



Brain Plasticity in Mammals: An Example for the Role of Comparative Medicine in the Neurosciences

Chiara La Rosa^{1,2} and Luca Bonfanti^{1,2*}

¹ Neuroscience Institute Cavalieri Ottolenghi, Turin, Italy, ² Department of Veterinary Sciences, University of Turin, Turin, Italy

OPEN ACCESS

Edited by:

Pablo Martín-Vasallo,
Universidad de La Laguna, Spain

Reviewed by:

Rosario Donato,
University of Perugia, Italy
Luis Miguel García-Segura,
Consejo Superior de Investigaciones
Científicas (CSIC), Spain

*Correspondence:

Luca Bonfanti
luca.bonfanti@unito.it

Specialty section:

This article was submitted to
Comparative and Clinical Medicine,
a section of the journal
Frontiers in Veterinary Science

Received: 17 September 2018

Accepted: 15 October 2018

Published: 01 November 2018

Citation:

La Rosa C and Bonfanti L (2018) Brain
Plasticity in Mammals: An Example for
the Role of Comparative Medicine in
the Neurosciences.
Front. Vet. Sci. 5:274.
doi: 10.3389/fvets.2018.00274

Comparative medicine deals with similarities and differences between veterinary and human medicine. All mammals share most basic cellular and molecular mechanisms, thus justifying murine animal models in a translational perspective; yet “mice are not men,” thus some biases can emerge when complex biological processes are concerned. Brain plasticity is a cutting-edge, expanding topic in the field of Neurosciences with important translational implications, yet, with remarkable differences among mammals, as emerging from comparative studies. In particular, adult neurogenesis (the genesis of new neurons from brain stem cell niches) is a life-long process in laboratory rodents but a vestigial, mostly postnatal remnant in humans and dolphins. Another form of “whole cell” plasticity consisting of a population of “immature” neurons which are generated prenatally but continue to express markers of immaturity during adulthood has gained interest more recently, as a reservoir of young neurons in the adult brain. The distribution of the immature neurons also seems quite heterogeneous among different animal species, being confined within the paleocortex in rodents while extending into neocortex in other mammals. A recent study carried out in sheep, definitely showed that gyrencephalic, large-sized brains do host higher amounts of immature neurons, also involving subcortical, white, and gray matter regions. Hence, “whole cell” plasticity such as adult neurogenesis and immature neurons are biological processes which, as a whole, cannot be studied exclusively in laboratory rodents, but require investigation in comparative medicine, involving large-sized, long-living mammals, in order to gain insights for translational purposes.

Keywords: comparative studies, sheep, immature neurons, translation, cerebral cortex, amygdala, comparative neuroanatomy

INTRODUCTION

Comparative medicine, namely, “the field of study concentrating on similarities and differences between veterinary and human medicine,” relies on the fact that “mice are not men” and thus laboratory rodents cannot always be the best models for human physiology and disease (1). Though mice and humans share striking biological similarities, mainly regarding basic molecular mechanisms which can be profitably explored by genetic manipulation, important differences and biases also emerge when complex biological processes are concerned. Dominant models may bias research directions or omit important context, possibly hampering reliable translational outcomes (2). In some cases, other animal models are required and the experience/expertise of the veterinary

sciences might be involved. Nevertheless, if veterinary and human medicine are steeped in a rich history of “One Medicine,” they have paradoxically parted ways, still not recognizing that they speak different dialects of the same language (1, 3). As stated by Schwabe (4), “the attempt to biologically pseudo-oligarchy-ize the human among species is one of the most pervasive mistakes of modern science,” and this is a pity for veterinary research, which seems to be intended to always remain in the background, not fully achieving the One Health purpose. The success of the One Health concept requires breaking down the interdisciplinary barriers that still separate human and veterinary medicine, but also ecological, evolutionary and environmental sciences (5). In this context, we strongly feel that many hidden opportunities do exist to develop engaging and useful themes falling in the landscape of the veterinary field of interest and being necessary to human health, not only referring to comparative pathology but also involving basic sciences. One of the most investigated topics in modern Neurosciences is neural plasticity, namely, the capability of the brain to change its structure/function throughout life in response to environmental challenges, thus being fundamental for species/individual adaptation and, notably, for brain aging and repair (6, 7). Due to its different features/types in relation to different neuroanatomies, environments and animal behavior, neural plasticity is a typical example of a complex biological process substantially differing among mammalian species (8, 9). In this perspective article we will address the reasons making brain plasticity an excellent topic for comparative medicine, also illustrating the current and future chances for developing it.

BRAIN PLASTICITY IN MAMMALS: WHY IS IT IMPORTANT?

In modern societies, neurological diseases, dementia and other age-related problems affecting the human brain represent a heavy health, social and economic burden (10). The progressive extension of human lifespan expectancy (11) will produce increasing numbers of individuals affected by neurological problems worldwide, thus making the issue of age-related dementia a global priority (12). Despite intensive research in Neuroscience, most brain pathologies involving progressive degeneration or functional loss of neurons still fall into the category of incurable diseases. Such failure is strictly linked to a well-known feature of the mammalian nervous tissue: its scarce capacity of renewing the cellular components through adulthood (most neurons are lifelong elements) and to undergo efficacious reparative processes after damage (7, 13, 14). In other words, despite thousands of papers published on the mechanisms of neural plasticity, our brain seems to suffer a substantial lack of structural plasticity. An explanation can be found in comparative medicine. Neurobiological research carried out on non-mammalian vertebrates clearly shows that fish, amphibians and reptiles do renew most neuronal populations throughout life and perform regeneration/repair after lesion (14, 15) (**Figure 1**). By contrast, both cell renewal and efficacious regenerative processes are not a natural property of the mammalian nervous

system, likely due to evolutionary constraints involving a combination of factors [among them: brain complexity, scarcity of stem cells, incapability of cell de-differentiation, role of immune system; (7, 13, 16)]. As a result, apart from widespread synaptic plasticity, which operates on the pre-existing neuronal elements and only in very small portions of the cell [dendritic spines, apical part of axons; see (17)], a “whole cell” plasticity involving changes in the overall shape and/or number of cells is a rare event. Usually, in mammals, the genesis of new neurons (adult neurogenesis) is highly spatially-restricted to two stem cell niches (the forebrain subventricular zone and the subgranular zone of the hippocampus; **Figure 1**). A third neurogenic zone is known to exist in the rodent hypothalamus, and other small spots of cell proliferation have been described to occur in other regions, yet none of them leading to fully structural/functional integration of the newlyborn neurons [reviewed in (18, 19)]. During the last three decades, most efforts of Neurodevelopmental research have been directed to foster residual “whole cell” plasticity in the mammalian nervous system, with the aim of reactivating intrinsic/silent programs of cell genesis/regeneration. Some cellular reactions can be experimentally-induced after lesion in murine models. For instance, activation of stem/progenitor cells in the neurogenic niches (20, 21) or local activation of neurogenic astrocytes (22, 23), can be elicited by inflammation/cell death and are followed by migration of neuroblasts toward the site of lesion. Though reminiscent of the first phases of reparative processes occurring in non-mammalian vertebrates (24), most of these tissue reactions ultimately do not lead to brain repair and cell replacement, as described in the widely plastic fish brain (reviewed in 15). Instead, they are followed by disappearance of the newlyborn cells, which have a transient existence (21, 23) during which they can exert “bystander” effects through the secretion of molecules/microvesicles in the site of lesion [reviewed in (7, 25)]. In parallel, different types of stem/progenitor cells have been harvested, cultured or produced *in vitro* (e.g., induced pluripotent stem cells) to be used for transplantation. Yet, no substantial efficacious treatments have been achieved in clinical trials for neurodegenerative diseases carried out in humans and based on stem cell treatments (26, 27). Out of 300 trials started before 2012 in humans, no one led to efficacious treatment and final approval (28), what consists of high costs in terms of money and time. Even the most prominent scientists working on these issues and trying to use plasticity to foster brain repair agree that it would be premature to launch clinical trials to use stem cells to treat neurological disorders and that further preclinical studies are needed (26, 27). On the whole, such efforts still clash against the above-mentioned evolutionary constraints, not taking into account an aspect which was underestimated since the beginning: the remarkable differences existing among mammalian species.

DIFFERENCES IN BRAIN PLASTICITY AMONG MAMMALS

Since their discovery in the nineties (29, 30), adult neural stem cells and their continuous renewal of neurons in the

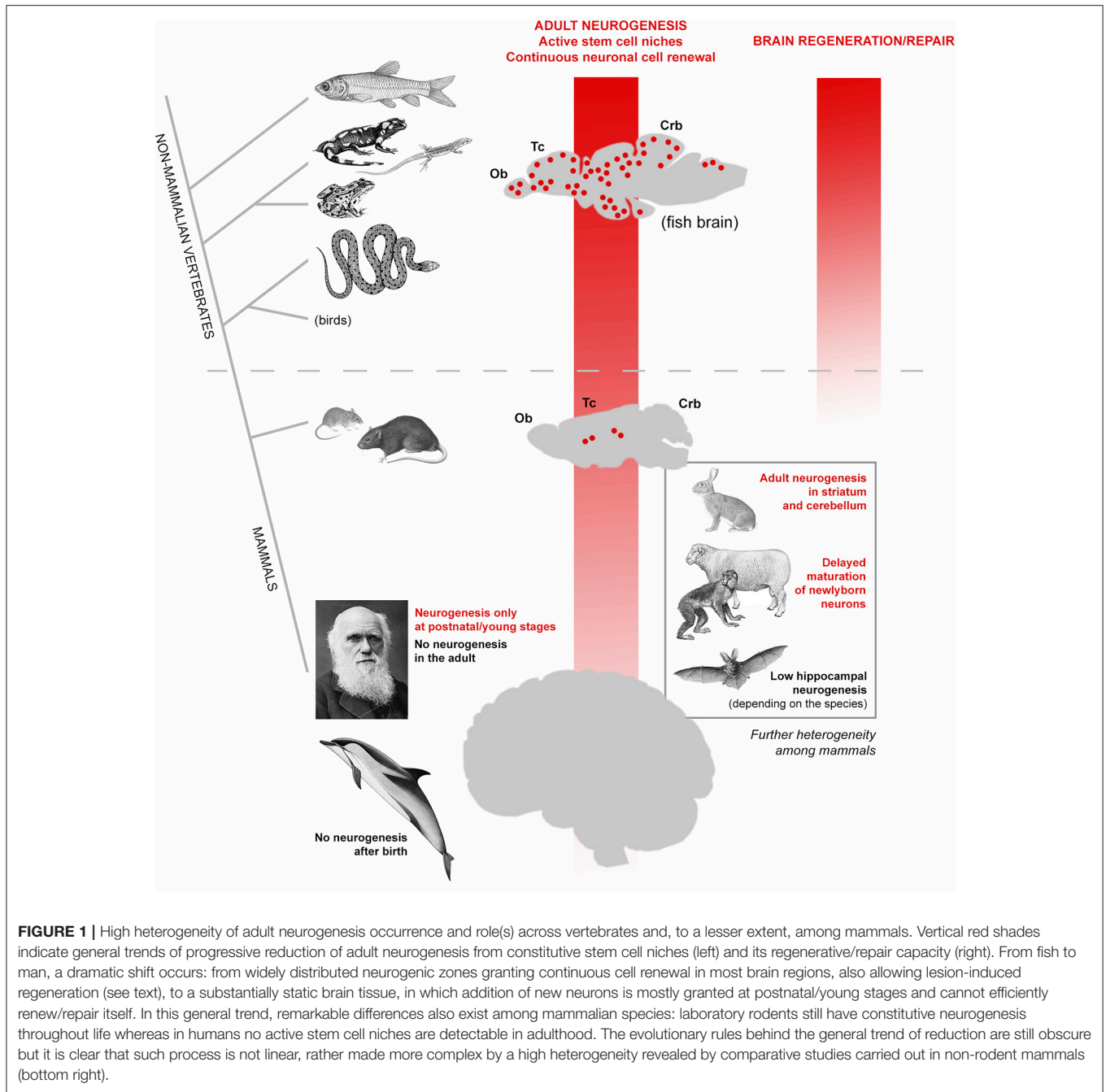


FIGURE 1 | High heterogeneity of adult neurogenesis occurrence and role(s) across vertebrates and, to a lesser extent, among mammals. Vertical red shades indicate general trends of progressive reduction of adult neurogenesis from constitutive stem cell niches (left) and its regenerative/repair capacity (right). From fish to man, a dramatic shift occurs: from widely distributed neurogenic zones granting continuous cell renewal in most brain regions, also allowing lesion-induced regeneration (see text), to a substantially static brain tissue, in which addition of new neurons is mostly granted at postnatal/young stages and cannot efficiently renew/repair itself. In this general trend, remarkable differences also exist among mammalian species: laboratory rodents still have constitutive neurogenesis throughout life whereas in humans no active stem cell niches are detectable in adulthood. The evolutionary rules behind the general trend of reduction are still obscure but it is clear that such process is not linear, rather made more complex by a high heterogeneity revealed by comparative studies carried out in non-rodent mammals (bottom right).

olfactory bulb and hippocampal dentate gyrus of rodents were intensively studied. Such research was fueled by the hope of using plasticity to replace lost/damaged neurons (both from endogenous—constitutive adult neurogenesis—and exogenous—cultured stem cells—sources). The progressive, marvelous results of such research (exceeding now 9,500 scientific papers in a PubMed search for “adult neurogenesis”) undoubtedly led to deep knowledge of the cellular and molecular mechanisms regulating the neural stem cell biology [reviewed in (31, 32)]. In parallel, studies carried out on mammalian species different from mice and rats started to show that adult

neurogenesis occurrence, extension, rate, behavioral role(s), and function(s) can be heterogeneous among mammals [reviewed in (8, 33, 34) **Figure 1**]. Beside striking neurogenic processes in unconventional regions of only some animal species [e.g., the striatum and cerebellum in rabbits; (35–38)], even the apparently conserved hippocampal neurogenesis revealed remarkable differences among mammals, such as in bats [very low levels and high species-specific differences in its rate; (39)] or in dolphins [absence of hippocampal neurogenesis; (40)]. Wild mammals show species-specific, rather stable hippocampal neurogenesis, the rapid adaptation of hippocampal neurogenesis

to experimental challenges being a characteristic of laboratory rodents (41).

Most importantly, adult neurogenesis appears to be highly reduced in humans with respect to rodents: while in mice and rats the genesis of new neurons is a lifelong process, though declining with age, in humans the forebrain and hippocampal stem cell niches are not more detectable at 2 and 7–13 years of age, respectively (42–44). Recent work carried out in dolphins, namely, mammals devoid of olfaction, and brain olfactory structures, showed that no periventricular neurogenesis is detectable after birth in these large-brained, long-living mammals (40, 45). Taken together, the results obtained in human and dolphins strengthen the view that neurogenic processes in mammals are strictly linked to specific physiological functions (e.g., olfaction) rather than to brain repair [see (46)].

THE “IMMATURE NEURON” STORY

In the nineties, in parallel with the first studies on adult neurogenesis, another population of cells which continue to express markers of immaturity throughout life (i.e., doublecortin–DCX–and polysialylated Neural Cell Adhesion Molecule–PSA–NCAM) was discovered in the cortical layer II of the rodent piriform cortex (47–49) (**Figure 2A**). Later, these cells were called “immature” neurons (51), since they are generated prenatally and considered as a potential reservoir of young neurons which might progressively mature to be integrated into circuits of the adult cerebral cortex at different ages, in a sort of “delayed neurogenesis” [hypotheses developed in (52, 53)]. Recently, by using a tamoxifen-inducible transgenic mouse model in which DCX-expressing cells can be permanently labeled with a green fluorescent protein, it was shown that most cortical immature neurons of the mouse piriform cortex actually mature across ages, being no more visible after ceasing to express the immaturity markers (54). Although analyses conducted with different cell and synaptic markers strongly suggest that some of these neurons become structurally integrated [(54, 55), in sheep], a clear and definitive proof that they can function into mature neural circuits is still lacking.

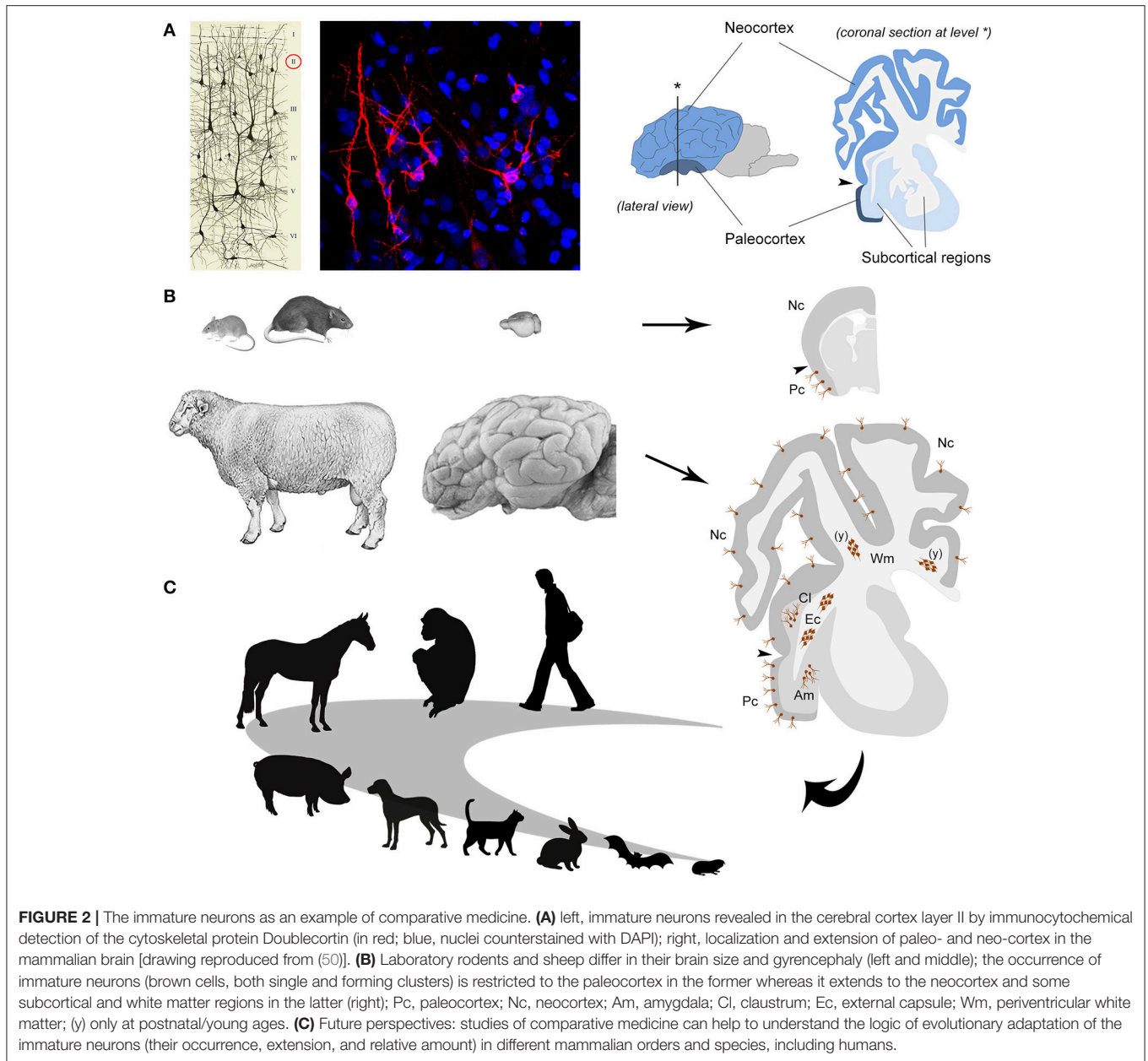
If further studies are needed to go deeper in the molecular mechanisms regulating the mature/immature states of the cells, other urgent questions are open on the side of comparative medicine, since we know from preliminary data that the topographical distribution of immature neurons can be different in some mammalian species. Whereas in mice and rats they are confined to the evolutionarily ancient cortex (paleocortex), in rabbit, guinea pig, some bat and non-human primate species, they have been found to extend into the neocortex (56–61). The comparative studies seem to reveal a turning point in mammalian brain plasticity, suggesting that immature neurons might be far less abundant in the species relying on high rates of lifelong neurogenesis, such as the laboratory rodents.

IMMATURE NEURONS IN SHEEP: THE IMPORTANCE OF COMPARATIVE NEUROSCIENCE

Sheep are mammals showing some features closer to humans than to rodents, such as relatively long lifespan [15–20 years] and larger, gyrencefalic brain [weight: 118–140 grams; gyrification index: 1.94–2.29; (62)]. At the same time, it is still possible to perform experiments usually carried out on mice and rats, including BrdU treatment and brain tissue perfusion, allowing to study neuronal plasticity at a cellular level in physiological and experimental conditions (55, 63). On these bases, we recently investigated the occurrence and anatomical distribution of immature neurons in adult sheep, showing that they are surprisingly abundant in this large-brained mammal with respect to rodents (55). Of importance, they not only extend into the whole neocortex but can also be found in several subcortical regions, including important gray matter nuclei (amygdala and claustrum, involved in emotion-related memories and consciousness) and white matter bundles [(55); **Figure 2B**]. The BrdU treatment (performed both during pregnancy and adulthood) allowed to prove that the DCX+ cells in all those regions were generated prenatally and do not divide in the adult, thus being actually immature neurons. These data support the hypothesis that immature neurons might be more represented in animals with relatively large and “complex” brains with respect to rodents, unlike neurogenic processes which become quite reduced in large-brained, long-living species (8, 64, 65). Furthermore, the sheep study establishes that immature neurons must not be considered restricted to the cerebral cortex layer II, implying that studying the other, subcortical cell populations will require non-rodent animal models.

Among the novel findings emerging from the sheep brain, were also large clusters of doublecortin-positive, non-newly generated cells in the dorsal (periventricular) white matter of very young animals (see **Figure 2B**). The same structures, reminiscent of “trapped” immature neurons, were present in neonatal dolphins (grouped with sheep in the order Cetartiodactyla), then disappearing in both species around puberty (66). This is quite different from what is known to occur in early postnatal brains of rodents and humans, characterized by the migration of isolated neuroblasts toward the cortex. Again, a remarkable heterogeneity of structural plasticity is confirmed in the brain of mammals, here explained by the different stages of development at birth (highly advanced in Cetartiodactyla with respect to rodents and humans).

Hence, studies carried out on non-rodent species reveal that large-brained mammals harbor different types of brain plasticity, especially considering the topographical distribution and amount of the “whole cell” plasticity. On these bases, laboratory rodents (while remaining a good model for molecular and cellular mechanisms) cannot be the right model for studying the outcome of some processes, such as adult neurogenesis and immature neurons, since they are not present in some brain regions and/or occur at different rates in small- and large brained mammals [see (65) for evolutionary hypotheses]. For instance, sheep might be used



to investigate behaviorally/environmentally-induced modulation of immature neurons in neocortical areas, amygdala and claustrum.

FUTURE PERSPECTIVES

Dealing with brain plasticity, the alternative scenarios opened by comparative results with respect to most research conducted on laboratory rodents strongly indicates a need for comparative medicine involving a wide range of animal species, including those of veterinary interest. It is naïve to think that after unraveling the cellular and molecular mechanisms of neural plasticity and brain stem cells in rodent models, the same

outcome in terms of behavior and outcome of their progeny, can be simply obtained in brains which, through million years, have evolved different neuroanatomy, size, regional complexity, functions [see (2, 9)], and in which even the putative existence/role(s) of different types of “natural” plasticity appear to be different (8). The dramatic drop in adult neurogenesis in some long-living mammals is a prominent example for the need for a wider “phylogenetic” view in order to fill the gap between mice and men (64, 65).

Accordingly, the emerging opportunity of immature neurons should be investigated further on by considering a wide range of mammals, including humans (**Figure 2C**). We still lack systematic quantitative, really comparable data on the distribution of these neurons in mammalian species which

widely differ in their neuroanatomy (brain size, gyrencephaly, cortical extension) and other internal/external features (lifespan, ecological niche, domestication vs. wildness, food habit). The question is: are the relative amount and topographical distribution of immature neurons homogeneous across mammals? If not: is there a trend/relationship with some of the above-mentioned anatomical/behavioral features? Moreover, it could be something major at stake. Some studies performed in sheep and macaque monkeys to determine the time course of adult neurogenesis in the olfactory bulb (sheep) and dentate gyrus of the hippocampus (both species), showed that neuroblast maturation is protracted over a minimum of a 3–6 months, respectively, thus resulting delayed several times longer than in rodents (67, 68). These data are in accord with different rates of cell proliferation markers (e.g., Ki-67 antigen) vs. immaturity markers (e.g., doublecortin) detection in rodent and non-rodent species, with a higher persistence of the immaturity markers in the latter [reviewed and discussed in (8)]. Taken together, these observations suggest that long time maturation and accumulation of “immature” neurons might be a feature of long-living species, even within the neurogenic sites. Yet, also in this case, the data available are still fragmentary and mostly qualitative. Only accurate and comparable quantitative analyses across different species could help to establish if this trend can be confirmed, and, in that case, sheep can be useful animal models to unravel the mechanisms of such delayed maturation (as medium-sized domestic animals, they can be employed in behavioral tests and subsequent non-invasive imaging techniques; (69, 70). Understanding whether the young neurons (both newborn and generated pre-natally but “immature”)

can mature differently in widely different mammals might be of paramount importance for translational perspectives.

In conclusion, brain structural “whole cell” plasticity is a typical example for the need of comparative medicine in order to unravel the evolutionary aspects determining different outcomes of the neurogenic/other plastic processes in different mammals, thus avoiding to slip into biases when figuring out translational outcomes based exclusively on the use of laboratory rodents.

AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

FUNDING

MIUR-PRIN2015 (grant 2015Y5W9YP).

ACKNOWLEDGMENTS

We thank MIUR-PRIN2015 (grant 2015Y5W9YP) for financial support, and the University of Turin (PhD program in Veterinary Sciences). We wish to thank several collaborators who across the years contributed, with lab work and/or scientific discussion, to develop our approach on brain plasticity in comparative medicine. Among them: Roberta Parolisi, Matteo Piumatti, Ottavia Palazzo, Alessandra Pecora, Paolo Peretto, Federico Luzzati, Bruno Cozzi, Irmgard Amrein, Lutz Slomianka, Hans-Peter Lipp, Chet Sherwood, Barbara Finlay.

REFERENCES

- Mobasheri A. Comparative medicine in the twenty-first century: where are we now and where do we go from here? *Front Vet Sci.* (2015) 2:2. doi: 10.3389/fvets.2015.00002
- Bolker JA. Animal models in translational research: Rosetta stone or stumbling block? *Bioessays* (2017) 39:1700089. doi: 10.1002/bies.201700089
- Cardiff RD, Ward JM, Barthold SW. ‘Onemedicine - onepathology’: are veterinary and human pathology prepared? *Lab Invest.* (2008) 88:18–26. doi: 10.1038/labinvest.3700695
- Schwabe CW. *Veterinary medicine and human health.* 3rd ed. Philadelphia: Williams & Wilkins, Baltimore, xix (1984). p. 1–680.
- Destoumieux-Garçon D, Mavingui P, Boetsch G, Boissier J, Darriet F, Duboz P, et al. The one health concept: 10 years old and a long road ahead. *Front Vet Sci.* (2018) 5:14. doi: 10.3389/fvets.2018.00014
- Vivar C, van Praag H. Running changes the brain: the long and the short of it. *Physiology* (2017) 32:410–24. doi: 10.1152/physiol.00017.2017
- Martino G, Pluchino S, Bonfanti L, Schwartz M. Brain regeneration in physiology and pathology: the immune signature driving therapeutic plasticity of neural stem cells. *Physiol Rev.* (2011) 91:1281–304. doi: 10.1152/physrev.00032.2010
- Lipp HP, Bonfanti L. Adult neurogenesis in mammals: variations and confusions. *Brain Behav Evol.* (2016) 87:205–21. doi: 10.1159/000446905
- Faykoo-Martinez M, Toor I, Holmes MM. Solving the neurogenesis puzzle: looking for pieces outside the traditional box. *Front Neurosci.* (2017) 11:505. doi: 10.3389/fnins.2017.00505
- Thakur KT, Albanese E, Giannakopoulos P, Jette N, Linde M, Prince MJ, et al. Neurological disorders. In: Patel V, Chisholm D, Dua T, Laxminarayan R, Medina-Mora ME, editors. *Mental, Neurological, and Substance Use Disorders: Disease Control Priorities, 3rd Edn, Vol 4.* Washington, DC: Chapter 5: The International Bank for Reconstruction and Development/The World Bank (2016). p. 87–108.
- Vaupel JW. Biodemography of human ageing. *Nature* (2010) 464:536–42. doi: 10.1038/nature08984
- WHO. (2012). *Dementia: A Public Health Priority.* Geneva: WHO.
- Bonfanti L. From hydra regeneration to human brain structural plasticity: a long trip through narrowing roads. *Sci World J.* (2011) 11:1270–99. doi: 10.1100/tsw.2011.113
- Alunni A, Bally-Cuif L. A comparative view of regenerative neurogenesis in vertebrates. *Development* (2016) 143:741–53. doi: 10.1242/dev.122796
- Ganz J, Brand M. Adult neurogenesis in fish. *Cold Spring Harb Perspect Biol.* (2016) 8:a019018. doi: 10.1101/cshperspect.a019018
- Weil ZM, Norman GJ, De Vries AC, Nelson RJ. The injured nervous system: A Darwinian perspective. *Prog Neurobiol.* (2008) 86:48–59. doi: 10.1016/j.pneurobio.2008.06.001
- Sale A, Berardi N, Maffei L. Environment and brain plasticity: Towards an endogenous pharmacotherapy. *Physiol Rev.* (2014) 94:189–234. doi: 10.1152/physrev.00036.2012
- Bonfanti L. The (real) neurogenic/gliogenic potential of the postnatal and adult brain parenchyma. *ISRN Neurosci.* (2013) 2013:354136. doi: 10.1155/2013/354136
- Feliciano DM, Bordey A, Bonfanti L. Noncanonical sites of adult neurogenesis in the mammalian brain. *Cold Spring Harb Perspect Biol.* (2015) 7:a018846. doi: 10.1101/cshperspect.a018846
- Arvidsson A, Collin T, Kirik D, Kokaia Z, Lindvall O. Neuronal replacement from endogenous precursors in the adult brain after stroke. *Nat Med.* (2002) 8:963–70. doi: 10.1038/nm747

21. Palma-Tortosa S, García-Culebras A, Moraga A, Hurtado OA, Perez-Ruiz A, Durán-Laforet V, et al. Specific features of SVZ neurogenesis after cortical ischemia: a longitudinal study. *Sci Rep.* (2017) 7:16343. doi: 10.1038/s41598-017-16109-7
22. Magnusson JP, Göritz C, Tatarishvili J, Dias DO, Smith EN, Lindvall O, et al. A latent neurogenic program in astrocytes regulated by Notch signaling in the mouse. *Science* (2014) 346:237–41. doi: 10.1126/science.346.6206.237
23. Nato G, Caramello A, Trova S, Avataneo V, Rolando C, Taylor V, et al. Striatal astrocytes produce neuroblasts in an excitotoxic model of Huntington's disease. *Development* (2015) 142:840–5. doi: 10.1242/dev.116657
24. Kyritsis N, Kizil C, Zocher S, Kroehne V, Kaslin J, Freudenreich D, et al. Acute inflammation initiates the regenerative response in the adult zebrafish brain. *Science* (2012) 338:1353–6. doi: 10.1126/science.1228773
25. Iraci N, Leonardi T, Gessler F, Vega B, Pluchino S. Focus on extracellular vesicles: Physiological role and signaling properties of extracellular membrane vesicles. *Int J Mol Sci.* (2016) 17:171. doi: 10.3390/ijms17020171
26. Lindvall O, Kokaia Z. Stem cells in human neurodegenerative disorders—time for clinical translation? *J Clin Invest.* (2010) 120:29–40. doi: 10.1172/JCI40543
27. Scolding NJ, Pasquini M, Reingold SC, Cohen JA. Cell-based therapeutic strategies for multiple sclerosis. *Brain* (2017) 140:2776–96. doi: 10.1093/brain/awx154
28. Donegà M, Giusto E, Cossetti C, Pluchino S. Systemic neural stem cell-based therapeutic interventions for inflammatory CNS disorders. In: Bonfanti L, editor. *Neural Stem Cells: New Perspectives*. Rijeka: INTECH (2013). p. 287–347.
29. Lois C, Alvarez-Buylla A. Long-distance neuronal migration in the adult mammalian brain. *Science* (1994) 264:1145–8.
30. Suhonen JO, Peterson DA, Ray J, Gage FH. Differentiation of adult hippocampus-derived progenitors into olfactory neurons in vivo. *Nature* (1996) 383:624–7. doi: 10.1038/383624a0
31. Aimone JB, Li Y, Lee SW, Clemenson GD, Deng W, Gage FH. Regulation and function of adult neurogenesis: from genes to cognition. *Physiol Rev.* (2014) 94:991–1026. doi: 10.1152/physrev.00004.2014
32. Lim DA, Alvarez-Buylla A. The adult ventricular-subventricular zone (V-SVZ) and olfactory bulb (OB) neurogenesis. *Cold Spring Harb Perspect Biol.* (2016) 8:a018820. doi: 10.1101/cshperspect.a018820
33. Bonfanti L, Peretto P. Adult neurogenesis in mammals – a theme with many variations. *Eur J Neurosci.* (2011) 34:930–50. doi: 10.1111/j.1460-9568.2011.07832.x
34. Amrein I. Adult hippocampal neurogenesis in natural populations of mammals. *Cold Spring Harb Perspect Biol.* (2015) 7:a021295. doi: 10.1101/cshperspect.a021295
35. Luzzati F, De Marchis S, Fasolo A, Peretto P. Neurogenesis in the caudate nucleus of the adult rabbit. *J Neurosci.* (2006) 26:609–21. doi: 10.1523/JNEUROSCI.4371-05.2006
36. Ponti G, Aimar P, Bonfanti L. Cellular composition and cytoarchitecture of the rabbit subventricular zone (SVZ) and its extensions in the forebrain. *J Comp Neurol.* (2006) 498:491–507. doi: 10.1002/cne.21043
37. Ponti G, Peretto P, Bonfanti L. Genesis of neuronal and glial progenitors in the cerebellar cortex of peripuberal and adult rabbits. *PLoS ONE* (2008) 3:e2366. doi: 10.1371/journal.pone.0002366
38. Luzzati F, Peretto P, Aimar P, Ponti G, Fasolo A, Bonfanti L. Glia-independent chains of neuroblasts through the subcortical parenchyma of the adult rabbit brain. *Proc Natl Acad Sci USA.* (2003) 100:13036–41. doi: 10.1073/pnas.1735482100
39. Amrein I, Dechmann DK, Winter Y, Lipp HP. Absent or low rate of adult neurogenesis in the hippocampus of bats (Chiroptera). *PLoS ONE* (2007) 2:e455. doi: 10.1371/journal.pone.0000455
40. Patzke N, Spocter MA, Karlsson KÆ, Bertelsen MF, Haagensen M, Chawana R, et al. In contrast to many other mammals, cetaceans have relatively small hippocampi that appear to lack adult neurogenesis. *Brain Struct Funct.* (2015) 220:361–83. doi: 10.1007/s00429-013-0660-1
41. Amrein I, Slomianka L, Poletava II, Bologova NV, Lipp HP. Marked species and age-dependent differences in cell proliferation and neurogenesis in the hippocampus of wild-living rodents. *Hippocampus* (2004) 14:1000–10. doi: 10.1002/hipo.20018
42. Sanai N, Nguyen T, Ihrie RA, Mirzadeh Z, Tsai H-H, Wong M, et al. Corridors of migrating neurons in the human brain and their decline during infancy. *Nature* (2011) 478:382–6. doi: 10.1038/nature10487
43. Sorrells SF, Paredes MF, Cebrian-Silla A, Sandoval K, Qi D, Kelley KW, et al. Human hippocampal neurogenesis drops sharply in children to undetectable levels in adults *Nature* (2018) 555:377–81. doi: 10.1038/nature25975
44. Cipriani S, Ferrer A, Arinica I, Kovacs G, Verney C, et al. Hippocampal radial glial subtypes and their neurogenic potential in human fetuses and healthy and Alzheimer disease adults. *Cereb Cortex.* (2018) 28:2458–78. doi: 10.1093/cercor/bhy096
45. Parolisi R, Cozzi B, Bonfanti L. (2017). Non-neurogenic SVZ-like niche in dolphins, mammals devoid of olfaction. *Brain Struct Funct.* (2015) 2:2625–39. doi: 10.1007/s00429-016-1361-3
46. Peretto P, Bonfanti L. Adult neurogenesis 20 years later: physiological function vs. brain repair. *Front Neurosci.* (2015) 9:71. doi: 10.3389/fnins.2015.00071
47. Seki T, Arai Y. Expression of highly polysialylated NCAM in the neocortex and piriform cortex of the developing and the adult rat. *Anat Embryol.* (1991) 184:395–401.
48. Bonfanti L, Olive S, Poulain DA, Theodosis DT. Mapping of the distribution of polysialylated neural cell adhesion molecule throughout the central nervous system of the adult rat: an immunohistochemical study. *Neuroscience* (1992) 49:419–36. doi: 10.1016/0306-4522(92)90107-D
49. Nacher J, Crespo C, McEwen BS. Doublecortin expression in the adult rat telencephalon. *Eur J Neurosci.* (2001) 14:629–44. doi: 10.1046/j.0953-816x.2001.01683.x
50. Palazzo O, La Rosa C, Piumatti M, Bonfanti L. Do large brains of long-living mammals prefer non-newly generated, immature neurons? *Neural Regen Res.* (2018) 13:633–4. doi: 10.4103/1673-5374.230282
51. Gómez-Climent MA, Castillo-Gómez E, Varea E, Guirado R, Blasco-Ibáñez JM, Crespo C, et al. A population of prenatally generated cells in the rat paleocortex maintains an immature neuronal phenotype into adulthood. *Cereb Cortex* (2008) 18:2229–40. doi: 10.1093/cercor/bhm255
52. Bonfanti L, Nacher J. New scenarios for neuronal structural plasticity in non-neurogenic brain parenchyma: the case of cortical layer II immature neurons. *Prog Neurobiol.* (2012) 98:1–15. doi: 10.1016/j.pneurobio.2012.05.002
53. König R, Benedetti B, Rotheneichner P, O'Sullivan A, Kreutzer C, Belles M, et al. Distribution and fate of DCX/PSA-NCAM expressing cells in the adult mammalian cortex: a local reservoir for adult cortical neuroplasticity? *Front Biol.* (2016) 11:193–213. doi: 10.1007/s11515-016-1403-5
54. Rotheneichner P, Belles M, Benedetti B, König R, Dannehl D, Kreutzer C, et al. Cellular plasticity in the adult murine piriform cortex: continuous maturation of dormant precursors into excitatory neurons. *Cereb Cortex* (2018) 28:2610–21. doi: 10.1093/cercor/bhy087
55. Piumatti M, Palazzo O, La Rosa C, Crociara P, Parolisi R, Luzzati F, et al. Non-newly generated, “immature” neurons in the sheep brain are not restricted to cerebral cortex. *J Neurosci.* (2017) 38:826–42. doi: 10.1523/JNEUROSCI.1781-17.2017
56. Xiong K, Luo DW, Patrylo PR, Luo XG, Struble RG, Clough RW, et al. Doublecortin-expressing cells are present in layer II across the adult guinea pig cerebral cortex: partial colocalization with mature interneuron markers. *Exp Neurol.* (2008) 211:271–82. doi: 10.1016/j.expneurol.2008.02.003
57. Cai Y, Xiong K, Chu Y, Luo DW, Luo XG, Yuan XY, et al. Doublecortin expression in adult cat and primate cerebral cortex relates to immature neurons that develop into GABAergic subgroups. *Exp Neurol.* (2009) 216:342–56. doi: 10.1016/j.expneurol.2008.12.008
58. Luzzati F, Bonfanti L, Fasolo A, Peretto P. DCX and PSA-NCAM expression identifies a population of neurons preferentially distributed in associative areas of different pallial derivatives and vertebrate species. *Cereb Cortex* (2009) 19:1028–41. doi: 10.1093/cercor/bhn145
59. Zhang XM, Cai Y, Chu Y, Chen EY, Feng JC, Luo XG, et al. Doublecortin-expressing cells persist in the associative cerebral cortex and amygdala in aged nonhuman primates. *Front Neuroanat.* (2009) 3:17. doi: 10.3389/fnro.05.017.2009
60. Chawana R, Patzke N, Alagaili AN, Bennett NC, Mohammed OB, Kaswera-Kyamakya C, et al. The distribution of Ki-67 and doublecortin

- immunopositive cells in the brains of three microchiropteran species, *Hipposideros fuliginosus*, *Triaenops persicus*, and *Asellia tridens*. *Anat Rec.* (2016) 299:1548–60. doi: 10.1002/ar.23460
61. Yang Y, Xie M-X, Li J-M, Hu X, Patrylo PR, Luo X-G, et al. Prenatal genesis of layer II doublecortin expressing neurons in neonatal and young adult guinea pig cerebral cortex. *Front Neuroanat.* (2015) 9:109. doi: 10.3389/fnana.2015.00109
62. Zilles K, Palomero-Gallagher N, Amunts K. Development of cortical folding during evolution and ontogeny. *Trends Neurosci.* (2013) 36:275–84. doi: 10.1016/j.tins.2013.01.006
63. Lévy F, Batailler M, Meurisse M, Migaud M. Adult neurogenesis in sheep: characterization and contribution to reproduction and behavior. *Front Neurosci.* (2017) 11:570. doi: 10.3389/fnins.2017.00570
64. Paredes MF, Sorrells SF, Garcia-Verdugo JM, Alvarez-Buylla A. Brain size and limits to adult neurogenesis. *J Comp Neurol.* (2016) 524:646–64. doi: 10.1002/cne.23896
65. Parolisi R, Cozzi B, Bonfanti L. Humans and dolphins: decline and fall of adult neurogenesis. *Front Neurosci.* (2018) 12:497. doi: 10.3389/fnins.2018.00497
66. La Rosa C, Parolisi R, Palazzo O, Lévy F, Meurisse M, Bonfanti L. Clusters of DCX+ cells “trapped” in the subcortical white matter of early postnatal *Cetartiodactyla* (*Tursiops truncatus*, *Stenella coeruleoalba* and *Ovis aries*). *Brain Struct Funct.* (2018) 223:3613–32. doi: 10.1007/s00429-018-1708-z
67. Kohler SJ, Williams NI, Stanton GB, Cameron JL, Greenough WT. Maturation time of new granule cells in the dentate gyrus of adult macaque monkeys exceeds six months. *Proc Natl Acad Sci USA.* (2011) 108:10326–31. doi: 10.1073/pnas.1017099108
68. Brus M, Meurisse M, Gheusi G, Keller M, Lledo PM, Levy F. Dynamics of olfactory and hippocampal neurogenesis in adult sheep. *J Comp Neurol.* (2013) 521:169–88. doi: 10.1002/cne.23169
69. Muehlemann T, Reefmann N, Wechsler B, Wolf M, Gygax L. In vivo functional near-infrared spectroscopy measures mood-modulated cerebral responses to a positive emotional stimulus in sheep. *Neuroimage* (2011) 54:1625–33. doi: 10.1016/j.neuroimage.2010.08.079
70. Torricelli A, Contini D, Dalla Mora A, Pifferi A, Re R, Zucchelli L, et al. Neurophotonic: non-invasive optical techniques for monitoring brain functions. *Funct Neurol.* (2014) 29:223–30. doi: 10.11138/FNeur/2014.29.4.223

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 La Rosa and Bonfanti. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.