% moringa levels, respectively. Lesion scoring in spleen and mid-gut ranged from 0 to 3; 0 refers to non or slight pathological lesions; 1 refers to mild changes; 2, moderate changes; and 3 represents severe changes. Data are expressed as means ± SEM. Different letters represent statistical differences between different treatments at p -value < 0.05 using three-way ANOVA test.

TABLE 8 | Goblet cells count (cell count/mm²) under normal health condition and following *A. hydrophila* infection.

Moringa levels	Normal health condition	24 h post infection	48 h post infection
C	142.00 ± 8.08 ^{cb}	171.33 ± 6.96 ^{ca}	168.67 ± 10.48 ^{ca}
T	221.33 ± 11.62 ^{ab}	242.67 ± 9.62 ^{aa}	229.33 ± 7.06 ^{aa}
M	196.67 ± 10.48 ^{bb}	211.33 ± 14.44 ^{ba}	221.33 ± 14.11 ^{ba}
Moringa effect	p value < 0.001		
Time effect	p value = 0.535		
Infection effect	p value = 0.045		
Moringa x time x infection	p value = 0.016		

C, T, and M denote the 0, 5, and 10% moringa levels, respectively. Data are expressed as means ± SEM. Lower case letters represent the statistically significant effect of moringa, while upper case letters represent statistically significant effect of infection at P < 0.05 using three-way ANOVA test.

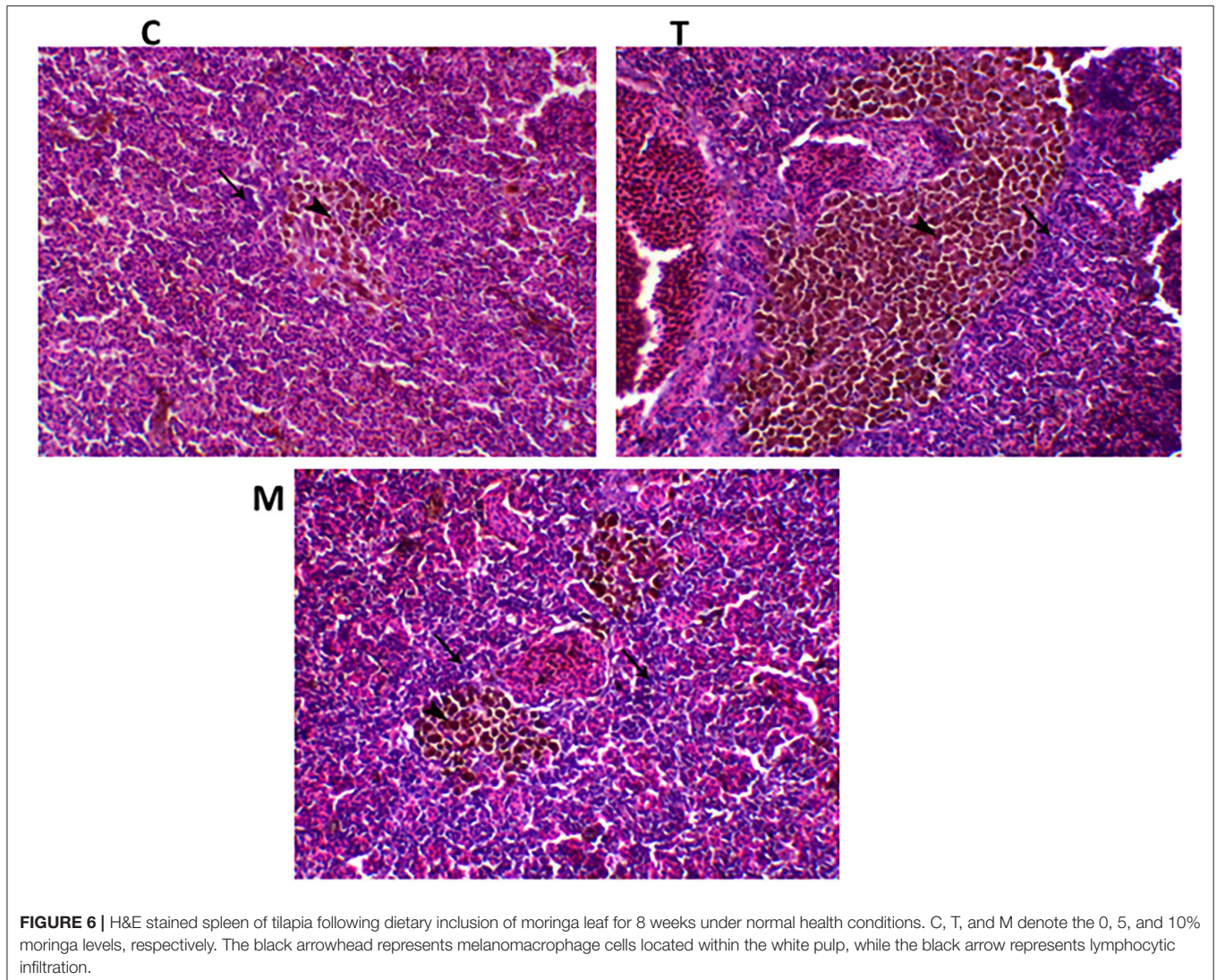
DISCUSSION

The current study's foremost results revealed that the dietary inclusion of *M. oleifera* leaf powder in mono-sex Nile tilapias increased the phagocytic and lysozyme activities with a marked lowering of the H/L ratio, especially following *A. hydrophila* challenge. Similar findings were reported for *M. oleifera*, in many fish species, such as increasing serum lysozyme activity, immunoglobulin M (IgM) level, phagocytic, peroxidase, and the respiratory burst activities along with the complement component 3 (C3) values [reviewed by Abdel-Latif et al. (18)]. These immune-enhancing effects of *M. oleifera* might be due to its abundant contents of many biologically active phytochemicals such as polyphenols, volatile oils, vitamins, such as vitamin E, A, and ascorbic acid, phenolic acids, flavonoids, and sterols (β -sitosterol) with potential antioxidant, antifungal, and antibacterial properties [reviewed by Abdel-Latif et al. (18)]. On the other hand, the phagocytic activity and index were not influenced by moringa feeding under normal health conditions. This response concurs with (41–43), who recorded that the direct application of different plant extracts did not improve the leucocyte activities and even reduced them at larger doses.

Furthermore, following *A. hydrophila* challenge, the effect on PI increased with increasing time where there was no change after 24 h post-challenge, but after 48 h of the challenging, PI was significantly elevated. The latter effect might increase the dose of microorganisms that increased with increasing time (44).

Since lysozyme is an effective mucolytic innate immunity mediator of leukocytic origin that has antibacterial activity and stimulates phagocytosis, it is a good indicator of the non-specific immune functions of fish (45). In the present study, under normal health conditions, and following *A. hydrophila* challenge, moringa dietary inclusion induced a significant dose and time-dependent higher lysozyme activity. This increasing response is likely correlated with the antibacterial properties of *M. oleifera* due to its high content of several immune-stimulant bioactive components like carotenoids, moringinine, vitamins, minerals, sterols, alkaloids, flavonoids, caffeoylquinic acids, and phytoestrogens (41, 46). Our results agreed with (22, 46, 47) findings that reported increasing lysozyme activities with increasing moringa levels in the diet.

Besides, the aforementioned effects were accompanied by, under normal conditions, significant upregulations of the *CAT* and *GPx*, relative mRNA levels. These anti-stress properties of *M. oleifera* might be explained by its high content of

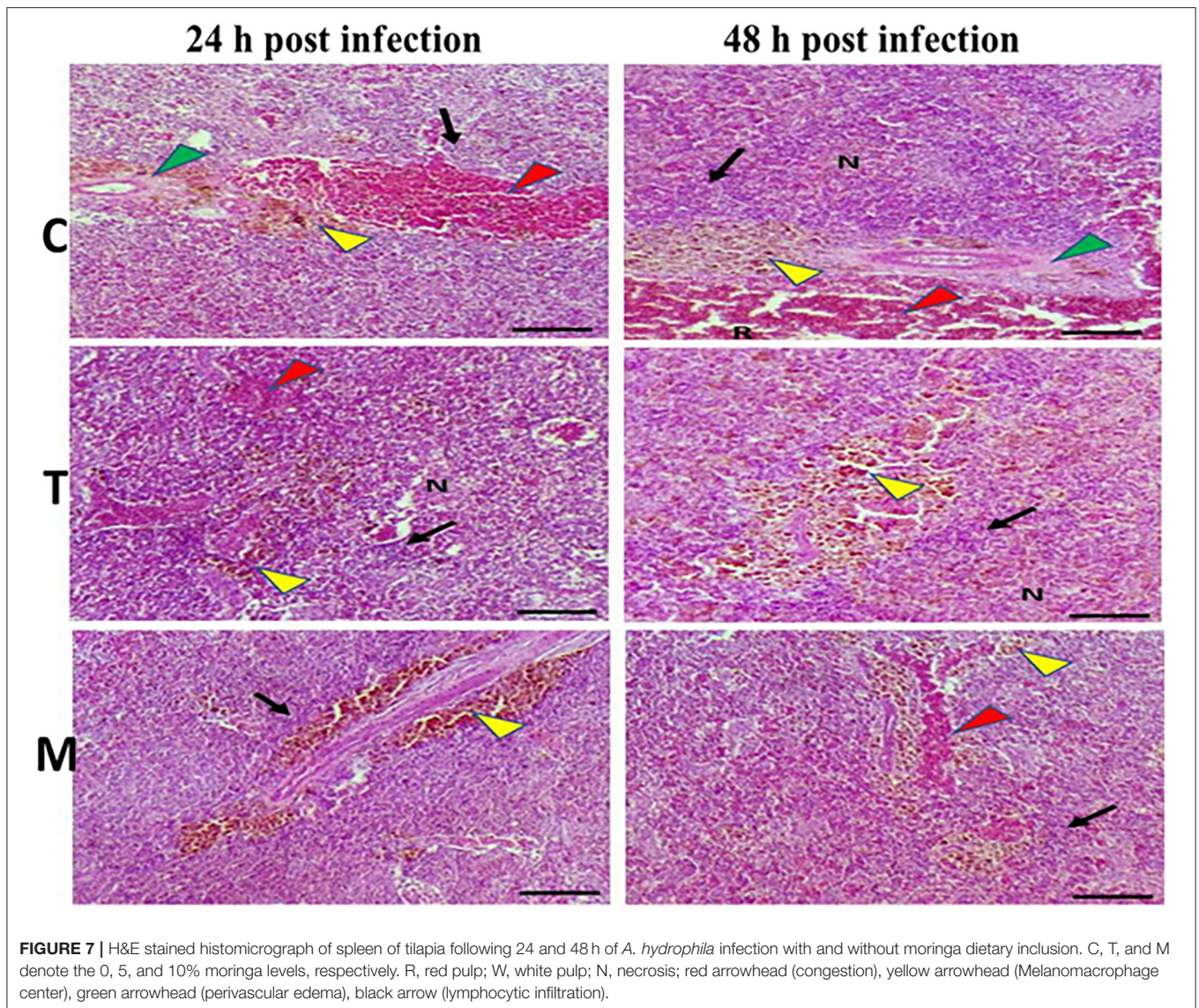


phytogenic constituents, which protect against oxidative and stress-associated damages (48). In this regard, the high content of moringa leaf extract of ascorbic acid, phenolics, and flavonoids like sitosterol, quercetin, and kaempferol restore glutathione, glutathione-S transferase (GST), glutathione peroxidase (GPx), and glutathione reductase (GR) levels by preventing lipid peroxidation and scavenging free radicals (49–51). Besides reducing the lipid peroxidation, increasing the antioxidants such as SOD and CAT activities following dietary *M. oleifera* (48) is also probably linked with improving liver health (28).

Moreover, the antioxidant characteristic of moringa is likely mediated via the stimulation of nuclear factor (erythroid-derived 2)-like 2 (Nrf2) transcriptional factor, which has a critical role in stimulating multiple antioxidants and chemoprotective genes (52). The activation of Nrf2 is due to the high content of isothiocyanates in *M. oleifera* leaves (53). In turn, Nrf2 triggers the antioxidant response element (ARE), which upregulates the transcription of many genes such as GST, stimulating a higher GPx response (52–54). Our findings are consistent with (29)

results, which reported that *M. oleifera* leaf extracts improved CAT, SOD, and GSH-Px activity in case of ammonia stress compared to the control.

While, in the case of *A. hydrophila* challenge, there was a marked upregulation of *CAT* and *GPx* transcripts which was significantly downregulated in the case of moringa fed groups. The noticed upregulation of *CAT* and *GPx* in the case of *A. hydrophila*-infected tilapias might be explained by *A. hydrophila* infection-induced stress response, which activates the Nrf2 response inducing more antioxidant synthesis mediated through ARE activation (52). Whereas the downregulated activities of *CAT* and *GPx* in the case of moringa-fed groups may be due to moringa bioactive components' direct potential antioxidant capabilities to inhibit the formation of hydroxyl radicals and stabilize free radicals or stabilize free radicals peroxy-radicals because they are strong electron donors (hydrogen) (41). Also, because of their ability to neutralize free radicals or decompose peroxides into oxygen and water (55). Besides, lowering the *CAT* and *GPx* expression level is possibly associated



with the anti-inflammatory characteristics of *M. oleifera* that reduces the reactive oxygen species (ROS) production due to reducing iNOS expression level, which was mediated by NF- κ B inactivation (52, 56).

In addition, the *M. oleifera* dietary inclusion stress relief response was associated with an alteration of leukocyte populations of the mono-sex Nile tilapias. Under normal health conditions, *M. oleifera* induced a dose-dependent increase and decrease in the lymphocytic and heterophils populations, respectively. This effect might explain the reduction of the H/L ratio. These results agreed with (57, 58) findings, which reported that aqueous moringa leaf extract stimulates a dose-dependent elevation in human lymphocyte viability and the count of CD4+ and CD8+ cells. On the other hand, following infection with *A. hydrophila*, there was no clear observed effect (only slight decreases) of moringa on leukocytic populations, which might correlate with moringa's properties to reduce the production

of inflammatory mediators (41). This is probably confirmed by reducing the leukocytic infiltration in spleen tissue.

To the best of our knowledge, *Moringa oleifera* leaves have anti-inflammatory propriety, which is not well investigated at the transcriptomic level in Nile tilapia. Also, there are no available records on moringa leaves dietary inclusion on proinflammatory cytokines expression levels under normal health conditions. All available reports are on its impact on the inflammatory response in case of stress and infection-associated conditions. So, our study represents the first report on the effects of moringa leaves dietary inclusion on the transcriptomic profile of inflammatory mediator genes, such as *IL-1 β* , *IL-8*, and *IFN- γ* under normal health and infection conditions. Under normal health conditions of mono-sex Nile tilapia, *M. oleifera* induced significant upregulations of the *IL-1 β* , *IL-8*, and *IFN- γ* mRNA transcripts. This effect might be linked to increasing the number of lymphocytes and their infiltration in the spleen, which increased with increasing

moringa levels in the diet. Moreover, this pro-inflammatory response might correlate with the high isothiocyanates content in moringa leaves (53). With its anti-inflammatory characteristics, it also promotes a pro-inflammatory response in some species, such as mice (59). The isothiocyanates associated with pro-inflammatory response are related to the activation of Th1 (59).

Besides, *A. hydrophila* challenge caused significant upregulations of *IL-1 β* , *IL-8*, and *IFN- γ* mRNA levels which might be associated with the activation of the toll-like receptors (TLRs) pathway by LPS content of *A. hydrophila* cell wall (60). The latter activates NF- κ B, which induces the expression of proinflammatory cytokines through a MyD88-independent pathway activation (60). The interesting reduction of this inflammatory response and downregulation of *IL-1 β* , *IL-8*, and *IFN- γ* mRNA levels in the case of moringa-fed groups are perhaps linked with the inactivation of NF- κ B via Nrf2 activation by the high content of moringa leaves of isothiocyanates (53). Therefore, further future investigations of moringa dietary inclusion effects on Th2 and anti-inflammatory response are recommended. In addition, lowering the *A. hydrophila*-associated inflammatory response in the case of moringa-fed tilapias might clarify the mitigation of inflammatory signs in the intestine and the degenerative changes that occurred due to *A. hydrophila*.

Conclusively, *M. oleifera* leaf powder dietary inclusion enhanced mono-sex Nile tilapia's immune response under normal health conditions through increasing the phagocytic and lysozyme activities with upregulations of the CAT and GPx, and *IL-1 β* , *IL-8*, and *IFN- γ* relative mRNA levels. Nevertheless, differential responses characterized by reducing the infection-associated inflammatory response as confirmed by downregulation of *IL-1 β* , *IL-8*, and *IFN- γ* transcription levels and improving the degenerative changes in intestinal and spleen tissues were documented when fish were exposed to *A. hydrophila* and fed *M. oleifera* leaf powder diet. These differential responses may be explained by the different interactions of *M. oleifera* active components with fish physiological conditions, age, and the immune cascade to induce immune stimulation (41).

CONCLUSION

The dietary inclusion of *M. oleifera* leaf powder to mono-sex Nile tilapia at 5% and 10% significantly improved the growth performance and modulated the immune response. The immune-modulatory response varied between normal health culturing conditions and in case of infection. Normally, *M. oleifera* stimulates the tilapia immune system manifested by increasing antioxidant activities, reducing stress H/L

ratio indicator, and inducing a pro-inflammatory response distinguished by upregulation of *IL-1 β* , *IL-8*, and *IFN- γ* transcripts. While, in the condition of infection, *M. oleifera* mainly acted to reduce the *A. hydrophila*-associated inflammatory and degenerative changes by downregulating the expression level of *IL-1 β* , *IL-8*, and *IFN- γ* . Further investigations are suggested to deeply realize the exact mechanisms that mediate the *M. oleifera* effects between normal and diseased conditions.

DATA AVAILABILITY STATEMENT

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

ETHICS STATEMENT

The animal study was reviewed and approved by the present trial was adopted on mono-sex Nile tilapia, *Oreochromis niloticus* following the standard operating steps approved by the Animal Care Committee, Faculty of Veterinary Medicine, Kafrelsheikh University, Egypt (KFS-2019/9). Written informed consent was obtained from the owners for the participation of their animals in this study.

AUTHOR CONTRIBUTIONS

SE-K, KE-N, and EM: conceptualization, data curation, resources, supervision, writing—original draft preparation, and writing—review and editing. RM, WA, CC-J, and EA: conceptualization, methodology, formal analysis, supervision, writing—original draft preparation, and writing—review and editing. MH and SA: data curation, investigation, and methodology. SE-K, KE-N, and EM: methodology and software. NA, MS, FA, ME-S, and EM: methodology and formal analysis. SE-K: methodology, software, and resources. KE-N: resources, data curation, and investigation. All authors contributed to the article and approved the submitted version.

ACKNOWLEDGMENTS

The authors are thankful for the financial support provided by the Fundação de Amparo à Pesquisa do Estado do Rio de Janeiro (FAPERJ) Brazil—grant number [E-26/200.891/2021] and the Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq)—grant number [313119/2020-1]. This project was also supported by the Deanship of Scientific Research (DSR), King Abdulaziz University, Jeddah. The authors acknowledge with thank DSR for technical support.

REFERENCES

1. Waagbø R, Remø SC. 7 - Functional diets in fish health management. In: Kibenge FSB, Powell MD, editors. *Aquaculture Health Management*. Academic Press (2020). p. 187–234.
2. Plant KP, LaPatra SE. Advances in fish vaccine delivery. *Dev Comp Immunol*. (2011) 35:1256–62. doi: 10.1016/j.dci.2011.03.007
3. Hamouda A, Moustafa E, Zayed M. Overview on the most prevailing bacterial diseases infecting *Oreochromis niloticus* at aswan fish hatchery, Egypt. *Adv Anim Vet Sci*. (2019) 7:950–61. doi: 10.17582/journal.aavs/2019/7.11.950.961

4. Austin B, Austin D. *Bacterial Fish Pathogens, Diseases of Farmed and Wild Fish Fourth Edition*. Chichester, UK: Praxis Publishing Ltd (2007).
5. El-Saadony MT, Alagawany M, Patra AK, Kar I, Tiwari R, Dawood MA, et al. The functionality of probiotics in aquaculture: an overview. *Fish Shellfish Immunol.* (2021) 117:36–52. doi: 10.1016/j.fsi.2021.07.007
6. El-Saadony MT, Saad AM, Taha TF, Najjar AA, Zbermawi NM, Nader MM, et al. Selenium nanoparticles from *Lactobacillus paracasei* HM1 capable of antagonizing animal pathogenic fungi as a new source from human breast milk. *Saudi J. Biol. Sci.* (2021) 28(12):6782–94. doi: 10.1016/j.sjbs.2021.07.059
7. El-Saadony MT, Zbermawi NM, Zbermawi NM, Burollus MA, Shafi ME, Alagawany M, et al. Nutritional aspects and health benefits of bioactive plant compounds against infectious diseases: a review. *Food Rev. Int.* (2021) 2021:1–23. doi: 10.1080/87559129.2021.1944183
8. Santos L, Ramos F. Analytical strategies for the detection and quantification of antibiotic residues in aquaculture fishes: a review. *Trends Food Sci Technol.* (2016) 52:16–30. doi: 10.1016/j.tifs.2016.03.015
9. Alagawany M, Abd El-Hack M, Farag M, Elnesr S, El-Kholy M, Saadeldin I, et al. Dietary supplementation of *Yucca schidigera* extract enhances productive and reproductive performances, blood profile, immune function, and antioxidant status in laying Japanese quails exposed to lead in the diet. *Poult. Sci.* (2018) 97:3126–37. doi: 10.3382/ps/pey186
10. Saeed M, Abd El-Hack ME, Alagawany M, Arain MA, Arif, M, et al. Chicory (*Cichorium intybus*) herb: Chemical composition, pharmacology, nutritional and health applications. *Int. J. Pharmacol.* (2017) 13:351–60. doi: 10.3923/ijp.2017.351.360
11. Aladailah SH, Khafaga AF, Abd El-Hack ME, Al-Gabri NA, Abukhalil MH, Alfwuaires MA, et al. *Spirulina platensis* ameliorates the sub chronic toxicities of lead in rabbits via anti-oxidative, anti-inflammatory, and immune stimulatory properties. *Sci Total Environ.* (2020) 701:134879. doi: 10.1016/j.scitotenv.2019.134879
12. Fuchs VI, Schmidt J, Slater MJ, Zentek J, Buck BH, Steinhagen D. The effect of supplementation with polysaccharides, nucleotides, acidifiers and *Bacillus* strains in fish meal and soy based diets on growth performance in juvenile turbot (*Scophthalmus maximus*). *Aquaculture.* (2015) 437:243–51. doi: 10.1016/j.aquaculture.2014.12.007
13. Dawood MAO, Koshio S, Esteban M. Beneficial roles of feed additives as immunostimulants in aquaculture: a review. *Rev Aquac.* (2018) 10:950–74. doi: 10.1111/raq.12209
14. Van Hai N. The use of medicinal plants as immunostimulants in aquaculture: a review. *Aquaculture.* (2015) 446:88–96. doi: 10.1016/j.aquaculture.2015.03.014
15. Abd El-HackME, AlagawanyM, ElrysAS, Desoky ESM, Tolba H, Swelum AA. Effect of forage *Moringa oleifera* L.(moringa) on animal health and nutrition and its beneficial applications in soil, plants and water purification. *Agriculture.* (2018) 8:145. doi: 10.3390/agriculture8090145
16. Saini RK, Sivanesan I, Keum Y-S. Phytochemicals of *Moringa oleifera*: a review of their nutritional, therapeutic and industrial significance. *3 Biotech.* (2016) 6:203. doi: 10.1007/s13205-016-0526-3
17. Shih MC, Chang CM, Kang SM, Tsai ML. Effect of different parts (leaf, stem and stalk) and seasons (summer and winter) on the chemical compositions and antioxidant activity of *Moringa Oleifera*. *Int J Mol Sci.* (2011) 12:6077–88. doi: 10.3390/ijms12096077
18. Abdel-Latif HM, Abdel-Daim MM, Shukry M, Nowosad J, Kucharczyk D. Benefits and applications of *Moringa oleifera* as a plant protein source in Aquafeed: a review. *Aquaculture.* (2022) 547:737369. doi: 10.1016/j.aquaculture.2021.737369
19. Leone A, Spada A, Battezzati A, Schiraldi A, Aristil J, Bertoli S. Cultivation, genetic, ethnopharmacology, phytochemistry and pharmacology of *Moringa oleifera* leaves: an overview. *Int J Mol Sci.* (2015) 16:12791–835. doi: 10.3390/ijms160612791
20. Abd El-Rahman HH, Abo-State HA, El-Nadi ASM, Abozaid H, Mohamed MI, Abdalla AE-FM. Growth performance, feed utilization and body composition of Nile Tilapia (*Oreochromis niloticus*) fingerlings fed moringa (*Moringa oleifera* Lam) Seed Meal. *J Fish Aquat Sci.* (2017) 12:36–41. doi: 10.3923/jfas.2017.36.41
21. Elabd H, Soror E, El-Asely A, El-Gawad EA, Abbass A. Dietary supplementation of Moringa leaf meal for Nile tilapia *Oreochromis niloticus*: effect on growth and stress indices. *Egypt J Aquat Res.* (2019) 45:265–71. doi: 10.1016/j.ejar.2019.05.009
22. Abd El-Gawad EA, El Asely AM, Soror EI, Abbass AA, Austin B. Effect of dietary *Moringa oleifera* leaf on the immune response and control of *Aeromonas hydrophila* infection in Nile tilapia (*Oreochromis niloticus*) fry. *Aquac Int.* (2020) 28:389–402. doi: 10.1007/s10499-019-00469-0
23. Arif M, Hayat Z, Abd El-Hack ME, Saeed M, Imran HM, Alowaimier AN, et al. Impacts of supplementing broiler diets with a powder mixture of black cumin, Moringa and chicory seeds. *South African J Animal Sci.* (2019) 49:564–72. doi: 10.4314/sajas.v49i3.17
24. Ashour EA, El-Kholy MS, Alagawany M, Abd El-Hack ME, Mohamed LA, Tufarelli V. Effect of dietary supplementation with *Moringa oleifera* leaves and/or seeds powder on production, egg characteristics, hatchability and blood chemistry of laying Japanese quails. *Sustainability.* (2020) 12:2463. doi: 10.3390/su12062463
25. Ozovehe BN. Growth performance, haematological indices and some biochemical enzymes of juveniles *Clarias gariepinus* (Burchell 1822) fed varying levels of *Moringa oleifera* leaf meal diet. *J. Aquac. Res. Dev.* (2013) 4:2. doi: 10.4172/2155-9546.1000166
26. Richter N, Siddhuraju P, Becker K. Evaluation of nutritional quality of moringa (*Moringa oleifera* Lam) leaves as an alternative protein source for Nile tilapia (*Oreochromis niloticus* L). *Aquaculture.* (2003) 217:599–611. doi: 10.1016/S0044-8486(02)00497-0
27. Hlophe S, Moyo N, A. comparative study on the use of *Pennisetum clandestinum* and *Moringa oleifera* as protein sources in the diet of the herbivorous *Tilapia rendalli*. *Aquac Int.* (2014) 22:1245–62. doi: 10.1007/s10499-013-9744-4
28. El-Kassas S, Abdo SE, Abosheashaa W, Mohamed R, Moustafa EM, Helal MA, et al. Growth performance, serum lipid profile, intestinal morphometry, and growth and lipid indicator gene expression analysis of mono-sex Nile tilapia fed *Moringa oleifera* leaf powder. *Aquac Rep.* (2020) 18:100422. doi: 10.1016/j.aqrep.2020.100422
29. Kaleo IV, Gao Q, Liu B, Sun C, Zhou Q, Zhang H, et al. Effects of *Moringa oleifera* leaf extract on growth performance, physiological and immune response, and related immune gene expression of *Macrobrachium rosenbergii* with *Vibrio anguillarum* and ammonia stress. *Fish & shellfish immunol.* (2019) 89:603–13. doi: 10.1016/j.fsi.2019.03.039
30. Shiau S-Y, Huang S-L. Influence of varying energy levels with two protein concentrations in diets for hybrid tilapia (*Oreochromis niloticus* × *O. aureus*) reared in seawater *Aquaculture.* (1990) 91:143–52. doi: 10.1016/0044-8486(90)90183-N
31. Omar A, Moustafa EM, Zayed MM. Identification and characterization of virulence associated genes from pathogenic *Aeromonas hydrophila* strains. *World Vet J.* (2016) 6:185–92. doi: 10.5455/wvj.20161084
32. Reed LJ, Muench H. A simple method of estimating fifty per cent endpoints. *Am J Epidemiol.* (1938) 27:493–7. doi: 10.1093/oxfordjournals.aje.a118408
33. Li J, Ni X, Liu Y, Lu C. Detection of three virulence genes *ahp*, *ahp* and *aerA* in *Aeromonas hydrophila* and their relationship with actual virulence to zebrafish. *J Appl Microbiol.* (2011) 110:823–30. doi: 10.1111/j.1365-2672.2011.04944.x
34. Augustave LC. *Study of Treated and Untreated Oil Based Drilling Waste Using a Biomarker Approach: Gill and Liver Histopathology in Atlantic Salmon (Salmo salar)*. Norway: University of Stavanger (2014).
35. Mahmoud MA, El-Rahim A, Abeer H, Mahrous KF, Abdelsalam M, Abu-Aita NA, et al. The impact of several hydraulic fracturing chemicals on Nile tilapia and evaluation of the protective effects of *Spirulina platensis*. *Environ Sci Pollut Res.* (2019) 26:19453–67. doi: 10.1007/s11356-019-05246-3
36. Ngamkala S, Futami K, Endo M, Maita M, Katagiri T. Immunological effects of glucan and *Lactobacillus rhamnosus* GG, a probiotic bacterium, on Nile tilapia *Oreochromis niloticus* intestine with oral *Aeromonas* challenges. *Fish Sci.* (2010) 76:833–40. doi: 10.1007/s12562-010-0280-0
37. Abo-Al-Ela HG, El-Nahas AF, Mahmoud S, Ibrahim EM. Vitamin C modulates the immunotoxic effect of 17 α -methyltestosterone in Nile tilapia. *Biochemistry.* (2017) 56:2042–50. doi: 10.1021/acs.biochem.6b01284
38. Lie O, Syed M, Solbu H. Improved agar plate assays of bovine lysozyme and haemolytic complement activity. *Acta Vet Scand.* (1986) 27:23–32. doi: 10.1186/BF03548556

39. Dawood MA, Moustafa EM, Gewaily MS, Abdo SE, AbdEl-Kader MF, SaadAllah MS, et al. Ameliorative effects of *Lactobacillus plantarum* L-137 on Nile tilapia (*Oreochromis niloticus*) exposed to deltamethrin toxicity in rearing water. *Aquat Toxicol.* (2020) 219:105377. doi: 10.1016/j.aquatox.2019.105377
40. Livak KJ, Schmittgen TD. Analysis of relative gene expression data using real-time quantitative PCR and the $2^{-\Delta\Delta CT}$ method. *Methods.* (2001) 25:402–8. doi: 10.1006/meth.2001.1262
41. García-Beltrán JM, Mansour AT, Alsaqufi AS, Ali HM, Esteban MÁ. Effects of aqueous and ethanolic leaf extracts from drumstick tree (*Moringa oleifera*) on gilthead seabream (*Sparus aurata* L) leucocytes, and their cytotoxic, antitumor, bactericidal and antioxidant activities. *Fish & Shellfish Immunol.* (2020) 106:44–55. doi: 10.1016/j.fsi.2020.06.054
42. Beltrán JMG, Espinosa C, Guardiola FA, Esteban MÁ. Dietary dehydrated lemon peel improves the immune but not the antioxidant status of gilthead seabream (*Sparus aurata* L). *Fish Shellfish Immunol.* (2017) 64:426–36. doi: 10.1016/j.fsi.2017.03.042
43. Othmen KB, Elfalleh W, Beltrán JMG, Esteban MÁ, Haddad M. An in vitro study of the effect of carob (*Ceratonia siliqua* L) leaf extracts on gilthead seabream (*Sparus aurata* L) leucocyte activities. Antioxidant, cytotoxic and bactericidal properties. *Fish shellfish immunol.* (2020) 99:35–43. doi: 10.1016/j.fsi.2020.02.005
44. Abdel-Mageid AD, Zaki AG, El Senosi YA, Fahmy HA, El Asely AM, Abo-Al-Ela HG, et al. Modulatory effect of lipopolysaccharide on immune-related gene expression and serum protein fractionation in grey mullet, *Mugil cephalus*. *Aquac Res.* (2020) 51:1643–52. doi: 10.1111/are.14510
45. Saurabh S, Sahoo P. Lysozyme: an important defence molecule of fish innate immune system. *Aquac Res.* (2008) 39:223–39. doi: 10.1111/j.1365-2109.2007.01883.x
46. Mansour AT, Miao L, Espinosa C, García-Beltrán JM, Francisco DCC, Esteban MÁ. Effects of dietary inclusion of *Moringa oleifera* leaves on growth and some systemic and mucosal immune parameters of seabream. *Fish Physiol Biochem.* (2018) 44:1223–40. doi: 10.1007/s10695-018-0515-z
47. Bisht M, Kumar A, Shah TK. Effect of *Moringa oleifera* leaf powder on skin mucosal immune responses and growth performance of guppy, *Poecilia reticulata* (Peter, 1860). *Aquac Res.* (2020) 51:4984–90. doi: 10.1111/are.14834
48. Kou X, Li B, Olayanju JB, Drake JM, Chen N. Nutraceutical or pharmacological potential of *Moringa oleifera* Lam. *Nutrients.* (2018) 10:343. doi: 10.3390/nu10030343
49. Singh D, Arya PV, Aggarwal VP, Gupta RS. Evaluation of antioxidant and hepatoprotective activities of *Moringa oleifera* Lam. leaves in carbon tetrachloride-intoxicated rats. *Antioxidants.* (2014) 3:569–91. doi: 10.3390/antiox3030569
50. Sharifudin SA, Fakurazi S, Hidayat MT, Hairuszah I, Aris Mohd Moklas M, Arulselvan P. Therapeutic potential of *Moringa oleifera* extracts against acetaminophen-induced hepatotoxicity in rats. *Pharm Biol.* (2013) 51:279–88. doi: 10.3109/13880209.2012.720993
51. Xu Y-B, Chen G-L, Guo M-Q. Antioxidant and anti-inflammatory activities of the crude extracts of *Moringa oleifera* from Kenya and their correlations with flavonoids. *Antioxidants.* (2019) 8:296. doi: 10.3390/antiox8080296
52. Jaja-Chimedza A, Graf BL, Simmler C, Kim Y, Kuhn P, Pauli GF, et al. Biochemical characterization and anti-inflammatory properties of an isothiocyanate-enriched moringa (*Moringa oleifera*) seed extract. *PLoS ONE.* (2017) 12:e0182658. doi: 10.1371/journal.pone.0182658
53. Waterman C, Cheng DM, Rojas-Silva P, Poulev A, Dreifus J, Lila MA, et al. Stable, water extractable isothiocyanates from *Moringa oleifera* leaves attenuate inflammation in vitro. *Phytochemistry.* (2014) 103:114–22. doi: 10.1016/j.phytochem.2014.03.028
54. Cheenpracha S, Park E-J, Yoshida WY, Barit C, Wall M, Pezzuto JM, et al. Potential anti-inflammatory phenolic glycosides from the medicinal plant *Moringa oleifera* fruits. *Bioorg Med Chem.* (2010) 18:6598–602. doi: 10.1016/j.bmc.2010.03.057
55. Shourbela R, El-Hawarry W, AM AE-L, Abo-Kora S. Potentiality of *Moringa oleifera* aqueous extract as a growth modulator and antistress in acute hypoxic Nile tilapia *Oreochromis niloticus*. *Iran. J. Fish Sci.* (2020) 19:67–84. doi: 10.22092/ijfs.2019.119208
56. Cui C, Chen S, Wang X, Yuan G, Jiang F, Chen X, et al. Characterization of *Moringa oleifera* roots polysaccharide MRP-1 with anti-inflammatory effect. *Int J Biol Macromol.* (2019) 132:844–51. doi: 10.1016/j.ijbiomac.2019.03.210
57. Nair S, Varalakshmi K. Anticancer, cytotoxic potential of *Moringa oleifera* extracts on HeLa cell line. *J Nat Pharm.* (2011) 2:138–42. doi: 10.4103/2229-5119.86260
58. Rachmawati I, Rifa'i M. In vitro immunomodulatory activity of aqueous extract of *Moringa oleifera* Lam leaf to the CD4+, CD8+ and B220+ cells in *Mus musculus*. *J Exp Life Sci.* (2014) 4:15–20. doi: 10.21776/ub.jels.2014.004.01.03
59. Martin SA, Król E. Nutrigenomics and immune function in fish: new insights from omics technologies. *Dev Comp Immunol.* (2017) 75:86–98. doi: 10.1016/j.dci.2017.02.024
60. Zhang J, Kong X, Zhou C, Li L, Nie G, Li X. Toll-like receptor recognition of bacteria in fish: ligand specificity and signal pathways. *Fish Shellfish Immunol.* (2014) 41:380–8. doi: 10.1016/j.fsi.2014.09.022

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 El-Kassas, Aljahdali, Abdo, Alaryani, Moustafa, Mohamed, Abosheashaa, Abdulraouf, Helal, Shafi, El-Saadony, El-Naggar and Conte-Junior. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.