



Atmospheric H₂S: Impact on Plant Functioning

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Hydrogen sulfide (H₂S) is an air pollutant present at high levels in various regions. Plants actively take up H₂S via the foliage, though the impact of the gas on the physiological functioning of plants is paradoxical. Whereas elevated H₂S levels may be phytotoxic, H₂S levels realistic for polluted areas can also significantly contribute to the sulfur requirement of the vegetation. Plants can even grow with H₂S as sole sulfur source. There is no relation between the rate of H₂S metabolism and the H₂S susceptibility of a plant, which suggests that the metabolism of H₂S does not contribute to the detoxification of absorbed sulfide. By contrast, there may be a strong relation between the rate of H₂S metabolism and the rate of sulfate metabolism: foliar H₂S absorbance may downregulate the metabolism of sulfate, taken up by the root. Studies with plants from the *Brassica* genus clarified the background of this downregulation. Simultaneously, these studies illustrated that H₂S fumigation may be a useful tool for obtaining insight in the regulation of sulfur homeostasis and the (signal) functions of sulfur-containing compounds in plants.

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INTRODUCTION

Hydrogen sulfide (H₂S) is a gaseous compound present in the global atmosphere (Watts, 2000). Together with sulfur dioxide (SO₂) and a variety of organo-sulfur gases, H₂S plays a pivotal role in shaping the earth's climate (Sipilä et al., 2010; Perraud et al., 2015). The gas naturally originates from volcanoes and geothermal vents as well as from wetlands, salt marshes and estuaries, where it is produced by bacteria during the anaerobic decay of organic sulfur compounds (Kanda and Tsuruta, 1995; Watts, 2000; Stern, 2005). The estimated natural emission of H₂S is 4.4 Tg per year, which is only a small fraction of the total natural sulfur gas emission, estimated at 52 Tg per year (Watts, 2000). SO₂ and dimethylsulfide (DMS) make up the majority of this emission: 23 and 24.5 Tg per year, respectively (Watts, 2000; Carn et al., 2017). Besides from natural sources, atmospheric H₂S originates from livestock production and industrial processes, such as the combustion of biomass and fossil fuels (Watts, 2000). The anthropogenic emission of sulfur gases is exceeding the natural sulfur gas emission and is currently estimated at 70–100 Tg per year (Klimont et al., 2013; Fioletov et al., 2016). Most of this sulfur is emitted as SO₂, though approximately 3 Tg per year is emitted as H₂S (Klimont et al., 2013; Fioletov et al., 2016).

The residence time of emitted H₂S in the atmosphere is short (approximately 15 days), since H₂S is rapidly oxidized by hydroxyl radicals and other atmospheric oxidants to SO₂ and finally sulfate (Trudinger, 1986). Consequently, in rural areas, H₂S concentrations are ranging between 0.02 and 0.3 nl L⁻¹ (Kellogg et al., 1972; Slatt et al., 1978; Beauchamp et al., 1984).

However, in regions with volcanic activity and in regions with polluting industrial or livestock production, H₂S concentrations may easily surpass the odor threshold level of 0.02 μl L⁻¹ (resulting in a distinct rotten egg smell; Beauchamp et al., 1984). Moreover, in the close vicinity of volcanoes and geothermal wells, atmospheric H₂S concentrations may even exceed 0.1 μl L⁻¹ (Ernst, 1997; Schulte et al., 1997; Baillie et al., 2016).

The impact of elevated H₂S levels on plants is paradoxical. Although high atmospheric H₂S concentrations may negatively affect plant growth and survival, the foliar uptake of H₂S may also substantially contribute to plant sulfur nutrition (De Kok, 1990; De Kok et al., 2007). This concise review presents an overview of the impact of atmospheric H₂S on the physiological functioning of plants.

UPTAKE AND EMISSION OF H₂S BY PLANTS

Atmospheric H₂S may be adsorbed at the leaf's surface, though H₂S uptake mainly occurs via stomata: the cuticle is hardly permeable for gases (Lendzian, 1984). The rate of foliar gas uptake can be described by Fick's law of diffusion: $J = \Delta c \times g$. In this equation, J represents the rate of gas uptake by the shoot (μmol cm⁻² s⁻¹), Δc the concentration gradient of the gas between the atmosphere and the shoot's interior (μmol cm⁻³), and g the diffusive conductance of the shoot to the gas (cm s⁻¹; Nobel, 1983; De Kok and Tausz, 2001; De Kok et al., 2007). The diffusive conductance of the shoot is predominantly determined by stomatal conductance and mesophyll conductance (Nobel, 1983; De Kok and Tausz, 2001; De Kok et al., 2007). Whereas stomatal conductance depends on the extent of stomatal aperture, mesophyll conductance depends on the physical and biochemical characteristics of a gas, *viz.* its solubility in the aqueous phase of the mesophyll, its reactivity with cellular components and in case of some gases (e.g., CO₂) its rate of metabolism (Nobel, 1983; De Kok and Tausz, 2001; De Kok et al., 2007). The diffusive conductance of the shoot is frequently expressed in mmol m⁻² s⁻¹. For conversion to cm s⁻¹, the volume of 1 mol of a gas at 1 atm and 20°C is 24.06 dm³ (Nobel, 1983). This corresponds to 41.6 mol m⁻³. Therefore, a gas conductance of 1 cm s⁻¹ at 1 atm and 20°C equals 41.6 mol m⁻³ × 0.01 m s⁻¹ = 416 mmol m⁻² s⁻¹.

The uptake of H₂S by plant shoots follows distinct kinetics, which greatly differ from the kinetics observed for other sulfur gases. For instance, SO₂ uptake rates generally increase linearly with atmospheric SO₂ concentration (Tausz et al., 1998; Van der Kooij and De Kok, 1998; De Kok and Tausz, 2001). Stomatal conductance is limiting SO₂ uptake rates, since the diffusive conductance of the shoot to SO₂ is often close to the stomatal conductance for water vapor (Tausz et al., 1998; Van der Kooij and De Kok, 1998; De Kok and Tausz, 2001). In accordance with this, the mesophyll conductance to SO₂ is high and therefore not limiting uptake rates. SO₂ has a high solubility in the aqueous phase of the mesophyll: it has a rather high Henry's law solubility constant of 1.23 M/atm at 25°C (De Bruyn et al., 1995). Moreover,

it rapidly reacts with mesophyll water, resulting in the formation of sulfurous acid (Tausz et al., 1998).

In contrast to SO₂, H₂S uptake rates follow saturation kinetics with respect to the atmospheric H₂S level (Figure 1; De Kok et al., 2002, 2007, 2009). These kinetics, which can be described by the Michaelis-Menten equation, are controlled by mesophyll conductance (Figure 1). At non-saturating atmospheric H₂S levels, uptake rates increase linearly with external H₂S concentration, since stomatal conductance is not affected upon exposure to H₂S concentrations <1 μl L⁻¹ (De Kok et al., 1989, 1991, 1997). However, at saturating atmospheric H₂S levels, mesophyll conductance prevents further increments in H₂S uptake rates (De Kok et al., 1989, 1991, 1997).

It is evident that at the pH of mesophyll cells (between 5 and 6.4) absorbed H₂S remains largely undissociated (H₂S → HS⁻ + H⁺; pK_a = 7.0), causing it to easily pass cellular membranes (Cope and Spedding, 1982; Mathai et al., 2009; Riahi and Rowley, 2014). H₂S is only slightly soluble in the mesophyll: it has a Henry's law solubility constant of 0.086 M/atm at 25°C (De Bruyn et al., 1995). However, mesophyll conductance appears to be strictly controlled by the rate of sulfide metabolism in the amino acid cysteine (De Kok et al., 1989, 1991, 1998). After absorbance, H₂S is incorporated with very high affinity in cysteine via the reaction of sulfide with O-acetylserine (OAS), catalyzed by the enzyme O-acetylserine(thiol)lyase (OAS-TL; De Kok et al., 2007, 2009). The activity of OAS-TL, the affinity of the enzyme for sulfide and the availability of OAS determine mesophyll conductance (De Kok et al., 2007, 2009). Consequently, in spinach (*Spinacia oleracea*) H₂S uptake rates were, in contrast to SO₂ uptake rates, strongly dependent on shoot temperature with lower uptake rates at lower temperatures (De Kok et al., 1991). Moreover, in spinach, the maximum H₂S uptake rate (JH₂S_{max}) could be enhanced by the direct supply of OAS

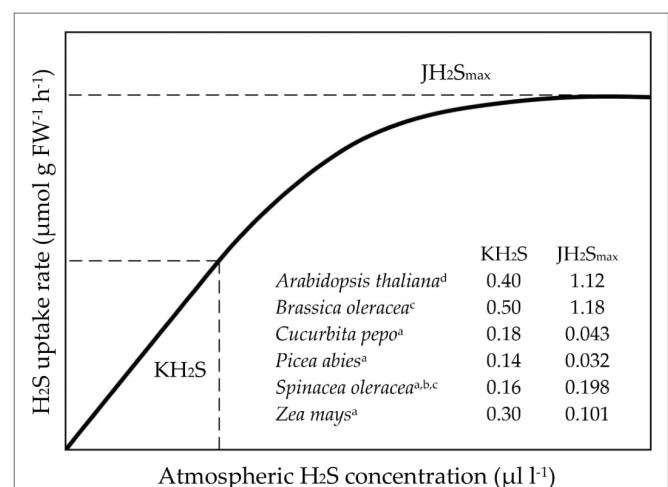


FIGURE 1 | The kinetics of foliar H₂S uptake. JH₂S_{max} represents the maximum uptake rate of H₂S and KH₂S the concentration at which ½JH₂S_{max} is reached. FW: fresh weight. Figure modified after De Kok et al. (2002). Data derived from De Kok et al. (1989^a, 1991^b, 1997^c) and Van der Kooij and De Kok (1998^d).

to foliar tissue (Buwalda et al., 1992). Notably, a second cysteine-producing reaction may further determine mesophyll conductance. Cysteine desulhydrases (DES) have significance in the degradation of cysteine, which results in the endogenous release of sulfide (Schütz et al., 1991). However, circumstantial evidence suggests that the reverse reaction may be relevant for foliar H₂S absorbance: in leaf homogenates of cucurbit plants (*Cucurbita pepo* spp.) DES assimilated atmospheric H₂S in cysteine by using ammonia and pyruvate as substrates (Schütz et al., 1991). Nevertheless, the significance of this reaction for intact plants remains to be studied (De Kok et al., 1998, 2007).

Maximum H₂S uptake rates and KH₂S values (*viz.* the H₂S concentration at which ½JH₂S_{max} is reached) differ considerably among species (Figure 1; De Kok et al., 2002). For example, whereas measured KH₂S values ranged from 0.14 to 0.50 μL⁻¹ H₂S, maximum H₂S uptake rates varied between 0.03 and 1.18 μmol H₂S g FW⁻¹ h⁻¹ (Figure 1). This variation corresponded with variation in the rate of sulfide incorporation in cysteine (De Kok et al., 2007). However, it remains elusive to what extent it also coincides with variation in sulfur growth requirement among species. It is unknown if, for instance, species with a high sulfur demand also have high H₂S uptake efficiencies.

Besides H₂S uptake, plants may also emit H₂S. It has been suggested that emission of H₂S to the atmosphere has significance in regulating sulfur homeostasis (Schröder, 1993). Plants may, temporarily, emit elevated levels of H₂S via their foliage into the atmosphere when exposed to excess sulfur in the form of SO₂, sulfate or cysteine (Rennenberg, 1984; De Kok, 1990; Schröder, 1993; Bloem et al., 2007). For instance, depending on the atmospheric SO₂ level, up to 15% of foliarly absorbed SO₂ could be re-emitted as H₂S (De Kok, 1990). However, to what extent H₂S evolution has significance in regulating sulfur homeostasis in the absence of excess sulfur is unclear. If plants were grown with a normal sulfur supply, the H₂S evolution rate generally constituted a negligible fraction of the total sulfur assimilation rate (Stulen and De Kok, 1993). For instance, in spruce (*Picea abies*), this fraction was less than 0.1% (Rennenberg et al., 1990). More likely, the rate of foliar H₂S emission just reflects the rate of sulfate reduction as well as the activity and sulfide-affinity of the cysteine synthesizing and degrading enzymes (*viz.* OAS-TL and DES, possibly together with other enzymes; Hell et al., 2002). Irrespective of this, minute H₂S emission levels might still have physiological significance in, for instance, plant stress protection. Emitted H₂S might possibly degrade leaf surface ozone (O₃; Schnug, 1997; Haneklaus et al., 2007). Furthermore, H₂S emission may have importance in the defense of plants against pathogen attacks (Bloem et al., 2007; Haneklaus et al., 2007).

PHYTOTOXICITY OF H₂S

The foliar absorbance of atmospheric H₂S may negatively affect plant functioning. H₂S is a very reactive compound and, similar to cyanide, it complexes with high affinity to the

metallo-groups in proteins (Mudd, 1979; Beauchamp et al., 1984; Martin and Maricle, 2015). In several plants, exposure to H₂S inhibited respiration, which could be explained by the reaction of H₂S with the heme-group of cytochrome *c* oxidase (Martin and Maricle, 2015). Additionally, in various plants the presence of H₂S repressed the activity of a broad group of (likely heme-containing) NADH-oxidizing enzymes (Maas and De Kok, 1988). Notably, these repressions may directly result in a lower plant growth and survival, but also indirectly: by disturbing energy homeostasis, H₂S presence may increase the susceptibility of plants for other environmental stressors. For instance, exposure to 0.25 μL⁻¹ H₂S decreased the freezing tolerance of the foliage of winter wheat (*Triticum aestivum*; Stuiver et al., 1992).

H₂S presence may also affect the activity of enzymes involved in photosynthetic CO₂ fixation and photosynthetic electron transport (Oliva and Steubing, 1976; Coyne and Bingham, 1978; De Kok et al., 1983a; Taylor and Selvidge, 1984). In isolated spinach chloroplasts H₂S exposure inhibited the photoreduction of NADP⁺ and upon illumination it initiated oxygen uptake by the chloroplasts (De Kok et al., 1983a). Sulfide-induced oxygen uptake by chloroplasts was sensitive to the herbicide DCMU (3-(3,4-dichlorophenyl)-1,1-dimethylurea) and prevented by the addition of superoxide dismutase to the chloroplast suspension. This indicated that sulfide was oxidized by chloroplasts, its oxidation being initiated by superoxide formed upon illumination at the reducing side of photosystem I (De Kok et al., 1983a). Nevertheless, reductions in photosynthesis are unlikely to be the primary basis of H₂S toxicity, since generally in intact plants photosynthesis was only reduced after prolonged exposure to toxic H₂S concentrations (Maas et al., 1987a, 1988; De Kok, 1989, 1990).

There is a large variation in H₂S susceptibility between species as well as between cultivars of the same species. Whereas continuous exposure to 0.03 μL⁻¹ H₂S (a level realistic for regions with industrial or agricultural pollution) negatively affected the growth of susceptible plants (e.g., various spinach cultivars), it stimulated the growth of several other plants on sulfur-sufficient soils [e.g., lettuce (*Lactuca sativa*), alfalfa (*Medicago sativa*), and sugar beet (*Beta vulgaris*); Thompson and Kats, 1978; De Kok, 1989, 1990]. Moreover, whereas in some plants (e.g., lettuce, sugar beet, and common grape vine; *Vitis vinifera*), visible leaf injury developed upon prolonged exposure to 0.3 μL⁻¹ H₂S (a level realistic for areas nearby, e.g., volcanoes), other plants remained unaffected (Thompson and Kats, 1978; De Kok, 1989, 1990). A small number of plants could actually tolerate H₂S levels as high as 20 μL⁻¹ [e.g., maiden silvergrass (*Miscanthus sinensis*), which inhabits zones very close to volcanoes; Mudd, 1979]. However, at higher H₂S levels, all plants quickly developed severe leaf necrosis and rapidly started wilting (Mudd, 1979).

Variation in susceptibility to the negative effects of H₂S may partly be associated with differences in the impact of H₂S on energy homeostasis. The *in vitro* cytochrome *c* oxidase activity was less affected by H₂S exposure in flooding-tolerant than in flooding-sensitive species (Martin and Maricle, 2015). Flooding can induce H₂S formation in soils, which may

explain this variation (Martin and Maricle, 2015). Furthermore, in spinach, the more susceptible a plant was for H₂S, the more the *in vitro* NADH oxidation capacity of shoots was decreased upon H₂S exposure (Maas and De Kok, 1988). In addition, in experiments with other plant species in which growth was not affected by H₂S presence, also the *in vitro* NADH-oxidation capacity was not affected (Stulen et al., 1990). Differences in H₂S tolerance could also be related to differences in plant morphology. In general, dicots appeared to be more susceptible to atmospheric H₂S than monocots (Stulen et al., 1990, 2000). In monocots, H₂S can hardly penetrate the shoot meristem, because the meristem is sheltered by a whorl of leaves (Stulen et al., 1990, 2000). This may cause monocots to be relatively H₂S tolerant, since carefully uncovering the shoot meristem of maize (*Zea mays*) increased its susceptibility to H₂S (Stulen et al., 1990, 2000). Whereas this uncovering did not affect the elongation rate of leaves, it delayed the initial leaf development from the meristem and it triggered cell deformations as well as chromosomal irregularities inside the meristem (Stulen et al., 1990, 2000). Notably, it remains elusive if other morphological traits (e.g., traits associated with leaf anatomy) also explain variation in H₂S phytotoxicity. Furthermore, it remains elusive if life-history traits (e.g., being annual or perennial) contribute to this variation. However, by contrast, it has been observed that variation in H₂S phytotoxicity is not directly interrelated to the capacity of a plant to metabolize sulfide in organic compounds (see the next section).

IMPACT OF H₂S ON PLANT SULFUR METABOLISM

Besides reducing growth and survival, plants may also benefit from the presence of atmospheric H₂S. Since sulfide is a substrate for cysteine synthesis, the gas can be used to synthesize proteins and other organic compounds (De Kok et al., 2007, 2009; Koralewska et al., 2008). Plants can even grow with atmospheric H₂S as the only sulfur source (*viz.* in the absence of root sulfate supply; Buchner et al., 2004; Aghajanzadeh et al., 2016). Atmospheric H₂S levels of 0.06 $\mu\text{L L}^{-1}$ were already sufficient to fully cover the organic sulfur requirement of curly kale (*Brassica oleracea*; Buchner et al., 2004). Since curly kale is characterized by an extraordinary high sulfur demand, H₂S levels realistic for polluted regions (e.g., regions with volcanic activity or intensive animal farming) may significantly contribute to the sulfur requirement of plants in general (Aghajanzadeh et al., 2014, 2015, 2016).

Typically, a significant part of absorbed H₂S (up to 30%) can be revealed in plants as water-soluble non-protein thiols (De Kok, 1990; Poortinga and De Kok, 1997). In shoots the content of these metabolites rapidly and strongly increases upon H₂S exposure (De Kok et al., 1983b, 1985; Maas et al., 1985; Poortinga and De Kok, 1997; Riemenschneider et al., 2005; Shahbaz et al., 2013). In the absence of atmospheric H₂S, water-soluble non-protein thiols constitute 2–4% of total sulfur present in tissues (Noctor et al., 2012). Generally, glutathione is the

most abundant water-soluble non-protein thiol with its content accounting for more than 90% of the total water-soluble non-protein thiol pool (Noctor et al., 2012). Glutathione is synthesized from cysteine in a two-step process. Cysteine first reacts with glutamate to yield γ -glutamylcysteine, which subsequently reacts with glycine to yield glutathione (Hawkesford and De Kok, 2006).

Depending on the species and cultivar, H₂S exposure increased shoot water-soluble non-protein thiol levels up to five-fold (De Kok et al., 1983b, 1985; Maas et al., 1985; Poortinga and De Kok, 1997; Riemenschneider et al., 2005; Shahbaz et al., 2013). Generally, thiol accumulations were stronger at higher atmospheric H₂S levels and the accumulations were not affected by plant age, exposure temperature or the applied light regime (De Kok, 1990). Thiol levels often started to increase within one or 2 h after the onset of H₂S exposure and maximum thiol levels were usually reached after one or 2 days of exposure, independent from the applied H₂S concentration (De Kok et al., 1985; Maas et al., 1987b,c; Stuijver et al., 1992; Poortinga and De Kok, 1997). Generally, thiol accumulations could not solely be attributed to enhanced glutathione levels. For instance, a 12-h exposure of different plant species to 0.25 $\mu\text{L L}^{-1}$ H₂S resulted not only in a two- to three-fold increase in shoot glutathione content, but also in a 8- to 37-fold increase in shoot cysteine content (Buwalda et al., 1988, 1993, 1994). Additionally, in the dark, H₂S exposure resulted in an accumulation of γ -glutamylcysteine (up to 20-fold) due to a limitation in glycine availability, which was caused by the absence of photorespiration in the dark (Buwalda et al., 1988, 1993). However, upon transition to the light, accumulated γ -glutamylcysteine rapidly disappeared as it was metabolized into glutathione (Buwalda et al., 1988, 1993).

Besides altering shoot thiol pools, H₂S exposure occasionally also alters root thiol pools. However, upon H₂S fumigation root thiol pools usually increased significantly less than shoot thiol pools (maximum two-fold) and the increases could often fully be ascribed to enhanced glutathione levels (De Kok et al., 1997; Poortinga and De Kok, 1997; Stuijver and De Kok, 1997; Tausz et al., 1998; Westerman et al., 2000a). In general, glutathione is predominantly present in plants in its reduced form (e.g., in spinach leaves >84%; De Kok et al., 1986). In spinach, the ratio between reduced and oxidized glutathione as well as the activity of glutathione reductase were not substantially altered by exposure to 0.25 $\mu\text{L L}^{-1}$ H₂S (De Kok et al., 1986; Tausz et al., 1998).

After termination of H₂S exposure, thiol levels decrease. In spinach, cysteine and glutathione levels in shoots decreased simultaneously (De Kok et al., 1985, 1986; Maas et al., 1987b; Buwalda et al., 1994). Moreover, a transition from dark to light immediately after the end of H₂S exposure resulted in a decrease in accumulated γ -glutamylcysteine and a simultaneous increase in glutathione, after which both cysteine and glutathione contents decreased at similar rates (Buwalda et al., 1994). Independent from the H₂S concentration that plants were exposed to, thiol levels restored to the levels of unexposed plants within 1 or 2 days (Buwalda et al., 1994). In spinach,

decreases in thiol levels were not associated with the foliar emission of H₂S, indicating that desulfhydration of cysteine was not involved in the loss of thiols (De Kok, 1989, 1990). Apparently, accumulated thiols were rapidly metabolized in other compounds.

It is yet unclear why H₂S exposure strongly enhances the size of the thiol pool. H₂S may partly be metabolized in cysteine and glutathione in another subcellular compartment than sulfate taken up by the root (*viz.* in the mitochondria and/or cytosol instead of the chloroplasts). In this way, the metabolism of atmospheric H₂S could be beyond the control of existing regulatory feedback mechanisms (Hesse et al., 1997; Saito et al., 1997).

It may be hypothesized that variation in the rate of sulfide incorporation in cysteine, glutathione, and other organic compounds (partly) explains variation in H₂S tolerance between plants. However, there is no relation between the H₂S uptake kinetics and the H₂S susceptibility of plants (*viz.* H₂S phytotoxicity; De Kok et al., 1989, 1998, 2002). Moreover, whereas in thale cress (*Arabidopsis thaliana*) OAS-TL knock-out mutants differed in leaf cysteine and glutathione concentrations from wild-type plants (when exposed to atmospheric H₂S concentrations up to 1 μL⁻¹), leaf sulfide concentrations and sulfide tolerance were not different (Birke et al., 2015). Seemingly, the metabolism of sulfide in organic compounds is not involved in the detoxification of absorbed H₂S. It is thus unlikely that differences in the capacity to metabolize H₂S explain differences in the phytotoxicity of H₂S.

It is also unlikely that changes in the size and composition of the thiol pool are directly explaining the phytotoxicity of H₂S. Enhanced glutathione levels may potentially deregulate the activity of various enzymes and cysteine may possibly react with metabolic aldehydes (Rennenberg, 1981). Nevertheless, various plants (especially monocots) can tolerate strongly elevated shoot cysteine and glutathione levels in the presence of H₂S without any negative impact on biomass production, even after prolonged H₂S exposure (De Kok, 1989, 1990; Stulen et al., 1990, 2000). In this perspective, it is noteworthy that enhanced glutathione levels may actually also be beneficial for plants. Glutathione has antioxidant properties and elevated glutathione levels may consequently increase the tolerance of plants to environmental stress (Noctor et al., 2012). However, the physiological significance of increased glutathione levels for plant stress protection remains elusive. Exposure of Chinese cabbage (*Brassica pekinensis*) to copper reduced biomass production (Shahbaz et al., 2014). The copper tolerance of Chinese cabbage was not affected by fumigation with subtoxic H₂S levels, even though this strongly enhanced water-soluble non-protein thiol levels (Shahbaz et al., 2014). Furthermore, in maize, the increased thiol levels upon subtoxic H₂S fumigation could not counteract the negative impact of salinity on growth (Ausma et al., 2017). Finally, exposure to subtoxic H₂S levels did not affect the freezing tolerance of the foliage of winter wheat (Stuiver et al., 1992).

Besides affecting water-soluble non-protein thiol levels, H₂S exposure may affect the levels of other sulfur metabolites.

For instance, in onion (*Allium cepa*) H₂S exposure resulted in an accumulation of secondary sulfur compounds (*viz.* alliinins or their precursors) in the shoot (Durenkamp et al., 2005). Similar to the thiol accumulations, onion could show strongly elevated levels of these compounds without any negative growth effects (Durenkamp et al., 2005). Thale cress accumulated thiosulfate in the shoot upon exposure to a high H₂S level (1 μL⁻¹; Birke et al., 2015). However, by contrast, exposure to a lower H₂S concentration (0.25 μL⁻¹) did not affect the thiosulfate content of several crop species (Buwalda et al., 1993). This suggests that thiosulfate formation is likely also not connected to the phytotoxicity of H₂S. In some species, H₂S exposure resulted in increased shoot sulfate levels (Durenkamp and De Kok, 2002, 2004, 2005; Durenkamp et al., 2005, 2007). Sulfate accumulation presumably occurs in the vacuole and is, therefore, probably likewise not involved in H₂S toxicity (Durenkamp et al., 2005). Enhanced sulfate levels upon H₂S exposure may be caused by the degradation of excessively accumulated organic sulfur compounds and/or by the oxidation of absorbed H₂S (via sulfide oxidases or via non-enzymatic processes; Durenkamp et al., 2005). Alternatively, it may be caused by a poor regulatory control of H₂S over the uptake and assimilation of sulfate (Durenkamp et al., 2005). However, in a plethora of tested plants, *viz.* common duckweed (*Lemna minor*), spinach, tobacco (*Nicotiana tabacum*), and *Brassica* species, there was a good regulatory control of H₂S over the metabolism of sulfate: in these plants H₂S absorbance downregulated sulfate uptake and assimilation (Brunold and Erismann, 1975; Herschbach et al., 1995a,b; De Kok et al., 2002). The background of this phenomenon has most extensively been investigated in *Brassica* seedlings.

IMPACT OF H₂S ON THE UPTAKE AND ASSIMILATION OF SULFATE IN BRASSICA

The genus *Brassica* contains various agriculturally relevant crops. *Brassica* species are characterized by a high content of secondary sulfur compounds (*viz.* glucosinolates), leading to an extraordinary high sulfur demand for growth (Aghajanzadeh et al., 2014, 2015). Additionally, *Brassica* species are characterized by a high tolerance to atmospheric H₂S. Biomass production of *Brassica* seedlings was only significantly reduced upon prolonged exposure to ≥0.4 μL⁻¹ H₂S (Westerman et al., 2000a).

Similar to observations for other species, in *Brassica*, cysteine and glutathione levels increased in the shoot upon H₂S exposure (at 0.8 μL⁻¹ H₂S, approximately 12- and 3.5-fold, respectively), whereas their content in the root was hardly affected (De Kok et al., 2000; Buchner et al., 2004; Koralewska et al., 2008). In *Brassica*, H₂S fumigation did additionally hardly affect the composition and size of the glucosinolate pool (Aghajanzadeh et al., 2014, 2015).

Brassica species may switch from sulfate taken up by the root to H₂S taken up by the shoot as sulfur source (De Kok et al., 1997, 2000; Westerman et al., 2000a,b; Buchner et al., 2004;

Koralewska et al., 2008). If these species were grown on a sulfate-rich medium, H₂S exposure caused a partial downregulation in the activity of transporters involved in root sulfate uptake and sulfate distribution to the shoot (Figure 2; De Kok et al., 1997, 1998, 2000; Westerman et al., 2000a,b; Buchner et al., 2004; Koralewska et al., 2008). This downregulation occurred within a few days after the onset of H₂S exposure and the extent of the downregulation was dependent on the applied H₂S concentration (Westerman et al., 2000a). However, in curly kale, this downregulation was never greater than 60% (reached at 0.2 μL⁻¹ H₂S; Westerman et al., 2000a). Higher repressions appeared unnecessary, since in curly kale the organic sulfur fraction constitutes approximately 60% of total sulfur, whereas sulfate makes up for the other 40% (De Kok et al., 2000; Westerman et al., 2000a). Consequently, the total sulfur and sulfate content of *Brassica* were usually hardly affected by H₂S fumigation (De Kok et al., 1997, 2000; Westerman et al., 2000a,b; Buchner et al., 2004; Koralewska et al., 2008).

Besides repressing sulfate uptake, H₂S exposure may lower sulfate assimilation rates (Figure 2). Briefly, during sulfate assimilation, which is located in plastids of both the root and shoot, sulfate is first converted to adenosine 5'-phosphosulfate (APS) by the enzyme ATP sulfurylase (ATPS; Hawkesford and De Kok, 2006). The majority of APS is

subsequently reduced to sulfite by the enzyme APS reductase (APR), which controls the rate of sulfate assimilation (Hawkesford and De Kok, 2006). Sulfite is, in-turn, reduced to sulfide by the enzyme sulfite reductase (SIR; Hawkesford and De Kok, 2006). Finally, sulfide is incorporated in the amino acid cysteine via a reaction with *O*-acetylserine (OAS), catalyzed by OAS-TL (Hawkesford and De Kok, 2006). In *Brassica*, exposure to H₂S decreased both the activity and expression of the enzyme APR in shoots and roots (up to 80% at 0.8 μL⁻¹ H₂S in curly kale; Westerman et al., 2001; De Kok et al., 2002; Aghajanzadeh et al., 2016). The expression and activity of the other enzymes involved in sulfate assimilation viz. ATPS, SIR, and OAS-TL were usually hardly affected by H₂S exposure (De Kok et al., 2000; Stuijver and De Kok, 2001; Westerman et al., 2001).

Evidently, in *Brassica*, there is a good regulatory control of H₂S over the utilization of sulfate. However, the shoot-to-root signals via which H₂S downregulates sulfate utilization remain elusive. Though it was proposed that glutathione, sulfate, and/or compounds from nitrogen and carbon metabolism (e.g., amino acids and carbohydrates) may be signal compounds, in *Brassica*, there were no clear correlations between the levels of these metabolites, the activity of the sulfate transporters and the activity of APR (Westerman et al., 2000a; Koralewska et al., 2008; Shahbaz et al., 2014). Yet, it was evident that root sulfate uptake in *Brassica* is strongly controlled by the sink capacity (viz. the sulfur status) of the shoot (Koralewska et al., 2009). Furthermore, it was clear that H₂S presence downregulates sulfate uptake via transcriptional and posttranscriptional mechanisms (Koralewska et al., 2008). Decreases in sulfate uptake capacity upon H₂S fumigation did not directly correlate with decreases in the expression of sulfate transporter 1;2 (Sultr1;2), which is in *Brassica* – at an ample sulfate supply – the main transporter responsible for root sulfate uptake (Koralewska et al., 2008).

When *Brassica* plants are deprived of sulfur, they induce multiple responses enabling an enhanced sulfur use efficiency. Sulfur deprivation rapidly induced the expression of transporters involved in root sulfate uptake, the transport of sulfate to the shoot and the vacuolar exchange of sulfate (Koralewska et al., 2007, 2008). At sulfate-sufficient conditions, generally only Sultr1;2 is responsible for root sulfate uptake (Buchner et al., 2004). However, sulfur deprivation resulted not only in an enhanced expression of Sultr1;2 but also in a strongly enhanced expression of sulfate transporter 1;1 (Sultr1;1; Buchner et al., 2004; Koralewska et al., 2007, 2008). The upregulated expression of these two sulfate transporters was accompanied by an increased sulfate uptake capacity of the root (Buchner et al., 2004; Koralewska et al., 2007, 2008). Additionally, sulfur deprivation quickly enhanced the expression and activity of APR in both roots and shoots (Koralewska et al., 2007, 2008; Shahbaz et al., 2014; Aghajanzadeh et al., 2016). Moreover, prolonged sulfur deprivation resulted in an altered shoot-to-root biomass partitioning in favor of that of the root (De Kok et al., 1997; Koralewska et al., 2007, 2008). In the absence of a sulfur supply, these changes were

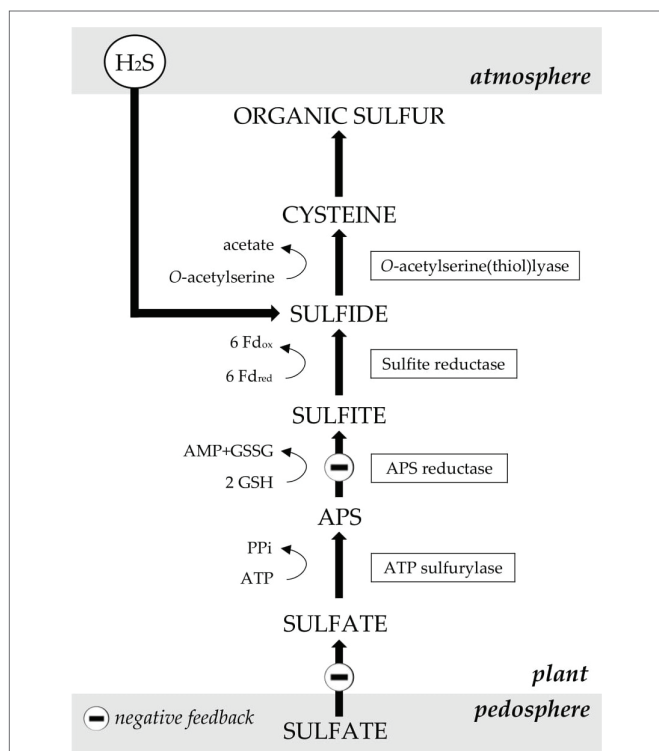


FIGURE 2 | The metabolism of sulfate and the presumed sites in *Brassica* at which H₂S absorbance downregulates sulfate uptake and assimilation (at an ample sulfate supply). APS: adenosine 5'-phosphosulfate; Fd_{red}, Fd_{ox}: reduced and oxidized ferredoxin; GSH, GSSG: reduced and oxidized glutathione. Figure modified after De Kok et al. (2009).

accompanied by decreases in sulfate and thiol content (De Kok et al., 1997; Buchner et al., 2004; Aghajanzadeh et al., 2016).

If sulfur-deprived *Brassica* plants were fumigated with $\geq 0.06 \mu\text{L}^{-1}$ H₂S, growth rate generally restored to the level of plants grown in the presence of sulfate (De Kok et al., 1997; Westerman et al., 2000a,b; Buchner et al., 2004; Aghajanzadeh et al., 2016). Additionally, the enhanced expression of APR and the sulfate transporters in the shoot was largely alleviated (Buchner et al., 2004; Aghajanzadeh et al., 2016). The levels of sulfate in these plants remained low, but the water-soluble non-protein thiol content of the shoot restored to the level of sulfate-sufficient plants (though the level in the roots remained slightly lower; Buchner et al., 2004; Aghajanzadeh et al., 2016). This confirmed that at atmospheric H₂S levels $\geq 0.06 \mu\text{L}^{-1}$ foliarly absorbed sulfide could fully replace sulfate taken up by the root as sulfur source for growth (De Kok et al., 1997; Westerman et al., 2000a,b; Buchner et al., 2004; Aghajanzadeh et al., 2016). Nevertheless, atmospheric H₂S exposure of sulfur-deprived plants had little effect on the expression of APR in the root (Buchner et al., 2004; Koralewska et al., 2008; Shahbaz et al., 2014). Furthermore, the expression of Sultr1;1 and Sultr1;2 as well as the sulfate uptake capacity of sulfur-deprived H₂S-fumigated plants were similar to those of sulfur-deprived plants (Buchner et al., 2004; Koralewska et al., 2008). In addition, the decrease in shoot-to-root biomass partitioning remained largely unaffected (Koralewska et al., 2008). Apparently, when no sulfate is present in the root environment, there is a poor shoot-to-root signaling for the regulation of sulfate utilization in *Brassica*. This suggests that the uptake and subsequent metabolism of sulfate in *Brassica* are at least partly controlled by the sulfate concentration in the root environment (besides the sulfur status of the plant itself).

CONCLUDING REMARKS

The impact of atmospheric H₂S on vegetation is paradoxical. On the one hand, H₂S presence may negatively affect plant growth and survival. On the other hand, plants can use the gas as a sulfur source for growth. H₂S levels found in polluted regions can significantly contribute to the sulfur demand of plants. There is no relation between the rate of H₂S metabolism in organic compounds and the H₂S susceptibility of plants, which suggests that metabolizing H₂S does not constitute a strategy to detoxify absorbed sulfide. By contrast, there may be a strong relation between the rate of H₂S metabolism and the rate of sulfate metabolism. The uptake and metabolism of H₂S may strongly downregulate the uptake and metabolism of sulfate.

Studies with *Brassica* have clarified the background of this downregulation. Simultaneously, these investigations illustrated that H₂S fumigation may be a useful tool for studying the regulation of sulfur homeostasis in plants. H₂S exposure induces changes in the expression and activity of enzymes involved in sulfate metabolism. Moreover, it alters the levels of sulfur metabolites. However, at subtoxic H₂S concentrations, biomass production is not affected. Therefore, at subtoxic H₂S

concentrations, changes in metabolic status are not the consequence of changes in growth (*viz.* the result of growth concentration or dilution), but instead the direct consequence of changes in sulfur utilization. Thus, relating changes in, e.g., metabolite content to changes in enzyme expression and activity can be used to unravel signal transduction pathways that control sulfur homeostasis.

In this way, H₂S fumigation may not only further clarify the regulation of sulfur metabolism in *Brassica*, but also the regulation of sulfur metabolism in other plants. For instance, it may clarify the regulation of sulfate uptake in monocots, in which Sultr1;1 appears to be mainly responsible for the primary uptake of sulfate (instead of Sultr1;2 as in *Brassica*; Buchner et al., 2010). Moreover, fumigation studies may help to elucidate the regulation of sulfate assimilation in C₄ plants. Research with the genus *Flaveria*, which contains C₃ and C₄ species, indicated that C₄ species may have a higher demand for reduced sulfur than C₃ species and that sulfate assimilation in C₄ species may have shifted to the roots compared to C₃ species (Gerlich et al., 2018).

The application of subtoxic H₂S levels may additionally help to clarify the physiological significance of sulfur in plants. Analogous to its function in animal physiology, it has been suggested that endogenous sulfide might also in plants function as a signal molecule that modulates plant immunity, senescence, and various other processes (reviewed by Li et al., 2016; Aroca et al., 2018; Hancock, 2018). However, in several studies that addressed the role of endogenous sulfide, plants were cultivated under suboptimal conditions (e.g., under low light intensities) and/or exposed to relatively high concentrations of sodium hydrosulfide (NaHS). If NaHS is added to a nutrient or tissue incubation solution at neutral pH, it will result in a short-term burst of H₂S, followed by the release of H₂S into the atmosphere (Lee et al., 2011). This means that plants are briefly exposed to relatively high H₂S levels in both the root and shoot environment, which may not only be phytotoxic, but which also may interfere with the regulation of sulfur homeostasis (De Kok et al., 2007; Martin and Maricle, 2015). Evidently, H₂S fumigation of plants provides another way to study the physiological significance of endogenous sulfide as a signal molecule. Foliar H₂S application may also be useful to investigate if organic sulfur compounds modulate physiological processes, since upon H₂S fumigation the content of several of these compounds may be strongly altered.

AUTHOR CONTRIBUTIONS

TA and LK have both made substantial, direct, and intellectual contributions to this work. Both approved it for publication.

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REFERENCES

- Aghajanzadeh, T., Hawkesford, M. J., and De Kok, L. J. (2014). The significance of glucosinolates for sulfur storage in Brassicaceae seedlings. *Front. Plant Sci.* 5:704. doi: 10.3389/fpls.2014.00704
- Aghajanzadeh, T., Hawkesford, M. J., and De Kok, L. J. (2016). Atmospheric H₂S and SO₂ as sulfur sources for *Brassica juncea* and *Brassica rapa*: regulation of sulfur uptake and assimilation. *Environ. Exp. Bot.* 124, 1–10. doi: 10.1016/j.envexpbot.2015.12.001
- Aghajanzadeh, T., Kopriva, S., Hawkesford, M. J., Koprivova, A., and De Kok, L. J. (2015). Atmospheric H₂S and SO₂ as sulfur source for *Brassica juncea* and *Brassica rapa*: impact on the glucosinolate composition. *Front. Plant Sci.* 6:924. doi: 10.3389/fpls.2015.00924
- Aroca, A., Gotor, C., and Romero, L. C. (2018). Hydrogen sulfide signaling in plants: emerging roles of protein persulfidation. *Front. Plant Sci.* 9:1369. doi: 10.3389/fpls.2018.01369
- Ausma, T., Parmar, S., Hawkesford, M. J., and De Kok, L. J. (2017). "Impact of atmospheric H₂S, salinity and anoxia on sulfur metabolism in *Zea mays*" in *Sulfur metabolism in higher plants: Fundamental, environmental and agricultural aspects*. eds. L. J. De Kok, M. J. Hawkesford, S. H. Haneklaus, and E. Schnug (Dordrecht: Springer), 93–101.
- Baillie, C. K., Kaufholdt, D., Karpinski, L. H., Schreiber, V., Hänsch, S., Evers, C., et al. (2016). Detoxification of volcanic sulfur surplus in plants: three different strategies of survival. *Environ. Exp. Bot.* 126, 44–54. doi: 10.1016/j.envexpbot.2016.02.007
- Beauchamp, R. O., Bus, J. S., Popp, J. A., Boreiko, C. J., and Andjelkovich, D. A. (1984). A critical review of the literature on hydrogen sulfide toxicity. *CRC Crit. Rev. Toxicol.* 13, 25–97. doi: 10.3109/10408448409029321
- Birke, H., De Kok, L. J., Wirtz, M., and Hell, R. (2015). The role of compartment-specific cysteine synthesis for sulfur homeostasis during H₂S exposure in *Arabidopsis*. *Plant Cell Physiol.* 56, 358–367. doi: 10.1093/pcp/pcu166
- Bloem, E., Haneklaus, S., Salac, I., Wickenhauser, P., and Schnug, E. (2007). Facts and fiction about sulfur metabolism in relation to plant-pathogen interactions. *Plant Biol.* 9, 596–607. doi: 10.1055/s-2007-965420
- Brunold, C., and Erisman, K. H. (1975). H₂S as sulfur source in *Lemma minor* L. II. Direct incorporation into cysteine and inhibition of sulfate assimilation. *Experientia* 31, 508–510. doi: 10.1007/BF01932426
- Buchner, P., Parmar, S., Kriegel, A., Carpentier, M., and Hawkesford, M. J. (2010). The sulfate transporter family in wheat: tissue-specific gene expression in relation to nutrition. *Mol. Plant* 3, 374–389. doi: 10.1093/mp/ssp119
- Buchner, P., Stuiver, C. E. E., Westerman, S., Wirtz, M., Hell, R., Hawkesford, M. J., et al. (2004). Regulation of sulfate uptake and expression of sulfate transporter genes in *Brassica oleracea* L. as affected by atmospheric H₂S and pedospheric sulfate nutrition. *Plant Physiol.* 136, 3396–3408. doi: 10.1104/pp.104.046441
- Buwalda, F., De Kok, L. J., and Stulen, I. (1992). The flux of atmospheric H₂S to spinach leaves can be affected by the supply of O-acetylserine. *Phyton* 32, 15–18.
- Buwalda, F., De Kok, L. J., and Stulen, I. (1993). Effects of atmospheric H₂S on thiol composition of crop plants. *J. Plant Physiol.* 142, 281–285. doi: 10.1016/S0176-1617(11)80423-2
- Buwalda, F., De Kok, L. J., and Stulen, I. (1994). The pool of water-soluble non-protein thiols is not regulated within narrow limits in spinach leaves exposed to atmospheric H₂S. *Plant Physiol. Biochem.* 32, 533–537.
- Buwalda, F., De Kok, L. J., Stulen, I., and Kuiper, P. J. C. (1988). Cysteine γ -glutamyl-cysteine and glutathione contents of spinach leaves as affected by darkness and application of excess sulfur. *Physiol. Plant.* 74, 663–668. doi: 10.1111/j.1399-3054.1988.tb02033.x
- Carn, S. H., Fioletov, V. E., McLinden, C. A., Li, C., and Krotkov, N. A. (2017). A decade of global volcanic SO₂ emissions measured from space. *Sci. Rep.* 7:44095. doi: 10.1038/srep44095
- Cope, D. M., and Spedding, D. J. (1982). Hydrogen sulphide uptake by vegetation. *Atmos. Environ.* 16, 349–353. doi: 10.1016/0004-6981(82)90452-8
- Coyne, P. I., and Bingham, G. E. (1978). Photosynthesis and stomatal light responses in snap beans exposed to hydrogen sulfide and ozone. *J. Air Pollut. Control Assoc.* 28, 1119–1123. doi: 10.1080/00022470.1978.10470715
- De Bruyn, W. J., Swartz, E., Hu, J. H., Shorter, J. A., Davidovits, P., Worsnop, D. R., et al. (1995). Henry's law solubilities and Setchenow coefficients for biogenic reduced sulfur species obtained from gas-liquid uptake measurements. *J. Geophys. Res.* 100, 7245–7251. doi: 10.1029/95JD00217
- De Kok, L. J. (1989). Responses of sulfur metabolism in plants to atmospheric hydrogen sulfide. *Phyton* 29, 189–201.
- De Kok, L. J. (1990). "Sulfur metabolism in plants exposed to atmospheric sulfur" in *Sulfur nutrition and sulfur assimilation in higher plants*. eds. H. Rennenberg, C. Brunold, L. J. De Kok, and I. Stulen (The Hague: SPB Academic Publishers), 111–130.
- De Kok, L. J., Bosma, W., Maas, F. M., and Kuiper, P. J. C. (1985). The effect of short-term H₂S fumigation on water-soluble sulfhydryl compounds and glutathione levels in spinach. *Plant Cell Environ.* 8, 189–194. doi: 10.1111/1365-3040.ep11604605
- De Kok, L. J., Durenkamp, M., Yang, L., and Stulen, I. (2007). "Atmospheric sulfur" in *Sulfur in plants: An ecological perspective*. eds. M. J. Hawkesford and L. J. De Kok (Berlin: Springer), 91–106.
- De Kok, L. J., Maas, F. M., Godeke, J., Haaksma, A. B., and Kuiper, P. J. C. (1986). Glutathione a tripeptide which may function as a temporary storage of excessive reduced sulphur in H₂S fumigated spinach plants. *Plant Soil* 91, 349–352. doi: 10.1007/BF02198121
- De Kok, L. J., Rennenberg, H., and Kuiper, P. J. C. (1991). The internal resistance in spinach shoots to atmospheric H₂S deposition is determined by metabolism processes. *Plant Physiol. Biochem.* 29, 463–470.
- De Kok, L. J., Stahl, K., and Rennenberg, H. (1989). Fluxes of atmospheric hydrogen sulfide to plant shoots. *New Phytol.* 112, 533–542. doi: 10.1111/j.1469-8137.1989.tb00348.x
- De Kok, L. J., Stuiver, C. E. E., Rubinig, M., Westerman, S., and Grill, D. (1997). Impact of atmospheric sulfur deposition on sulfur metabolism in plants: H₂S as sulfur source for sulfur deprived *Brassica oleracea* L. *Bot. Acta* 110, 411–419. doi: 10.1111/j.1438-8677.1997.tb00657.x
- De Kok, L. J., Stuiver, C. E. E., and Stulen, I. (1998). "Impact of atmospheric H₂S on plants" in *Responses of plant metabolism to air pollution and global change*. eds. L. J. De Kok and I. Stulen (Leiden: Backhuys Publishers), 41–63.
- De Kok, L. J., Stuiver, C. E. E., Westerman, S., and Stulen, I. (2000). "Atmospheric H₂S as plant sulfur source: interaction with pedospheric sulfur nutrition – a case study with *Brassica oleracea* L." in *Sulfur nutrition and sulfur assimilation in higher plants: Molecular, biochemical and physiological aspects*. eds. C. Brunold, H. Rennenberg, L. J. De Kok, I. Stulen, and J. C. Davidian (Bern: Paul Haupt), 41–55.
- De Kok, L. J., Stuiver, C. E. E., Westerman, S., and Stulen, I. (2002). "Elevated levels of hydrogen sulfide in the plant environment: nutrient or toxin" in *Air pollution and biotechnology in plants*. eds. K. Omasa, H. Saji, S. Youssefian, and N. Kondo (Tokyo: Springer-Verlag), 201–213.
- De Kok, L. J., and Tausz, M. (2001). "The role of glutathione in plant reaction and adaptation to air pollutants" in *Significance of glutathione to plant adaptation to the environment*. eds. D. Grill, M. Tausz, and L. J. De Kok (Dordrecht: Kluwer Academic), 185–201.
- De Kok, L. J., Thompson, C. R., and Kuiper, P. J. C. (1983a). Sulfide-induced oxygen uptake by isolated spinach chloroplasts catalyzed by photosynthetic electron transport. *Physiol. Plant.* 59, 19–22. doi: 10.1111/j.1399-3054.1983.tb06564.x
- De Kok, L. J., Thompson, C. R., Mudd, J. B., and Kats, G. (1983b). Effect of H₂S fumigation on water-soluble sulfhydryl compounds in shoots of crop plants. *Z. Pflanzenphysiol.* 111, 85–89. doi: 10.1016/S0044-328X(83)80076-2
- De Kok, L. J., Yang, L., Stuiver, C. E. E., and Stulen, I. (2009). "Negative vs. positive functional plant responses to air pollution: a study establishing cause-effect relationships to SO₂ and H₂S" in *Air quality and ecological impacts, relating sources to effects*. ed. A. Legge (Amsterdam: Elsevier), 121–134.
- Durenkamp, M., and De Kok, L. J. (2002). The impact of atmospheric H₂S on growth and sulfur metabolism of *Allium cepa* L. *Phyton* 42, 55–63.
- Durenkamp, M., and De Kok, L. J. (2004). Impact of pedospheric and atmospheric sulphur nutrition on sulphur metabolism of *Allium cepa* L. a species with a potential sink capacity for secondary sulphur compounds. *J. Exp. Bot.* 55, 1821–1830. doi: 10.1093/jxb/erh187
- Durenkamp, M., and De Kok, L. J. (2005). Low levels of H₂S may replace sulfate as sulfur source in sulfate-deprived onion. *Phyton* 45, 69–77.
- Durenkamp, M., De Kok, L. J., and Kopriva, S. (2007). Adenosine 5' phosphosulphate reductase is regulated differently in *Allium cepa* L. and

- Brassica oleracea* L. upon exposure to H₂S. *J. Exp. Bot.* 58, 1571–1579. doi: 10.1093/jxb/erm031
- Durenkamp, M., Posthumus, F. S., Stuiver, C. E. E., and De Kok, L. J. (2005). “Metabolism of atmospheric sulfur gases in onion” in *Plant responses to air pollution and global change*. eds. K. Omasa, I. Nouchi, and L. J. De Kok (Tokyo: Springer), 3–11.
- Ernst, W. H. O. (1997). “Life-history syndromes and the ecology of plants from high sulphur-habitats” in *Sulphur metabolism in higher plants: Molecular ecophysiological and nutritional aspects*. eds. W. J. Cram, L. J. De Kok, I. Stulen, C. Brunold, and H. Rennenberg (Leiden: Backhuys Publishers), 131–146.
- Fioletov, V. E., McLinden, C. A., Krotkov, N., Li, C., Joiner, J., Theys, N., et al. (2016). A global catalogue of large SO₂ sources and emissions derived from the ozone monitoring system. *Atmos. Chem. Phys.* 16, 11497–11519. doi: 10.5194/acp-16-11497-2016
- Gerlich, S. C., Walker, B. J., Krueger, S., and Kopriva, S. (2018). Sulfate metabolism in *C₄ Flaveria* species is controlled by the root and connected to serine biosynthesis. *Plant Physiol.* 178, 565–582. doi: 10.1104/pp.18.00520
- Hancock, J. T. (2018). Hydrogen sulfide and environmental stresses. *Environ. Exp. Bot.* 161, 50–56. doi: 10.1016/j.envexpbot.2018.08.034
- Haneklaus, S., Bloem, S., and Schnug, E. (2007). “Sulfur and plant disease” in *Mineral nutrition and plant diseases*. eds. L. Datnoff, W. Elmer, and D. Huber (St. Paul: APS Press), 101–118.
- Hawkesford, M. J., and De Kok, L. J. (2006). Managing sulphur metabolism in plants. *Plant Cell Environ.* 29, 382–395. doi: 10.1111/j.1365-3040.2005.01470.x
- Hell, R., Jost, R., Berkowitz, O., and Wirtz, M. (2002). Molecular and biochemical analysis of the enzymes of cysteine biosynthesis in the plant *Arabidopsis thaliana*. *Amino Acids* 22, 245–257. doi: 10.1007/s007260200012
- Herschbach, C., De Kok, L. J., and Rennenberg, H. (1995a). Net uptake of sulphate and its transport to the shoot in spinach plants fumigated with H₂S or SO₂: does atmospheric sulphur affect the ‘inter-organ’ regulation of sulphur nutrition? *Bot. Acta* 108, 41–46. doi: 10.1111/j.1438-8677.1995.tb00829.x
- Herschbach, C., De Kok, L. J., and Rennenberg, H. (1995b). Net uptake of sulphate and its transport to the shoot in tobacco plants fumigated with H₂S or SO₂. *Plant Soil* 175, 75–84. doi: 10.1007/BF02413012
- Hesse, H., Lipke, J., Altman, T., and Hoefgen, R. (1997). “Expression analysis and subcellular localization of cysteine synthase isoforms from *Arabidopsis thaliana*” in *Sulphur metabolism in higher plants: Molecular ecophysiological and nutritional aspects*. eds. W. J. Cram, L. J. De Kok, I. Stulen, C. Brunold, and H. Rennenberg (Leiden: Backhuys Publishers), 227–230.
- Kanda, K. I., and Tsuruta, H. (1995). Emissions of sulfur gases from various types of terrestrial higher plants. *Soil Sci. Plant Nutr.* 41, 321–328. doi: 10.1080/00380768.1995.10419589
- Kellogg, W. W., Cadle, R. D., Allen, E. R., Lazrus, A. L., and Martell, E. A. (1972). The sulfur cycle. *Science* 175, 587–596. doi: 10.1126/science.175.4022.587
- Klimont, Z., Smith, S. J., and Cofala, J. (2013). The last decade of global anthropogenic sulfur dioxide: 2000–2011 emissions. *Environ. Res. Lett.* 8:014003. doi: 10.1088/1748-9326/8/1/014003
- Koralewska, A., Posthumus, F. S., Stuiver, C. E. E., Buchner, P., and De Kok, L. J. (2007). The characteristic high sulfate content in *Brassica oleracea* is controlled by the expression and activity of sulfate transporters. *Plant Biol.* 9, 654–661. doi: 10.1055/s-2007-965438
- Koralewska, A., Stuiver, C. E. E., Posthumus, F. S., De Kok, L. J., and Hawkesford, M. J. (2009). “The up-regulated sulfate uptake capacity, but not the expression of sulfate transporters is strictly controlled by the sink capacity in curly kale” in *Sulfur metabolism in plants: Regulatory aspects, significance of sulfur in the food chain, agriculture and the environment*. eds. A. Sirko, L. J. De Kok, S. Haneklaus, M. J. Hawkesford, H. Rennenberg, K. Saito, E. Schnug, and I. Stulen (Leiden: Backhuys Publishers), 61–68.
- Koralewska, A., Stuiver, C. E. E., Posthumus, F. S., Kopriva, S., Hawkesford, M. J., and De Kok, L. J. (2008). Regulation of sulfate uptake, expression of the sulfate transporters Sultr1,1 and Sultr1,2, and APS reductase in Chinese cabbage (*Brassica pekinensis*) as affected by atmospheric H₂S nutrition and sulfate deprivation. *Funct. Plant Biol.* 35, 318–327. doi: 10.1071/FP07283
- Lee, Z. W., Zhou, J., Chen, C. S., Zhao, Y., Tan, C. H., Li, L., et al. (2011). The slow-releasing hydrogen sulfide donor, GYY4137, exhibits novel anti-cancer effects *in vitro* and *in vivo*. *PLoS One* 6:e21077. doi: 10.1371/journal.pone.0021077
- Lenzian, K. L. (1984). “Permeability of plant cuticles to gaseous air pollutants” in *Gaseous air pollutants and plant metabolism*. eds. M. J. Koziol, and F. R. Whatley (London: Butterworths), 77–81.
- Li, Z. G., Min, X., and Zhou, Z. H. (2016). Hydrogen sulfide: a signal molecule in plant cross-adaptation. *Front. Plant Sci.* 26:1621. doi: 10.3389/fpls.2016.01621
- Maas, F. M., and De Kok, L. J. (1988). In vitro NADH oxidation as an early indicator for growth reductions in spinach exposed to H₂S in the ambient air. *Plant Cell Physiol.* 29, 523–526. doi: 10.1093/oxfordjournals.pcp.a077524
- Maas, F. M., De Kok, L. J., Hoffmann, I., and Kuiper, P. J. C. (1987a). Plant responses to H₂S and SO₂ fumigation. I. Effects on growth, transpiration and sulfur content of spinach. *Physiol. Plant.* 70, 713–721. doi: 10.1111/j.1399-3054.1987.tb04329.x
- Maas, F. M., De Kok, L. J., and Kuiper, P. J. C. (1985). The effect of H₂S fumigation on various spinach (*Spinacia oleracea* L.) cultivars. Relation between growth inhibition and accumulation of sulphur compounds in the plant. *J. Plant Physiol.* 119, 219–226. doi: 10.1016/S0176-1617(85)80181-4
- Maas, F. M., De Kok, L. J., Peters, J. L., and Kuiper, P. J. C. (1987c). A comparative study on the effects of H₂S and SO₂ fumigation on the growth and accumulation of sulfate and sulphydryl compounds in *Trifolium pratense* L., *Glycine max* Merr. and *Phaseolus vulgaris* L. *J. Exp. Bot.* 38, 1459–1469. doi: 10.1093/jxb/38.9.1459
- Maas, F. M., De Kok, L. J., Strik-Timmer, W., and Kuiper, P. J. C. (1987b). Plant responses to H₂S and SO₂ fumigation. II. Differences in metabolism of H₂S and SO₂ in spinach. *Physiol. Plant.* 70, 722–728. doi: 10.1111/j.1399-3054.1987.tb04330.x
- Maas, F. M., van Loo, E. N., and Van Hasselt, P. R. (1988). Effect of long-term H₂S fumigation on photosynthesis in spinach. Correlation between CO₂ fixation and chlorophyll a fluorescence. *Physiol. Plant.* 72, 77–83. doi: 10.1111/j.1399-3054.1988.tb06625.x
- Martin, N. M., and Maricle, B. R. (2015). Species-specific enzymatic tolerance of sulfide toxicity in plant roots. *Plant Physiol. Biochem.* 88, 36–41. doi: 10.1016/j.plaphy.2015.01.007
- Mathai, J. C., Missner, A., Kügler, P., Saparov, S. M., Zeidel, M. L., Lee, J. K., et al. (2009). No facilitator required for membrane transport of hydrogen sulfide. *Proc. Natl. Acad. Sci. USA* 106, 16633–16638. doi: 10.1073/pnas.0902952106
- Mudd, J. B. (1979). “Effects on vegetation and aquatic animals” in *Hydrogen sulfide*. Ed. Assembly of Life Sciences (U.S.) Subcommittee on Hydrogen Sulfide (Baltimore: University Park Press), 67–79.
- Nobel, P. S. (1983). *Biophysical plant physiology and ecology*. (San Francisco: Freeman and Co.).
- Noctor, G., Mhamdi, A., Chaouch, S., Han, Y., Neukermans, J., Marquez-Garcia, B., et al. (2012). Glutathione in plants: an integrated overview. *Plant Cell Environ.* 35, 454–484. doi: 10.1111/j.1365-3040.2011.02400.x
- Oliva, M., and Steubing, L. (1976). Untersuchungen über der beeinflussung von photosynthese, respiration und wasserhaushalt durch H₂S bei *Spinacia oleracea*. *Angew. Bot.* 50, 1–17.
- Perraud, V., Horne, J. R., Martinez, A. S., Kalinowski, J., Meinardi, S., Dawson, M. L., et al. (2015). The future of airborne sulfur-containing particles in the absence of fossil fuel sulfur dioxide emissions. *Proc. Natl. Acad. Sci. USA* 112, 13514–13519. doi: 10.1073/pnas.1510743112
- Poortinga, A. M., and De Kok, L. J. (1997). “Uptake of atmospheric H₂S by *Spinacia oleracea* L. and consequences for thiol content and composition in shoots and roots” in *Sulphur metabolism in higher plants: Molecular, ecophysiological and nutritional aspects*. eds. W. J. Cram, L. J. De Kok, I. Stulen, C. Brunold, and H. Rennenberg (Leiden: Backhuys Publishers), 285–288.
- Rennenberg, H. (1981). Differences in the use of cysteine and glutathione as sulfur source in photoheterotrophic tobacco suspension cultures. *Z. Pflanzenphysiol.* 105, 31–40. doi: 10.1016/S0044-328X(81)80005-0
- Rennenberg, H. (1984). The fate of excess sulfur in higher plants. *Ann. Rev. Plant Physiol.* 35, 121–153. doi: 10.1146/annurev.pp.35.060184.001005
- Rennenberg, H., Huber, B., Schröder, P., Stahl, K., Haunold, W., Georgii, H. W., et al. (1990). Emission of volatile sulfur compounds from spruce trees. *Plant Physiol.* 92, 560–564. doi: 10.1104/pp.92.3.560
- Riahi, S., and Rowley, C. N. (2014). Why can sulfide permeate cell membranes? *J. Am. Chem. Soc.* 136, 15111–15113. doi: 10.1021/ja508063s
- Riemenschneider, A., Nikiforova, V., Hoefgen, R., De Kok, L. J., and Papenbrock, J. (2005). Impact of elevated H₂S on metabolite levels, activity of enzymes

- and expression of genes involved in cysteine metabolism. *Plant Physiol. Biochem.* 43, 473–483. doi: 10.1016/j.plaphy.2005.04.001
- Saito, K., Takahashi, H., Takagi, Y., Inoue, K., and Nojima, M. (1997). “Molecular characterization and regulation of cysteine synthase and serine acetyltransferase from plants” in *Sulphur metabolism in higher plants: Molecular, ecophysiological and nutritional aspects*. eds. W. J. Cram, L. J. De Kok, I. Stulen, C. Brunold, and H. Rennenberg (Leiden: Backhuys Publishers), 235–238.
- Schnug, E. (1997). “Significance of sulphur for the quality of domesticated plants” in *Sulphur metabolism in higher plants: Molecular, ecophysiological and nutritional aspects*. eds. W. J. Cram, L. J. De Kok, I. Stulen, C. Brunold, and H. Rennenberg (Leiden: Backhuys Publishers), 109–130.
- Schröder, P. (1993). “Plants as sources of atmospheric sulfur” in *Sulfur nutrition and assimilation in higher plants: Regulatory, agricultural and environmental aspects*. eds. L. J. De Kok, I. Stulen, H. Rennenberg, C. Brunold, and W. E. Rauser (The Hague: SPB Academic Publishing), 253–270.
- Schulte, M., Herschbach, C., and Rennenberg, H. (1997). “Long term effects of naturally elevated CO₂, H₂S and SO₂ on sulphur allocation in *Quercus*” in *Sulphur metabolism in higher plants: Molecular, ecophysiological and nutritional aspects*. eds. W. J. Cram, L. J. De Kok, I. Stulen, C. Brunold, and H. Rennenberg (Leiden: Backhuys Publishers), 289–291.
- Schütz, B., De Kok, L. J., and Rennenberg, H. (1991). Thiol accumulation and cysteine desulfhydrase activity in H₂S-fumigated leaves and leaf homogenates of cucurbit plants. *Plant Cell Physiol.* 32, 733–736. doi: 10.1093/oxfordjournals.pcp.a078138
- Shahbaz, M., Parmar, S., Stuijver, C. E. E., Hawkesford, M. J., and De Kok, L. J. (2013). Copper toxicity and sulfur metabolism in Chinese cabbage are affected by UV radiation. *Environ. Exp. Bot.* 88, 60–70. doi: 10.1016/j.envexpbot.2012.04.007
- Shahbaz, M., Stuijver, C. E. E., Posthumus, F. S., Parmar, S., Hawkesford, M. J., and De Kok, L. J. (2014). Copper toxicity in Chinese cabbage is not influenced by plant sulphur status, but affects sulphur metabolism-related gene expression and the suggested regulatory metabolites. *Plant Biol.* 16, 68–78. doi: 10.1111/plb.12019
- Sipilä, M., Berndt, T., Petäjä, T., Brus, D., Vanhanen, J., Stratmann, F., et al. (2010). The role of sulfuric acid in atmospheric nucleation. *Science* 327, 1243–1246. doi: 10.1126/science.1180315
- Slatt, B. J., Natusch, D. F. S., Prospero, J. M., and Savoie, D. L. (1978). Hydrogen sulfide in the atmosphere of the northern equatorial Atlantic ocean and its relation to the global sulfur cycle. *Atmos. Environ.* 12, 981–991. doi: 10.1016/0004-6981(78)90342-6
- Stern, D. I. (2005). Global sulfur emissions from 1850 to 2000. *Chemosphere* 58, 163–175. doi: 10.1016/j.chemosphere.2004.08.022
- Stuijver, C. E. E., and De Kok, L. J. (1997). “Atmospheric H₂S as sulphur source for sulphur deprived *Brassica oleracea* L and *Hordeum vulgare* L.” in *Sulphur metabolism in higher plants: Molecular, ecophysiological and nutritional aspects*. eds. W. J. Cram, L. J. De Kok, I. Stulen, C. Brunold, and H. Rennenberg (Leiden: Backhuys Publishers), 293–294.
- Stuijver, C. E. E., and De Kok, L. J. (2001). Atmospheric H₂S as sulfur source for *Brassica oleracea*: kinetics of H₂S uptake and activity of O-acetylserine (thiol)lyase as affected by sulfur nutrition. *Environ. Exp. Bot.* 46, 29–36. doi: 10.1016/S0098-8472(01)00080-6
- Stuijver, C. E. E., De Kok, L. J., and Kuiper, P. J. C. (1992). Freezing tolerance and biochemical changes in wheat shoots as affected by H₂S fumigation. *Plant Physiol. Biochem.* 30, 47–55.
- Stulen, I., and De Kok, L. J. (1993). “Whole plant regulation of sulfate uptake and metabolism - a theoretical approach and comparison with current ideas on regulation of nitrogen metabolism” in *Sulfur nutrition and assimilation in higher plants: Regulatory, agricultural and environmental aspects*. eds. L. J. De Kok, I. Stulen, H. Rennenberg, C. Brunold, and W. E. Rauser (The Hague: SPB Academic Publishing), 77–91.
- Stulen, I., Posthumus, F. S., Amâncio, S., and De Kok, L. J. (1990). Why is H₂S not phytotoxic in monocots? *Physiol. Plant.* 7:123.
- Stulen, I., Posthumus, F. S., Amâncio, S., Masselink-Beltman, I., Müller, M., and De Kok, L. J. (2000). “Mechanism of H₂S phytotoxicity” in *Sulfur nutrition and sulfur assimilation in higher plants: Molecular, biochemical and physiological aspects*. eds. C. Brunold, H. Rennenberg, L. J. De Kok, I. Stulen, and J. C. Davidian (Bern: Paul Haupt), 381–383.
- Tausz, M., Van der Kooij, T. A. W., Müller, M., De Kok, L. J., and Grill, D. (1998). “Uptake and metabolism of oxidized and reduced sulfur pollutants by spruce trees” in *Responses of plant metabolism to air pollution and global change*. eds. L. J. De Kok, and I. Stulen (Leiden: Backhuys Publishers), 457–460.
- Taylor, G. E., and Selvidge, W. J. (1984). Phytotoxicity in bush bean of five sulfur-containing gases released from advanced fossil energy technologies. *J. Environ. Qual.* 13, 224–230. doi: 10.2134/jeq1984.00472425001300020010x
- Thompson, C. R., and Kats, G. (1978). Effect of continuous H₂S fumigation on crop and forest plants. *Environ. Sci. Technol.* 12, 550–553. doi: 10.1021/es60141a001
- Trudinger, P. A. (1986). “Chemistry of the sulfur cycle” in *Sulfur in agriculture*. ed. M. A. Tabatabai (Madison: American Society of Agronomy), 1–22.
- Van der Kooij, T. A. W., and De Kok, L. J. (1998). “Kinetics of deposition of SO₂ and H₂S to shoots of *Arabidopsis thaliana* L.” in *Responses of plant metabolism to air pollution and global change*. eds. L. J. De Kok and I. Stulen (Leiden: Backhuys Publishers), 479–481.
- Watts, S. F. (2000). The mass budgets of carbonyl sulfide, dimethyl sulfide, carbon disulfide and hydrogen sulfide. *Atmos. Environ.* 34, 761–779. doi: 10.1016/S1352-2310(99)00342-8
- Westerman, S., De Kok, L. J., Stuijver, C. E. E., and Stulen, I. (2000a). Interaction between metabolism of atmospheric H₂S in the shoot and sulfate uptake by the roots of curly kale (*Brassica oleracea*). *Physiol. Plant.* 109, 443–449. doi: 10.1034/j.1399-3054.2000.100411.x
- Westerman, S., Stulen, I., Suter, M., Brunold, C., and De Kok, L. J. (2001). Atmospheric H₂S as sulfur source for *Brassica oleracea*: consequences for the activity of the enzymes of the assimilatory sulfate reduction pathway. *Plant Physiol. Biochem.* 39, 425–432. doi: 10.1016/S0981-9428(01)01258-X
- Westerman, S., Weidner, W., De Kok, L. J., and Stulen, I. (2000b). Effect of H₂S exposure on ³⁵S-sulfate uptake, transport and utilization in curly kale. *Phyton* 40, 293–302.

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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