



# Pathophysiological Consequences of At-Risk Alcohol Use; Implications for Comorbidity Risk in Persons Living With Human Immunodeficiency Virus

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At-risk alcohol use is a significant risk factor associated with multisystemic pathophysiological effects leading to multiorgan injury and contributing to 5.3% of all deaths worldwide. The alcohol-mediated cellular and molecular alterations are particularly salient in vulnerable populations, such as people living with HIV (PLWH), diminishing their physiological reserve, and accelerating the aging process. This review presents salient alcohol-associated mechanisms involved in exacerbation of cardiometabolic and neuropathological comorbidities and their implications in the context of HIV disease. The review integrates consideration of environmental factors, such as consumption of a Western diet and its interactions with alcohol-induced metabolic and neurocognitive dyshomeostasis. Major alcohol-mediated mechanisms that contribute to cardiometabolic comorbidity include impaired substrate utilization and storage, endothelial dysfunction, dysregulation of the renin-angiotensin-aldosterone system, and hypertension. Neuroinflammation and loss of neurotrophic support in vulnerable brain regions significantly contribute to alcohol-associated development of neurological deficits and alcohol use disorder risk. Collectively, evidence suggests that at-risk alcohol use exacerbates cardiometabolic and neurocognitive pathologies and accelerates biological aging leading to the development of geriatric comorbidities manifested as frailty in PLWH.

**Keywords:** alcohol, HIV, cardiometabolic comorbidity, neuropathology, diet

## DEVELOPMENT OF ALCOHOL USE DISORDER

Globally, at-risk alcohol use is the seventh leading cause for morbidity and mortality accounting for 5.3% of all deaths, and the leading cause of disability-adjusted life-years (DALYs) among individuals 15–49 years of age (Burton and Sheron, 2018). Alcohol use disorder (AUD) is a chronic, relapsing condition often characterized by a parallel emergence of neurological deficits and negative emotional states (e.g., cognitive dysfunction, depression, pain) along with a significant escalation of alcohol use (Edwards and Koob, 2010). Consequently, the progression from recreational to at-risk drinking is thought to involve a motivational transition from positive reinforcement (i.e., drinking for pleasure) to negative reinforcement (i.e., drinking to avoid or mask the unpleasant symptoms of withdrawal). However, continued drinking at high levels over long periods of time

not only continues to damage the brain and other organs, but also locks the individual into a motivational cycle of binge/intoxication, withdrawal, and craving/anticipation of renewed drinking during repeated abstinence (Koob, 2021). The primary driver of excessive alcohol drinking stems from a dysregulation of motivational behaviors regulated by the brain. Specific brain reinforcement centers (including the central amygdala and prefrontal cortex described below) have evolved to process adaptive interactions with natural rewards encountered in our environment (including the pursuit of food, sex, and social cooperation). However, the adaptive function of these brain regions and goal-directed behaviors can be compromised by multiple genetic and environmental factors, including the excessive use of alcohol.

Epidemiological data using Diagnostic and Statistical Manual of Psychiatric Disorders (DSM-V) criteria indicate that 13.9% of the United States of America (United States) population met criteria for AUD over the past year (Grant et al., 2015). AUD significantly decreases life expectancy and increases the risk of mortality from mental disorders (10-fold), and from cardiovascular diseases and cancers (two-fold) (Roerecke and Rehm, 2014). The increased risk of mortality associated with at-risk alcohol use results from alcohol-induced end organ injury spanning cardiopulmonary, gastrointestinal, immune, adipose, musculoskeletal, and nervous systems.

## Relevance of Alcohol Drinking Patterns

Drinking in moderation, is defined by the 2015–2020 Dietary Guidelines for Americans as consumption of up to one drink per day for women and up to two drinks per day for men. Low risk drinking as defined by the National Institute on Alcohol Abuse and Alcoholism (NIAAA) is no more than 3 drinks per day and no more than 7 drinks per week for women, and no more than 4 drinks per day and no more than 14 drinks per week for men. Drinking in moderation and low risk drinking are not associated with a significant increase in risk for alcohol-induced comorbidities. In contrast, binge drinking defined as drinking to elevate blood alcohol concentrations to 0.08% (80 mg/dl) or higher, generally resulting from drinking 4 or 5 drinks over a 2-h time frame is considered at-risk and categorized as extreme when the consumption is twofold or greater than the gender-specific thresholds (i.e., 10 or more standard drinks for men, and 8 or more for women) (Yeomans et al., 2003; Alcohol Research, Current Reviews, and Editorial Staff., 2018). Alcohol consumption is discouraged in certain groups of people including underage individuals (less than 21 years of age), women who are pregnant or trying to become pregnant, those who take medications that interact with alcohol, and those with existing medical conditions, including human immunodeficiency virus (HIV) as discussed in this review (Alcohol Research, Current Reviews, and Editorial Staff., 2018).

While the contribution of at-risk alcohol consumption to organ injury is well recognized, the definition of at-risk drinking is complicated by different patterns of consumption, the types of alcoholic beverages consumed across geographical regions (World Health Organization, 2021), and the inconsistent definition of a standard drink across countries, ranging from 8

g in Iceland to 20 g in Austria (Kalinowski and Humphreys, 2016). In addition, there appears to be wide variability for defining rates of low to moderate-risk drinking across nations, ranging from 98 to 140 g per week for women and 150–280 g per week for men (Kalinowski and Humphreys, 2016). Several factors are considered in the classification of alcohol drinking patterns, including the frequency and amount of alcohol intake, number of episodes of acute intoxication, and number of alcohol binges (Wetterling et al., 1999). According to the World Health Organization (WHO), prevalence of at-risk drinking is measured by heavy episodic drinking, defined as consuming 60 g of alcohol or more on at least one occasion in the past 30 days. In the United States, where a standard drink is 14 g, heavy episodic drinking is defined by consumption of 4.25 standard drinks. Worldwide, approximately 18% of the adult population report heavy episodic drinking (Poznyak, 2018).

## AT-RISK ALCOHOL USE IN CHRONIC DISEASE: RELEVANCE TO PERSONS LIVING WITH HUMAN IMMUNODEFICIENCY VIRUS

The Centers of Disease Control (CDC) estimates that over 50% of PLWH in the U.S. are 50 years of age or older (CDC, 2018). As life expectancy of PLWH continues to rise and approach that of the general population, the frequency of maladaptive behaviors increases as well. Maladaptive coping resulting from psychosocial stressors, including stigma associated with living with HIV, is associated with poorer immune status, increased viral load over time, faster disease progression, and higher rates of mortality (Cruess et al., 2003; Leserman, 2008). At-risk alcohol use is among the principal maladaptive coping behaviors in PLWH, and AUD frequently occurs in PLWH (Wardell et al., 2018).

AUD may exacerbate the risk for geriatric comorbidities including cardiometabolic syndrome (CMS) and neurological deficits (Kahl and Hillemecher, 2016; Sullivan and Pfefferbaum, 2019; Edwards et al., 2020) among PLWH. Data collected from an ongoing New Orleans Alcohol and HIV (NOAH) longitudinal clinical study (Ferguson et al., 2020) show that lifetime alcohol exposure positively associates with Phenotypic Frailty Index and a 52-item deficit index after adjustment for subject demographics, HIV-related covariates, smoking, and history of other substance use (Maffei et al., 2020). Premature decline in functional status has been reported in PLWH (Piggott et al., 2016). Data show that the prevalence of a frailty-related phenotype (unintentional weight loss, exhaustion, weakness, slow walking speed, low physical activity) in 55-year-old HIV-infected men is comparable to that of HIV-uninfected men > 65 years old, strongly supporting accelerated biological aging in PLWH and its likelihood of associated enhanced risk for comorbidities (Desquilbet et al., 2007).

The principal mechanisms underlying alcohol addiction, alcohol-induced organ injury, and alcohol-related liver injury have been extensively reviewed (Molina et al., 2014b; Massey et al., 2015; Aberg et al., 2020; Arab et al., 2020;

Maddur and Shah, 2020; Rasineni et al., 2020; Koob, 2021; Osná et al., 2021). Here we focus on salient mechanisms of alcohol-induced risk and exacerbation of CMS and neuropathological comorbidities and discuss their implications in the context of HIV infection and their role in exacerbating or accelerating development of associated comorbidities in PLWH.

## MECHANISMS OF ALCOHOL-INDUCED END ORGAN INJURY

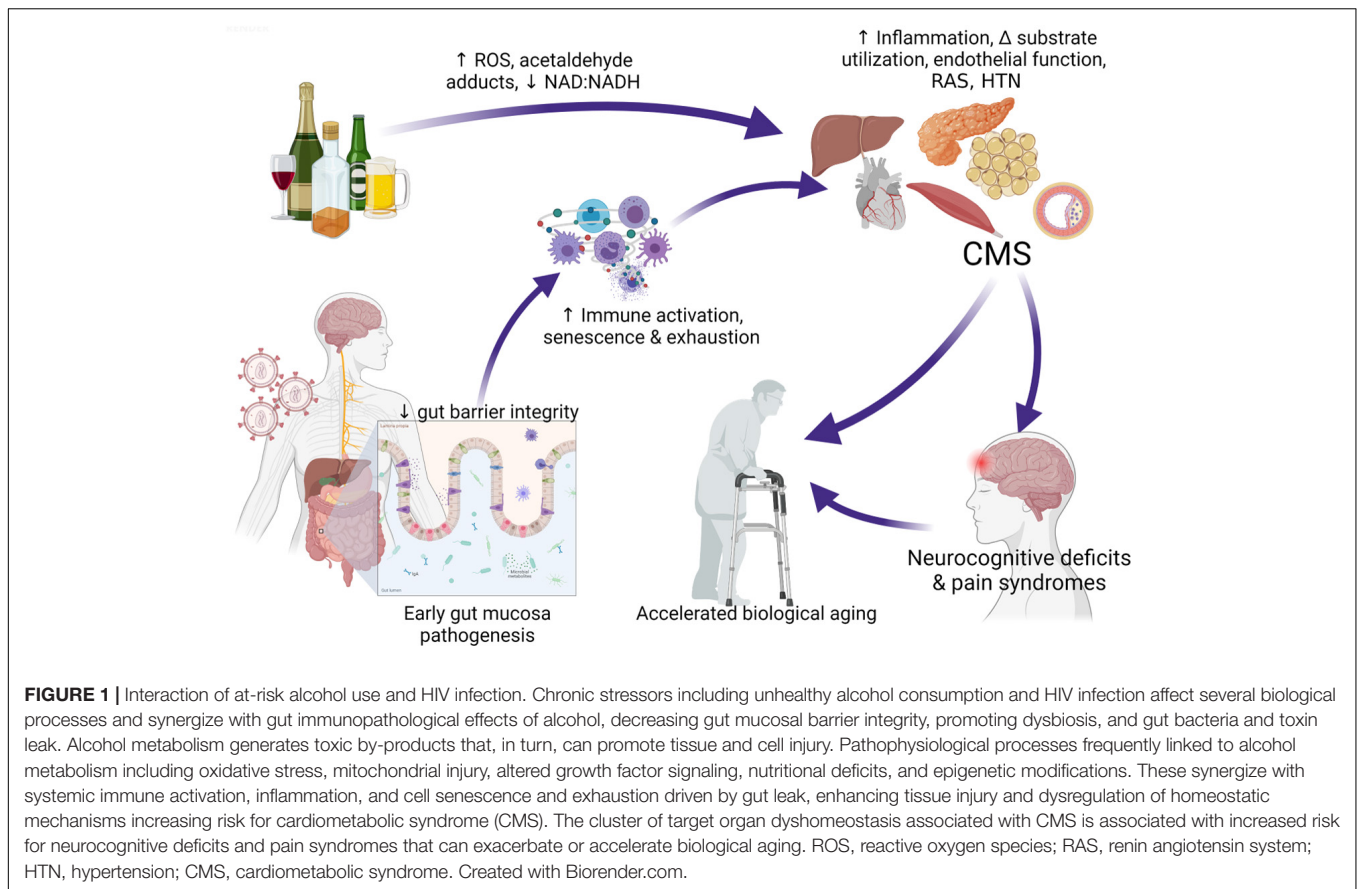
Alcohol-induced tissue injury leading to increased risk of comorbidities result from a combination of pathophysiological processes frequently linked to alcohol metabolism including oxidative stress, mitochondrial injury, altered growth factor signaling, nutritional deficits, and epigenetic modifications (Molina et al., 2014b). Alcohol metabolism can occur in virtually all tissues but occurs predominantly in the liver and generates toxic by-products that, in turn, can promote tissue and cell injury (Souza-Smith et al., 2016). Following first-pass metabolism of a small fraction of ingested alcohol in the stomach, most (92–95%) alcohol consumed is degraded by alcohol dehydrogenase (ADH) and aldehyde dehydrogenase (ALDH), forming acetaldehyde and acetate. Alcohol breakdown to acetaldehyde by ADH is associated by nicotinamide adenine dinucleotide (NAD<sup>+</sup>) reduction to NADH and a resulting decreased NAD<sup>+</sup>: NADH ratio that decreases availability of mitochondrial glutathione (mGSH), and cellular antioxidant reserve. The highly reactive product of ADH; acetaldehyde, can react with proteins and cell membranes producing acetaldehyde–protein adducts that also contribute to tissue injury. Acetaldehyde metabolism to acetate by ALDH2 in the mitochondria, further decreases cellular NAD<sup>+</sup>: NADH ratio. In addition to ADH conversion of alcohol to acetaldehyde; alcohol is also metabolized by cytochrome P450, producing reactive oxygen species (ROS). Thus, alcohol and its metabolites, the generation of ROS, and formation of acetaldehyde adducts contribute to cell and tissue injury (Cederbaum, 2012). Oxidative stress and mitochondrial dysfunction are both implicated in HIV disease pathogenesis (Aukrust et al., 2003; Schank et al., 2021) suggesting that in combination, at-risk heavy alcohol use and HIV are likely to impose a greater oxidative burden on tissues. Chronic oxidative stress affects several biological processes and synergize with gut immunopathological effects of alcohol (Gu et al., 2021; Shukla et al., 2021) to decrease gut mucosal barrier integrity and promote dysbiosis (Maffei et al., 2021). The resulting gut leak promotes systemic immune activation, inflammation, and cell senescence, enhancing tissue injury and dysregulation of homeostatic mechanisms exacerbating the risk for comorbidities in PLWH (Katz et al., 2015; Maffei et al., 2020; **Figure 1**).

### Interactions of At-Risk Alcohol Use and Human Immunodeficiency Virus, Risk for Cardiometabolic Syndrome

Cardiometabolic syndrome is a combination of alterations including glucose intolerance, dyslipidemia, hypertriglyceridemia, central adiposity, and hypertension,

that is now recognized by the WHO as a disease entity (Saljoughian, 2016). CMS increases mortality risk due to coronary heart disease, myocardial infarction, stroke, and type 2 diabetes (Castro et al., 2003) and is one of the geriatric comorbidities prevalent in PLWH (Woldu et al., 2020). Lifestyle behaviors including at-risk alcohol use, consumption of diets rich in fat and sugar, low physical activity, and behavioral and psychosocial stressors are associated with increased risk for CMS and type 2 diabetes. Alcohol use impacts risk for CMS following a U or J shape curve (Alkerwi et al., 2009; Baliunas et al., 2009; Koloverou et al., 2015; Lai et al., 2019). However, this is not generalizable across geographical locations or ethnic groups. Moderate alcohol consumption is shown to be protective in studies from the United States (Kerr and Ye, 2010) or Sweden (Rasouli et al., 2013), but not in studies conducted in Japan (Teratani et al., 2012). Data relating alcohol to all-cause and cardiovascular (CV) morbidity and mortality from more than 600,000 current drinkers without a history of CV disease, indicated a much lower threshold for amount of alcohol consumed than currently accepted (Wood et al., 2018). In a prospective, longitudinal study, male sex, being physically active, and in good health status were independently associated with light to moderate drinking. Though there was an apparent protective effect of light to moderate drinking on mortality after adjusting for age, sex, risk factors, and cardiovascular events, the relationship was no longer significant when adjusting for physical activity and perceived health status (Muscari et al., 2015). These results suggest that the protective effect of moderate alcohol use is most prominent among individuals with a healthier lifestyle (Koloverou et al., 2015). Moreover, compelling evidence suggest that risk for CMS increases with increasing alcohol consumption (Vieira et al., 2016). Alcohol-using PLWH have higher odds of displaying lipodystrophy (Cheng et al., 2009; Molina et al., 2014a) and altered adipokine profiles including that of leptin (Srdic et al., 2017) and adiponectin (Lee and Kwak, 2014; Langkilde et al., 2018). Alterations in these adipokines have been linked to metabolic dysregulation in obese subjects (Harwood, 2012; Lee and Kwak, 2014), in PLWH on ART (Jan et al., 2004; Lagathu et al., 2005), and to impaired insulin signaling (Jan et al., 2004; Stankov and Behrens, 2010). CMS is a comorbidity commonly associated with cognitive deficits in PLWH (Pope et al., 2020; Pasipanodya et al., 2021), and cardiometabolic health is linked to improved cognitive function in aging PLWH (Saloner et al., 2019).

Diet quality is influenced by the pattern of alcohol use, being the poorest among subjects who consumed the highest quantity of alcohol (Breslow et al., 2006), among women who were current drinkers, and among both men and women as the amount of alcohol consumption increased (Breslow et al., 2010). The interaction of unhealthy alcohol use with an unhealthy diet is likely to further enhance risk for comorbidities. Diet quality among PLWH (Muhammad et al., 2019; Weiss et al., 2019) is significantly lower than recommended. In a large single-site study among PLWH in the US, dietary recall demonstrated that the average carbohydrate intake was twice the recommended amount and protein consumption was a third of recommendations



and was also associated with a trend for increased fat intake (Webel et al., 2017). This dietary pattern, referred to as a Western Diet has been linked to immune and metabolic dysfunction (Childs et al., 2019). PLWH have a lower healthy eating index (HEI) score compared to HIV- subjects, with decreased scores for consumption of seafood and plant proteins and beneficial fatty acids, along with increased consumption of foods with non-nutritional calories including alcohol (Weiss et al., 2019). The HEI assesses dietary patterns and their degree of adherence to the Dietary Guidelines for Americans (Guenther et al., 2008), and scores  $\leq 50$  are inversely correlated with reduced risk of all-cause mortality in the general population (Sotos-Prieto et al., 2017a,b). Increased insulin, cholesterol, and triglycerides in PLWH are associated with increased total dietary energy intake with high total and saturated fat (Batterham et al., 2000). Chronic binge alcohol in simian immunodeficiency virus (SIV) infected macaques decreases total caloric intake and alters nutrient selection, decreasing nitrogen intake and balance as the disease progresses (Molina et al., 2006). The self-imposed carbohydrate rich, low nitrogen diet likely contributes to decreased muscle mass observed in end stage disease. Changes in dietary intake can influence micronutrient availability and in addition modulate circulating and tissue growth factors (Childs et al., 2019). Hence, direct, and indirect alcohol-mediated effects as well as diet quality and composition can significantly contribute to risk for CMS in PLWH.

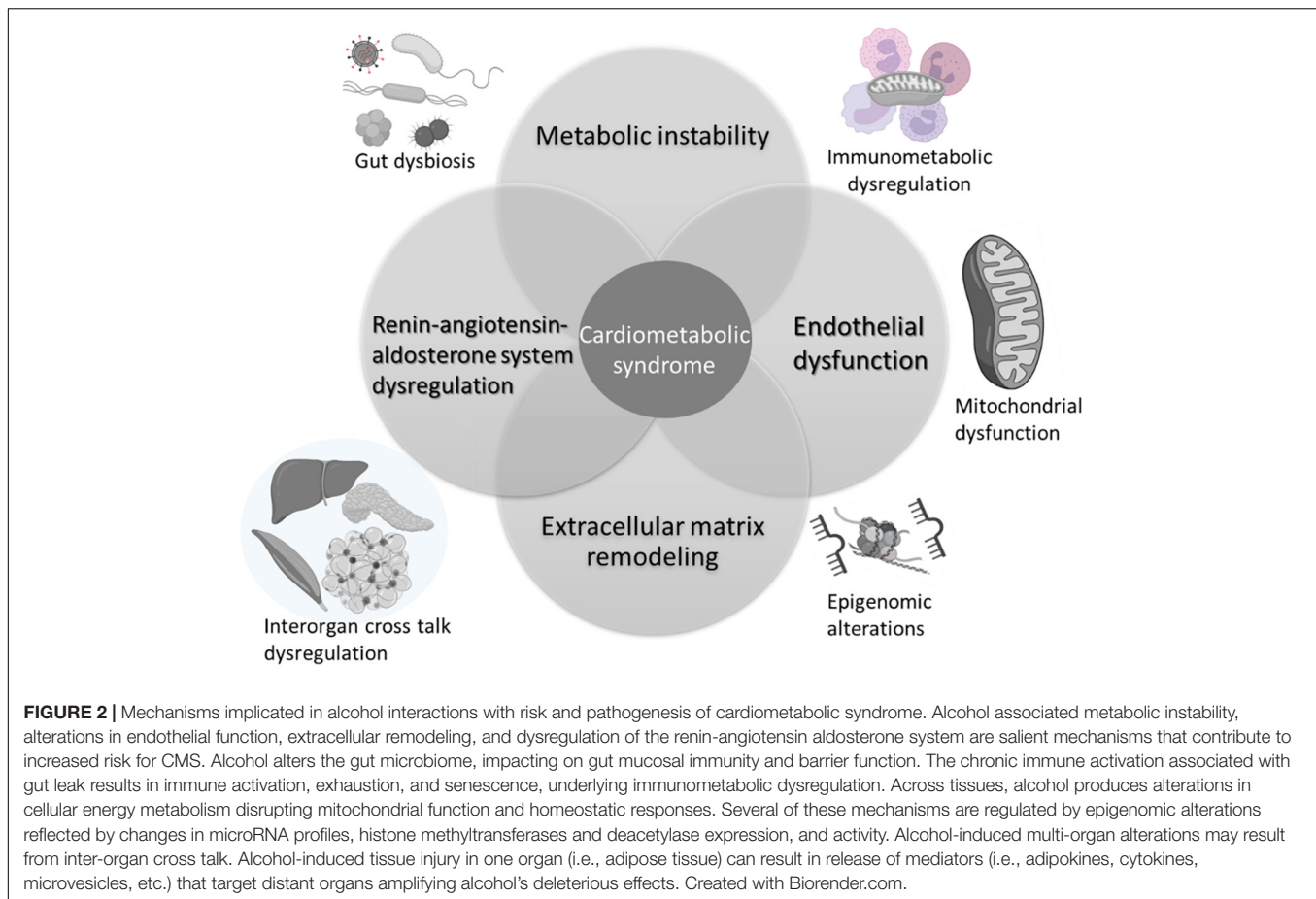
## MECHANISMS IMPLICATED IN ALCOHOL-HUMAN IMMUNODEFICIENCY VIRUS INTERACTIONS INCREASING RISK AND PATHOGENESIS OF CARDIOMETABOLIC SYNDROME

The pathophysiology of CMS is multifactorial, involving multiple organs and several independent and interdependent pathways. Several mechanisms impacted by at-risk alcohol use can potentially increase the risk and/or exacerbate CMS in PLWH (Figure 2). The most salient ones are discussed below.

### Interference With Energy Substrate Utilization and Storage

Alcohol-induced carbohydrate, lipid, and protein metabolic dysregulation is due to either direct effects of alcohol and its metabolites on liver, adipose tissue, and skeletal muscle (SKM), or due to indirect effects resulting from immune dysregulation (Figure 3). Alcohol consumption significantly decreases gluconeogenesis partly due to impaired hepatic utilization of lactic acid, glycerol, and alanine as gluconeogenic substrates (Steiner et al., 2015a) and a reduction in glycolytic and gluconeogenic hepatic enzyme activity (Baranyai and Blum, 1989; Mokuda et al., 2004). Alcohol impairs insulin signaling





including decreased phosphorylation of the insulin receptor, insulin receptor substrate (IRS)-1, and AKT, and decreases GLUT4 membrane translocation in peripheral insulin sensitive tissues (Steiner et al., 2015a). Reports in the literature indicate that chronic alcohol feeding (Kang et al., 2007b) as well as ART (Boccarda, 2008) result in the development of insulin resistance, one of the main components of CMS. The mechanisms underlying insulin resistance and dysglycemia; manifested as an increased frequency of glucose intolerance or frank diabetes mellitus in PLWH (Willig and Overton, 2016; Natsag et al., 2017; Noumegni et al., 2017), remain poorly understood. Chronic subclinical inflammation has been proposed as a likely factor contributing to metabolic dyshomeostasis seen in ART-treated PLWH (Longenecker et al., 2013; Beires et al., 2018).

SKM is the major insulin-sensitive tissue in the body and is a major player in maintaining glucose homeostasis. Thus, alcohol-induced alterations in SKM homeostasis can potentially have a significant impact on metabolic dysregulation. The alcohol-mediated decrease in SKM protein synthesis (LeCapitaine et al., 2011; Steiner and Lang, 2019) together with alterations in the mammalian target of rapamycin (mTOR) signaling pathway (Hong-Brown et al., 2012) result in alcoholic myopathy; which together with the alcohol-associated alterations in synthesis and breakdown of carbohydrates and lipids, contribute to metabolic instability and increased risk for CMS (Figure 3).

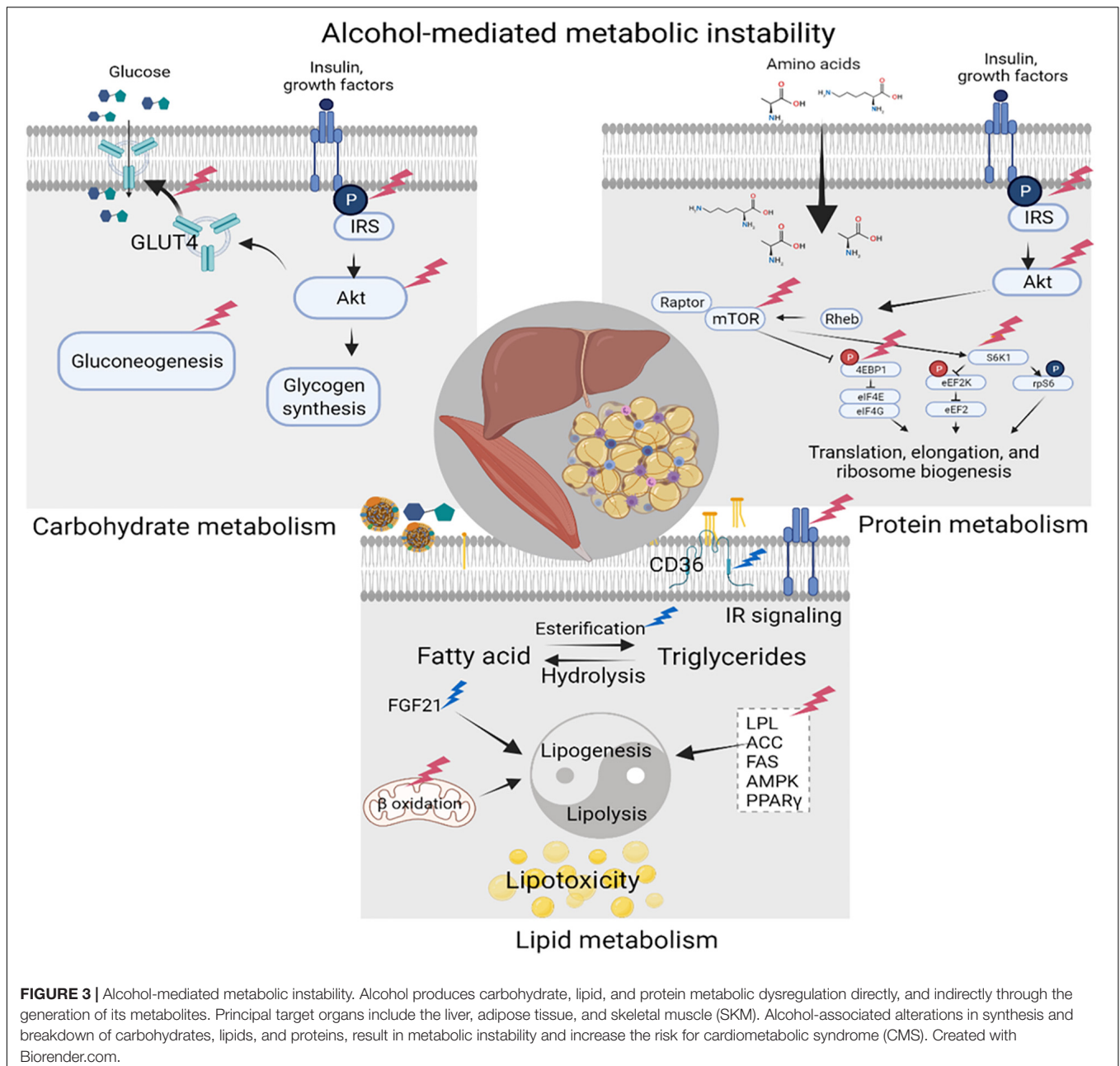
These alcohol-mediated alterations in SKM and adipose mass, and carbohydrate storage, can be reflected in anthropometric changes as well as in alterations of circulating levels of glucose, triglycerides, and cholesterol, as briefly discussed in the next section.

### Alterations in Body Composition

Central or abdominal obesity is associated with increased insulin resistance (Klaus et al., 2009; Govers, 2015; Jankie and Pinto Pereira, 2021). Heavy (20 to less than 60 g/day) and very heavy (60 g/day) alcohol use increases waist circumference and abdominal fat accumulation (Churilla et al., 2014; Hirakawa et al., 2015). Though the exact mechanisms are unclear, proposed alcohol-associated mechanisms include decreased leptin and glucagon like peptide -1, alterations in sex steroid hormones, loss of lean body mass, and increased lipolysis and fatty acid flux (Riserus and Ingelsson, 2007; Sumi et al., 2019). In the post-ART era, obesity is emerging as a critical problem in PLWH (Godfrey et al., 2019), with increased body mass index (BMI) associated with increased risk of CV disease and diabetes mellitus among PLWH (Achhra et al., 2018).

### Alterations in Glycemic Control

Reports on the effects of alcohol on glycemic control are mixed. Low to moderate alcohol use inhibits hepatic glycogenolysis



and gluconeogenesis, decreases fasting insulin and HbA1c, and increases insulin sensitivity (Hong et al., 2009; Koloverou et al., 2015; Schrieke et al., 2015; Vieira et al., 2016). Clinical studies show that alcohol decreases circulating basal insulin secretion (Bonnet et al., 2012) and decreases circulating insulin and c-peptide response to glucose (Patto et al., 1993). Preclinical studies show that chronic binge alcohol significantly impairs endocrine pancreatic response to a glucose load in SIV-infected macaques (Ford et al., 2016; Simon et al., 2021). Similarly, rodents on a chronic alcohol diet have decreased circulating insulin levels (Rasineni et al., 2019), and decreased pancreatic expression of glucokinase, glucose transporter-2 (Kim et al., 2010), and gamma-aminobutyric acid (GABA) receptors (Wang et al., 2014).

These alcohol-induced alterations can potentially contribute to decreased insulin release following a glucose load. This is aligned with studies showing that *in vitro* alcohol exposure decreases glucose-stimulated insulin secretion (GSIS) from human (Dragan et al., 2017) and rodent (Wang et al., 2014; Rasineni et al., 2019) pancreatic islets and increases  $\beta$ -cell apoptosis (Dembele et al., 2009; Kim et al., 2010). These findings support a role for alcohol-mediated impairment of pancreatic endocrine function, particularly of the integrity of  $\beta$ -cell response to glucose as an underlying mechanism of glucose dyshomeostasis. In addition, preclinical studies show that adiponectin, an insulin sensitizing adipokine with anti-inflammatory properties is decreased with at-risk alcohol use (Ford et al., 2016; Steiner and Lang, 2017).

A potential factor in alcohol-mediated decrease in adiponectin levels may be increased resistin levels in both clinical studies and animal models of alcohol use. Resistin suppresses adiponectin secretion and stimulates lipolysis, releasing fatty acids and glycerol into circulation (Chen et al., 2014). However, results from clinical studies are inconclusive, with both low to moderate and at-risk alcohol use increasing adiponectin (Steiner and Lang, 2017; Nova et al., 2019). Overall, alcohol-associated alterations in adipocytokine profile may contribute to impaired glucose homeostasis. Moreover, alcohol-mediated impairment of glycolytic pathways is associated with increased formation of advanced glycation end products (AGEs), which together with endothelial dysfunction, inflammatory responses, and oxidative stress discussed below contribute to the development of hypertension and atherosclerosis, important components of CMS (Vasdev et al., 2006). Recent data show that in PLWH at-risk alcohol use increased the likelihood of meeting the clinical criteria for prediabetes/diabetes (Primeaux et al., 2021). Moreover, HOMA- $\beta$  cell function negatively associated with AUDIT-C, phosphatidyl ethanol (PEth), and Timeline Followback (TLFB), suggesting that alcohol use is associated with impaired endocrine pancreatic function (Simon et al., 2020). Others have also reported a high incidence of diabetes in well controlled PLWH (Bratt et al., 2021), associated with increased oxidative stress (Bastard et al., 2019).

### Dysregulation of Lipid Homeostasis

Chronic alcohol use results in a dose-response positive relationship with high density lipoprotein-cholesterol (HDLc) and negative relationship with low density lipoprotein (LDL) (Shuval et al., 2012). The alcohol-mediated biological mechanisms contributing to lowering of cholesterol levels include increased transport of lipoproteins and lipoprotein lipase activity (Shuval et al., 2012), and increased eNOS activity that helps transport of HDL-C (Hirakawa et al., 2015; Wakabayashi, 2019). At-risk alcohol use increases triglyceride levels especially among women (Hirakawa et al., 2015; Vieira et al., 2016) and moderate alcohol is associated with lower triglyceride levels (Kovar and Zemankova, 2015). Alcohol-mediated mechanisms that contribute to increased triglycerides include adipose tissue lipolysis, and hepatic synthesis of large very low density lipoprotein (VLDL) particles through the increased expression of microsomal triglyceride transfer protein (Klop et al., 2013). The interactions of alcohol with high fat diet further exacerbate hypertriglyceridemia by regulating hypothalamic peptides that increase appetite and energy intake (Barson et al., 2010).

Alcohol directly and indirectly alters the balance of lipogenesis and lipolysis and thus dysregulates lipid metabolism (Figure 3). Loss in adipose tissue mass resulting from chronic alcohol consumption is partly due to an increase in triglyceride turnover without significant alteration in triglyceride synthesis (Kang et al., 2007a; Zhong et al., 2012). Alcohol-induced activation of lipolysis and release of free fatty acids (FFAs) that are taken up by the liver has been proposed as a central mechanism of alcohol-induced steatohepatitis. Overall, alcohol alters hepatic lipid flux by increasing free fatty acid uptake from the diet and from adipose tissue lipolysis; stimulating lipogenesis from glycolytic

end products; and dysregulating  $\beta$ -oxidation. Moreover, alcohol upregulates expression of fatty acid transporters particularly CD36/FAT promoting fatty acid uptake, and esterification of free fatty acids into triglycerides (You and Arteel, 2019). Alcohol dysregulates lipolysis through inhibitory effects on insulin action (Yki-Jarvinen et al., 1988), but not through enhanced adrenergic effects (Lang et al., 2014). In addition, alcohol increases plasma and adipose tissue expression of fibroblast growth factor 21 (FGF21), a known stimulus for lipolysis (Zhao et al., 2015). In contrast, alcohol decreases expression and activation of several lipogenic enzymes (Steiner and Lang, 2017). Thus, alcohol-associated dysregulation of lipid homeostasis results from an imbalance between lipolysis and lipogenesis. Dyslipidemia is highly prevalent among PLWH (Grunfeld et al., 1992), and this has been attributed to the infection itself, and some types of ART drugs. Early following HIV seroconversion, total, HDL, LDL cholesterol levels decrease. Following the initiation of ART, the profile shifts to increased total and LDL cholesterol to pre-infection levels, with persistent low HDL levels (Riddler et al., 2003). Alcohol-using PLWH have higher odds of presenting with lipodystrophy (Cheng et al., 2009). CVD risk factors, including dyslipidemia and hypertension, are determinants of reduced life expectancy in PLWH (Freiberg and Kraemer, 2010).

### Dysregulation of Blood Pressure

In addition to alcohol-induced metabolic dysregulation or instability, alcohol-associated risk for hypertension is also an important contributor to CMS (Saljoughian, 2016). Some studies report higher prevalence of hypertension in PLWH than in uninfected individuals, with a higher frequency in individuals 50 years of age or older (Xu et al., 2017; Fahme et al., 2018). Furthermore, hypertension in PLWH is associated with heavy alcohol use (Ikeda et al., 2013). Multiple mechanisms affected by alcohol and HIV infection can contribute to blood pressure dysregulation. Alcohol-induced dysregulation of vasoconstrictor levels, vasoreactivity, and endothelial integrity are all possible mechanisms contributing to altered blood pressure regulation. Alcohol at low doses produces transient vasodilation. In contrast, at higher doses and with binge drinking, alcohol leads to vasoconstriction (Rekik et al., 2002; Kloner and Rezkalla, 2007) and chronic heavy alcohol use increases the risk of hypertension (Kahl and Hillemecher, 2016; Vieira et al., 2016). Several mechanisms contribute to alcohol-associated hypertension including genetic predisposition for an *ALDH2* polymorphism (Hu et al., 2014), activation of renin angiotensin aldosterone system (RAAS), central adrenergic activity, increased sympathetic flow, vascular smooth muscle tone, shifts in baroreceptor reflex sensitivity, and decreased nitric oxide (NO) release (Husain et al., 2014; Kahl and Hillemecher, 2016). In addition, alcohol increases both cytosolic free calcium ( $\text{Ca}^{2+}$ ) and cellular  $\text{Ca}^{2+}$  uptake by direct upregulation of voltage-gated  $\text{Ca}^{2+}$  channels, inhibits  $\text{Ca}^{2+}$ -adenosine triphosphatase, and sodium-potassium ion pumps, thus increasing sensitivity to endogenous vasoconstrictors and exacerbating hypertension (Altura and Altura, 1982; Husain et al., 2014). The magnitude of increase in blood pressure with heavy alcohol use averages about 5–10 mm Hg with



greater increases in systolic than diastolic pressure (Clark, 1985). The association between alcohol use and hypertension is stronger among older individuals (Burke et al., 1992; Riserus and Ingelsson, 2007), which may be due to enhanced sensitivity to sympathoadrenal activation (Husain et al., 2014). Thus, alcohol-associated risk for hypertension results from alterations in endothelial function and in the levels and activity of endogenous mediators of vasoreactivity as discussed in the next section.

## Endothelial Dysfunction

Several alterations in vascular integrity and responsiveness may contribute to increased risk for hypertension and vascular instability. The vascular endothelium integrates humoral and hemodynamic signals modulating vasomotor tone and adjusting blood flow to the local tissue needs. Alcohol exerts biphasic effects on endothelial function (Puddey et al., 2001; **Figure 4**). Acute alcohol consumption produces immediate and transient vasodilation (Bau et al., 2005; Oda et al., 2020) followed by increased blood pressure later after alcohol consumption resulting in a biphasic effect of alcohol on blood pressure. The acute alcohol-induced vasodilation is mainly attributed to the increased expression of endothelial nitric oxide synthase (eNOS) and NO (Venkov et al., 1999). This mechanism has been proposed in cardioprotective benefits of moderate alcohol consumption (Abou-Agag et al., 2005; Toda and Ayajiki, 2010). Chronic and at-risk alcohol use is associated with hyperactivation of the hypothalamo-pituitary-adrenal (HPA) axis and increased glucocorticoid release. Excess glucocorticoid action can also contribute to endothelial dysfunction and accelerate the atherogenic process (Bau et al., 2005). Overall, at-risk alcohol use increases the risk of coronary artery disease, and hemorrhagic and ischemic stroke. Endothelial dysfunction is a predictor of CV disease and precedes atheromatous plaque formation (Schachinger and Zeiher, 2000). In addition, cytotoxicity resulting from at-risk alcohol use through generation of peroxynitrite, resulting from reaction of NO with superoxide; a product of alcohol-induced oxidative stress (Pacher et al., 2007), may also contribute to endothelial dysfunction. PLWH have significant impairment in endothelial function reflected in significantly lower flow-mediated vasodilation compared to uninfected controls, and present with elevated levels of markers of endothelial dysfunction (Rethy et al., 2020). Though evidence supports a deleterious effect of HIV infection on endothelial function, definitive studies remain limited and inconclusive.

## Dysregulation of Vasoconstrictor Levels and Vasoreactivity

At-risk alcohol use may alter the balance of endogenous vasoconstrictor levels including angiotensin II, endothelins, and norepinephrine (Tsuji et al., 1992), contributing to the increased risk for hypertension. Studies demonstrate that alcohol consumption increases overall activity of RAAS (Jing et al., 2008; Yarlioglues et al., 2019). Plasma renin activity is increased in individuals with heavy alcohol consumption (Husain et al., 2014) and angiotensin converting enzyme (ACE) activity in serum of hospitalized individuals with history of chronic alcohol

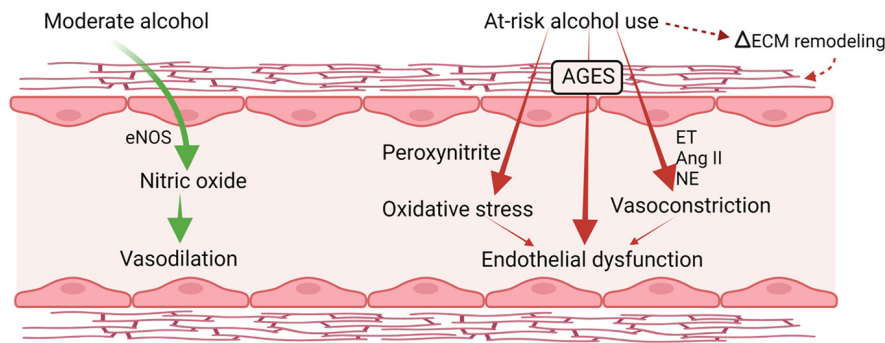
consumption is higher than that of healthy controls (Okuno et al., 1986). Similarly, some evidence supports a role for increased RAAS activation in development of hypertension and metabolic syndrome in PLWH (Srinivasa et al., 2015; Masenga et al., 2020). The importance of increased activation of the Ang II type 1 receptor (AT1) to the development of alcoholic cardiomyopathy has been shown in preclinical studies (Cheng et al., 2006). Furthermore, activation of the AT1 receptor by chronic alcohol activates NADPH oxidase producing endothelial injury (Warnholtz et al., 1999; Gonzaga et al., 2018; Assis et al., 2020). Alcohol increases levels of renin, aldosterone, and angiotensin II (Jing et al., 2008), circulating renin and ACE activity, and left ventricular myocyte AT1 receptor expression in individuals with at-risk alcohol use (Husain et al., 2014). In addition, at-risk alcohol use results in increased plasma vasopressin levels indicating increased sympathetic stimulation (Chan and Sutter, 1983). Together with alterations in endothelial function, the alcohol-induced dysregulation of circulating levels of endogenous vasoactive mediators and their respective receptors promotes alterations in blood pressure regulation, increasing the risk for hypertension.

## Alcohol-Human Immunodeficiency Virus Interactions Increase Risk for Neurological Impairment in Persons Living With Human Immunodeficiency Virus

CMS has been linked to neurocognitive impairment in the general population (Yaffe et al., 2004; Panza et al., 2010), particularly in the presence of inflammation. More recently, this link has been examined in the context of HIV infection, demonstrating an additive effect of HIV and CMS on neurocognitive dysfunction, including apathy and decreased executive function (Pope et al., 2020), and impairments in the neurocognitive domains of learning and fine motor skills (Yu et al., 2019). The neurological consequences of chronic HIV infection span both the peripheral and central nervous systems, and despite the success of ART to control viral loads, HIV infection and treatment continue to promote neurological and psychiatric co-morbidities that worsen disease outcomes and overall quality of life. Neurocognitive disorders are highly prevalent in PLWH, and HIV-associated neurocognitive disorder (HAND) and co-occurring AUD can exacerbate these deficits (Fama et al., 2016). HAND can include deficits in attention, memory, and executive function. ARTs have successfully extended life expectancy of PLWH but are not fully protective against the neurocognitive symptoms of HAND and therefore greater cognitive impairment may surface with aging (Alakkas et al., 2019).

The cognitive and behavioral deficits associated with excessive drinking are commonly attributed to long-lasting neuroadaptations and functional changes to neuronal circuitry, as both former and current AUD subjects demonstrate cognitive impairments including deficits in working memory (Kopera et al., 2012), executive functioning (Green et al., 2010), and impulsivity (Li et al., 2009), which all contribute to maladaptive decision-making, including relapse to alcohol seeking during attempted abstinence periods. Interestingly, such deficits partially but incompletely overlap between men and women





**FIGURE 4 |** Alcohol-induced biphasic effects on endothelial function. Acutely, moderate doses of alcohol increase production of nitric oxide and result in transient vasodilation. Chronic, heavy alcohol consumption produces oxidative stress and activates vasoconstrictor pathways including endothelins (ET) and angiotensin II formation (Ang II) that lead to endothelial dysfunction and altered response to vasoactive mediators. ET, endothelin; Ang II, angiotensin II; NE, norepinephrine; eNOS, endothelial nitric oxide synthase; AGES, advance glycation end products; ECM, extracellular matrix. Created with Biorender.com.

(Fama et al., 2020). From a neuroanatomical perspective, these chronic alcohol induced-cognitive impairments are associated with selective brain damage in vulnerable areas such as the prefrontal cortex (Pfefferbaum et al., 1998; Le Berre et al., 2014) and hippocampus (Mira et al., 2019). Heavy alcohol use may also predispose individuals to cognitive disorders such as Alzheimer's disease (AD), as frequent drinkers display higher levels of AD-related biomarkers in their cerebrospinal fluid (Wang et al., 2021). In addition, a rapidly emerging area of interest is how sleep disturbances impact neurological function and may interact with cognitive deficits to promote AUD (Laniepce et al., 2021).

Neuroinflammation is hypothesized to be a primary mechanism of HAND-associated cognitive impairment (Garvey et al., 2014). However, it is becoming increasingly recognized that neuroinflammation alone cannot explain the neurological consequences of HIV (Gelman, 2015). In a murine model of HIVE, neuropathology persists despite suppressed neuroinflammation and CNS viral replication. Preclinical studies suggest that disruptions in growth factor expression and signaling likely contributes to HIV neuropathology and cognitive impairments (Maxi et al., 2016). Chronic binge alcohol unmasks neurocognitive deficits in SIV-infected, non-ART treated macaques (Winsauer et al., 2002). Furthermore, disruptions in growth factor expression and signaling as potential mechanisms contributing to cognitive impairments have been reported. Brain-derived neurotrophic factor (BDNF) is a neuroprotective factor regulating synaptic plasticity and cognitive function. Reduced BDNF signaling through TrkB, including ERK and Akt pathways, have been shown to decrease neuronal plasticity and are linked to HIV-associated neurocognitive decline (Michael et al., 2020). *In vitro* studies have indicated that the HIV peptide *Tat* induces downregulation of *BDNF* and other CREB-regulated genes. Alcohol consumption can further reduce BDNF levels and impair BDNF signaling through TrkB receptors (John MacLennan et al., 1995). Interestingly, though ART attenuates expression of microglial markers of neuroinflammation in the frontal cortex and monocyte/macrophage markers of neuroinflammation in the basal ganglia, it did not ameliorate the alcohol-associated inhibition of growth factor signaling in the

frontal cortex of SIV-infected macaques. These findings suggest that while ART may be effective in reducing neuroinflammation associated with infection and alcohol, it is not sufficient to attenuate the deficits in BDNF signaling and may explain the persistence of HAND despite widespread ART use (Maxi et al., 2019). These results support the need for continued research into strategies to prevent HIV-associated neurocognitive decline, including the potential to target neuroinflammation and growth factor signaling as new therapies are developed.

### Neuropathological Comorbidities Associated With At-Risk Alcohol Use and Human Immunodeficiency Virus

In addition to the direct and indirect alcohol-mediated peripheral tissue injury implicated in pathophysiology of CMS, at-risk alcohol use also impacts both peripheral and central nervous system physiology. This is perhaps best represented by alcohol's ability to regulate the conscious experience of pain through its interactions with both ascending and descending nociceptive circuitry (Egli et al., 2012). Importantly, chronic pain affects approximately 20% of adults worldwide and approximately 100 million Americans (Nahin, 2015), a number that will likely increase over the next few decades given an aging U.S. population. Chronic or neuropathic pain is prevalent in approximately 57% of PLWH, significantly impairing quality of life (Ellis et al., 2010). PLWH with HIV-related pain also have higher rates of depression, anxiety, and insomnia, compared to those without neuropathy symptoms (Phillips et al., 2014), indicating that neurological pain symptoms may ultimately transform into CNS pathophysiology including cognitive dysfunction and psychiatric illness. Preclinical studies show significant heightened pain sensitivity in response to systemic injection of HIV viral particles (Guindon et al., 2019; Bagdas et al., 2020). SIV infection in pigtail macaques induces inflammatory cell infiltration in the spinal cord, a frequent finding in chronic pain conditions (Mangus et al., 2015).

Evidence also suggests that diet and metabolic factors influence the development and chronicity of pain symptoms.

Diets containing high amounts of omega-6 fatty acids (common in Western diets) exacerbate nociceptive sensitivity in preclinical animal models of neuropathic and inflammatory pain (Boyd et al., 2021). Importantly, Western diets appear to prolong recovery from a pain-related injury in animals, although recovery was accelerated by switching animals to an anti-inflammatory diet (Totsch et al., 2018). Similar dietary interventions have also demonstrated efficacy in reducing pain symptoms in humans (Lattanzio and Imbesi, 2018). A review of global studies also highlighted links between affective disorders, metabolic syndrome, and chronic pain (Bica et al., 2017), strongly suggesting that negative affect serves as a moderator of these relationships. Convergent evidence suggests a strong underlying influence of chronic inflammation in these integrated disease processes, providing a mechanistic basis for how HIV and alcohol use promotes and sustains these conditions.

### At-Risk Alcohol Use, Chronic Pain, and Pain-Related Negative Affect

While the analgesic effects of alcohol have been known for some time (Thompson et al., 2017; Cucinello-Ragland and Edwards, 2021), excessive alcohol exposure damages elements of the peripheral nervous system producing a characteristic small fiber painful neuropathy (Maiya and Messing, 2014), and the resulting increased nociceptive sensitivity (termed hyperalgesia) is hypothesized to contribute to an increased motivational drive to drink (Egli et al., 2012). In addition to the direct effects of alcohol, abstinence from heavy drinking can also increase affective pain sensitivity (termed hyperkatifeia) as part of a larger motivational withdrawal syndrome (Edwards et al., 2012; Koob, 2021) that may also promote the use of alcohol for affective/emotional pain management. Indeed, self-reports of alcohol use specifically for pain are common. Problem drinkers of both sexes report more severe pain symptoms compared to non-drinkers, and also a higher incidence of using alcohol to manage their pain (Brennan et al., 2005). Interestingly, use of alcohol to manage pain symptoms presaged a worsening of alcohol drinking-related comorbidities, including diabetes (Brennan et al., 2005).

Acute pain represents an adaptive sensory process vital to protecting our bodies from damage. In contrast, chronic or unrelieved pain represents a profound negative emotional experience that can have a powerful influence on brain reinforcement mechanisms, possibly facilitating the transition to AUD in vulnerable individuals (Zale et al., 2021). As support for this conceptualization, there are strong associations among alcohol consumption, chronic pain, and pain-related disability (Witkiewitz et al., 2015a,b; Yeung et al., 2020). Evidence suggests that chronic pain may be predictive of future at-risk alcohol use. In one prospective epidemiological study, self-reported pain interference (or how pain disrupts daily life activities) was predictive of AUD development (McDermott et al., 2018). Additional preclinical research suggests that the relationships between individual levels of alcohol drinking and resultant chronic pain relief may change over time

(Adrienne McGinn et al., 2020), although the neurobiological basis of these links is currently unknown.

In addition to chronic pain, important bi-directional relationships exist between at-risk alcohol drinking and other negative affective conditions (e.g., stress, anxiety, depression) that represent additional risks for psychiatric co-morbidities. Importantly, people with chronic pain and high negative affect report higher pain severity and pain interference compared to people with only one of the two disorders (Arnouk et al., 2006). Such symptoms may precede or emerge within the development of more severe forms of AUD (Gilpin et al., 2015). For example, the development of negative affective states comprises a portion of the constellation of symptoms of alcohol withdrawal, and as such, likely contribute to negative reinforcement processes that drive continued or escalated drinking over time (Tolomeo et al., 2021). Another intense line of preclinical and clinical investigation concerns factors that precede and may presage the development of at-risk drinking. These range from neurological conditions including traumatic brain injury (Phillips et al., 2014; Mayeux et al., 2015; Adams et al., 2020; Schindler et al., 2021) to pre-existing psychiatric disorders such as post-traumatic stress disorder (Whitaker et al., 2014; Langdon et al., 2016; Livingston et al., 2021).

### Confounding Factors Impacting Alcohol-Associated Risk for Cardiometabolic Syndrome and Cognitive Deficits

Several biological factors may contribute to alcohol-mediated tissue injury and risk for CMS, including the pattern and type of alcohol use, sex, existing underlying comorbid conditions, and alcohol-induced alterations in target organ milieu (i.e., dysregulation of the extracellular matrix). Both acute and chronic alcohol consumption decrease total brain glucose uptake and the rate of glucose utilization (Volkow et al., 2013), and this may be associated with decreased neuronal activity. These alcohol-associated alterations in brain glucose metabolism may have significant implications for increased risk of cognitive dysfunction, particularly in PLWH (Hammoud et al., 2018; Ge et al., 2021). Binge drinking increases the risk of CMS, type 2 diabetes, stroke, and coronary artery disease compared to continuous alcohol consumption (Lindtner et al., 2013; Hong et al., 2015), and more frequent moderate drinking is associated with more favorable outcomes than occasional or weekly drinking. Moreover, occasional heavy drinking rather than regular heavy drinking is associated with central obesity and hyperglycemia (Wakabayashi, 2014). Increased drinking frequency was associated with increased triglycerides, hyperglycemia, blood pressure, and abdominal obesity only among men (Schroder et al., 2007), and hyperglycemia and hypertension in women (Lee, 2012).

The type of alcohol consumed, and the rate of alcohol metabolism may have a differential impact on CMS risk. Wine that is rich in polyphenols is most associated with decreased CMS incidence (Beulens et al., 2012; Rasouli et al., 2013), and beer in larger doses increases the odds ratio of having a higher waist-hip ratio together with elevated blood pressure and triglycerides (Koloverou et al., 2015; Vieira et al., 2016).

Polyphenols found in alcohol, especially resveratrol, are anti-inflammatory and increase eNOS activity, which not only allows for vasodilation but also increases HDL-c transport. Polyphenols also decrease accumulation of 3-nitotyrosine, and PARylated (poly ADP-ribosylated) proteins associated with diabetes (Drel and Sybirna, 2010). Alcohol metabolism differs between sexes. Females have higher percentage of body fat, lower water content, and lower gastric ADH activity than males and these sex-specific differences result in higher blood alcohol concentrations in women than in men given consumption of a similar amount of alcohol (Cowan et al., 1996).

Emerging evidence suggests that Western diets commonly consumed in the U.S. worsen both neurocognitive (Martin and Davidson, 2014; Davidson et al., 2019) and pain symptomatology (Boyd et al., 2021). Thus, the neurobiological interaction of excessive alcohol drinking, cognition, and pain in the context of WD consumption represents a critical area of research and public health interest.

Although alcohol-mediated alterations in extracellular matrix (ECM) remodeling, characterized by an imbalance of extracellular matrix protein synthesis and degradation leading to overt fibrosis has been well described in alcohol-related liver injury (Poole and Arteel, 2016; Dolin and Arteel, 2020); more recently, dysregulation of matrix proteins has been reported in adipose, cardiac, SKM, brain, and lung (Sueblinvong et al., 2014; Steiner et al., 2015b; Mouton et al., 2016; Ninh et al., 2019). This altered ECM phenotype has functional consequences that can lead to or exacerbate tissue dysfunction and increase risk for CMS. The major proteins that make up the ECM include collagens, fibronectin, laminin, elastin, and proteoglycans and the continuous remodeling of the matrix relies on the expression and activity of metalloproteases and their tissue inhibitors (Poole and Arteel, 2016). Studies using both *in vivo* and *in vitro* preclinical models, and postmortem samples of people with AUD have shown that alcohol dysregulates matrix proteins including laminins, collagen, PAI-1, and tissue plasminogen activator (tPA) (Trindade et al., 2016; Rubio-Araiz et al., 2017; Go et al., 2020). In parallel, alcohol increases AGEs (Vasdev et al., 2006) and the irreversible cross-links of collagens and elastin are unorganized and have dysfunctional ECM fiber distribution (Zieman et al., 2005). AGEs also quench NO and generate peroxynitrite contributing to endothelial dysfunction. In addition, alcohol-mediated oxidative stress induces transforming growth factor beta-1 (TGF $\beta$ 1) (Sueblinvong et al., 2014), which in turn activates Smad signaling impacting target genes involved in maintaining ECM homeostasis such as PAI-1, urokinase plasminogen activator (uPA), and collagen (Seth et al., 2010). Alcohol dysregulates ECM remodeling producing a profibrotic milieu in SKM (Dodd et al., 2014; Simon et al., 2015) and adipose tissue (Ford et al., 2018) and these alterations in the matrix are associated with decreased SKM metabolic function and regenerative capacity (LeCapitaine et al., 2011; Simon et al., 2014; Duplanty et al., 2018; Ford et al., 2018) in SIV-infection. Thus, alcohol-induced dysregulation of the ECM contributes to tissue injury and risk of CMS. The ECM plays an important role in brain development, maturation of neural circuits, and adult neuroplasticity, suggesting that processes that affect

composition or turnover of brain ECM could impair brain function and contribute to development of neuropsychiatric or neurodegenerative disease (Lubbers et al., 2014; Senkov et al., 2014). Studies show that alcohol affects the regulation of brain ECM through various mechanisms throughout the lifespan. Because of the important role the ECM plays in synaptic processes affected by alcohol exposure, brain ECM remodeling may be an important contributor to the pathophysiology of AUD (Lasek, 2016) and is an area worthy of further investigation.

Taken together, data from clinical and preclinical studies strongly suggest that neuropathological comorbidities in PLWH result from a constellation of pathophysiological mechanisms including metabolic dyshomeostasis, neuroinflammation (Perkins et al., 2019; Delery and Edwards, 2020), and loss of neurotrophic support (Somkuwar et al., 2016; Maxi et al., 2019; Silva-Pena et al., 2019). Comorbid at-risk alcohol use exacerbates these neuropathological processes (Zahr, 2018) and a greater understanding of these mechanisms will lead to new and more effective pharmacological and behavioral strategies for treating AUD (Ray et al., 2021).

## CONCLUSION

The multisystemic pathophysiological effects of at-risk alcohol use alter underlying cellular and organ homeostasis predisposing the host for increased risk for comorbidities. These alterations are particularly salient in vulnerable populations such as those with chronic diseases, diminishing physiological reserve and increasing vulnerability to tissue injury resulting from direct and indirect effects of alcohol. In PLWH, at-risk alcohol use exacerbates cardiometabolic and neurocognitive pathologies that together with chronic use of ART lead to development of geriatric comorbidities manifested in frailty. Overall, these data strongly support the association of AUD with accelerated biological aging and enhanced risk for comorbidities. Interventions aimed at diminishing at-risk alcohol use are urgently needed in this vulnerable population. Greater understanding of the underlying mechanisms and their specific contribution to comorbidity risk is likely to identify therapeutic targets to ameliorate tissue injury and protect organ systems from the combined impact of HIV infection, chronic ART, and alcohol-mediated tissue injury.

## AUTHOR CONTRIBUTIONS

LS, SE, and PM conceived and drafted the review, edited, and approved the final version. All authors contributed to the article and approved the submitted version.

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## REFERENCES

- Aberg, F., Farkkila, M., and Mannisto, V. (2020). Interaction between alcohol use and metabolic risk factors for liver disease: a critical review of epidemiological studies. *Alcohol. Clin. Exp. Res.* 44, 384–403. doi: 10.1111/acer.14271
- Abou-Agag, L. H., Khoo, N. K., Binsack, R., White, C. R., Darley-Usmar, V., Grenett, H. E., et al. (2005). Evidence of cardiovascular protection by moderate alcohol: role of nitric oxide. *Free Radic. Biol. Med.* 39, 540–548. doi: 10.1016/j.freeradbiomed.2005.04.007
- Achhra, A. C., Sabin, C., Ryom, L., Hatleberg, C., Antonella D'aminio, M., De Wit, S., et al. (2018). Body mass index and the risk of serious Non-AIDS events and all-cause mortality in treated HIV-Positive individuals: D: a: D cohort analysis. *J. Acquir. Immune Defic. Syndr.* 78, 579–588. doi: 10.1097/QAI.0000000000001722
- Adams, R. S., Campbell-Sills, L., Stein, M. B., Sun, X., Larson, M. J., Kessler, R. C., et al. (2020). The association of lifetime and deployment-acquired traumatic brain injury with postdeployment binge and heavy drinking. *J. Head Trauma Rehabil.* 35, 27–36. doi: 10.1097/HTR.0000000000000508
- Adrienne, McGinn, M., Edwards, K. N., and Edwards, S. (2020). Chronic inflammatory pain alters alcohol-regulated frontocortical signaling and associations between alcohol drinking and thermal sensitivity. *Neurobiol. Pain* 8:100052. doi: 10.1016/j.nypai.2020.100052
- Alakkas, A., Ellis, R. J., Watson, C. W., Umlauf, A., Heaton, R. K., Letendre, S., et al. (2019). White matter damage, neuroinflammation, and neuronal integrity in HAND. *J. Neurovirol.* 25, 32–41. doi: 10.1007/s13365-018-0682-9
- Alcohol Research, Current Reviews, and Editorial Staff. (2018). Drinking patterns and their definitions. *Alcohol Res.* 39, 17–18.
- Alkerwi, A., Boutsen, M., Vaillant, M., Barre, J., Lair, M. L., Albert, A., et al. (2009). Alcohol consumption and the prevalence of metabolic syndrome: a meta-analysis of observational studies. *Atherosclerosis* 204, 624–635. doi: 10.1016/j.atherosclerosis.2008.10.036
- Altura, B. M., and Altura, B. T. (1982). Microvascular and vascular smooth muscle actions of ethanol, acetaldehyde, and acetate. *Fed Proc.* 41, 2447–2451.
- Arab, J. P., Arrese, M., and Shah, V. H. (2020). Gut microbiota in non-alcoholic fatty liver disease and alcohol-related liver disease: current concepts and perspectives. *Hepato. Res.* 50, 407–418. doi: 10.1111/hepr.13473
- Arnou, B. A., Hunkeler, E. M., Blasey, C. M., Lee, J., Constantino, M. J., Fireman, B., et al. (2006). Comorbid depression, chronic pain, and disability in primary care. *Psychosom. Med.* 68, 262–268. doi: 10.1097/01.psy.0000204851.15499.fc
- Assis, V. O., Gonzaga, N. A., Silva, C. B. P., Pereira, L. C., Padovan, C. M., and Tirapelli, C. R. (2020). Ethanol withdrawal alters the oxidative state of the heart through AT1-Dependent mechanisms. *Alcohol Alcohol.* 55, 3–10. doi: 10.1093/alcalc/aggz101
- Aukrust, P., Muller, F., Svardal, A. M., Ueland, T., Berge, R. K., and Froland, S. S. (2003). Disturbed glutathione metabolism and decreased antioxidant levels in human immunodeficiency virus-infected patients during highly active antiretroviral therapy-potential immunomodulatory effects of antioxidants. *J. Infect. Dis.* 188, 232–238. doi: 10.1086/376459
- Bagdas, D., Paris, J. J., Carper, M., Wodarski, R., Rice, A. S. C., Knapp, P. E., et al. (2020). Conditional expression of HIV-1 tat in the mouse alters the onset and progression of tonic, inflammatory and neuropathic hypersensitivity in a sex-dependent manner. *Eur. J. Pain* 24, 1609–1623. doi: 10.1002/ejp.1618
- Baliunas, D. O., Taylor, B. J., Irving, H., Roerecke, M., Patra, J., Mohapatra, S., et al. (2009). Alcohol as a risk factor for type 2 diabetes: a systematic review and meta-analysis. *Diabetes Care* 32, 2123–2132. doi: 10.2337/dc09-0227
- Baranyai, J. M., and Blum, J. J. (1989). Quantitative analysis of intermediary metabolism in rat hepatocytes incubated in the presence and absence of ethanol with a substrate mixture including ketoleucine. *Biochem. J.* 258, 121–140. doi: 10.1042/bj2580121
- Barson, J. R., Morganstern, I., and Leibowitz, S. F. (2010). Galanin and consummatory behavior: special relationship with dietary fat, alcohol and circulating lipids. *Exp. Suppl.* 102, 87–111. doi: 10.1007/978-3-0346-0228-0\_8
- Bastard, J. P., Couffignal, C., Fellahi, S., Bard, J. M., Mentre, F., Salmon, D., et al. (2019). Diabetes and dyslipidaemia are associated with oxidative stress independently of inflammation in long-term antiretroviral-treated HIV-infected patients. *Diabetes Metab.* 45, 573–581. doi: 10.1016/j.diabet.2019.02.008
- Batterham, M. J., Garsia, R., and Greenop, P. A. (2000). Dietary intake, serum lipids, insulin resistance and body composition in the era of highly active antiretroviral therapy 'Diet FRS Study'. *AIDS* 14, 1839–1843. doi: 10.1097/00002030-200008180-00020
- Bau, P. F., Bau, C. H., Naujorks, A. A., and Rosito, G. A. (2005). Early and late effects of alcohol ingestion on blood pressure and endothelial function. *Alcohol* 37, 53–58. doi: 10.1016/j.alcohol.2005.10.034
- Beires, M. T., Silva-Pinto, A., Santos, A. C., Madureira, A. J., Pereira, J., Carvalho, D., et al. (2018). Visceral adipose tissue and carotid intima-media thickness in HIV-infected patients undergoing cART: a prospective cohort study. *BMC Infect. Dis.* 18:32. doi: 10.1186/s12879-017-2884-9
- Beulens, J. W., Van Der Schouw, Y. T., Bergmann, M. M., Rohrmann, S., Schulze, M. B., Buijsse, B., et al. (2012). Alcohol consumption and risk of type 2 diabetes in European men and women: influence of beverage type and body size the EPIC-InterAct study. *J. Intern. Med.* 272, 358–370. doi: 10.1111/j.1365-2796.2012.02532.x
- Bica, T., Castello, R., Toussaint, L. L., and Monteso-Curto, P. (2017). Depression as a risk factor of organic diseases: an international integrative review. *J. Nurs. Scholarsh.* 49, 389–399. doi: 10.1111/jnu.12303
- Boccaro, F. (2008). Cardiovascular complications and atherosclerotic manifestations in the HIV-infected population: type, incidence and associated risk factors. *AIDS* 22(Suppl. 3), S19–S26. doi: 10.1097/01.aids.0000327512.76126.6e
- Bonnet, F., Disse, E., Laville, M., Mari, A., Hojlund, K., Anderwald, C. H., et al. (2012). Moderate alcohol consumption is associated with improved insulin sensitivity, reduced basal insulin secretion rate and lower fasting glucagon concentration in healthy women. *Diabetologia* 55, 3228–3237. doi: 10.1007/s00125-012-2701-3
- Boyd, J. T., Lococo, P. M., Furr, A. R., Bendele, M. R., Tram, M., Li, Q., et al. (2021). Elevated dietary omega-6 polyunsaturated fatty acids induce reversible peripheral nerve dysfunction that exacerbates comorbid pain conditions. *Nat. Metab.* 3, 762–773. doi: 10.1038/s42255-021-00410-x
- Bratt, G., Brannstrom, J., Missalidis, C., and Nystrom, T. (2021). Development of type 2 diabetes and insulin resistance in people with HIV infection: prevalence, incidence and associated factors. *PLoS One* 16:e0254079. doi: 10.1371/journal.pone.0254079
- Brennan, P. L., Schutte, K. K., and Moos, R. H. (2005). Pain and use of alcohol to manage pain: prevalence and 3-year outcomes among older problem and non-problem drinkers. *Addiction* 100, 777–786. doi: 10.1111/j.1360-0443.2005.01074.x
- Breslow, R. A., Guenther, P. M., and Smothers, B. A. (2006). Alcohol drinking patterns and diet quality: the 1999-2000 national health and nutrition examination survey. *Am. J. Epidemiol.* 163, 359–366. doi: 10.1093/aje/kwj050
- Breslow, R. A., Guenther, P. M., Juan, W., and Graubard, B. I. (2010). Alcoholic beverage consumption, nutrient intakes, and diet quality in the US adult population, 1999-2006. *J. Am. Diet. Assoc.* 110, 551–562. doi: 10.1016/j.jada.2009.12.026
- Burke, V., Beilin, L. J., German, R., Grosskopf, S., Ritchie, J., Puddey, I. B., et al. (1992). Association of lifestyle and personality characteristics with blood pressure and hypertension: a cross-sectional study in the elderly. *J. Clin. Epidemiol.* 45, 1061–1070. doi: 10.1016/0895-4356(92)90146-E
- Burton, R., and Sheron, N. (2018). No level of alcohol consumption improves health. *Lancet* 392, 987–988. doi: 10.1016/S0140-6736(18)31571-X
- Castro, J. P., El-Atat, F. A., Mcfarlane, S. I., Aneja, A., and Sowers, J. R. (2003). Cardiometabolic syndrome: pathophysiology and treatment. *Curr. Hypertens. Rep.* 5, 393–401. doi: 10.1007/s11906-003-0085-y
- CDC (2018). *Estimated HIV Incidence and Prevalence in the United States 2010–2015HIV Surveillance Supplemental Report*. Atlanta, GA: CDC.
- Cederbaum, A. I. (2012). Alcohol metabolism. *Clin. Liver. Dis.* 16, 667–685. doi: 10.1016/j.cld.2012.08.002
- Chan, T. C., and Sutter, M. C. (1983). Ethanol consumption and blood pressure. *Life Sci.* 33, 1965–1973. doi: 10.1016/0024-3205(83)90734-8
- Chen, N., Zhou, L., Zhang, Z., Xu, J., Wan, Z., and Qin, L. (2014). Resistin induces lipolysis and suppresses adiponectin secretion in cultured human visceral adipose tissue. *Regul. Pept.* 194–195, 49–54. doi: 10.1016/j.regpep.2014.10.001
- Cheng, C. P., Cheng, H. J., Cunningham, C., Shihabi, Z. K., Sane, D. C., Wannenburg, T., et al. (2006). Angiotensin II type 1 receptor blockade



- prevents alcoholic cardiomyopathy. *Circulation* 114, 226–236. doi: 10.1161/CIRCULATIONAHA.105.596494
- Cheng, D. M., Libman, H., Bridden, C., Saitz, R., and Samet, J. H. (2009). Alcohol consumption and lipodystrophy in HIV-infected adults with alcohol problems. *Alcohol* 43, 65–71. doi: 10.1016/j.alcohol.2008.09.004
- Childs, C. E., Calder, P. C., and Miles, E. A. (2019). Diet and immune function. *Nutrients* 11:1933. doi: 10.3390/nu11081933
- Churilla, J. R., Johnson, T. M., Curls, R., Richardson, M. R., Boyer, W. R., Devore, S. R., et al. (2014). Association between alcohol consumption patterns and metabolic syndrome. *Diabetes Metab. Syndr.* 8, 119–123. doi: 10.1016/j.dsx.2014.04.001
- Clark, L. T. (1985). Alcohol-induced hypertension: mechanisms, complications, and clinical implications. *J. Natl. Med. Assoc.* 77, 385–389.
- Cowan, J. M. Jr., Weathermon, A., Mccutcheon, J. R., and Oliver, R. D. (1996). Determination of volume of distribution for ethanol in male and female subjects. *J. Anal. Toxicol.* 20, 287–290. doi: 10.1093/jat/20.5.287
- Cruss, D. G., Pettito, J. M., Leserman, J., Douglas, S. D., Gettes, D. R., Ten Have, T. R., et al. (2003). Depression and HIV infection: impact on immune function and disease progression. *CNS Spectr.* 8, 52–58. doi: 10.1017/S1092852900023452
- Cucinello-Ragland, J. A., and Edwards, S. (2021). Neurobiological aspects of pain in the context of alcohol use disorder. *Int. Rev. Neurobiol.* 157, 1–29. doi: 10.1016/bs.irn.2020.09.001
- Davidson, T. L., Jones, S., Roy, M., and Stevenson, R. J. (2019). The cognitive control of eating and body weight: it's more than what you "Think". *Front. Psychol.* 10:62. doi: 10.3389/fpsyg.2019.00062
- Delery, E. C., and Edwards, S. (2020). Neuropeptide and cytokine regulation of pain in the context of substance use disorders. *Neuropharmacology* 174:108153. doi: 10.1016/j.neuropharm.2020.108153
- Dembele, K., Nguyen, K. H., Hernandez, T. A., and Nyomba, B. L. (2009). Effects of ethanol on pancreatic beta-cell death: interaction with glucose and fatty acids. *Cell Biol. Toxicol.* 25, 141–152. doi: 10.1007/s10565-008-9067-9
- Desquilbet, L., Jacobson, L. P., Fried, L. P., Phair, J. P., Jamieson, B. D., Holloway, M., et al. (2007). HIV-1 infection is associated with an earlier occurrence of a phenotype related to frailty. *J. Gerontol. A Biol. Sci. Med. Sci.* 62, 1279–1286. doi: 10.1093/gerona/62.11.1279
- Dodd, T., Simon, L., Lecapitaine, N. J., Zabaleta, J., Mussell, J., Berner, P., et al. (2014). Chronic binge alcohol administration accentuates expression of pro-fibrotic and inflammatory genes in the skeletal muscle of simian immunodeficiency virus-infected macaques. *Alcohol. Clin. Exp. Res.* 38, 2697–2706. doi: 10.1111/acer.12545
- Dolin, C. E., and Arteel, G. E. (2020). The matrisome, inflammation, and liver disease. *Semin Liver Dis.* 40, 180–188. doi: 10.1055/s-0039-3402516
- Dragan, N., Micic, M., Dimitrijević-Srečković, V., Kerkez, M., and Nikolić, B. (2017). Effect of alcohol on insulin secretion and viability of human pancreatic islets. *Srp. Arh. Celok. Lek.* 145, 159–164. doi: 10.2298/SARH160204023N
- Drel, V. R., and Sybirna, N. (2010). Protective effects of polyphenolics in red wine on diabetes associated oxidative/nitrate stress in streptozotocin-diabetic rats. *Cell Biol. Int.* 34, 1147–1153. doi: 10.1042/CBI20100201
- Duplanty, A. A., Siggins, R. W., Allerton, T., Simon, L., and Molina, P. E. (2018). Myoblast mitochondrial respiration is decreased in chronic binge alcohol administered simian immunodeficiency virus-infected antiretroviral-treated rhesus macaques. *Physiol. Rep.* 6:e13625. doi: 10.14814/phy2.13625
- Edwards, S., and Koob, G. F. (2010). Neurobiology of dysregulated motivational systems in drug addiction. *Future Neurol.* 5, 393–401. doi: 10.2217/fnl.10.14
- Edwards, S., Vendruscolo, L. F., Gilpin, N. W., Wojnar, M., and Witkiewitz, K. (2020). Alcohol and pain: a translational review of preclinical and clinical findings to inform future treatment strategies. *Alcohol. Clin. Exp. Res.* 44, 368–383. doi: 10.1111/acer.14260
- Edwards, S., Vendruscolo, L. F., Schlosburg, J. E., Misra, K. K., Wee, S., Park, P. E., et al. (2012). Development of mechanical hypersensitivity in rats during heroin and ethanol dependence: alleviation by CRF(1) receptor antagonism. *Neuropharmacology* 62, 1142–1151. doi: 10.1016/j.neuropharm.2011.11.006
- Egli, M., Koob, G. F., and Edwards, S. (2012). Alcohol dependence as a chronic pain disorder. *Neurosci. Biobehav. Rev.* 36, 2179–2192. doi: 10.1016/j.neubiorev.2012.07.010
- Ellis, R. J., Rosario, D., Clifford, D. B., McArthur, J. C., Simpson, D., Alexander, T., et al. (2010). Continued high prevalence and adverse clinical impact of human immunodeficiency virus-associated sensory neuropathy in the era of combination antiretroviral therapy: the CHARTER Study. *Arch. Neurol.* 67, 552–558. doi: 10.1001/archneurol.2010.76
- Fahme, S. A., Bloomfield, G. S., and Peck, R. (2018). Hypertension in HIV-infected adults: novel pathophysiologic mechanisms. *Hypertension* 72, 44–55. doi: 10.1161/HYPERTENSIONAHA.118.10893
- Fama, R., Le Berre, A. P., and Sullivan, E. V. (2020). Alcohol's Unique effects on cognition in women: a 2020 (Re)view to envision future research and treatment. *Alcohol Res.* 40:3. doi: 10.35946/arcv.v40.2.03
- Fama, R., Sullivan, E. V., Sassoon, S. A., Pfefferbaum, A., and Zahr, N. M. (2016). Impairments in component processes of executive function and episodic memory in alcoholism, HIV infection, and HIV infection with alcoholism comorbidity. *Alcohol. Clin. Exp. Res.* 40, 2656–2666. doi: 10.1111/acer.13250
- Ferguson, T. F., Theall, K. P., Brashear, M., Maffei, V., Beauchamp, A., Siggins, R. W., et al. (2020). Comprehensive assessment of alcohol consumption in people living with HIV (PLWH): the new orleans alcohol use in HIV study. *Alcohol. Clin. Exp. Res.* 44, 1261–1272. doi: 10.1111/acer.14336
- Ford, S. M. Jr., Simon Peter, L., Berner, P., Cook, G., Vande Stouwe, C., et al. (2018). Differential contribution of chronic binge alcohol and antiretroviral therapy to metabolic dysregulation in SIV-infected male macaques. *Am. J. Physiol. Endocrinol. Metab.* 315, E892–E903. doi: 10.1152/ajpendo.00175.2018
- Ford, S. M. Jr., Simon, L., Vande Stouwe, C., Allerton, T., Mercante, D. E., et al. (2016). Chronic binge alcohol administration impairs glucose-insulin dynamics and decreases adiponectin in asymptomatic simian immunodeficiency virus-infected macaques. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 311, R888–R897. doi: 10.1152/ajpregu.00142.2016
- Freiberg, M. S., and Kraemer, K. L. (2010). Focus on the heart: alcohol consumption, HIV infection, and cardiovascular disease. *Alcohol Res. Health* 33, 237–246.
- Garvey, L. J., Pavese, N., Politis, M., Ramlackhansingh, A., Brooks, D. J., Taylor-Robinson, S. D., et al. (2014). Increased microglia activation in neurologically asymptomatic HIV-infected patients receiving effective ART. *AIDS* 28, 67–72. doi: 10.1097/01.aids.0000432467.54003.f7
- Ge, I., Kirschen, G. W., and Wang, X. (2021). Shifted dynamics of glucose metabolism in the hippocampus during aging. *Front. Aging Neurosci.* 13:700306. doi: 10.3389/fnagi.2021.700306
- Gelman, B. B. (2015). Neuropathology of HAND With suppressive antiretroviral therapy: encephalitis and neurodegeneration reconsidered. *Curr. HIV/AIDS Rep.* 12, 272–279. doi: 10.1007/s11904-015-0266-8
- Gilpin, N. W., Herman, M. A., and Roberto, M. (2015). The central amygdala as an integrative hub for anxiety and alcohol use disorders. *Biol. Psychiatry* 77, 859–869. doi: 10.1016/j.biopsych.2014.09.008
- Go, B. S., Sirohi, S., and Walker, B. M. (2020). The role of matrix metalloproteinase-9 in negative reinforcement learning and plasticity in alcohol dependence. *Addict. Biol.* 25:e12715. doi: 10.1111/adb.12715
- Godfrey, C., Bremer, A., Alba, D., Apovian, C., Koethe, J. R., Koliwad, S., et al. (2019). Obesity and fat metabolism in human immunodeficiency virus-infected individuals: immunopathogenic mechanisms and clinical implications. *J. Infect. Dis.* 220, 420–431. doi: 10.1093/infdis/jiz118
- Gonzaga, N. A., Do Vale, G. T., Parente, J. M., Yokota, R., De Martinis, B. S., Casarini, D. E., et al. (2018). Ethanol withdrawal increases blood pressure and vascular oxidative stress: a role for angiotensin type 1 receptors. *J. Am. Soc. Hypertens.* 12, 561–573. doi: 10.1016/j.jash.2018.03.012
- Govers, E. (2015). Obesity and insulin resistance are the central issues in prevention of and care for comorbidities. *Healthcare (Basel)* 3, 408–416. doi: 10.3390/healthcare3020408
- Grant, B. F., Goldstein, R. B., Smith, S. M., Jung, J., Zhang, H., Chou, S. P., et al. (2015). The alcohol use disorder and associated disabilities interview Schedule-5 (AUDADIS-5): reliability of substance use and psychiatric disorder modules in a general population sample. *Drug Alcohol Depend* 148, 27–33. doi: 10.1016/j.drugalcdep.2014.11.026
- Green, A., Garrick, T., Sheedy, D., Blake, H., Shores, E. A., and Harper, C. (2010). The effect of moderate to heavy alcohol consumption on neuropsychological performance as measured by the repeatable battery for the assessment of neuropsychological status. *Alcohol. Clin. Exp. Res.* 34, 443–450. doi: 10.1111/j.1530-0277.2009.01108.x
- Grunfeld, C., Pang, M., Doerrler, W., Shigenaga, J. K., Jensen, P., and Feingold, K. R. (1992). Lipids, lipoproteins, triglyceride clearance, and cytokines in

- human immunodeficiency virus infection and the acquired immunodeficiency syndrome. *J. Clin. Endocrinol. Metab.* 74, 1045–1052. doi: 10.1210/jcem.74.5.1373735
- Gu, M., Samuelson, D. R., Taylor, C. M., Molina, P. E., Luo, M., Siggins, R. W., et al. (2021). Alcohol-associated intestinal dysbiosis alters mucosal-associated invariant T-cell phenotype and function. *Alcohol. Clin. Exp. Res.* 45, 934–947. doi: 10.1111/acer.14589
- Guenther, P. M., Reedy, J., and Krebs-Smith, S. M. (2008). Development of the healthy eating Index-2005. *J. Am. Diet. Assoc.* 108, 1896–1901. doi: 10.1016/j.jada.2008.08.016
- Guindon, J., Blanton, H., Brauman, S., Donckels, K., Narasimhan, M., and Benamar, K. (2019). Sex differences in a rodent model of HIV-1-Associated neuropathic pain. *Int. J. Mol. Sci.* 20:1196. doi: 10.3390/ijms20051196
- Hammoud, D. A., Sinharay, S., Steinbach, S., Wakim, P. G., Geannopoulos, K., Traino, K., et al. (2018). Global and regional brain hypometabolism on FDG-PET in treated HIV-infected individuals. *Neurology* 91, e1591–e1601. doi: 10.1212/WNL.0000000000006398
- Harwood, H. J., and Jr. (2012). The adipocyte as an endocrine organ in the regulation of metabolic homeostasis. *Neuropharmacology* 63, 57–75. doi: 10.1016/j.neuropharm.2011.12.010
- Hirakawa, M., Arase, Y., Amakawa, K., Ohmoto-Sekine, Y., Ishihara, M., Shiba, M., et al. (2015). Relationship between alcohol intake and risk factors for metabolic syndrome in men. *Intern. Med.* 54, 2139–2145. doi: 10.2169/internalmedicine.54.2736
- Hong, J., Smith, R. R., Harvey, A. E., and Nunez, N. P. (2009). Alcohol consumption promotes insulin sensitivity without affecting body fat levels. *Int. J. Obes. (Lond)* 33, 197–203. doi: 10.1038/ijo.2008.266
- Hong, S. W., Linton, J. A., Shim, J. Y., and Kang, H. T. (2015). High-risk drinking is associated with a higher risk of diabetes mellitus in Korean men, based on the 2010–2012 KNHANES. *Alcohol* 49, 275–281. doi: 10.1016/j.alcohol.2015.02.004
- Hong-Brown, L. Q., Brown, C. R., Kazi, A. A., Navaratnarajah, M., and Lang, C. H. (2012). Rag GTPases and AMPK/TSC2/Rheb mediate the differential regulation of mTORC1 signaling in response to alcohol and leucine. *Am. J. Physiol. Cell Physiol.* 302, C1557–C1565. doi: 10.1152/ajpcell.00407.2011
- Hu, N., Zhang, Y., Nair, S., Culver, B. W., and Ren, J. (2014). Contribution of ALDH2 polymorphism to alcoholism-associated hypertension. *Recent Pat. Endocr. Metab. Immune Drug Discov.* 8, 180–185. doi: 10.2174/1872214808666141020162000
- Husain, K., Ansari, R. A., and Ferder, L. (2014). Alcohol-induced hypertension: mechanism and prevention. *World J. Cardiol.* 6, 245–252. doi: 10.4330/wjc.v6.i5.245
- Ikeda, M. L., Barcellos, N. T., Alencastro, P. R., Wolff, F. H., Brandao, A. B., Fuchs, F. D., et al. (2013). Association of blood pressure and hypertension with alcohol consumption in HIV-infected white and nonwhite patients. *ScientificWorldJournal* 2013:169825. doi: 10.1155/2013/169825
- Jan, V., Cervera, P., Maachi, M., Baudrimont, M., Kim, M., Vidal, H., et al. (2004). Altered fat differentiation and adipocytokine expression are inter-related and linked to morphological changes and insulin resistance in HIV-1-infected lipodystrophic patients. *Antivir. Ther.* 9, 555–564.
- Jankie, S., and Pinto Pereira, L. M. (2021). Targeting insulin resistance with selected antidiabetic agents prevents menopausal associated central obesity, dysglycemia, and cardiometabolic risk. *Post Reprod Health* 27, 45–48. doi: 10.1177/2053369120982753
- Jing, L., Li, W. M., Zhou, L. J., Li, S., Kou, J. J., and Song, J. (2008). Expression of renin-angiotensin system and peroxisome proliferator-activated receptors in alcoholic cardiomyopathy. *Alcohol. Clin. Exp. Res.* 32, 1999–2007. doi: 10.1111/j.1530-0277.2008.00781.x
- John, MacLennan, A., Leea, N., and Walker, D. W. (1995). Chronic ethanol administration decreases brain-derived neurotrophic factor gene expression in the rat hippocampus. *Neurosci. Lett.* 197, 105–108. doi: 10.1016/0304-3940(95)11922-J
- Kahl, K. G., and Hillemecher, T. (2016). The metabolic syndrome in patients with alcohol dependency: current research and clinical implications. *Prog. Neuropsychopharmacol. Biol. Psychiatry* 70, 49–56. doi: 10.1016/j.pnpb.2016.05.001
- Kalinowski, A., and Humphreys, K. (2016). Governmental standard drink definitions and low-risk alcohol consumption guidelines in 37 countries. *Addiction* 111, 1293–1298. doi: 10.1111/add.13341
- Kang, L., Chen, X., Sebastian, B. M., Pratt, B. T., Bederman, I. R., Alexander, J. C., et al. (2007a). Chronic ethanol and triglyceride turnover in white adipose tissue in rats: inhibition of the anti-lipolytic action of insulin after chronic ethanol contributes to increased triglyceride degradation. *J. Biol. Chem.* 282, 28465–28473. doi: 10.1074/jbc.M705503200
- Kang, L., Sebastian, B. M., Pritchard, M. T., Pratt, B. T., Previs, S. F., and Nagy, L. E. (2007b). Chronic ethanol-induced insulin resistance is associated with macrophage infiltration into adipose tissue and altered expression of adipocytokines. *Alcohol. Clin. Exp. Res.* 31, 1581–1588. doi: 10.1111/j.1530-0277.2007.00452.x
- Katz, P. S., Siggins, R. W., Porretta, C., Armstrong, M. L., Zea, A. H., Mercante, D. E., et al. (2015). Chronic alcohol increases CD8+ T-cell immunosenescence in simian immunodeficiency virus-infected rhesus macaques. *Alcohol* 49, 759–765. doi: 10.1016/j.alcohol.2015.09.003
- Kerr, W. C., and Ye, Y. (2010). Relationship of life-course drinking patterns to diabetes, heart problems, and hypertension among those 40 and older in the 2005 U.S. National Alcohol Survey. *J. Stud. Alcohol Drugs* 71, 515–525. doi: 10.15288/jsad.2010.71.515
- Kim, J. Y., Song, E. H., Lee, H. J., Oh, Y. K., Park, Y. S., Park, J. W., et al. (2010). Chronic ethanol consumption-induced pancreatic {beta}-cell dysfunction and apoptosis through glucokinase nitration and its down-regulation. *J. Biol. Chem.* 285, 37251–37262. doi: 10.1074/jbc.M110.142315
- Klaus, J. R., Hurwitz, B. E., Llabre, M. M., Skyler, J. S., Goldberg, R. B., Marks, J. B., et al. (2009). Central obesity and insulin resistance in the cardiometabolic syndrome: pathways to preclinical cardiovascular structure and function. *J. Cardiometab Syndr.* 4, 63–71. doi: 10.1111/j.1559-4572.2008.00038.x
- Kloner, R. A., and Rezkalla, S. H. (2007). To drink or not to drink? that is the question. *Circulation* 116, 1306–1317. doi: 10.1161/CIRCULATIONAHA.106.678375
- Klop, B., Do Rego, A. T., and Cabezas, M. C. (2013). Alcohol and plasma triglycerides. *Curr. Opin. Lipidol.* 24, 321–326. doi: 10.1097/MOL.0b013e3283606845
- Koloverou, E., Panagiotakos, D. B., Pitsavos, C., Chrysohoou, C., Georgousopoulou, E. N., Metaxa, V., et al. (2015). Effects of alcohol consumption and the metabolic syndrome on 10-year incidence of diabetes: the ATTICA study. *Diabetes Metab.* 41, 152–159. doi: 10.1016/j.diabet.2014.06.003
- Koob, G. F. (2021). Drug addiction: hyperkatifeia/negative reinforcement as a framework for medications development. *Pharmacol. Rev.* 73, 163–201. doi: 10.1124/pharmrev.120.000083
- Kopera, M., Wojnar, M., Brower, K., Glass, J., Nowosad, I., Gmaj, B., et al. (2012). Cognitive functions in abstinent alcohol-dependent patients. *Alcohol* 46, 665–671. doi: 10.1016/j.alcohol.2012.04.005
- Kovar, J., and Zemankova, K. (2015). Moderate alcohol consumption and triglyceridemia. *Physiol. Res.* 64, S371–S375. doi: 10.33549/physiolres.933178
- Lagathu, C., Kim, M., Maachi, M., Vigouroux, C., Cervera, P., Capeau, J., et al. (2005). HIV antiretroviral treatment alters adipokine expression and insulin sensitivity of adipose tissue *in vitro* and *in vivo*. *Biochimie* 87, 65–71. doi: 10.1016/j.biochi.2004.12.007
- Lai, Y. J., Hu, H. Y., Lee, Y. L., Ko, M. C., Ku, P. W., Yen, Y. F., et al. (2019). Frequency of alcohol consumption and risk of type 2 diabetes mellitus: a nationwide cohort study. *Clin. Nutr.* 38, 1368–1372. doi: 10.1016/j.clnu.2018.06.930
- Lang, C. H., Derdak, Z., and Wands, J. R. (2014). Strain-dependent differences for suppression of insulin-stimulated glucose uptake in skeletal and cardiac muscle by ethanol. *Alcohol. Clin. Exp. Res.* 38, 897–910. doi: 10.1111/acer.12343
- Langdon, K. J., Fox, A. B., King, L. A., King, D. W., Eisen, S., and Vogt, D. (2016). Examination of the dynamic interplay between posttraumatic stress symptoms and alcohol misuse among combat-exposed Operation Enduring Freedom (OEF)/Operation Iraqi Freedom (OIF) Veterans. *J. Affect. Disord.* 196, 234–242. doi: 10.1016/j.jad.2016.02.048
- Langkilde, A., Tavenier, J., Danielsen, A. V., Eugen-Olsen, J., Therkildsen, C., Jensen, F. K., et al. (2018). Histological and molecular adipose tissue changes are related to metabolic syndrome rather than lipodystrophy in human immunodeficiency virus-infected patients: a cross-sectional study. *J. Infect. Dis.* 218, 1090–1098. doi: 10.1093/infdis/jiy284
- Laniece, A., Lahbairi, N., Cabe, N., Pitel, A. L., and Rauchs, G. (2021). Contribution of sleep disturbances to the heterogeneity of cognitive and brain

- alterations in alcohol use disorder. *Sleep Med. Rev.* 58:101435. doi: 10.1016/j.smrv.2021.101435
- Lasek, A. W. (2016). Effects of ethanol on brain extracellular matrix: implications for alcohol use disorder. *Alcohol. Clin. Exp. Res.* 40, 2030–2042. doi: 10.1111/acer.13200
- Lattanzio, S. M., and Imbesi, F. (2018). Fibromyalgia syndrome: a case report on controlled remission of symptoms by a dietary strategy. *Front. Med. (Lausanne)* 5:94. doi: 10.3389/fmed.2018.00094
- Le Berre, A. P., Pitel, A. L., Chanraud, S., Beaunieux, H., Eustache, F., Martinot, J. L., et al. (2014). Chronic alcohol consumption and its effect on nodes of frontocerebellar and limbic circuitry: comparison of effects in France and the United States. *Hum. Brain Mapp.* 35, 4635–4653. doi: 10.1002/hbm.22500
- LeCapitaine, N. J., Wang, Z. Q., Dufour, J. P., Potter, B. J., Bagby, G. J., Nelson, S., et al. (2011). Disrupted anabolic and catabolic processes may contribute to alcohol-accentuated AIDS-associated wasting. *J. Infect. Dis.* 204, 1246–1255. doi: 10.1093/infdis/jir508
- Lee, K. (2012). Gender-specific relationships between alcohol drinking patterns and metabolic syndrome: the Korea National Health and Nutrition Examination Survey 2008. *Public Health Nutr.* 15, 1917–1924. doi: 10.1017/S136898001100365X
- Lee, S., and Kwak, H. B. (2014). Role of adiponectin in metabolic and cardiovascular disease. *J. Exerc. Rehabil.* 10, 54–59. doi: 10.12965/jer.140100
- Leserman, J. (2008). Role of depression, stress, and trauma in HIV disease progression. *Psychosom. Med.* 70, 539–545. doi: 10.1097/PSY.0b013e3181777a5f
- Li, C. S., Luo, X., Yan, P., Bergquist, K., and Sinha, R. (2009). Altered impulse control in alcohol dependence: neural measures of stop signal performance. *Alcohol. Clin. Exp. Res.* 33, 740–750. doi: 10.1111/j.1530-0277.2008.00891.x
- Lindtner, C., Scherer, T., Zielinski, E., Filatova, N., Fasshauer, M., Tonks, N. K., et al. (2013). Binge drinking induces whole-body insulin resistance by impairing hypothalamic insulin action. *Sci. Transl. Med.* 5:170ra114. doi: 10.1126/scitranslmed.3005123
- Livingston, N. A., Farmer, S. L., Mahoney, C. T., Marx, B. P., and Keane, T. M. (2021). The role of PTSD symptom clusters and criterion in predicting future high-risk drug and alcohol use among returning veteran men and women. *Psychol. Serv.* Online ahead of print. doi: 10.1037/ser0000538
- Longenecker, C. T., Jiang, Y., Yun, C. H., Debanne, S., Funderburg, N. T., Lederman, M. M., et al. (2013). Perivascular fat, inflammation, and cardiovascular risk in HIV-infected patients on antiretroviral therapy. *Int. J. Cardiol.* 168, 4039–4045. doi: 10.1016/j.ijcard.2013.06.059
- Lubbers, B. R., Smit, A. B., Spijker, S., and Van Den Oever, M. C. (2014). Neural ECM in addiction, schizophrenia, and mood disorder. *Prog. Brain Res.* 214, 263–284. doi: 10.1016/B978-0-444-63486-3.00012-8
- Maddur, H., and Shah, V. H. (2020). Alcohol and liver function in women. *Alcohol Res.* 40:10. doi: 10.35946/arcr.v40.2.10
- Maffei, V. J., Ferguson, T. F., Brashear, M. M., Mercante, D. E., Theall, K. P., Siggins, R. W., et al. (2020). Lifetime alcohol use among persons living with HIV is associated with frailty. *AIDS* 34, 245–254. doi: 10.1097/QAD.0000000000002426
- Maffei, V. J., Siggins, R. W., Luo, M., Brashear, M. M., Mercante, D. E., Taylor, C. M., et al. (2021). Alcohol use is associated with intestinal dysbiosis and dysfunctional CD8+ T-Cell phenotypes in persons with human immunodeficiency virus. *J. Infect. Dis.* 223, 1029–1039. doi: 10.1093/infdis/jiaa461
- Maiya, R. P., and Messing, R. O. (2014). Peripheral systems: neuropathy. *Handb. Clin. Neurol.* 125, 513–525. doi: 10.1016/B978-0-444-62619-6.00029-X
- Mangus, L. M., Dorsey, J. L., Laast, V. A., Hauer, P., Queen, S. E., Adams, R. J., et al. (2015). Neuroinflammation and virus replication in the spinal cord of simian immunodeficiency virus-infected macaques. *J. Neuropathol. Exp. Neurol.* 74, 38–47. doi: 10.1097/NEN.0000000000000148
- Martin, A. A., and Davidson, T. L. (2014). Human cognitive function and the obesogenic environment. *Physiol. Behav.* 136, 185–193. doi: 10.1016/j.physbeh.2014.02.062
- Masenga, S. K., Elijovich, F., Koethe, J. R., Hamooya, B. M., Heimburger, D. C., Munsaka, S. M., et al. (2020). Hypertension and metabolic syndrome in persons with HIV. *Curr. Hypertens. Rep.* 22:78. doi: 10.1007/s11906-020-01089-3
- Massey, V. L., Beier, J. I., Ritzenthaler, J. D., Roman, J., and Arteel, G. E. (2015). Potential role of the Gut/Liver/Lung axis in alcohol-induced tissue pathology. *Biomolecules* 5, 2477–2503. doi: 10.3390/biom5042477
- Maxi, J. K., Dean, M., Zabaleta, J., Reiss, K., Bagby, G. J., Nelson, S., et al. (2016). Chronic binge alcohol administration dysregulates hippocampal genes involved in immunity and neurogenesis in simian immunodeficiency virus-infected macaques. *Biomolecules* 6:43. doi: 10.3390/biom6040043
- Maxi, J. K., Mercante, D., Foret, B., Oberhelman, S., Ferguson, T. F., Bagby, G. J., et al. (2019). Chronic binge alcohol-associated differential brain region modulation of growth factor signaling pathways and neuroinflammation in simian immunodeficiency virus-infected male macaques. *Alcohol Alcohol.* 54, 477–486. doi: 10.1093/alcac/agz056
- Mayeux, J. P., Teng, S. X., Katz, P. S., Gilpin, N. W., and Molina, P. E. (2015). Traumatic brain injury induces neuroinflammation and neuronal degeneration that is associated with escalated alcohol self-administration in rats. *Behav. Brain Res.* 279, 22–30. doi: 10.1016/j.bbr.2014.10.053
- McDermott, K. A., Joyner, K. J., Hakes, J. K., Okey, S. A., and Cogle, J. R. (2018). Pain interference and alcohol, nicotine, and cannabis use disorder in a national sample of substance users. *Drug Alcohol Depend.* 186, 53–59. doi: 10.1016/j.drugalcdep.2018.01.011
- Michael, H., Mpfofana, T., Ramlall, S., and Oosthuizen, F. (2020). The role of brain derived neurotrophic factor in HIV-Associated neurocognitive disorder: from the bench-top to the bedside. *Neuropsychiatr. Dis. Treat* 16, 355–367. doi: 10.2147/NDT.S232836
- Mira, R. G., Tapia-Rojas, C., Perez, M. J., Jara, C., Vergara, E. H., Quintanilla, R. A., et al. (2019). Alcohol impairs hippocampal function: from NMDA receptor synaptic transmission to mitochondrial function. *Drug Alcohol Depend* 205:107628. doi: 10.1016/j.drugalcdep.2019.107628
- Mokuda, O., Tanaka, H., Hayashi, T., Ooka, H., Okazaki, R., and Sakamoto, Y. (2004). Ethanol stimulates glycogenolysis and inhibits both glycogenesis via gluconeogenesis and from exogenous glucose in perfused rat liver. *Ann. Nutr. Metab.* 48, 276–280. doi: 10.1159/000080463
- Molina, P. E., Bagby, G. J., and Nelson, S. (2014a). Biomedical consequences of alcohol use disorders in the HIV-infected host. *Curr. HIV Res.* 12, 265–275. doi: 10.2174/1570162X12666140721121849
- Molina, P. E., Gardner, J. D., Souza-Smith, F. M., and Whitaker, A. M. (2014b). Alcohol abuse: critical pathophysiological processes and contribution to disease burden. *Physiology (Bethesda)* 29, 203–215. doi: 10.1152/physiol.00055.2013
- Molina, P. E., McNurlan, M., Rathmacher, J., Lang, C. H., Zambell, K. L., Purcell, J., et al. (2006). Chronic alcohol accentuates nutritional, metabolic, and immune alterations during asymptomatic simian immunodeficiency virus infection. *Alcohol. Clin. Exp. Res.* 30, 2065–2078. doi: 10.1111/j.1530-0277.2006.00252.x
- Mouton, A. J., Ninh, V. K., El Hajj, E. C., El Hajj, M. C., Gilpin, N. W., and Gardner, J. D. (2016). Exposure to chronic alcohol accelerates development of wall stress and eccentric remodeling in rats with volume overload. *J. Mol. Cell Cardiol.* 97, 15–23. doi: 10.1016/j.yjmcc.2016.04.010
- Muhammad, J. N., Fernandez, J. R., Clay, O. J., Saag, M. S., Overton, E. T., and Willig, A. L. (2019). Associations of food insecurity and psychosocial measures with diet quality in adults aging with HIV. *AIDS Care* 31, 554–562. doi: 10.1080/09540121.2018.1554239
- Muscari, A., Bianchi, G., Conte, C., Forti, P., Magalotti, D., Pandolfi, P., et al. (2015). No direct survival effect of light to moderate alcohol drinking in community-dwelling older adults. *J. Am. Geriatr. Soc.* 63, 2526–2533. doi: 10.1111/jgs.13837
- Nahin, R. L. (2015). Estimates of pain prevalence and severity in adults: United States, 2012. *J. Pain* 16, 769–780. doi: 10.1016/j.jpain.2015.05.002
- Natsag, J., Erlandson, K. M., Sellmeyer, D. E., Haberman, S. A., Margolick, J., Jacobson, L. P., et al. (2017). HIV infection is associated with increased fatty infiltration of the thigh muscle with aging independent of fat distribution. *PLoS One* 12:e0169184. doi: 10.1371/journal.pone.0169184
- Ninh, V. K., El Hajj, E. C., Mouton, A. J., and Gardner, J. D. (2019). Prenatal alcohol exposure causes adverse cardiac extracellular matrix changes and dysfunction in neonatal mice. *Cardiovasc. Toxicol.* 19, 389–400. doi: 10.1007/s12012-018-09503-8
- Noumegni, S. R., Bigna, J. J., Ama Moor, Epse Nkegoum, V. J., Nansseu, J. R., Assah, F. K., et al. (2017). Relationship between estimated cardiovascular disease risk



- and insulin resistance in a black African population living with HIV: a cross-sectional study from Cameroon. *BMJ Open* 7:e016835. doi: 10.1136/bmjopen-2017-016835
- Nova, E., San Mauro-Martin, I., Diaz-Prieto, L. E., and Marcos, A. (2019). Wine and beer within a moderate alcohol intake is associated with higher levels of HDL-c and adiponectin. *Nutr. Res.* 63, 42–50. doi: 10.1016/j.nutres.2018.12.007
- Oda, N., Kajikawa, M., Maruhashi, T., Kishimoto, S., Yusoff, F. M., Goto, C., et al. (2020). Endothelial function is preserved in light to moderate alcohol drinkers but is impaired in heavy drinkers in women: flow-mediated Dilation Japan (FMD-J) study. *PLoS One* 15:e0243216. doi: 10.1371/journal.pone.0243216
- Okuno, F., Arai, M., Ishii, H., Shigeta, Y., Ebihara, Y., Takagi, S., et al. (1986). Mild but prolonged elevation of serum angiotensin converting enzyme (ACE) activity in alcoholics. *Alcohol* 3, 357–359. doi: 10.1016/0741-8329(86)90053-4
- Osna, N. A., Ganesan, M., Seth, D., Wyatt, T. A., Kidambi, S., and Kharbanda, K. K. (2021). Second hits exacerbate alcohol-related organ damage: an update. *Alcohol Alcohol* 56, 8–16. doi: 10.1093/alcac/aga085
- Pacher, P., Beckman, J. S., and Liaudet, L. (2007). Nitric oxide and peroxynitrite in health and disease. *Physiol. Rev.* 87, 315–424. doi: 10.1152/physrev.00029.2006
- Panza, F., Frisardi, V., Capurso, C., Imbimbo, B. P., Vendemiale, G., Santamato, A., et al. (2010). Metabolic syndrome and cognitive impairment: current epidemiology and possible underlying mechanisms. *J. Alzheimers. Dis.* 21, 691–724. doi: 10.3233/JAD-2010-091669
- Pasipanodya, E. C., Montoya, J. L., Campbell, L. M., Hussain, M. A., Saloner, R., Paolillo, E. M., et al. (2021). Metabolic risk factors as differential predictors of profiles of neurocognitive impairment among older HIV+ and HIV- Adults: an observational study. *Arch. Clin. Neuropsychol.* 36, 151–164. doi: 10.1093/arclin/acz040
- Patto, R. J., Russo, E. K., Borges, D. R., and Neves, M. M. (1993). The enteroinsular axis and endocrine pancreatic function in chronic alcohol consumers: evidence for early beta-cell hypofunction. *Mt. Sinai J. Med.* 60, 317–320.
- Perkins, A. E., Varlinskaya, E. I., and Deak, T. (2019). From adolescence to late aging: a comprehensive review of social behavior, alcohol, and neuroinflammation across the lifespan. *Int. Rev. Neurobiol.* 148, 231–303. doi: 10.1016/bs.irm.2019.08.001
- Pfefferbaum, A., Sullivan, E. V., Rosenbloom, M. J., Mathalon, D. H., and Lim, K. O. (1998). A controlled study of cortical gray matter and ventricular changes in alcoholic men over a 5-year interval. *Arch. Gen. Psychiatry* 55, 905–912. doi: 10.1001/archpsyc.55.10.905
- Phillips, T. J. C., Brown, M., Ramirez, J. D., Perkins, J., Woldeamanuel, Y. W., Williams, A. C. C., et al. (2014). Sensory, psychological, and metabolic dysfunction in HIV-associated peripheral neuropathy: a cross-sectional deep profiling study. *Pain* 155, 1846–1860. doi: 10.1016/j.pain.2014.06.014
- Piggott, D. A., Erlandson, K. M., and Yarasheski, K. E. (2016). Frailty in HIV: epidemiology, biology, measurement, interventions, and research needs. *Curr. HIV/AIDS Rep.* 13, 340–348. doi: 10.1007/s11904-016-0334-8
- Poole, L. G., and Arteel, G. E. (2016). Transitional remodeling of the hepatic extracellular matrix in alcohol-induced liver injury. *Biomed. Res. Int.* 2016:3162670. doi: 10.1155/2016/3162670
- Pope, C. N., Montoya, J. L., Vasquez, E., Perez-Santiago, J., Ellis, R., Mccutchan, J. A., et al. (2020). Association of HIV serostatus and metabolic syndrome with neurobehavioral disturbances. *J. Neurovirol.* 26, 888–898. doi: 10.1007/s13365-020-00878-5
- Poznyak, V. (2018). *Global Status Report on Alcohol and Health 2018*. Switzerland: World Health Organization.
- Primeaux, S. D., Simon, L., Ferguson, T. F., Levitt, D. E., Brashear, M. M., Yeh, A., et al. (2021). Alcohol use and dysglycemia among people living with human immunodeficiency virus (HIV) in the alcohol & metabolic comorbidities in PLWH: evidence driven interventions (ALIVE-Ex) study. *Alcohol. Clin. Exp. Res.* 45, 1735–1746. doi: 10.1111/acer.14667
- Puddey, I. B., Zilkens, R. R., Croft, K. D., and Beilin, L. J. (2001). Alcohol and endothelial function: a brief review. *Clin. Exp. Pharmacol. Physiol.* 28, 1020–1024. doi: 10.1046/j.1440-1681.2001.03572.x
- Rasineni, K., Srinivasan, M. P., Balamurugan, A. N., Kaphalia, B. S., Wang, S., Ding, W. X., et al. (2020). Recent advances in understanding the complexity of alcohol-induced pancreatic dysfunction and pancreatitis development. *Biomolecules* 10:669. doi: 10.3390/biom10050669
- Rasineni, K., Thomes, P. G., Kubik, J. L., Harris, E. N., Kharbanda, K. K., and Casey, C. A. (2019). Chronic alcohol exposure alters circulating insulin and ghrelin levels: role of ghrelin in hepatic steatosis. *Am. J. Physiol. Gastrointest. Liver Physiol.* 316, G453–G461. doi: 10.1152/ajpgi.00334.2018
- Rasouli, B., Ahlbom, A., Andersson, T., Grill, V., Midthjell, K., Olsson, L., et al. (2013). Alcohol consumption is associated with reduced risk of Type 2 diabetes and autoimmune diabetes in adults: results from the Nord-Trøndelag health study. *Diabet. Med.* 30, 56–64. doi: 10.1111/j.1464-5491.2012.03713.x
- Ray, L. A., Green, R., and Grodin, E. (2021). Neurobiological models of alcohol use disorder in humans. *Am. J. Psychiatry* 178, 483–484. doi: 10.1176/appi.ajp.2021.21030310
- Rekik, M., El-Mas, M. M., Mustafa, J. S., and Abdel-Rahman, A. A. (2002). Role of endothelial adenosine receptor-mediated vasorelaxation in ethanol-induced hypotension in hypertensive rats. *Eur. J. Pharmacol.* 452, 205–214. doi: 10.1016/S0014-2999(02)02304-X
- Rethy, L., Feinstein, M. J., Sinha, A., Achenbach, C., and Shah, S. J. (2020). Coronary microvascular dysfunction in HIV: a review. *J. Am. Heart Assoc.* 9, e014018. doi: 10.1161/JAHA.119.014018
- Riddler, S. A., Smit, E., Cole, S. R., Li, R., Chmiel, J. S., Dobs, A., et al. (2003). Impact of HIV infection and HAART on serum lipids in men. *JAMA* 289, 2978–2982. doi: 10.1001/jama.289.22.2978
- Riserus, U., and Ingelsson, E. (2007). Alcohol intake, insulin resistance, and abdominal obesity in elderly men. *Obesity (Silver Spring)* 15, 1766–1773. doi: 10.1038/oby.2007.210
- Roerecke, M., and Rehm, J. (2014). Cause-specific mortality risk in alcohol use disorder treatment patients: a systematic review and meta-analysis. *Int. J. Epidemiol.* 43, 906–919. doi: 10.1093/ije/dyu018
- Rubio-Araiz, A., Porcu, F., Perez-Hernandez, M., Garcia-Gutierrez, M. S., Aracil-Fernandez, M. A., Gutierrez-Lopez, M. D., et al. (2017). Disruption of blood-brain barrier integrity in postmortem alcoholic brain: preclinical evidence of TLR4 involvement from a binge-like drinking model. *Addict. Biol.* 22, 1103–1116. doi: 10.1111/adb.12376
- Saljoughian, M. (2016). Cardiometabolic syndrome: a global health issue. *US Pharmacist* 41, 19–21.
- Saloner, R., Campbell, L. M., Serrano, V., Montoya, J. L., Pasipanodya, E., Paolillo, E. W., et al. (2019). Neurocognitive SuperAging in older adults living with HIV: demographic, neuromedical and everyday functioning correlates. *J. Int. Neuropsychol. Soc.* 25, 507–519. doi: 10.1017/S135617719000018
- Schachinger, V., and Zeiher, A. M. (2000). Atherosclerosis-associated endothelial dysfunction. *Z Kardiol.* 89(Suppl. 9), IX/70–74. doi: 10.1007/s003920070033
- Schank, M., Zhao, J., Moorman, J. P., and Yao, Z. Q. (2021). The impact of HIV- and ART-Induced mitochondrial dysfunction in cellular senescence and aging. *Cells* 10:174. doi: 10.3390/cells10010174
- Schindler, A. G., Baskin, B., Juarez, B., Janet Lee, S., Hendrickson, R., Pagulayan, K., et al. (2021). Repetitive blast mild traumatic brain injury increases ethanol sensitivity in male mice and risky drinking behavior in male combat veterans. *Alcohol. Clin. Exp. Res.* 45, 1051–1064. doi: 10.1111/acer.14605
- Schrieks, I. C., Heil, A. L., Hendriks, H. F., Mukamal, K. J., and Beulens, J. W. (2015). The effect of alcohol consumption on insulin sensitivity and glycemic status: a systematic review and meta-analysis of intervention studies. *Diabetics Care* 38, 723–732.
- Schroder, H., Morales-Molina, J. A., Bermejo, S., Barral, D., Mandoli, E. S., Grau, M., et al. (2007). Relationship of abdominal obesity with alcohol consumption at population scale. *Eur. J. Nutr.* 46, 369–376. doi: 10.1007/s00394-007-0674-7
- Senkov, O., Andjus, P., Radenovic, L., Soriano, E., and Dityatev, A. (2014). Neural ECM molecules in synaptic plasticity, learning, and memory. *Prog. Brain Res.* 214, 53–80. doi: 10.1016/B978-0-444-63486-3.00003-7
- Seth, D., D'souza El-Guindy, N. B., Apte, M., Mari, M., Dooley, S., Neuman, M., et al. (2010). Alcohol, signaling, and ECM turnover. *Alcohol. Clin. Exp. Res.* 34, 4–18. doi: 10.1111/j.1530-0277.2009.01060.x
- Shukla, P. K., Meena, A. S., Dalal, K., Canelas, C., Samak, G., Pierre, J. F., et al. (2021). Chronic stress and corticosterone exacerbate alcohol-induced tissue injury in the gut-liver-brain axis. *Sci. Rep.* 11:826. doi: 10.1038/s41598-020-80637-y
- Shuval, K., Finley, C. E., Chartier, K. G., Balasubramanian, B. A., Gabriel, K. P., and Barlow, C. E. (2012). Cardiorespiratory fitness, alcohol intake, and metabolic syndrome incidence in men. *Med. Sci. Sports Exerc.* 44, 2125–2131. doi: 10.1249/MSS.0b013e3182640c4e
- Silva-Pena, D., Garcia-Marchena, N., Alen, F., Araos, P., Rivera, P., Vargas, A., et al. (2019). Alcohol-induced cognitive deficits are associated with decreased



- circulating levels of the neurotrophin BDNF in humans and rats. *Addict. Biol.* 24, 1019–1033. doi: 10.1111/adb.12668
- Simon, L., Ferguson, T. F., Vande Stouwe, C., Brashear, M. M., Primeaux, S. D., Theall, K. P., et al. (2020). Prevalence of insulin resistance in adults living with HIV: implications of alcohol use. *AIDS Res. Hum. Retroviruses* 36, 742–752. doi: 10.1089/aid.2020.0029
- Simon, L., Hollenbach, A. D., Zabaleta, J., and Molina, P. E. (2015). Chronic binge alcohol administration dysregulates global regulatory gene networks associated with skeletal muscle wasting in simian immunodeficiency virus-infected macaques. *BMC Genomics* 16:1097. doi: 10.1186/s12864-015-2329-z
- Simon, L., Lecapitaine, N., Berner, P., Vande Stouwe, C., Mussell, J. C., Allerton, T., et al. (2014). Chronic binge alcohol consumption alters myogenic gene expression and reduces *in vitro* myogenic differentiation potential of myoblasts from rhesus macaques. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 306, R837–R844. doi: 10.1152/ajpregu.00502.2013
- Simon, L., Torres, D., Saravia, A., Levitt, D. E., Vande Stouwe, C., Mcgarrah, H., et al. (2021). Chronic binge alcohol and ovariectomy-mediated impaired insulin responsiveness in SIV-infected female rhesus macaques. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 321, R699–R711. doi: 10.1152/ajpregu.00159.2021
- Somkuwar, S. S., Fannon, M. J., Staples, M. C., Zamora-Martinez, E. R., Navarro, A. I., Kim, A., et al. (2016). Alcohol dependence-induced regulation of the proliferation and survival of adult brain progenitors is associated with altered BDNF-TrkB signaling. *Brain Struct. Funct.* 221, 4319–4335. doi: 10.1007/s00429-015-1163-z
- Sotos-Prieto, M., Bhupathiraju, S. N., and Hu, F. B. (2017a). Changes in diet quality and total and cause-specific mortality. *N. Engl. J. Med.* 377, 1304–1305.
- Sotos-Prieto, M., Bhupathiraju, S. N., Mattei, J., Fung, T. T., Li, Y., Pan, A., et al. (2017b). Association of changes in diet quality with total and cause-specific mortality. *N. Engl. J. Med.* 377, 143–153. doi: 10.1056/NEJMoa1613502
- Souza-Smith, F. M., Lang, C. H., Nagy, L. E., Bailey, S. M., Parsons, L. H., and Murray, G. J. (2016). Physiological processes underlying organ injury in alcohol abuse. *Am. J. Physiol. Endocrinol. Metab.* 311, E605–E619. doi: 10.1152/ajpendo.00270.2016
- Srdic, D., Khawla, A. M., Soldatovic, I., Nikolic, J., Jevtic, D., Nair, D., et al. (2017). Correlation of leptin, adiponectin, and resistin levels in different types of lipodystrophy in HIV/AIDS patients. *Metab. Syndr. Relat. Disord.* 15, 153–159. doi: 10.1089/met.2016.0068
- Srinivasa, S., Fitch, K. V., Wong, K., Torriani, M., Mayhew, C., Stanley, T., et al. (2015). RAAS activation is associated with visceral adiposity and insulin resistance among HIV-infected patients. *J. Clin. Endocrinol. Metab.* 100, 2873–2882. doi: 10.1210/jc.2015-1461
- Stankov, M. V., and Behrens, G. M. (2010). Contribution of inflammation to fat redistribution and metabolic disturbances in HIV-1 infected patients. *Curr. Pharm. Des* 16, 3361–3371. doi: 10.2174/138161210793563473
- Steiner, J. L., and Lang, C. H. (2017). Alcohol, adipose tissue and lipid dysregulation. *Biomolecules* 7:16. doi: 10.3390/biom7010016
- Steiner, J. L., and Lang, C. H. (2019). Ethanol acutely antagonizes the refeeding-induced increase in mTOR-dependent protein synthesis and decrease in autophagy in skeletal muscle. *Mol. Cell. Biochem.* 456, 41–51. doi: 10.1007/s11010-018-3488-4
- Steiner, J. L., Crowell, K. T., and Lang, C. H. (2015a). Impact of alcohol on glycemic control and insulin action. *Biomolecules* 5, 2223–2246. doi: 10.3390/biom5042223
- Steiner, J. L., Pruznak, A. M., Navaratnarajah, M., and Lang, C. H. (2015b). Alcohol differentially alters extracellular matrix and adhesion molecule expression in skeletal muscle and heart. *Alcohol. Clin. Exp. Res.* 39, 1330–1340. doi: 10.1111/acer.12771
- Sueblinvong, V., Kerchberger, V. E., Saghafi, R., Mills, S. T., Fan, X., and Guidot, D. M. (2014). Chronic alcohol ingestion primes the lung for bleomycin-induced fibrosis in mice. *Alcohol. Clin. Exp. Res.* 38, 336–343. doi: 10.1111/acer.12232
- Sullivan, E. V., and Pfefferbaum, A. (2019). Brain-behavior relations and effects of aging and common comorbidities in alcohol use disorder: a review. *Neuropsychology* 33, 760–780. doi: 10.1037/neu0000557
- Sumi, M., Hisamatsu, T., Fujiyoshi, A., Kadota, A., Miyagawa, N., Kondo, K., et al. (2019). Association of alcohol consumption with fat deposition in a community-based sample of Japanese men: the Shiga Epidemiological Study of Subclinical Atherosclerosis (SESSA). *J. Epidemiol.* 29, 205–212. doi: 10.2188/jea.JE20170191
- Teratani, T., Morimoto, H., Sakata, K., Oishi, M., Tanaka, K., Nakada, S., et al. (2012). Dose-response relationship between tobacco or alcohol consumption and the development of diabetes mellitus in Japanese male workers. *Drug Alcohol Depend* 125, 276–282. doi: 10.1016/j.drugalcdep.2012.03.002
- Thompson, T., Oram, C., Correll, C. U., Tsermentseli, S., and Stubbs, B. (2017). Analgesic effects of alcohol: a systematic review and meta-analysis of controlled experimental studies in healthy participants. *J. Pain* 18, 499–510. doi: 10.1016/j.jpain.2016.11.009
- Toda, N., and Ayajiki, K. (2010). Vascular actions of nitric oxide as affected by exposure to alcohol. *Alcohol Alcohol.* 45, 347–355. doi: 10.1093/alcal/agg028
- Tolomeo, S., Macfarlane, J. A., Baldacchino, A., Koob, G. F., and Steele, J. D. (2021). Alcohol binge drinking: negative and positive valence system abnormalities. *Biol. Psychiatry Cogn. Neurosci. Neuroimaging* 6, 126–134. doi: 10.1016/j.bpsc.2020.09.010
- Totsch, S. K., Meir, R. Y., Quinn, T. L., Lopez, S. A., Gower, B. A., and Sorge, R. E. (2018). Effects of a standard american diet and an anti-inflammatory diet in male and female mice. *Eur. J. Pain* 22, 1203–1213. doi: 10.1002/ejp.1207
- Trindade, P., Hampton, B., Manhaes, A. C., and Medina, A. E. (2016). Developmental alcohol exposure leads to a persistent change on astrocyte secretome. *J. Neurochem.* 137, 730–743. doi: 10.1111/jnc.13542
- Tsuji, S., Kawano, S., Michida, T., Masuda, E., Nagano, K., Takei, Y., et al. (1992). Ethanol stimulates immunoreactive endothelin-1 and -2 release from cultured human umbilical vein endothelial cells. *Alcohol. Clin. Exp. Res.* 16, 347–349. doi: 10.1111/j.1530-0277.1992.tb01389.x
- Vasdev, S., Gill, V., and Singal, P. K. (2006). Beneficial effect of low ethanol intake on the cardiovascular system: possible biochemical mechanisms. *Vasc. Health Risk Manag.* 2, 263–276. doi: 10.2147/vhrm.2006.2.3.263
- Venkov, C. D., Myers, P. R., Tanner, M. A., Su, M., and Vaughan, D. E. (1999). Ethanol increases endothelial nitric oxide production through modulation of nitric oxide synthase expression. *Thromb Haemost* 81, 638–642. doi: 10.1055/s-0037-1614538
- Vieira, B. A., Luft, V. C., Schmidt, M. I., Chambless, L. E., Chor, D., Barreto, S. M., et al. (2016). Timing and type of alcohol consumption and the metabolic syndrome - ELSA-Brasil. *PLoS One* 11:e0163044. doi: 10.1371/journal.pone.0163044
- Volkow, N. D., Kim, S. W., Wang, G. J., Alexoff, D., Logan, J., Muench, L., et al. (2013). Acute alcohol intoxication decreases glucose metabolism but increases acetate uptake in the human brain. *Neuroimage* 64, 277–283. doi: 10.1016/j.neuroimage.2012.08.057
- Wakabayashi, I. (2014). Frequency of heavy alcohol drinking and risk of metabolic syndrome in middle-aged men. *Alcohol. Clin. Exp. Res.* 38, 1689–1696. doi: 10.1111/acer.12425
- Wakabayashi, I. (2019). Relationships between alcohol intake and cardiovascular risk factors in middle-aged men with hypo-HDL cholesterolemia. *Clin. Chim. Acta* 495, 94–99. doi: 10.1016/j.cca.2019.04.006
- Wang, S., Luo, Y., Feng, A., Li, T., Yang, X., Nofech-Mozes, R., et al. (2014). Ethanol induced impairment of glucose metabolism involves alterations of GABAergic signaling in pancreatic beta-cells. *Toxicology* 326, 44–52. doi: 10.1016/j.tox.2014.10.005
- Wang, Z. T., Li, K. Y., Tan, C. C., Xu, W., Shen, X. N., Cao, X. P., et al. (2021). Associations of alcohol consumption with cerebrospinal fluid biomarkers of Alzheimer's disease pathology in cognitively intact older adults: the CABLE study. *J. Alzheimers. Dis.* 82, 1045–1054. doi: 10.3233/JAD-210140
- Wardell, J. D., Shuper, P. A., Rourke, S. B., and Hendershot, C. S. (2018). Stigma, coping, and alcohol use severity among people living with HIV: a prospective analysis of bidirectional and mediated associations. *Ann. Behav. Med.* 52, 762–772. doi: 10.1093/abm/kax050
- Warnholtz, A., Nickenig, G., Schulz, E., Macharzina, R., Brasen, J. H., Skatchkov, M., et al. (1999). Increased NADH-oxidase-mediated superoxide production in the early stages of atherosclerosis: evidence for involvement of the renin-angiotensin system. *Circulation* 99, 2027–2033. doi: 10.1161/01.CIR.99.15.2027
- Webel, A. R., Sattar, A., Funderburg, N. T., Kinley, B., Longenecker, C. T., Labbato, D., et al. (2017). Alcohol and dietary factors associate with gut integrity and inflammation in HIV-infected adults. *HIV Med.* 18, 402–411. doi: 10.1111/hiv.12442

- Weiss, J. J., Sanchez, L., Hubbard, J., Lo, J., Grinspoon, S. K., and Fitch, K. V. (2019). Diet quality is low and differs by sex in people with HIV. *J. Nutr.* 149, 78–87. doi: 10.1093/jn/nxy241
- Wetterling, T., Veltrup, C., Driessen, M., and John, U. (1999). Drinking pattern and alcohol-related medical disorders. *Alcohol Alcohol.* 34, 330–336. doi: 10.1093/alcalc/34.3.330
- Whitaker, A. M., Gilpin, N. W., and Edwards, S. (2014). Animal models of post-traumatic stress disorder and recent neurobiological insights. *Behav. Pharmacol.* 25, 398–409. doi: 10.1097/FBP.0000000000000069
- Willig, A. L., and Overton, E. T. (2016). Metabolic complications and glucose metabolism in HIV infection: a review of the evidence. *Curr. HIV/AIDS Rep.* 13, 289–296. doi: 10.1007/s11904-016-0330-z
- Winsauer, P. J., Moerschbaecher, J. M., Brauner, I. N., Purcell, J. E., Lancaster, J. R. Jr., et al. (2002). Alcohol unmasks simian immunodeficiency virus-induced cognitive impairments in rhesus monkeys. *Alcohol. Clin. Exp. Res.* 26, 1846–1857. doi: 10.1111/j.1530-0277.2002.tb02492.x
- Witkiewitz, K., Mccallion, E., Vowles, K. E., Kirouac, M., Frohe, T., Maisto, S. A., et al. (2015a). Association between physical pain and alcohol treatment outcomes: the mediating role of negative affect. *J. Consult. Clin. Psychol.* 83, 1044–1057. doi: 10.1037/ccp0000033
- Witkiewitz, K., Vowles, K. E., Mccallion, E., Frohe, T., Kirouac, M., and Maisto, S. A. (2015b). Pain as a predictor of heavy drinking and any drinking lapses in the COMBINE study and the UK Alcohol Treatment Trial. *Addiction* 110, 1262–1271. doi: 10.1111/add.12964
- Woldu, M., Minzi, O., and Engidawork, E. (2020). Prevalence of cardiometabolic syndrome in HIV-infected persons: a systematic review. *J. Diabetes Metab. Disord.* 19, 1671–1683. doi: 10.1007/s40200-020-00552-x
- Wood, A. M., Kaptoge, S., Butterworth, A. S., Willeit, P., Warnakula, S., Bolton, T., et al. (2018). Risk thresholds for alcohol consumption: combined analysis of individual-participant data for 599 912 current drinkers in 83 prospective studies. *Lancet* 391, 1513–1523. doi: 10.1016/S0140-6736(18)30134-X
- World Health Organization (2021). *Global Health Observatory Data Repository*. Geneva: World Health Organization.
- Xu, Y., Chen, X., and Wang, K. (2017). Global prevalence of hypertension among people living with HIV: a systematic review and meta-analysis. *J. Am. Soc. Hypertens.* 11, 530–540. doi: 10.1016/j.jash.2017.06.004
- Yaffe, K., Kanaya, A., Lindquist, K., Simonsick, E. M., Harris, T., Shorr, R. I., et al. (2004). The metabolic syndrome, inflammation, and risk of cognitive decline. *JAMA* 292, 2237–2242. doi: 10.1001/jama.292.18.2237
- Yarlioglu, M., Yalcinkaya, D., Oksuz, F., Celik, I. E., Duran, M., and Murat, S. N. (2019). Possible effect of alcohol consumption on aortic dilatation by inducing renin-angiotensin-aldosterone system. *Angiology* 70, 978–979. doi: 10.1177/0003319719857381
- Yeomans, M. R., Caton, S., and Hetherington, M. M. (2003). Alcohol and food intake. *Curr. Opin. Clin. Nutr. Metab. Care* 6, 639–644. doi: 10.1097/00075197-200311000-00006
- Yeung, E. W., Lee, M. R., Mcdowell, Y., Sher, K. J., and Gizer, I. R. (2020). The association between alcohol consumption and pain interference in a nationally representative sample: the moderating roles of gender and alcohol use disorder symptomatology. *Alcohol. Clin. Exp. Res.* 44, 645–659. doi: 10.1111/acer.14282
- Yki-Jarvinen, H., Koivisto, V. A., Ylikahri, R., and Taskinen, M. R. (1988). Acute effects of ethanol and acetate on glucose kinetics in normal subjects. *Am. J. Physiol.* 254, E175–E180. doi: 10.1152/ajpendo.1988.254.2.E175
- You, M., and Arteel, G. E. (2019). Effect of ethanol on lipid metabolism. *J. Hepatol.* 70, 237–248. doi: 10.1016/j.jhep.2018.10.037
- Yu, B., Pasipanodya, E., Montoya, J. L., Moore, R. C., Gianella, S., Mccutchan, A., et al. (2019). Metabolic syndrome and neurocognitive deficits in HIV infection. *J. Acquir. Immune Defic. Syndr.* 81, 95–101. doi: 10.1097/QAI.0000000000001964
- Zahr, N. M. (2018). The aging brain with HIV infection: effects of alcoholism or hepatitis C comorbidity. *Front. Aging Neurosci.* 10:56. doi: 10.3389/fnagi.2018.00056
- Zale, E. L., Powers, J. M., and Ditre, J. W. (2021). Cognitive-Affective transdiagnostic factors associated with vulnerability to alcohol and prescription opioid use in the context of pain. *Alcohol. Res.* 41:08. doi: 10.35946/arc.v41.1.08
- Zhao, C., Liu, Y., Xiao, J., Liu, L., Chen, S., Mohammadi, M., et al. (2015). FGF21 mediates alcohol-induced adipose tissue lipolysis by activation of systemic release of catecholamine in mice. *J. Lipid Res.* 56, 1481–1491. doi: 10.1194/jlr.M058610
- Zhong, W., Zhao, Y., Tang, Y., Wei, X., Shi, X., Sun, W., et al. (2012). Chronic alcohol exposure stimulates adipose tissue lipolysis in mice: role of reverse triglyceride transport in the pathogenesis of alcoholic steatosis. *Am. J. Pathol.* 180, 998–1007. doi: 10.1016/j.ajpath.2011.11.017
- Zieman, S. J., Melenovsky, V., and Kass, D. A. (2005). Mechanisms, pathophysiology, and therapy of arterial stiffness. *Arterioscler Thromb. Vasc. Biol.* 25, 932–943. doi: 10.1161/01.ATV.0000160548.78317.29

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